Supplementary Figures

Figure S1. Distribution of renal cells in our combined cohort by (a) known sampling location, (b) sequencing technology, and (c) donor sex. Cells with unknown sampling location are excluded from this figure.





Figure S2: Uniform manifold approximation and projection for dimension reduction (UMAP) and t-distributed stochastic neighbor embedding (t-SNE) visualizations of each dataset generated using the code provided by authors in their publication or sent to us directly for (A) Lake et al., (B) Liao et al., (C) Menon et al., (D) Wu et al., (E) Young et al.

(A)



(B)









(E)



Figure S3. Barcharts showing (a) the distribution of cell counts by harmonized cell type and (b) the proportion of cells from each study for each harmonized cell type.

(A)



Harmonized Cell Type



(B)



Figure S4. UMAP visualizations of the combined dataset (a) before batch correction, (b) after batch correction, and (c) after batch correction colored by harmonized cell type.



(B)

Cell Type Monocytes, Macrophages & Other Myeloid 10 Neutrophil **Proximal Tubule** Mast 5 -Urothelium B, Plasma, & Plasmacytoid intUMAP_2 0 Parietal Epithelium, Late Proximal Tubule & Descending Loop of Henle 10 Ascending Loop of Henle Natural Killer & T Distal Convoluted & Connecting Tubules Raw -5 Podocyte Principal Endothelium 140 Perivascular & Mesangium -10 -15 o intUMAP_1 -10 -5 10 5

(C)

Figure S5: Schematic of SVM outlier detection with 4,257 cells removed from further analysis due to poor alignment with their original annotated cell type. UMAP of cells within the "Proximal Tubule" harmonized cell type before (left) and after (right) SVM outlier detection. Cells circled in red are poorly aligned with the remaining cells of their harmonized cell type, a trend observed with all other harmonized cell types.



Contribution to Harmonized Cell Type Counts by Study									
Harmonized Cell Type	Lake et al.	Liao et al.	Menon et al.	Wu et al.	Young et al.	Total			
Ascending Loop of Henle	3620	0	1835	581	0	6036			
B, Plasma, & Plasmacytoid	0	42	129	0	69	240			
Distal Convoluted & Connecting Tubules	344	309	745	183	0	1581			
Endothelium	717	0	2308	0	1956	4981			
Fibroblasts	191	0	51	0	28	270			
Intercalated	976	48	1336	215	153	2728			
Mast	0	0	0	0	22	22			
Monocytes, Macrophages, & Other Myeloid	16	490	1307	0	616	2429			
Natural Killer & T	0	351	3582	0	1672	5605			
Neutrophil	0	0	0	0	77	77			
Parietal Epithelium, Late Proximal Tubule, & Descending Loop of Henle	980	71	738	2230	568	4587			
Perivascular & Mesangium	265	0	654	0	107	1026			
Podocytes	468	0	174	138	33	813			
Principal	2157	82	1429	426	59	4153			
Proximal Tubule	1950	14157	6639	0	84	22830			
Urothelium	0	0	0	0	486	486			

 Table S1. Distribution of 57,864 renal cells used in our analyses, following SVM-based exclusion of low-quality cells.

Figure S6. Heatmap of each classifier's rejection rate on (a) Menon et al., (b) Lake et al., (c) Liao et al. and (d) Wu et al with respect to each cell type.

(a)

	XGB	RF	MLP	KNN	SVC	
Ascending Loop of Henle	1.1	3.2	0.82	1.1	0.44	5
B, Plasma, & Plasmacytoid	1.6	1.6	2.3	0	0.78	
Distal Convoluted & Connecting Tubules	1.6	3.4	3.1	2.3	1.3	4
Endothelium	0.35	0.34	0.56	0.26	0.087	
Fibrobasts	2	2	5.9	2	0	
Intercalated	1.9	4.4	4.6	2.6	1.6	-3
Monocyte	0.38	0.61	0.54	0	0	
Natural Killer & T	0.028	0.22	0.28	0.056	0.056	2
Parietal Epithelium, Late Proximal Tubule, & Descending Loop of Henle	1.6	5.1	2	4.6	0.81	-
Perivascular & Mesangium	1.2	3.2	1.8	1.2	0.31	
Podocytes		3.4	3.4		3.4	1
Principal	0.98	1.3	0.77	0.42	0.28	
Proximal Tubule	0.1	0.24	0.27	0.03	0.075	0

(b)

	XGB	RF	MLP	KNN	SVC	
Ascending Loop of Henle	16	7.8	6.9		0.88	
Distal Convoluted & Connecting Tubules	0.29	3.2	3.5	0.58	1.2	14
Endothelium	2.4	2.1	2.2	1.3	0.42	12
Fibrobasts	4.2	18	13	4.7	7.9	10
Intercalated	0.82		1.8	2	1.3	10
Monocytes, Macrophages, & Other Myeloid	12	6.2	0	0	6.2	8
Parietal Epithelium, Late Proximal Tubule, & Descending Loop of Henle	6.6	3.7	5.8	4.8	1.8	6
Perivascular & Mesangium	0	4.2	1.5	0	0.76	Ŭ
Podocytes	3.6	8.3	5.1	1.9	4.9	4
Principal	1.6	3.7	2.5	1	1.1	2
Proximal Tubule	1.5	2	4.9	0.51	2.1	
						-0

(c)

	XGB	RF	MLP	KNN	SVC	
B, Plasma, & Plasmacytoid	0	4.8	2.4	0	0	7
Distal Convoluted & Connecting Tubules	0	0	0	0	0	6
Intercalated	0	0	2.1	0	0	5
Monocytes, Macrophages, & Other Myeloid	0.83	0.82		0.82	0.82	4
Natural Killer & T	0.28	3.1		0.28	0.28	3
Parietal Epithelium, Late Proximal Tubule, & Descending Loop of Henle	8.5	4.2	7	5.6	4.2	2
Principal	0	0	0	0	0	1
Proximal Tubule	2.9		2.8	2.9	2.7	1
						-0

(d)

	XGB	RF	MLP	KNN	SVC	
Ascending Loop of Henle	1.8	4.3	2.5	2.4	2.6	7
Distal Convoluted & Connecting Tubules		2.2	1.1		1.1	5
Intercalated	4.7	7.4	7	4.7	4.7	4
Parietal Epithelium, Late Proximal Tubule, & Descending Loop of Henle	3.2	3.1	3.3	1.9	2.3	3
Podocytes			0	0	1.4	2
Principal	3.5	3.5	0.96	3.3	1.6	1
						-0

Figure S	7. Heatmap	of each c	lassifier's F1	score on Liao	et al. with	respect to	each cell type.
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	XGB	RF	MLP	KNN	SVC	
B, Plasma, & Plasmacytoid	0.78	0.78	0.79	0.7	0.73	0.9
Distal Convoluted & Connecting Tubules	0.78	0.89	0.86	0.85	0.9	0.8
Intercalated		0.9	0.98	0.98	0.98	0.7
Monocytes, Macrophages, & Other Myeloid	0.99	1	0.98	0.99	0.99	0.6
Natural Killer & T	0.96	0.95	0.96	0.97	0.96	0.5
Parietal Epithelium, Late Proximal Tubule, & Descending Loop of Henle	0.15	0.16	0.17	0.16	0.15	0.4
Principal	0.99	0.99	0.98	0.99	0.99	0.3
Proximal Tubule	0.96	0.94	0.96	0.96	0.96	0.2



Figure S8: Schematic of our study snakemake pipeline.

Figure S9: Evidence for exclusion of sample "kidney1" from Liao et al. Violin plot of the expression of a mitochondrial gene in each of the three samples from Liao et al showing a uniform abnormally high distribution of kidney1 (left). UMAP of cells in the "Proximal Tubule" harmonized cell type, colored by sample of origin showing that while most samples align with one another, cells from sample kidney1 appear distinct from the group and have poor alignment with the other cells in the same cell type (right).



Evidence for Exclusion of Sample 'kidney1'



