

Golgi-localized Ring Finger Protein 121 is necessary for MYCN-driven neuroblastoma tumorigenesis

Belamy B. Cheung^{1,2,*,#}, Ritu Mittra^{1,#}, Jayne Murray¹, Qian Wang¹, Janith A. Seneviratne¹, Mukesh Raipuria¹, Iris Poh Ling Wong¹, David Restuccia¹, Andrew Gifford¹, Alice Salib¹, Selina Sutton¹, Libby Huang¹, Parisa Vahidi Ferdowsi¹, Joanna Tsang¹, Eric Sekyere¹, Chelsea Mayoh^{1,2}, Lin Luo³, Darren L. Brown³, Jennifer L. Stow³, Shizhen Zhu⁴, Richard J. Young⁵, Benjamin J Solomon⁵, Stephane Chappaz⁶, Benjamin Kile⁷, Andrew Kueh^{8,9}, Marco J Herold^{8,9}, Douglas J Hilton^{8,9}, Tao Liu^{1,10}, Murray D. Norris^{1,10}, Michelle Haber^{1,2}, Daniel R. Carter^{1,11}, Michael W. Parker^{12,13}, Glenn M. Marshall^{1, 14,*}

Joint first authors

*Corresponding Authors: Belamy B. Cheung, Children's Cancer Institute Australia, UNSW Sydney, PO Box 81, Randwick NSW 2031, Australia. Phone: 61(02) 9385 2450, Fax: 61 (02) 9662 6584, E-mail: bcheung@ccia.unsw.edu.au
or Glenn M. Marshall, Sydney Children's Hospital, Level 1, South Wing, High Street, Randwick 2031, NSW Australia. Phone: 61 (02) 9382 1721, Fax: (02) 9382 1789, E-mail: glenn.marshall@health.nsw.gov.au

Supplementary Figure S1

Chromatograph of *Rnf121*

A to C substitution at co-ordinate
102031527 on chromosome 7 in the 4th
transmembrane domain of RNF121

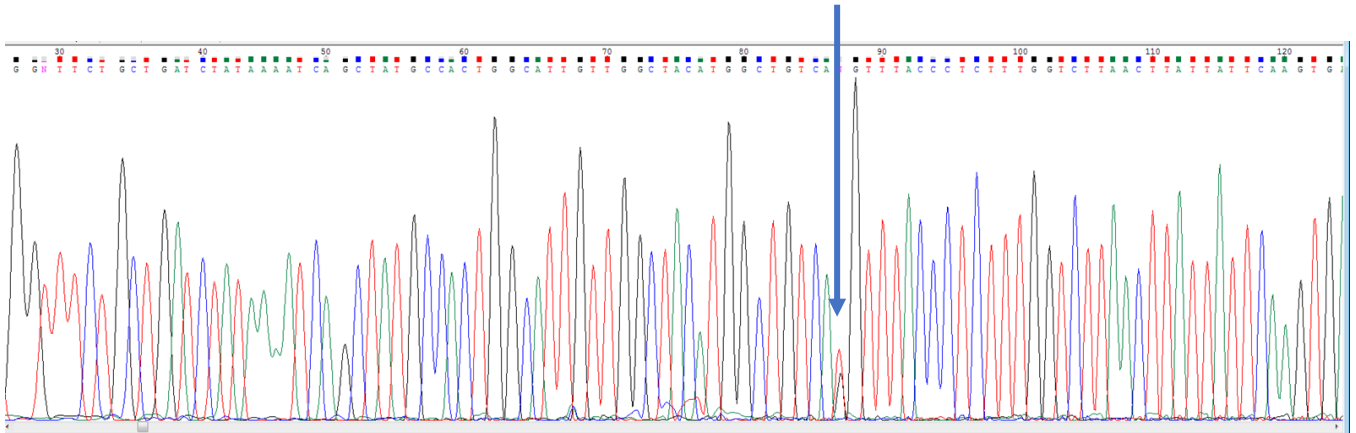
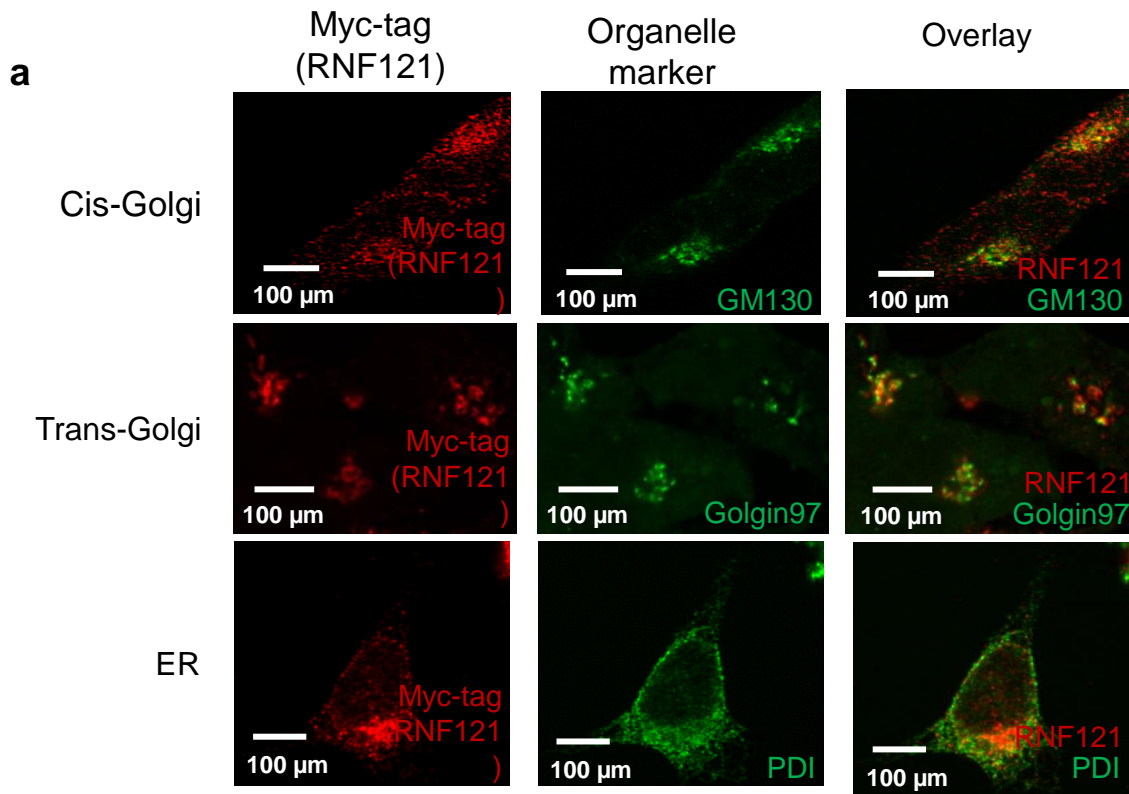


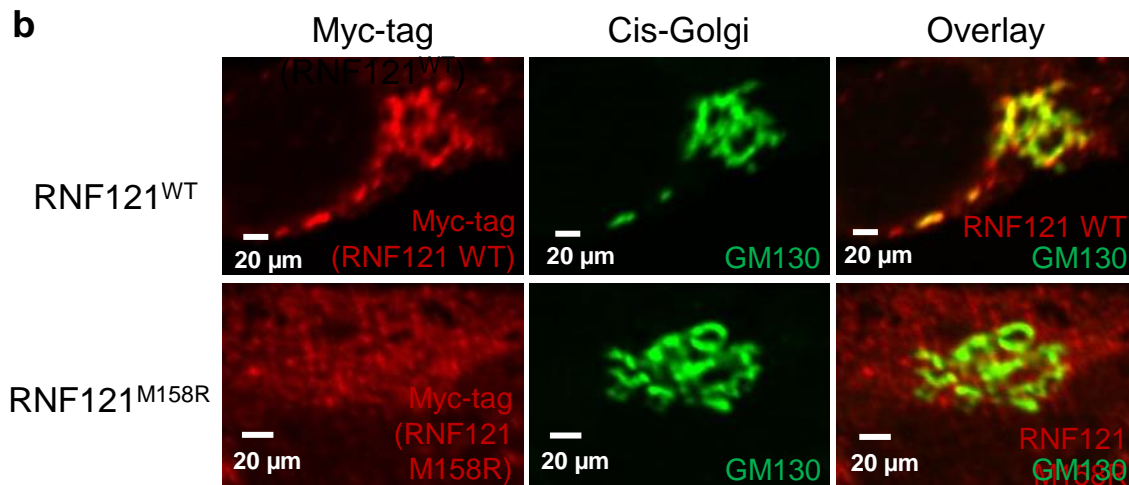
Fig. S1 A germline RNF121M158R mutation is associated with the tumor suppressed phenotype in Th-MYCN mice. A representative chromatograph of mouse chromosome 7 DNA encoding the germline *Rnf121* gene sequence from a 1929-derived and tumor-suppressed mouse showing heterozygosity of A and C at position 102031527 which is predicted to result in a methionine to arginine amino acid substitution at position 158 in the RNF121 protein.

Supplementary Figure S2

SK-N-BE(2)-C



SK-N-BE(2)-C



c

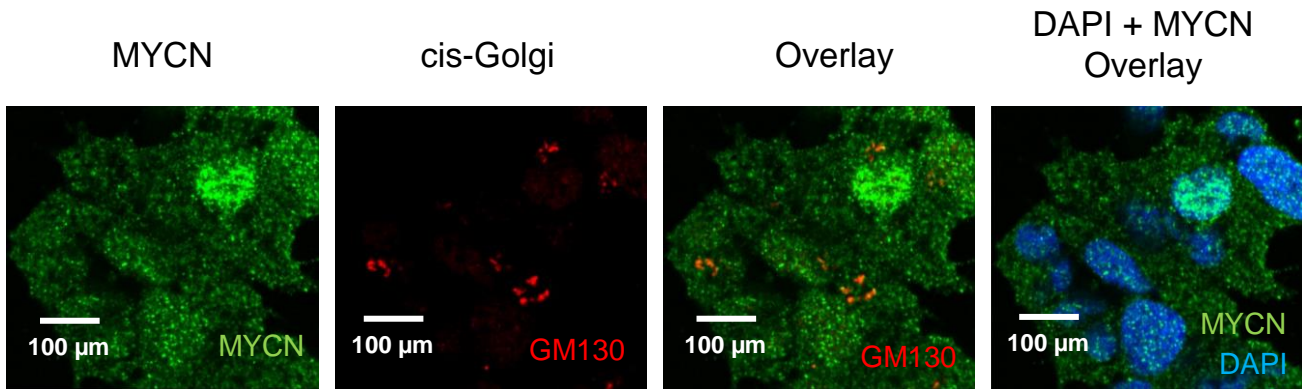
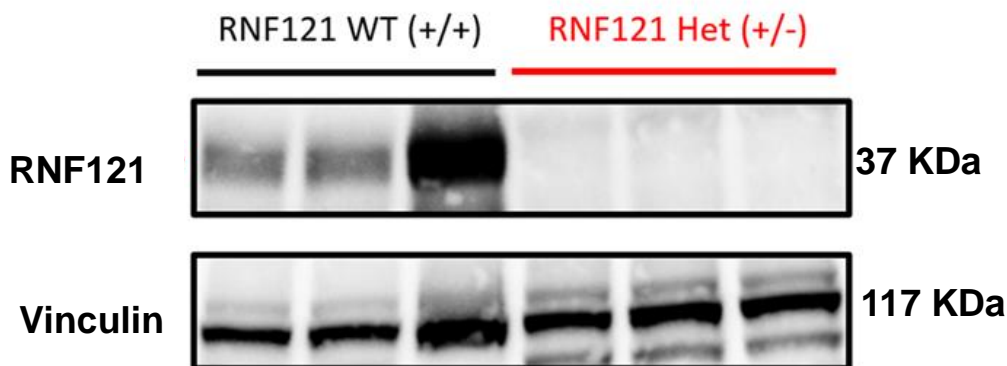


Fig. S2 *RNF121*^{M158R} mutation disrupts localisation to the *cis*-Golgi. **a** Immunofluorescent staining using: an anti-Myc-tag antibody for the transfected Myc-tagged *RNF121*^{WT} protein, an anti-GM130 antibody as a *cis*-Golgi marker, an anti-Golgin-97 antibody as a *trans*-Golgi marker, and an anti-PDI antibody as an endoplasmic reticulum (ER) marker in SK-N-BE(2)-C neuroblastoma cells. Scale bar: 100 μ m. **b** SK-N-BE(2)-C cells were transiently transfected with Myc-tagged *RNF121*^{WT} or mutant *RNF121*^{M158R} for 48 hours. Representative images of immunofluorescent staining with an anti-Myc-tag antibody for *RNF121* protein and a GM130 antibody for *cis*-Golgi are displayed. Scale bar: 20 μ m. **c** immunofluorescent staining using an anti-MYCN antibody to detect MYCN protein, an anti-GM310 antibody to detect *cis*-Golgi, and DAPI to detect nuclei. Scale bar: 100 μ m

Supplementary Figure S3

a



b

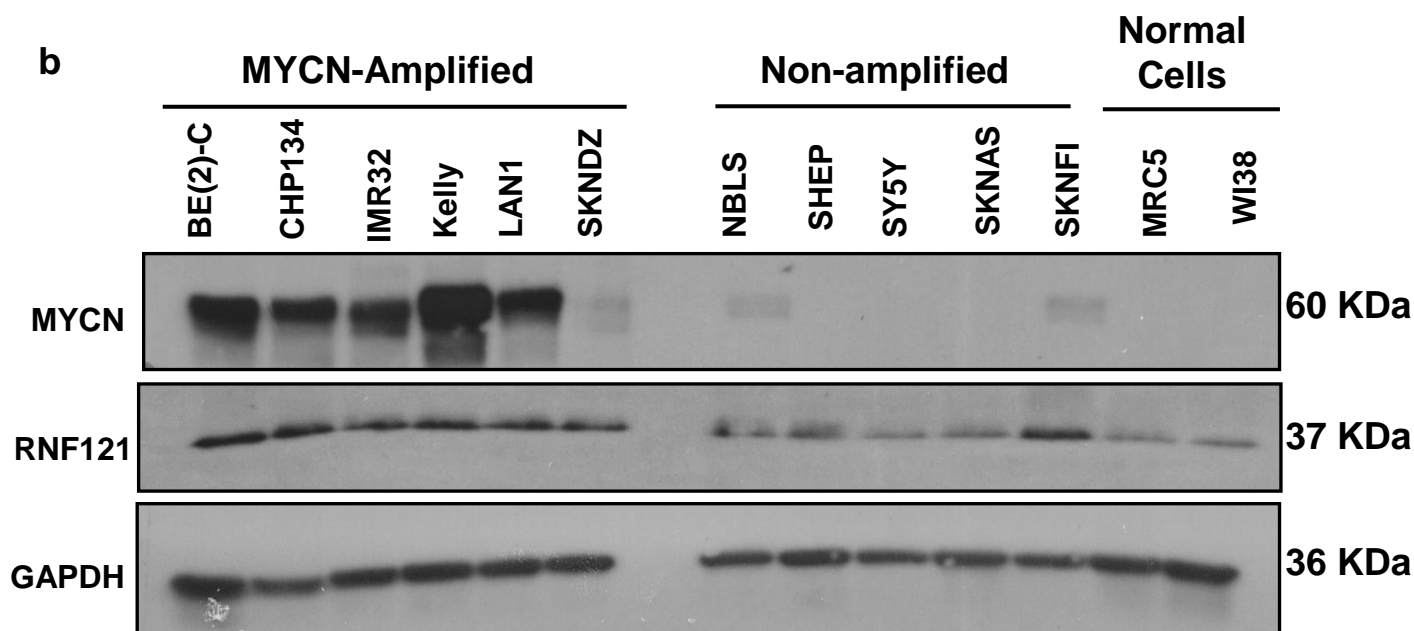
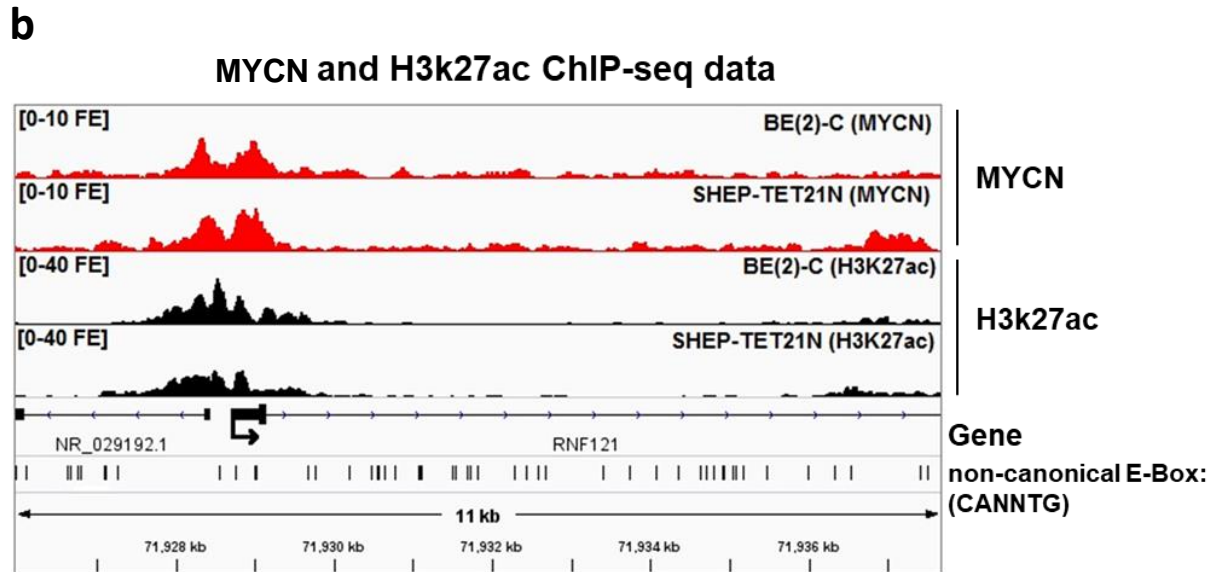
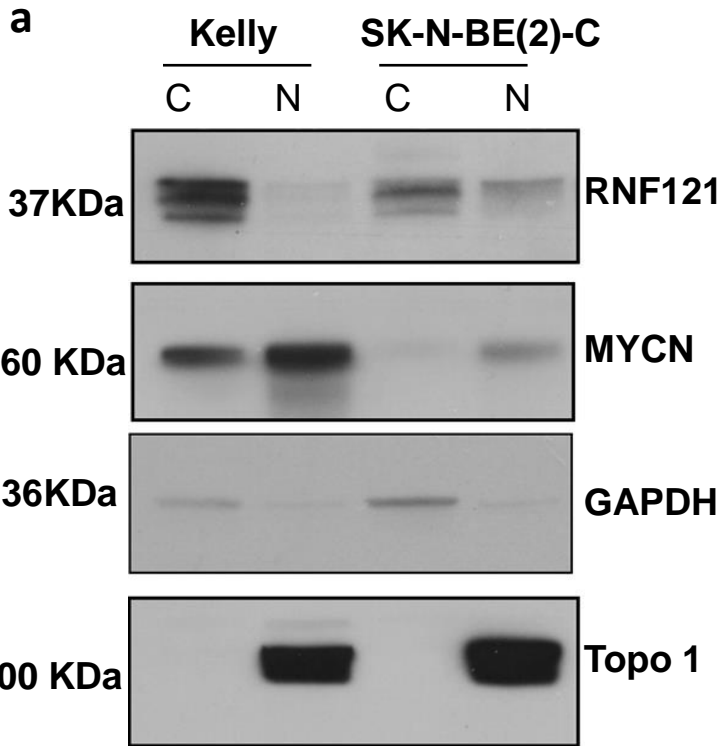


Fig. S3 The effects of RNF121 deletion, mutation or overexpression in murine and human neuroblastoma cells. **a** Immunoblot analysis of whole protein of tumour samples from Hemizygous *RNF121*^{+/-} knockout (n=3) or *RNF121*^{WT} (n=3) mice crossed with *TH-MYCN*^{+/+} mice for RNF121 expression using a murine anti-RNF121^{WT} antibody. Vinculin expression was used as a loading control. **b** Immunoblot for MYCN and RNF121 protein expression using specific antibodies in a range of neuroblastoma cell lines and normal human fibroblast cells, MRC-5 and WI-38.

Supplementary Figure S4



Supplementary Figure S4

c

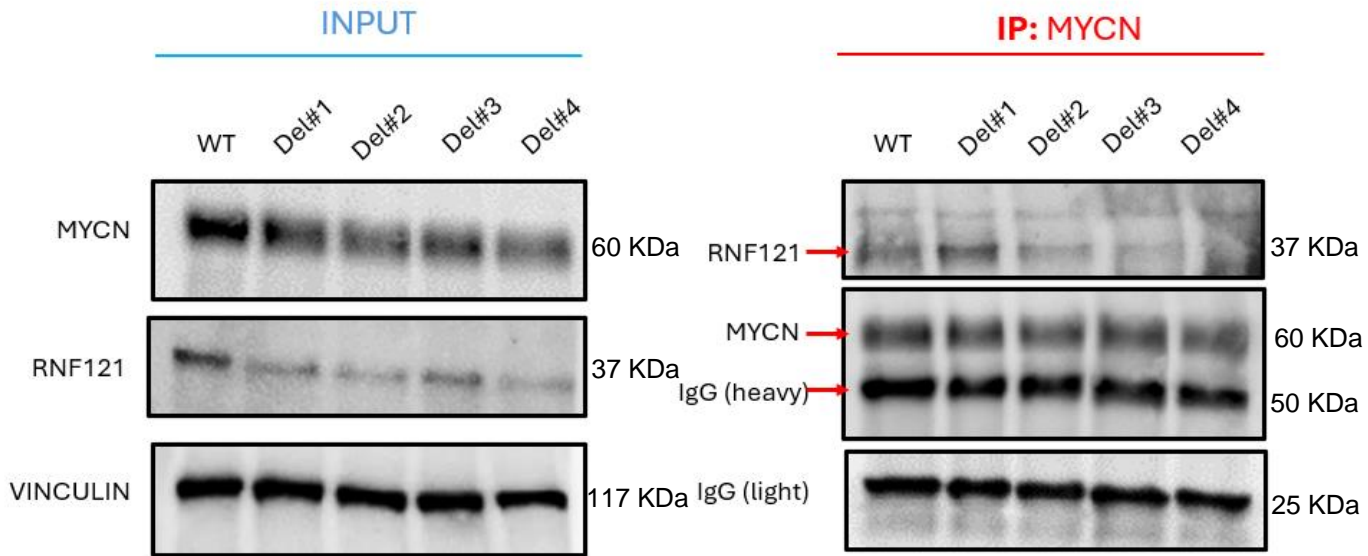
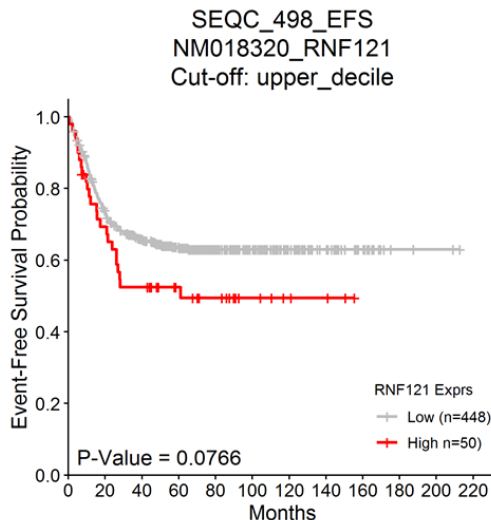
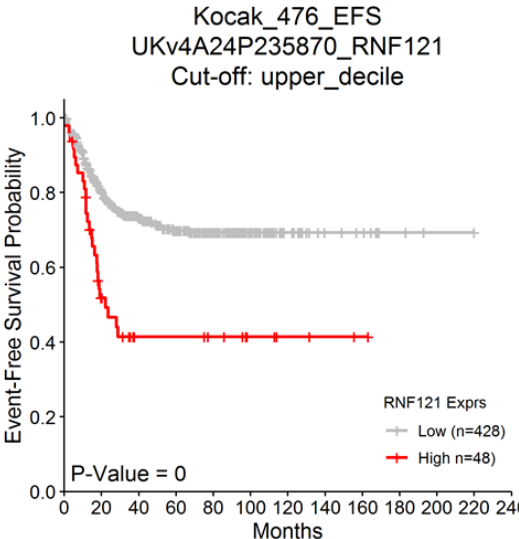


Fig. S4 RNF121 localises to the cytoplasm and is a transcriptional target for MYCN. **a** Immunoblotting showing RNF121 and MYCN expression in Kelly and SK-N-BE(2)-C cells using fractionated protein from the nuclear (N) and cytosolic (C) subcellular fractions. RNF121 and MYCN protein expression were detected using RNF121- and MYCN-specific antibodies. GAPDH- and Topoisomerase 1(Topo1)-specific antibodies were used as loading controls in the cytoplasmic and the nuclear compartments, respectively. **b** MYCN and H3k27ac ChIP-seq profiles covering the *RNF121* gene promoter region for the two neuroblastoma cell lines SK-N-BE(2)-C and SH-EP TET21N²⁴. The vertical axes on each track graph represents a scale of fold enrichment (FE) values (compared to genomic input) and the horizontal axes the gene body including the transcription start site (TSS) and proximal promoter. The TSS of RNF121 is marked by a thick arrow in the gene track, exons are indicated by thick rectangles, introns are represented by lines and the directions of transcription are marked by arrows along the gene body. Non-canonical MYCN E-Boxes (CANNTG) are marked under the gene track by vertical lines.

Supplementary Figure S5

a



b

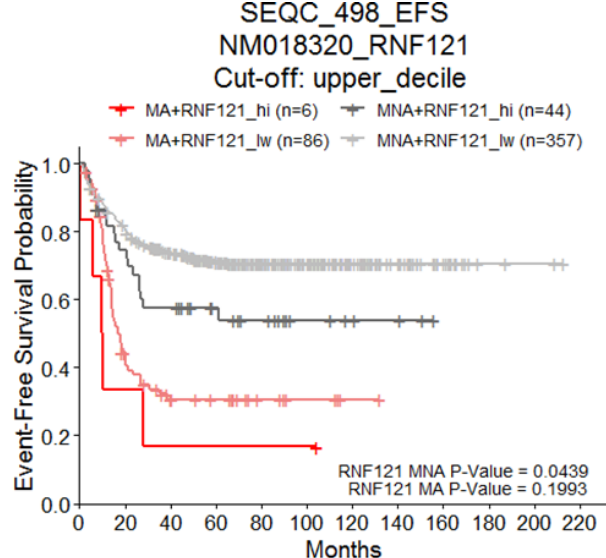
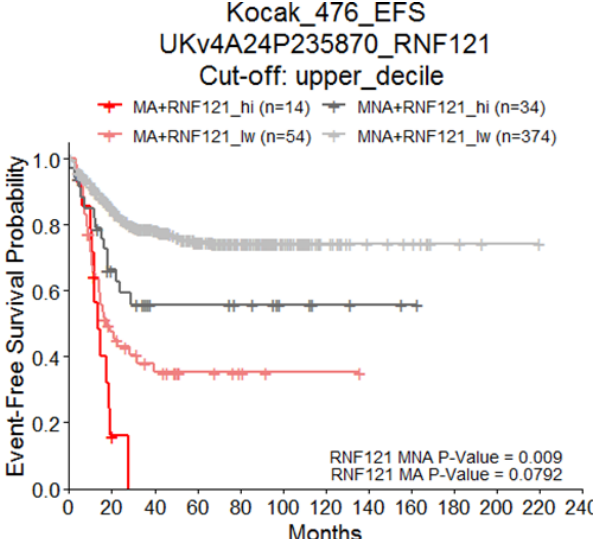
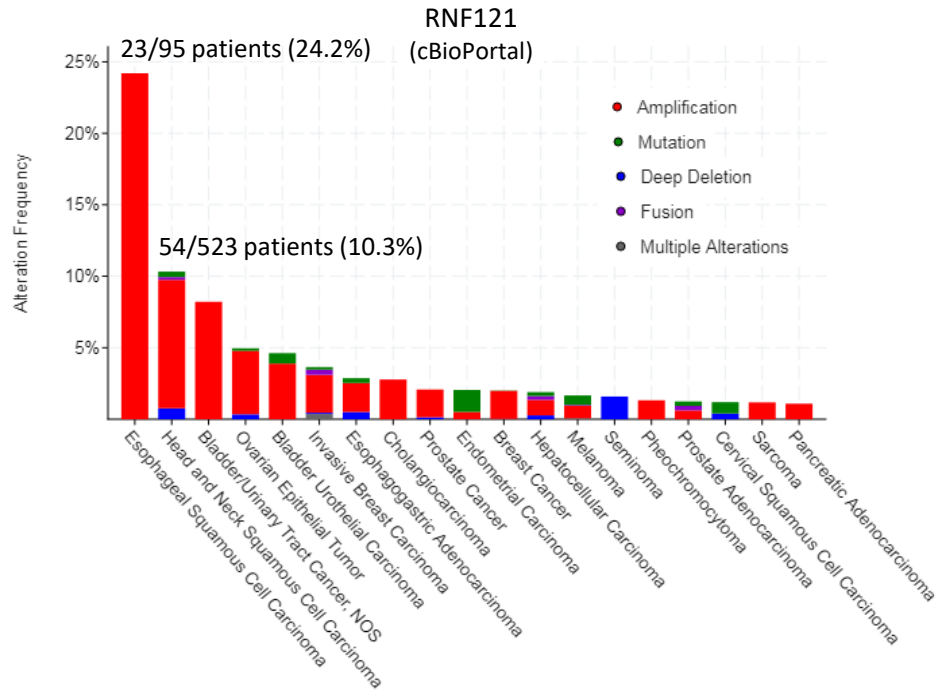


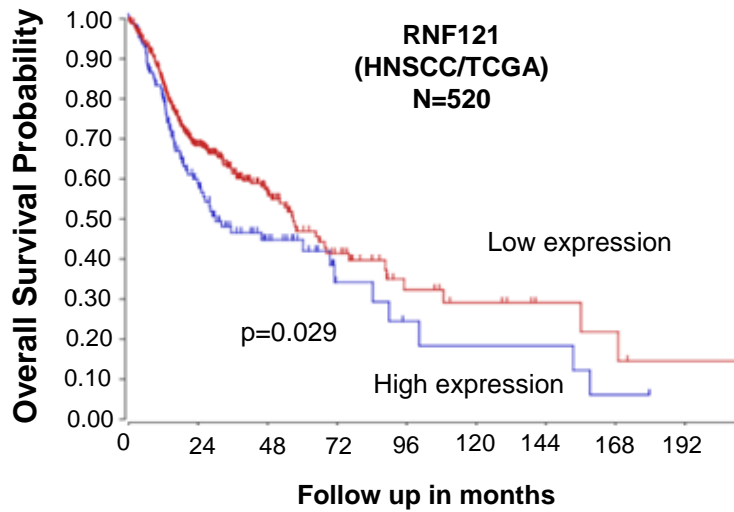
Fig. S5 High RNF121 mRNA expression associates with poor patient prognosis. a Kaplan-Meier curves of Event-free Survival probability for patients from the Kocak (n = 476) or SEQC neuroblastoma cohort (n = 498) subgrouped above or below the upper-decile of *RNF121* mRNA expression level. The p-values are from two-sided log-rank tests. **b** Kaplan-Meier curves of Event-free Survival probability for patients from the Kocak (n = 476) or SEQC neuroblastoma cohort (n = 498) subdivided by *RNF121* mRNA expression and *MYCN* amplification status (Amplified; MA, Non-amplified; MNA). Patients were subgrouped above or below the upper decile of *RNF121* expression level (High; *RNF121_hi*, Low; *RNF121_lw*). The p-values are from two-sided log-rank tests between each *RNF121* (lw, hi) expression group after subdividing cohorts by *MYCN* amplification status. P-values were adjusted using the Benjamini–Hochberg method to account for multiple comparisons. **c** Kelly cells were transiently transfected for 48 hours with *RNF121*^{WT} or the *RNF121* deletion mutants 1-4 (with mutants 3 and 4 lacking helix 5). Endogenous *MYCN* co-IP was performed, followed by immunoblotting with antibodies to recognize *MYCN* or *RNF121* and Vinculin.

Supplementary Figure S6

a



b

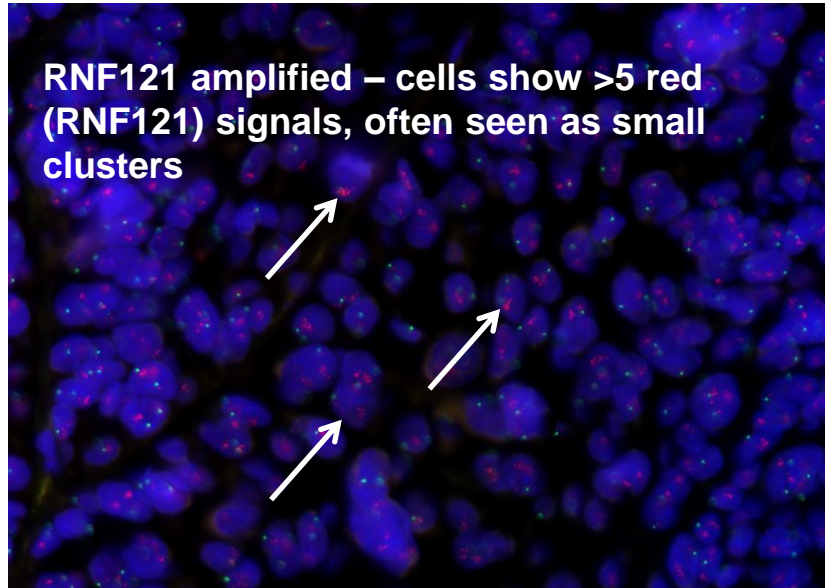


Supplementary Figure 6

c

12/66 (18.2%) cases scored as RNF121 gene amplified

RNF121 FISH results (LSCC TMA's 1A+2A, Empire Genomics probes)		
	#	%
Total # scored	66	
hemi gene loss	1	1.5
hemi gene loss	1	
gene disomy	30	45.5
disomy	25	
chr trisomy	4	
chr polysomy	1	
gain	23	34.8
gain	16	
gain+chr trisomy	6	
gain+chr polysomy	1	
amplification	12	18.2
amplification	7	
amplification+chr trisomy	2	
amplification+chr polysomy	3	



d

19/46 (41.3%) cases scored as Myc gene amplified

Myc FISH results (LSCC TMA's 1A+2A, Vysis probes)		
	#	%
Total # scored	46	
gene disomy	12	26.1
disomy	10	
chr trisomy	1	
chr polysomy	1	
gain	15	32.6
gain	5	
gain+chr trisomy	6	
gain+chr polysomy	4	
amplification	19	41.3
amplification	11	
amplification+chr trisomy	4	
amplification+chr polysomy	4	

Fig. S6 *RNF121*^{WT} amplification is frequent in head and neck squamous cell cancer (HNSCC). **a** The frequency of *RNF121*^{WT} copy number alterations (Amplification, Mutation, Deep Deletion, Fusion and Multiple Alterations) by cancer type from the cBio Cancer Genomics Portal (cBio portal, <http://cbioportal.org>). **b** Kaplan-Meier curves of Overall Survival probability of patients from the TCGA HNSCC cohort (n = 520) subdivided by *RNF121* gene expression, where patients were grouped based on the upper-decile of gene expression values. The p-values are from two-sided log-rank tests between each group. **c** Left panel: Fluorescence *in situ* hybridisation (FISH) scoring results for tumour cores from 66 HNSCC patients analysed for *RNF121* gene amplification. Right panel: Representative images of fluorescence in situ hybridisation of *RNF121* amplification. Arrows point to cells with increased number of *RNF121* copies (red signals). **d** FISH scoring results for tumour cores from 46 HNSCC patients analysed for c-*Myc* gene amplification.

Supplementary Figure 7 (unprocessed immunoblots)

Figure 2c

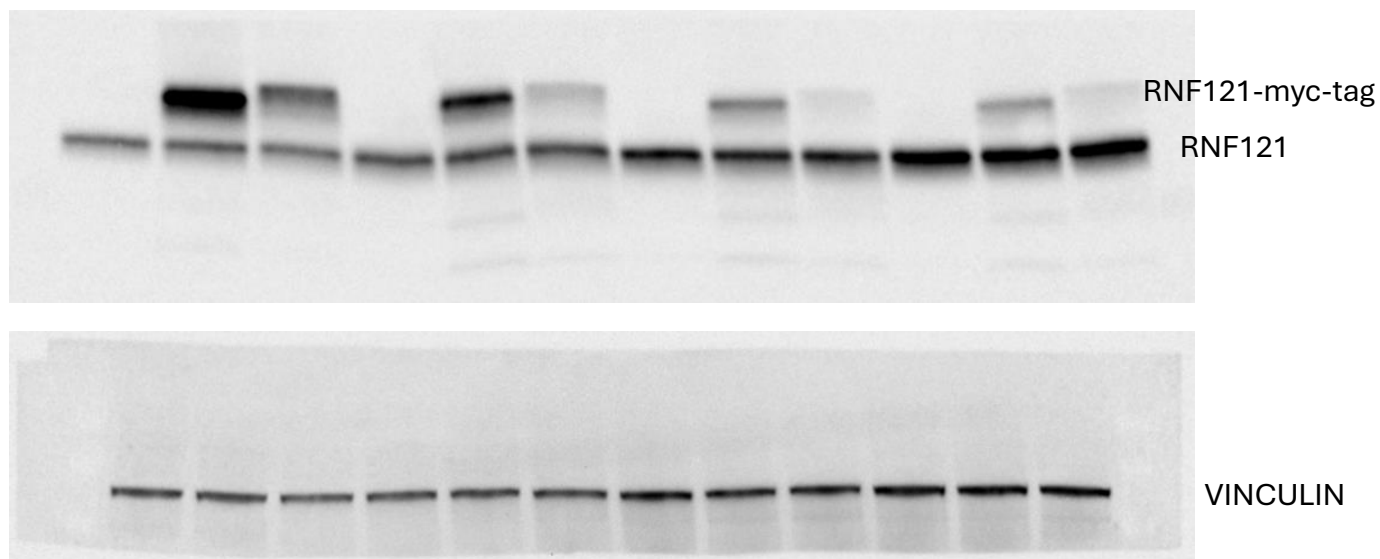


Figure 2d

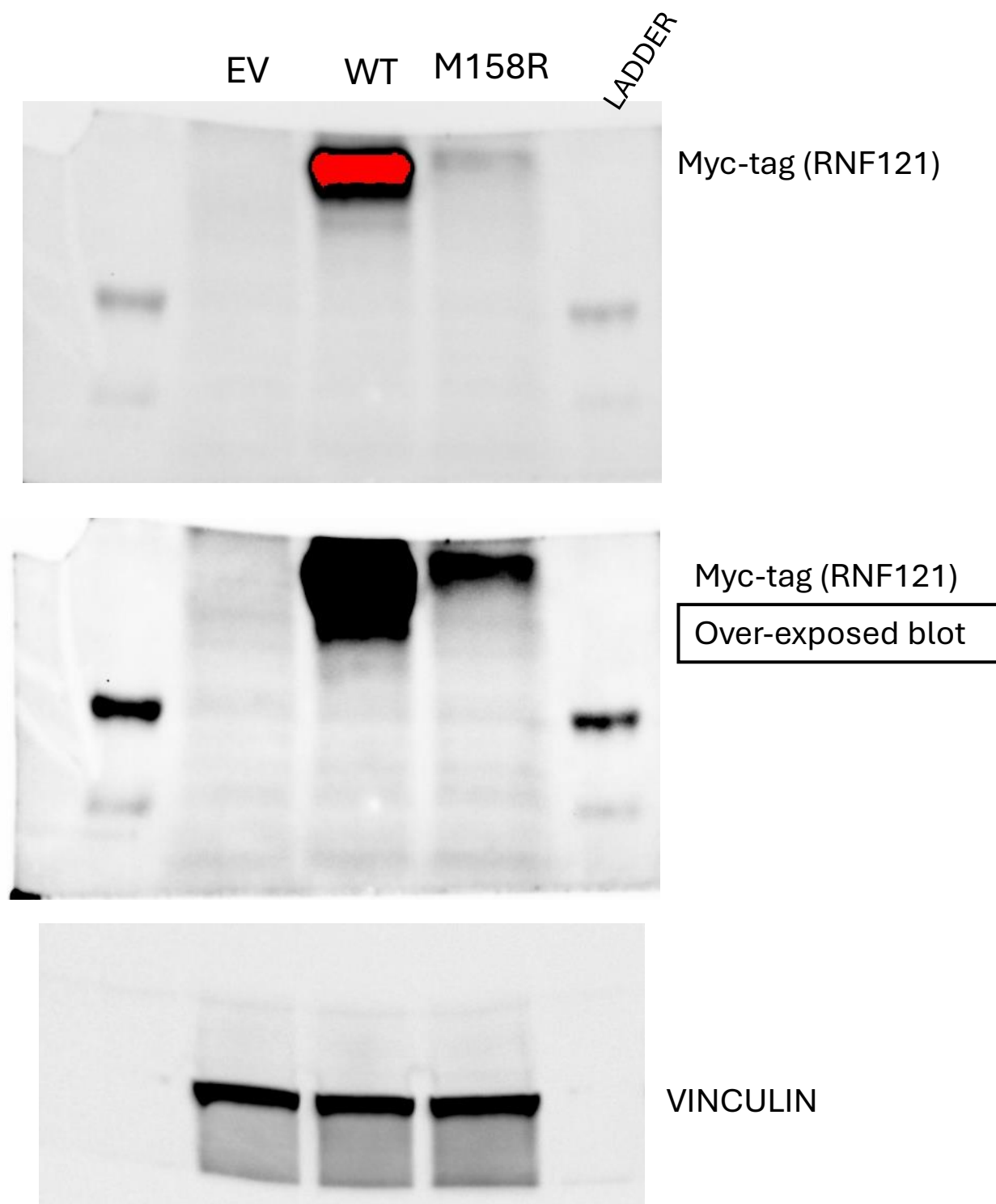


Figure 4c

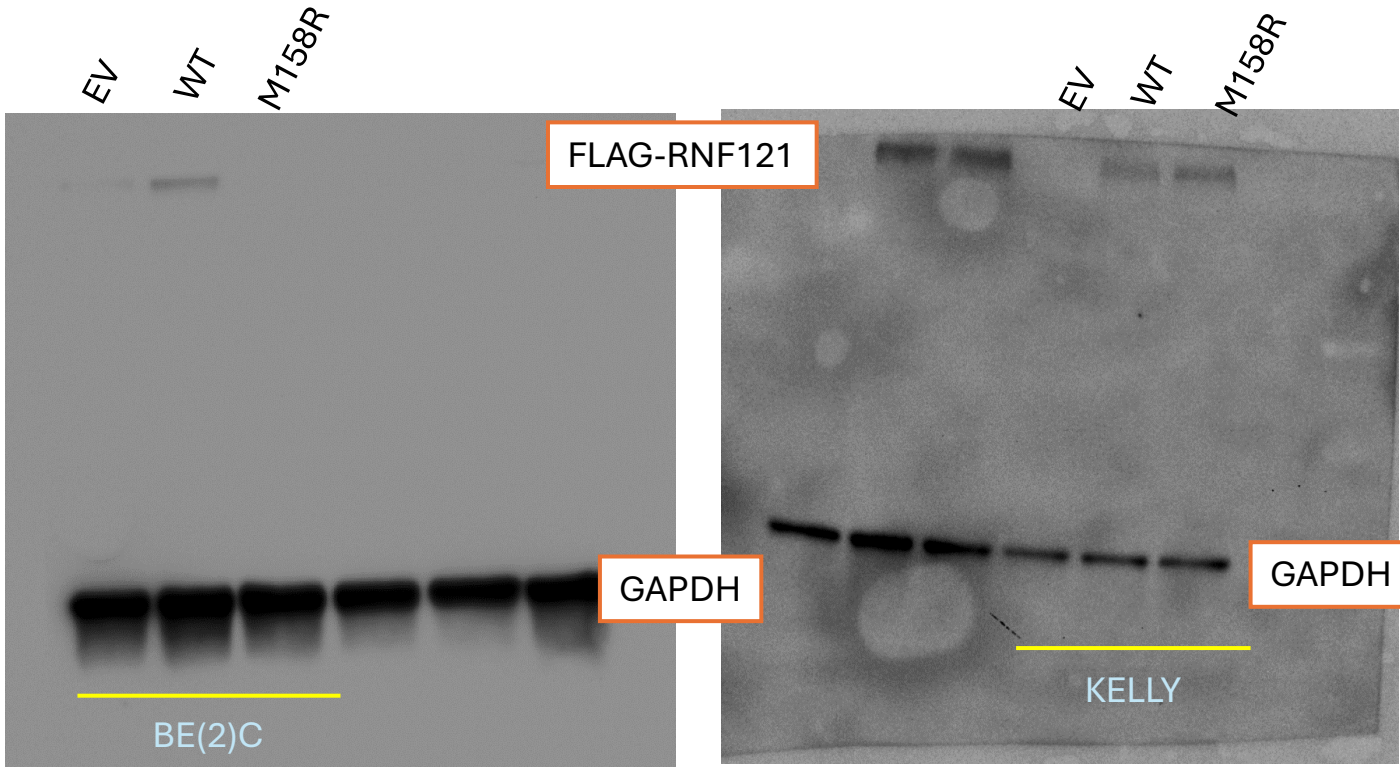


Figure 4d

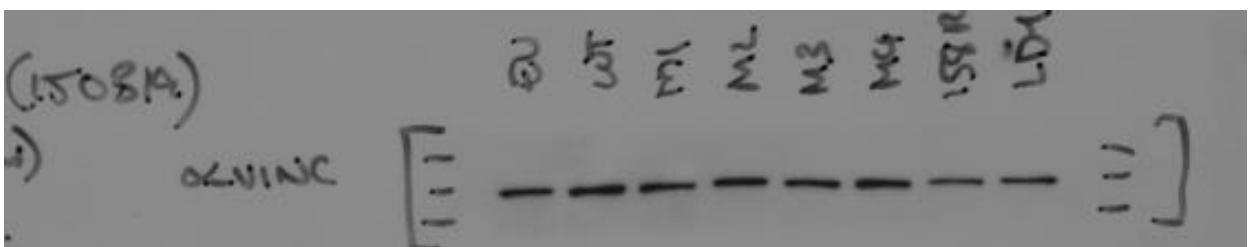
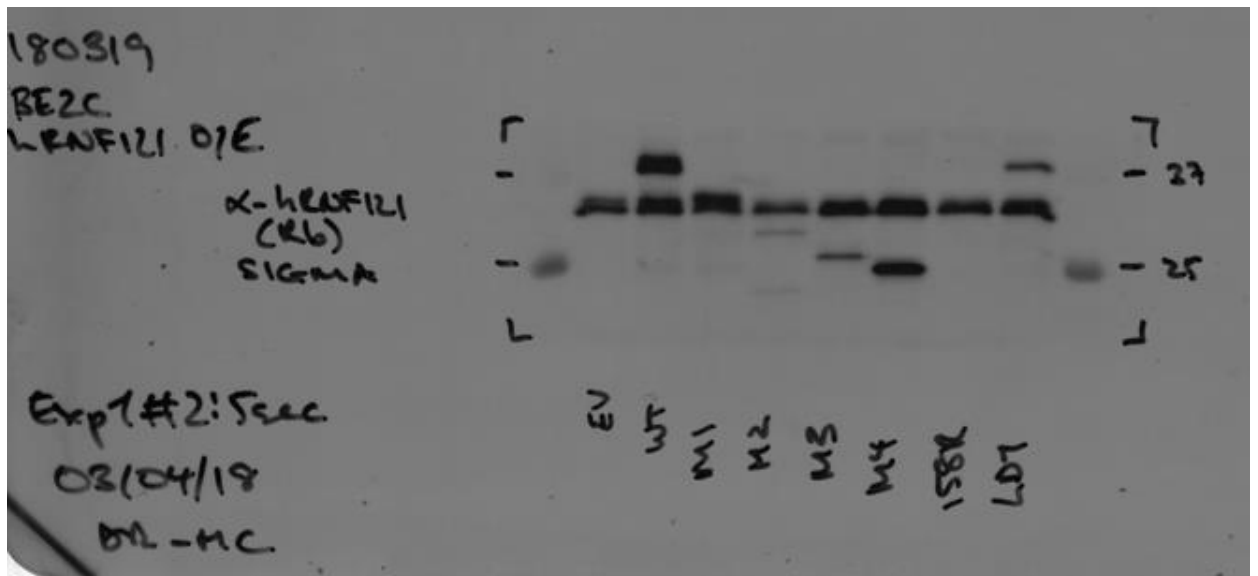
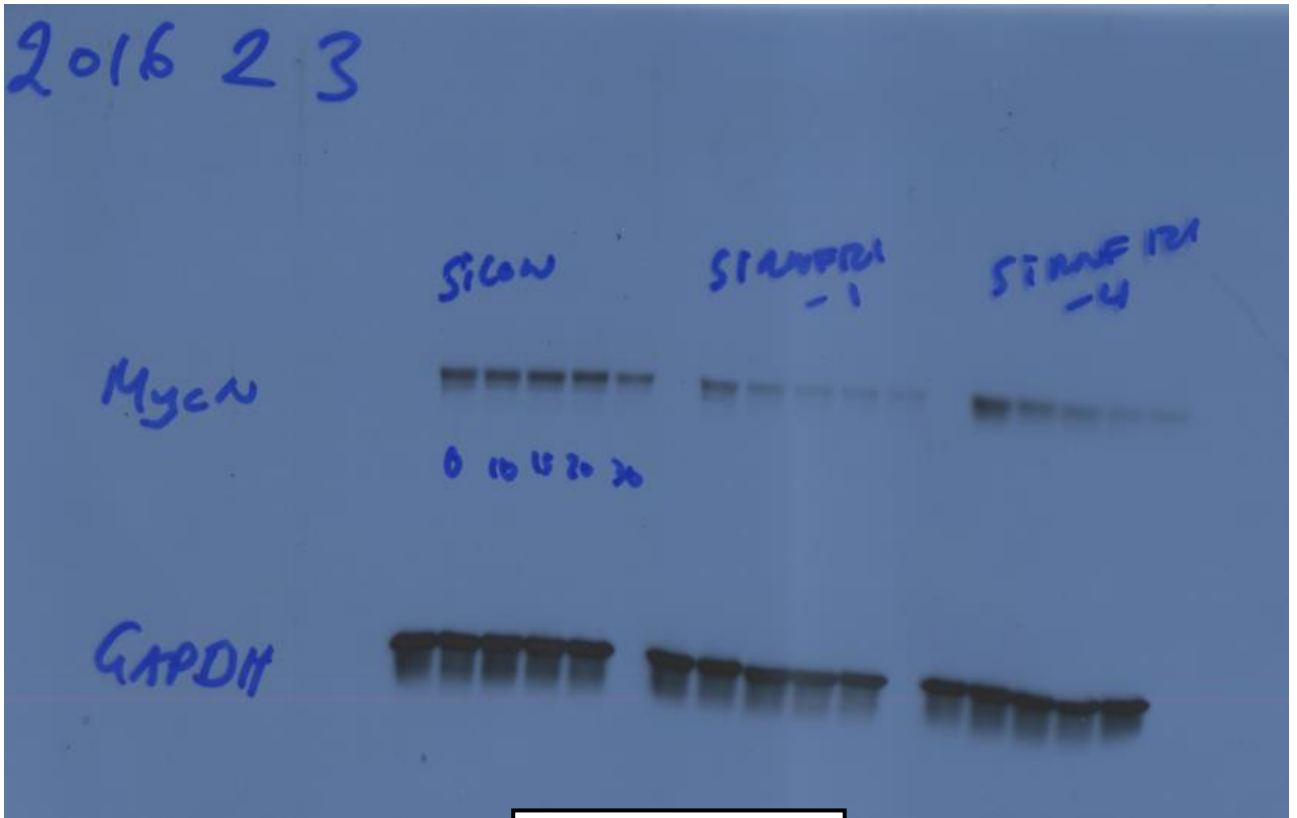
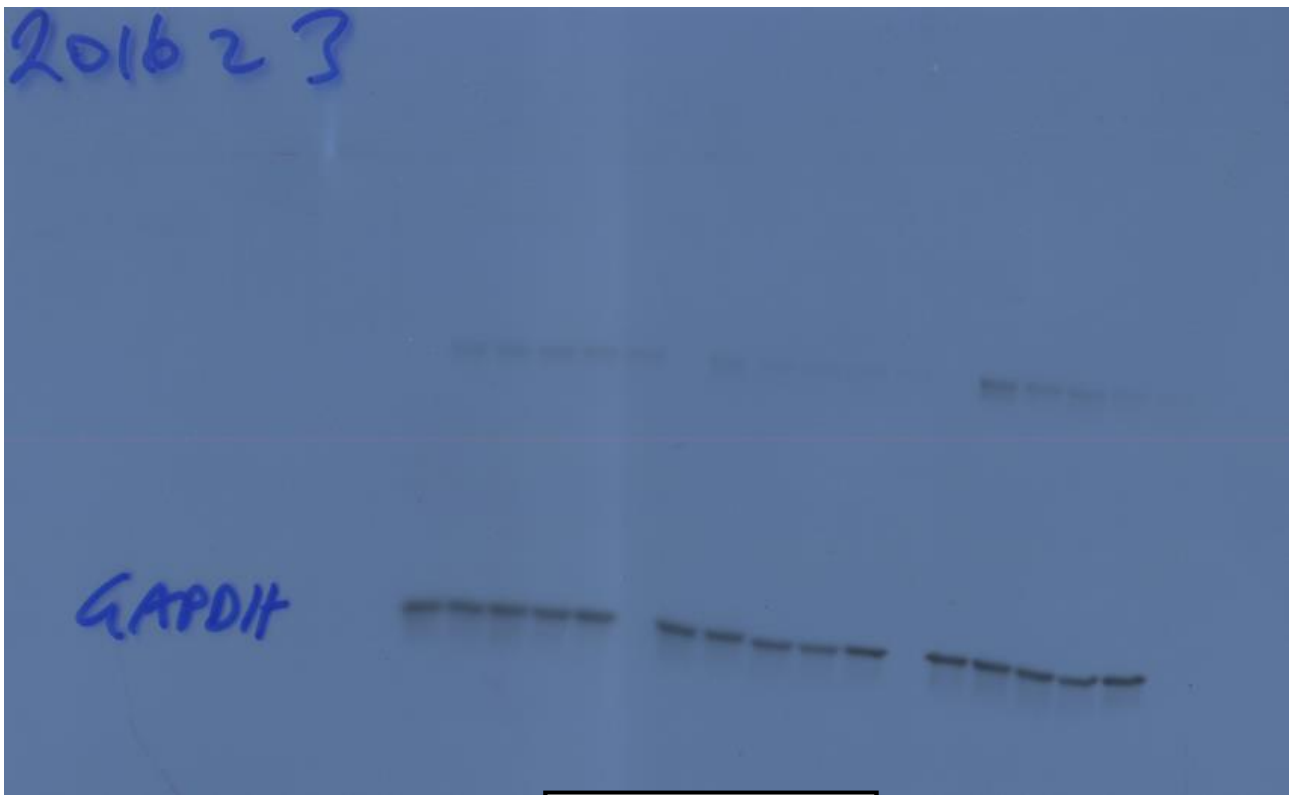


Figure 5a



High-exposure



Low-exposure

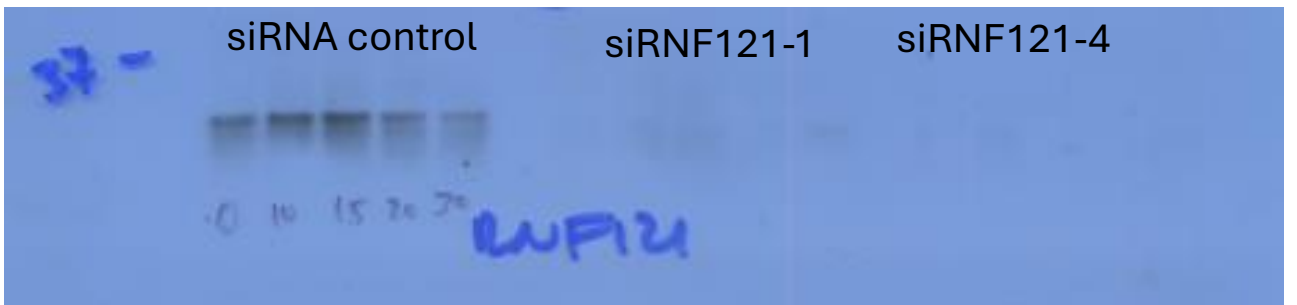


Figure 5b

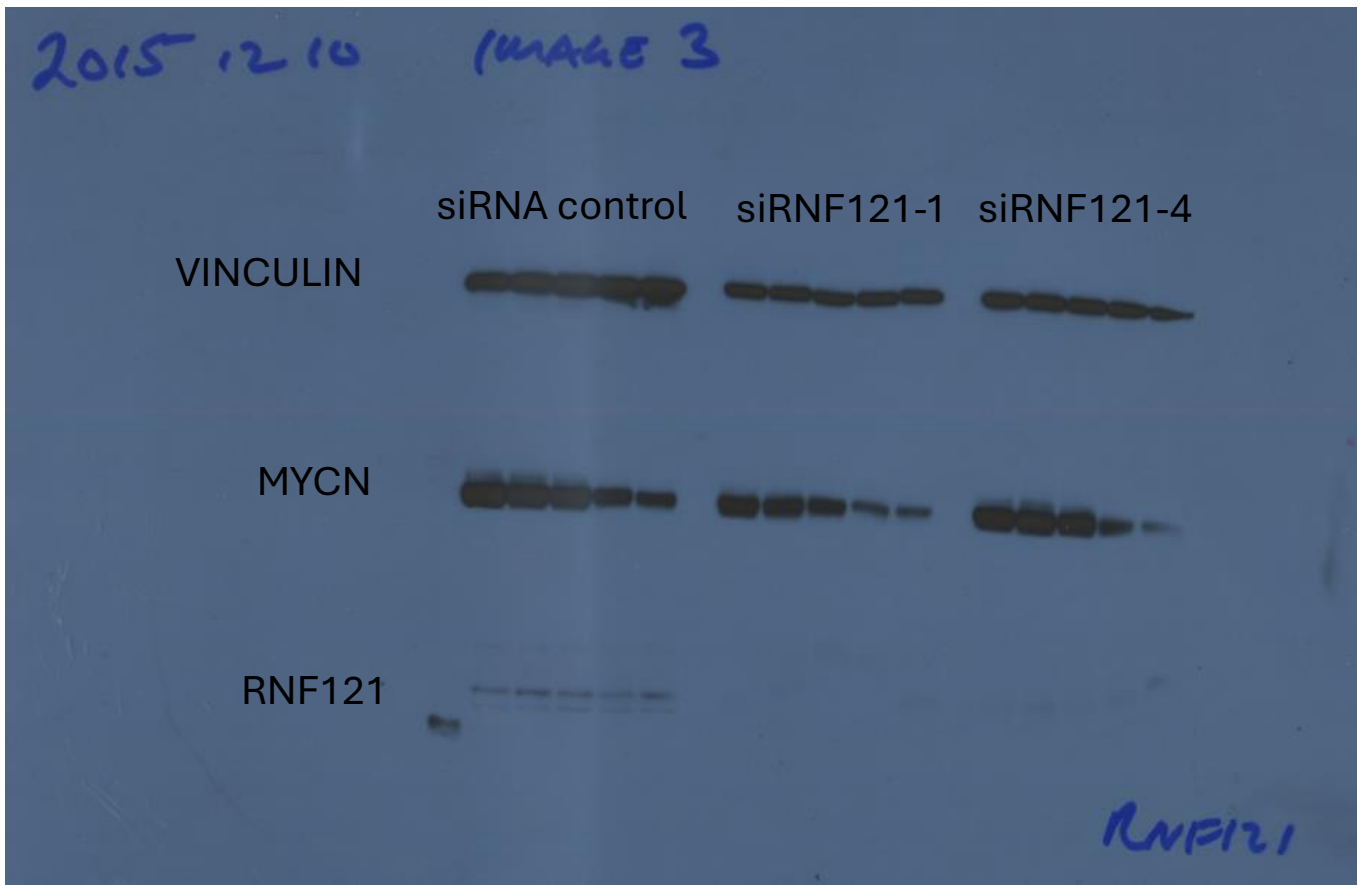
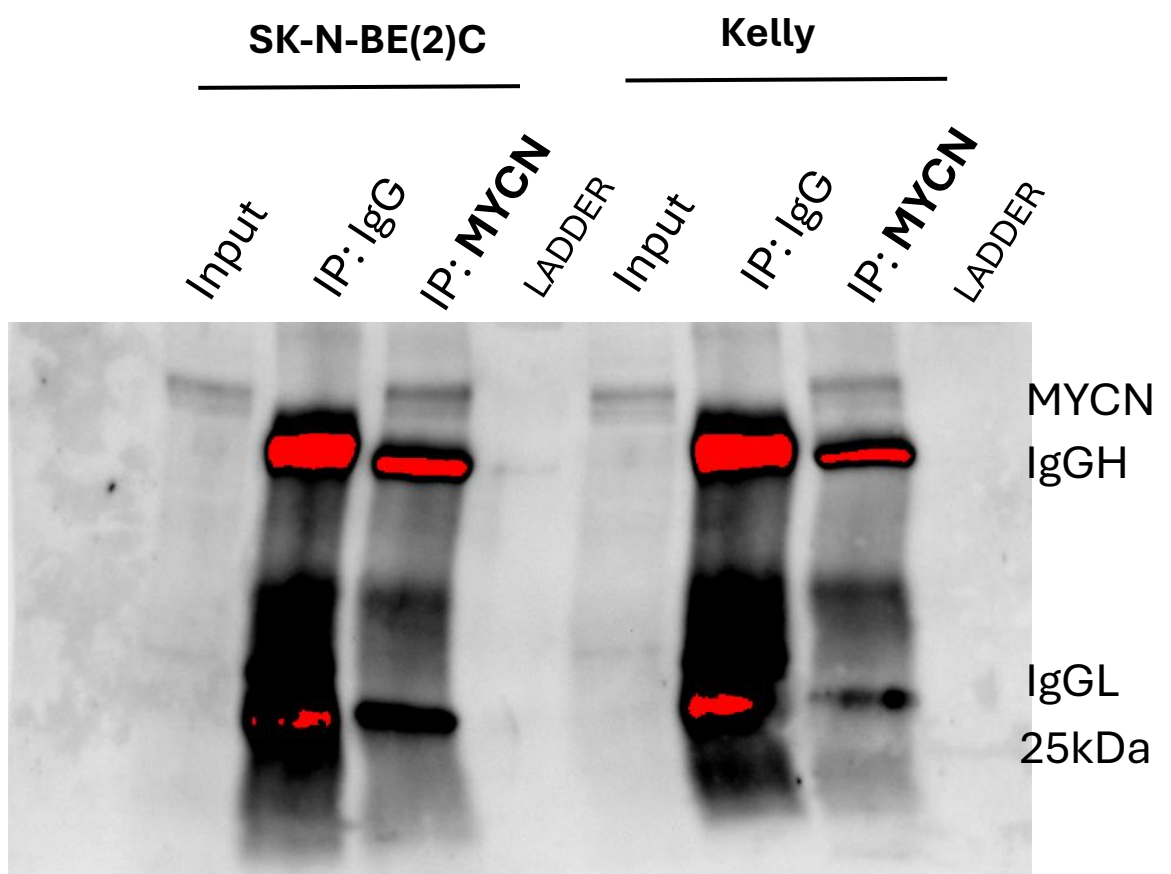
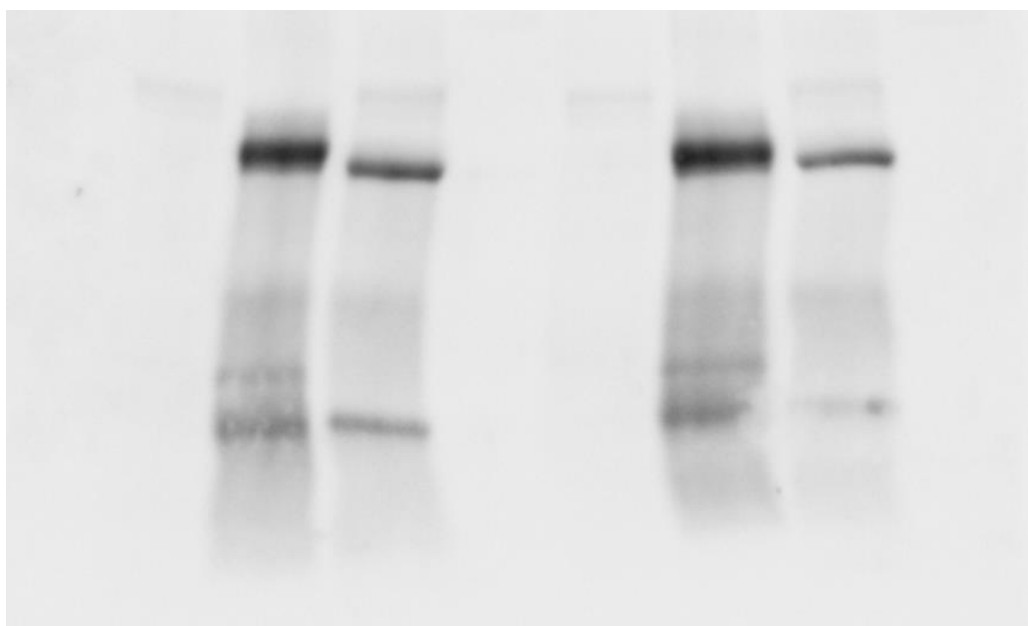


Figure 5g



Over-exposed blot showing **MYCN pulldown** (red was corrected in processed image)

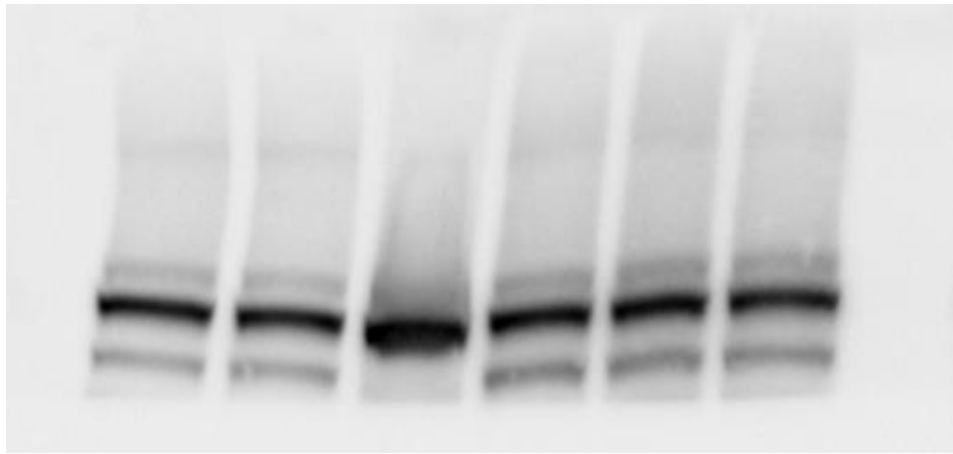


Low-exposure whole blot showing MYCN pulldown

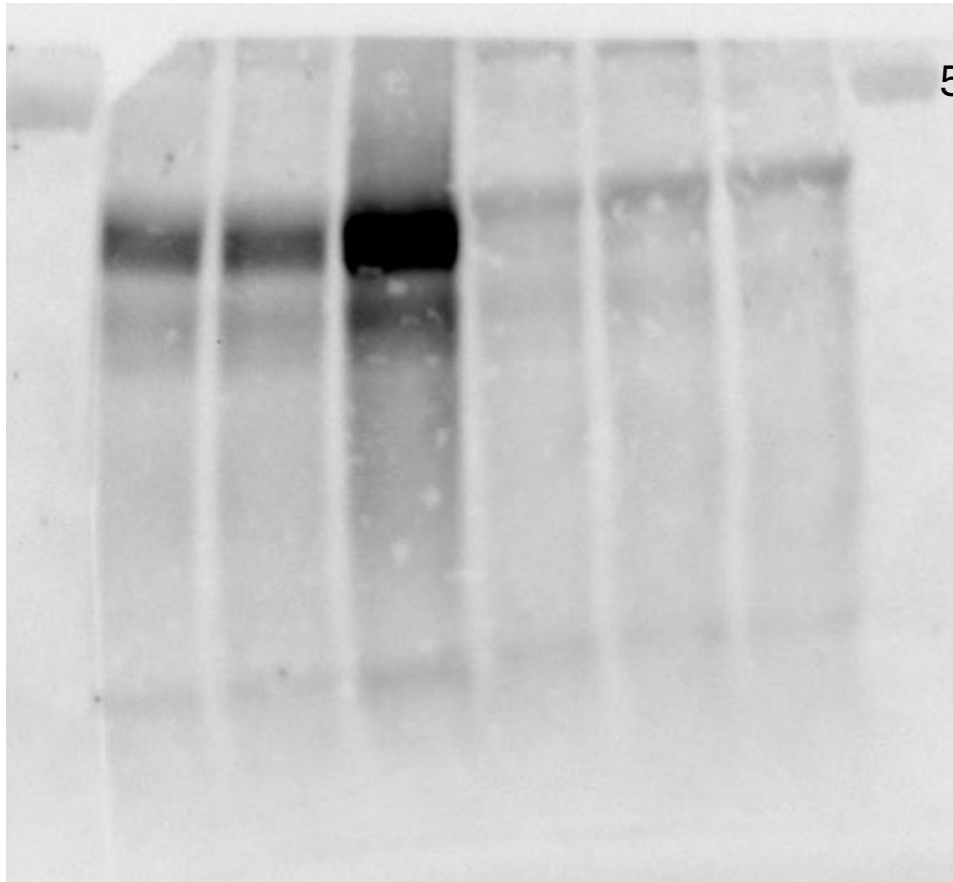


Blot was cut, stripped, and **re-probed for RNF121** to check for co-immunoprecipitation

Supplementary figure **S3a**



Vinculin



50kDa

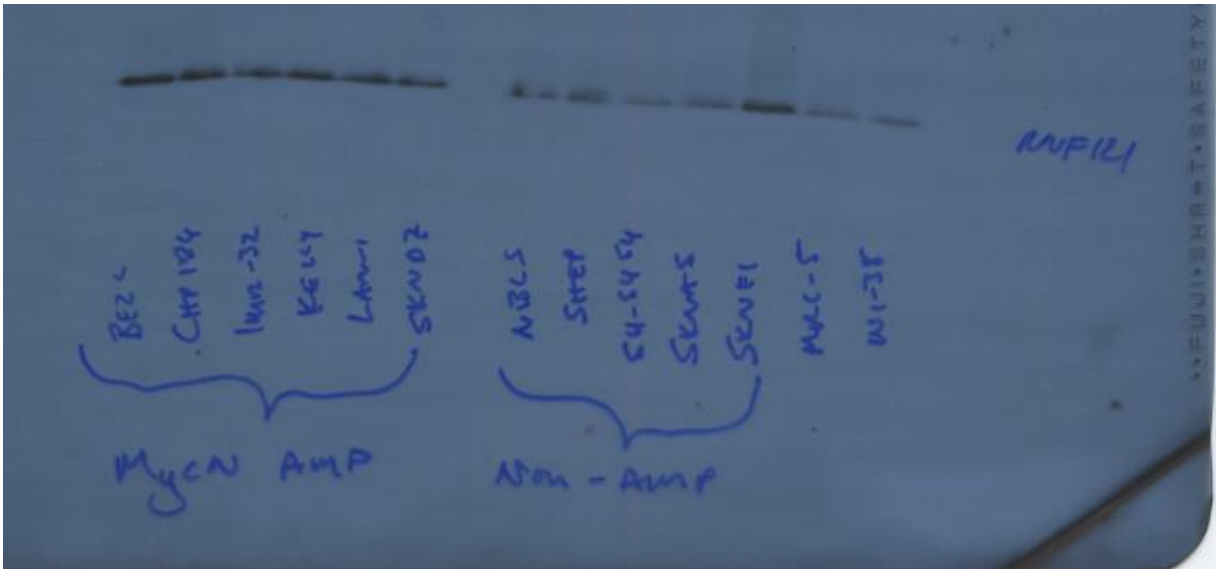
RNF121

Whole blot cut and probed for Vinculin (on top)
and RNF121 in bottom half

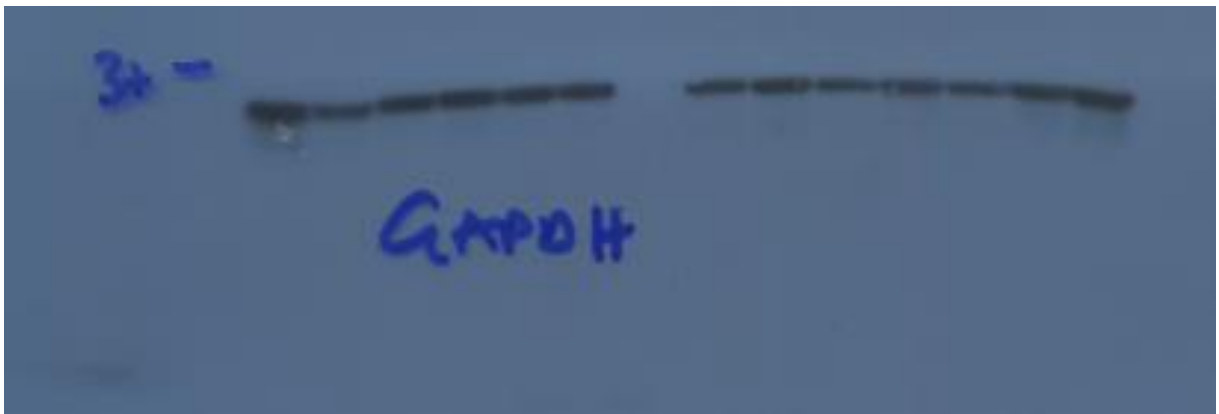
Supplementary figure **S3b**



MYCN expression across cell line panel (with GAPDH **over-exposed**)

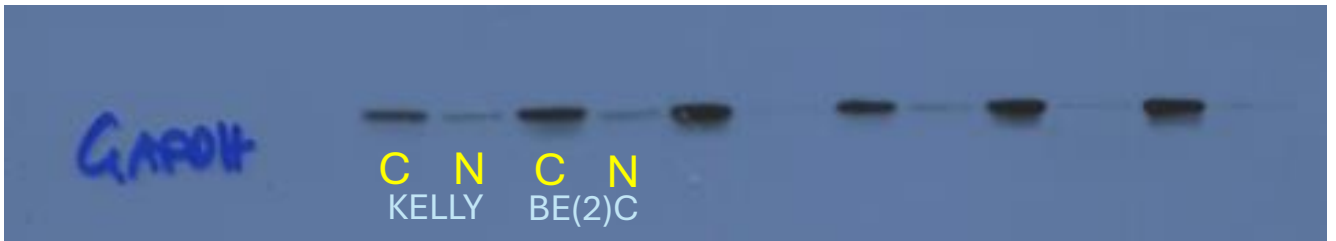
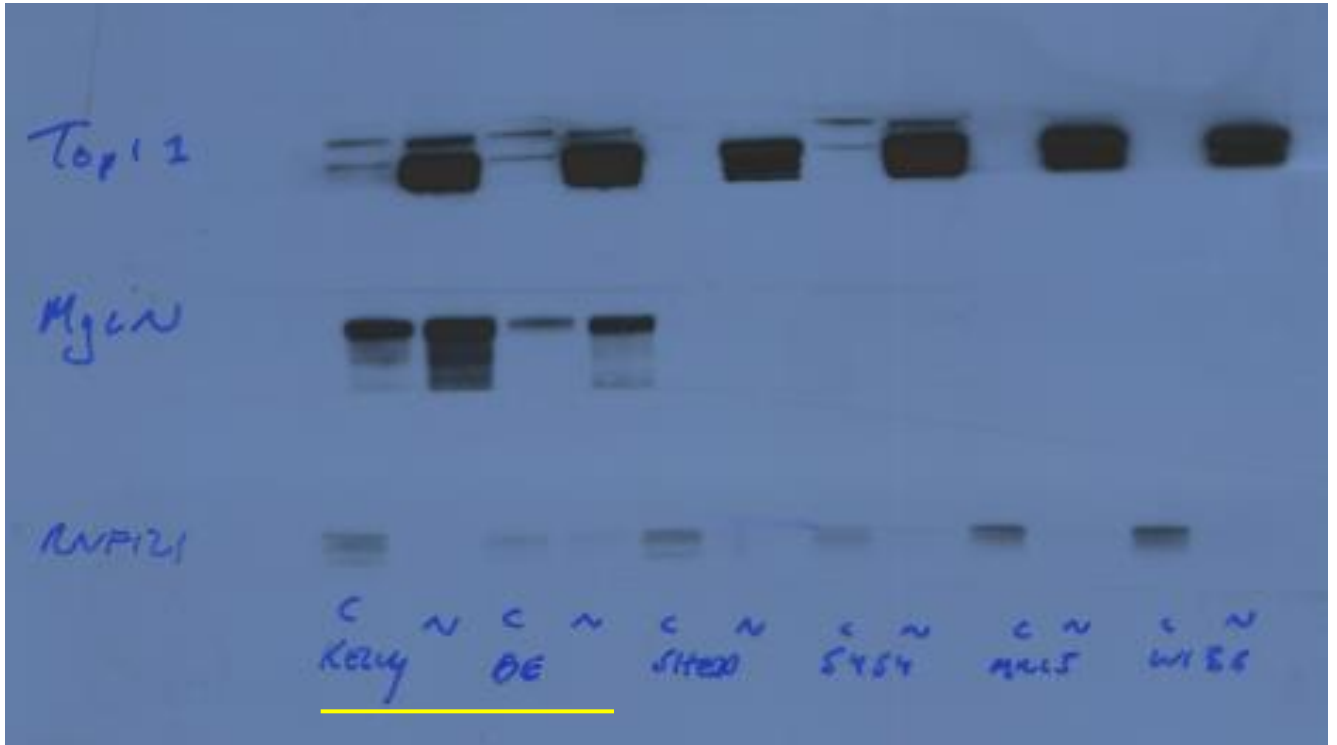


RNF121 expression across cell line panel



GAPDH low-exposure

Supplementary figure S4a

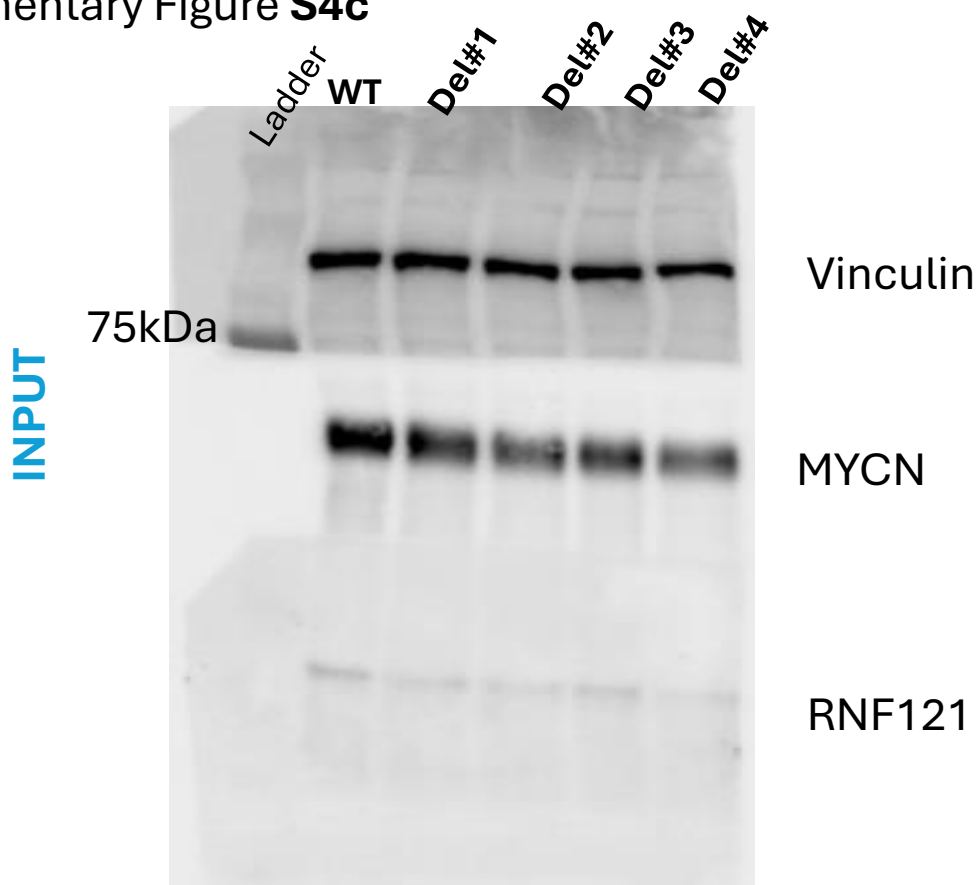


Only Kelly and BE(2)-C data was taken for processed figure

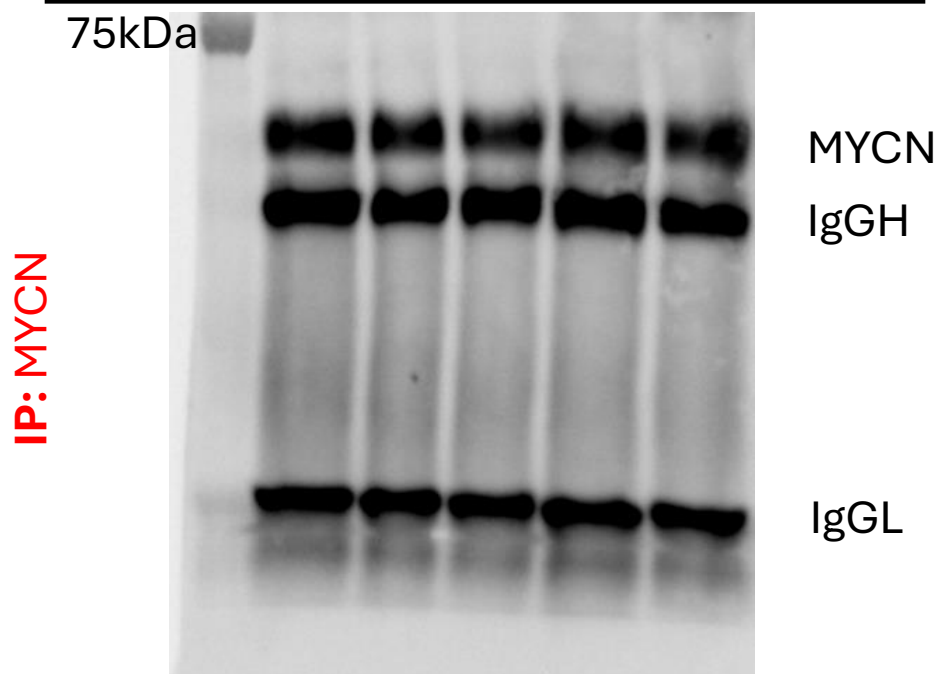


Over-exposed for RNF121 expression

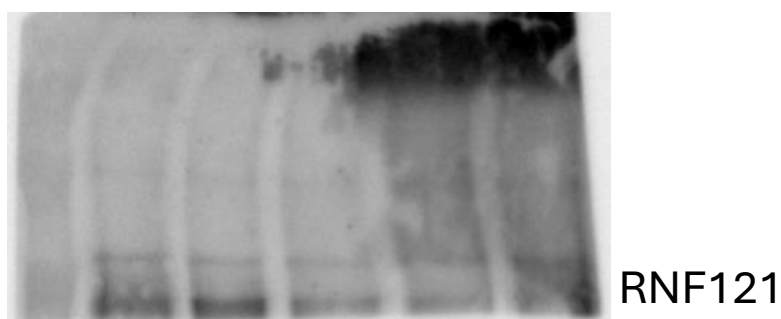
Supplementary Figure **S4c**



INPUT immunoblot **strips put together** (cut to probe for different antibodies – MYCN ladder caused over-exposure and was cut before imaging)



Immunoblot probed for **MYCN pulldown** (high-exposure)



Immunoblot cut, stripped, and probed for **RNF121 pulldown** (highest-exposure)