

## Supplementary materials

**Multi-assessed green sustainable chromatographic resolution of nicotine and caffeine; application to in-vitro release from a new quick mist mouth spray co-formula**

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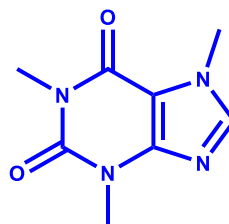
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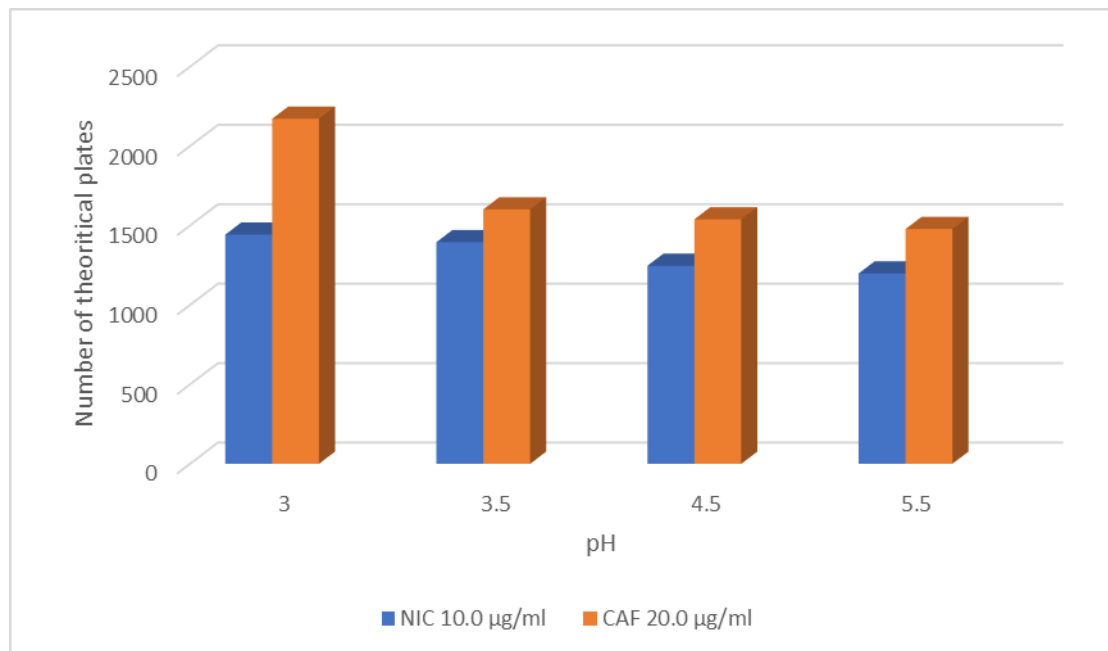


**Nicotine (NIC)**

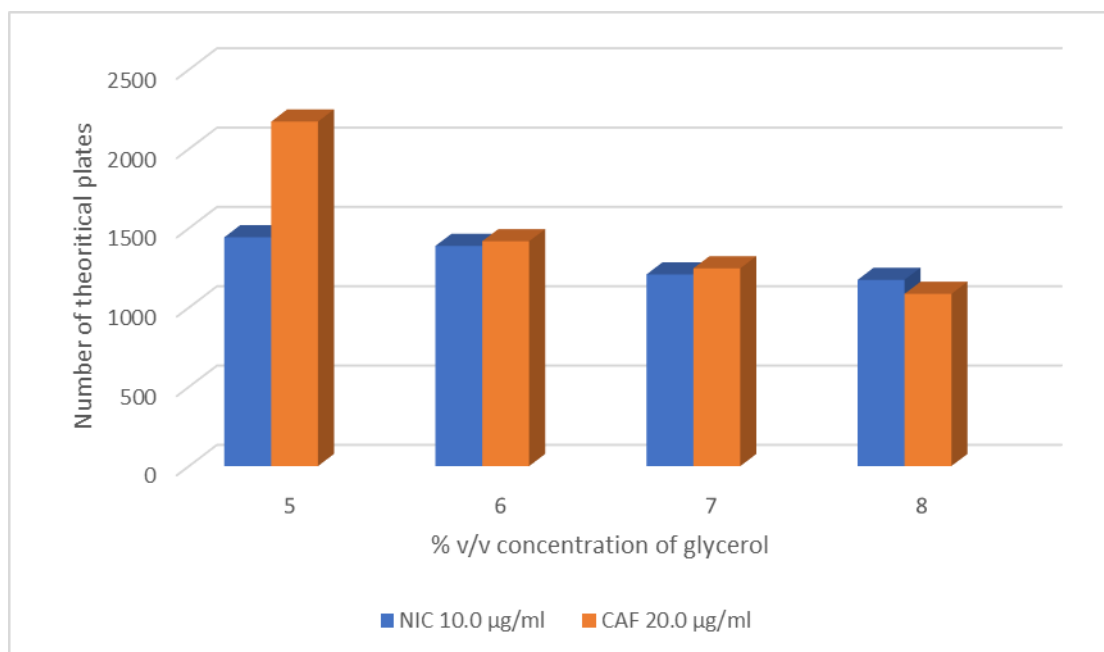


**Caffeine (CAF)**

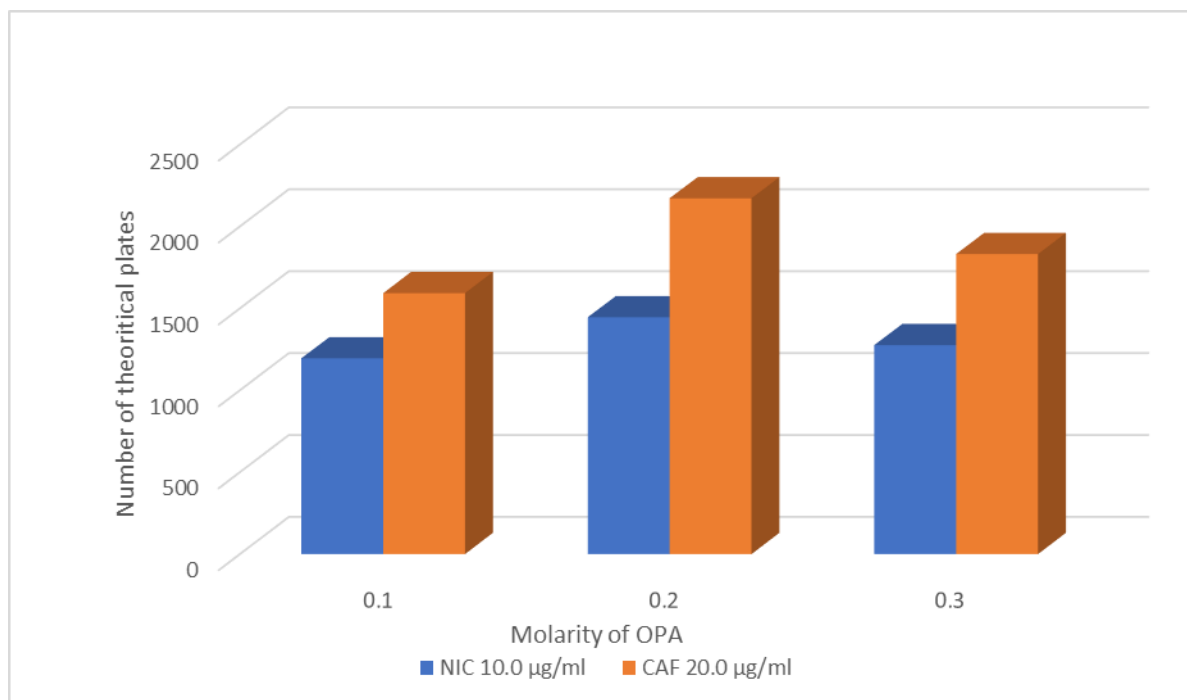
**Fig. S1.** Structural formula of nicotine (NIC) and caffeine (CAF)



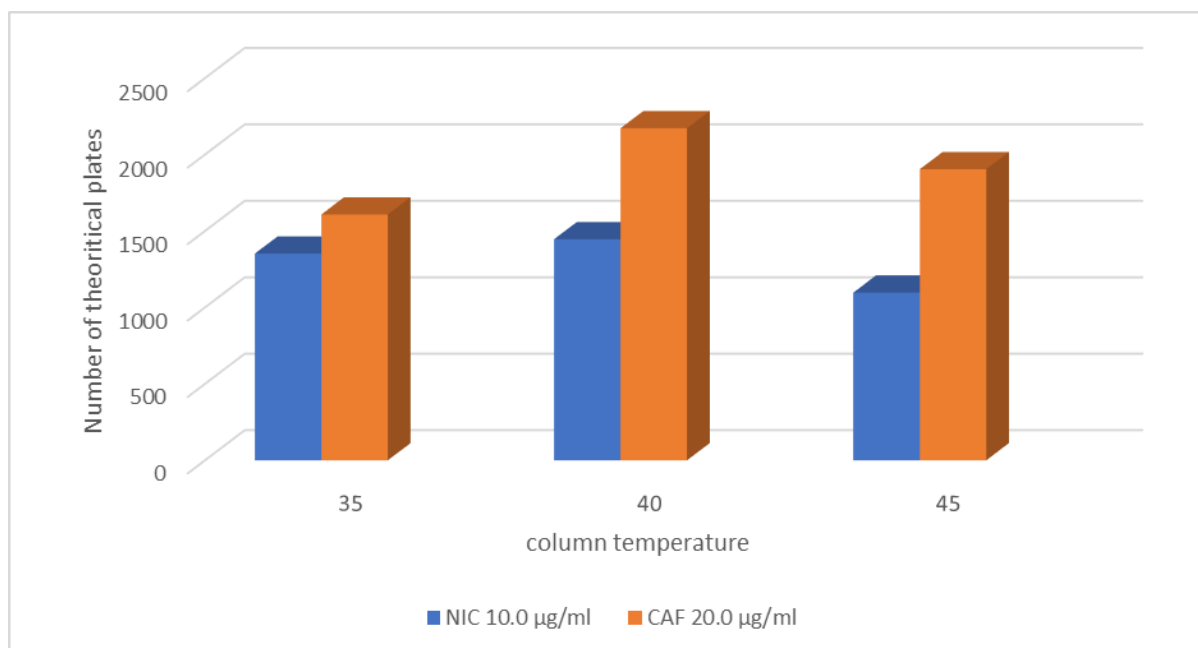
**Fig. S2.** Effect of different pH on the number of theoretical plates of NIC 10.0 µg/ml and CAF 20.0 µg/ml using mobile phase consisting of Glycerol: 0.2 M OPA, (5: 95% v/v). Flow rate, 1.0 mL/min, UV detection at 260 nm; column temperature, 40 degrees Celsius.



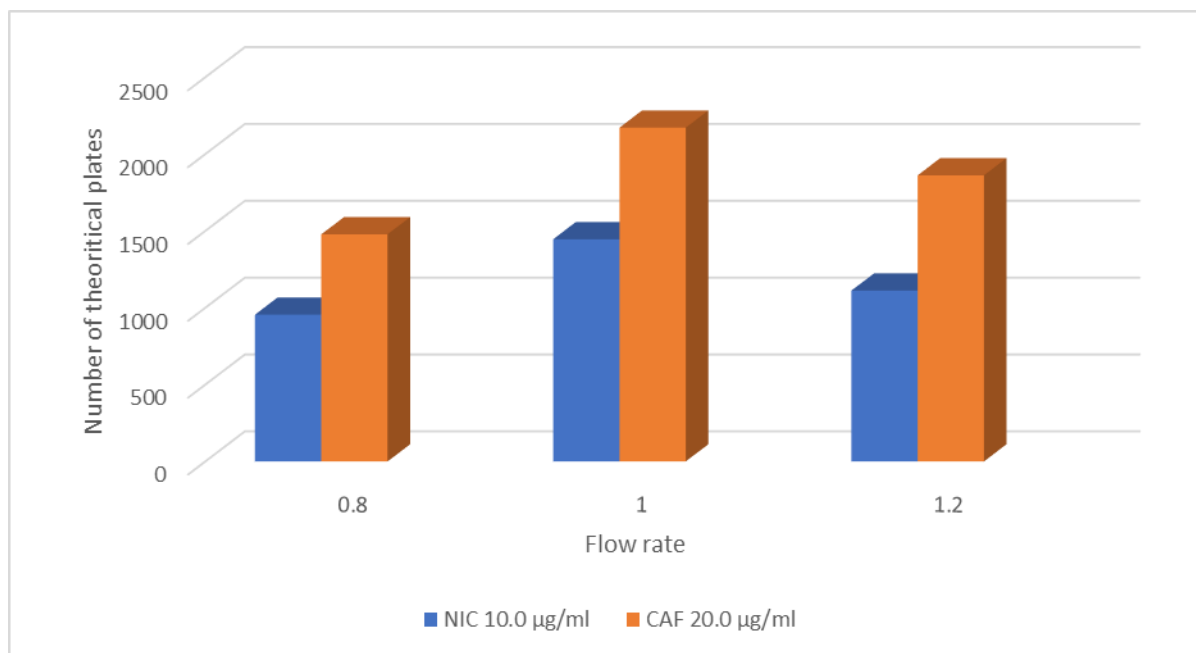
**Fig. S3.** Effect of different % v/v concentration of glycerol on the number of theoretical plates of NIC 10.0 µg/ml and CAF 20.0 µg/ml using a mobile phase containing 0.2 M OPA pH 3.0, Flow rate, 1.0 mL/min, column temperature, 40 degrees Celsius.



**Fig. S4.** Effect of different molarity of OPA on the number of theoretical plates of NIC 10.0 µg/ml and CAF 20.0 µg/ml using a mobile phase mobile phase consisting of Glycerol: 0.2 M OPA, (5: 95% v/v), pH 3.0, Flow rate, 1.0 mL/min, column temperature, 40 degrees Celsius.



**Fig. S5.** Effect of column temperature in Celsius degree on the number of theoretical plates of NIC 10.0 µg/ml and CAF20.0 µg/ml using mobile phase consisting of Glycerol: 0.2 M OPA, (5: 95% v/v), pH 3.0. Flow rate, 1.0 mL/min, UV detection at 260 nm.



**Fig. S6.** Effect of flow rate on the number of theoretical plates of NIC 10.0 µg/ml and CAF 20.0 µg/ml using mobile phase consisting of Glycerol: 0.2 M OPA, (5: 95% v/v), pH 3.0. UV detection at 260 nm, column temperature, 40 degrees Celsius.

**Table S1. Composition of 0.4% (w/w) NIC and 4% (w/w) CAF QMS**

<b>Ingredients</b>	<b>Amount %(w/w)</b>
<b>NIC</b>	0.4
<b>CAF</b>	4
<b>Ethanol</b>	15
<b>Propylene glycol</b>	10
<b>Butylated hydroxytoluene</b>	0.1
<b>Distilled Water to</b>	100



**Table S2: Optimization of the chromatographic parameters for separation of mixture of NIC and CAF by the proposed green chromatographic method.**

Parameter	No. of theoretical plates (N)		Mass distribution ratio (Dm)		Resolution (Rs)	Pressure	Relative retention ( $\alpha$ )	
	NIC	CAF	NIC	CAF	Rs	p.	$\alpha$	
pH	<u>3.0</u>	<u>1445.64</u>	<u>2171.9</u>	<u>1.33</u>	<u>2.6</u>	<u>5.64</u>	<u>1960</u>	<u>1.95</u>
	3.5	1394.7	1601.1	0.98	2.54	4.86	2143	1.51
	4.5	1246.5	1538.8	0.84	2.46	4.88	2613	1.57
	5.5	1198.1	1477.9	0.78	2.5	5.11	2876	1.61
Percentage Ratio of [Glycerol: 0.2 M OPA]	<u>(5:95)</u>	<u>1445.64</u>	<u>2171.9</u>	<u>1.33</u>	<u>2.6</u>	<u>5.64</u>	<u>1960</u>	<u>1.95</u>
	(6:94)	1387.9	1418.2	1.08	2.78	4.91	2589	2.58
	(7:93)	1207.7	1246.5	0.833	2.5	4.66	2789	2.9
	<u>(8:92)</u>	<u>1173.9</u>	<u>1085.5</u>	<u>0.769</u>	<u>2.23</u>	<u>4.15</u>	<u>3098</u>	<u>3.1</u>
Molarity of OPA	0.1	1194.74	1592.8	1.21	2.54	4.83	1960	2.1
	<u>0.2</u>	<u>1445.64</u>	<u>2171.9</u>	<u>1.33</u>	<u>2.6</u>	<u>5.64</u>	<u>1960</u>	<u>1.55</u>
	0.3	1275.3	1831.4	0.833	2.5	4.93	2148	2.8
Temp.	35	1352	1608.1	1.08	2.83	4.88	2780	2.61
	<u>40</u>	<u>1445.64</u>	<u>2171.9</u>	<u>1.33</u>	<u>2.6</u>	<u>5.64</u>	<u>1960</u>	<u>1.55</u>
	45	1097.3	1903.5	1.37	2.8	5.59	1890	2.03
Flow rate	0.8	955.2	1477.9	0.73	2.26	4.62	1597	3.09
	<u>1.0</u>	<u>1445.64</u>	<u>2171.9</u>	<u>1.33</u>	<u>2.6</u>	<u>5.64</u>	<u>1960</u>	<u>2.77</u>
	1.2	1111.8	1862	1.12	3.12	5.33	2464	1.55

Where: Number of theoretical plates (N) =  $5.45(t_R/W_{h/2})^2$ ,

- $W_{h/2}$  is the half peak width
- Resolution (R) =  $2 \Delta t_R/W_1 + W_2$ ,
- $W_1$  and  $W_2$  are the peaks width of the two components measured at their bases.
- Relative retention ( $\alpha$ ) =  $Dm_2/Dm_1$ ,

Mass distribution ratio (Dm) =  $t_R - t_m / t_m$ ,  $t_R$  is time of retention of the measured substance measured from the injection point and  $t_m$  is the retention time of a non-retained marker.

**Table S3: Robustness results for determination of NIC and CAF by the proposed green chromatographic method.**

Parameter	Value	NIC			CAF		
		% Recovery	Mean % recovery $\pm$ SD	%RSD	% Recovery	Mean % recovery $\pm$ SD	%RSD
<b>pH</b>	2.9	99.52			102.54		
	<u>3.0</u>	101.02	100.49 $\pm$ 0.84	0.84	99.76	101.03 $\pm$ 1.41	1.40
	3.1	100.94			100.78		
<b>Glycerol concentration %</b>	4.6	98.42			97.51		
	<u>5.0</u>	98.94	99.19 $\pm$ 0.92	0.93	98.74	98.28 $\pm$ 0.67	0.68
	5.4	100.21			98.58		
<b>Temperature (<math>^{\circ}</math>C)</b>	38	98.51			100.81		
	<u>40</u>	100.78	100.34 $\pm$ 1.65	1.64	101.92	101.87 $\pm$ 1.03	1.01
	42	101.73			102.87		
<b>Flow rate</b>	0.9	98.24			99.52		
	<u>1.0</u>	100.58	98.78 $\pm$ 1.6	1.62	98.74	99.67 $\pm$ 1.01	1.01
	1.1	97.53			100.75		

**Table S4. Kinetic modeling of release data from NIC/CAF QMS and trade marketing formulations through cellophane membrane.**

Drug	Formula	R <sup>2</sup>			Release Order	Korsmeyer et al		Main transport mechanism
		Zero-Order	First Order	Higuchi Model		R <sup>2</sup>	n	
NIC	Gum	0.6004	0.3249	0.7647	Diffusion	0.6440	1.26	Super case II
	QMS	0.7885	0.6233	0.9348		0.9078	0.62	Non-Fickian
CAF	Patch	0.8141	0.7058	0.9699		0.9513	0.50	
	QMS	0.7580	0.6178	0.9313		0.9111	0.56	

**Table S5. Assessment of green analytical performance of the developed HPLC/UV method using analytical Eco-scale.**

Proposed method		Reported method [21]	
Reagents	PPs	Reagents	PPs
Glycerol	0	Acetonitrile	4
OPA	3	Methanol	5
TEA	2	Diethyl amine	2
		Acetic acid	4
	Σ5		Σ15
<b>Instruments</b>		<b>Instruments</b>	
HPLC/UV	1	HPLC/UV	1
Occupational hazards	0	Occupational hazards	0
waste	3	waste	5
	Σ4		Σ6
<b>Total PPs: 9</b>		<b>Total PPs: 21</b>	
<b>Score: 91 (excellent green analysis)</b>		<b>Score: 79 (acceptable green analysis)</b>	

**Table S6. Comparison of the developed green HPLC method to reported methods**

<b>Method</b>	<b>Mobile phase</b>	<b>Run time</b>	<b>Analysis throughput</b>	<b>Sample</b>	<b>Limitations</b>	<b>Ref.</b>
<b>HPLC-UV</b>	Water: methanol: buffer acetate (pH 4.66): acetonitrile: acetic acid (50:29:20:2:1, v/v)	10 min	6 samples per hour	Meconium	-Utilization of hazardous solvents. -Long runtime.	[19]
<b>HPLC-UV</b>	Methanol: buffer (5 mM sodium octane sulphonate, 10 mM citric acid adjusted to pH 5.8 with triethylamine) (20:80)	> 22 min	2 samples per hour	Human milk	-Utilization of hazardous solvents. -Long runtime. - high buffer loads.	[20]
<b>GC-MS/MS</b>	Helium 5.0	> 11 min	5 samples per hour	Chocolate	Utilization of highly sophisticated and expensive instrumentation. Tedious sample preparation.	[21]
<b>HPLC-UV</b>	Glycerol: orthophosphoric acid (0.2 M) adjusted to pH 3.0 using 0.05 M triethylamine (5:95, v/v)	5 min	12 samples per hour	Dosage forms and artificial saliva.	-	This work