

Miktoarm star-polypept(o)ide-based polyion complex micelles for the delivery of large nucleic acids

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Synthesis

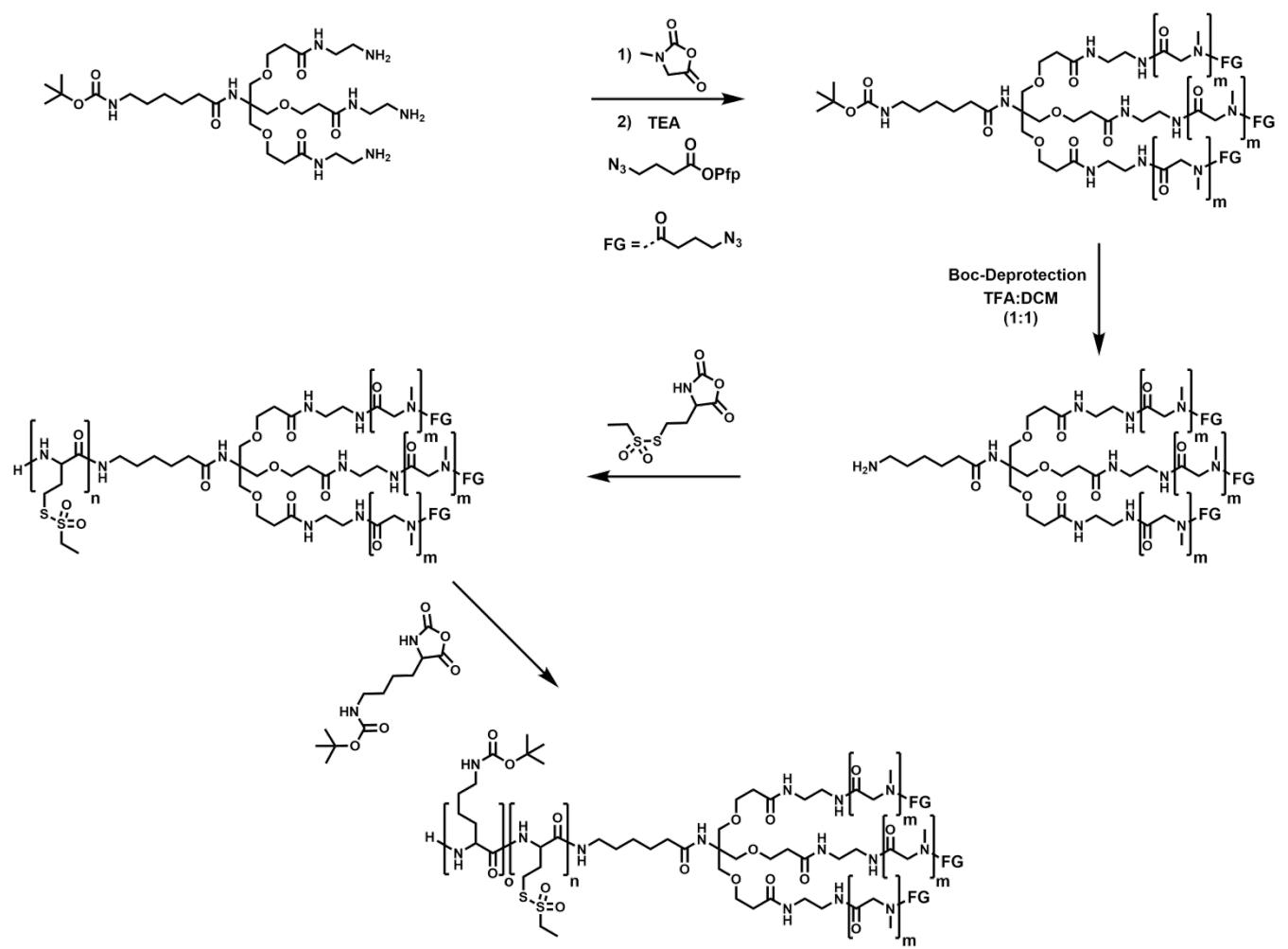


Figure S1. Synthetic pathway for the realization of cross-linkable AB_3 PeptoMiktoStars.

Monomers

Sarcosine-*N*-carboxyanhydride (Sar-NCA)

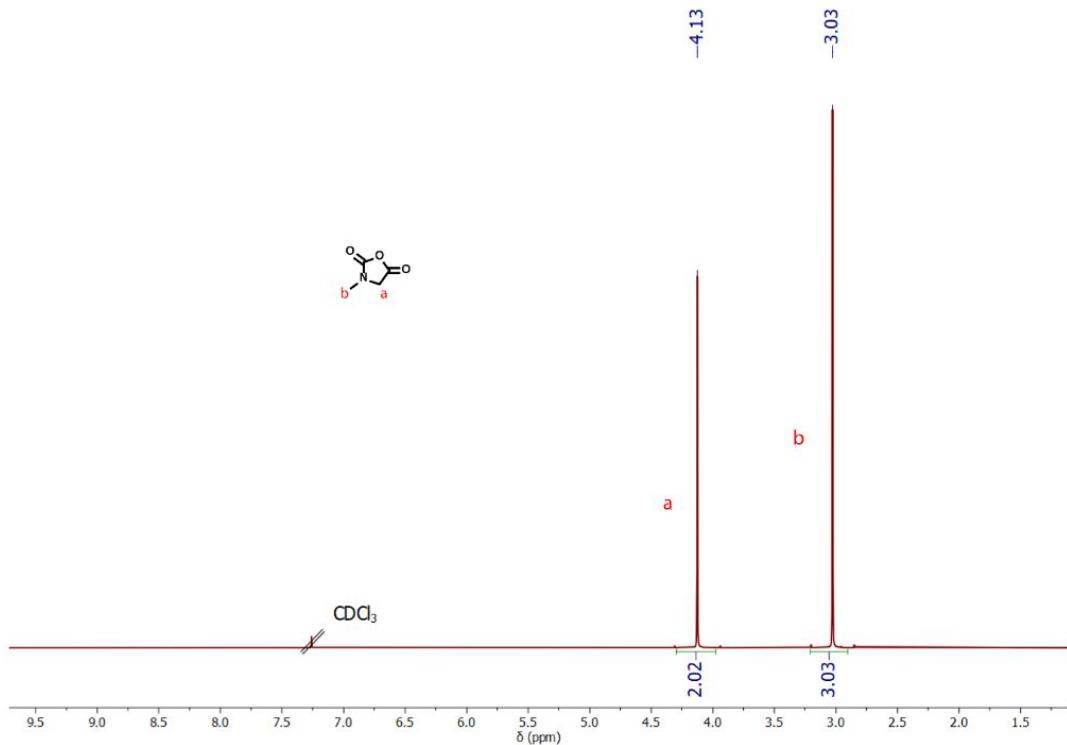


Figure S2. ^1H NMR spectrum of sarcosine-*N*-carboxyanhydride (Sar-NCA) in CDCl_3 .

S-Ethylsulfonyl-*L*-homocysteine-*N*-carboxyanhydride (*Hcy(SO*₂*Et*)*-NCA*)

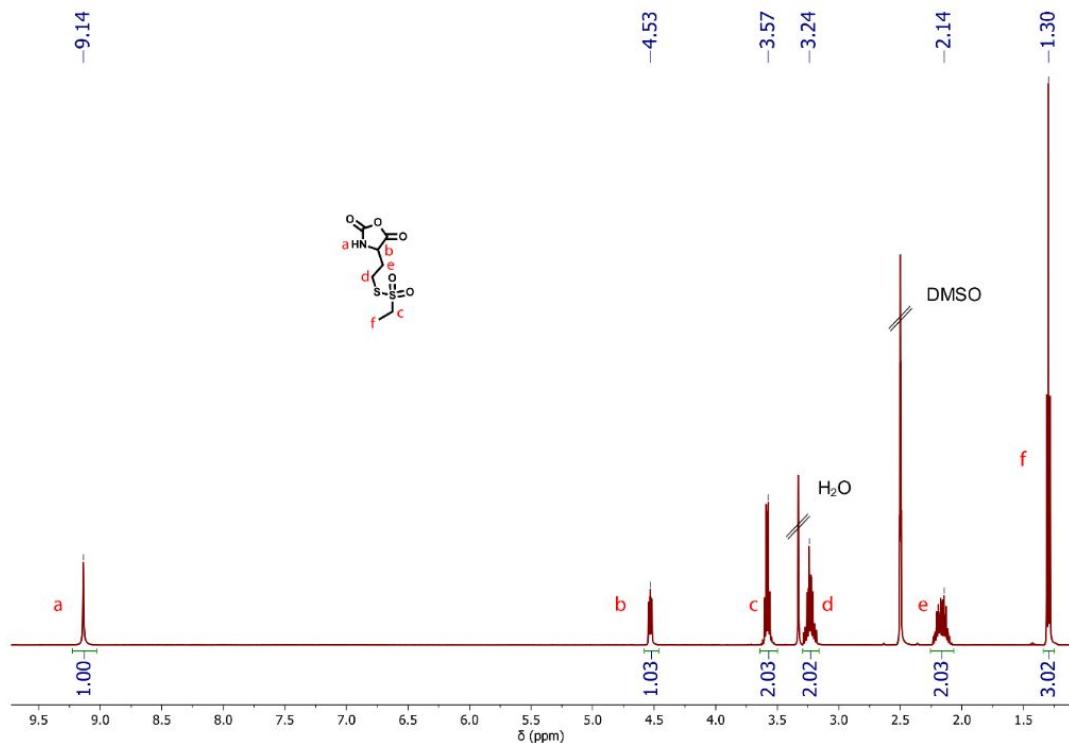


Figure S3. ¹H NMR spectrum of *S*-ethylsulfonyl-*L*-homocysteine-*N*-carboxyanhydride (*Hcy(SO*₂*Et*)*-NCA*) in *DMSO-d*₆.

N-*ε*-*tert*-butyloxycarbonyl-L-lysine-*N*-carboxyanhydride (*Lys(Boc)-NCA*)

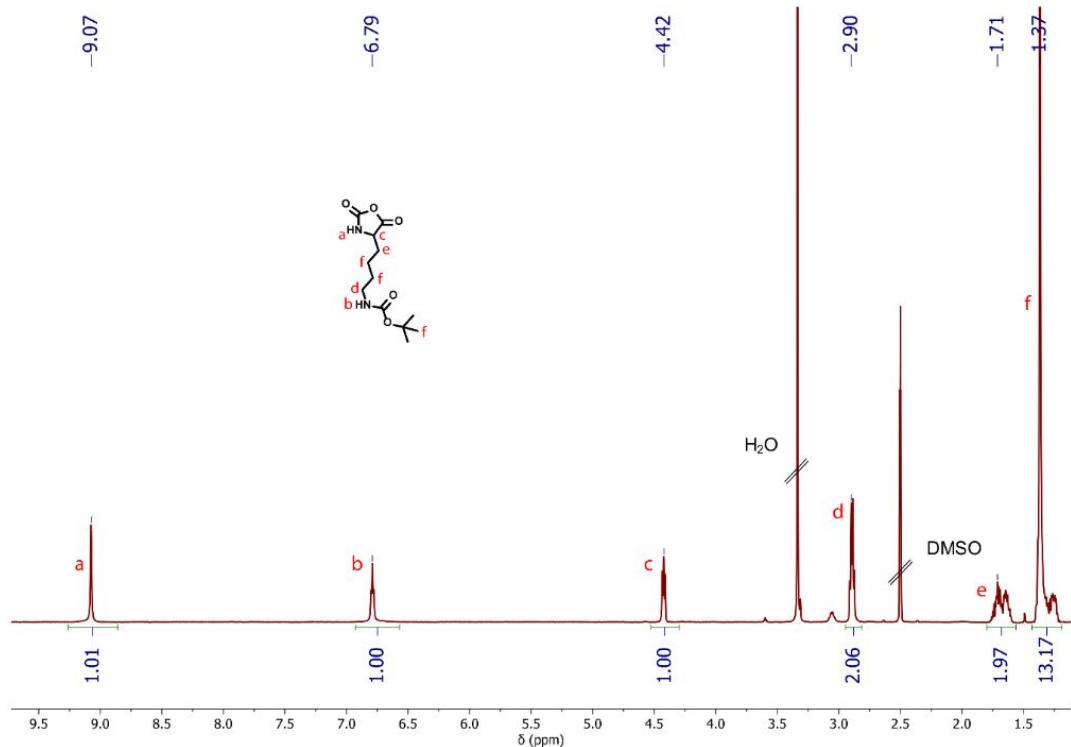


Figure S4. ^1H NMR spectrum of *N*-*ε*-*tert*-butyloxycarbonyl-L-lysine-*N*-carboxyanhydride (*Lys(Boc)-NCA*) in $\text{DMSO}-d_6$.

Initiators

Tetrafunctional Initiator

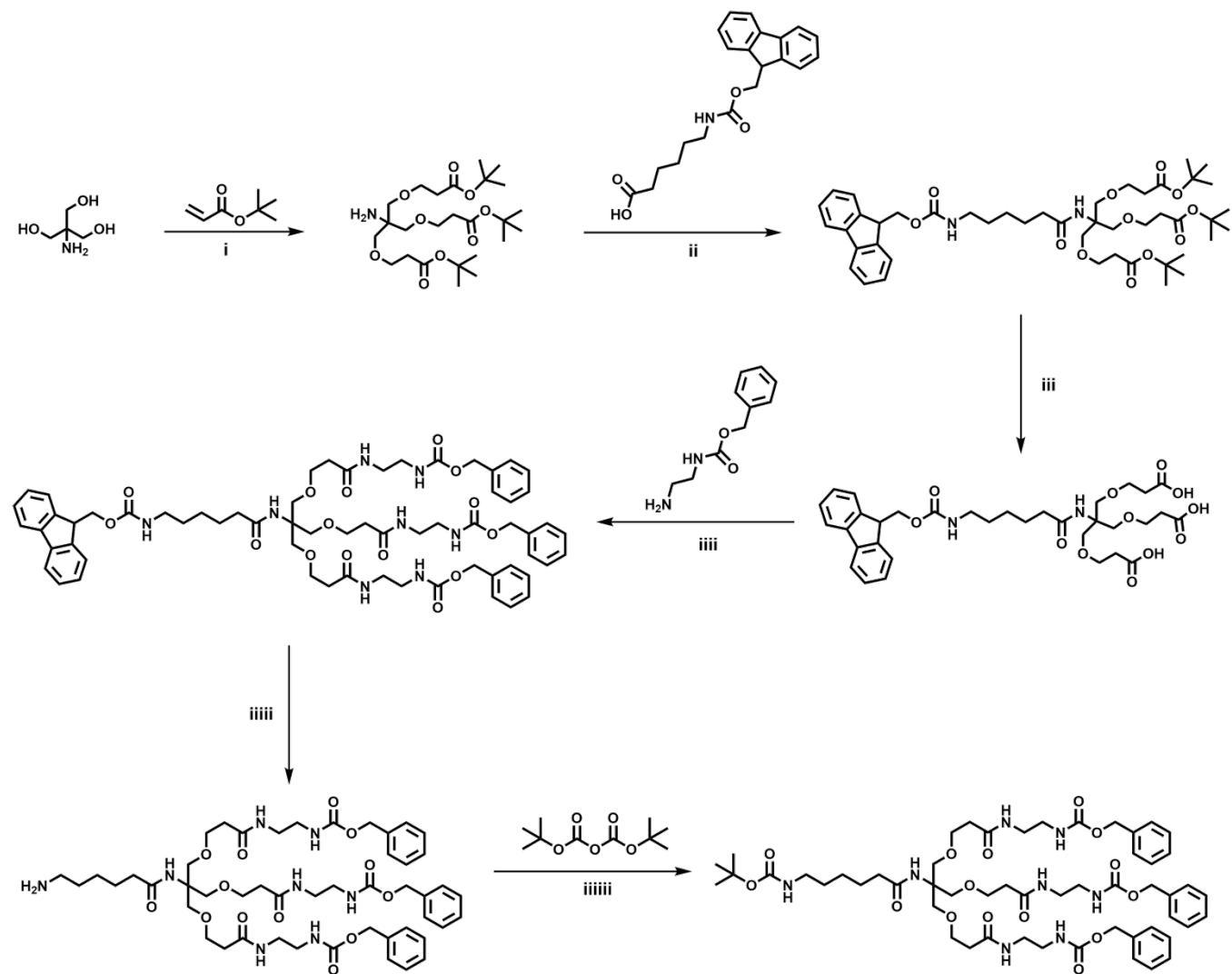


Figure S5. Synthesis of the tetrafunctional initiator Boc-Ahx-Tris{[2-(*N*-Cbz-ethylendiaminecarbonyl)ethoxy]methyl}methyl-amide. i) NaOH, DMSO, 15 °C, 24 h, yield 30 %. ii) PyBOP, DIPEA, THF, rt, yield: 93 %. iii) TFA/DCM (1/1), 0 °C, 1 h, yield: quantitative. iv) PyBOP, DIPEA, DMF/DCM (1/1), rt, 24 h, yield: 90 %. v) DMF/Piperidine (4/1), 0 °C, 1 h, yield: quantitative, vi) TEA, DCM, rt, 24 h, yield: 92 %.

*Synthesis of Tris{[2-(*tert*-butoxycarbonyl)ethoxy]methyl}methylamine (i)*

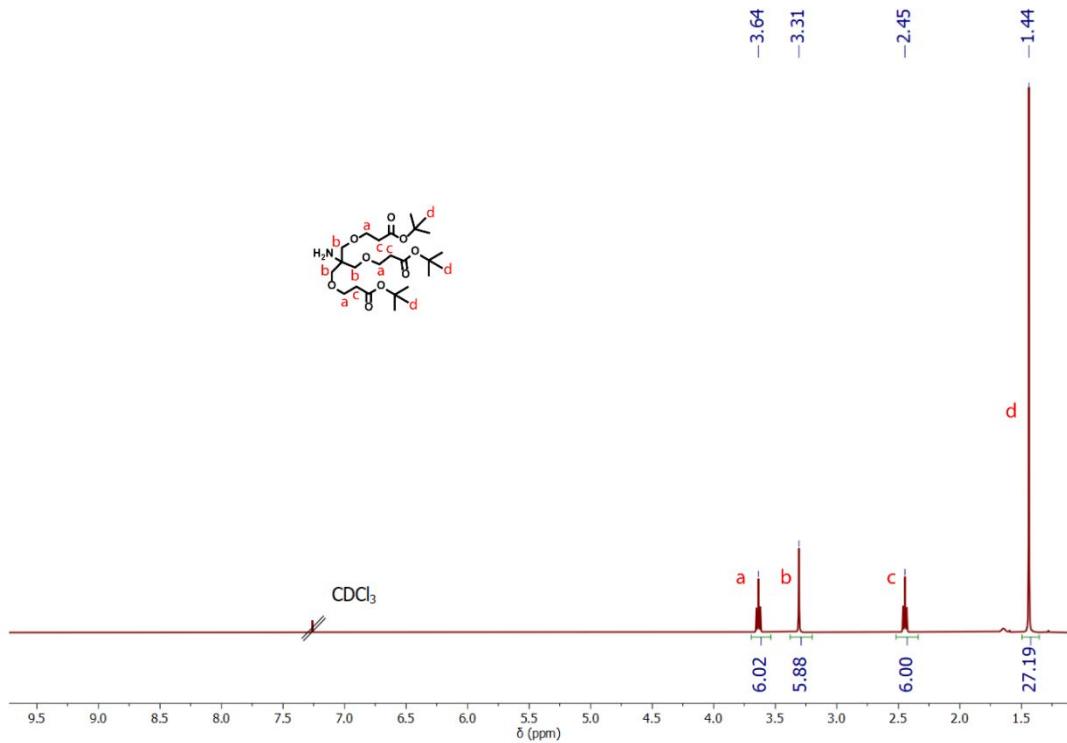


Figure S6. ^1H NMR spectrum of (i) Tris{[2-(*tert*-butoxycarbonyl)ethoxy]methyl}methylamine in CDCl_3 .

Synthesis of Fmoc-Ahx-Tris{[2-(tert-butoxycarbonyl)ethoxy]methyl}methylamide (ii)

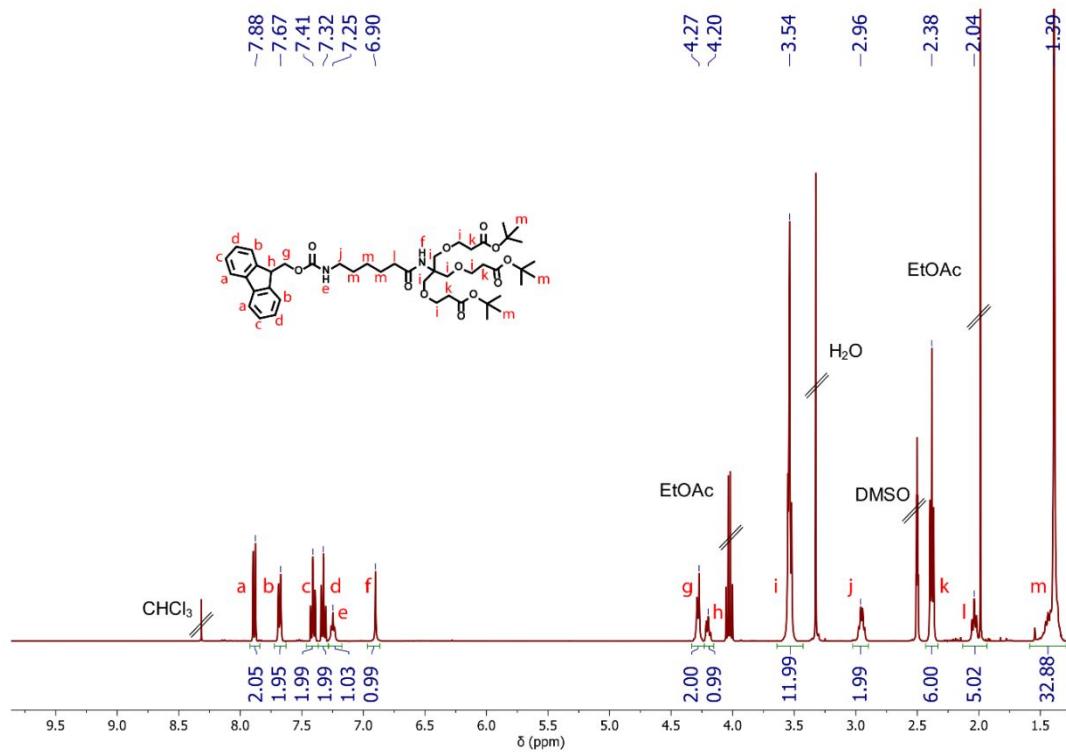


Figure S7. ^1H NMR spectrum of (ii) Fmoc-Ahx-Tris{[2-(tert-butoxycarbonyl)ethoxy]methyl}methylamide in $\text{DMSO}-d_6$.

Synthesis of Fmoc-Ahx-Tris[2-(carboxyethoxy)methyl]methylamide (iii)

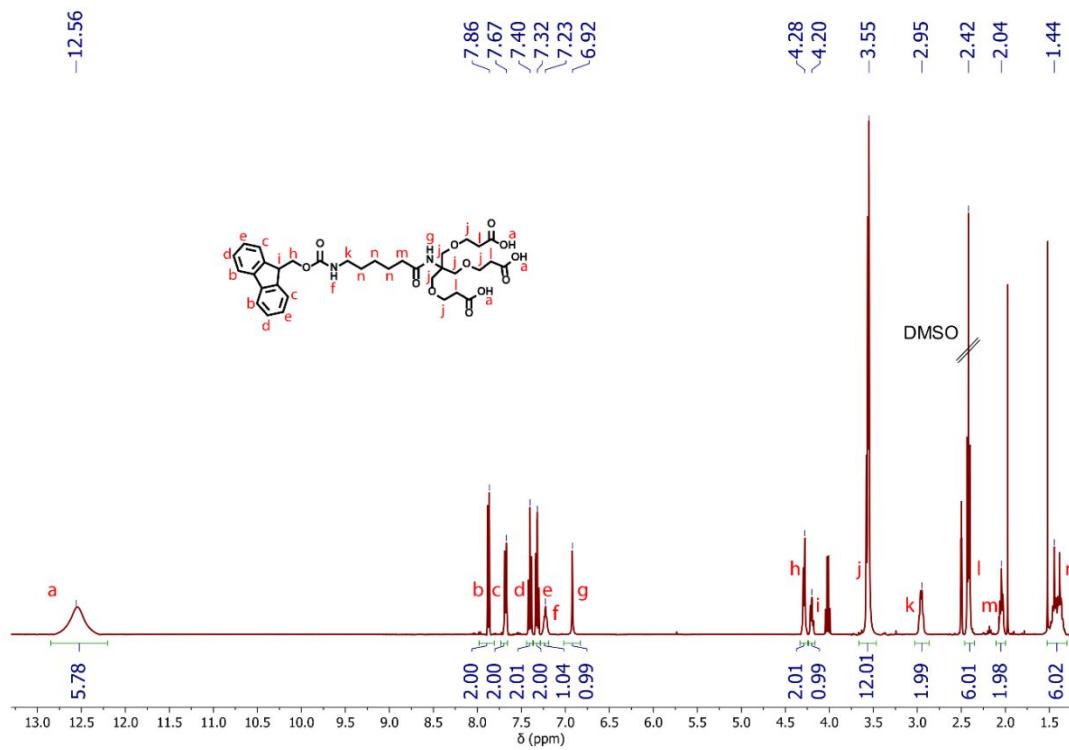


Figure S8. ^1H NMR spectrum of (iii) Fmoc-Ahx-Tris[2-(carboxyethoxy)methyl]methylamide in $\text{DMSO}-d_6$.

Synthesis of Fmoc-Ahx-Tris{[2-(N-Cbz-ethylendiaminecarbonyl)ethoxy]methyl}methylamide (iii)

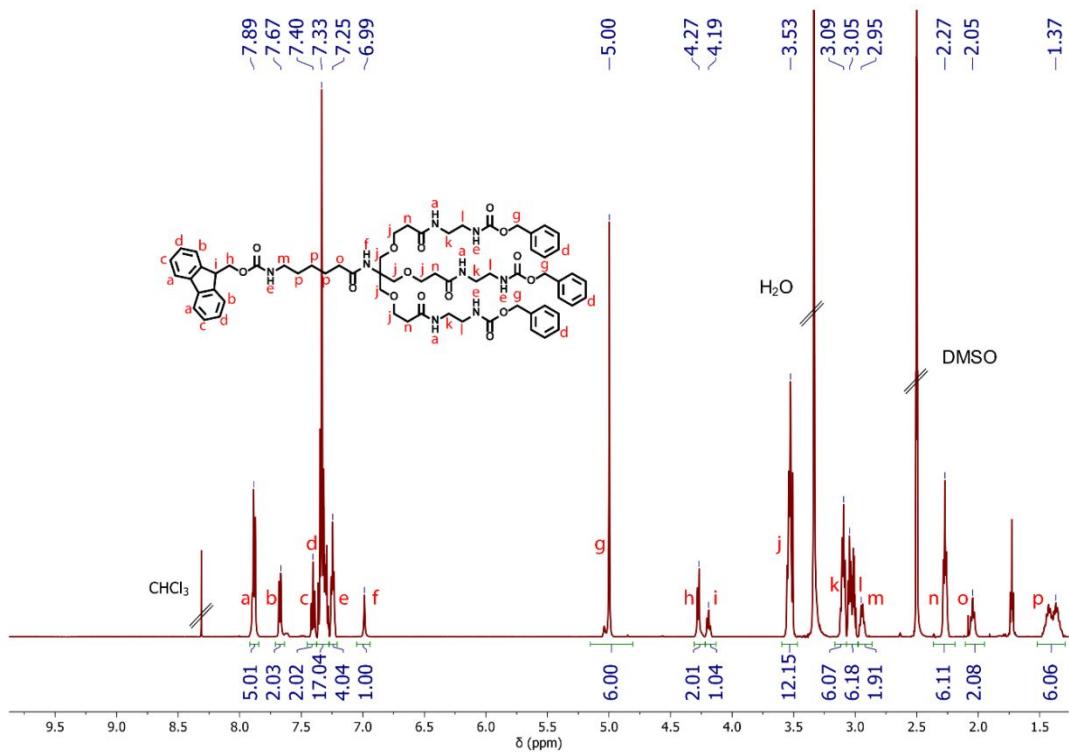


Figure S9. ^1H NMR spectrum of (iii) Fmoc-Ahx-Tris{[2-(*N*-Cbz-ethylendiaminecarbonyl)ethoxy]methyl}methylamide in $\text{DMSO}-d_6$.

Synthesis of Ahx-Tris{[2-(N-Cbz-ethylendiaminecarbonyl)ethoxy]methyl}methylamide (iiii)

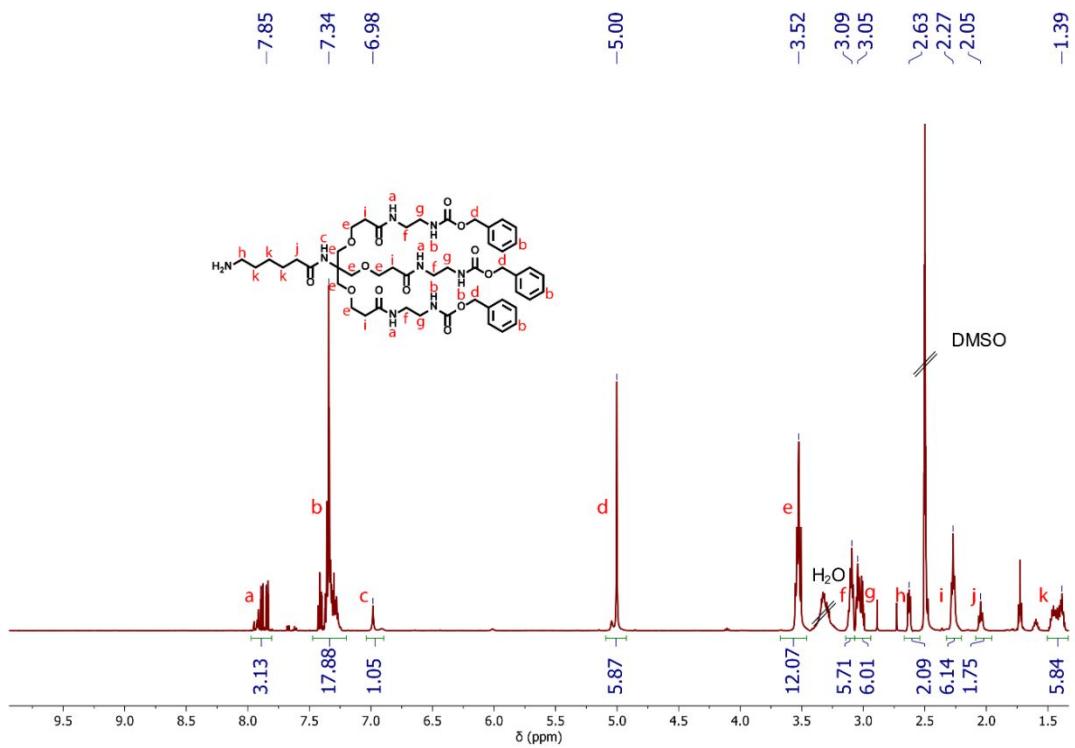


Figure S10. ^1H NMR spectrum of (iiii) Ahx-Tris{[2-(N-Cbz-ethylendiaminecarbonyl)ethoxy]methyl}methylamide in $\text{DMSO}-d_6$.

Synthesis of Boc-Ahx-Tris{[2-(*N*-Cbz-ethylendiaminecarbonyl)ethoxy]methyl}methylamide (iiiiii)

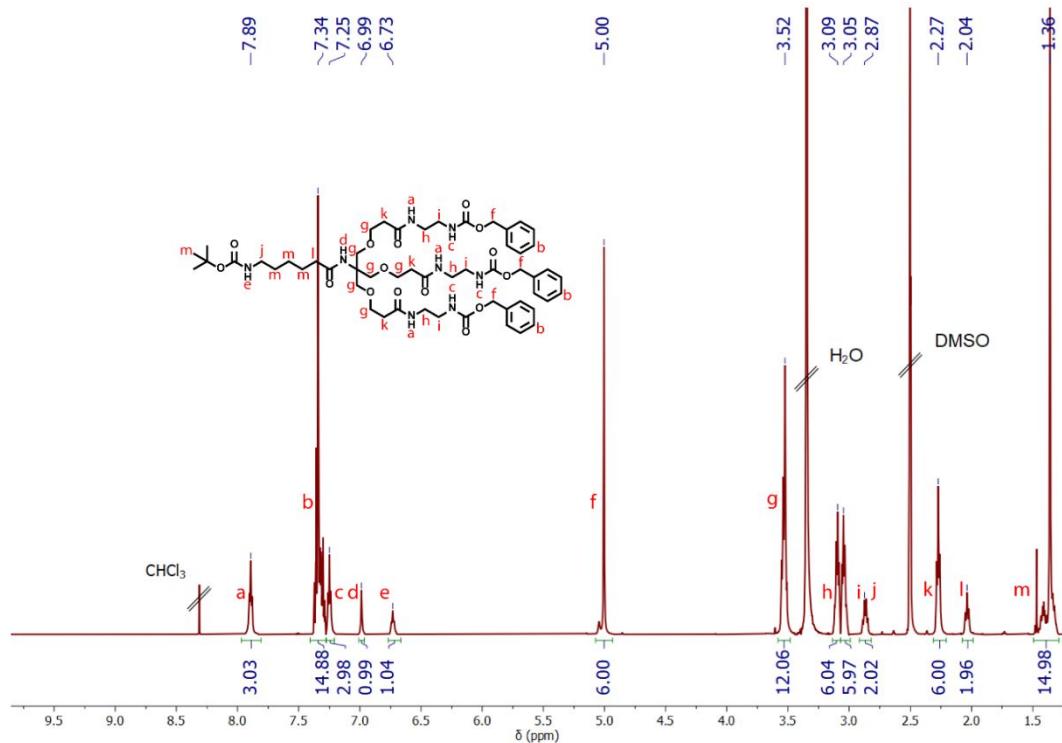


Figure S11. ^1H NMR spectrum of iiii) Boc-Ahx-Tris{[2-(*N*-Cbz-ethylendiaminecarbonyl)ethoxy]methyl}methylamide) in $\text{DMSO}-d_6$.

Heptafunctional Initiator

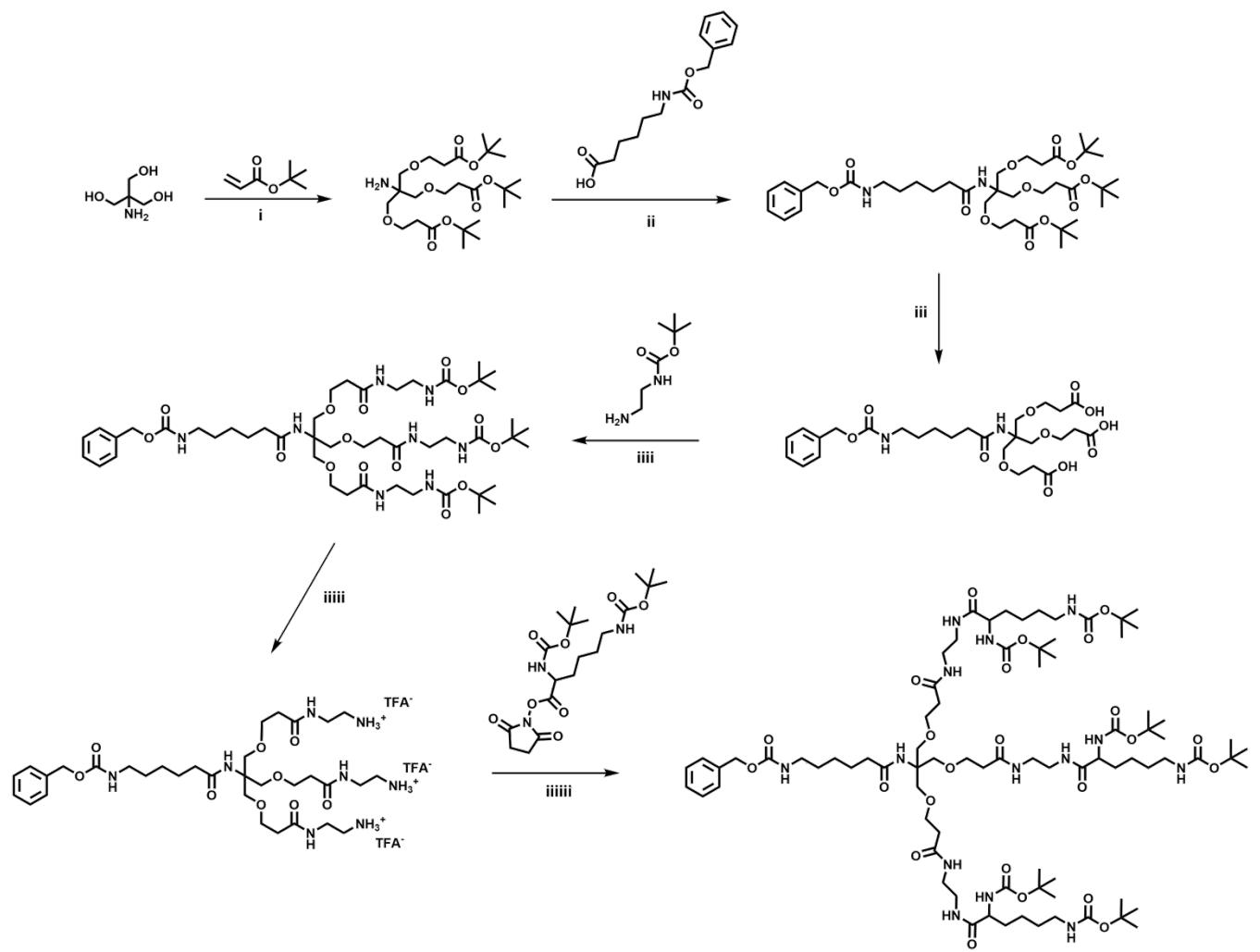


Figure S12. Synthesis of the Heptafunctional initiator. i) $\text{NaOH}, \text{DMSO}, 15^\circ\text{C}, 24\text{ h}$, yield 30 %. ii) $\text{PyBOP}, \text{DIPEA}, \text{THF}, \text{rt}$, yield: 91 %. iii) $\text{TFA/DCM (1/1)}, 0^\circ\text{C}, 1\text{ h}$, yield: quantitative. iv) $\text{PyBOP}, \text{DIPEA}, \text{DMF/DCM (2/1)}, \text{rt}, 24\text{ h}$, yield: 55 %. v) $\text{TFA/DCM (1/1)}, 10^\circ\text{C}, 1\text{ h}$, yield: quantitative. vi) $\text{DIPEA}, \text{DCM/DMF (1/1)}, \text{rt}, 24\text{ h}$, yield: 63 %.

Synthesis of Cbz-Ahx-Tris{[2-(tert-butoxycarbonyl)ethoxy]methyl}methylamide (ii)

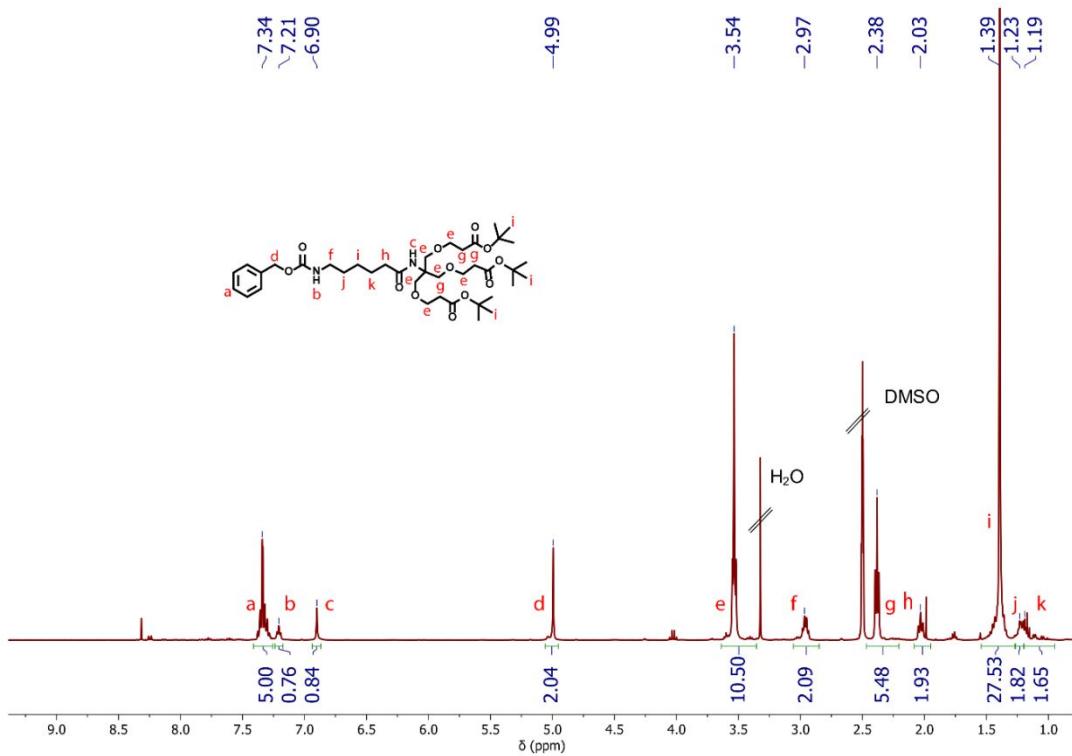


Figure S13. ^1H NMR spectrum of (ii) Cbz-Ahx-Tris{[2-(tert-butoxycarbonyl)ethoxy]methyl}methylamide in $\text{DMSO}-d_6$.

Synthesis of Cbz-Ahx-Tris[2-(carboxyethoxy)methyl]methylamide (iii)

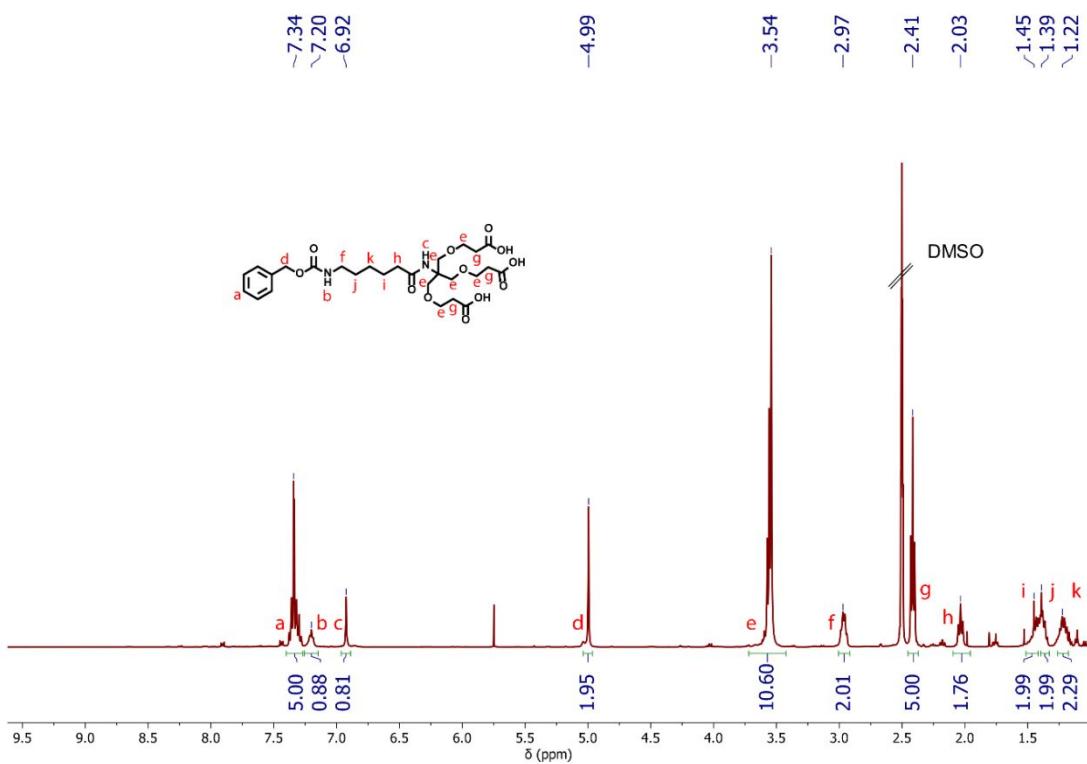


Figure S14. ^1H NMR spectrum of (iii) Cbz-Ahx-Tris[2-(carboxyethoxy)methyl]methylamide in DMSO- d_6 .

Synthesis of Cbz-Ahx-Tris{[2-(N-Boc-ethylendiaminecarbonyl)ethoxy]methyl}methylamide (iiii)

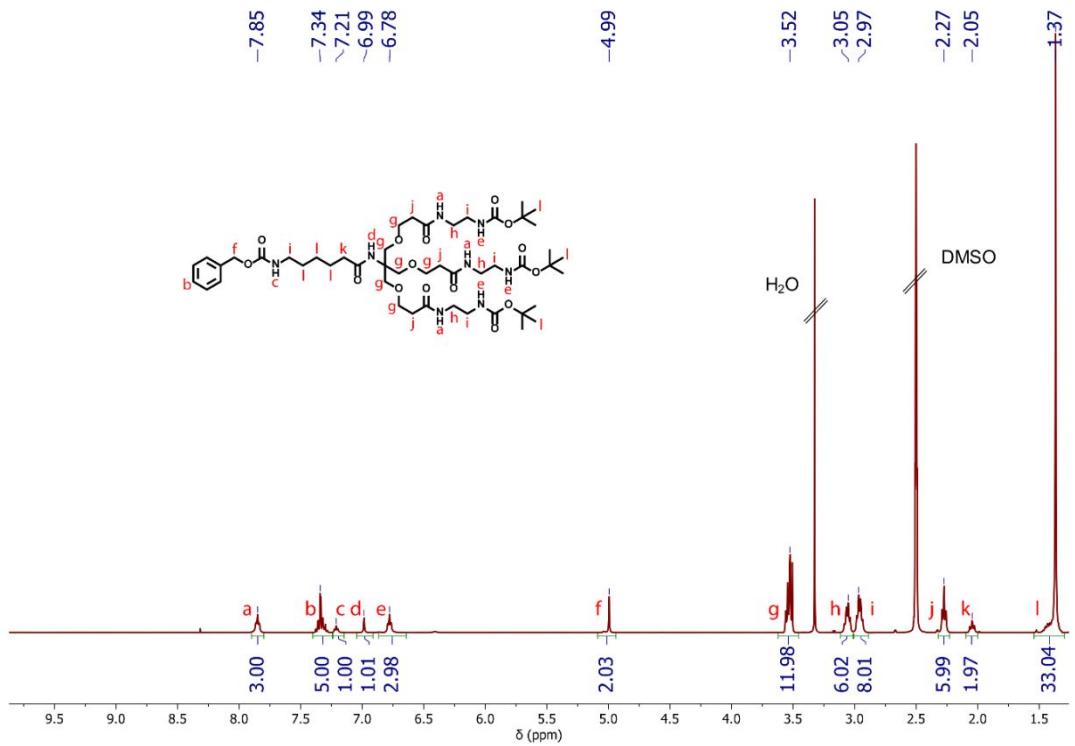


Figure S15. ^1H NMR spectrum of (iiii) Cbz-Ahx-Tris{[2-(N-Boc-ethylendiaminecarbonyl)ethoxy]methyl}methylamide in $\text{DMSO}-d_6$.

Boc-Deprotection of the tetrafunctional initiator (iiii)

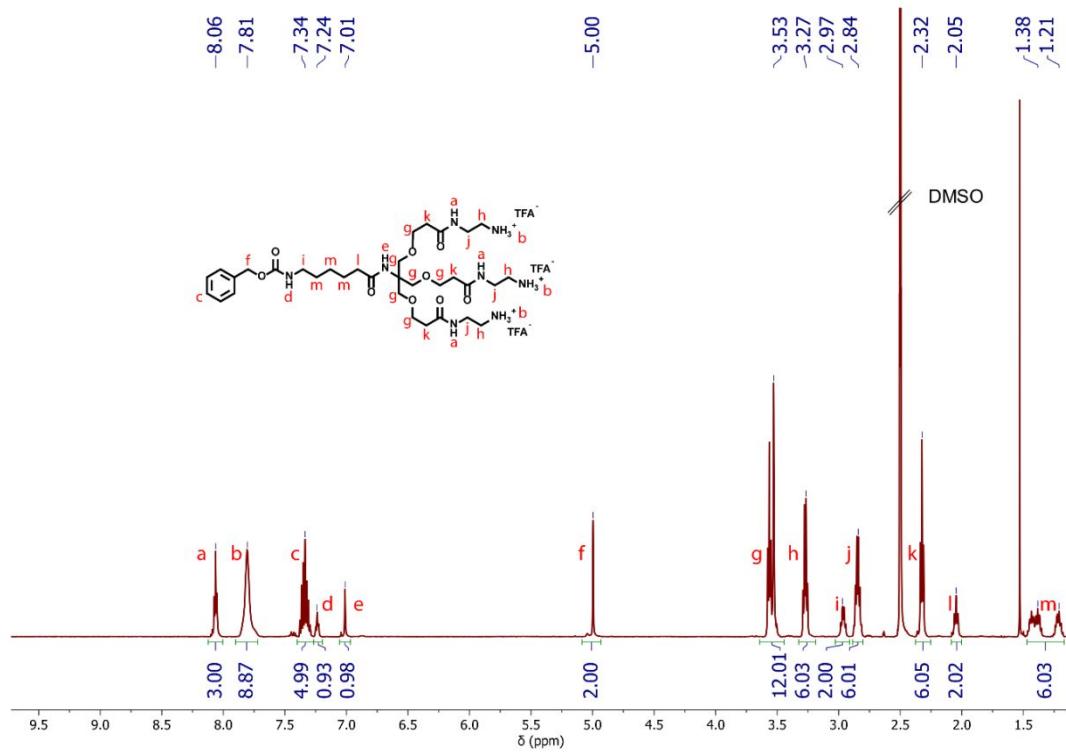


Figure S16. ¹H NMR spectrum of (iiii) the Boc-deprotected tetrafunctional initiator in DMSO-*d*₆.

Synthesis of the heptafunctional initiator (iiiiii)

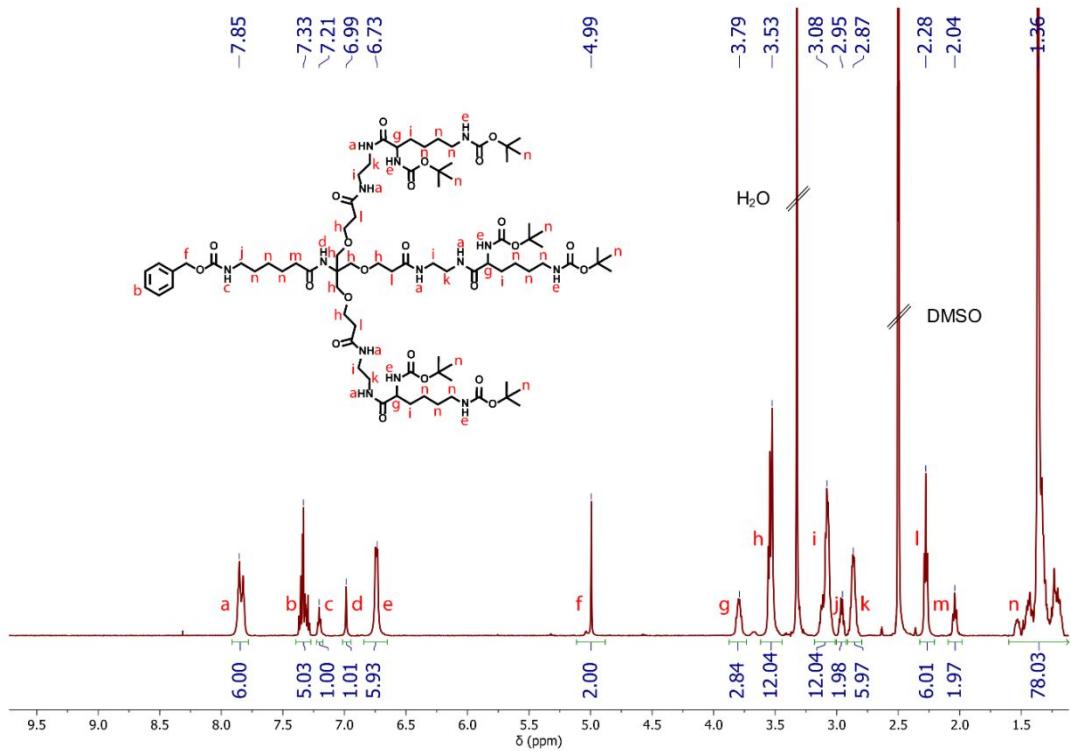


Figure S17. ^1H NMR spectrum of (iiiiii) the heptafunctional initiator in $\text{DMSO}-d_6$.

Initiator Deprotection for Macroinitiator Synthesis

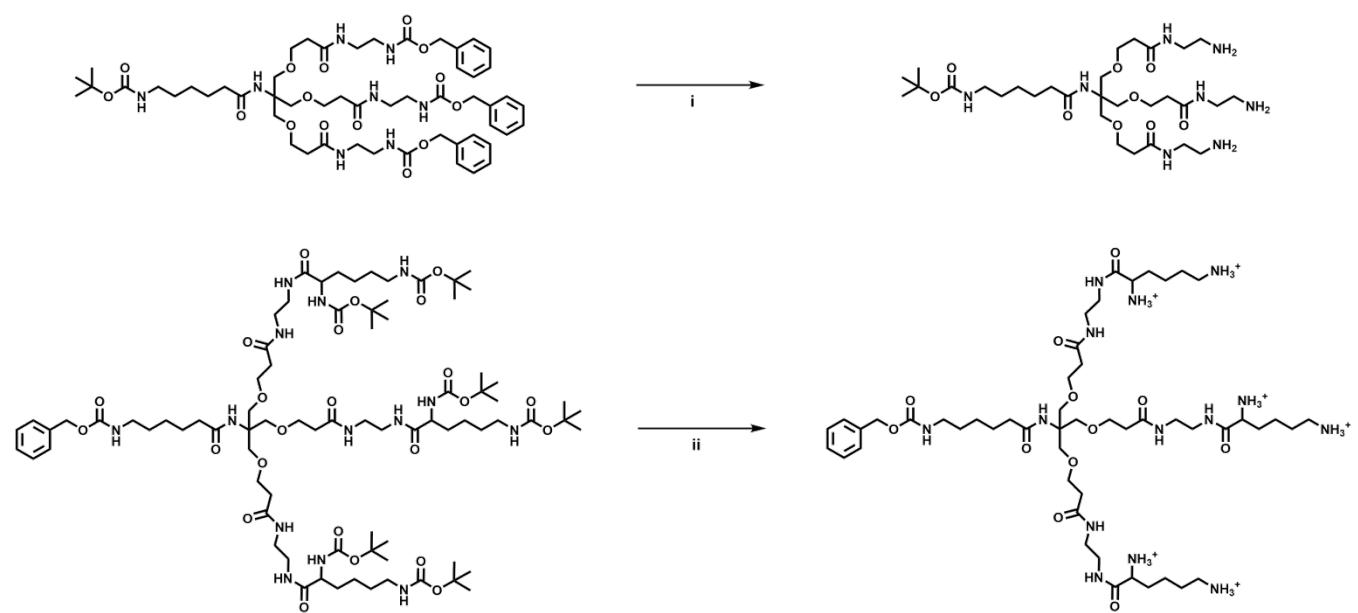


Figure S18. Applied deprotection methods as prerequisite for macroinitiator synthesis. i) Pd/C (10 wt%), H_2 , MeOH , rt, 2d, yield: quantitative. ii) TFA/DCM (1/1), 0 °C, 1 h, yield: quantitative.

Cbz-deprotection of the tetrafunctional initiator (i)

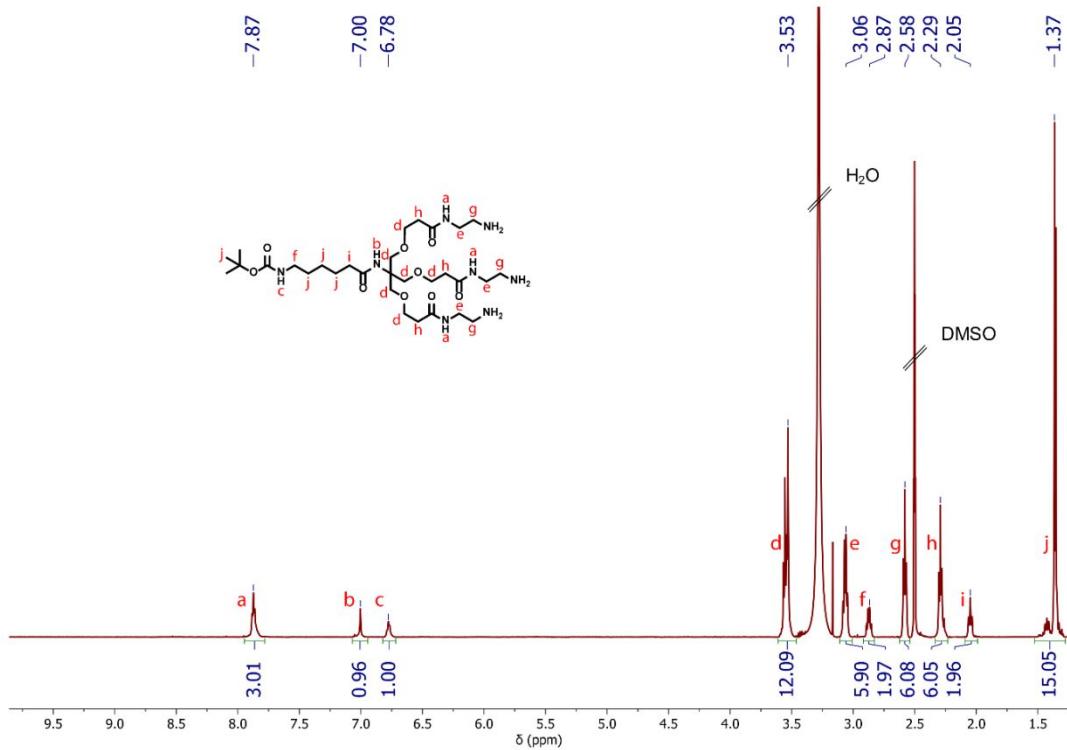


Figure S19. ^1H NMR spectrum of i) Cbz-deprotected tetrafunctional initiator in $\text{DMSO}-d_6$.

Boc-deprotection of the heptafunctional initiator (ii)

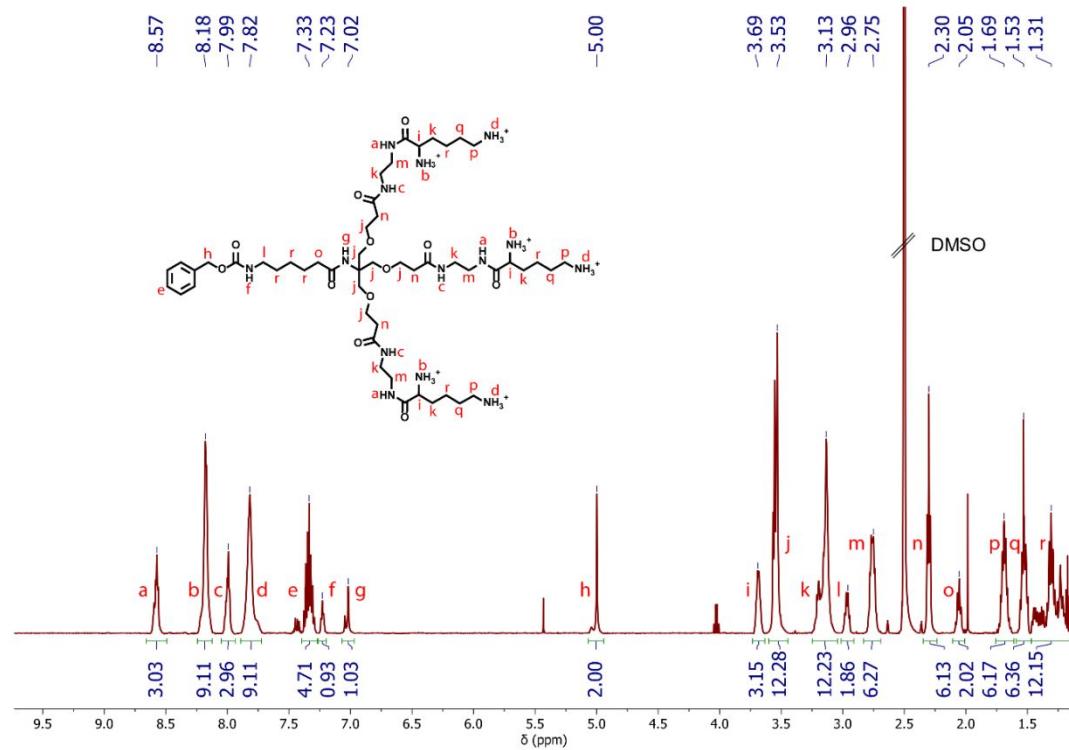


Figure S20. ¹H NMR spectrum of ii) Boc-deprotected heptafunctional initiator in *DMSO-d*₆.

Polypeptoid Macroinitiators

*Three-Arm Macroinitiator Synthesis (Boc-(*p*Sar₁₀₀-N₃)₃)*

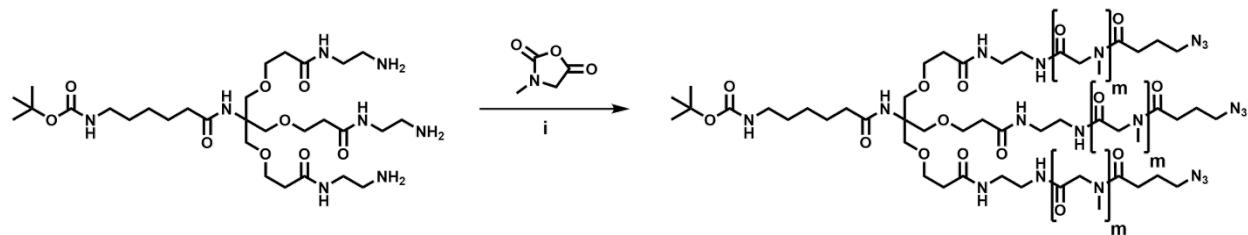


Figure S21. Synthesis of the three-arm macroinitiator with *m* = 100 i) DMF, 0 °C, 3d; pentafluorophenyl-4-azido-butanoat, DIPEA, rt, 1d, yield: 77 %.

Synthesis of three-arm pSar-macroinitiator (i)

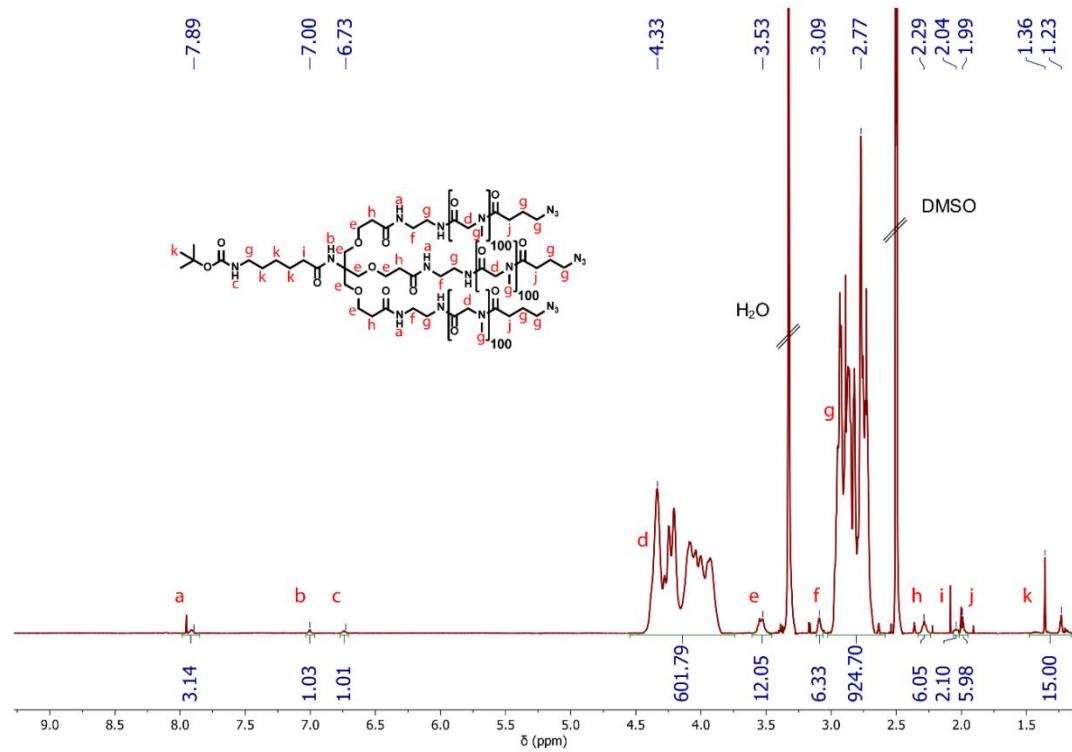


Figure S22. ^1H NMR spectrum of i) three-arm macroinitiator in $\text{DMSO}-d_6$.

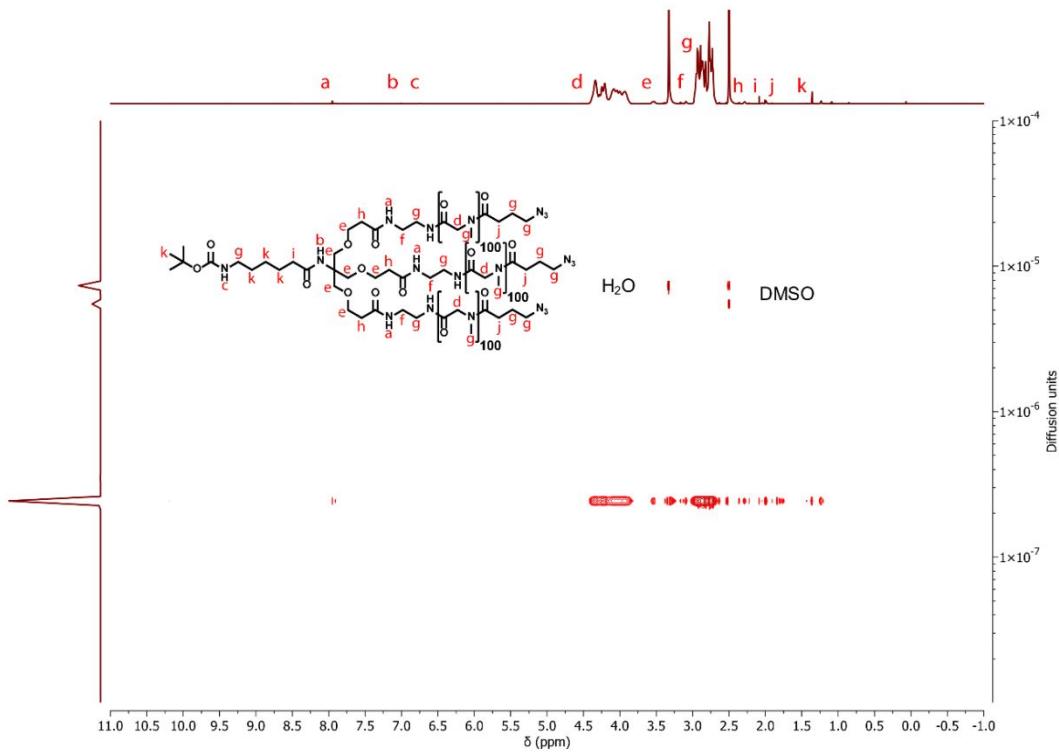


Figure S23. ¹H DOSY spectrum of i) three-arm macroinitiator in *DMSO-d*₆.

*Six-Arm Macroinitiator Synthesis (Cbz-(*p*Sar₅₀-N₃)₆)*

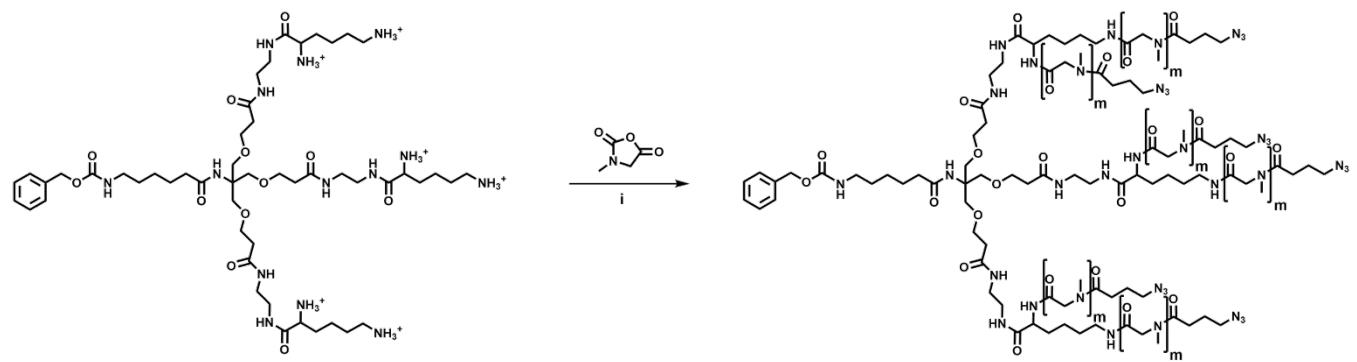


Figure S24. Synthesis of the six-arm macroinitiators with $m = 50$ i) DMF, DIPEA, 0 °C, 3d;

pentafluorophenyl-4-azido-butanoat, DIPEA, rt, 1d, yield: 72 %.

Synthesis of six-arm pSar-macroinitiator (i)

Boc-Deprotection of the Three-Arm Macroinitiator ($\text{NH}_2\text{-}(p\text{Sar}_{100}\text{-N}_3)_3$)

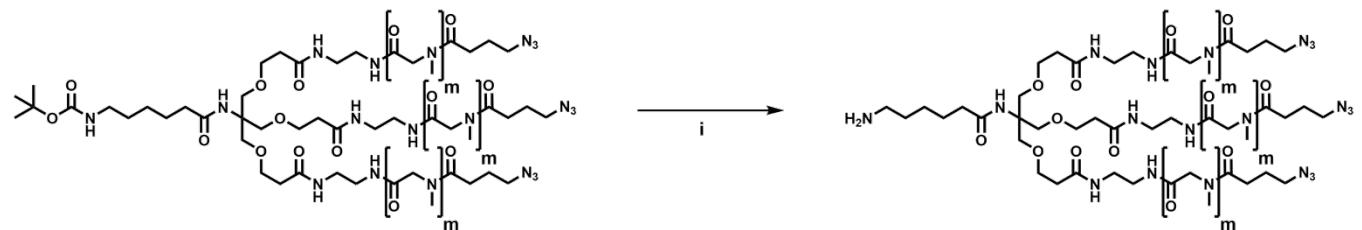


Figure S25. Boc-deprotection of the three-arm macroinitiator i) TFA/H₂O (1/1), 0 °C, 2 h, yield: 65%.

*Boc-deprotection of three-arm macroinitiator Boc-(*p*Sar₁₀₀-N₃)₃ (i)*

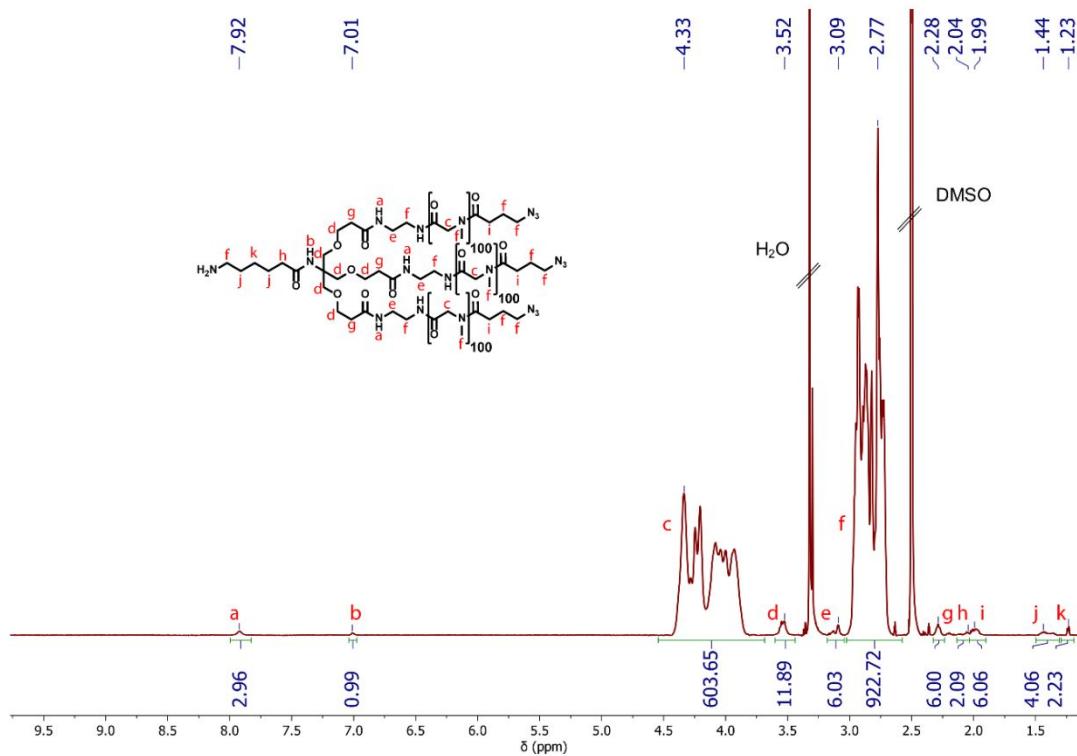


Figure S26. ¹H NMR spectrum of i) deprotected three-arm macroinitiator in DMSO-*d*₆.

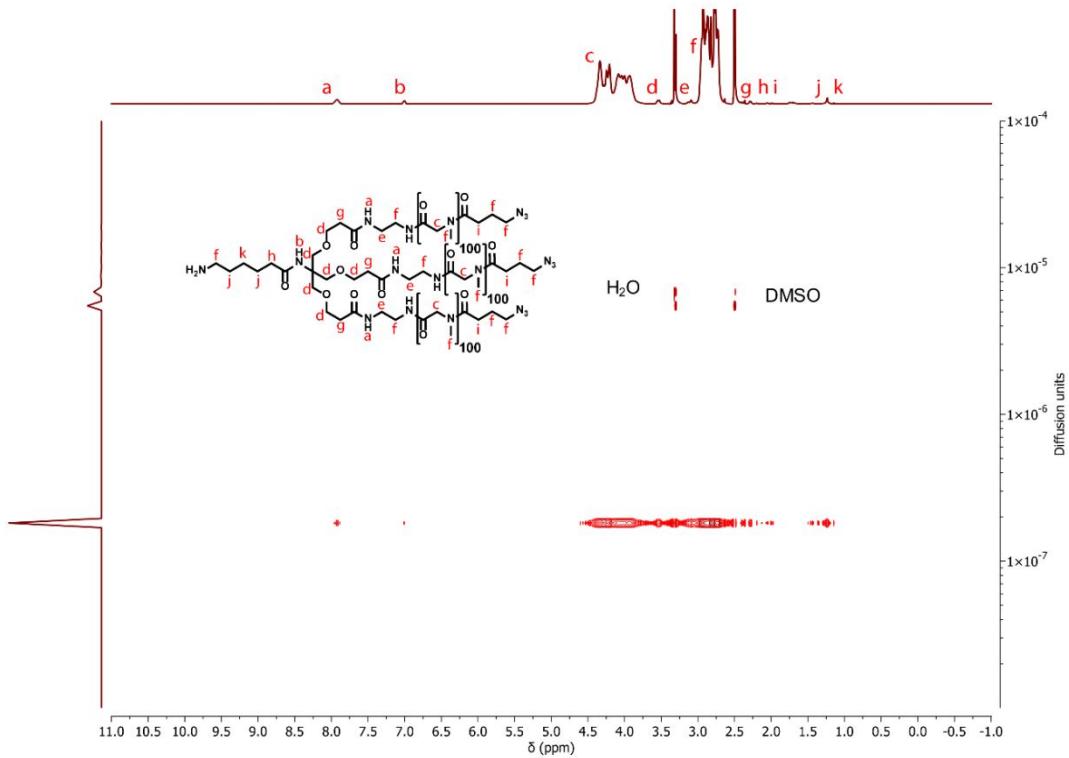


Figure S27. DOSY spectrum of i) deprotected three-arm macroinitiator in $\text{DMSO}-d_6$.

Cbz-Deprotection of the Six-Arm Macroinitiator ($NH_2-(pSar_{50}-N_3)_6$)

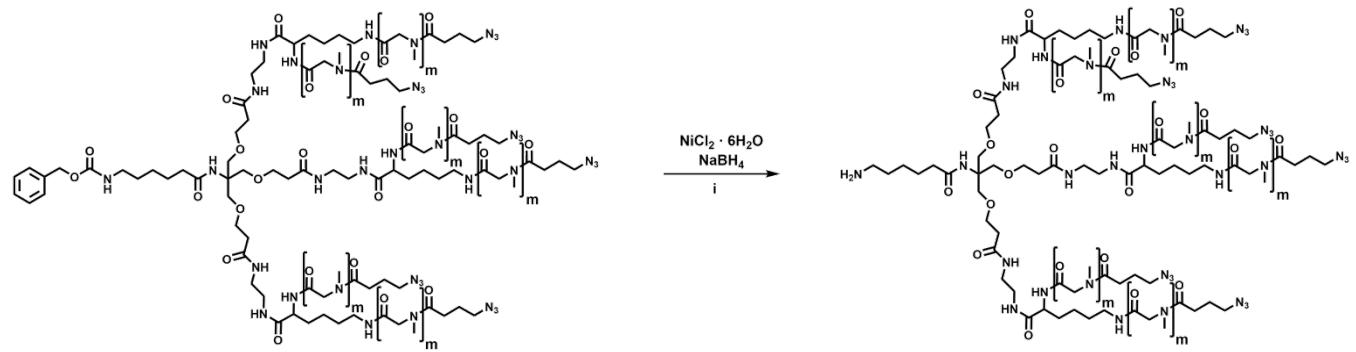


Figure S28. Cbz-deprotection of the six-arm macroinitiator i) MeOH, 0 °C, 1 h, rt, 48 h yield: 63%.

*Cbz-deprotection of six-arm macroinitiator Cbz-(*p*Sar₅₀-N₃)₆ (i)*

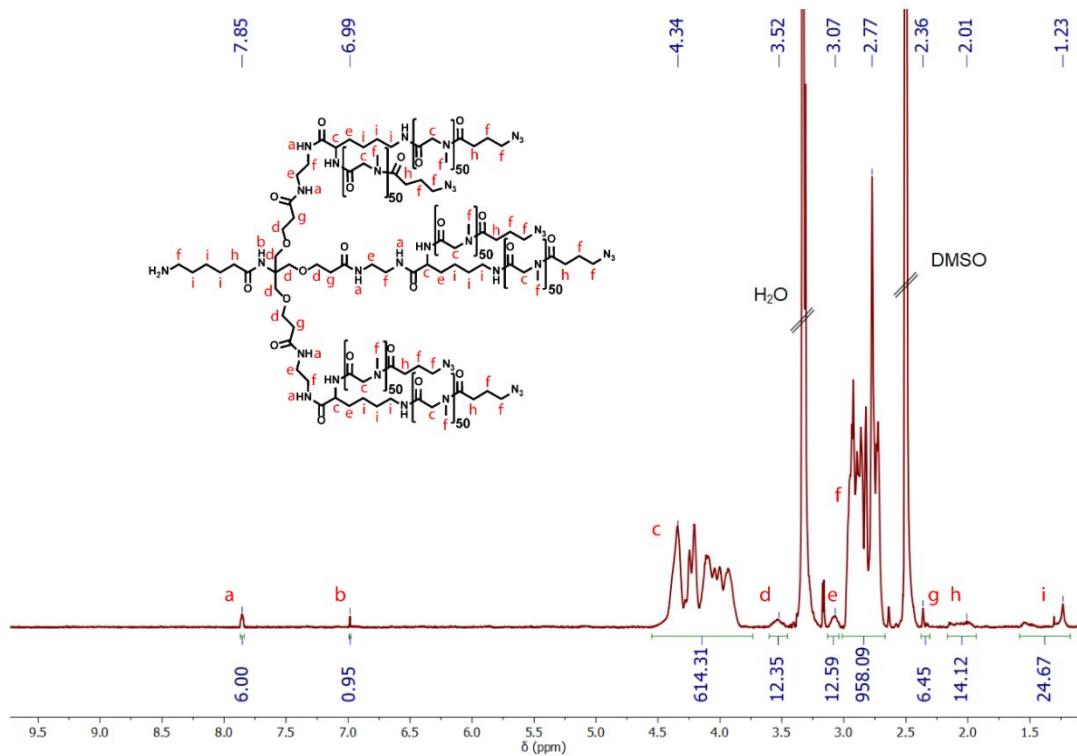


Figure S29. ¹H NMR spectrum of i) deprotected six-arm macroinitiator in DMSO-*d*₆.

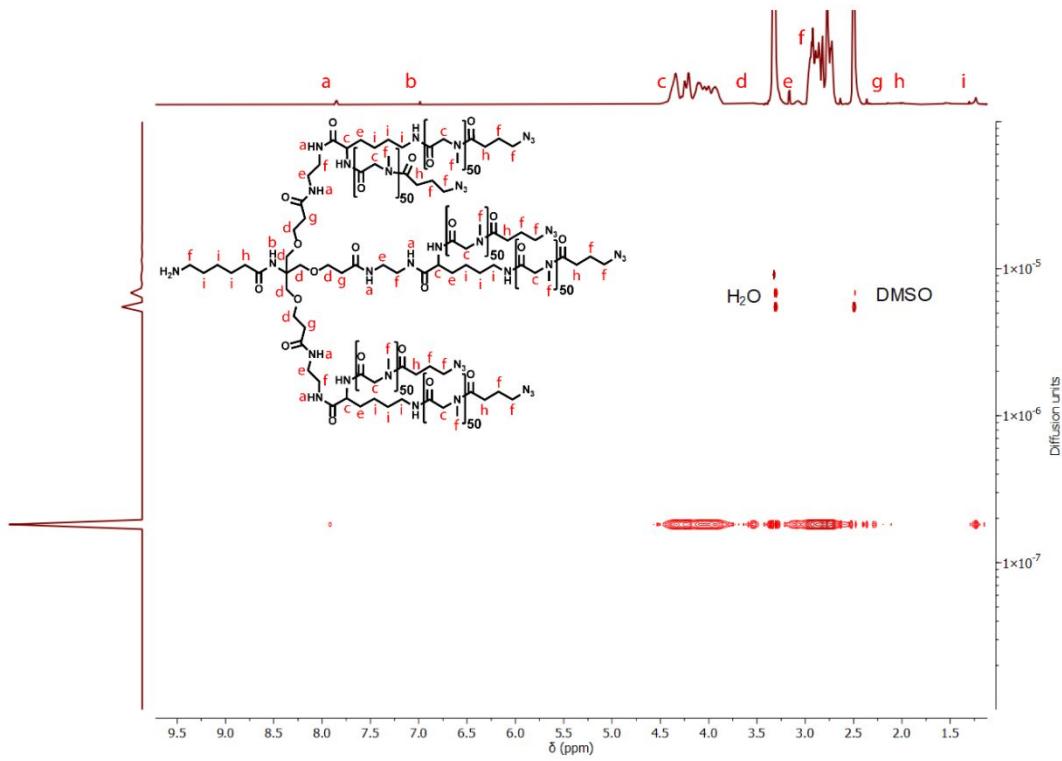


Figure S30. ^1H NMR spectrum of i) deprotected six-arm macroinitiator in $\text{DMSO}-d_6$.

Cross-linkable and Complexing Miktoarm Star Polymers (PeptoMiktoStars)

AB₃ PeptoMiktoStar (pLys(Boc)₂₀pHcy(SO₂Et)₂₀(pSar₁₀₀-N₃)₃)

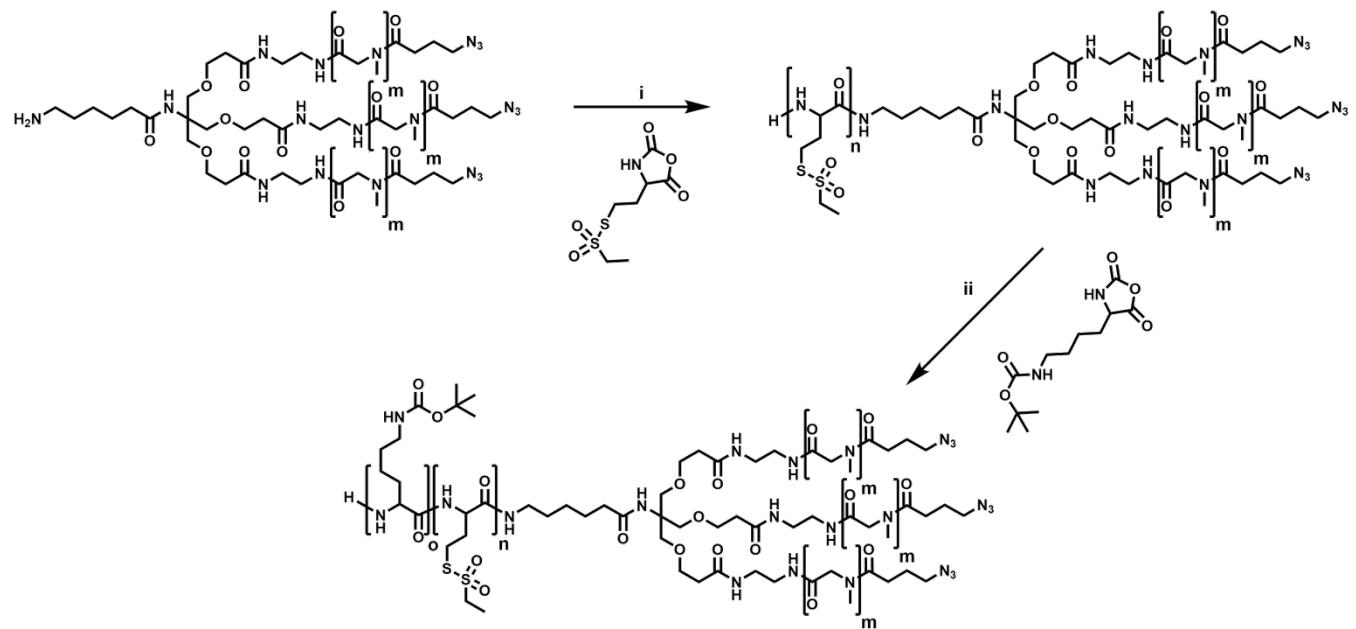


Figure S31. Introduction of both polypeptide blocks for the synthesis of AB₃ PeptoMiktoStars with m = 100, n = 20 and o = 20, i) DMF, -10 °C, 7 d, yield: 78%; ii) DMF, 0 °C, 3 d, yield: 80 %.

Introduction of the pHcy(SO₂Et)-block - (pHcy(SO₂Et)₂₀(pSar₁₀₀-N₃)₃ (i)

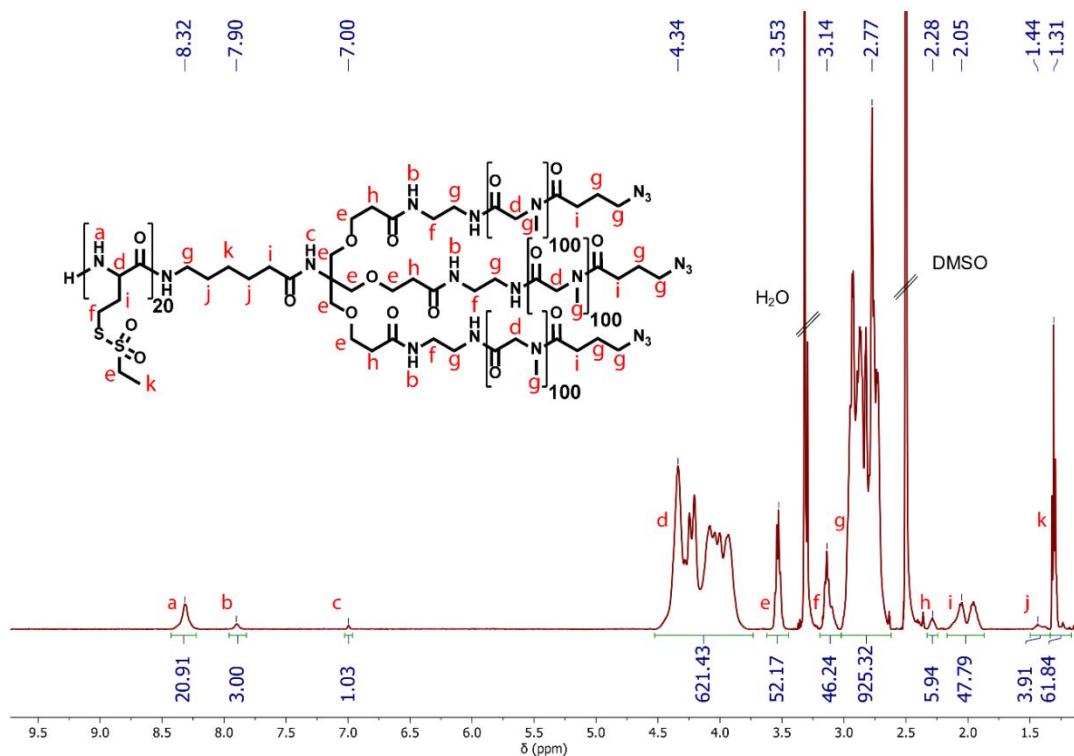


Figure S32. ¹H NMR spectrum of i) AB₃ PeptoMiktoStar pHcy(SO₂Et)₂₀(pSar₁₀₀-N₃)₃ in DMSO-*d*₆.

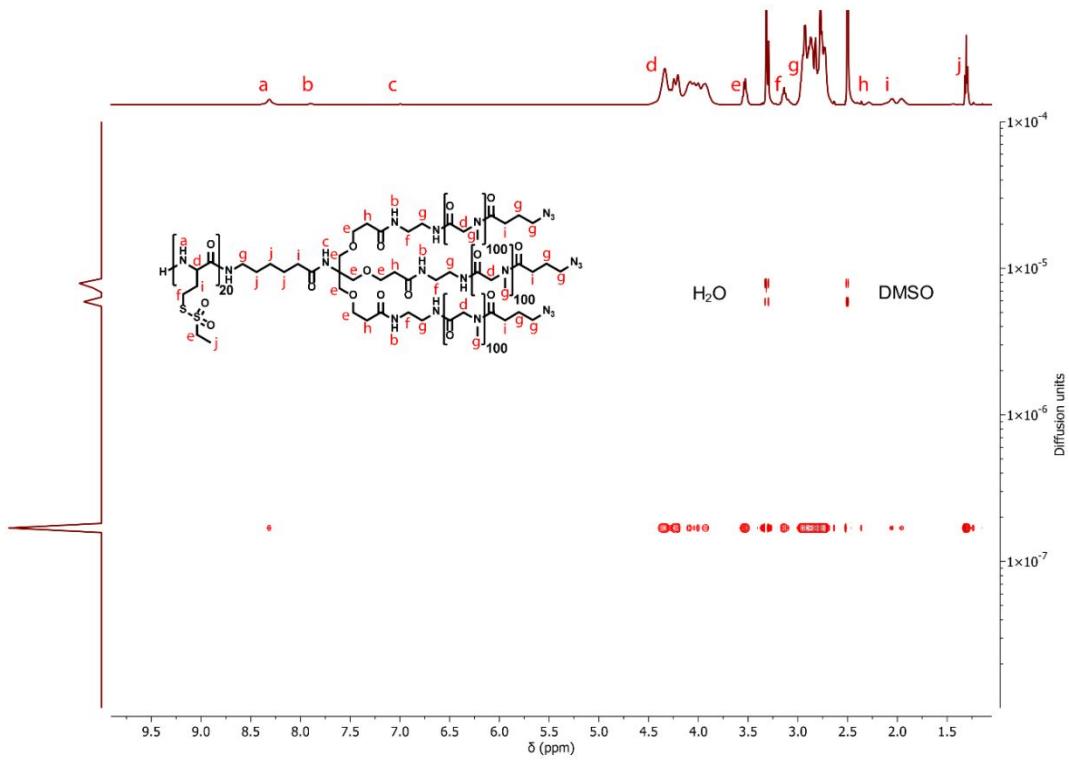


Figure S33. DOSY spectrum of i) AB_3 PeptoMiktoStar pHcy(SO_2Et)₂₀(pSar₁₀₀-N₃)₃ in DMSO-*d*₆.

Introduction of the pLys(Boc)-block - (pLys(Boc)₂₀pHcy(SO₂Et)₂₀(pSar₁₀₀-N₃)₃ (ii)

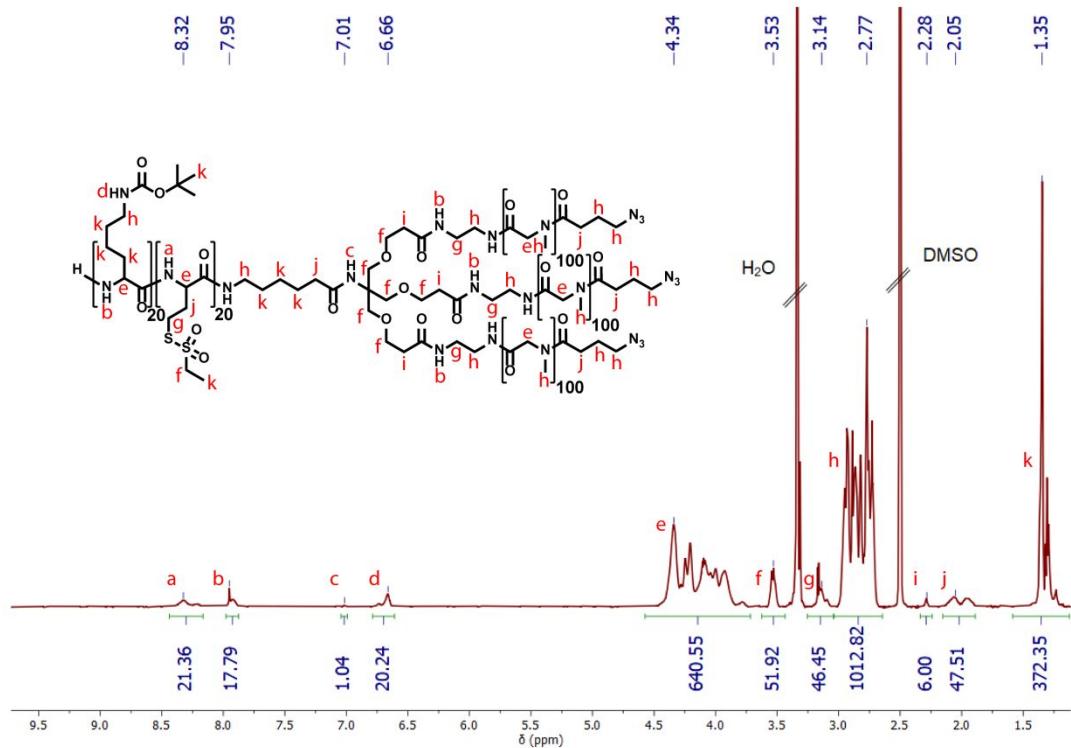


Figure S34. ¹H NMR spectrum of ii) AB₃ PeptoMiktoStar pLys(Boc)₂₀pHcy(SO₂Et)₂₀(pSar₁₀₀-N₃)₃ in DMSO-*d*₆.

Deprotected AB₃ PeptoMiktoStar (pLys₂₀pHcy(SO₂Et)₂₀(pSar₁₀₀-N₃)₃)

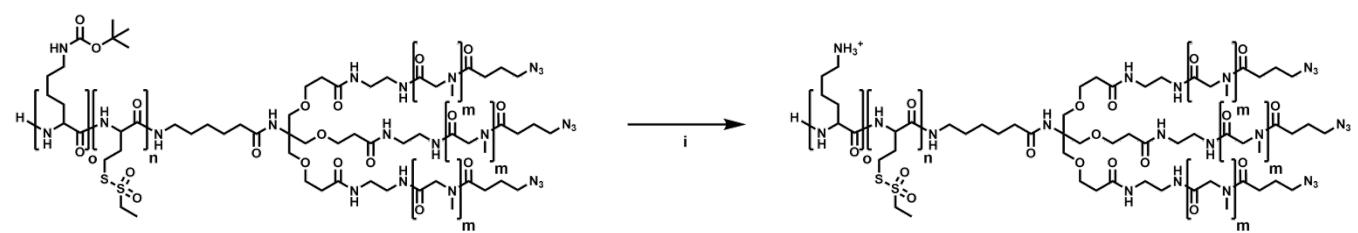


Figure S35. Boc-Deprotection of AB₃ PeptoMiktoStars with m = 100, n = 20 and o = 20 i)

TFA/H₂O (1/1), 2h, 0 °C, overnight, rt, yield: 85%.

*AB*₆ PeptoMiktoStar (*p*Lys(*Boc*)₂₀*p*Hcy(*SO*₂*Et*)₂₀(*p*Sar₅₀-N₃)₆)

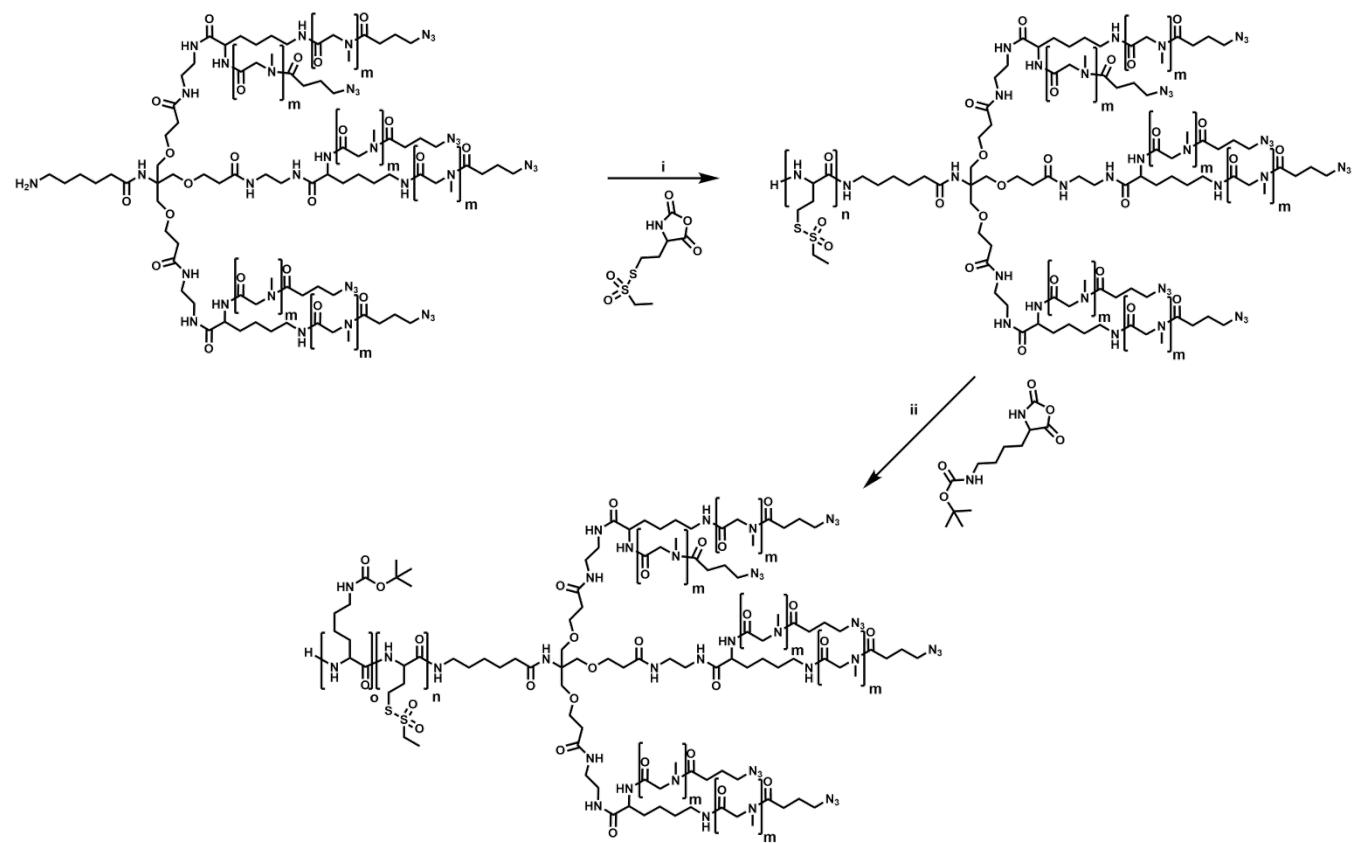


Figure S36. Introduction of both polypeptide blocks for the synthesis of *AB*₆ PeptoMiktoStars with m = 50, n = 20 and o = 20, i) DMF, -10 °C, 7 d, yield: 67%; ii) DMF, 0 °C, 3 d, yield: 72%.

Introduction of the pHcy(SO₂Et)-block - (pHcy(SO₂Et)₂₀(pSar₅₀-N₃)₆ (i)

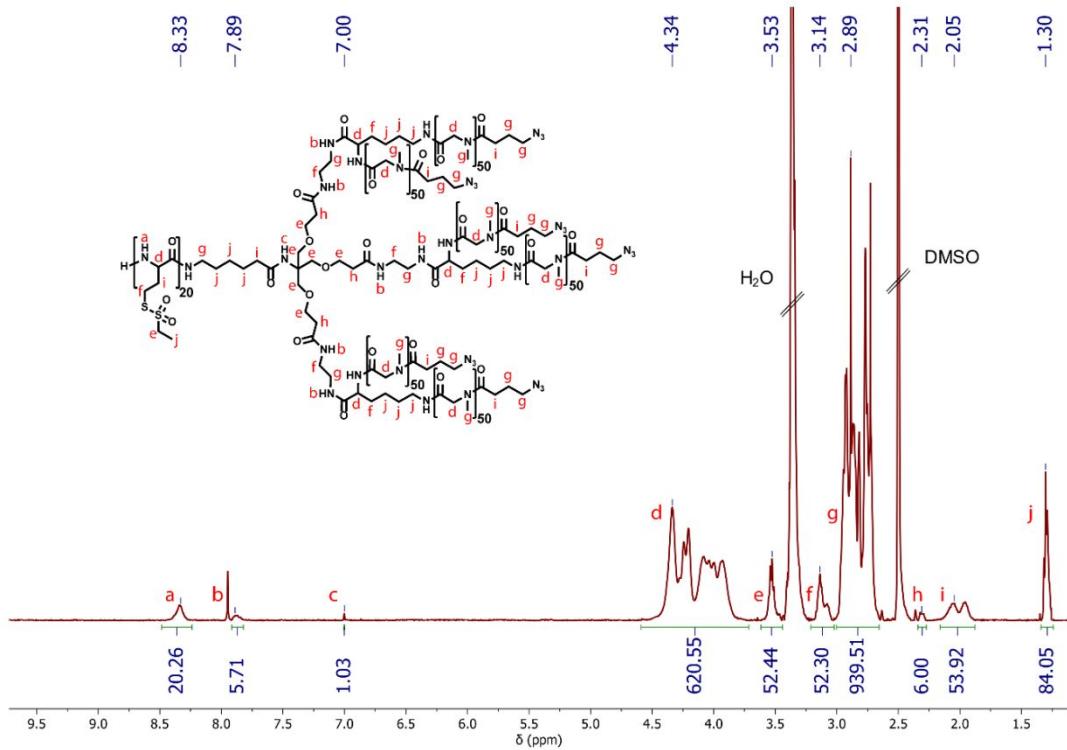


Figure S37. ¹H NMR spectrum of i) AB₆ PeptoMiktoStar pHcy(SO₂Et)₂₀(pSar₅₀-N₃)₆ in DMSO-*d*₆.

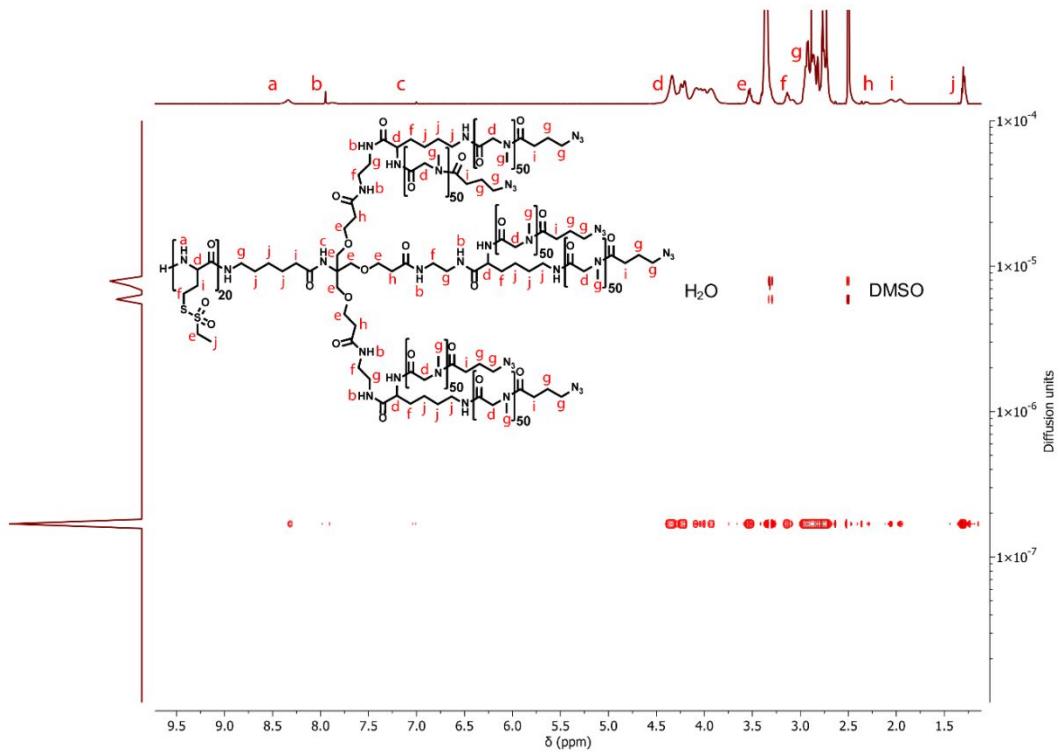


Figure S38. DOSY spectrum of i) AB₆ PeptoMiktoStar pHcy(SO₂Et)₂₀(pSar₅₀-N₃)₆ in DMSO-*d*₆.

Introduction of the pLys(Boc)-block - (pLys(Boc)₂₀pHcy(SO₂Et)₂₀(pSar₅₀-N₃)₆ (ii)

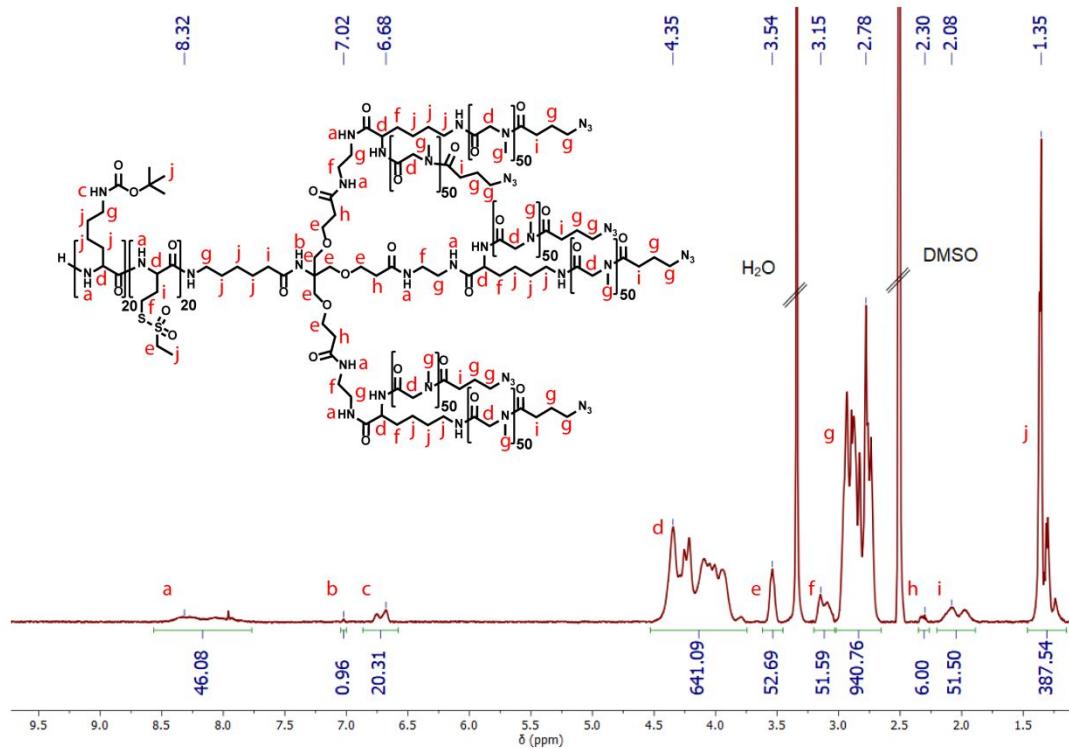


Figure S39. ¹H NMR spectrum of ii) AB₆ PeptoMiktoStar pLys(Boc)₂₀pHcy(SO₂Et)₂₀(pSar₅₀-N₃)₆ in DMSO-*d*₆.

Deprotected AB₆ PeptoMiktoStar (pLys₂₀pHcy(SO₂Et)₂₀(pSar₅₀-N₃)₆)

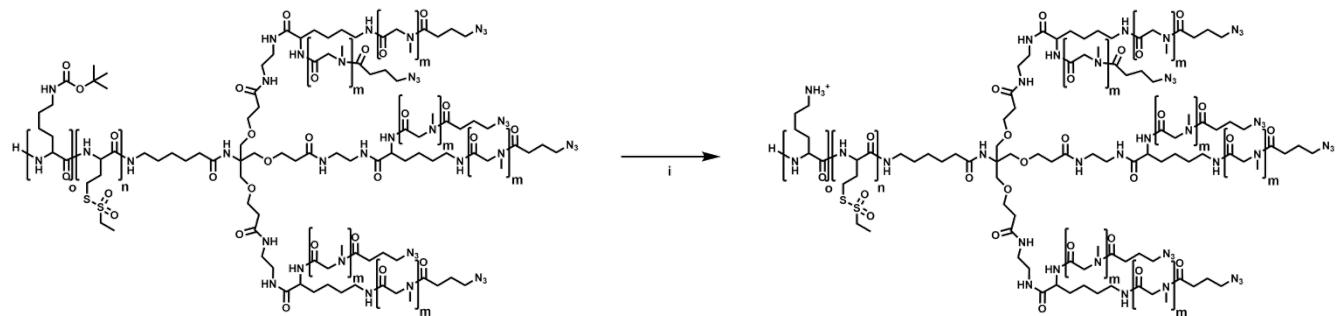


Figure S40. Boc-Deprotection of AB_6 PeptoMiktoStars with $m = 50$, $n = 20$ and $o = 20$ i)

TFA/H₂O (1/1), 2h, 0 °C, overnight, rt, yield: 77 %.

Further Reagents

Pentafluorophenyl-4-azidobutanoat

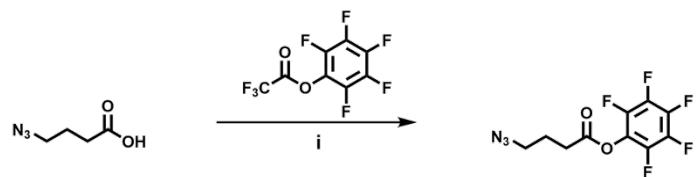


Figure S41. Synthesis of the azide functionalization agent pentafluorophenyl-4-azidobutanoate.(i)

TEA, THF, rt, 1d, yield: 83%.

Synthesis of Pentafluorophenyl-4-azidobutanoat (i)

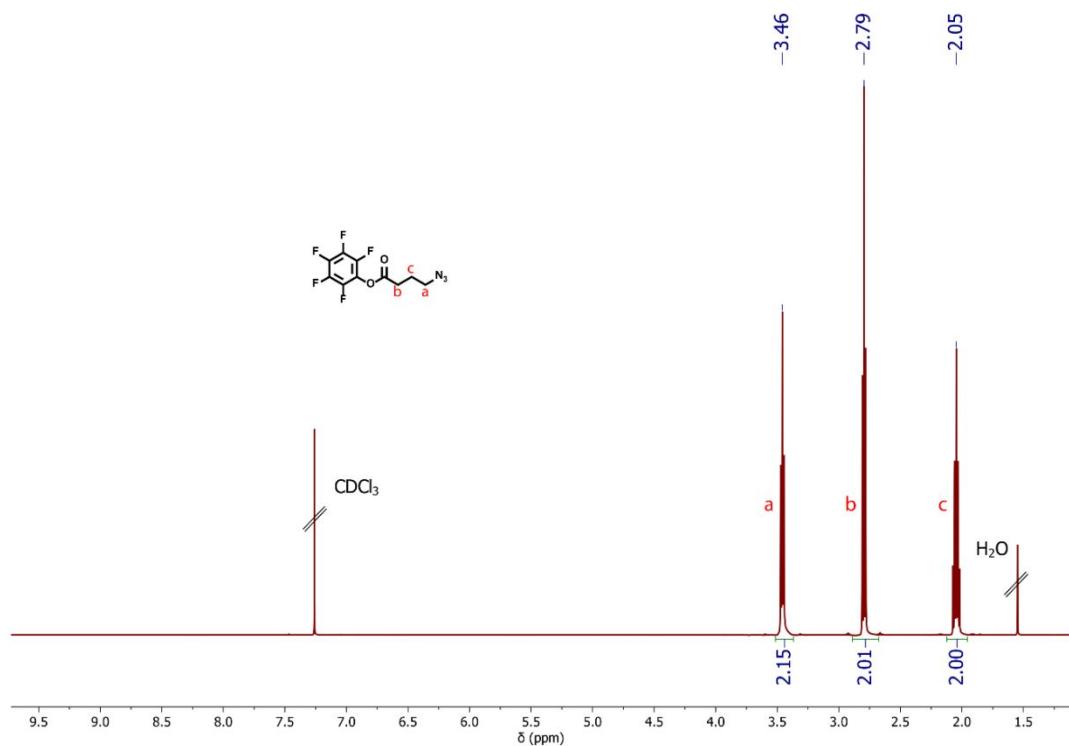


Figure S42. ^1H NMR spectrum of pentafluorophenyl-4-azidobutanoate in CDCl_3 .

Biological test

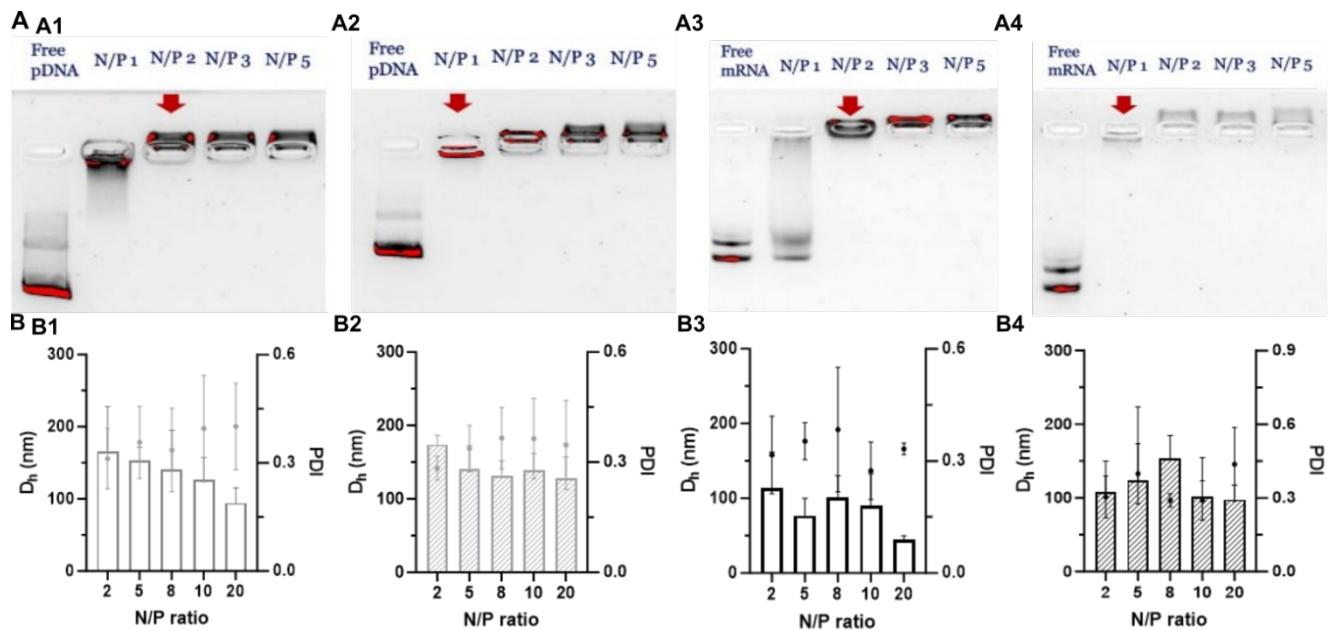


Figure S43. Characterization of pDNA/ABn and mRNA/ABn PICMs. (A) Complexation ability of (A1, A2) pDNA and (A3, A4) mRNA by (A1, A3) AB₃ or (A2, A4) AB₆ triblock copolymer using agarose gel electrophoresis. Lane 1, naked/free (A1, A2) pDNA or (A3, A4) mRNA, lane 2-5, (A1, A2) pDNA/ABn PICMs at N/P 1, 2, 3 and 5, (A3, A4) mRNA/ABn PICMs at N/P 1, 2, 3 and 5. (B) Size and size distribution of (B1, B2) pDNA/ABn PICMs and (B3, B4) mRNA/ABn PICMs complexed by triblock copolymer of (B1, B3) AB₃ or (B2, B4) AB₆ in HEPES buffer (10 mM, pH7.4) measured by DLS. All data was averaged from three independent experiments ($n \geq 9$).

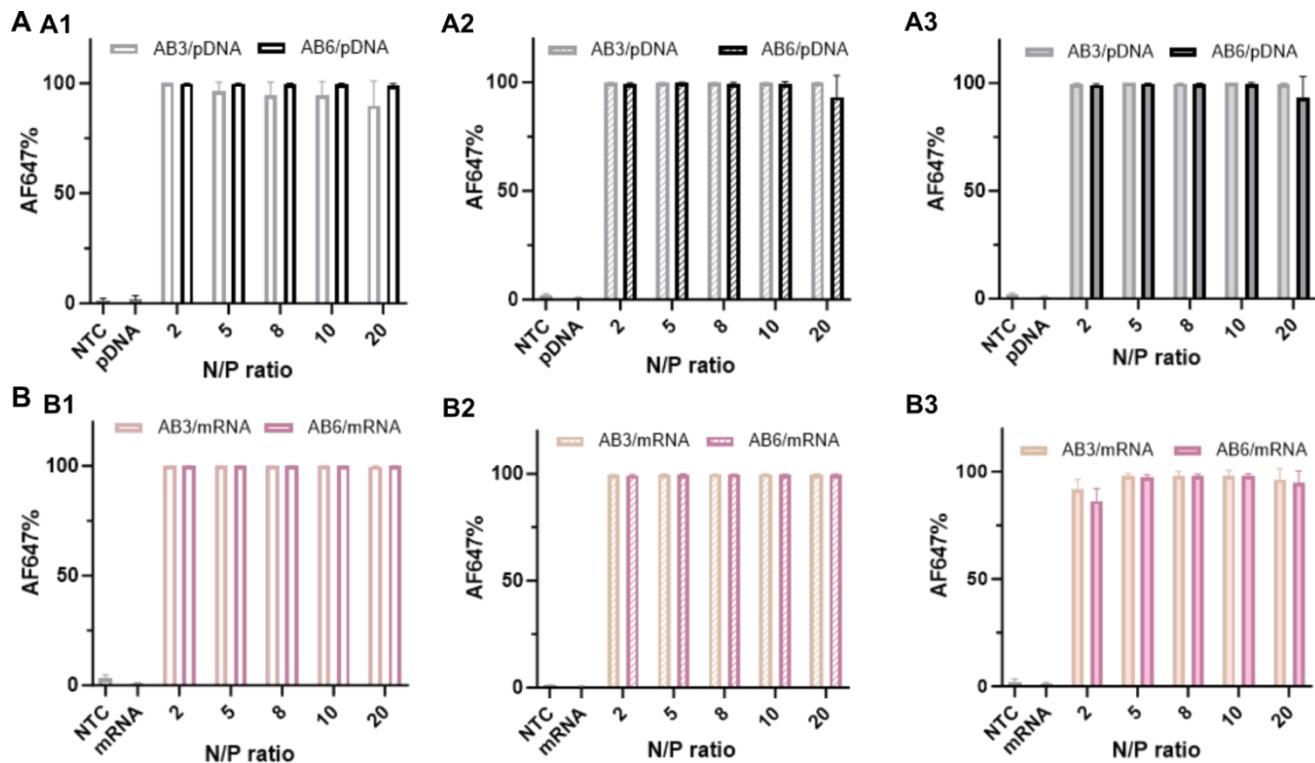


Figure S44. *In vitro* evaluation of (A) pDNA/AB_n PICMs and (B) mRNA/AB_n PICMs on (A1, B1) D1 cells and (A2, B2, A3, B3) Jurkat T cells. (A1) D1 cells and (A2) Jurkat T cells were exposed to pDNA/AB_n PICMs (final pDNA concentration of 0.2 µg/well) in fresh culture medium for 48 h. (A3) pDNA/AB_n PICMs were incubated with Jurkat T cells in Opti-MEM for 48 h. At the end of incubation, cells were collected for flow cytometry measurements. (B1) D1 cells and (B2) Jurkat T cells were exposed to mRNA/AB_n PICMs in fresh culture medium for 24 h. (B3) mRNA/AB_n PICMs were incubated with Jurkat T cells in Opti-MEM for 24 h. At the end of incubation, cells were collected for flow cytometry measurements. In each well, the final pDNA or mRNA concentration is 0.2 µg/well. All data was averaged from three independent experiments ($n \geq 9$).

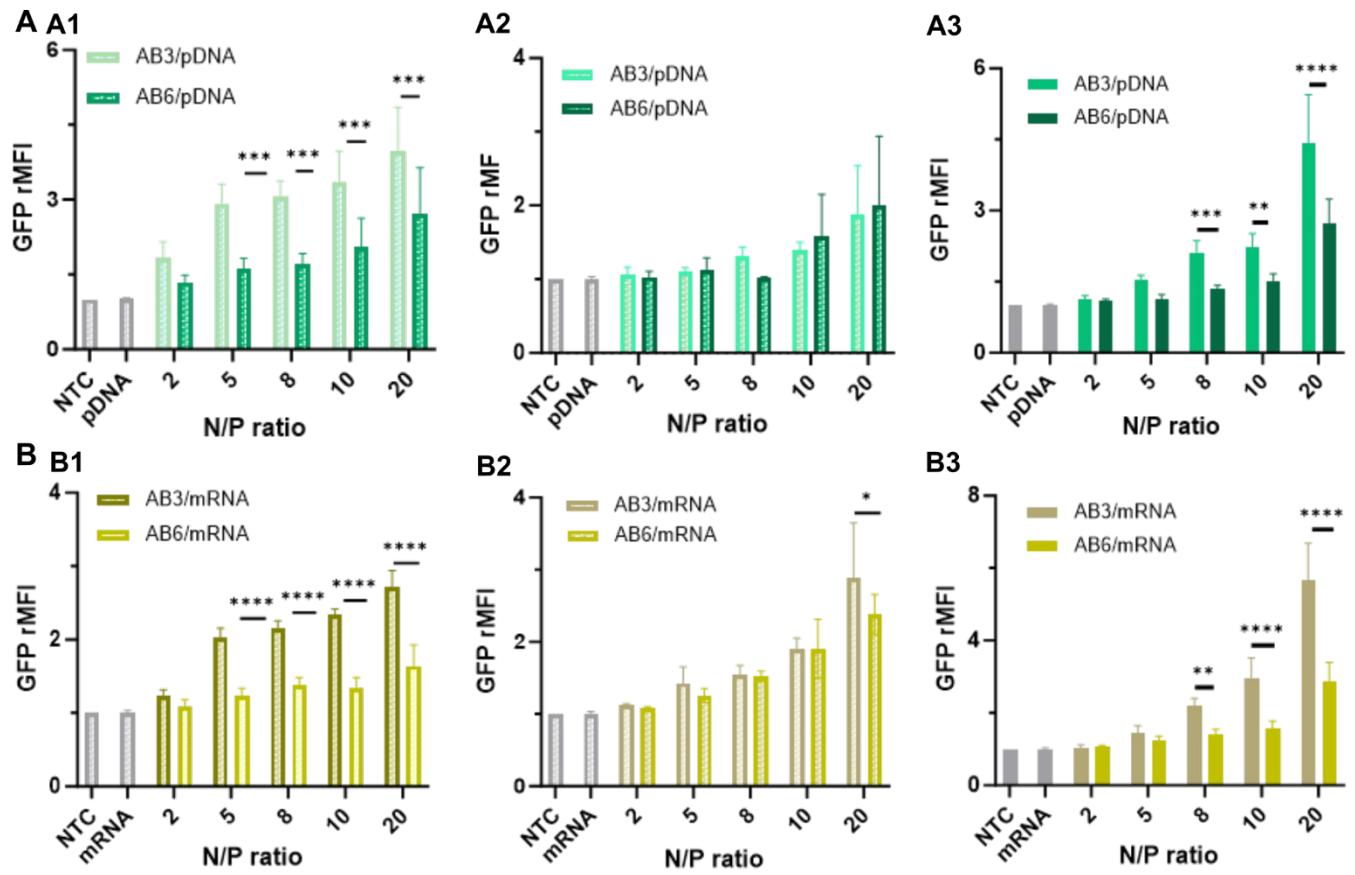


Figure S45. *In vitro* evaluation of (A) pDNA/AB_n PICMs and (B) mRNA/AB_n PICMs on (A1, B1) D1 cells and (A2, B2, A3, B3) Jurkat T cells. (A1) D1 cells and (A2) Jurkat T cells were exposed to pDNA/AB_n PICMs (final pDNA concentration of 0.2 µg/well) in fresh culture medium for 48 h. (A3) pDNA/ABn PICMs were incubated with Jurkat T cells in Opti-MEM for 48 h. At the end of incubation, cells were collected for flow cytometry measurements. (B1) D1 cells and (B2) Jurkat T cells were exposed to mRNA/AB_n PICMs in fresh culture medium for 24 h. (B3) mRNA/AB_n PICMs were incubated with Jurkat T cells in Opti-MEM for 24 h. At the end of incubation, cells were collected for flow cytometry measurements. In each well, the final pDNA or mRNA concentration is 0.2 µg/well. All data was averaged from three independent experiments ($n \geq 9$).

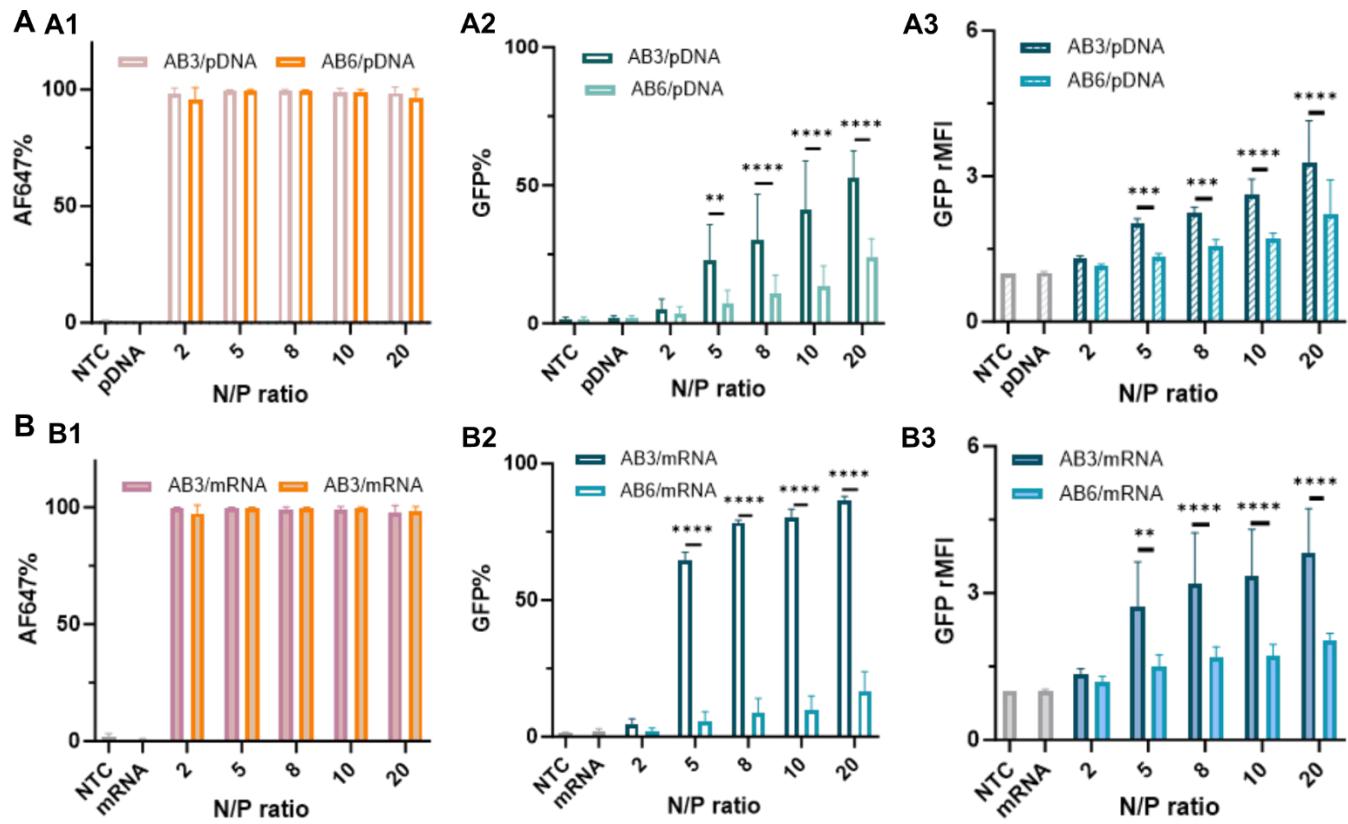


Figure S46. *In vitro* evaluation of (A) pDNA/AB_n PICMs and (B) mRNA/AB_n PICMs on D1 cells. (A) D1 cells cultured in Opti-MEM were exposed to pDNA/AB_n PICMs (A1) for 24 h to examine cellular uptake and (A2, A3) for 48 h to examine transfection efficiency (final pDNA concentration of 0.2 µg/well). (B) D1 cells cultured in Opti-MEM were exposed to mRNA/AB_n PICMs for 24 h (B1) for cellular uptake and (B2, B3) for transfection efficiency (final mRNA concentration of 0.2 µg/well). At the end of incubation, cells were collected for flow cytometry measurements. All data was averaged from three independent experiments ($n \geq 9$).

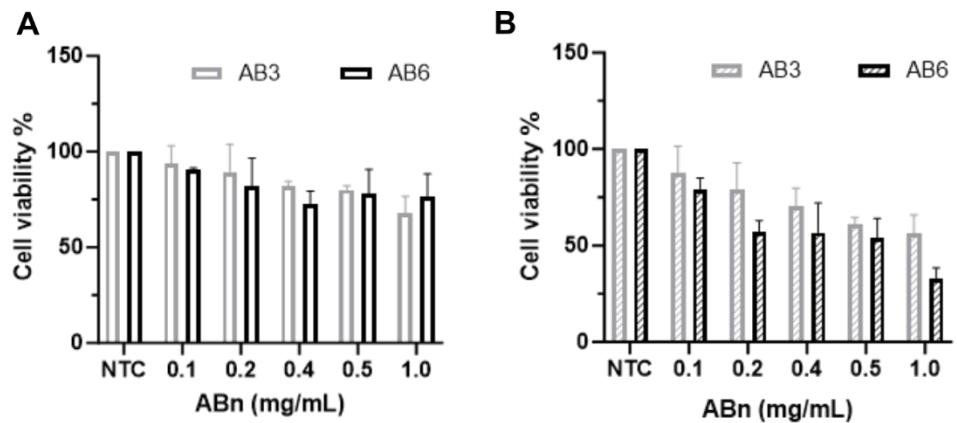


Figure S47. Cell viability of (A) D1 cell and (B) Jurkat T cell upon 24 hours' incubation with blank AB3 and blank AB6 at a final concentration of 0.1, 0.2, 0.4, 0.5 and 1.0 mg/mL. All data was averaged from three independent experiments ($n \geq 9$).