

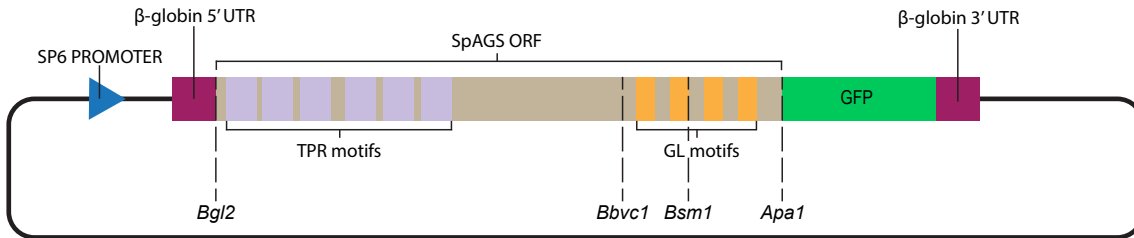
Emura et al.,

“The evolutionary modifications of a GoLoco motif in the AGS protein facilitate micromere formation in the sea urchin embryo”

Supplementary Figures and Legends

Supplemental Figure S1

A



B

S. purpuratus AGS Protein Sequence

TPR1 TPR2
 MQSEASCMELALEGERLCKAGNCREGVRYLEAAVEVGTDDLKLSAIYSQLGNAYFYLEDYGKALTYHKHDL

TPR3 117aa, before here removed in 1F
 TLATSIGDRLGEAKASGNIGNTLKVLGKFDEAICCCQRHLDISRELSEKVGEGRALYNLGNVYHAKGKHAGRS

TPR4
 GHQDPGDFPQEVTDCLKQAIQFYEANLAIVRELGDRAAQGRACGNLGNTHYLLGNFETAIQFHTERLAIKE

222aa, before here removed in 2F TPR5 TPR6
 FGDKA AERRAYSNLGNACVFMVQFEVAAKYKSLHIARQLGELAMEAQACYSLGNTYLLREYKAVEYHS

301aa, before here removed in 3F
 RHMEIAQQLNDRVGEGRACWSLGNHAHTSLGNHEKALHYATLHLQISREVDRTGEVTAKMNLQDLQTLFG

ISTADLSEVSTTVQTPLQESKGARPRRRSMENLELVAMTPEKTEIVQVPRKIRPGSKLKLKNGGESKDKQS

BbvC1 RE site GL1 (removed in Δ1)
 HKDSKESVSSAKNSVPAPSAKASQTSARKGGEPQMLDDADNFFEALSRFQSNRMDEQRCSFGRLQKKLAD

GL2 (removed in Δ2) Bsm1 RE site GL3 (removed in Δ3)
 EEGNGLPEKEELLDEIAKLQGSRLNEQRAF SVKRLPGLPLRANEDVVGKLLAKGERAEPD DDFEMIIIRCQG

GL4 (removed in Δ4)
 ARIEDQRSTLPIQAPAPTVPDEDFFSLIQRIQSKRIEEQRSIAPWEKSGTSCVCFYDYDTSRTEGTACCVYSRP

659aa
 VQESHL

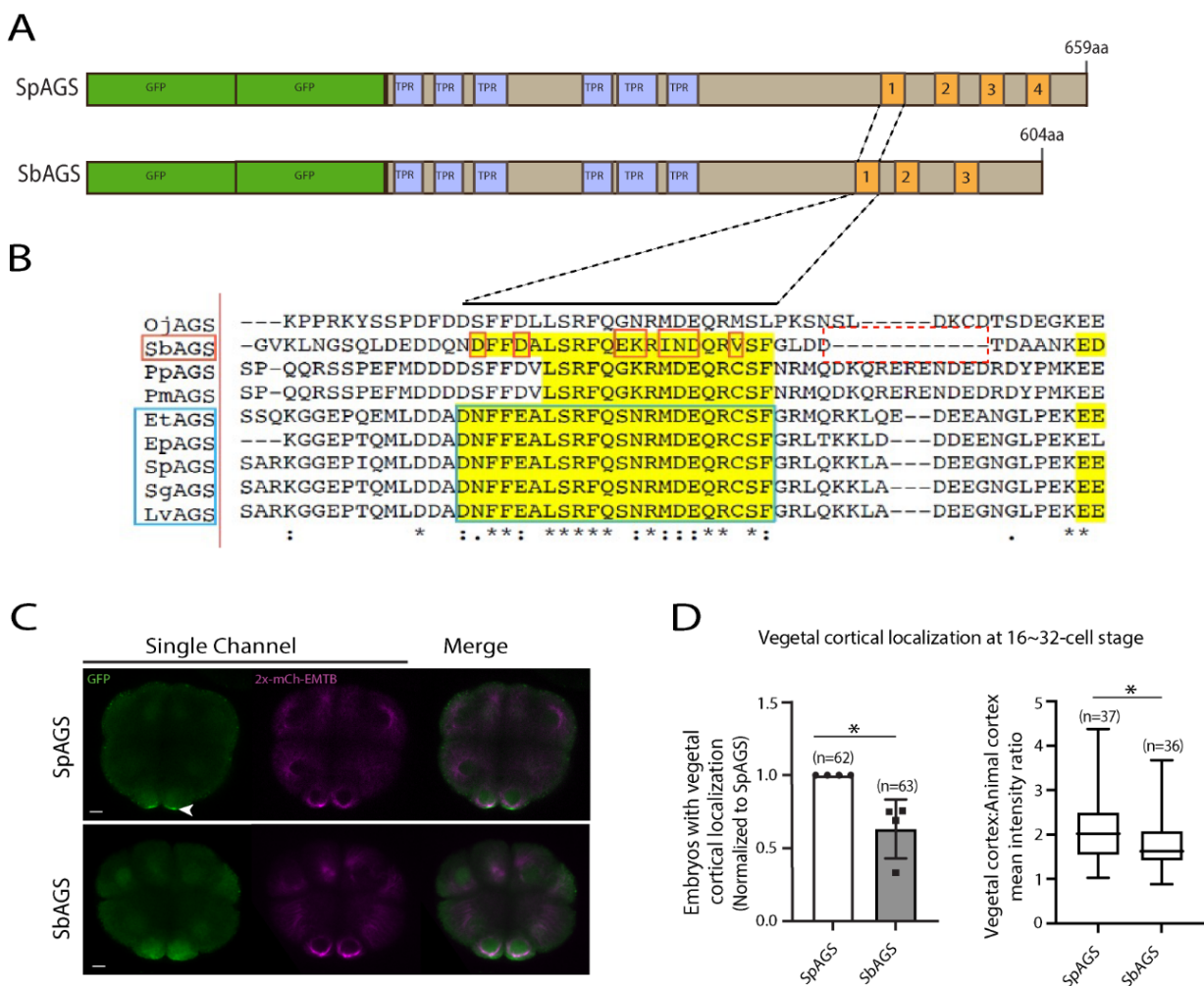
Supplemental Figure 1. Dissection of SpAGS motifs. **A**, The construct design of SpAGS-GFP, including the restriction enzyme sites used to prepare SpAGS mutants. **B**, The protein sequence of SpAGS. Predicted domains are labeled based on NCBI blast results, and the sequence portions deleted for each N-terminal construct are marked. The sequences for each GL motif used for deletion or swapping are indicated in orange. The internal restriction enzyme sites for *BbvCI* and *BsmI* are shown in green.

Supplemental Figure 2 Echinoderm AGS sequence alignment



Supplemental Figure 2. Echinoderm AGS sequence alignment. All AGS are similar in the N-terminus with the predicted conserved TPR motifs (blue) but are highly variable in the C-terminus with the predicted GL motifs (yellow).

Supplemental Figure 3



Supplemental Figure 3. SbAGS does not fully localize at the vegetal cortex. **A**, Design of 2XGFP-AGS constructs that contain AGS orthologs from two different species tested in this study, namely, *S. purpuratus* (Sp; sea urchin), and *S. briareus* (Sb; sea cucumber). TPR motifs are marked in blue, and GL motifs are in orange. **B**, Alignment of GL1 motif sequences among echinoderms. **C**, Single Z-slice confocal images of sea urchin (Sp) or sea cucumber (Sb) embryos at 16-cell stage showing localization of 2x-GFP-AGS. Embryos were injected with 0.2-0.3 μ g/ μ l stock of GFP-AGS mRNA and 0.25 μ g/ μ l stock of 2x-mCherry-EMTB mRNA. The white arrowhead indicates vegetal cortical localization of GFP-AGS. **D**, Left graph, the number of embryos with vegetal cortical localization of 2x-GFP-AGS in 16~32-cell embryos was scored and normalized to that of the control group (SpAGS). Right graph, the ratio of the vegetal cortex-to-animal cortex mean intensity. Statistical analysis was performed against the control (SpAGS) by *t*-test. *n* indicates the total number of embryos scored. * represents *p*-value < 0.05. Each experiment was performed at least two independent times. Error bars represent standard error. Scale bars=10 μ m.

Supplemental Figure 4 SpAGS, EtAGS and PmAGS linker domain sequence alignment

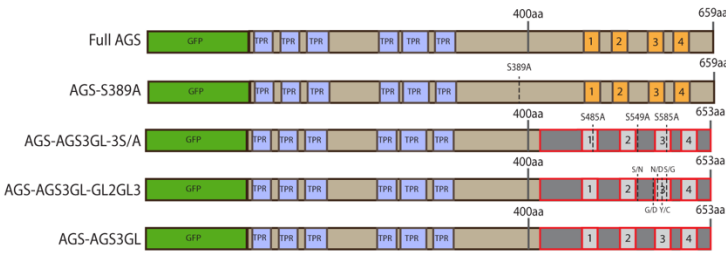
A

Yellow, CK1 ; Blue, Aurora ; Green, CMGC/CDK

PmAGS GDKTGE^YLTQMNLADLR^YTVLGLDKDPC^YSDGLLQNA^YSK^YTKLKEKELN^YR^YQR^YRS^YME^YRLELV
 SpAGS GDRTGE^YVTA^YMN^YLQDL^YQL^YTLFGI^YSTADL^YSEV^YST^Y--V^YQ^YPL^YQE^YSK^YGAR^YPR^YRR^YRS^YME^YNLELV
 EtAGS GDRTGE^YVTA^YMN^YLADL^YKTL^YFGI^YTGDL^YSHD^YV^YTAA^Y--CPP^YAM^YQ^YPK^YGAT^YPR^YRR^YRS^YME^YNLELV
 :*:***:*** **:***:*** * **:*:***:***

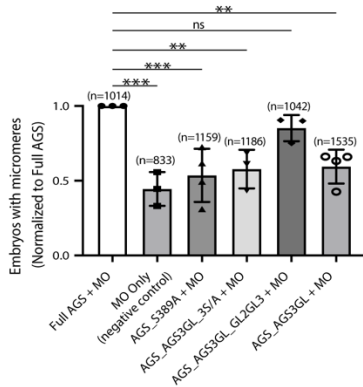
PmAGS **TM**TPDKNAAATANGKPKQSAGAKFKRK--GSNKEKLSRKSSNTSS^YSSSSGESAACLRT
 SpAGS **AM**TP^YE^YKT^YEIV^YQ^YVP^YK^YR^YK^YIRP^Y--GSKL^YKL^YK^YNG^YGES^YK^YDK^YQ^YSH^YK^YDS^YKE^YSV^YSS^YAK^YNS^YVP^YAP^YSK^YAK
 EtAGS **AM**TP^YE^YKN^YT^YDS^YAV^Y--PNG^YKL^YK^YPT^YG^YSK^YK^YRR^YNG^YKE^Y---KLS^YR^YK^YG^YNT^YST^YG^YSS^YNS^YSS^YSS^YAP^YS^YQ^YAS
 :***:***: . : *:* * : . *:*: * * * : . * . .

B



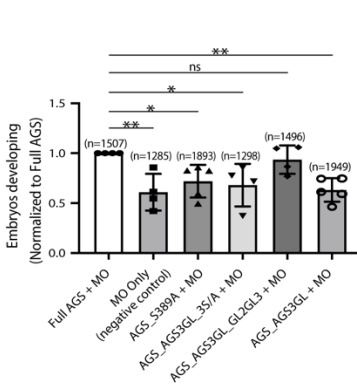
E

Micromere formation at 16~32-cell stage

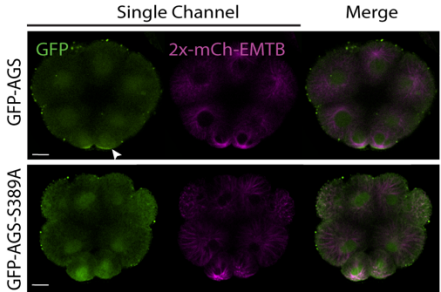


F

Development at 2 dpf

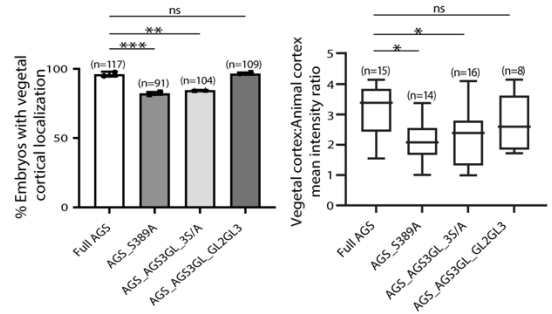


C

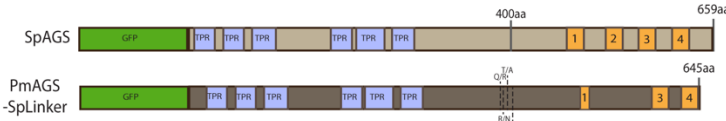


D

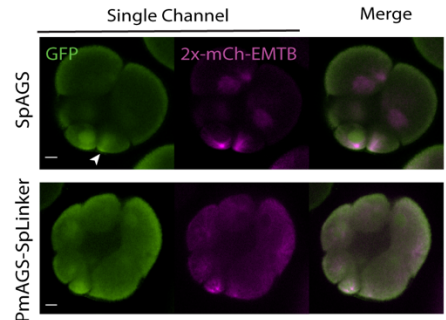
Vegetal cortical localization at 16~32-cell stage



G

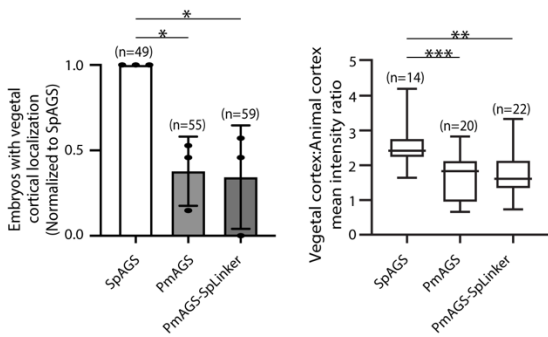


H



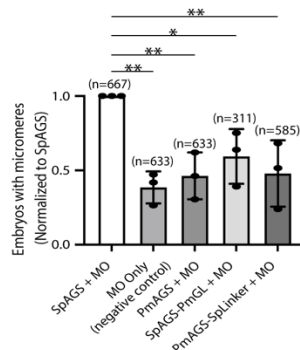
I

Vegetal cortical localization at 16~32-cell stage



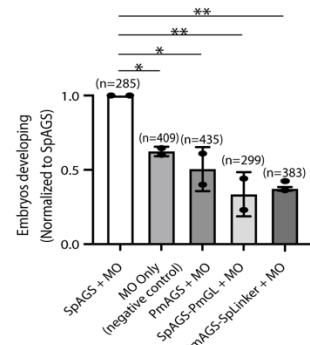
J

Micromere formation at 16~32-cell stage



K

Development at 2 dpf



Supplemental Figure 4. The linker domain and GL2-GL3 regions are important for AGS localization and function. **A**, Alignment of linker domain between echinoderms including sea urchin (SpAGS), pencil urchin (EtAGS), and sea star (PmAGS). Bold letters represent the conserved core linker domain. The yellow, blue, or green highlights indicate the CK1, Aurora, or CMGC/CDK phosphorylation sites predicted by GPS 6.0. The red letters indicate PmAGS amino acids mutated to those of SpAGS to construct the PmAGS-SpLinker mutant. **B**, Design of GFP-AGS mutant constructs tested in this study. TPR motifs are marked in blue, and GL motifs are in orange. The brown section indicates the SpAGS sequence, and the red and grey boxes show the non-sea urchin (non-SpAGS) introduced at the C-terminus. The dotted lines represent single amino acid mutations. **C**, Single z-slice confocal images of sea urchin embryos at 8~16-cell stage showing localization of GFP-AGS-S389A mutant. Embryos were injected with 0.3 μ g/ μ l stock of GFP-AGS mRNA and 0.25 μ g/ μ l stock of 2x-mCherry-EMTB mRNA. The white arrowhead indicates vegetal cortical localization of GFP-AGS. **D**, % of the embryos with vegetal cortical localization of GFP-AGS mutants (left) and the ratio of the vegetal cortex-to-animal cortex mean intensity (right) in 16~32-cell embryos. Statistical analysis was performed against Full AGS by One-Way ANOVA. **E-F**, Embryos were injected with 0.15 μ g/ μ L stock of GFP-AGS mRNAs and 0.75mM SpAGS MO. The number of embryos forming micromeres (E) and developing to gastrula or pluteus stage (F) were scored, and each of which was then normalized to that of the Full AGS. Statistical analysis was performed by One-Way ANOVA. **G**, Design of GFP-AGS constructs tested in this study from *S. purpuratus* (Sp) and *P. miniata* (Pm). TPR motifs are marked in blue, and GL motifs are in orange. The dotted lines represent single amino acid mutations. **H**, Single Z-slice confocal images of sea urchin embryos at 8~16-cell stage showing localization of GFP-AGS. Embryos were injected with 0.3 μ g/ μ l stock of GFP-AGS mRNAs and 0.25 μ g/ μ l stock of 2x-mCherry-EMTB mRNA. The white arrowhead indicates vegetal cortical localization of GFP-AGS. **I**, The number of embryos with vegetal cortical localization of GFP-AGS mutants in 16~32-cell embryos was scored and normalized to that of the SpAGS (left graph). Right graph shows the ratio of the vegetal cortex-to-animal cortex mean intensity. Statistical analysis was performed against SpAGS by One-Way ANOVA. **J-K**, Embryos were injected with 0.3 μ g/ μ l stock of GFP-AGS mRNAs and 0.75mM SpAGS MO. The number of embryos making micromeres (J) and developing to gastrula or pluteus stage (K) were scored and normalized to that of the SpAGS. Statistical analysis was performed by One-Way ANOVA. n indicates the total number of embryos scored. * represents p-value<0.05, ** p-value < 0.01, and *** p-value < 0.001. Each experiment was performed at least two independent times. Error bars represent standard error. Scale bars=10 μ m.

Supplemental Figure S5 GoLoco Domain sequence alignment

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SpAGS_GL 461 RKGGEPIQMLDDADNFFEALSRFQSNRMDEQRCSEFGRLQKKLA-----
DmPins_GL 455 EKLRQAQATRKAKDDFFEMLSRFSQSRMDDQRCSEIKVNPAGAPAVATGATRKPL----
PmAGS_GL 479 PQQRSSEPEFMDDDDSFFDVLSRFQGKRMDEQRCSEFNRMQDKQRERE-----
EtAGS_GL 459 QKGGEPQEMLLDADNFFEALSRFQSNRMDEQRCSEFGRMQRKLE-----
HsLGN_GL 468 NSQRKISADTIGDEGFFDLLSRFQSNRMDDQRCCLQEKNCHTASTTTSTPPKMLKT
HsAGS3_GL 460 PRTSIPRAPSSDEECFFDLLTKFQSSRMDDQRCPLDDGQAGAAEATAAPTLEDRI--A
      : **: **: *..***:*** :

SpAGS_GL DEENGLPEKELLDEIAKLQSRLNEQRAFSVKRPLPGLPGLR-ANE---DVVGK---LL
DmPins_GL -----VQQNSLFVDPTNLPGLKSPSSANPSAIGHGPLAR
PmAGS_GL NDEDRDYPMKEELLNQIASFQSRLNEQRSS----ICNLPGLKTNNE---EVLGQ---LL
EtAGS_GL DEEANLPEKELLDEIAKLQSRLNEQRAFSQRLPGLPGLR-ANE---NVLGK---LL
HsLGN_GL SSVPVVSPNTDEFFLDLLASSQSRLLDDQRAS----FSNLPGLRLTQNSQ-SVLS----HL
HsAGS3_GL QPSMTASPQTEEFFDLLIASSQSRLLDDQRAS----VGLPGLRITHSNA-GHLR---G-
      *: .****: . :

SpAGS_GL AKGERAEPDDFFEMIRCOGARIEDQRSTLP-I-----
DmPins_GL SATTTQPDDFFLDMLMRCQGSRLEEQRSELPRPNVTMDAEAEAPPRSVPEAAVPGAPRG
PmAGS_GL QKGNQGVPDDFFDLMLMRCQGSRINDQRSEPP-V-----
EtAGS_GL AKGDLAVPDDFFEMLMRCQARIEDQRSTLP-M-----
HsLGN_GL MTNDNKEADEDEFFDILVKCQSRLDDQRCAPPPA-----
HsAGS3_GL -HGEPQEPDDEFFFMLIKIQSSRIDDQRCPPPDV-----
      .:****:****: *

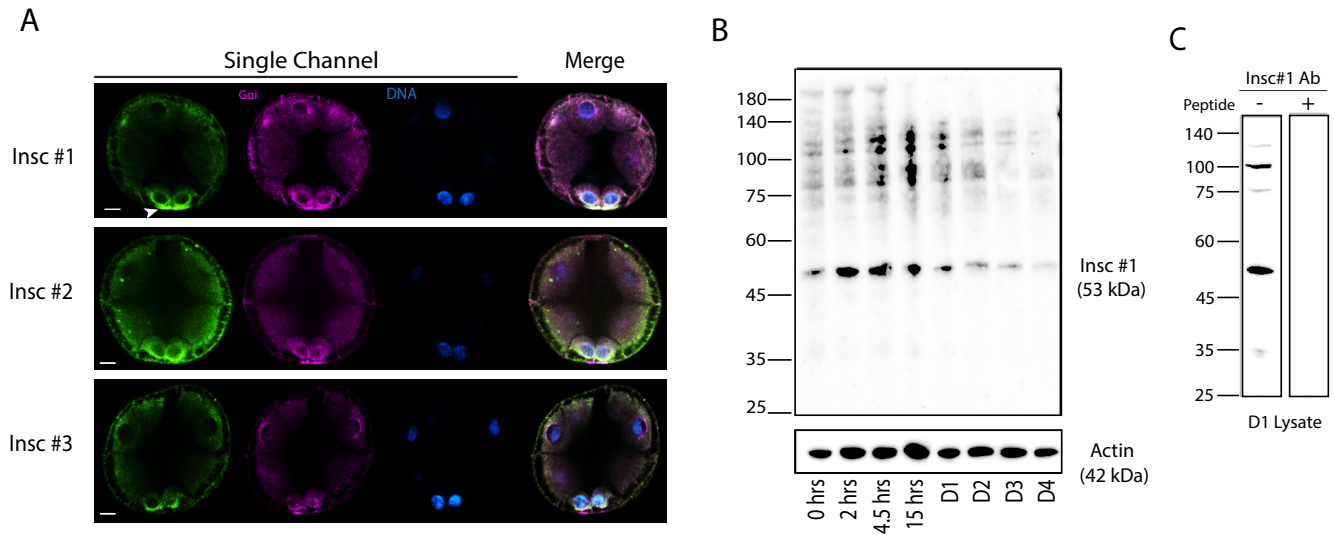
SpAGS_GL -QAPAPTVPDEFFSLIQRIQSKRIEEQRSIAPWEKGSGT-----SCVCFYDYDTSR
DmPins_GL QTGRGATVPDEFFSLIMKVQSGRMEDQRASIPFRNANNNSRS-----NNNG
PmAGS_GL -LQPAPTVPDEFFFALIQRVQSKRMDAQRSDKGPQEQTKT-----SD-----
EtAGS_GL -PTPAPTVPDEFFFTLIQRIQSRRIEEQRTIPPWEKGPKP-----SDK-----
HsLGN_GL -TTKGPTVPDEFFSLILRSQGKRMDEQRVLLQRDQNRDTDFGLKDFLQNNALLEFKNSG
HsAGS3_GL -LPRGPTMPDEFFSLIQRVQAKRMDEQRVDLAGGPEQGAGPPEPQQC-----QPG
      .:*****:***:*.*:**

SpAGS_GL TEGTACCVYSRPVQESL
DmPins_GL SAGGAGK-----
PmAGS_GL -----
EtAGS_GL -----
HsLGN_GL KKSADH-----
HsAGS3_GL AS-----

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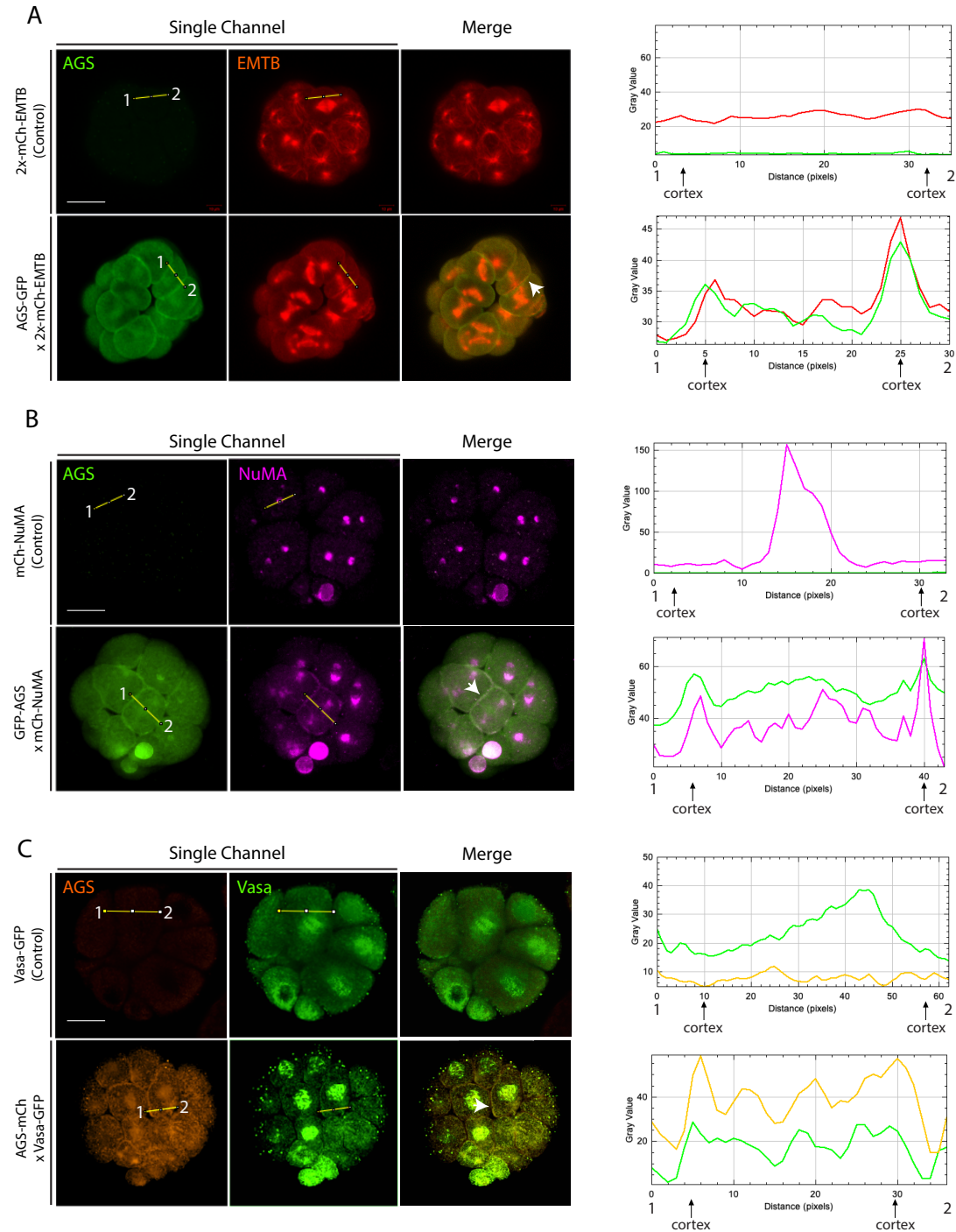
Supplemental Figure 5. Alignment of C-terminus GoLoco domain sequences used for chimeric mutants. Sea urchin *S. purpuratus* (SpAGS_GL), *Drosophila* (DmPins_GL), sea star *P. miniata* (PmAGS_GL), pencil urchin *E. tribuloides* (EtAGS_GL), human *H. sapiens* LGN (HsLGN_GL) and human *H. sapiens* AGS3 (HsAGS3_GL). Bold letters indicate GoLoco motif sequences. The green highlight indicates additional serine amino acid present uniquely in HsAGS3_GL and mutated to Alanine in AGS_AGS3GL_3S/A mutant. The highlighted amino acids between GL2 and GL3 and within GL3 are those mutated to match HsLGN_GL in AGS_AGS3GL_GL2GL3 mutant.

Supplemental Figure 6



Supplemental Figure 6. Insc protein expression during embryonic development. **A**, Endogenous Insc protein localization by immunofluorescence. Embryos were stained with three Insc antibodies (green) designed for different Insc amino acid sequence sections. Embryos were stained with Gai antibody (magenta) and Hoechst dye (blue). During the 16-cell stage, all antibodies show signal enriched at the vegetal pole. With #2 and #3 antibodies, some non-specific cortex signals were also observed around the entire embryo. **B**, Insc immunoblot analysis. Embryos were collected at 0, 2, 4.5, 15, 24, 48, 72, and 96 hours post fertilization and subjected to immunoblot with Insc #1 antibody. Actin (42kDa) was used as a loading control. The expected size of Insc is 53kDa. **C**, Peptide competition assay with Insc #1 antibody. The 24-hour lysate was used. Each experiment was performed at least two independent times. Scale bars=10 μ m.

Supplemental Figure S7



Supplemental Figure 7. SpAGS colocalizes with micromere-specific fate determinants. **A**, Embryos were co-injected with 2x-mCherry-EMTB (0.5 μ g/ μ l stock) mRNA with or without GFP-AGS (0.5 μ g/ μ l stock) mRNA. **B**, Embryos were co-injected with mCherry-NuMA (0.15 μ g/ μ l) mRNA with or without GFP-AGS (0.5 μ g/ μ l stock). **C**, Embryos were co-injected with Vasa-GFP (1 μ g/ μ l stock) mRNA with or without AGS-mCherry (0.5 μ g/ μ l stock) mRNA. The intensity of each signal, from one cortex to the other, was measured and plotted from point 1 to 2 on the corresponding graph (right) using *ImageJ*. White arrows indicate the cortical colocalization of each construct. All images represent over 50% of the embryos observed (n=30 or larger) per group. Scale bars=20 μ m.

Supplemental Figure 8 Sea urchin SpDlg and sea star PmDlg sequence alignment

SpDlg	-----	0
PmDlg	MN NIEEWEYEDIIILNRGGAGLGFSIAGGVNDNPHVPDDPSIFITKLI PGGAAAVGGRRLRLM	60
SpDlg	-----MVLRRWPRNPSTGK TLSIELIKGSKGLGFS	30
PmDlg	DV IIRVNETEIHNVPHQNAVDALKKTGSIVNLVVRRRRQRRRTGDVIRIKLVKGT KGLGFS	120
	: : * * : * * . . : * : * * : * * * * * * *	
SpDlg	IAGGCGNQHVPGDNGIFVTKIIDGGAEQDGR LQVGDKITAVGDN N LVEVTHE DAVAVLK	90
PmDlg	IAGGCGNQHVAGDNGIFVTKIIDGGAQM DGNLQIGDKIIAVGNDKLEEVTHE NAVAVLK	180
	**** * * * * * * * * * * * * * * * : * * : * * * * * * * * * * * * * * * *	
SpDlg	ATSNR VLLTVIKNATLLAGTQMAPYSQNDENAQP PDVSNVDMSAANPVYAT-----	141
PmDlg	ATSEVVLLTVIKGAVISYSPSPAS I-----TSQPDVSTMQSSPPPPYPEAIVSSVAGV	234
	*** : * * * * * * * . : : * : : * * * * * * * : * : * * *	
SpDlg	--MQPTLMQEPRSSDLQELDGAFGTNVAFGT N VANGANGANGSNGEPPSYNRVTNSQPP	198
PmDlg	SLQSEPAL-QRPTPPHQEMSMMNPLTYHQPPQDTAYGSM-----KMKHVPPSP	284
	: * : * * * : * * . . : : : * * : : : * * *	
SpDlg	NAFNNTPAKIFIQNDDGIPREPRQVVLNKGATGLGFNIVGGEDGEGIFISFILAGGVAD	258
PmDlg	TANYEGNGVKPVQVNDDFPREARTVVLNKGATGLGFNIVGGEDGEGIFISFILAGGVAD	344
	. * : . * : *	
SpDlg	L S G A L R R G D Q I L A V N S K D L V N A T H E D A A L A L K G A G Q V V T I D A Q Y K P E E Y N C F E A K I Q S L R	318
PmDlg	L S G E L K R G Q I L S V N K K D L R G A T H E D A A L A L K G T G Q T V L I E A Q Y K P E E Y N R F E A K I Q S L R	404
	*** : *	
SpDlg	EEMM Q N S V S - - P T G S L R T S A K R T L F V R T L F D Y D K S R D S G L P S Q G L S F D F G D I I H V T N A S D	376
PmDlg	EEMM N S S V S S T T G S L R T S A K R S L Y V R A L F D Y D K T K D S G L P S Q G L S F N Y G D I L H V T N A S D	464
	*** : *	
SpDlg	DEW W Q A R H I L P N G E G E G I G I I P S K R R V E R K E K A R L R S V K F S G N K D R N N I Q E P A G T V N S K G	436
PmDlg	DEW W Q A K Q V L P T G E E G E M G V I P S K R R V E K R E R A R L K S V K F S G R G G S - - - L E S K G S M N E K R	521
	*** : *	
SpDlg	KKGFF-RKF K K N R D S D Q E T S D F E P F T S N A S D S E S S Y R N E E V I L S Y E A V V Q Q E I K Y T R P V I	495
PmDlg	KKGFFSRKFKKGRSDLDTS DAEHITSNASDES S Y R N D E M I L S Y E G V V Q Q E V K Y T R P V I	581
	*** *	
SpDlg	I L G P G K D R I N D D L I S E M P D E F G S C V P H T T R A R P H E V D G R D Y H F V E S R E Q M E K D I Q N H L E	555
PmDlg	I L G S G K D R V N D D L I S E M F E K F G S C V P H T T R P R P H E V D K R D Y H F V E S R E Q M E K D I Q D H L E	641
	*** *	
SpDlg	I E A G Q Y N E N L Y G T S V A S V K E V S E K G K H C I L D V S G N A I K R L Q V A Q L Y P I A I F L K P K S V E S I	615
PmDlg	I E A G Q Y N E N L Y G T S V A S V R E V A D K G K H C I L D V S G N A I R R L Q V A N L H P I A I F I R P K S V E S V	701
	*** : *	
SpDlg	V E M N R R V S E D Q A R K T Y E R A L K L E Q E F L E Y F T A I I L G D T I D D I Y Q Q S K E I I H E Q S G P T I W I	675
PmDlg	M E M N R R M S R E Q A E K T Y E R A I K L E I E F G E F F T A I I S G D T L D E I Y Y K C K E V I H D Q S G S T I W V	761
	: * * * * * * * : * * . *	
SpDlg	PAKEKL 681	
PmDlg	PAKDKL 767	
	*** : * *	

PDZ
SH3
GUK

Supplemental Figure 8. Sea urchin (SpDlg) and sea star (PmDlg) sequence alignment. Blue, yellow, and green highlights indicate the PDZ, SH3, and GUK domains, respectively.

Supplemental Figure 9 Sea urchin SpInsc, pencil urchin EtInsc and sea star PmInsc sequence alignment

PmInsc	-----MNGFNES-ALAGWLADDVIEHMTDEDSVVRWLQDLRTMTECECMCILQG	49
SpInsc	-----MSDN---KNTESFP-EKKLMQRCDVDSVQQWLIDIRLMPEQECTVVMET	45
EtInsc	MTCLYGVVRTMYGPPAPVICEEKLPTSTSRFQLTEVRSVQQWLSDLQGMMECECN-ILET	59
	* . : . :: : ** : ** * : * * * * : :	
PmInsc	KPIRTSAHDMAKVTSARDNIYILRQRASSISADFAKLYKLDSGSWDQVH-WLCLSLTI	108
SpInsc	KPIHNSPLNLENLAATVKE SVYRLQSIKDGFLSLSQIYMCLDLSLQ-----	92
EtInsc	KPIHNSPLNLENVAATKENIYRLRNRAENLARELQQLYHKLTSGTYSQRLRPAVLALTL	119
	***:. * : : : : : : : * * : . . : : : * * .	
PmInsc	RLRFLHECTS YVPTMPLHLFQRQQTVLEECGRLVKHAESFEELNGTLPALPTLNVLSS	168
SpInsc	-----SVK--LFIQSLTT	103
EtInsc	HVRSCLEECNEYCPDF-LHMKEKQQVVIDAMATLVKNFEMYDNSKGCELD--SLIHSLTS	176
	: : * : :	
PmInsc	LGKAFNDLVHMVQGI LIQT VVRSIHESASEYNIKIAINS IHCLSEDDSEIRRLLAKEGAI	228
SpInsc	VAKAIKAYLEMVQ GALVVRKLVDMVQGAATELNVKFAVLTVHSLAQDGVWLRRLLIQEDTM	163
EtInsc	LGNALNSIRQTVQSALVVRKLVMMIQDSASELNVKFAVITAHSLSQDSIWLRRILIQENAM	236
	: : * : : . ** . * : : * : : * * * : * : * : * . : * * * : * : :	
PmInsc	QALLQLCQQASL--NAMKPMALKTVANLNCNVAEGRQLELERASGVEYLSEILSSPTATESV	286
SpInsc	RALLAICRI TTFSIRSIPTALEIVSILC TEVEGRAELEKVGVGALSEILASPTILEAV	223
EtInsc	GALFAVCRI TTLSIPSIVTALQAI STLCAEPEGRVELEKIGGLGFLGELSSPSTSEAV	296
	** : * : : : : ** : : * * * * * : . * : * * * * * : * : *	
PmInsc	KAEAA SVIAQITSPNIDNYHLLGFIEYMEDLLKDLTALSGSASSSSVFIATSAIANIT	346
SpInsc	KKEAASVLA EITS PDADIFHRMLCFIEHMEDLLRNLTVLC DQTS SPGVF LVATTAIANIT	283
EtInsc	KLEAASVLA EITS PDINSYHRMLCFIEHMEDLLKNLTALCDQTSNPLVFLVSASAIANIT	356
	* * * * * : * : * * * : : * : * * * * * : * * * * * . * * : : : * * * * *	
PmInsc	FMDSMACDC LASFHTARALVKGCNNGMANTLYCKDQVATILANMAVTDQCRKEIMVSGGT	406
SpInsc	FMDAMASDY LAEFQTA EVLIQGYLKGKAPSIYSKHQVITIANISCSKSCREQIVSSGGM	343
EtInsc	FMDTMSDF LAMFNATILIDGYTSGKAQNMYSKDQVVTIANMSGSEKREQI IKAGGM	416
	*** : * : * * * * * : * : * . * * : * : * * * * * : : * * * * * : * *	
PmInsc	ELLVDFLHTRALVHPSSAK--KGLKGSGLSGGCSGGKQDVVMSSSCEKVHQKAAIALA	463
SpInsc	NILAE LLQGV PSTSRPTSLSRKLSFDGN-----VSLGQMGEESAYEDIYQKAATALG	395
EtInsc	KVLVELLHTRAPFTTVKTNQVN---QRA-----DSKAACGMASTCEHVHQKSAIALG	465
	: * : : : : : : * : * : * * * * * * *	
PmInsc	RLARDQDSAE LILELDAI PRLVELCCNRRERNDSEFVLVACLAALRKLHNLGSPVFKSS	523
SpInsc	RLCQDYETCKMAVDLKVVSRLADLCKHARERNHRSDVLVACLAALRRIHSHIGSSPLKEN	455
EtInsc	RLCQDHD TAMMAVDLGVIPRLVDMCRHRRERNDSEFVLVACLAALRKLH SVVGASKFEPK	525
	*** : * : : : * : : * * : * : * * * . * * * * * : * : * * : :	
PmInsc	DVDQLIKPRLIDSYLMCSATK--ESFV	548
SpInsc	DVEQLIQPRLMESFLHCSTTVRRETFV	482
EtInsc	ALQQLIRPRLIESFVTCSSIGSTETFV	552
	: * * * * * : * : * * : * : *	

LBD (LGN-binding domain)

Supplemental Figure 9. Sea urchin (SpInsc), pencil urchin (EtInsc), and sea star (PmInsc) sequence alignment. The blue highlight indicates the LBD domain.