Tomato 26S Proteasome subunit RPT4a regulates ToLCNDV transcription and activates hypersensitive response in tomato

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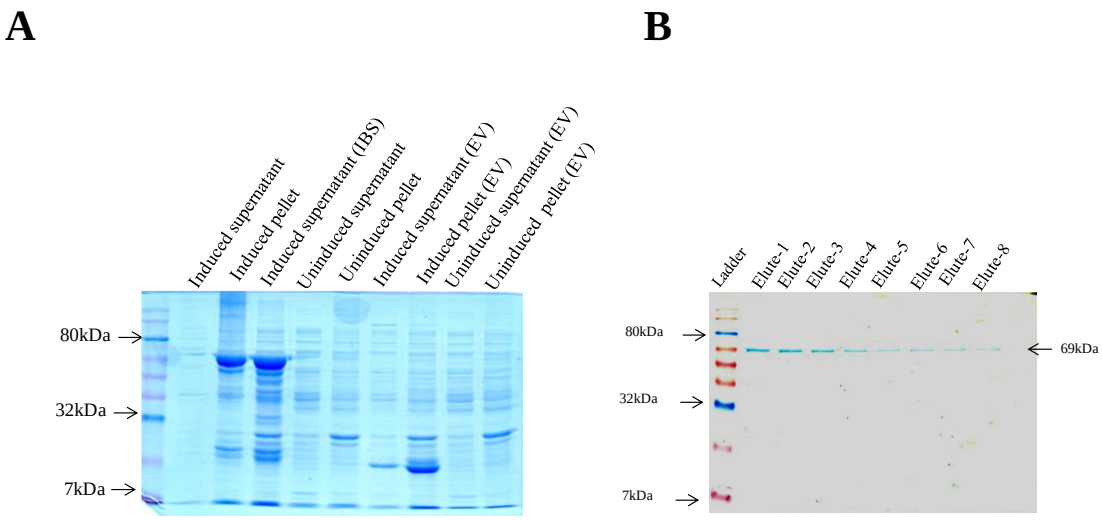
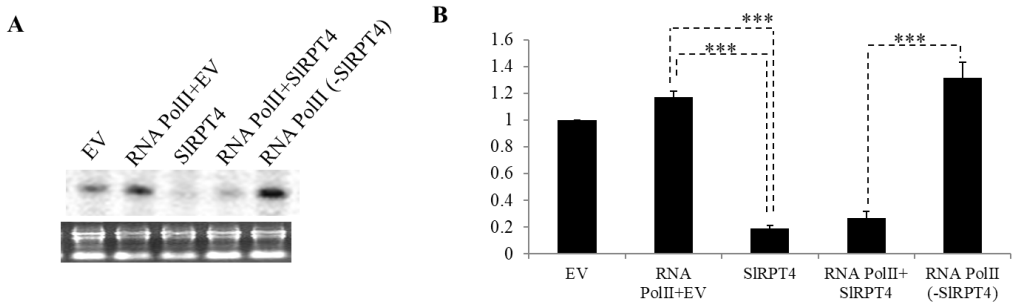
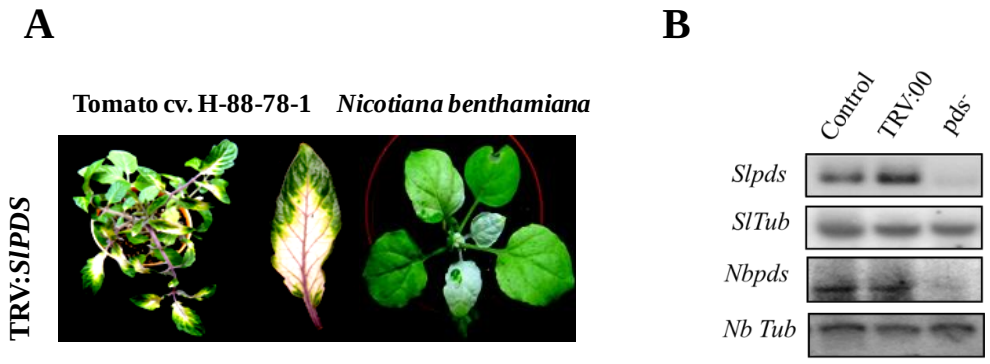


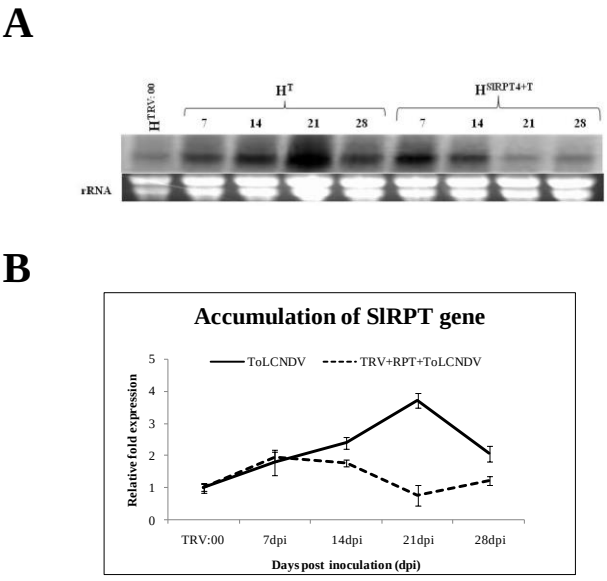
Supplementary Figure 1. Bacterial expression and purification of SlRPT4a. (A) Coomassie brilliant blue stained 12% SDS-PAGE containing induced and uninduced SlRPT4-GST fusion protein (69kDa); (B) Purified SlRPT4 protein. Band represents fractions of GSTaffinity purification of SlRPT4-GST fusion protein.



Supplementary Figure 2. Accumulation of Coat protein (CP) specific transcripts, (A) Northern hybridization showing the Relative accumulation of CP transcripts in the leaf samples infiltrated with empty vector (EV), SlRPT4-cmyc and RNA Pol II-3-gfp construct alone, and co-infiltrated with RNA Pol II-3-gfp and SlRPT4-muc construct. Fragment corresponding to ToLCNDV-CP gene was used as probe. Total RNA is shown as equivalent loading in the experiment. Data depicts means±SD of three independent experiments (n=3); *, $P<0.05$; **, $P<0.01$; ***, $P<0.001$.

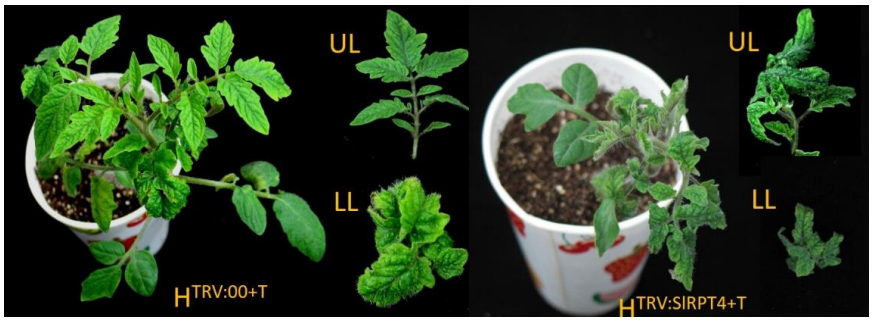


Supplementary Figure 3. TRV-based VIGS in tomato and *Nicotiana benthamiana*. (A) Phenotype of tomato and *N. benthamiana* plant at 21 day post-silencing. Fragments corresponding to *Slpds* and *Nbpds* were used to produce TRV-based gene silencing constructs. After 21 day post silencing typical leaf bleaching symptoms were observed. (B), Northern blot analysis to evaluate the relative level of pds gene in control (without virus, mock or silencing treatments), vector infiltrated (TRV:00) and *pds* silenced (pds-) plants. Tubulin gene from tomato and Nicotiana were used as internal control.



Supplementary Figure 4. Accumulation of SlRPT4 in H^T (ToLCNDV) and $H^{SlRPT4+T}$ (TRV:SlRPT4+ToLCNDV). (A) Northern hybridization to evaluate the accumulation of *SlRPT* transcript. (B) Relative accumulation of *SlRPT4* in the experimental samples. Tomato plant infected with TRV: 00 vector was used as negative control. Bars show standard deviations (±SD). Ethidium bromide-stained total RNA has been shown as the equivalent loading control of the experiment.

A



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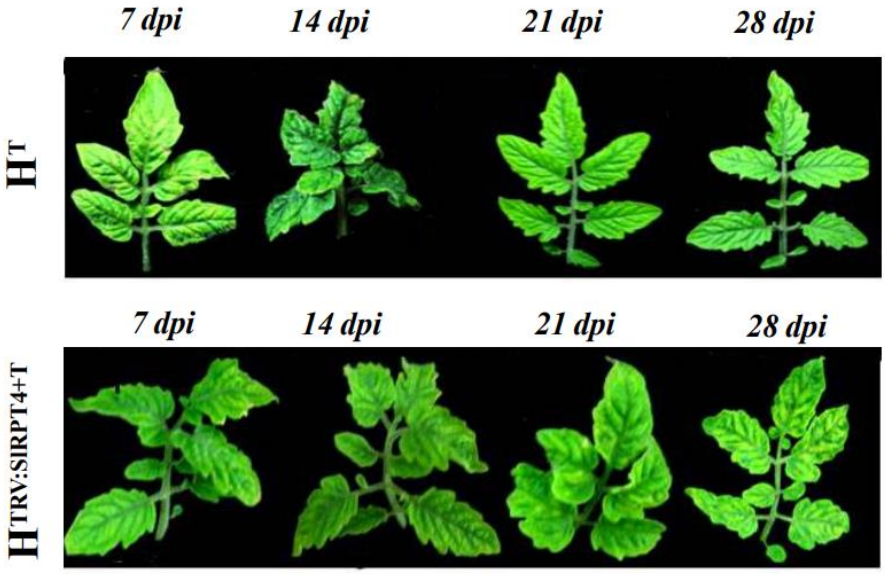
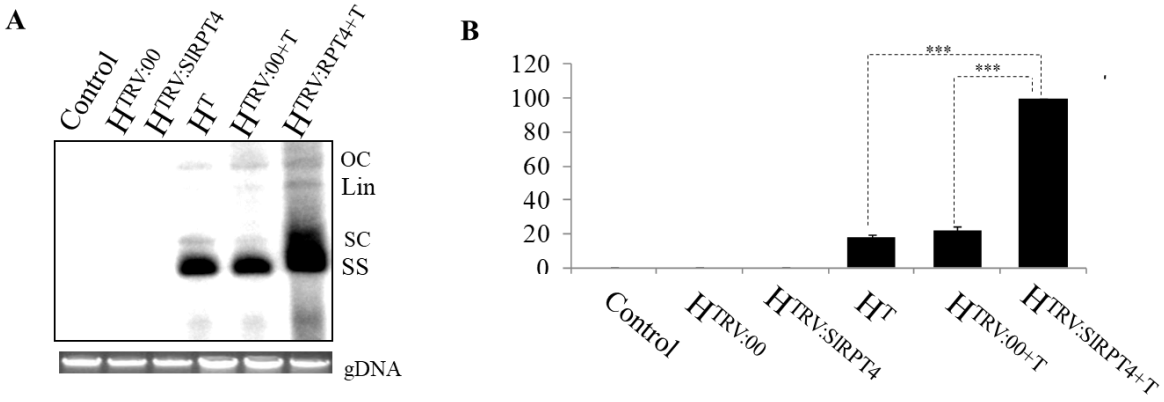
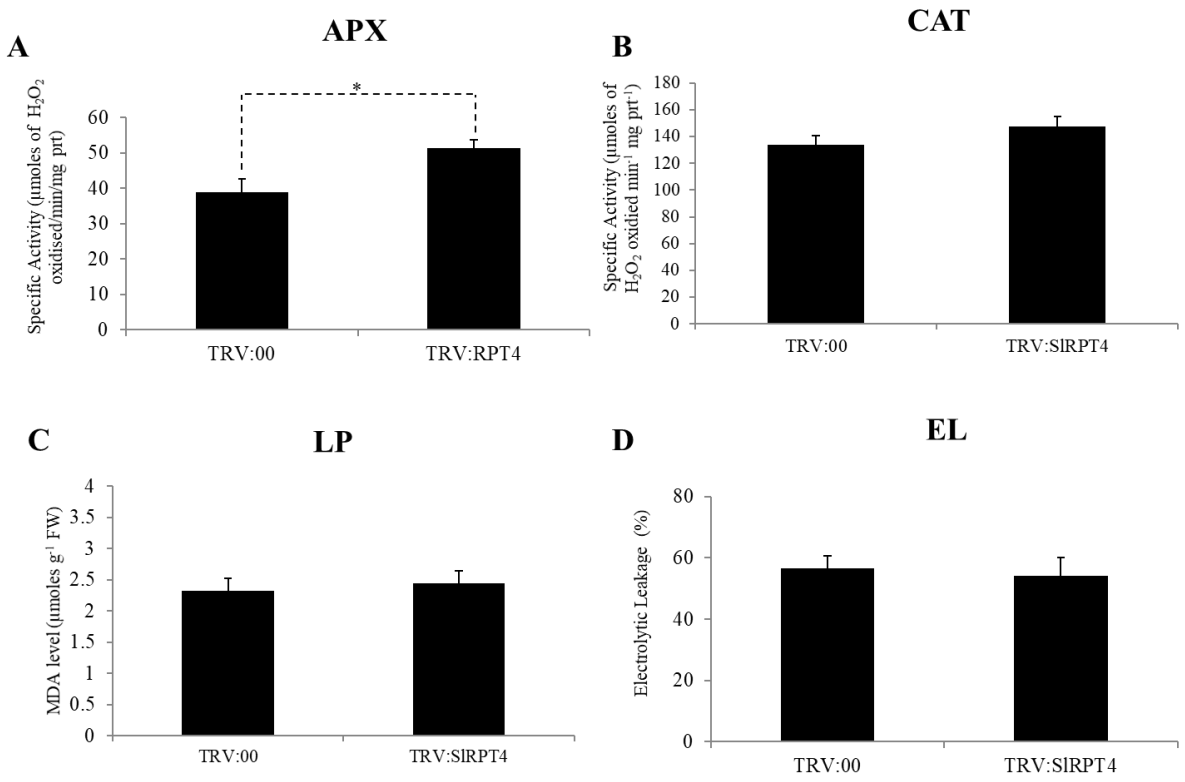


Supplementary Figure 5. Phenotype of Mock and SlRPT4 silenced cv. H-88-78-1 at 21 day post ToLCNDV infection. (A) Symptom remission. Systemic leaves showed symptom recovery in mock plants, however SlRPT4 silenced cv. H-88-78-1 was failed to recover from the ToLCNDV infection. $H^{TRV:00+T}$, mock plant infected with ToLCNDV; $H^{TRV:SlRPT4+T}$, SlRPT4 silenced plant infected with ToLCNDV, UL, upper leaf; LL, lower leaf. (B) Comparison of progression of leaf curl symptom between Control (H^{T}) and *SlRPT4* silenced cv. H-88-78-1 ($H^{TRV:SlRPT4+T}$) at 7-28 dpi of ToLCNDV.



Supplementary Figure 6. Accumulation of DNA-B specific ToLCNDV molecule. Southern blot of tomato genomic DNA from all experimental plants were hybridized with ToLCNDV-BC1 (encoding Movement proteins) gene specific probe. Replicative forms of ToLCNDV genome are designate as open circular (OC), linear (Lin), supercoiled (SC) and single strand (SS). TRV:00 infiltrated H-88-78-1 was taken as a mock control. Ethidium bromide stained DNA from each experiment were shown as equivalent loading. (C) Relative accumulation of viral DNA in the samples H^{T} and $H^{SlRPT4+T}$ at different time points. Data depicts means±SD of three independent experiments (n=3); *, $P<0.05$; **, $P<0.01$; ***, $P<0.001$.



Supplementary Figure 7. Estimation of antioxidant enzyme activity in cv. H-88-78-1. (A) Specific activity of APX was measured as 1 µmol of ascorbate oxidized min^{-1}. (B) Specific activity of CAT was measured as 1 µmol H_2O_2 oxidized min^{-1}. (C) Levels of lipid peroxidation expressed in terms of MDA concentration. (D) Percentage electrolytic leakage. Data depicts means±SD of three independent experiments (n=3); *, $P<0.05$; **, $P<0.01$; ***, $P<0.001$. Mock, TRV:00 infiltrated cv. H-88-78-1; TRV:SlRPT4; SlRPT4 silenced cv. H-88-78-1.

Supplementary Table-S1. List of primers used in the study

Primers used for bacterial expression of SlRPT4 protein		
Primer Name	**Forward Primer Sequence**	**Reverse Primer Sequence**
pGEX4:RPT4	CGGGATCCATGGCGACCGAAGAAGACG	CGGAATTCTTATTCCTTGCCAAAATCAG
Primers used for VIGS		
pTRV-Slpds	CCGCTCGAGCTGACGAGCTTTCGATGCAGTGC	CGGGATCCATATATGGGACATTTATCACAGGA
pTRV-SlRPT4	CCGCTCGAGGTAATGCAGCCAACTCTTTCTCT	CGGGATCCCCTCTACTATATTACACCCCGTCCT
Primers used for Southern blot analysis		
Coat protein	ACAGAAAACCCAGAATGTACAGAA	CAACATTAAGGCATTTTCAGTATG
BC1	GTTTTGTGTCCCCCTCCGTCA	GTTTTGTGTCCCCCTCCGTCA
Primers used for transient expression analysis		
pCAMBIA1302: SlRPT4	CATGCCATGGTAATGGCGACCGAAGACG	GGACTAGTTTATTCCTTGCCAAAATCAGCA
Primers used for Northern blot analysis		
Rep gene	TTTAAAGTGCTTTAGATAGTG	CACAATTACTTGTGTGGACAT
Coat Protein	ATGAAATTCACGCTACATGGCCTA	CGTTGAAATGATGATATCTGCTGG
SlRPT4	CCGCTCGAGGTAATGCAGCCAACTCTTTCTCT	CGGGATCCCCTCTACTATATTACACCCCGTCCT
Slpds	CCGCTCGAGCTGACGAGCTTTCGATGCAGTGC	CGGGATCCATATATGGGACATTTATCACAGGA
Nbpds	TAAACCCTGACGAGCTTTCGATGC	TTTATCACAGGAACTCCCACTAGC
αTubulin	TCAAACCTCAAAGAAGCTGTCA	ACAATTTATCCCTCACCACAGG
Primers used for EMSA and ChIP assay		
DNA-A-IR_EMSA (2592-47)	AAAACTTGTCGTTTTGATT	TGGTTGAGGGCCCACCTAAA
DNA-B-IR_EMSA (2617-67)	ACACCATATGGCATTATTGTAAT	AACGGCGTGCAATGATTACGC
DNA-A-Rep_EMSA (1939-2046)	GACTATGCTTATGGGCCTAAA	CCATTTTCAATTTCATCCT
IR_ChIP (2592-47)	AAAACTTGTCGTTTTGATT	TGGTTGAGGGCCCACCTAAA
pENTER-RPT4	CACCATGGCGACCGAAGAAGACGCCG	TTCCTTGCCAAAATCAGCACTGTAG
pENTER-RNA Pol-II-3	CACCATGGAGGGCGTTTCGTACCAG	TTAACCTCCACGCATATGAGCGCCCA
Actin 7	CTGGTGTGTGATAATGGAACG	GCTTCATCACCAACATACGC