

Supplemental Figure 1. TNFAIP3 (A20) expression in human AML and its role in chemotherapy. (**A**) Recurrent mutations in TCGA AML patients grouped by initial response to chemotherapy or induction failure. (**B**) Initial response to 7+3 chemotherapy vs induction failure in young (<60 years old) and older (>60 years old) AML Alliance cohort patients that have either a low or mid/high NF- κ B signature: Young, NF- κ B low (n = 249); young, NF- κ B mid/high (n = 438); older, NF- κ B low (n = 33); older NF- κ B mid/high (n = 153). (**C**) Expression (grey) and mutation frequency (blue) of TNFAIP3/A20 across adult cancers from TCGA. AML is highlighted in red. (**D**) Expression of TNFAIP3/A20 across pediatric cancers from the PeCan database. AML is highlighted in red. (**E**) Mutations or AML subtype in patients with AML from TCGA (left) or BEAT-AML (right) clustered in Group 1 (low A20 expression) and Group 2 (high A20 expression). P values were determined with hypergeometric testing. Source data are provided as a source data file.



Supplemental Figure 2. TNFAIP3 (A20) expression in human AML and its role in chemotherapy sensitivity. (A) Sensitivity of AML PDX samples to either doxorubicin (left) or cytarabine (right) at 1.56, 6.25, 25, and 100 nM. A >67% reduction or >25% reduction in viability was required to be classified as resistant to 100 nM doxorubicin or 100 nM cytarabine, respectively. (B) TNFAIP3/A20 expression in AML PDX samples that are either resistant or sensitive to doxorubicin (left) or cytarabine (right). A >67% reduction or >25% reduction in viability was required to be classified as resistant to 100 nM doxorubicin in viability was required to be classified as resistant to 100 nM doxorubicin or 100 nM cytarabine, respectively. (C) Correltation of TNFAIP3/A20 mRNA expression and relative inhibition of viability of PDX AML samples (n =12) treated with 100 nM doxorubicin. (D) Relative inhibition of cell viability in A20^{High} and A20^{Low} PDX AML samples treated with doxorubicin (100 nM). (E) TNFAIP3/A20 expression in AML PDX samples that are either resistant or sensitive to venetoclax (Ven) and azacitidine (Aza) (adapted from Pei et al, Cancer Discovery, 2020). (F) Cell viability (trypan blue exclusion) of MLL-AF9;FLT3-ITD cells expressing an empty vector or A20 following treatment with doxorubicin. Error bars represent the standard error of the mean. *, P < 0.05. Source data are provided as a source data file.



Supplemental Figure 3. Characterization of A20-deficient AML mouse models. (**A**) Genotyping of A20+/+, A20+/f, and A20f/f Rosa CreER treated with vehicle (100% ethanol) or 1 μ M 4-OHT for 48 hours. (**B-E**) Quantitative PCR showing relative A20 mRNA expression in A20+/+, A20+/f, and A20f/f Rosa CreER treated with vehicle (100% ethanol) or 1 μ M 4-OHT for 48 hours in either the MLL-AF9 (B), MN1 (C), or shp53;KRAS^{G12V} (D) leukemic models or healthy lineage negative bone marrow (E). (**F-I**) Immunoblot showing A20 protein abundance in A20+/+, A20+/f, and A20f/f Rosa CreER treated with vehicle (100% ethanol) or 1 μ M 4-OHT for 48 hours in the MLL-AF9 (F), MN1 (G), or shp53;KRAS^{G12V} (H) AML models or healthy lineage negative BM (I). (J) H&E staining of tibia, spleen, and liver sections from moribund mice that were transplanted with either A20+/+ MLL-AF9 or A20f/f MLL-AF9 cells followed by tamoxifen treatment. Source data are provided as a source data file.



Supplemental Figure 4. Characterization of hematopoiesis in A20-deficient mice. (A) Overview of A20 floxed (fl) RosaCreER healthy bone marrow transplant. Four weeks after transplant, recipient mice were injected intraperitoneally with 1 mg tamoxifen daily for 5 days to induce recombination of A20. (B) Peripheral blood counts in recipient mice before tamoxifen treatment and at 16 and 45 days post-tamoxifen. White blood cells (WBC); red blood cells (RBC); platelets (PLT). Source data are provided as a source data file.



MLL-AF9

Supplemental Figure 5. Characterization of necroptosis in A20-deficient AML cells. (**A**) Correlation of TNFα and A20 mRNA expression in primary AML samples (adapted from BEAT-AML dataset). (**B**) Immunoblot of RIPK3 activation in MOLM14 cells transduced with two different shRNAs for A20 (shA20) or scrambled control (shControl). (**C**) Necroptosis was assayed in A20^{High} AML patient-derived samples (JM62) using the lactate dehydrogenase (LDH) release assay. (**D**) Immunoblot of IKKβ activation and Caspase 3 cleavage in A20+/+, A20+/-, and A20-/- MLL-AF9 AML cells. (**E**) Transmission electron microscopy cell death morphology controls in A20f/f MLL-AF9 AML cells. Apoptosis was induced by treating cells with 125 μM hydrogen peroxide for 16 hours. Necroptosis was induced by treating cells with 200 ng/mL TNF, 100 nM Birinapant, and 5 μM z-VAD-fmk for 16 hours. Error bars represent the standard error of the mean. *, P < 0.05. For each immunoblot (**B**,**D**), all samples derive from the same experiment, but different gels for each antibody were processed in parallel. Source data are provided as a source data file.



Supplemental Figure 6. Evaluation of necroptosis signaling in A20-deficient AML cells. (**A-B**) Viable cells by trypan blue exclusion in A20+/+ and A20f/f MLL-AF9 that received 1 uM 4-OHT and were treated with 2 uM z-VAD-fmk pan-caspase inhibitor (A) or 10 nM IKK NF- κ B inhibitor (B). (**C**) Colony forming assay of MV4;11 cells transduced with shRNA to A20 or scrambled control that were seeded into methylcellulose with 30 μ M Necrostatin-1. Colonies were counted after 8 days. (**D**) Colony forming assay of A20+/+ and A20f/f MLL-AF9 cells that were treated with 1 μ M 4-OHT for 48 hours, then seeded into methylcellulose with 1 μ M 4-OHT. Colonies were counted at day 7 before collecting and reseeding an equal cell number into methylcellulose with 1 uM 4-OHT or vehicle. (**E**) Immunoblot of A20+/+RIPK3-/-, A20f/fRIPK3-/-, and A20f/fRIPK3+/+ MLL-AF9 cells following treatment with 1 μ M 4-OHT or vehicle for 48 hours. (**F**) Immunoblot showing RIPK3 expression in MV4;11 following sgRIPK3. (**G**) Sequence trace of MV4;11 reference genomic DNA sequence (top) and genomic DNA sequences for two different MV4;11 sgRIPK3 clones. The sgRIPK3 sequence is highlighted in grey.Error bars represent the standard error of the mean. For each immunoblot (**E**,**F**), all samples derive from the same experiment, but different gels for each antibody were processed in parallel. Source data are provided as a source data file.



Supplemental Figure 7. Effects of chemotherapy on AML cells. (**A-B**) Relative number of live cells by trypan blue exclusion in MLL-AF9 mouse cells and CD34+ MLL-AF9;FLT3-ITD human cells following doxorubicin (A) or cytarabine treatment (B) for 24 hours. (**C**) Proliferation assay showing live cells by trypan blue exclusion in CD34+ MLL-AF9;FLT3-ITD cells treated with the indicated concentrations of doxorubicin. (**D**) Proliferation assay showing live cells by trypan blue exclusion in CD34+ MLL-AF9;FLT3-ITD cells treated with 250 nM cytarabine. (**E**) A20 protein expression was evaluated on CD34+ MLL-AF9;FLT3-ITD cells recovered following 5 day treatment with the indicated concentrations of doxorubicin. (**F**) A20 protein expression was evaluated in CD34+ MLL-AF9;FLT3-ITD cells expressed vector or A20 following treatment with azacitine (+, 2.5 μ M; ++, 5 μ M) or venetoclax (+, 0.5 μ M; ++, 1 μ M). For each immunoblot (**E**,**F**), all samples derive from the same experiment, but different gels for each antibody were processed in parallel. Source data are provided as a source data file.