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Last updated by author(s):	May 26, 2024

Reporting Summary

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our Editorial Policies and the Editorial Policy Checklist.

For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section

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n/a	Confirmed
	$oxed{x}$ The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
	🗴 A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
	The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.
x	A description of all covariates tested
	🕱 A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
	For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i>) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted Give <i>P</i> values as exact values whenever suitable.
x	For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
×	For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
x	Estimates of effect sizes (e.g. Cohen's d, Pearson's r), indicating how they were calculated
'	Our web collection on <u>statistics for biologists</u> contains articles on many of the points above.

Software and code

Policy information about availability of computer code

Data collection

One-photon fluorescence imaging data were collected using an Andor Zyla 4.2 sCMOS camera. Fly trajectories on the trackball were monitored using two optical USB pen mice (i-Pen Mouse; Finger System).

Data analysis

We used MATLAB (v2018b and v2020b; Mathworks) to analyze the imaging and fly behavior data. We wrote custom MATLAB scripts to implement the computational model (see Code Availability Statement, with Github address).

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio guidelines for submitting code & software for further information.

Data

Policy information about <u>availability of data</u>

All manuscripts must include a <u>data availability statement</u>. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our policy

The raw voltage imaging data are available from the corresponding authors upon reasonable request. The analyzed voltage imaging traces are available at Zenodo: 10.5281/zenodo.10998457.

Research involving human participants, their data, or biological material

Policy information about studies with <u>human participants or human data</u>. See also policy information about <u>sex, gender (identity/presentation)</u>, and sexual orientation and race, ethnicity and racism.

Reporting on sex and gender

Use the terms sex (biological attribute) and gender (shaped by social and cultural circumstances) carefully in order to avoid confusing both terms. Indicate if findings apply to only one sex or gender; describe whether sex and gender were considered in study design; whether sex and/or gender was determined based on self-reporting or assigned and methods used. Provide in the source data disaggregated sex and gender data, where this information has been collected, and if consent has been obtained for sharing of individual-level data; provide overall numbers in this Reporting Summary. Please state if this information has not been collected.

Report sex- and gender-based analyses where performed, justify reasons for lack of sex- and gender-based analysis.

Reporting on race, ethnicity, or other socially relevant groupings

Please specify the socially constructed or socially relevant categorization variable(s) used in your manuscript and explain why they were used. Please note that such variables should not be used as proxies for other socially constructed/relevant variables (for example, race or ethnicity should not be used as a proxy for socioeconomic status).

Provide clear definitions of the relevant terms used, how they were provided (by the participants/respondents, the researchers, or third parties), and the method(s) used to classify people into the different categories (e.g. self-report, census or administrative data, social media data, etc.)

Please provide details about how you controlled for confounding variables in your analyses.

Population characteristics

Describe the covariate-relevant population characteristics of the human research participants (e.g. age, genotypic information, past and current diagnosis and treatment categories). If you filled out the behavioural & social sciences study design questions and have nothing to add here, write "See above."

Recruitment

Describe how participants were recruited. Outline any potential self-selection bias or other biases that may be present and how these are likely to impact results.

Ethics oversight

Identify the organization(s) that approved the study protocol.

Note that full information on the approval of the study protocol must also be provided in the manuscript.

Field-specific reporting

Please select the one belo	w that is the best fit for your research.	If you are not sure, read the appropriate sections before making your selection.
x Life sciences	Behavioural & social sciences	Ecological, evolutionary & environmental sciences

For a reference copy of the document with all sections, see nature.com/documents/nr-reporting-summary-flat.pdf

Life sciences study design

All studies must disclose on these points even when the disclosure is negative.

Data exclusions

For imaging experiments, data exclusion was performed only in the event of poor imaging quality (e.g. low signal-to-noise ratio, substantial brain motion, weak fluorescence signals) or for unhealthy animals (e.g. that failed to move on the trackball). In behavioral experiments, we similarly excluded the data from unhealthy animals that were unable to walk properly on the trackball.

Sample sizes were chosen using our own and published empirical measurements (Huang et al., Nat. Comm. 2018; Hige et al., Nature 2015) to

Replication

Sample size

Each experiment was replicated in at least 10 individual flies. In total, we used >500 flies across the entire study.

Randomization

There was no formal randomization procedure, but flies were informally chosen in a random manner for all studies. Please see the "Fly stocks" section of the Methods.

Blinding

Experimenters were not blind to the genotypes of the animals used. However, all data collection and analyses were performed automatically using computer software that was uniformly applied to all flies irrespective of their genotypes. Please see the "Fly stocks" section of the Methods.

Reporting for specific materials, systems and methods

gauge effect magnitudes. Please see the 'Statistical analyses' section of the Methods.

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

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Materials & experime	ntal systems Methods	
n/a Involved in the study	n/a Involved in the study	
X Antibodies	ChIP-seq	
x Eukaryotic cell lines	Flow cytometry	
Palaeontology and a	archaeology MRI-based neuroimaging	
Animals and other o	organisms	
Clinical data		
Dual use research o	f concern	
X Plants		
Animals and othe	r research organisms	
Policy Information about <u>st</u> Research Laboratory animals	Drosophila melanogaster, female, aged 3-8 days. Detailed information is described in the Methods section. The full list of fly lines used is provided in the Extended Data Table 1.	
Wild animals	No wild animals were used in this study.	
Reporting on sex	We used only female flies in this study.	
Field-collected samples	No field-collected animals were used in this study.	
Ethics oversight	Institutional approval of experimental procedures and ethics oversight was not required for studies of fruit flies.	
Note that full information on t	he approval of the study protocol must also be provided in the manuscript.	
Dlants		
Plants		
Seed stocks	Report on the source of all seed stocks or other plant material used. If applicable, state the seed stock centre and catalogue number. If plant specimens were collected from the field, describe the collection location, date and sampling procedures.	
Novel plant genotypes	Describe the methods by which all novel plant genotypes were produced. This includes those generated by transgenic approaches, gene editing, chemical/radiation-based mutagenesis and hybridization. For transgenic lines, describe the transformation method, the	

Describe the methods by which all novel plant genotypes were produced. This includes those generated by transgenic approaches, gene editing, chemical/radiation-based mutagenesis and hybridization. For transgenic lines, describe the transformation method, the number of independent lines analyzed and the generation upon which experiments were performed. For gene-edited lines, describe the editor used, the endogenous sequence targeted for editing, the targeting guide RNA sequence (if applicable) and how the editor

Authentication

was applied.

Describe any authentication procedures for each seed stock used or novel genotype generated. Describe any experiments used to assess the effect of a mutation and, where applicable, how potential secondary effects (e.g. second site T-DNA insertions, mosiacism, off-target gene editing) were examined.