

Fig. S1. Quality control plots for the *He* scRNA-seq atlas. (A) Violin plots showing the distribution of the number of RNA features (unique gene transcripts) in each cell for each sample time point. (B) Violin plots showing the distribution of the number of RNA transcripts in each cell for each sample time point.

Fig. S2. Dot plot of cell type marker gene expression patterns. ~3-6 enriched marker genes (see Table S1) for each cell type in the scRNA-seq atlas were selected, and the average expression of the genes in each cell type was plotted.

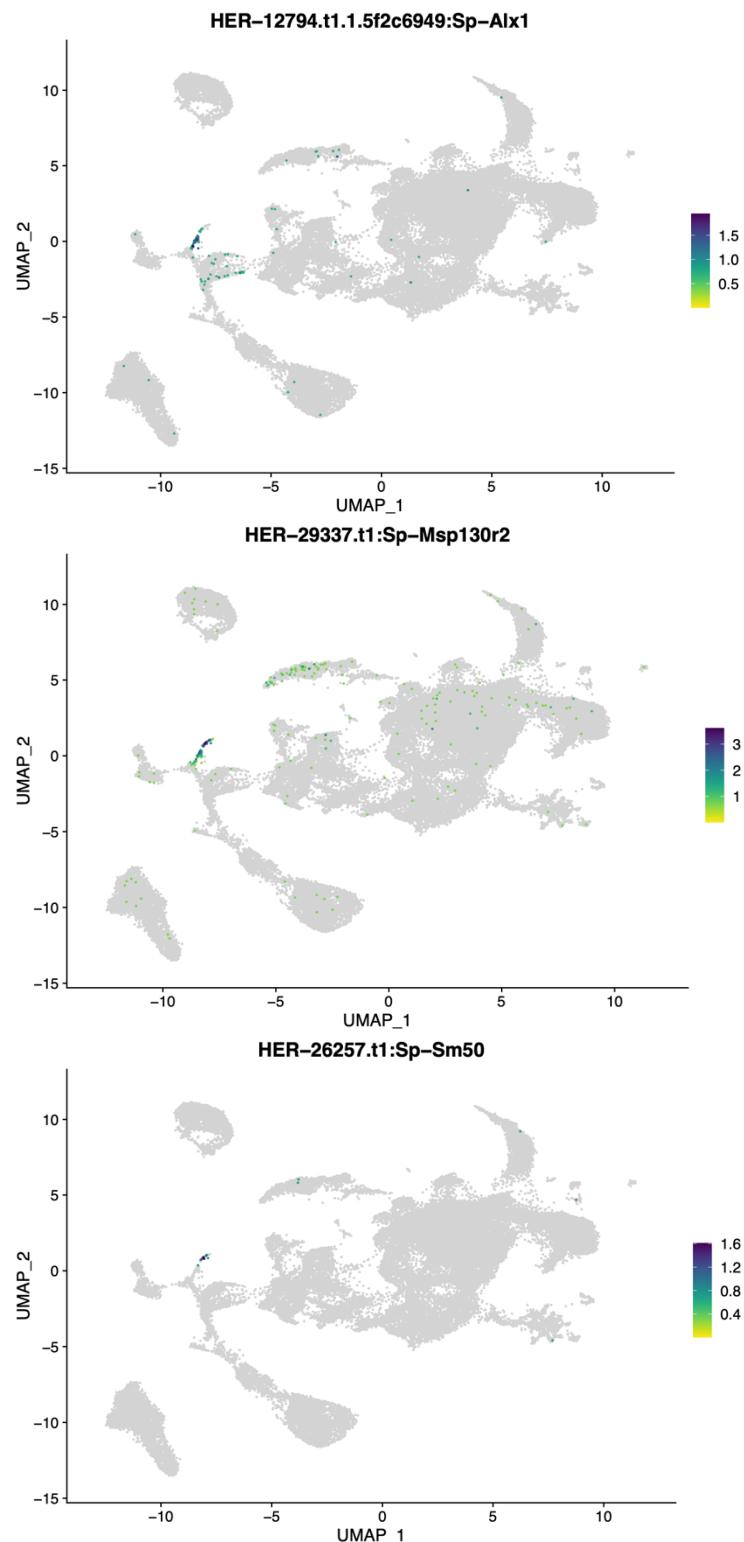


Fig. S3. Expression patterns of skeletogenic cell marker genes. These UMAPs show the expression patterns of three of the marker genes used to annotate the skeletogenic cell lineage in the *He* scRNA-seq atlas.

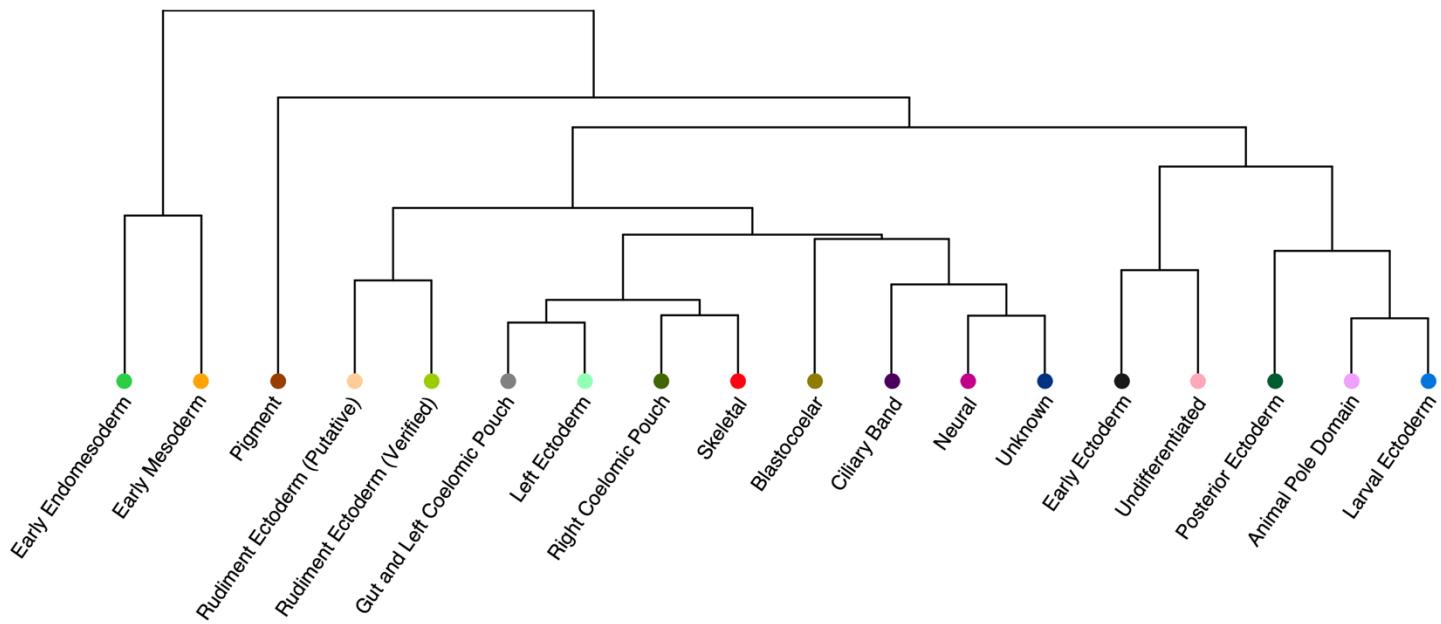


Fig. S4. Phylogenetic tree of cell types in the scRNA-seq atlas. This plot was generated using Seurat's BuildClusterTree function, which generates the phylogenetic tree using pairwise distances between the cell types in gene expression space.

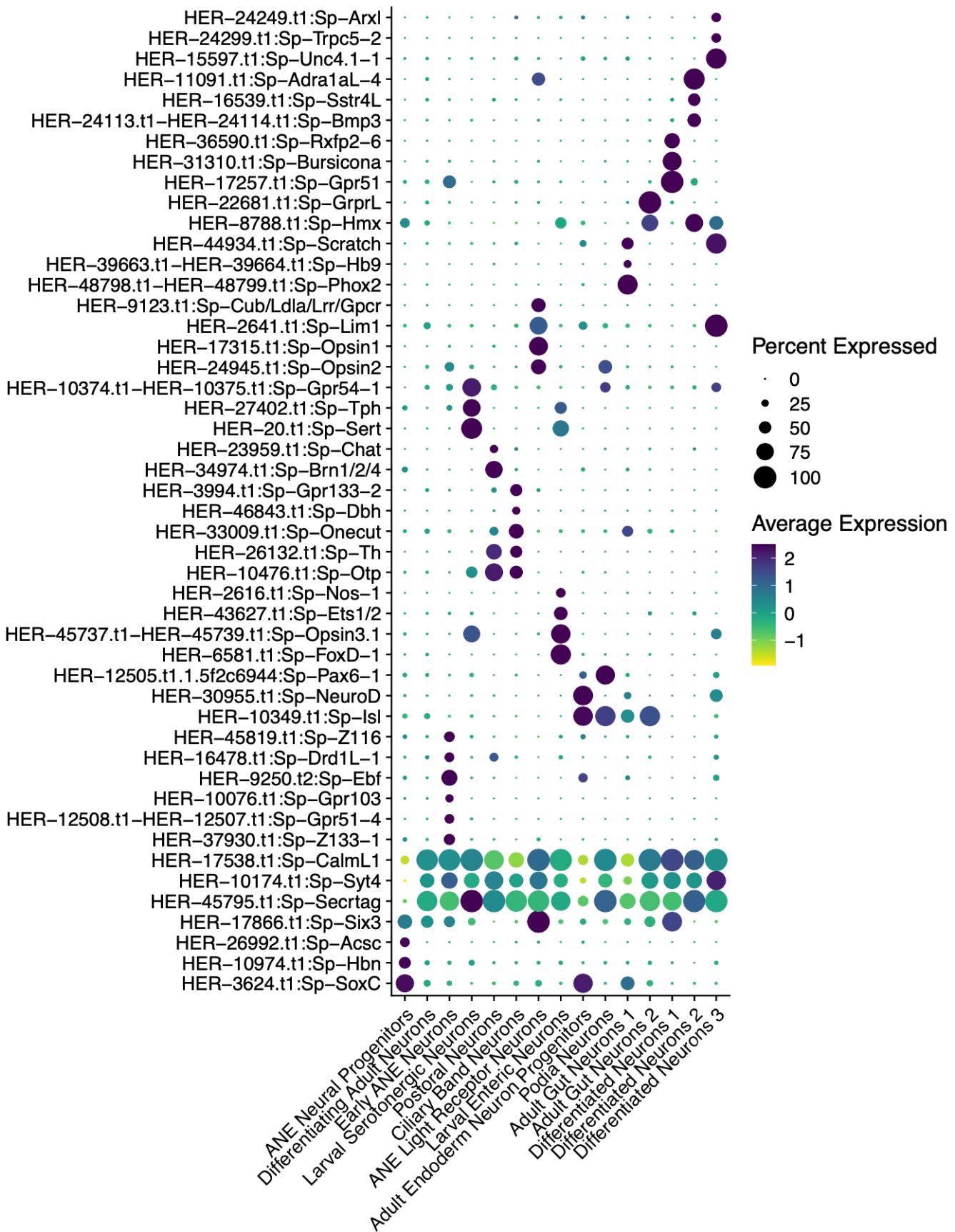
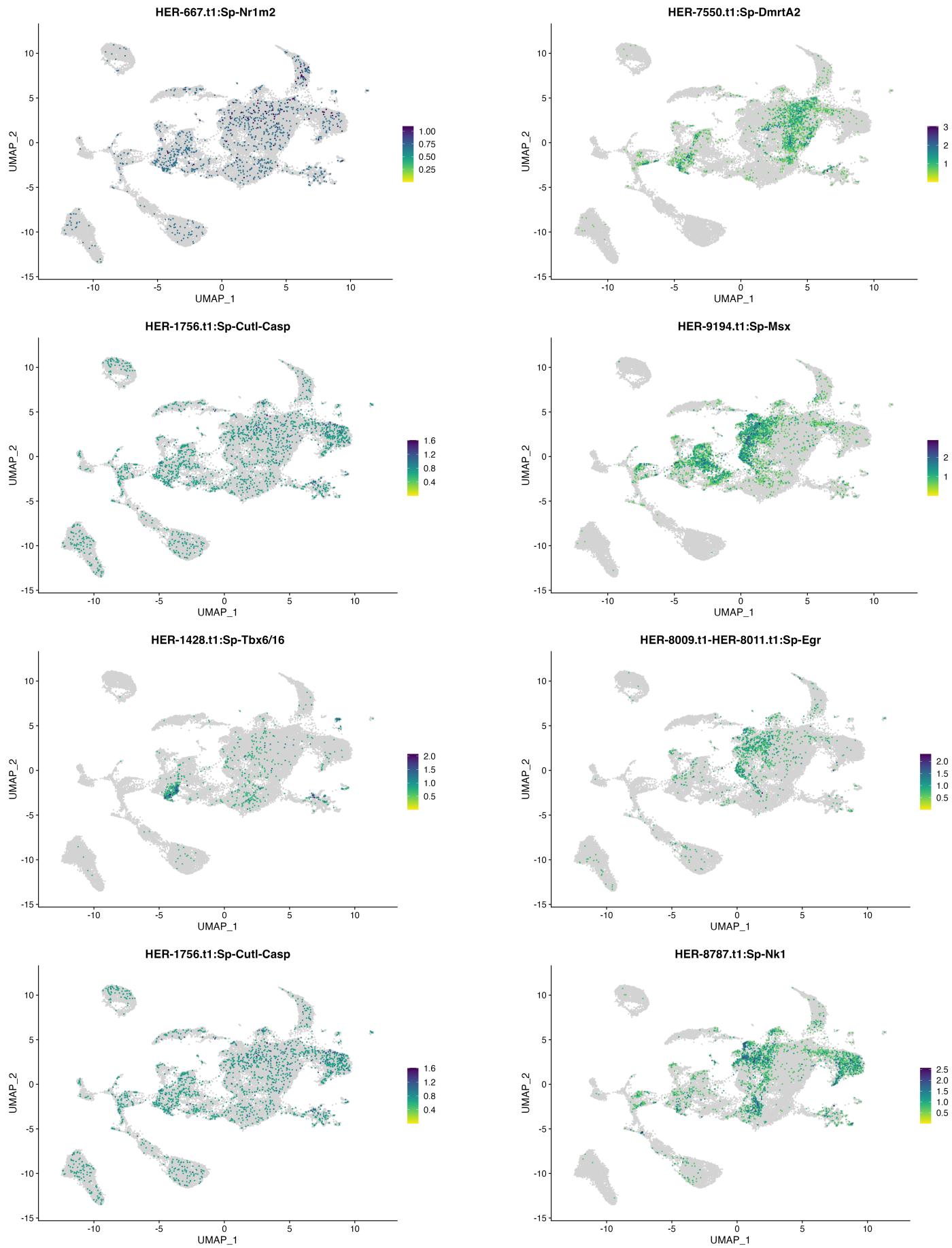
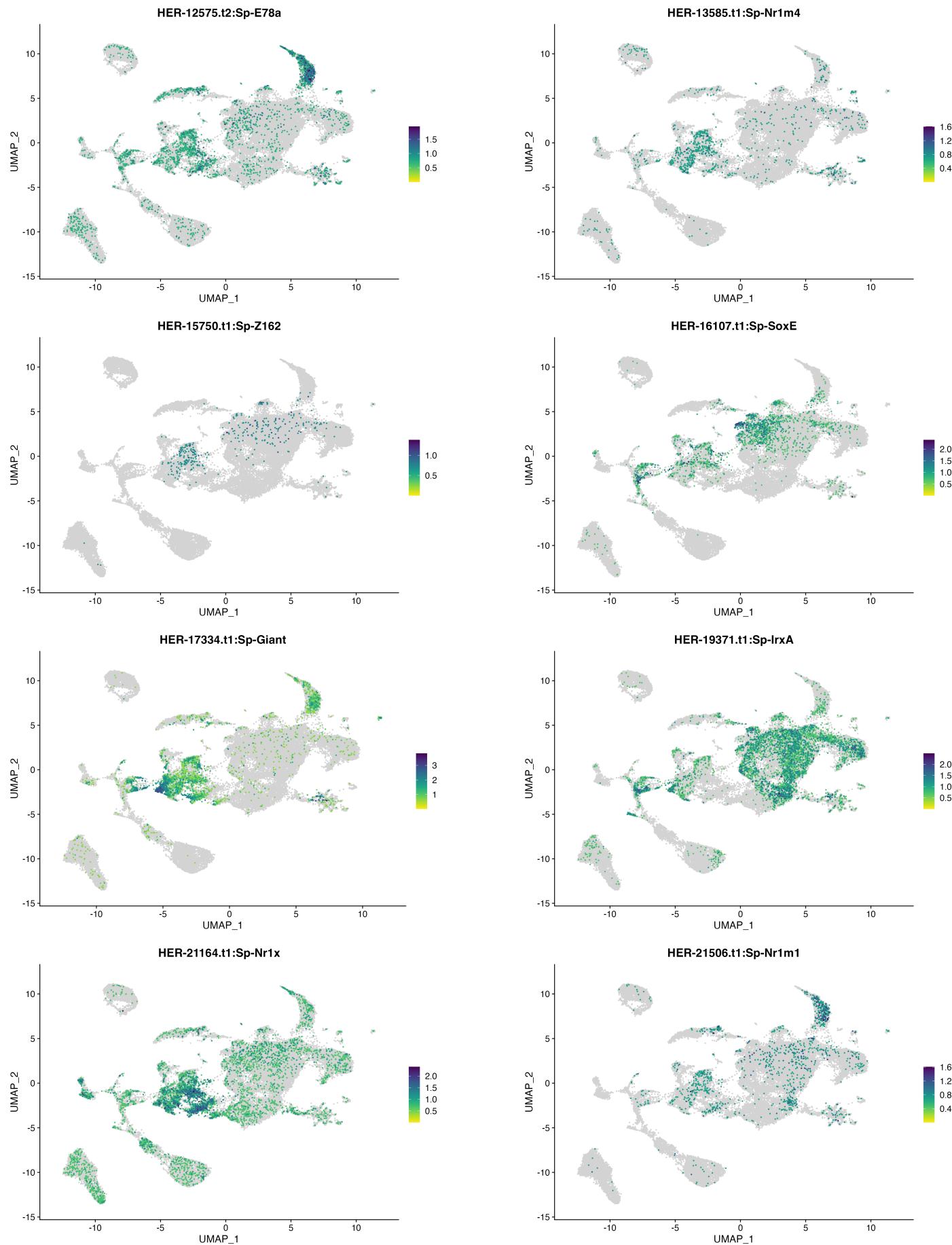
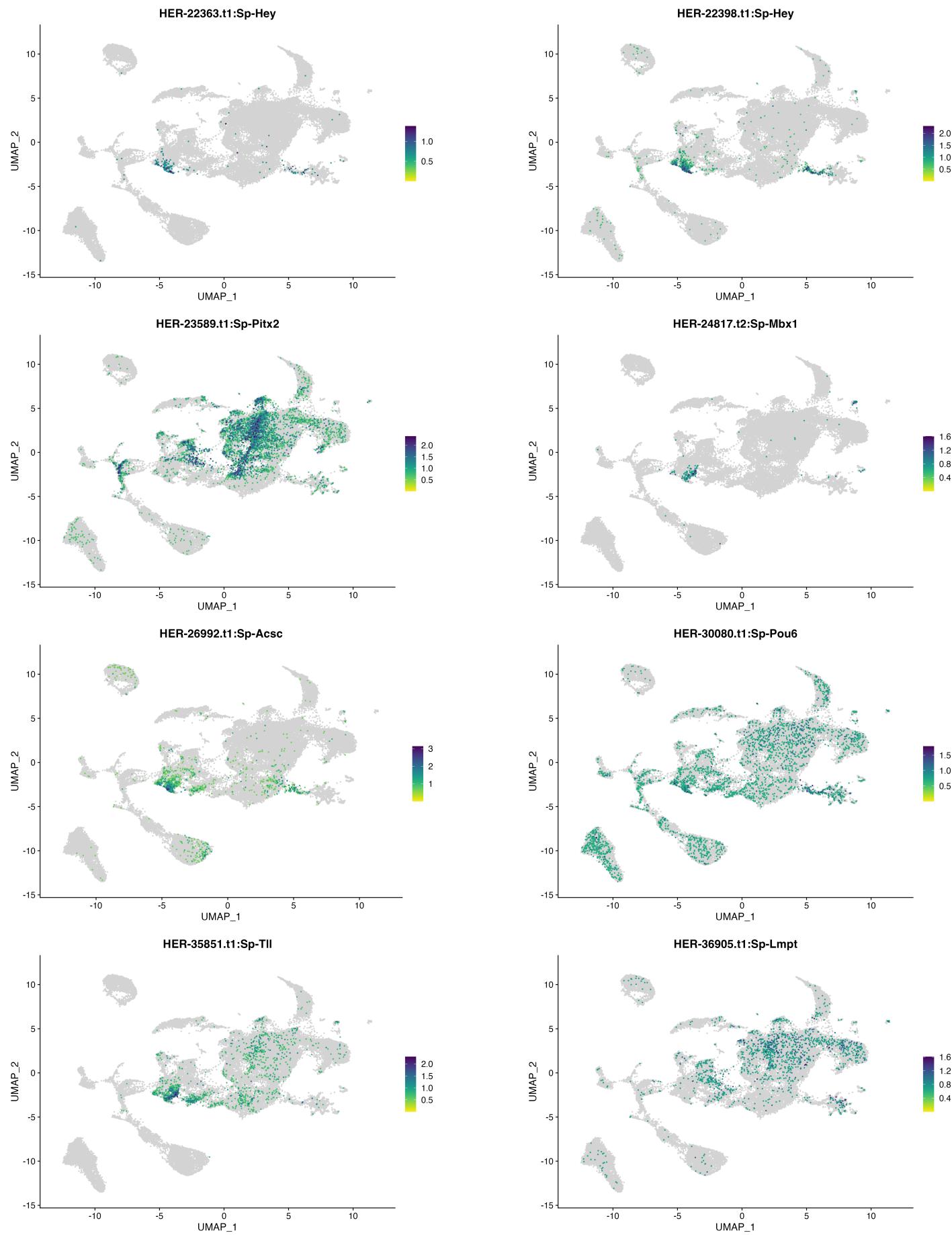
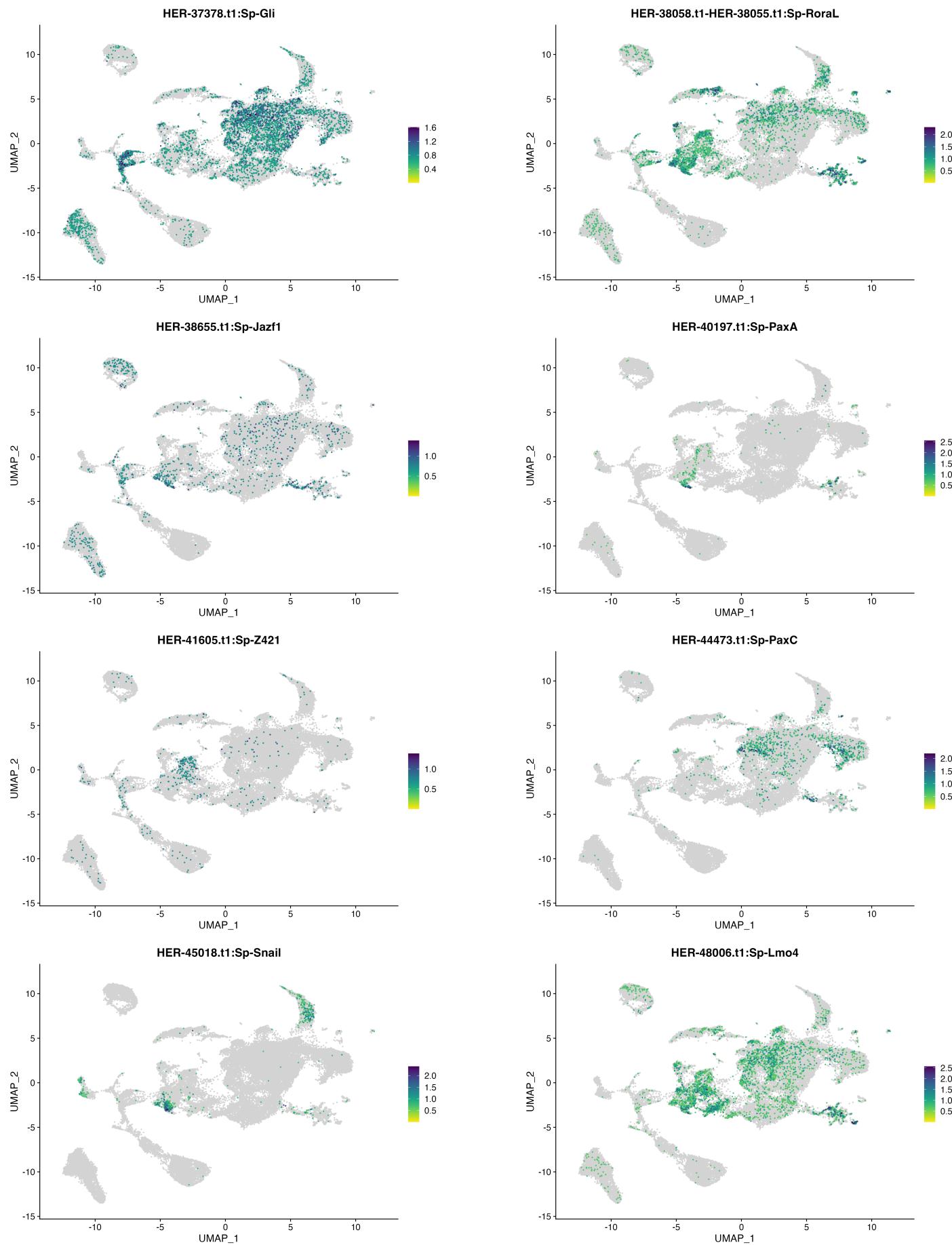


Fig. S5. Dot plot of neural cell type marker gene expression patterns. ~3-6 enriched marker genes (see Table S2) for each neural cell type in the scRNA-seq atlas were selected, and the average expression of the genes in each cell type was plotted.









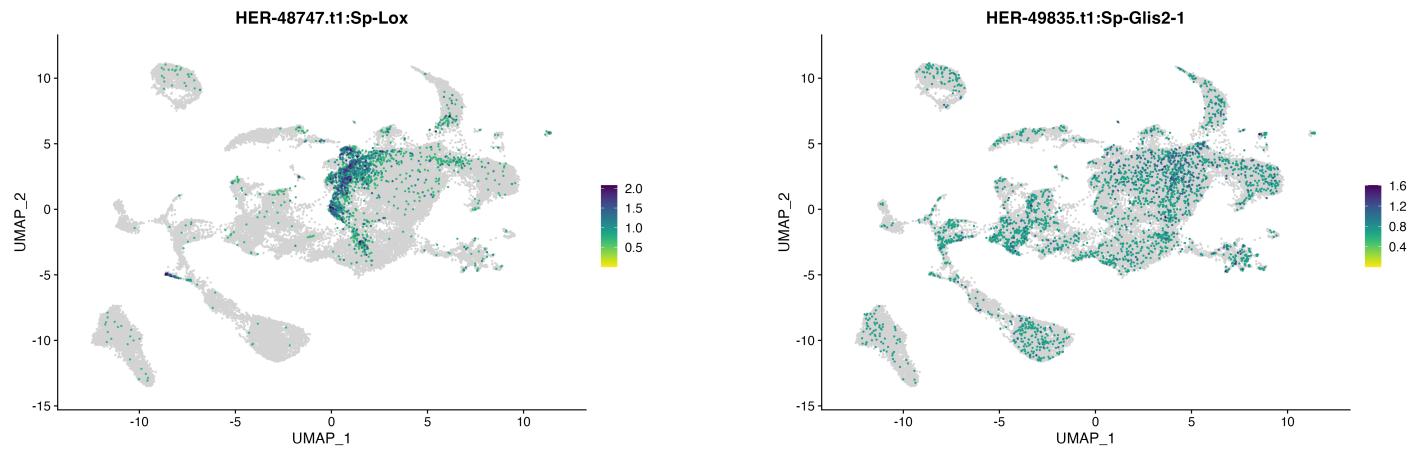


Fig. S6. Expression patterns of the candidate transcription factors for adult rudiment development.

Each UMAP shows the scRNA-seq expression pattern of one of the 33 candidate transcription factors that was shown to be enriched in adult rudiment tissues (rudiment ectoderm and posterior ectoderm).

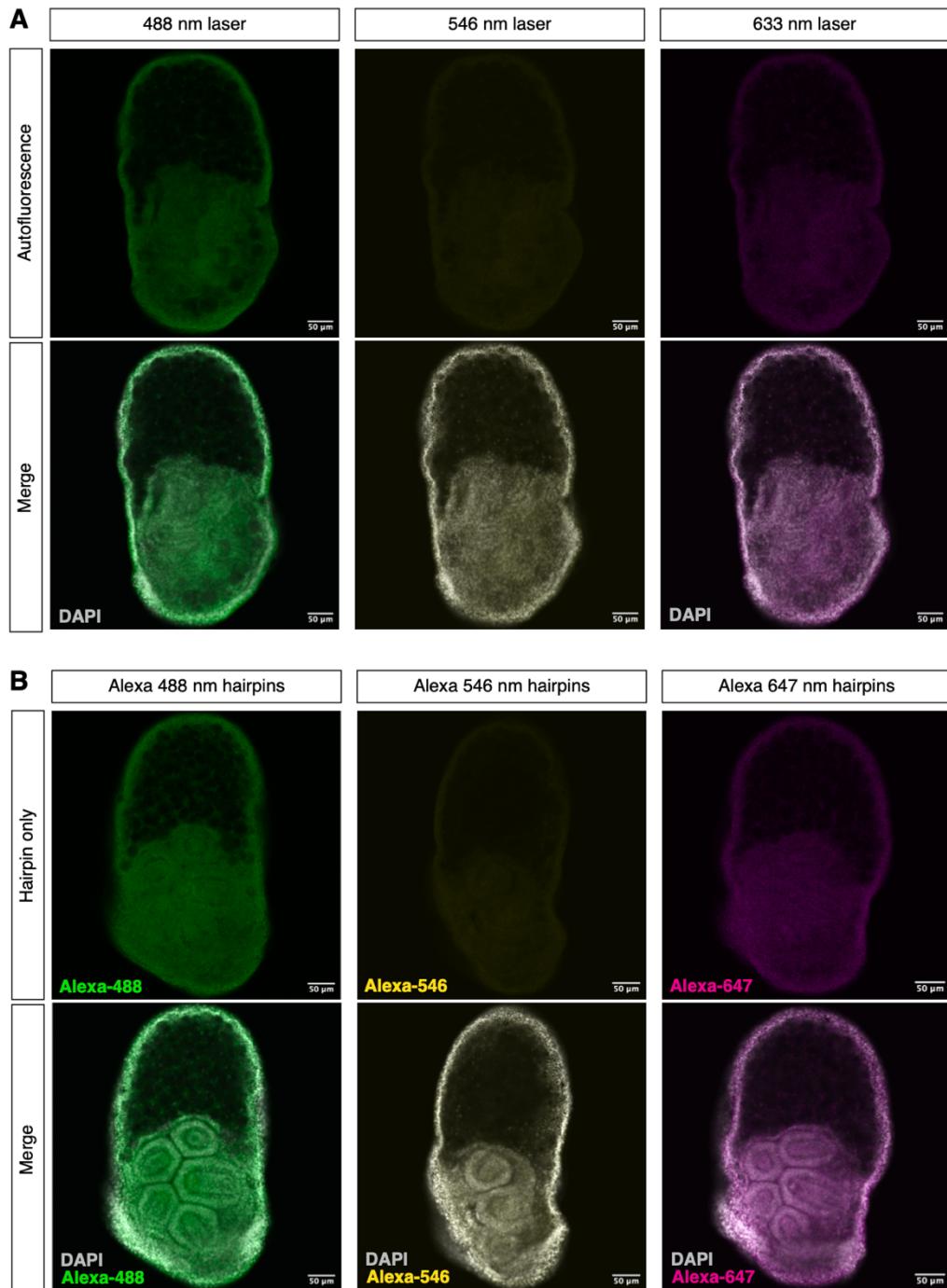


Fig. S7. Control experiments to verify HCR specificity. (A) HCR micrographs of 53 hpf *He* larvae that were incubated with no probes and no hairpins. (B) HCR micrographs of 53 hpf *He* larvae that were incubated with no probes. Neither condition resulted in localized fluorescent patterns, confirming the specificity of the probes used in this study.

Table S1. Cell type marker genes and references for the full scRNA-seq atlas

Available for download at

<https://journals.biologists.com/dev/article-lookup/doi/10.1242/dev.203015#supplementary-data>

Table S2. Neural cell type marker genes and references for the neural-only scRNA-seq atlas

Available for download at

<https://journals.biologists.com/dev/article-lookup/doi/10.1242/dev.203015#supplementary-data>

Table S3. List of putative transcriptional regulators of adult rudiment development

Available for download at

<https://journals.biologists.com/dev/article-lookup/doi/10.1242/dev.203015#supplementary-data>

Table S4. Probe sequences used in HCR experiments

Available for download at

<https://journals.biologists.com/dev/article-lookup/doi/10.1242/dev.203015#supplementary-data>

- Wei, Z., Yaguchi, J., Yaguchi, S., Angerer, R. C. and Angerer, L. M. (2009). The sea urchin animal pole domain is a Six3-dependent neurogenic patterning center. *Development*, 136, 7, 1179-1189. <https://doi.org/10.1242/dev.032300>
- Wessel, G. M., Zaydfudim, V., Hsu, Y. T. J., Laidlaw, M. and Brooks, J. M. (2000). Direct molecular interaction of a conserved yolk granule protein in sea urchins. *Dev Growth Differ*, 42, 5, 507-517. <https://doi.org/10.1046/j.1440-169x.2000.00534.x>
- Yaguchi, J., Angerer, L. M., Inaba, K. and Yaguchi, S. (2012). Zinc finger homeobox is required for the differentiation of serotonergic neurons in the sea urchin embryo. *Developmental Biology*, 363, 1, 74-83. <https://doi.org/10.1016/j.ydbio.2011.12.024>
- Yaguchi, J. and Yaguchi, S. (2021). Sea urchin larvae utilize light for regulating the pyloric opening. *Bmc Biology*, 19, 1. <https://doi.org/10.1186/s12915-021-00999-1>
- Yaguchi, S. and Katow, H. (2003). Expression of tryptophan 5-hydroxylase gene during sea urchin neurogenesis and role of serotonergic nervous system in larval behavior. *J Comp Neurol*, 466, 2, 219-229. <https://doi.org/10.1002/cne.10865>
- Yaguchi, S., Taniguchi, Y., Suzuki, H., Kamata, M. and Yaguchi, J. (2022). Planktonic sea urchin larvae change their swimming direction in response to strong photoirradiation. *Plos Genet*, 18, 2. <https://doi.org/10.1371/journal.pgen.1010033>
- Yaguchi, S., Yaguchi, J., Angerer, R. C. and Angerer, L. M. (2008). A Wnt-FoxQ2-nodal pathway links primary and secondary axis specification in sea urchin embryos. *Dev Cell*, 14, 1, 97-107. <https://doi.org/10.1016/j.devcel.2007.10.012>