

Supplementary Materials for  
**Stem cell–homing biomimetic hydrogel promotes the repair of osteoporotic  
bone defects through osteogenic and angiogenic coupling**

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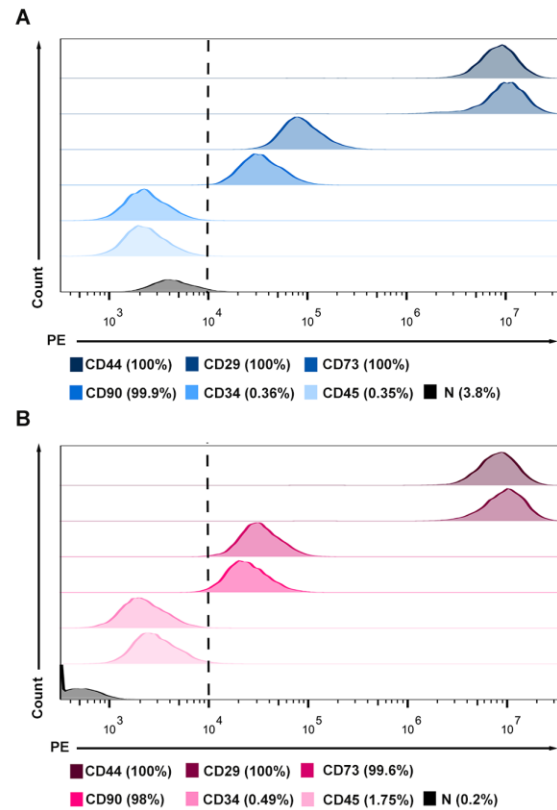
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*Sci. Adv.* **10**, eadq6700 (2024)  
DOI: 10.1126/sciadv.adq6700

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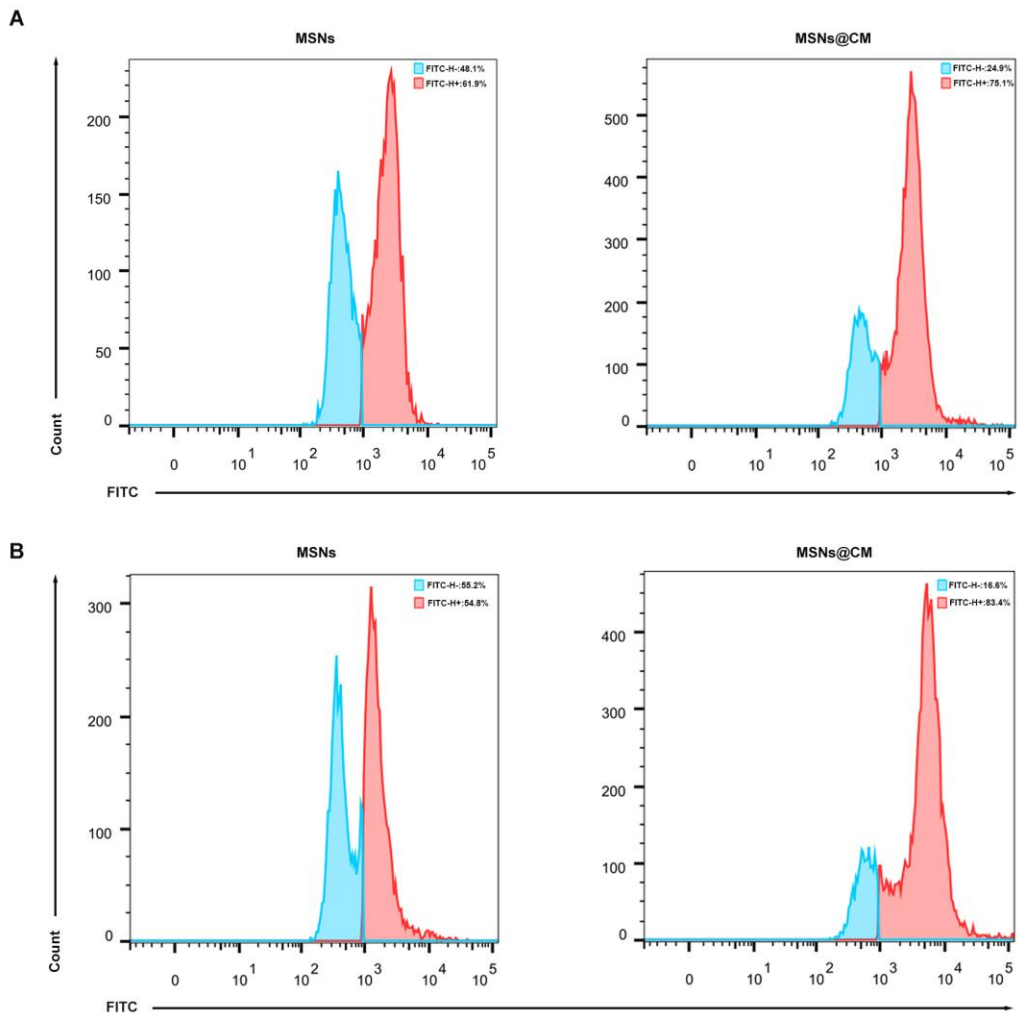
Figs. S1 to S7  
Tables S1 and S2

## Supplementary Information



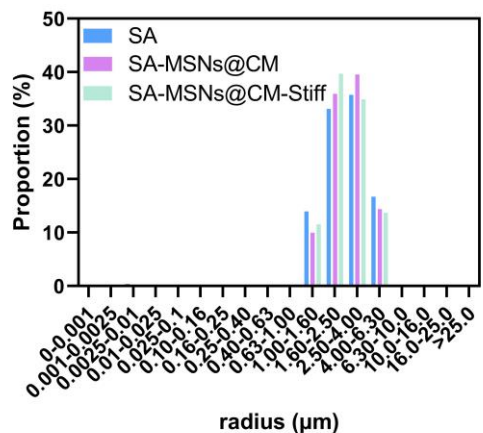
**Fig. S1. Identification of BMSCs from different groups by flow cytometry.**

(A) Flow cytometry showed BMSCs from osteoporotic rats were positive for CD44 (100%), CD29 (100%), CD73 (100%), and CD90 (99.9%), negative for CD34 (0.36%) and CD45 (0.35%). (B) Flow cytometry showed BMSCs from osteoporotic rats were positive for CD44 (100%), CD29 (100%), CD73 (99.6%), and CD90 (98%), negative for CD34 (0.49%) and CD45 (1.75%).  $n = 3$  for each group.

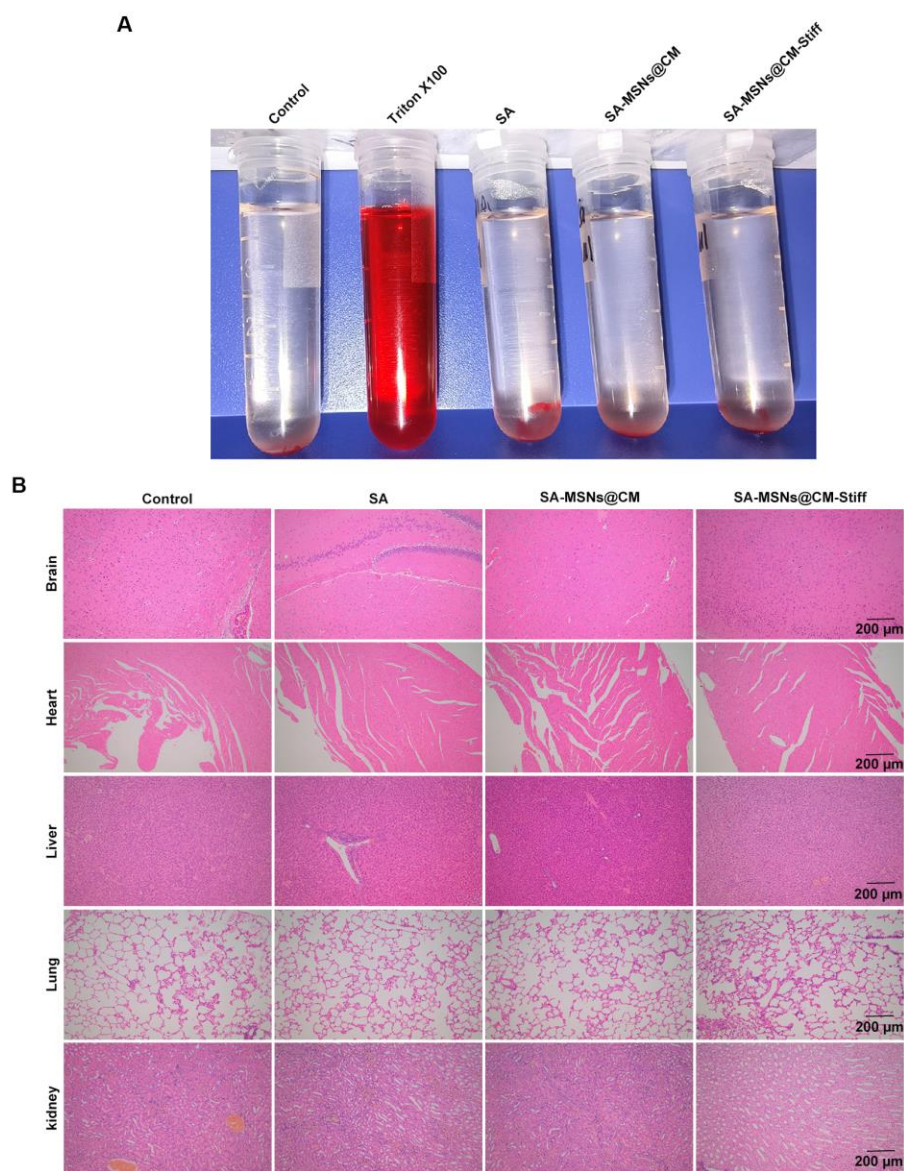


**Fig. S2. Analyzing the impact of cell membrane coverage on MSNs.**

(A) Flow cytometric analysis of MSNs (coated by BMSCs and HUVECs membranes) uptake by BMSCs. (B) Flow cytometric analysis of MSNs (coated by BMSCs and HUVECs membranes) uptake by HUVECs.

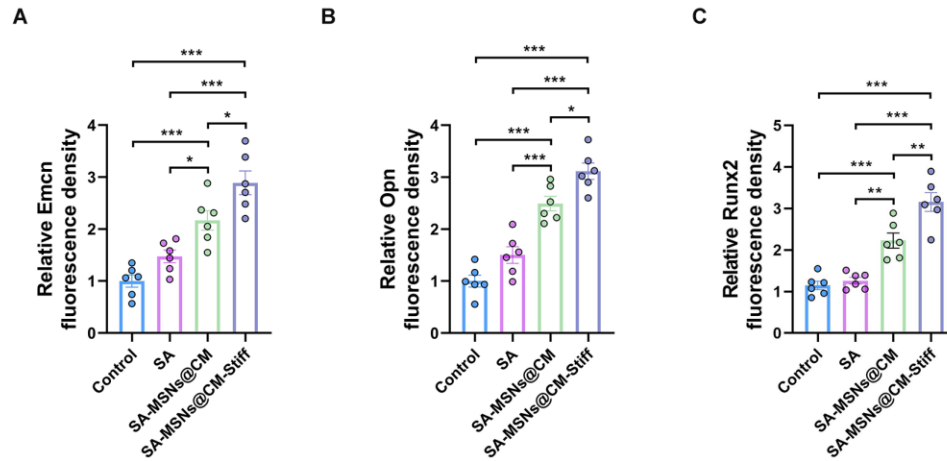


**Fig. S3. Pore size distributions of SA, SA-MSNs@CM and SA-MSNs@CM-Stiff.** The pore size distribution of SA, SA-MSNs@CM and SA-MSNs@CM-Stiff was measured by low-frequency magnetic resonance.



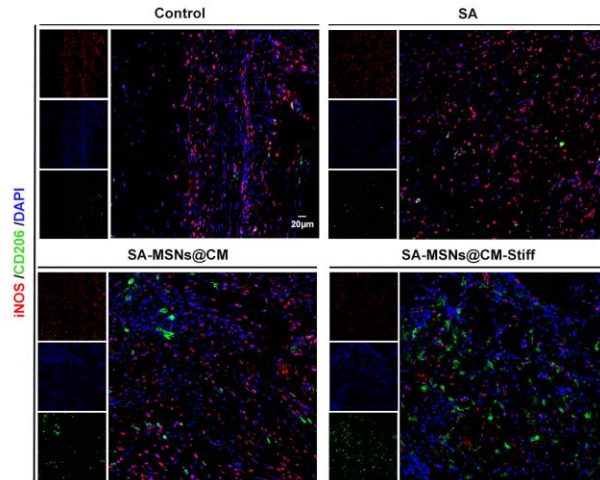
**Fig. S4. Hydrogels biocompatibility testing.**

(A) Erythrocyte haemolysis assay to evaluate the biocompatibility of the hydrogel. (B) Hematoxylin and eosin staining and histological analysis of main organs from hydrogels-treated rats. n = 3 for each group.

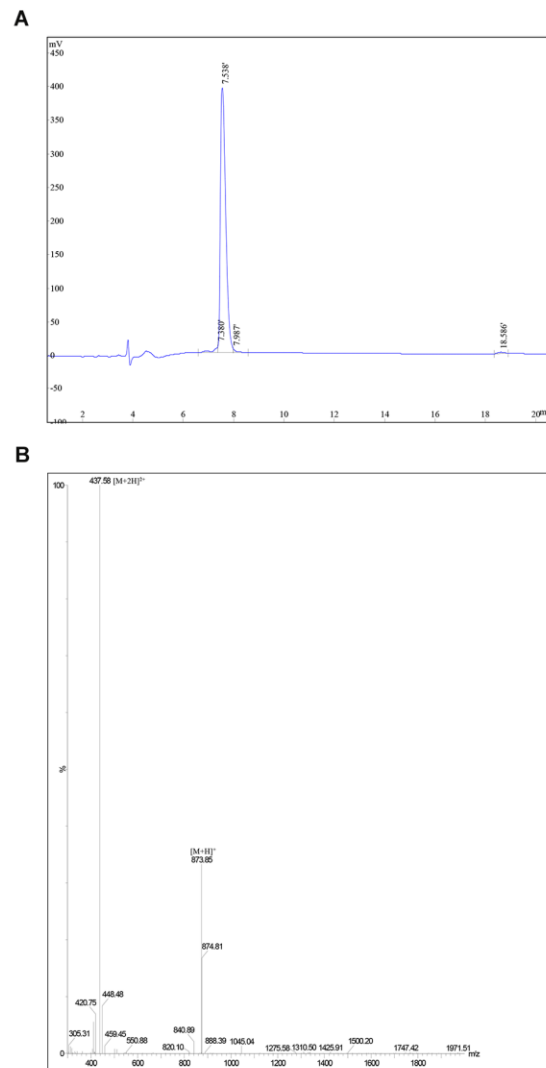


**Fig. S5. SA-MSNs@CM-Stiff induces BMSCs recruitment and promotes osseointegration *in vivo*.**

(A) Relative Emcn fluorescence density (Statistical analysis for Figures 8G). (B) Relative Opn fluorescence density (Statistical analysis for Figures 8H). (C) Relative Runx2 fluorescence density (Statistical analysis for Figures 8I).  $n = 6$  for each group. Error bars denote mean  $\pm$  SEM; ns, no significance;  $*p < 0.05$ ,  $**p < 0.01$ , and  $***p < 0.001$ .



**Fig. S6. SA-MSNs@CM-Stiff regulates macrophage polarization *in vivo*.** Representative immunofluorescence staining of iNOS and CD206 in the femoral condyle of an osteoporotic rat eight weeks after hydrogel implantation (scale bar: 20 μm); n = 6.



**Fig. S7. Analytical HPLC trace and electrospray ionization mass spectrometry (ESI-MS) analysis of bone marrow-homing peptide.**

(A) Analytical HPLC trace of bone marrow-homing peptide. (B) ESI-MS analysis of bone marrow-homing peptide.



**Table S1. Primary antibodies information for western blotting.**

<b>Antibody</b>	<b>Host</b>	<b>Dilution ratio</b>	<b>Company</b>	<b>Item number</b>
anti-Nanog	Rabbit	1:1000	ProteinTech	14295-1-AP
anti-Sox2	Rabbit	1:1000	ProteinTech	11064-1-AP
anti-Oct4	Rabbit	1:1000	ProteinTech	11263-1-AP
anti-P21	Rabbit	1:1000	ProteinTech	10355-1-AP
anti-P53	Rabbit	1:1000	ProteinTech	10442-1-AP
anti-P16	Rabbit	1:600	ProteinTech	10883-1-AP
anti-eNOS	Rabbit	1:1000	ABclonal	A20985
anti-p-eNOS	Rabbit	1:1000	ProteinTech	28939-1-AP
anti-sGC	Rabbit	1:1000	ProteinTech	11936-1-AP
anti-PKG	Rabbit	1:1000	ProteinTech	21646-1-AP
anti-Runx2	Rabbit	1:1000	Abways Technology	CY5864
anti-Opn	Rabbit	1:1000	ProteinTech	25715-1-AP
anti-CD31	Mouse	1:1000	ProteinTech	66065-2-Ig
anti-Emcn	Rabbit	1:1000	Invitrogen	PA5-115178
anti-CD86	Rabbit	1:1000	ProteinTech	13395-1-AP
anti-CD206	Rabbit	1:1000	ProteinTech	18704-1-AP
anti-Gapdh	Rabbit	1:1000	ProteinTech	60004-1-Ig

**Table S2. Primary antibodies information for Immunofluorescence.**

Antibody	Host	Dilution ratio	Company.	Item number
anti- $\gamma$ -H2AX	Rabbit	1:200	Abcam	ab81299
anti-Runx2	Rabbit	1:200	Abways Technology	CY5864
anti-Opn	Rabbit	1:200	ProteinTech	25715-1-AP
anti-CD90	Mouse	1:200	Invitrogen	14-0900-81
anti-CD31	Mouse	1:1000	ProteinTech	66065-2-Ig
anti-Emcn	Rabbit	1:1000	Invitrogen	PA5-115178
anti-CD68	Mouse	1:200	ProteinTech	66231-2-Ig
anti-CD86	Rabbit	1:200	ProteinTech	13395-1-AP
anti-CD206	Rabbit	1:200	ProteinTech	18704-1-AP
anti-iNOS	Mouse	1:50	Invitrogen	MA5-47726