

Supplementary Information for

**Structures of the ribosome bound to EF-Tu–isoleucine
tRNA elucidate the mechanism of AUG avoidance**

Mariia Yu. Rybak¹ and Matthieu G. Gagnon^{1,2,3,4,*}

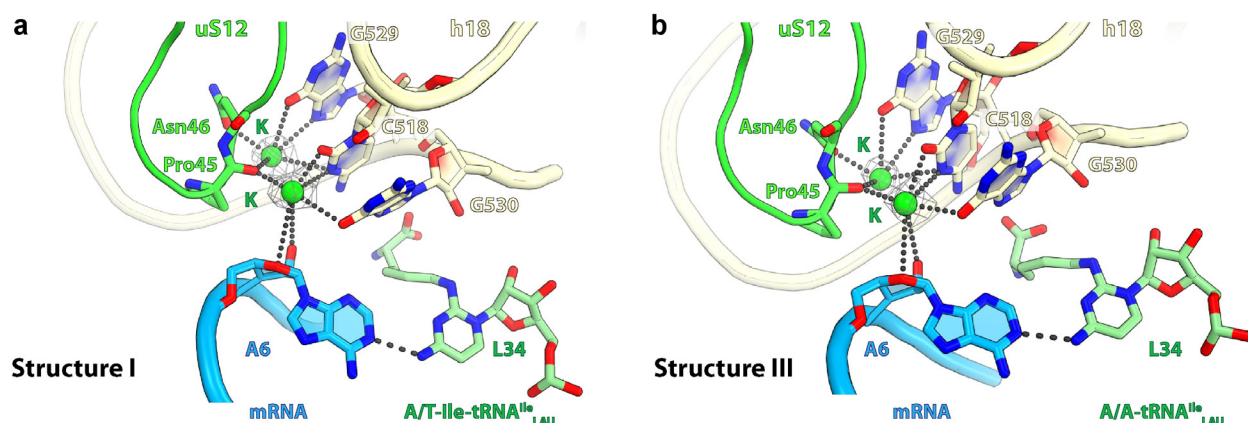
¹ Department of Microbiology and Immunology, University of Texas Medical Branch,
Galveston, Texas 77555, USA.

² Department of Biochemistry and Molecular Biology, University of Texas Medical Branch,
Galveston, Texas 77555, USA.

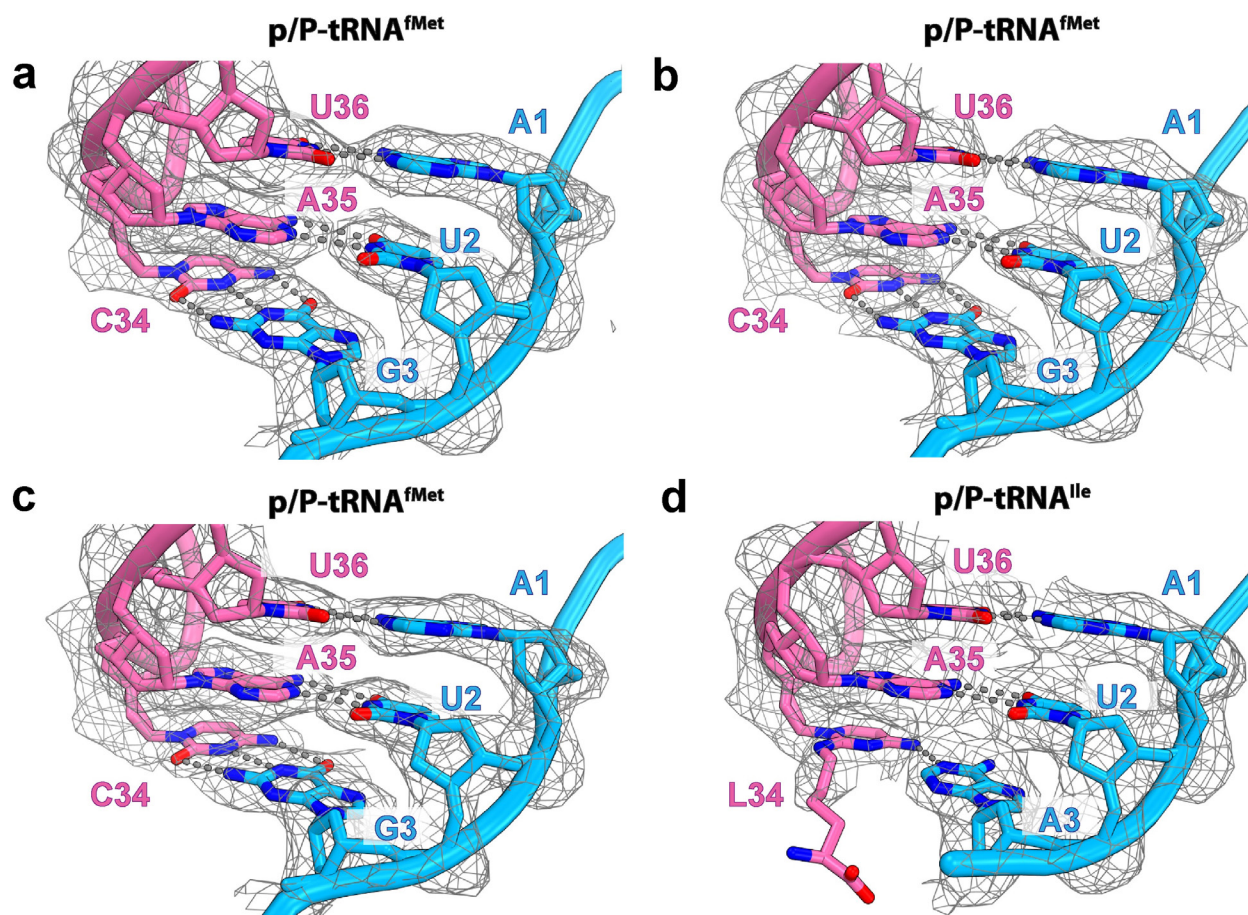
³ Sealy Center for Structural Biology and Molecular Biophysics, University of Texas Medical
Branch, Galveston, Texas 77555, USA.

⁴ Institute for Human Infections and Immunity, University of Texas Medical Branch, Galveston,
Texas 77555, USA.

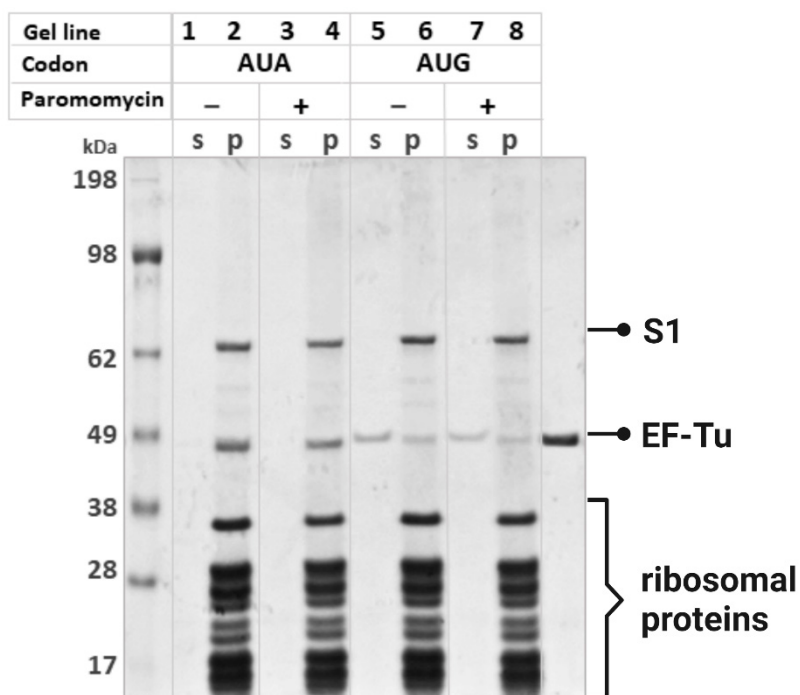
*email: magagnon@utmb.edu



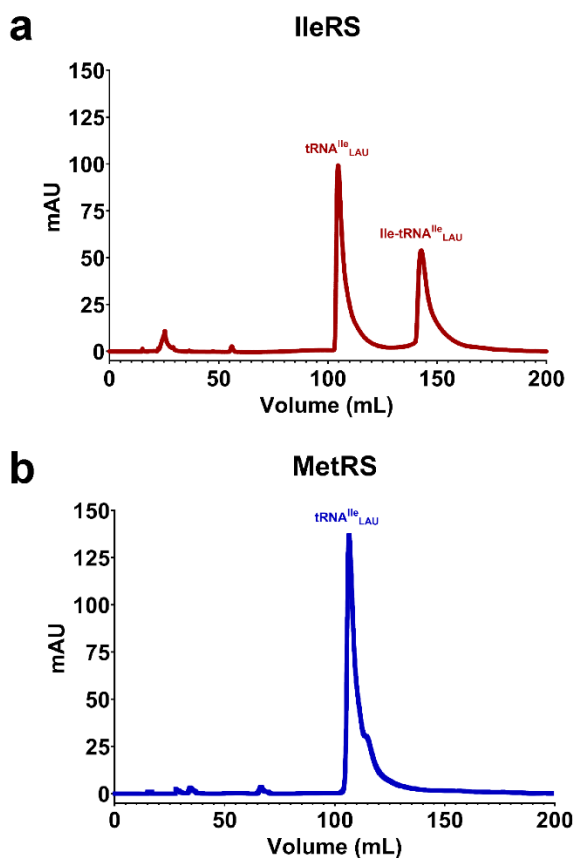
Supplementary Fig. 1. Potassium ions in the ribosome decoding center. Stabilization of the decoding center by potassium ions (green spheres) upon binding of the A-site tRNA^{Ile}_{LAU} in structures I (a) and III (b), as previously observed in an elongation ribosome complex¹. The stabilization of the third nucleotide of the A-site codon (A6) by potassium in both structures suggests that the ASL of the tRNA^{Ile}_{LAU} is fully accommodated into the A site. The Coulomb potential density of the K⁺ ions (gray mesh) is contoured at 2.9σ. The gray dashed lines indicate putative hydrogen bonds.



Supplementary Fig. 2. Interactions between the P-site tRNA and mRNA in structures I-IV. In structures I (a), II (b) and III (c), initiator tRNA^{fMet} is bound to the P-site AUG codon, while in structure IV (d), tRNA^{Ile}_{LAU} is bound to the P-site AUA codon. The Coulomb potential density (gray mesh) is contoured at 2.9σ . Putative hydrogen bonds are indicated by the gray dashed lines.



Supplementary Fig. 3. Co-sedimentation assay assessing the binding of EF-Tu•Ile-tRNA^{Ile}_{LAU} to A-site AUA or AUG programmed 70S ribosome complexes. Binding of the *E. coli* ternary complex of EF-Tu, guanosine 5'- β , γ -methylenetriphosphate, and Ile-tRNA^{Ile}_{LAU} to the 70S ribosome programmed with the cognate AUA (lanes 1-4) or near-cognate AUG (lanes 5-8) codon in the A site, analyzed by SDS-PAGE after centrifugation through a sucrose cushion. Gel lanes labeled (s) and (p) are proteins in the supernatant and pellet fractions, respectively. EF-Tu•GDP•Ile-tRNA^{Ile}_{LAU} binds to the 70S ribosome with the AUA codon programmed in the A site similarly in the absence (lane 2) or presence (lane 4) of paromomycin. EF-Tu•GDP•Ile-tRNA^{Ile}_{LAU} binds to the 70S ribosome programmed with the near-cognate AUG codon in A site with low apparent affinity with or without paromomycin (lanes 6 and 8). The results are representative of three independent experiments.



Supplementary Fig. 4. Aminoacylation reactions of tRNA^{Ile}_{LAU} analyzed by reverse-phase HPLC. Aminoacylation of tRNA^{Ile}_{LAU} by *E. coli* IleRS (**a**) or *E. coli* MetRS (**b**). The purified tRNA^{Ile}_{LAU} is only aminoacylated by IleRS and does not show any peak for aminoacylated-tRNA^{Ile} following incubation with MetRS, confirming that lysidinylated-tRNA^{Ile} is recognized exclusively by IleRS, as reported².

Supplementary References

1. Rozov, A. et al. Importance of potassium ions for ribosome structure and function revealed by long-wavelength X-ray diffraction. *Nat. Commun.* **10**, 2519 (2019).
2. Nakanishi, K. et al. Structural basis for translational fidelity ensured by transfer RNA lysidine synthetase. *Nature* **461**, 1144-1148 (2009).