A novel immunomodulating peptide with potential to complement oligodeoxynucleotide-mediated adjuvanticity in vaccination strategies

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Supplementary information

Figure S1: Refers to manuscript **Figure 1a and 1b** CD14⁺/CD14^{neg} (Monocyte-derived Dendritic Cells)



Debris and multicellular events were excluded, then viable (live) cells gated and CD11c⁺ cells identified. Within the CD11c⁺ population, CD14⁺ and CD14^{neg} cell populations were identified.

Figure S2: Refers to manuscript **Figure 1c and 1d** CD14⁺/CD14^{neg} (Monocyte-derived Dendritic Cells)



Debris and multicellular events were excluded, then viable (live) cells gated and CD11c⁺ cells identified. Within the CD11c⁺ population, CD14⁺ and CD14^{neg} cell populations were identified.

Figure S3: Refers to manuscript **Figure 1e** Ki67 (Monocyte-derived Dendritic Cells)



Figure S4: Refers to manuscript **Figure 2c and 2d** Ki67 (Unstimulated human PBMC cultures: CD4⁺ T cells)



Debris and multicellular events were excluded, then viable (live) cells gated. CD4⁺ cells selected for analysis of Ki67. L-15800 and D-15800 treatment groups were cultured across different plates, with vehicle control groups included on each plate.





Debris and multicellular events were excluded, then viable (live) cells gated. CD4⁺ cells selected for analysis of CD25. L-15800 and D-15800 treatment groups were cultured across different plates, with vehicle control groups included on each plate.

Figure S6: Refers to manuscript **Figure 2g – 2j** Ki67 and CD25 (Unstimulated human PBMC cultures: CD3^{neg} CD56⁺ cells)



Debris and multicellular events were excluded, then viable (live) cells gated. $CD3^{neg}CD56^+$ cells selected for analysis of Ki67 and CD25.

Figure S7: Refers to manuscript **Figure 2k and 2l** CD40L (Unstimulated human PBMC cultures: CD4⁺ T cells)



Debris and multicellular events were excluded, then viable (live) cells gated and CD4⁺ cells selected for analysis of CD40L. L-15800 and D-15800 treatment groups were cultured across different plates, with vehicle control groups included on each plate.

Figure S8: Refers to manuscript Figure 2m

CD40L (Stimulated human PBMC cultures: CD4⁺ T cells)



Debris and multicellular events were excluded, then viable (live) cells gated. CD4⁺ T cells selected for analysis of CD40L. L-15800 and D-15800 treatment groups were cultured across different plates, with vehicle control groups included on each plate.

Figure S9: Refers to manuscript Figure 2n

CD40L (Unstimulated human PBMC cultures: CD3^{neg} CD56⁺ cells)



Debris and multicellular events were excluded, then viable (live) cells gated and CD3^{neg}CD56⁺ cells selected for analysis of CD40L. L-15800 and D-15800 treatment groups were cultured across different plates, with vehicle control groups included on each plate.

Figure S10: Refers to manuscript Figure 3b Cell viability (unstimulated PBMCs)



Debris and multicellular events were excluded, then viable (live) cells gated.





Debris and multicellular events were excluded, viable (live) cells selected and CD19⁺ cells identified for analysis of IgM expression.

Figure S12 – Refers to manuscript Figure 5a and 5b:

Elution profile of 15800 peptide isomers and sera from a Phenomenex[™] Jupiter 5 µm, C4, 300Å column



The percentage and rate of degradation of the L-15800 and D-15800 peptide isomers in both human serum (HS) and foetal bovine serum (FBS) at 37°C over 48 hours were determined by area under the curve (AUC) analysis of the integrated peak area observed at T=0 with those observed at the T=1, 4, 24 & 48 h time points.

i, Elution profile of L- / D- 15800 peptide isomers. **ii**, Elution profile of FBS. **iii**, Elution profile of L- / D- 15800 peptide isomers combined with FBS. **iv**, Elution profile of HS. **v**, Elution profile of L- / D- 15800 peptide isomers combined with HS.