





^{·····}S R Y P V S S I E A A R I ·····





Supplementary Figure 1. a Scatter plots for TR of genes in low and high mTOR in *Tsc1*^{-/-} MEFs. $Tsc1^{-/-} + mock$ (high mTOR) vs $Tsc1^{-/-} + Torin 1$ (100 nM, 24 hours; low mTOR) were compared. b Additional examples of RNA-Seq read alignments for genes showing enriched intronic APA events in high mTOR Tsc1-- MEFs. The alignments were color-coded as indicated in the figure. The yellow box highlights the regions with the intronic APA event. c and d Real-time quantitative PCR (RT-qPCR) validation of genes showing dynamic intronic APA events upon the changes of mTOR signaling in cells. RNAi knockdown of mTOR kinase was conducted using Tsc1-/- MEFs and the changes of TR for selected genes were tested for intronic APA events. The selection of genes was based on the enriched intronic APA events in *Tsc1*^{-/-} or WT MEFs. The Y-axis scale is shown in the log scale. Four technical repeats were conducted, and students' t-tests were performed for statistical analysis. The data are presented as the mean (SD). e Overlap of intronic APA genes across the investigated breast cancer cell lines. Overlapping intronic APA events among three breast cancer cell lines with mock or Torin 1 treatment were analyzed. f Enriched KEGG pathways by intronic APA events in three breast cancer cell lines. Enrichment was analyzed by the comparison between mock and Torin 1 treated cells. g A schematic presentation of intronic APA and the formation of new ORFs based on intron regions. h A SRM workflow for the identification of new peptide sequences from intron regions. i Peptide sequences that are produced from intron regions due to intronic APA. Peptides were identified and verified by SRM.





COAD (Colon Adenocarcinoma) HNSC (Head-Neck Squamous Cell Carcinoma)









Supplementary Figure 2. a Collection of tumor samples and normal tissues from 10 types of cancer in the TCGA data. b Differential expression analyses for genes with annotated intronic APA events in normal tissues and tumor samples. TCGA-COAD (Colon Adenocarcinoma), TCGA-HNSC (Head and Neck Squamous Cell Carcinomas), TCGA-LUSC (Lung Squamous Cell Carcinoma), TCGA-LIHC (Liver Hepatocellular Carcinoma), TCGA-STAD (Stomach adenocarcinoma), TCGA-PRAD (Prostate Adenocarcinoma), and TCGA-THCA (Thyroid carcinoma) data analyses are shown. The x-axis presents the significance of intronic APA events calculated by -log₁₀(*p*-value). *p*-values were determined by students' t-test. The y-axis shows the fold changes of gene expression in tumors over normal samples. Red dots indicate the genes showing significant intronic APA events conserved in 80% or more of tumor samples (i.e., 80% or more of tumor samples have higher TRs than the mean TR of normal tissue samples). Blue dots indicate the genes showing significant APA events that 80% or more of tumor samples have lower TRs than the mean TR of normal tissue samples. c Exemplary RNA-Seq read alignments from BRCA data for H2AZ2 and LRRFIP1 genes. d Exemplary RNA-Seq read alignments from BRCA data for CXCL12 and DST genes. e Scatter plots for intronic APA events in COAD, HNSC, LUSC, LIHC, STAD, PRAD, and THCA. The TR mean for genes with significant intronic APA events is color-coded. Genes showing intronic APA events in more than 88% of samples are color-coded as blue (normal) or red (tumor). Genes with intronic APA events in 80-88% of samples are shown in cyan (normal) or orange (tumor). f A heatmap for the KEGG pathways that are enriched by intronic APA events. The KEGG pathways that are unique to each cancer type are displayed. The color scale represents $-\log_{10}(p-value)$ for pathways enriched in tumor and $\log_{10}(p-value)$ for pathways enriched in normal: red-colored KEGG pathways are enriched in tumor samples and blue-colored KEGG pathways are enriched in normal tissues.





Supplementary Figure 3. a Examples of intronic APA events that are unique to each cancer type.b Schematic presentation of Pfam domain swapping in tumors and normal tissues by intronic APA events. The WD-40 domain containing proteins are shown as an example.



Supplementary Figure 4. a The boxplots show four examples that intronic APA events are correlated with hormone receptor phenotypes but not the corresponding gene expression levels in BRCA data. The *p*-values were determined by unpaired t-test. *NGEF* TR *p*=1.25e-10, Gene expression (GE) *p*=0.603; *NGEF* TR *p*=6.60e-8, Gene expression (GE) *p*=0.747; *HTATIP2* TR *p*=1.95e-6, GE *p*=0.964; *GTF2IRD2* TR *p*=6.90e-12, GE *p*=0.540; *TVP23C* TR *p*=3.61e-6, GE *p*=0.721; *FAM120C* TR *p*=2.09e-15, GE *p*=0.536; *ICAM3* TR *p*=9.46e-11, GE *p*=0.802; *RIMKLB* TR *p*=3.36e-10, GE *p*=0.640. **b** Violin plots illustrate multiple exemplary genes demonstrating significant intronic APA events but not significant differential gene expression in cancer stages. **c** Kaplan-Meier (KM) plots illustrate the correlation between the TR of selected genes (*VRK3*, *EPB41L5* and *PDLIM7*) and the disease-free rate or survival rate of cancer patients in STAD, LIHC and KIRC.



Supplementary Figure 5. a The distribution of relative positions of the 3'-last exon of intronic APA transcripts. The graph was generated using all annotated intronic APA transcripts and their corresponding full-length transcripts registered in the current genome annotation. **b** Histogram displays for the distribution of intronic APA position in pan-cancer data.