Expanded View Figures

Figure EV1. Single-cell profiling of the developing and postnatal liver reveals a cell population shift from embryonic erythropoiesis to postnatal immune surveillance in Jag1^{*/+}, Jag1^{Ndt/+}, and Jag1^{Ndt/Hdr} mice.

(A) Violin plots of read counts (nCount_RNA), unique gene counts (nFeature_RNA), and percentage of mitochondrial mRNA (percent-mt) across the cell types identified in the $Jag1^{H/r/+}$, $Jag1^{Ndr/+}$, and $Jag1^{Ndr/Ndr}$ livers at E16.5 and P3 (n = 2 each). (B) Genotype, stage, or organ contribution to the composition of the scRNAseq dataset of E16.5 and P3 $Jag1^{+/+}$, $Jag1^{Ndr/+}$, and $Jag1^{Ndr/Ndr}$ mice. (C) Dot plot of the SCT-normalized mRNA marker expression of 78 DEGs characteristic for the individual cell types identified in (B). (D) Bar graph representing the relative proportion of parenchymal and Kupffer cells (clusters 1-6) versus non-parenchymal cells (clusters 7-20) in dissociated $Jag1^{+/+}$, $Jag1^{Ndr/+}$, and $Jag1^{Ndr/Ndr}$ livers at E16.5 and P3 (n = 2 each). Graph represents mean ± SD. (E) Graph depicting relative cell type contribution in dissociated $Jag1^{+/+}$, $Jag1^{Ndr/+}$, and $Jag1^{Ndr/Ndr}$ livers at E16.5 and P3 (n = 2 each). Graph cell count.





Figure EV2. Altered gene expression profile of hepatocytes and whole livers in Jag1^{Ndr/Ndr} animals.

(A) Box and whiskers plot of the percentage mitochondrial mRNA (percent.mt), unique gene counts (nFeature_RNA), and read counts (nCount_RNA) across the hepatocyte-like cell types identified in the $Jag1^{+/+}$, $Jag1^{Ndr/+}$, and $Jag1^{Ndr/+dr}$ livers by scRNA seq at E16.5 and P3 (n = 2 each). Center line indicates Median (50th percentile). The lower and upper bound of box indicate Q1, and Q3 (25th and 75th percentiles), respectively. Lower and upper whisker indicate min and max values within 1.5 * IQR below Q1 and above Q3, respectively. (B) Heatmap of the top 8 mRNA markers for each cluster. (C) PCA distribution of the bulk RNAseq samples from Andersson et al, 2018, re-analyzed in Fig. 2 and Fig. EV2. (D) Volcano plot of the DEGs of $Jag1^{Ndr/Ndr}$ vs. $Jag1^{+/+}$ liver at P10. Pro-inflammatory markers of activated hepatocytes are highlighted. Significance was confirmed via Benjamini-Hochberg *P* value correction. (E) Top29 GO pathways enriched in the $Jag1^{Ndr/Ndr}$ livers, NES (normalized enrichment score) indicates overlap of genes across the pathways. Significance was confirmed via false-discovery rate (FDR) statistic with 25% cutoff.



Figure EV3. The proportions of early embryonic hematopoietic progenitor populations are not altered in Jag1^{Ndr/Ndr} embryo proper (EP) and yolk sac (YS).

(A) Schematic of the experiment. E9.5 embryos were harvested and yolk sac (YS) and embryo proper (EP) were processed separately for flow cytometry analysis. (B) Gating strategy for identification of the Erythrocyte progenitors (Eryp), Macrophage progenitors (MFp), Megakaryocyte progenitors (MKp), and Erythro-myeloid progenitors (EMP) after dead cell exclusion. (C, D) Relative proportion of live Eryp, EMP, MFp, and MKp cells from Jag1^{Ndr/Mdr} (n = 7), Jag1^{Ndr/+} (n = 9), and Jag1^{+/+} (n = 8) yolk sac (C) and whole E9.5 embryos (D). Graph represents mean ± SD. (E, F) Dot plot of the re-analyzed median scaled In-normalized mRNA expression of Notch signaling components in human (E) and mouse (F') thymic cell populations from Park et al, 2020. One-way ANOVA, multiple comparison with Bonferroni method; n.s., no significant difference.



Figure EV4. Jag1^{Ndr/Ndr} T cells are not autoimmune as shown by the transfer to Rag1^{-/-} hosts.

(A) Schematic of the experiment. (B) Survival curve of the $Rag1^{-/-}$ animals over the course of 8 weeks following transfer with $Jag1^{+/+}$ (n = 6) or $Jag1^{Ndr/Ndr}$ (n = 8) T cells. (C) Relative quantification of $Jag1^{+/+} \rightarrow Rag1^{-/-}$ and $Jag1^{Ndr/Ndr} \rightarrow Rag1^{-/-}$ mouse weight normalized to its original value on day 0 after T cell transfer over 8 weeks (1 = 100% of original weight, mean \pm SD, n = 6-10 mice). (D) Representative H&E staining of intestinal sections (duodenum—top, ileum—bottom) performed 8 weeks after T cell transfer. (E) Comparison of the intestinal inflammatory score calculated using the H&E-stained slides of intestinal sections from the $Jag1^{+/+} \rightarrow Rag1^{-/-}$ (n = 4) and $Jag1^{Ndr/Ndr} \rightarrow Rag1^{-/-}$ (n = 5) mice. (F-H) Representative immunofluorescent images of sections from $Jag1^{+/+} \rightarrow Rag1^{-/-}$ (right) mice intestine after DSS, stained for LCK and FOXP3, counterstained with Hoechst (F). Quantification of average LCK⁺ T cell count/Hoechst area (G), and LCK⁺/FOXP3⁺ Treg/LCK⁺ ratio (H) Means \pm SD, Statistical analysis was performed by unpaired, two-tailed Student's t-test.



Figure EV5. T cells, present in the periportal area after bile duct ligation of Jag1^{Ndr/Ndr} transplanted mice, are enriched in patients with ALGS and anti-correlate with fibrosis.

(A) Representative Sirius red staining in Zone 3 of $Jag1^{+/+} \rightarrow Rag1^{-/-}$ and $Jag1^{Ndt/Ndr} \rightarrow Rag1^{-/-}$ mice after BDL. (B, C) Western blot of total LCK, inactive Tyr505 LCK and β -ACTIN levels in LLL lysates from $Jag1^{+/+} \rightarrow Rag1^{-/-}$ and $Jag1^{Ndt/Ndr} \rightarrow Rag1^{-/-}$ mice (n = 3 each) after BDL (B) and respective quantification (C). Graph represents mean \pm SD. (D, E) Representative immunofluorescent images of cryosections from the left lateral lobe of $Jag1^{+/+} \rightarrow Rag1^{-/-}$ (left) and $Jag1^{Ndt/Ndr} \rightarrow Rag1^{-/-}$ (right) mice after BDL treatment, stained with antibodies against Lck and SMA (D), and respective quantification of Lck⁺ in the periportal area (E), (n = 3 each). Graph represents mean \pm SD, Statistical analysis was performed by unpaired, two-tailed Student's t-test. (F) List of patient samples from Andersson et al, 2018, re-analyzed in this study. Asterisk indicates severity not significance. (G) Heatmap of gene sets over- and under-represented in ALGS and control patients. Significance was confirmed via false-discovery rate (FDR) statistic with 25% cutoff. (H) Pearson correlation of liver damage marker *LGALS3BP*, markers of lymphocytes *CD19*, *CD4*, *CD8A*, *FOXP3* and markers of fibrosis CO*L11*, *LOX* mRNA expression in patients with ALGS and control patients (cholestasis type 2, (N)ES (normalized) enrichment score. Fisher Exact test was used for the gene set enrichment analysis; LLL left lateral lobe, CV central vein, PV portal vein, BD bile duct, HA hepatic artery.