Supporting Information

CaY@C2n: Exploring Molecular Qubits with Ca-Y Metal-Metal Bond

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EXPERIMENTAL DETAILS

Synthesis and Isolation of $CaY@C_{2n}$ **(2n = 80 and 82). The carbon soot containing** calcium and yttrium EMFs were produced by a modified Krätschmer-Huffman directcurrent arc discharge method. The graphite rods, each of which was filled with 0.33 g of CaO, 0.67 g of Y₂O₃ and 2.13 g of graphite powder (molar ratio of Ca : Y : C = 1 : 1 : 30), were vaporized in the arcing chamber under 200 Torr He atmosphere. In total, about 300 graphite rods were burned. The carbon soot was collected and soaked in CS_2 solution for 24 h. CS_2 was removed by a rotary evaporator and then the resulting powder was redissolved in toluene for subsequent HPLC separation. The separation and purification of CaY@C_{2n} (2n = 80 and 82) were achieved by multistage HPLC procedures. Multiple HPLC columns, including Buckyprep M (25 mm \times 250 mm, Cosmosil, Nacalai Tesque Inc.), Buckyprep (10 mm × 250 mm, Cosmosil, Nacalai Tesque, Japan), and 5PBB (10 mm \times 250 mm, Cosmosil, Nacalai Tesque, Japan), were utilized in the procedures.

X-ray Crystallographic Study. Black block cocrystals of $CaY@C_{82}$ [Ni^I(OEP)] $2C_6H_6$ were obtained by slow diffusion from a benzene solution of Ni^I(OEP) into a CS₂ solution of CaY(∂ C₈₂. Single-crystal X-ray data of $CaY@C_s(6)-C₈₂$ were collected at 173 K on a diffractometer (Bruker D8 Venture) equipped with a CCD collector. The multiscan method was used for absorption correction. The structures were solved using the intrinsic phasing methods and refined on F² using full-matrix least-squares using the SHELXL2018 crystallographic software packages. Hydrogen atoms were inserted at calculated positions and constrained with isotropic thermal parameters.

Spectroscopic Studies. The positive-ion mode matrix-assisted laser desorption/ionization time-of-flight (Bruker, Germany) was employed for mass characterization. The UV-vis-NIR absorption spectra of purified $CaY@C_{2n}$ (2n = 80 and 82) were measured in CS_2 solution at room temperature with a Cary 5000 UV-vis-NIR spectrophotometer (Agilent, U.S.). The CW-EPR spectra of the purified $CaY@C_{2n}$ $(2n = 80$ and 82) were measured at the X-band $(\omega = 9.14 \text{ GHz})$ in CS₂ solution with a

JES-X320 spectrometer (JEOL, Japan).

X-band pulse EPR experiments. All X-band pulse EPR experiments were conducted on the CIQTEK EPR100 spectrometer equipped with 1.8 T bipolar Helmholtz electromagnet with high quality resonator, dielectric resonator and dual-mode resonator at the Instrumentation and Service Center for Molecular Sciences, Westlake University. A small amount of $CaY@C_s(6)-C₈₂$ dissolved in toluene was vacuum sealed in 4 mm (external diameter) thick wall quartz tube frozen in liquid nitrogen. Before measurements, the magnetic field of instrument was calibrated by a standard DPPH sample.

For all pulse EPR measurements, $\pi/2$ and π pulses with lengths of 20 ns and 40 ns were chosen to maximize echo intensity besides Rabi oscillations. These pulse lengths were optimized by a three-pulse nutation sequence (nutation pulse $-t - \pi/2 - \tau - \pi - \tau$ − echo), and the microwave attenuation was changed to match the pulse lengths making the first trough position at 40 ns at nutation curve between the intensity of the echo and the length of the nutation pulse. The shot repetition time (SRT) in all experiments was set to be longer than $5T_1$ to ensure that all spins have enough time to return to the thermal equilibrium state. To reduce the influence of noise as much as possible, the threshold for the ends of the echo integrator gate was set about 1/3 of echo amplitude from the baseline.

The echo-detected field sweep (EDFS) spectrum was obtained with the Hahn echo sequence $(\pi/2 - \tau - \pi - \tau -$ echo) where τ was 200 ns by changing the magnetic field with 512 data points, 2048 shots per point and sweeps twice at 90 K. In this experiment, two-step phase cycling $(+x, +x, -x, +x)$ was used to cancel background drift and defense pulse. Integration of the echo was plotted against the magnetic field, giving an EDFS spectrum. The collected EDFS spectrum was fitted using Easyspin on MATLAB R2023a.

The spin–lattice relaxation time (T_1) was measured by employing the inversion recovery sequence $(\pi - t - \pi/2 - \tau - \pi - \text{echo})$ where τ was fixed at 200 ns with 512 data points at the magnetic field with the maximum EDFS intensity (approximately 354

mT). The first length of t was 11 μs and the increment was adjusted to make the flat end of the curve accounts for 1/3 - 1/4 of the entire curve at variable temperature from 10 K to 170 K. In order to cancel background drift, defense pulse and other interference echoes, four-step phase cycling $(+x, +x, +x, +x, -x, +x, -x, +x, -x, -x, -x, +x)$ was conducted. Integration of the echo was plotted against t, giving the inversion recovery curve. The obtained curves were fitted by a mono-exponential decay function:

$$
I = ae^{-\frac{t}{T_1}} + I_0
$$

Where *I* is echo intensity, a is a pre-factor, and *I*₀ represents baseline drift.

The spin decoherence time (T_2) was characterized by the Hahn echo sequence $(\pi/2)$ $-\tau - \pi - \tau$ – echo) with 512 data points at the magnetic field with the maximum EDFS intensity (approximately 354 mT). τ stared at 200 ns and the increment was adjusted to make the flat end of the curve accounts for 1/3 - 1/4 of the entire curve at variable temperature from 10 K to 170 K. In order to cancel background drift and defense pulse, two-step phase cycling $(+x, +x; -x, +x)$ was employed. Integration of the echo was plotted against 2τ , giving the Hahn echo decay curve. The obtained curves were fitted by a stretched exponential decay function:

$$
I = ae^{-\left(\frac{2\tau}{T_2}\right)^q} + I_0
$$

Where *I* is echo intensity, a is a pre-factor, q is the stretch factor, and I_0 represents baseline drift.

The Rabi oscillations were implemented by a three-pulse nutation sequence (nutation pulse $-t - \pi/2 - \tau - \pi - \tau$ – echo) with 1024 data points at the magnetic field with the maximum EDFS intensity (approximately 354 mT) and temperature of 30 K. Here the delay time t was set as 40 μs which is greater than $5T_2$ and τ was 300 ns. The first length of nutation pulse was 10 ns and the increment was 1 ns. In order to cancel background drift, defense pulse and other interference echoes, four-step phase cycling $(+x, +x, +x; +x, -x, +x; -x, +x, -x, -x, -x, -x)$ was conducted. Different Rabi oscillations curves were measured by changing the microwave attenuations (0, 3, 6, 9, 12, 15 dB) where microwave power was matched by varying pulse length. Integration of the echo

was plotted against the length of nutation pulse, giving Rabi oscillations curves. The obtained nutation curves were transformed to the frequency domain spectrum by subtracting baseline with linear fitting, processing with the Hamming window function, filling zeroes and performing fast Fourier transform (FFT) with OriginPro 2021. The observed peak frequency was assigned as the Rabi frequency. The Rabi frequencies were plotted with $10^{-\frac{A}{20 \text{ dB}}}$ (*A* represents the microwave attenuations in the unit of dB), which reflects the ratio between the magnetic field of the output microwave $(B_{MW\,output})$ and the input microwave $(B_{MW\; input})$.

The combination-peak electron spin echo envelope modulation (CP-ESEEM) vs. τ_1 experiment was conducted by seven-pulse sequence $(\pi/2 - \tau_1 - \pi/2 - t - \pi - t - \pi/2)$ $-\tau_1 - \pi/2 - t_s - \pi/2 - \tau_2 - \pi - \tau_2$ – echo) where the sequence of $\pi/2 - t_s - \pi/2 - \tau_2 - \pi$ $-\tau_2$ – echo as remote echo detection was used to measure echo modulation for τ_1 values shorter than the dead time.¹ The t_s and τ_2 were fixed at 8040 ns and 300 ns, respectively. τ_1 started at 80 ns and was incremented by 4 ns per step till 200 ns. For each τ_1 , t started at 400 ns and was incremented by 8 ns with 512 data points. This experiment was carried out with 512 shots per point and at the magnetic field with the maximum EDFS intensity (approximately 354 mT) and temperature of 30 K. In order to cancel background drift, defense pulse and other interference echoes, eight‐step phase cycling $(+x, +x, +x, +x, +x, +x, +x, -x, +x, -x, +x, +x, +x, +x, +x, +x, -x, -x, +x, +x, -x, +x, -x, -x,$ ‐x, +x, +x, +x, +x, +x; +x, +x, +x, ‐x, +x, +x, +x; ‐x, +x, +x, ‐x, +x, +x, +x; +x, ‐x, +x, $-x, +x, +x, +x; -x, -x, +x, -x, +x, +x, +x)$ was conducted. For each τ_1 , integration of the echo was plotted against the t, giving time‐domain CP‐ESEEM spectrum. The obtained time‐domain CP‐ESEEM spectrum was transformed to the frequency-domain spectrum by subtracting baseline with exponential decay fitting, processing with the Hamming window function, filling zeroes and performing fast Fourier transform (FFT) by CIQTEK EPR100 software. The frequency‐domain CP‐ESEEM spectrum was plotted against τ_1 , giving a two-dimensional CP-ESEEM spectrum.

The HYSCORE experiment was performed by seven-pulse sequence $(\pi/2 - \tau_1$ $\pi/2 - t_1 - \pi - t_2 - \pi/2 - \tau_1 - \pi/2 - t_s - \pi/2 - \tau_2 - \pi - \tau_2$ – echo) where the sequence of $\pi/2$ – $t_s - \pi/2 - \tau_2 - \pi - \tau_2$ – echo as remote echo detection was used to measure echo modulation. The t_s and τ_2 were fixed at 8040 ns and 300 ns, respectively. τ_1 was fixed at 164 ns, which is the optimal delay time obtained by CP-ESEEM vs. τ_1 . Both t_1 and t_2 started at 400 ns and were incremented 16 ns per step independently with 128 date points. This experiment was carried out with 400 shots per point and at the magnetic field with the maximum EDFS intensity (approximately 354 mT) and temperature of 30 K. In order to cancel background drift, the defense pulse and other interference echoes, eight‐step phase cycling (+x, +x, +x, +x, +x, +x, +x; ‐x, +x, +x, +x, +x, +x, +x; +x, ‐x, +x, +x, +x, +x, +x; ‐x, ‐x, +x, +x, +x, +x, +x; +x, +x, +x, ‐x, +x, +x, +x; ‐x, +x, $+x$, $-x$, $+x$, $+x$, $+x$, $-x$, $+x$, $-x$, $+x$, $+x$, $+x$, $-x$, $-x$, $+x$, $-x$, $+x$, $+x$, $+x$) was conducted. For each combination of t_1 and t_2 , integration of the echo was plotted against the t_1 and t₂, giving time-domain HYSCORE spectrum. The obtained HYSCORE spectrum was transformed to frequency-domain spectrum by subtracting baseline with polynomial fitting, processing with the Hamming window function, filling zero and performing two-dimensional fast Fourier transform (FFT) in two dimensions by CIQTEK EPR100 software.

Computational Details. Kohn-Sham density functional theory (DFT) calculations were performed with the Amsterdam Density Functional (ADF, v. 2019) package.² The PBE0 exchange correlation functional³⁻⁵ was used in conjunction with all-electron triple- ζ polarized (TZP) Slater-type orbital (STO) basis sets quality.^{6, 7} Scalar relativistic (SR) zero-order regular approximation (ZORA) was included for relativistic effects.⁸ 'D3' dispersion corrections by Grimme were also considered.^{9, 10} ESR calculations were performed using an all-electron basis set with ADF. Calculations of redox properties were performed using the BP86 functional¹¹ and introducing solvent effects by means of the Conductor like Screening Model (COSMO),¹² as implemented in ADF using o-dichlorobenzene as a solvent.13

High performance liquid chromatography (HPLC) separation process of $CaY@C_s(6)-C₈₂$. Toluene was used as mobile phase in the multiple-step HPLC procedure. The first step was performed on a Buckyprep-M column ($25 \text{ mm} \times 250 \text{ mm}$, Cosmosil Nacalai Tesque) (Figure S1 (a)). The fraction from 18 to 23.5 min (marked in red) containing $CaY(\partial C_{82}$ was collected and then injected into the Buckyprep column $(10 \text{ mm} \times 250 \text{ mm}, \text{Cosmosil}, \text{Nacalai Tesque})$ for the second step. In the second step, the fraction from 46 to 52 min (marked in orange) was collected for the next step of HPLC separation (Figure S1 (b)). The third step was conducted on a 5PBB column (10 $mm \times 250$ mm, Cosmosil, Nacalai Tesque) and the fraction from 87 to 96 min (marked in green) was collected (Figure S1 (c)). Then, this fraction was injected into a Buckyprep column (10 mm \times 250 mm, Cosmosil, Nacalai Tesque) in recycle mode for the fourth step (Figure S1 (d)). The fraction marked in blue was collected and reinjected into the Buckyprep column (10 mm \times 250 mm, Cosmosil, Nacalai Tesque) in recycle mode for the fifth step. After that, the fraction marked in pink, namely pure $CaY@C_{82}$, was finally obtained (Figure S1 (e)). Figure S1 (f) shows the purity of the isolated $CaY@C_{82}$.

Figure S1. (a-e) HPLC profiles showing the isolation procedures of CaY@C₈₂. (f) The MALDI-TOF mass spectrum of the purified $CaY@C_{82}$.

High performance liquid chromatography (HPLC) separation process of $CaY@C_{2v}(5)-C₈₀$ Toluene was used as mobile phase in the multiple-step HPLC procedure. The first step was performed on a Buckyprep-M column (25 mm \times 250 mm, Cosmosil Nacalai Tesque) (Figure S2 (a)). The fraction from 18 to 23.5 min (marked in red) containing $CaY@C_{80}$ was collected and then injected into the Buckyprep column $(10 \text{ mm} \times 250 \text{ mm}, \text{Cosmosil}, \text{Nacalai Tesque})$ for the second step. In the second step, the fraction from 46 to 52 min (marked in orange) was collected for the next step of HPLC separation (Figure S2 (b)). The third step was conducted on a 5PBB column (10 $mm \times 250$ mm, Cosmosil, Nacalai Tesque) and the fraction from 73 to 79 min (marked in green) was collected (Figure S2 (c)). Then, this fraction was injected into a Buckyprep column (10 mm \times 250 mm, Cosmosil, Nacalai Tesque) in recycle mode for the fourth step. After that, the fraction marked in blue, namely pure $CaY@C_{80}$, was finally obtained (Figure S2 (d)). Figure S2 (e) shows the purity of the isolated $CaY@C_{80}$.

Figure S2. (a-d) HPLC profiles showing the isolation procedures of $CaY@C_{80}$. (e) The MALDI-TOF mass spectrum of the purified $CaY@C_{80}$.

Figure S3. HPLC chromatogram of purified $CaY@C_{82}$ (a) and $CaY@C_{80}$ (b) on a Buckyprep column with toluene as the eluent (HPLC conditions: $\lambda = 310$ nm; flow rate = 4 mL/min). The insets show the positive-ion mode MALDI-TOF mass spectra and expansions of the corresponding experimental isotopic distribution of $CaY(\partial_{\theta}C_{82}$ (a) and $CaY@C_{80}$ (b) in comparison with the theoretical one.

Figure S4. Drawing of all the Ca and Y disordered sites and their mirror-related counter-parts (Ca1_m and Y1_m) in CaY@C_{*s*}(6)-C₈₂.

Figure S5. UV-vis-NIR absorption spectrum of CaY@C₈₂ (a) and CaY@C₈₀ (b). The inset shows the photograph of purified $CaY@C_{82}$ (a) and $CaY@C_{80}$ (b) dissolved in $CS₂$ solution.

	Metal site \vert Ca1/Ca1 _m	$Y1/Y1_m$	Ca2	Ca ₃	
\vert Occupancy \vert 0.3157(17) \vert 0.3157(17) \vert 0.172(3) \vert 0.196(3) \vert 0.172(3) \vert 0.196(3)					

Table S1. Atom occupancy of metal sites in $CaY@C_s(6)-C₈₂$.

Table S2. Crystallographic information of $CaY@C_{82}$ [Niⁿ(OEP)] $2C_6H_6$

	$CaY@C_{82}$ [Ni ⁿ (OEP)] $2C_6H_6$
Empirical formula	$C130H56CaN4NiY$
Formula weight	1861.48
Crystal system	monoclinic
Space group	C2/m
a, \AA	25.2749(9)
b, \AA	15.2299(5)
c, \AA	19.9217(7)
α , deg	90
β , deg	95.247(2)
γ , deg	90
Volume, Å ³	7636.4(5)
Z	$\overline{4}$
T, K	173(2)
F(000)	3804
ρ , g·cm ⁻³	1.619
θ , deg	2.227-77.942
R_1 (all data)	0.1348
wR_2 (all data)	0.2804
$R_1(I > 2.0\sigma(I))$	0.1069
$wR_2(I > 2.0\sigma(I))$	0.2589
parameters	1040
Goodness-of-fit indicator	1.083
$R_{\rm int}$	0.0463

Figure S6. DFT-optimized geometry of CaY@*Cs*(6)-C82 cage. a) Ca-Y distance (in Å) is indicated. b) Ca-C and Y-C distances (in Å) are represented.

Figure S7. DFT-optimized geometry of $CaY@C_{2v}(5)$ -C₈₀ cage. a) Ca-Y distance (in Å) is indicated. b) Ca-C and Y-C distances (in Å) are represented.

Table S3. Relative energies (Erel, in kcal·mol-1) and Ca-Y distances (in Å) for different orientations of the Ca and Y metal atoms inside the $C_{2v}(5)$ -C₈₀ cage. Orientations according to Figure S8.

Orientation	Erel	$dist(Ca-Y)$
A	0.0	3.841
B	42	3.866
C	12.2	3.543
D	21.8	3.982

Figure S8. Different orientations of Ca and Y metal atoms within the $C_{2v}(5)$ -C₈₀ cage. Ca and Y are indicated in each DFT-optimized geometry.

Table S4. Relative energies (Erel, in kcal·mol-1) and Ca-Y distances (in Å) for different orientations of the Ca and Y metal atoms inside the $C_s(6)$ -C₈₂ cage. Orientations according to Figure S9.

Orientation	Erel	$dist(Ca-Y)$
\mathbf{A}	0.0	3.705
B	3.5	3.695
C	8.8	3.617
D	8.9	3.610
E	9.1	4.040
F	10.5	3.746
G	16.8	3.970
H	27.1	3.697
I	31.4	3.668

Figure S9. Different orientations of Ca and Y metal atoms within the $C_s(6)$ -C₈₂ cage. Ca and Y are indicated in each DFT-optimized geometry.

Figure S10. a) DFT-optimized geometry of $CaY@C_{2v}(5)-C_{80}$ cage. b) Potential electrostatic map of $C_{2\nu}(5)$ -C₈₀⁴. c) Isosurfaces (\pm 0.03 au) of the LUMO and LUMO+1 for $C_{2\nu}(5)$ - C_{80} .

Figure S11. a) DFT-optimized geometry of $CaY@C_s(6)-C₈₂$ cage. b) Potential electrostatic map of $C_s(6)$ - C_{82} ⁴. c) Isosurfaces (\pm 0.03 au) of the LUMO and LUMO+1 for $C_s(6)$ - C_{82} .

Figure S12. Molecular orbital (MO) diagrams for $CaY@C_{2v}(5)-C_{80}$ (left) and $CaY@C_s(6)-C₈₂$ (right). The energy levels of the delocalized sigma orbital $a₁$ (for alpha- and beta-spins) are drawn in red and the associated MO isosurfaces $(\pm 0.03 \text{ a.u.})$ with the corresponding MO energy (in eV) are shown on the side. The energy levels for the highest four electrons (in black) from the cage are also represented.

Figure S13. Representation of the correlation between the hyperfine coupling constant (in MHz) and the atomic Mulliken spin density of Y atom for all the $X-Y@C_{2n}$ systems shown in Table 1. Correlation coefficient *r* is given inside the plot.

Figure S14. Different X-ray orientations of Ca and Y metal atoms within the $C_s(6)$ -C₈₂ cage with their corresponding DFT-optimized structures. The metal displacement during the DFT-optimization of Ca and Y is also represented. For both metals, lighter colour atoms correspond to the X-ray position and darker colour atoms to the DFToptimized position. Ca and Y are indicated in each geometry.

Figure S15. Spin density distributions for the oxidized $CaY@C_s(6)-C₈₂⁺$ and reduced $CaY@C_s(6)-C₈₂$ (triplet states) with an isosurface of ± 0.002 a.u. Both oxidation and reduction are predicted to take place on the carbon cage and not in the single-electron occupied Ca-Y bonding orbital. The closed-shell singlet states are 6 kcal mol-1 higher in energy than the triplets for both the oxidized $CaY@C_s(6)-C₈₂$ ⁺ and reduced $CaY@C_s(6)-C₈₂$ systems. The first oxidation potential (in o-dcb, +0.40 V) and the first reduction potential (-0.89 V), as well as the electrochemical gap (1.29 V), are comparable to those computed for $Sc_2O@C_s(6)$ -C₈₂ (1.18 V) and $Sc_2S@C_s(6)$ -C₈₂ (1.23 V), which share the same cage.¹⁴ Therefore, the unpaired electron in the sigma bond seems to be rather well protected by the carbon cage.

xzy Coordinates

$CaY@C_{2\nu}(5)-C_{80}$

$CaY@C_s(6)-C₈₂$

Figure S16. The X-band CW spectra of (a) $CaY@C_s(6)$ -C₈₂ and (b) $CaY@C_{2v}(5)$ -C₈₀ measured in CS_2 solution at 290 K. Each spectrum was fitted using Easyspin 6.0.0 in MATLAB R2024a with a $S = 1/2$ electron spin system coupled with a $I = 1/2$ nuclear spin. The spectrum lines were found to exhibit the Lorentzian lineshape. For $CaY@C_s(6)-C₈₂$, the fitted g_{iso} , *A*, and linewidth are 1.981855(2), 260.42(1) MHz, and 0.0989(3) mT, respectively. For $CaY@C_{2v}(5)-C_{80}$, the fitted g_{iso} , *A* and linewidth are 1.982660(2), 251.65(1) MHz, and 0.0994(3) mT, respectively.

Figure S17. (a) Inversion recovery curves and (b) Hahn echo decay curves of $CaY@C_s(6)-C₈₂$ dissolved in toluene at variable temperature from 10 K to 170 K.

Figure S18. Inversion recovery curves of $CaY@C_s(6)-C₈₂$ dissolved in toluene via fitting with a mono-exponential decay at 10, 30, 60, 90, 120, 150 K. The black lines are the fitting curves.

Figure S19. Hahn echo decay curves of $CaY@C_s(6)$ -C₈₂ dissolved in toluene via fitting with stretched exponential decay at 10, 30, 60, 90, 120, 150 K. The black lines are the fitting curves.

Figure S20. a) Two-dimensional CP-ESEEM vs. τ_1 spectrum and b) the corresponding superimposed intensity across all τ_1 values from 80 to 200 ns at 30 K for CaY@C_{*s*}(6)- C_{82} dissolved in toluene.

Figure S21. The frequency domain spectrum of Rabi oscillations after fast Fourier transformation at different microwave attenuations for $CaY@C_s(6)-C₈₂$ dissolved in toluene.

Table S5. Summary of spin–lattice relaxation time (T_1) , spin decoherence time (T_2) and corresponding stretch factor q at variable temperature from 10 K to 170 K for $CaY@C_s(6)-C₈₂$ dissolved in toluene.

Temperature (K)	T_1 (μ s)	T_2 (μ s)	$\mathbf q$
10	$8.093(40) \times 10^3$	6.52(2)	2.36(2)
15	$3.507(19) \times 10^3$	6.79(2)	2.49(3)
20	$1.929(10) \times 10^3$	7.25(2)	2.69(2)
25	1.213 (6) \times 10 ³	7.37(1)	2.64(2)
30	$8.01(5) \times 10^{2}$	7.54(1)	2.67(2)
40	$4.01(3) \times 10^2$	7.74(2)	2.67(2)
50	$2.42(1) \times 10^2$	7.68(2)	2.61(2)
60	157.2(8)	7.50(1)	2.53(2)
70	108.2(5)	7.18(1)	2.38(2)
80	77.9(5)	6.49(2)	2.09(2)
90	56.8(2)	5.28(2)	1.73(2)
100	45.6(2)	3.66(2)	1.39(1)
120	26.0(2)	1.22(2)	1.12(2)
140	19.2(3)	0.74(4)	0.72(2)
150	16.0(3)	0.61(5)	0.74(3)
160	12.2(4)	0.69(6)	0.86(5)
170	9.5(4)	0.78(6)	1.07(9)

Figure S22. Comparison of the yield of $Y_2@C_{82}$ and $CaY@C_{82}$. The figure on the left shows the chromatogram of the third step of the separation of $CaY@C_{82}$ and the right one is the corresponding MALDI-TOF-MS spectrum. The yield of $CaY@C_{82}$ is estimated to be about 1/20 of that of the $Y_2@C_{82}$.

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