

Supplementary Materials for

Same rule, different genes: *Blimp1* is a pair-rule gene in the milkweed bug  
*Oncopeltus fasciatus*

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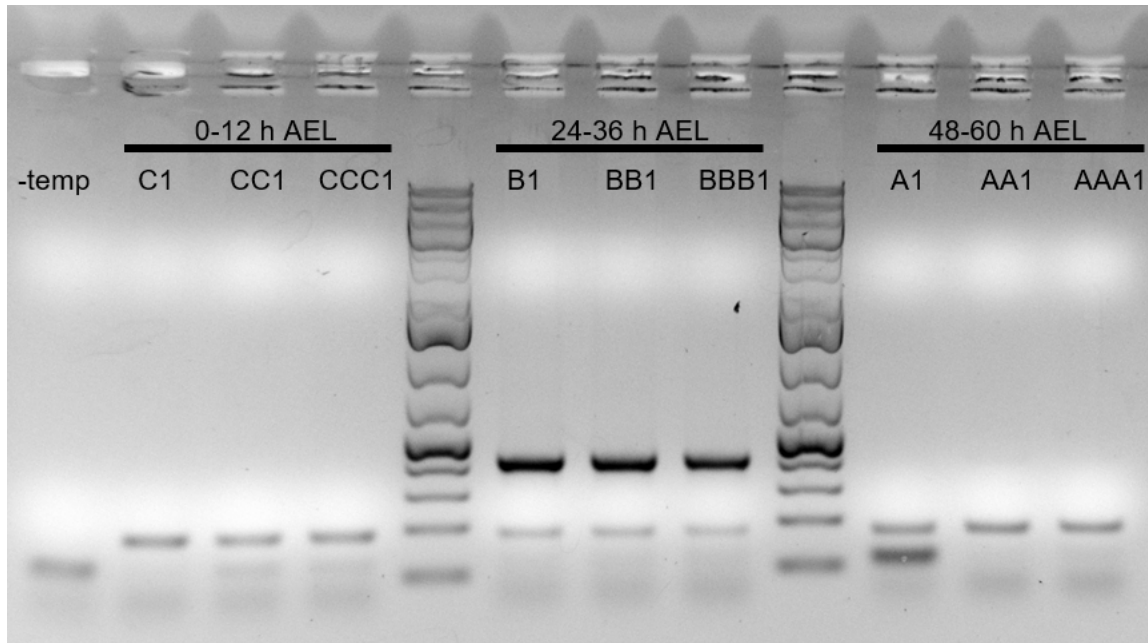
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**The PDF file includes:**

Figs. S1 to S6  
Tables S1 and S2  
Legends for data S1 and S2

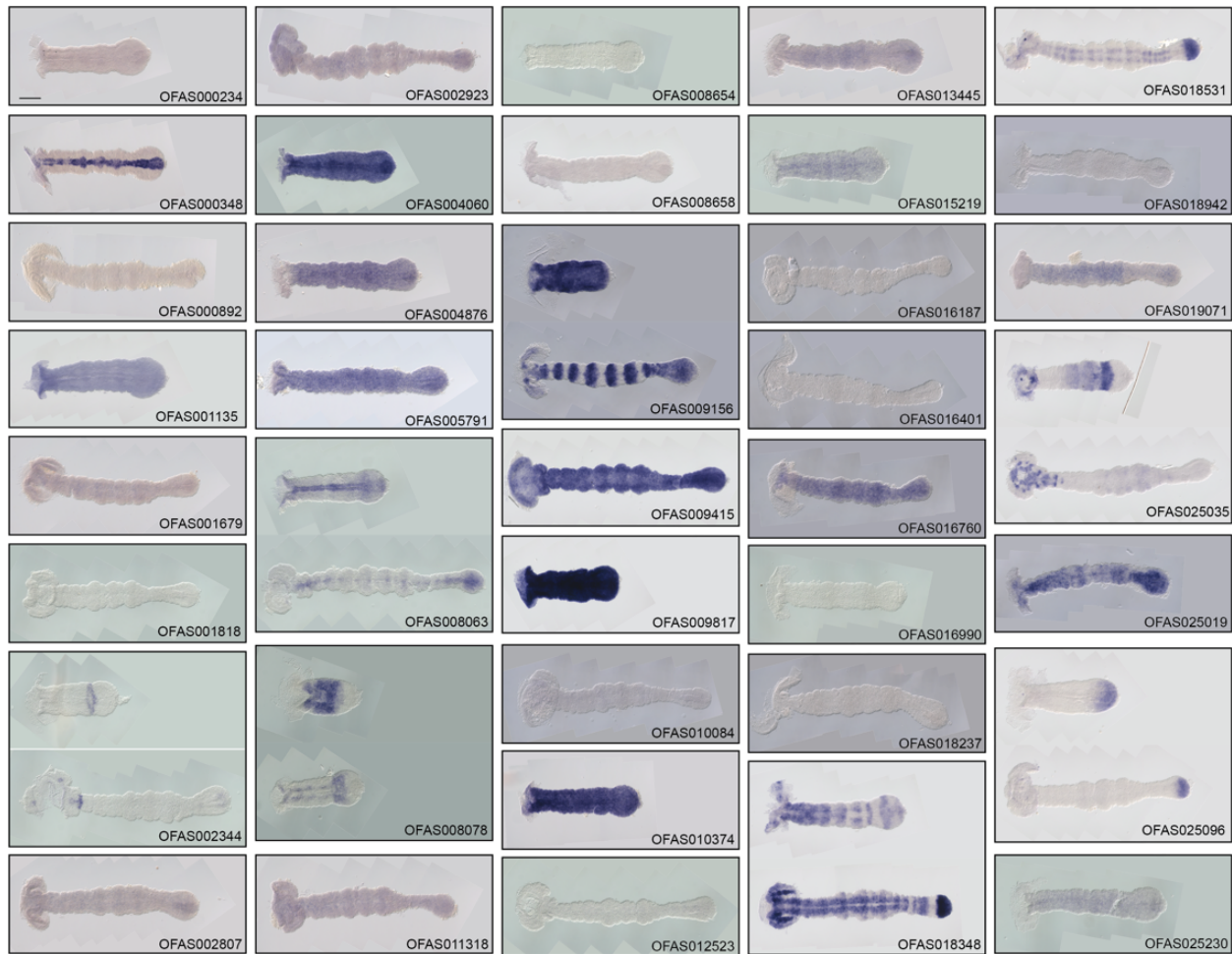
**Other Supplementary Material for this manuscript includes the following:**

Data S1 and S2



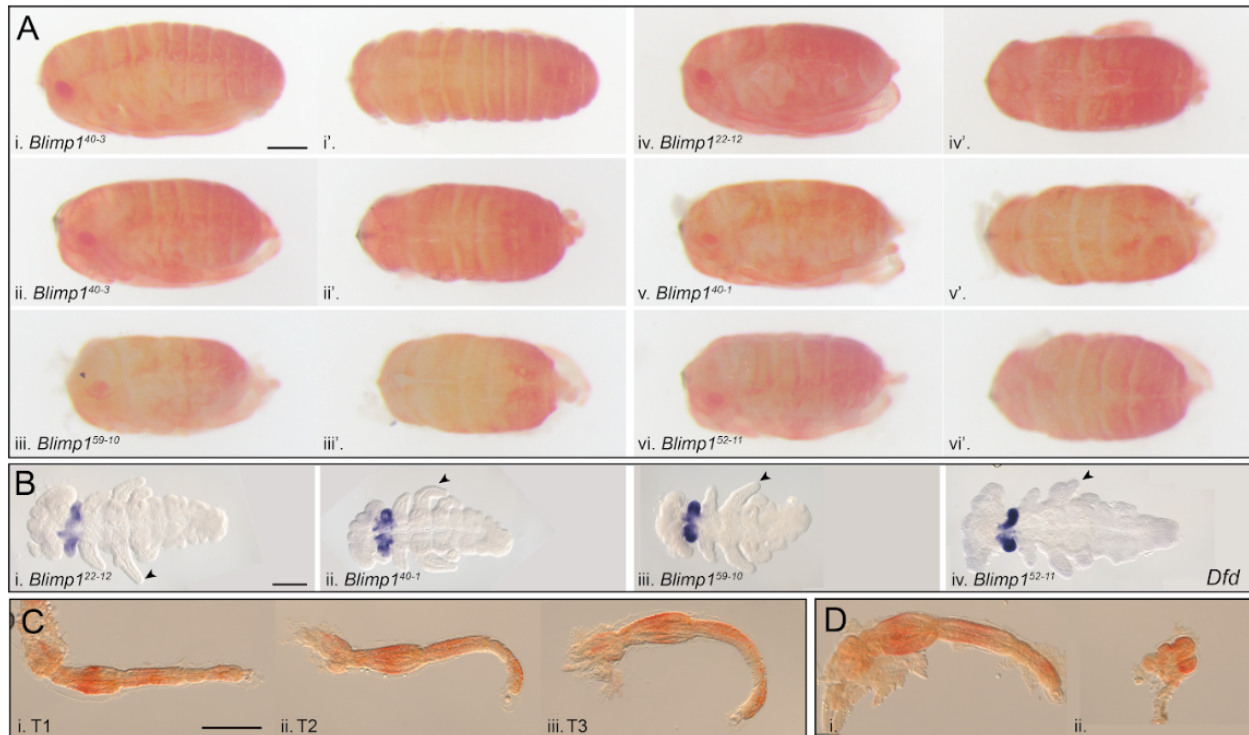
**Fig. S1. RT-PCR of *E75A* across three embryonic time points**

An 366 bp amplicon of *E75A* was amplified at 0-12, 24-36, and 48-60 hours after egg laying (h AEL) to validate staging for RNA-seq. A 173 bp amplicon of *actin* was simultaneous amplified as a positive control. The first lane is a no template control. Three replicates were performed per time point.



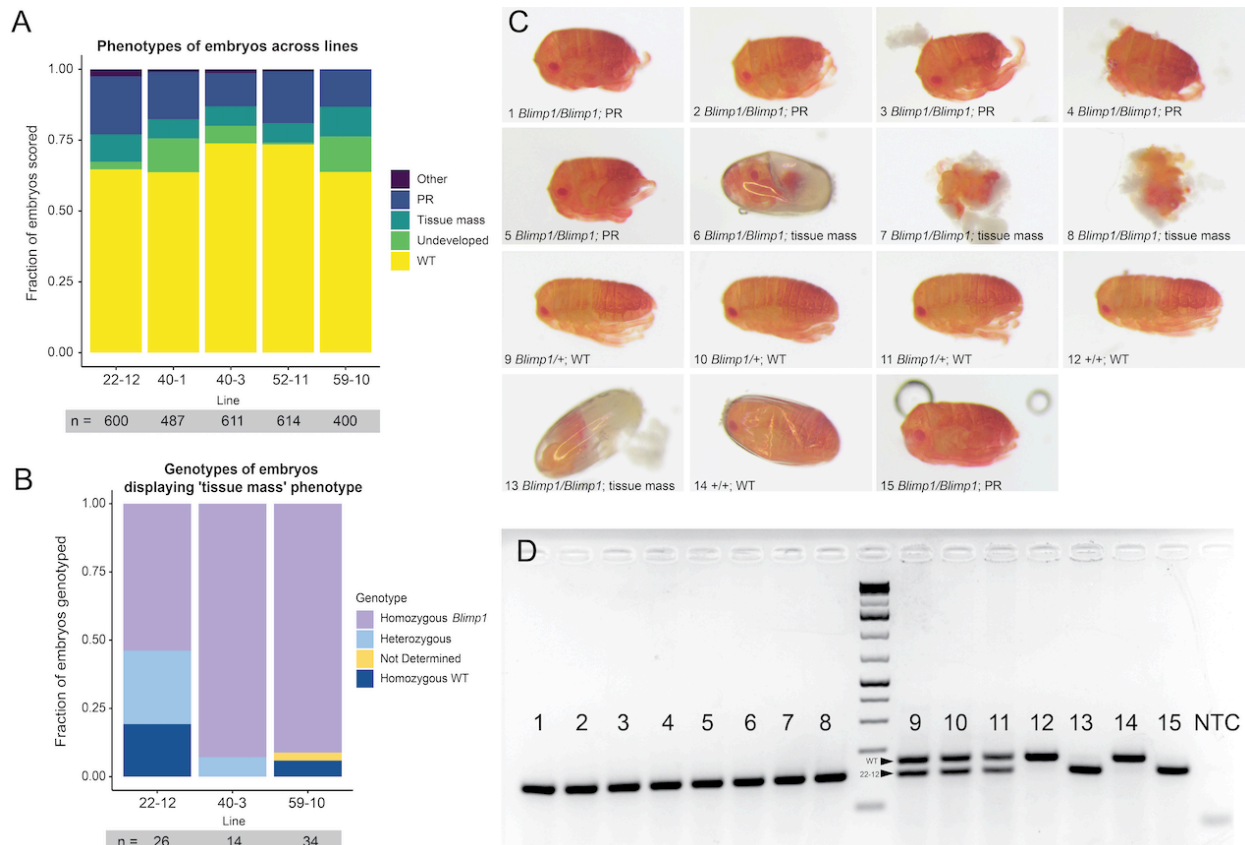
**Fig. S2. In situ hybridization screen for *E75A*-coexpressed genes.**

Expression patterns identified in the initial screen *Of-E75A*-coexpressed genes are shown. These 38 genes are in addition to the 8 presented in Fig. 1 for a total of 46 transcription factor-encoding genes screened by in situ hybridization to date. Expression patterns are shown at 24-48 h AEL for the following genes: OFAS000234, OFAS000348, OFAS000892, OFAS001135, OFAS001679, OFAS001818, OFAS002344 (annotated as *Knirps2*), OFAS002807, OFAS002923, OFAS004060, OFAS004876, OFAS005791, OFAS008063 (annotated as *retained*), OFAS008078 (annotated as *SoxN*), OFAS008654, OFAS008658, OFAS009156 (annotated as *cubitus interruptus-partial*), OFAS009415, OFAS009817, OFAS010084, OFAS010374, OFAS011318, OFAS012523, OFAS013445, OFAS015219, OFAS016187, OFAS016401, OFAS016760, OFAS016990, OFAS018237, OFAS018348 (annotated as *hunchback*), OFAS018531 (annotated as *Dichaete*), OFAS018942, OFAS019071, OFAS025035 (annotated as *giant*), OFAS025019 (annotated as *extramacrochaetae*), OFAS025096 (annotated as *caudal*), and OFAS025230. Scale bar corresponds to 200  $\mu$ m. Genes are named based on published annotations (Panfilio et al. 2019).



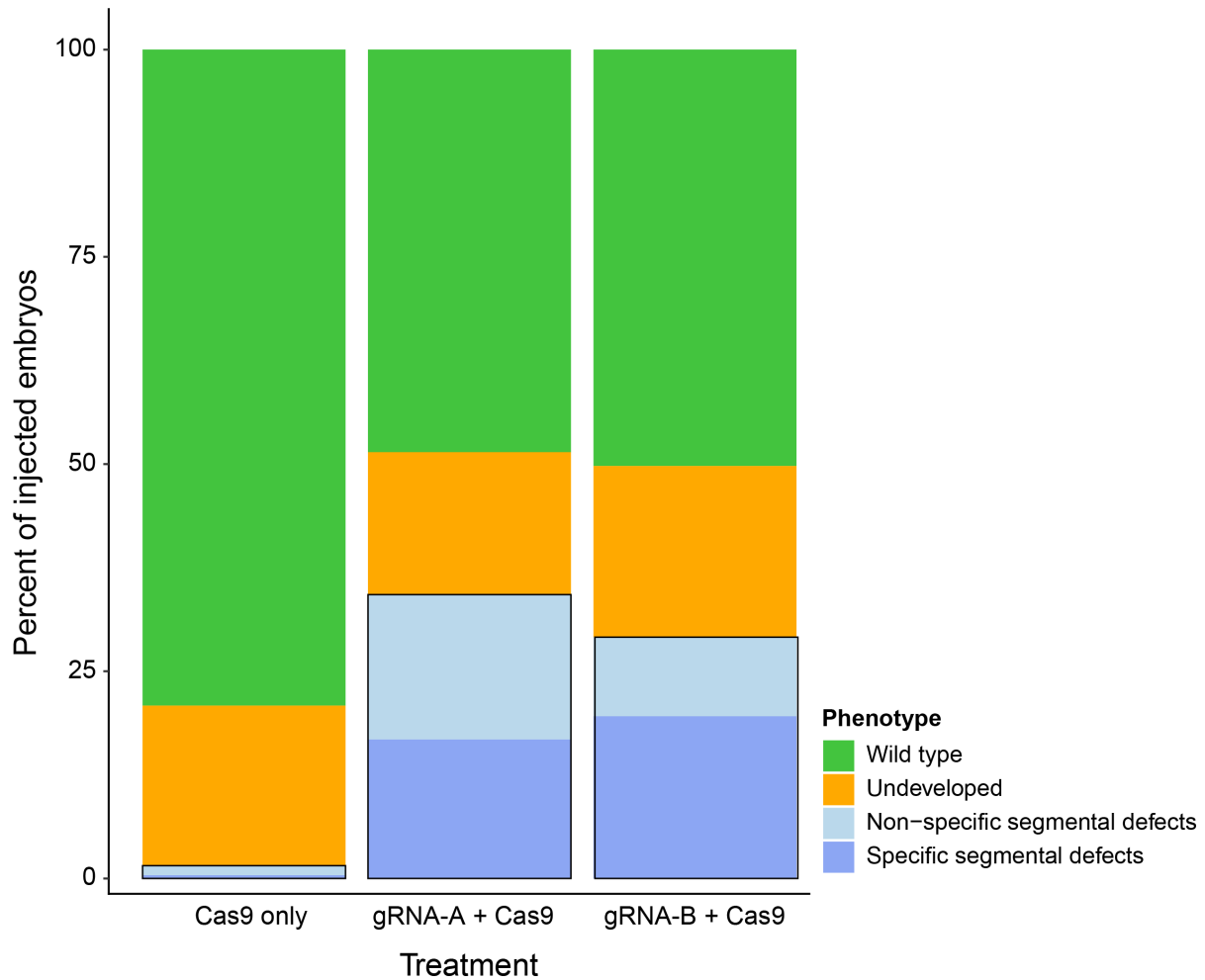
**Fig. S3. Pair-rule phenotypes in all *Of-Blimp1* mutant lines**

All embryos are offspring of self-crosses for the indicated line. **A)** Pre-hatchling phenotypes. i) Wildtype-like *Of-Blimp1*<sup>40-3</sup> offspring, Pair-rule defects for: (ii) *Of-Blimp1*<sup>40-3</sup>, (iii) *Of-Blimp1*<sup>59-10</sup>, (iv) *Of-Blimp1*<sup>22-12</sup>, (v) *Of-Blimp1*<sup>40-1</sup>, and (vi) *Of-Blimp1*<sup>52-11</sup> offspring. i-vi) lateral view; i'-vi') dorsal view. Scale bar corresponds to 200  $\mu$ m. **B)** *Dfd* expression in embryos from line (i) *Of-Blimp1*<sup>22-12</sup>, (ii) *Of-Blimp1*<sup>40-1</sup>, (iii) *Of-Blimp1*<sup>59-10</sup>, (iv) *Of-Blimp1*<sup>52-11</sup>. Arrowhead indicates fused appendage. **C)** T1 (i), T2 (ii), and T3 (iii) leg dissected from a wild type-like *Of-Blimp1*<sup>22-12</sup> individual. **D)** Leg (i) and nub-like appendage (ii) dissected from a presumptive *Of-Blimp1*<sup>22-12</sup> homozygote. Scale bars represent 200  $\mu$ m.



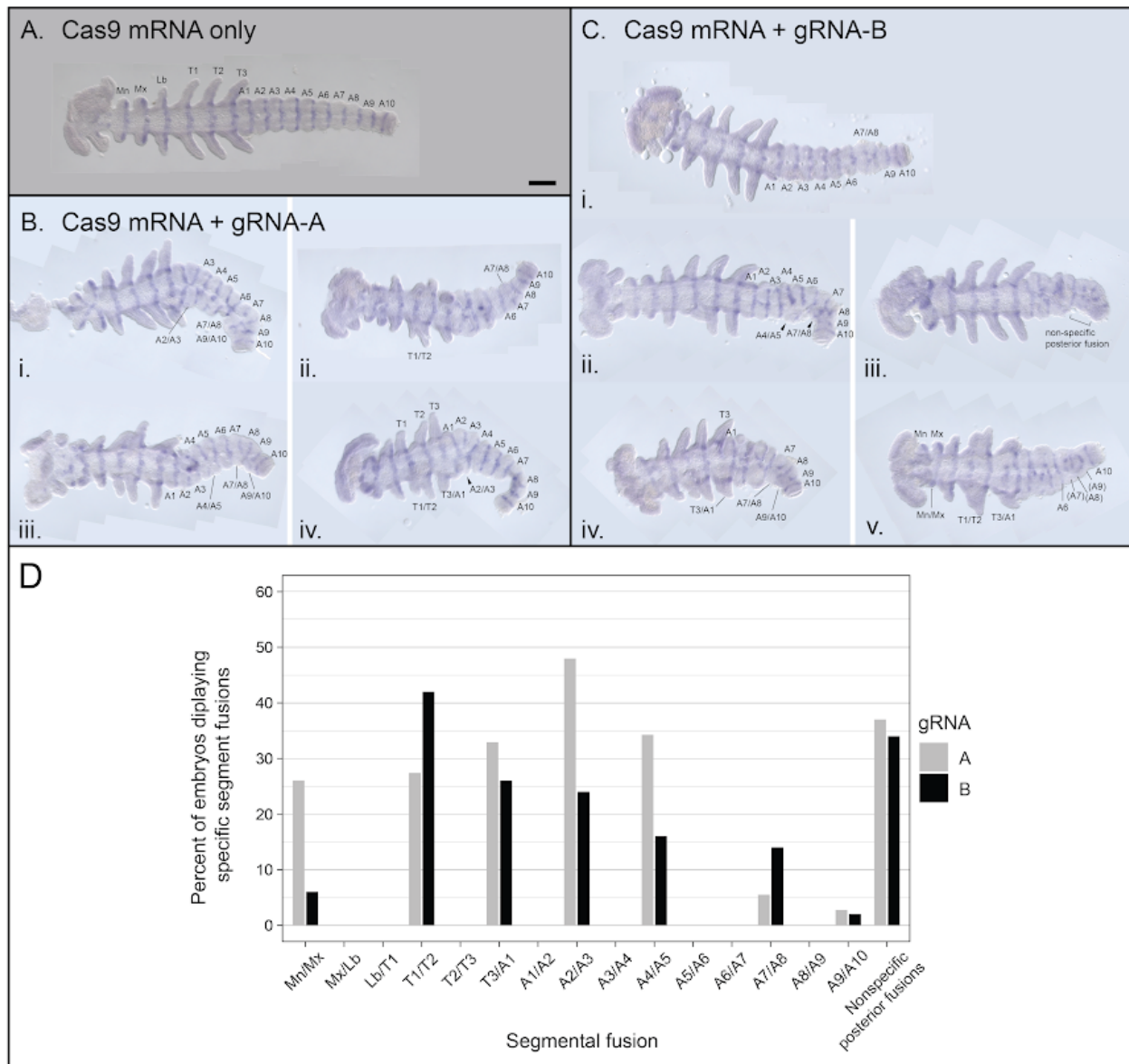
**Fig. S4. Phenotyping and genotyping 'tissue mass' phenotype across lines.**

**A)** Graph showing overall phenotypes of embryos derived from self-crosses of indicated *Blimp 1* mutant lines. A large portion of embryos from all five lines displayed an unexpected 'tissue mass' phenotype. Number of embryos examined from each line are presented in a gray box beneath the plot. **B)** Genotypes of embryos displaying 'tissue mass' phenotype from lines *Blimp1*<sup>22-12</sup>, *Blimp1*<sup>40-3</sup>, and *Blimp1*<sup>59-10</sup>. While most 'tissue mass' embryos are *Blimp1* homozygotes, many—particularly from line *Blimp1*<sup>22-12</sup>—are heterozygous or homozygous WT, suggesting that this phenotype is not caused by *Blimp1* mutation, but may be due to a linked off-target mutation. The number of embryos genotyped from each line are displayed in a gray box beneath the plot. **C)** 15 *Blimp1*<sup>22-12</sup> embryos that were phenotyped prior to being sacrificed for genotyping. Embryos 6-8 and 13 display the 'tissue mass' phenotype. Embryos are labeled: Genotype; Phenotype. **D)** Genotyping gel from embryos in C. A single lower band is present for *Blimp1*<sup>22-12</sup> homozygotes (1-8, 13, and 15), a single upper band is present for samples homozygous for the WT allele (12 and 14), and both bands are present in heterozygotes (9-11). NTC = no template control.



**Fig. S5. Frequencies of phenotypes observed after somatic *Of-Blimp1* CRISPR/Cas9 somatic mutagenesis**

Embryos injected with Cas9 mRNA only (n=259), Cas9 mRNA and *Of-Blimp1* gRNA-A (n=698), or Cas9 mRNA and *Of-Blimp1* gRNA-B (n=639) were broadly categorized as wild type, undeveloped (no clear structures, likely killed by injection or unfertilized), having non-specific segmental defects (defects were too severe or segment loss too complete to accurately identify affected segments), or as having specific partial segmental defects. Only embryos in this last category could be accurately identified and used for analysis presented in main text Fig. 6. The percentage of embryos displaying any segmental defects (both specific and non-specific) is outlined in the plot.



**Fig. S6. Expression of *invected (inv)* in *Of-Blimp1* somatic mutants**

**A)** An embryo injected with Cas9 mRNA only, displaying wildtype segment morphology and a wildtype *inv* expression pattern. **B)** Embryos injected with Cas9 mRNA and *Of-Blimp1* gRNA-A. (i) Embryo displaying disrupted segment boundaries, including fusion of segments A2/A3, A7/A8, and A9/A10 on one side. (ii) Embryo displaying fusion of segments T1 and T2, A7 and A8. (iii) Embryo displaying A4/A5, A7/A8, and A9/A10 segmental fusions. (iv) An embryo displaying T1/T2, T3/A1, and A2/A3 fusions on one side. **C)** Embryos injected with Cas9 mRNA and *Of-Blimp1* gRNA-B. (i) An embryo displaying fusion of segments A7 and A8. (ii) An embryo displaying A4/A5 and A7/A8 segmental fusions. (iii) An embryo displaying reduced *inv* expression and posterior curling in the posteriormost segments (bracket), which was scored as having nonspecific posterior defects. (iv) An embryo displaying T3/A1, A7/A8, and A9/A10 segmental fusions. (v) An embryo with Mn/Mx, T1/T2 and T3/A1 segmental fusions, and nonspecific posterior fusions. In the region of A7 through A9, *inv* expression did not form complete stripes. All embryos were fixed at 67-71 h AEL. **D)** Plot displaying the percent of each segmental fusion observed in embryos injected with Cas9 mRNA and gRNA-A (gray) or -B

(black). Nonspecific posterior fusions refer to segment fusions in the region of A7 through A10 which could not be specifically identified, which were marked by reduced or disrupted *inv* expression and often by curling of the posterior abdomen. Scale bar represents 200  $\mu\text{m}$ .



**Table S1.**

Primer sequences used to amplify probe template for in situ hybridization screen of genes co-expressed with *Of-E75A*.

<b>Primer name</b>	<b>Primer Sequence</b>	<b>Amplicon (bp)</b>
ofas008150-F	GCAGTGTATCTAGTACTGGACC	1,164
ofas008150-RT7	taatacgactcactatagggagaGTAGTCGGAGTATGCAGGAG	
ofas013944-F	GCACCAATGAAGAATCAGGACTTCCAAAGATG	605
ofas013944-RT7	taatacgactcactatagggagaCTTCTGATGACCAAGCACTGTTTCCATCTTG	
ofas000035-F	GCTTGATTAACCTGTTGCAGAAGCAGGGTC	686
ofas000035-RT7	taatacgactcactatagggagaCGTGATGCCCTTCGTGGTAGAGGATCG	
ofas012059-F	CTAGAAGCCCTCTTCGCCAAGAC	518
ofas012059-RT7	taatacgactcactatagggagaGTAAGTGGTATTCAGGATACGATCCCGAGG	
ofas025181-F	GGGACTAAGTGTGAGAAGTGCAGTTTAAGC	711
ofas025181-RT7	taatacgactcactatagggagaGAGTGCGAAGTCTGTGAGAGCCTTCC	
ofas025116-F	GAAGGAGAAGTTCGAAGAATGC	685
ofas025116-RT7	taatacgactcactatagggagaGTGCAACTTGATCTCGGTG	
ofas025203-F	GAGTTGGAAGAACGTGATCTCTGGAGAAG	601
ofas025203-RT7	taatacgactcactatagggagaCCAACCAGCCACTTTCATATGCTGTTG	
ofas003956-F	GATGAAGAAGGAATCAGACGGAACCTCC	642
ofas003956-RT7	taatacgactcactatagggagaGAGGTGCTTCAGAACCACTATAGGACTTATTGAC	
ofas000234-F	CTAGTTCTGATAGTAGTGCAGGTTCTGTGTCATC	677
ofas000234-RT7	taatacgactcactatagggagaCTGGAGACCAGCAGGAAGTCCC	
ofas000348-F	GATTAGCCATGACGAAAGTGCCGTCCTTC	727
ofas000348-RT7	taatacgactcactatagggagaGCCCTTCACTCCTCTCCTCGTTGC	

ofas000892-F	GTTATAGAGCCATCCAAGGAACCTAC	717
ofas000892-RT7	taatacgactcactatagggagaCAGATGCTGTTCCAAGGCAT TG	
ofas001135-F	GAGTGGTTTTAGGACATTTGAATC	339
ofas001135-RT7	taatacgactcactatagggagaCCATTTTCAAACCTTGCAACA AC	
ofas001679-F	GCCATCGATAATAAAATTGAGCAGGC	598
ofas001679-RT7	taatacgactcactatagggagaCAA GTG GAA GGG TAT GCT ACG C	
ofas001818-F	CTGGTCCTAAATGTGTTGTGTTGG	833
ofas001818-RT7	taatacgactcactatagggagaCTGTCCACAATTAGTGGCAA GAATTAC	
ofas002344-F-2	GAGCCAGCGGCAGGATTCAC	516
ofas002344-RT7	taatacgactcactatagggagaCCTGTGGTTGAAGTTGTCGT CGTAGCAG	
ofas002807-F	GTGACCATAGAAGAAAATGGGATACAGAAG	447
ofas002807-RT7	taatacgactcactatagggagaCTTTCATCCTCTGTTCCAAGT CATAGTCTC	
ofas002923-F	GTCCGTAATTCTAGTGTGCCTAAGATGGAGC	733
ofas002923-RT7	taatacgactcactatagggagaCCACTTTTGGCATGTTCCAT TACTTGCTG	
ofas004060-F	GCAGATGACTGGGGTGGATG	960
ofas004060-RT7	taatacgactcactatagggagaCAGAGTCACAACCTCTCACGT CG	
ofas004876-F	GACTCCTGTTTCGACGTAAACGAAGGGAAAC	599
ofas004876-RT7	taatacgactcactatagggagaGGTAACCATTGTCTTCTTCA GAGGACCACACAG	
ofas005791-F	GGATTTCCAACCAGAAGAGGTGACGGTAAC	568
ofas005791-RT7	taatacgactcactatagggagaGACAACCATAGCTGCTTCCA CGAGG	
ofas008063-F	GAAATACTTGTACCCCTACGAATGTG	575
ofas008063-RT7	taatacgactcactatagggagaCATTATCTGGGTAATGTCTC TTAGGTG	

ofas008078-F	GTACCATCGCCGGCAGCCAAG	916
ofas008078-RT7	taatacgactcactatagggagaGCGTGTGGATGCCCTGCG	
ofas008654-F	CAGCACTACTGTTTACGTTGG	880
ofas008654-RT7	taatacgactcactatagggagaGATGATCCATGTCCATCGC	
ofas008658-F	GAGATTGGAGCCAAGGTGTTATAAGACG	334
ofas008658-RT7	taatacgactcactatagggagaCCTTTGAGCTAAGTCTAACG GGGC	
ofas009156-F	GACTAGCAAGCCCTAGACCTC	996
ofas009156-RT7	taatacgactcactatagggagaCTCCTCCATCTTCTGAGCCA C	
ofas009415-F	GACAGCAGAGGCACCTAAATCTG	771
ofas009415-RT7	taatacgactcactatagggagaCCATCCACAATAACTGAATG AGCC	
ofas009817-F	GAATTCTGGAGCAAATTCGAGGAG	866
ofas009817-RT7	taatacgactcactatagggagaCTGAGTTCTTCAAATGCGTT CC	
ofas010084-F	GCATATGAGGATAAGACACAGTGATG	842
ofas010084-RT7	taatacgactcactatagggagaCGAACTAATCTGTGCTAACT TATCTGC	
ofas010374-F	CAGACCAATCATCTTGTACTGTAGGAATATGGGC TG	801
ofas010374-RT7	taatacgactcactatagggagaCAGGTGGTTCATTTCCATAC CAAATTGTCTACTCC	
ofas011318-F	GTGCAAGTGCCTAAAGACTGCCCAATATC	851
ofas011318-RT7	taatacgactcactatagggagaGCGACACTCCAAGAGAACA CTCGACC	
ofas012523-F	CGAATTATAGCGGCAGAGTTGC	857
ofas012523-RT7	taatacgactcactatagggagaGAGAAGCTCAGTTGGATTGT TCAC	
ofas013445-F	CTCTTCTGTGATTACAGTGCTCCTTATGCAAGTAA C	641
ofas013445-RT7	taatacgactcactatagggagaGCAAGACTCAACTGTACTGT ATTCGCAATGTGG	

ofas015219-F	GACCTGCATCAAGATAGAGGAGC	957
ofas015219-RT7	taatacgactcactatagggagaGTCTGAATTATGGGCTGCTG AAC	
ofas016187-F	GTAAATAACACTGTAGATCCCCTGGCTTCAG	757
ofas016187-RT7	taatacgactcactatagggagaCTATAAGAGTTTCACTCAAT GGGCGAATGAGTTC	
ofas016401-F	GACTCAGTCAGGTGCTCGTCCTAATATTTGG	501
ofas016401-RT7	taatacgactcactatagggagaCCAGAGAGGAGAGATTTCGG TAACTGGC	
ofas016760-F	GGAAATTTAGAGACATTGATGTGGACACCTAACC AC	604
ofas016760-RT7	taatacgactcactatagggagaGCCACCATACTTCTTCCAA TAACACCAAC	
ofas016990-F	CGAAACAGTGACTIONTCGGCG	847
ofas016990-RT7	taatacgactcactatagggagaGAGGTCCGATCCGCTCTG	
ofas018237-F	CAGAGGACGAGTTTGGTGACAGTG	879
ofas018237-RT7	taatacgactcactatagggagaCGGTATGCTTAGACATAATG TGTCTCTTCATGC	
ofas018348-F	CCAGCATGACGGGAGGGGTG	1,025
ofas018348-RT7	taatacgactcactatagggagaCGCCGCTGTCTTCAACCATA GG	
ofas018531-F	GTCAGGTTACAGTTTCCCATG	722
ofas018531-RT7	taatacgactcactatagggagaGTCTTCTTAGGCTGTCCATG TC	
ofas018942-F	GAAGCGAAAGCACAGGAGTAGATC	405
ofas018942-RT7	taatacgactcactatagggagaGGAGGCTTGATAGTTCCATC GTC	
ofas019071-F	CCACCCTTCAAGGCAGAAATGC	436
ofas019071-RT7	taatacgactcactatagggagaGAAGTCCTTGGCAAATAAAT AGGTTCACTAGG	
ofas025035-F	GCAGGAGTACTGGACCTAC	553
ofas025035-RT7	taatacgactcactatagggagaCGTAGAACATTCGCCTCATG TG	

ofas025019-F	GAAAGTGTCAAATGTTTGTGGTTCCTC	303
ofas025019-RT7	taatacgactcactatagggagaCAAGAGACGGGTGGTACGATG	
ofas025096-F	CCTCTGAGAGTATGTACTATCCTCAG	462
ofas025096-RT7	taatacgactcactatagggagaCTTGTCGTTCTGACAATCCAAG	
ofas025230-F	CCTGTGAAAGAAGAGACACCTTTG	905
ofas025230-RT7	taatacgactcactatagggagaGTATGTTTGGCGAGTATGTGTCTATTC	

**Table S2. Primer sequences not related to expression pattern screen**

Primer sequences used to amplify probe template for in situ hybridization screen of genes co-expressed with *Of-E75A*.

Primer name	Primer Sequence	Notes
Of-Blimp-gDNaseq-F1	GCAATACAATGCAATAGGCTTG	Used for sequencing
Of-Blimp-gDNaseq-R2	GCAAATCTCACACTGATGAGG	
Of-Blimp-exon10-R	GTTGGCTGATACTCATAGTAGTATTGAG	
Of-Blimp-exon8-F	GTTCCAGTAAGCCCTGATTCAAC	Used for sequencing and screening
Cas9-mRNA-F-T7	taatacgactcactatagggagaGGTGGGAGGTCTATATAAGCAG	To amplify template for <i>Cas9</i> mRNA; amplicon: 4,300 bp
Cas9-mRNA-R	GCTGATCAGCGGGTTTAAACTC	
Of-Blimp-gRNA-A	ttaatacgactcactataggTCAAGGATGGATGGTTGCTGgttttagagctagaaatag	Used with gRNArev to amplify template for <i>Of-Blimp1</i> gRNA-A
Of-Blimp-gRNA-B	ttaatacgactcactataggTAAGGGTAGTCGGA GTATGCgttttagagctagaaatag	Used with gRNArev to amplify template for <i>Of-Blimp1</i> gRNA-B
gRNArev	AAAAGCACCGACTCGGTGCC	Used to amplify templates for gRNA synthesis
Of-Blimp-screening-F1	CAGTTCTCGTAAGCCCAGAGCC	Used for screening
Of-Blimp-screening-R1	ATCTCAGTAGGACTACTCGGCGG	
Of-Blimp-screening-R2	CCGAGATCCTGTAGGTGAGAGTG	
Dmel-Blimp-gRNA-F	TATATAGGAAAGATATCCGGGTGAAC TTCgtggttgatgcatgcaGTTTTAGAGCTA GAAATAGCAAG	Used to construct gRNA expression plasmid
Dmel-Blimp-gRNA-R	ATTTTAACTTGCTATTTCTAGCTCTAA AACtctacgtgattcgtatgacGACGTTAAATTG	

	AAAATAGGTC	
Dmel-Blimp-5-HDR-F	cgacggccagtgaattcgagctcggtacccGTGGGTC AGTAGTAGGAACTCTC	To amplify 5' homology arm for <i>Dmel-Blimp1</i> HDR template; amplicon: 1,097 bp
Dmel-Blimp-5-HDR-R	gctcgaattaacatGGCGACTATAGCTAGAT TGTTGCTCAG	
Dmel-Blimp-3-HDR-F	catcaatgtatcttaAAGGTACATGGTACATGG TATATGGAG	To amplify 3' homology arm for <i>Dmel-Blimp1</i> HDR template; amplicon: 942 bp
Dmel-Blimp-3-HDR-R	atgcctgcaggtcgactctagaggatccccGCTAGGC GATACTAGTTCAACAGATG	
Dmel-Blimp-HDR-insert-F	ctagctatagtcgccATGGTTAATTCGAGCTC GCC	To amplify <i>3XP3&gt;EGFP&gt;SV40</i> transgene from plasmid; amplicon: 1,245 bp
Dmel-Blimp-HDR-insert-R	tgtaccatgtaccttTAAGATACATTGATGAGT TTGGACAAAC	
ofas008150-F	GCAGTGTATCTAGTACTGGACC	To amplify <i>Of-Blimp1</i> ( <i>OFAS008150</i> ) probe template; amplicon: 1,164 bp
ofas008150-RT7	taatacgactcactatagggagaGTAGTCGGAGTA TGCAGGAG	
ofas008150-FT7	taatacgactcactatagggagaGCAGTGTATCTA GTAAGGACC	
OfE75AF_New	tcaagagggactccatacac	To amplify <i>Of-E75A</i> probe template; amplicon: 413 bp
OfE75ART7_New	taatacgactcactatagggagaTGTACCAGGACT CTTGCTCT	
Of-slp F2	CTTACAGCTACAACGCCCTC	To amplify <i>Of-slp</i> probe template; amplicon: 1,476 bp
Of-slp3UTR-R1T7	taatacgactcactatagggagaTGAGGAATGTGA CGACTTTAGG	
Of-enRT7	taatacgactcactatagggagaTGTCTTCCTTGCT CTTGCTCT	To amplify <i>Of-inv</i> probe template

Of-enF	aatcggatgtagtgaggatg	(primers were named before gene identified as <i>inv</i> ; forward primer is found closest to <i>inv</i> in genome, reverse primer is not specific to either gene); amplicon:
Of-Ubx-F	GGATTCTACGGTTCACATCACC	To amplify <i>Of-Ubx</i> probe template; amplicon: 544 bp
Of-Ubx-RT7	taatacgactcactatagggagaGATTGGCAGGCTGTTGATGG	
Of-lab-F	GTGAGCAGGAGAACTACTGCCAG	To amplify <i>Of-lab</i> probe template; amplicon: 712 bp
Of-lab-RT7	taatacgactcactatagggagaGAGGTGCTGGAGTGTGAGAG	
Of-AbdB-F	GACCGAGCGGCAGGTGAAG	To amplify Of-AbdB probe template; amplicon: 602 bp
Of-AbdB-RT7	taatacgactcactatagggagaGTTGAATGAGGCCTGCGG	
Dmel-Blimp1-F	GAGCTAATGGTTTGGTACTGCAAG	To amplify Dmel-Blimp1 probe template; amplicon: 798 bp
Dmel-Blimp1-RT7	taatacgactcactatagggagaCTCATTCTTGTCCGTTTCGCAAATGG	
Of-Blimp1-dsRNA2-FT7	taatacgactcactatagggagaGCAGCAGCATGTGGCTAGGAAATG	To amplify Of-Blimp1 dsRNA #2 template; amplicon: 626 bp
Of-Blimp1-dsRNA2-RT7	taatacgactcactatagggagaACCAAGAGGAAC TTCAGTTGGCTG	
turbo GFP 1R_T7_2	ttaatacgactcactataggCTCGGTGTTGCTGTGATCCTC	To amplify <i>tGFP</i> control dsRNA; amplicon: 636 bp
T7-tGFP-F	taatacgactcactatagggagaATGGAGAGCGACGAGAGC	
Of-Blimp1-upstream-2	GTCATGGTAATTTAGAGTGAAGATCATAGAAG	To amplify more complete Blimp1 cDNA; amplicon: 3,539 bp
Of-Blimp1-downstream-2	CAGTCGATTTGATTGGTAGCAG	



Of-E75a-3-RT7	taatacgactcactatagggagaGCAAGCGCGGCA GAGTACTG	Used to amplify <i>Of-E75A</i> - RT-PCR to determine correct staging for RNA-seq; Amplicon: 366 bp, but T7 promoters add an additional 46 bp (band size = 412 bp)
Of-E75a-F-T7-2	taatacgactcactatagggagaTCGGCACTCGCC CGGTTATCA	
F_Of_actin	ATGGTCGGTATGGGACAGAA	Used to amplify <i>Of-actin</i> as positive control for RT-PCR; Amplicon: 173 bp
R_Of_actin	TGTTCTTCAGGGGCAACTCT	

**Data S1. (separate file)**

Sanger-sequenced DNA fragments

**Data S2. (separate file)**

Data set including all *Oncopeltus* gene models found to be differentially expressed and passed to WGCNA for clustering into co-expression modules, as well as their module membership.