

Supporting Information for

Modeling extrahepatic hepatitis E virus infection in induced human primary neurons

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SI References

Other supporting materials for this manuscript include the following:

Movies S1

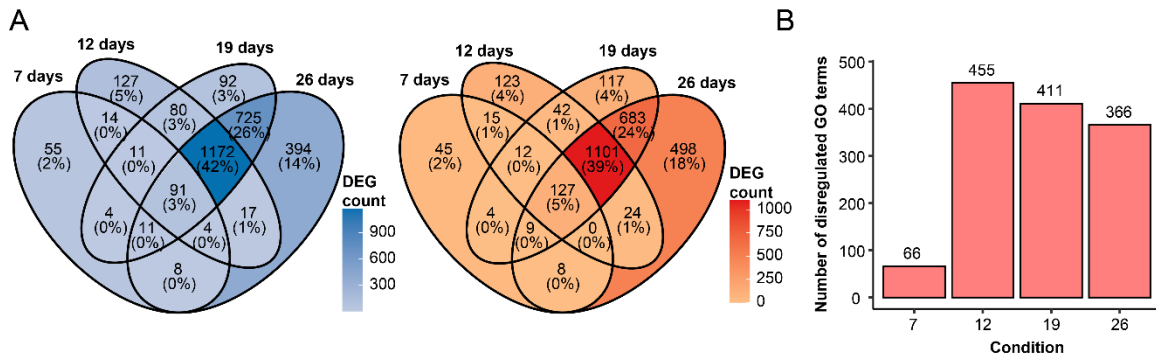


Fig. S1. Regulation of genes during the neuron differentiation process. (A) Venn diagram of significant down-regulated differentially expressed genes (DEGs, blue) and up-regulated DEGs (red) at different stages of differentiation compared to the earliest time point of 4 days. **(B)** Overview of the number of significant deregulated Gene ontology (GO) terms during the differentiation process.

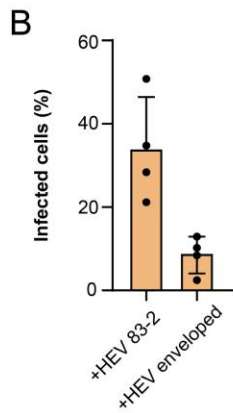
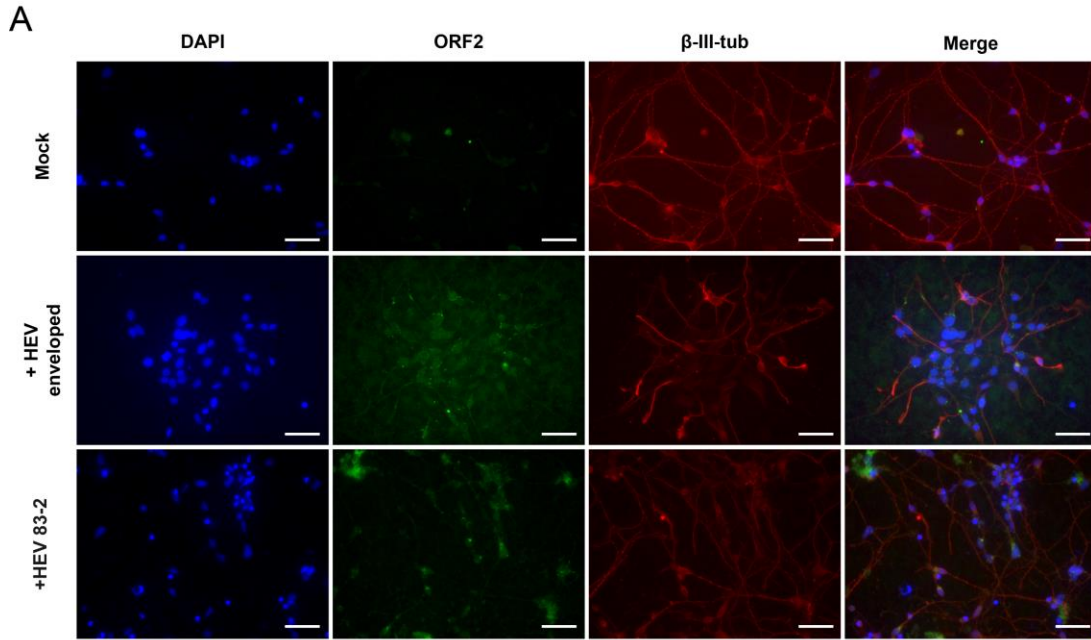


Fig. S2. Infection of human induced primary neurons with quasi-enveloped HEVcc Kernow-C1 p6 strain and wild boar HEV-3 strain 83-2. (A) Immunofluorescence staining was performed for uninfected (Mock) and infected induced primary neurons (iPNs) with quasi-enveloped Kernow-C1 p6 strain and the wild boar HEV-3 strain 83-2. 4',6-diamidino-2-phenylindole (DAPI) was used to stain the nuclei (blue), a polyclonal rabbit anti-HEV antiserum to stain the ORF2-encoded capsid protein (green) and an anti- β -III-tubulin (β -III-Tub) antibody to stain the neuronal cytoskeleton. Scalebars represent 50 μ m. (B) The susceptibility of neurons to both HEV strains was quantified by measuring the ORF2 signal using CellProfiler.

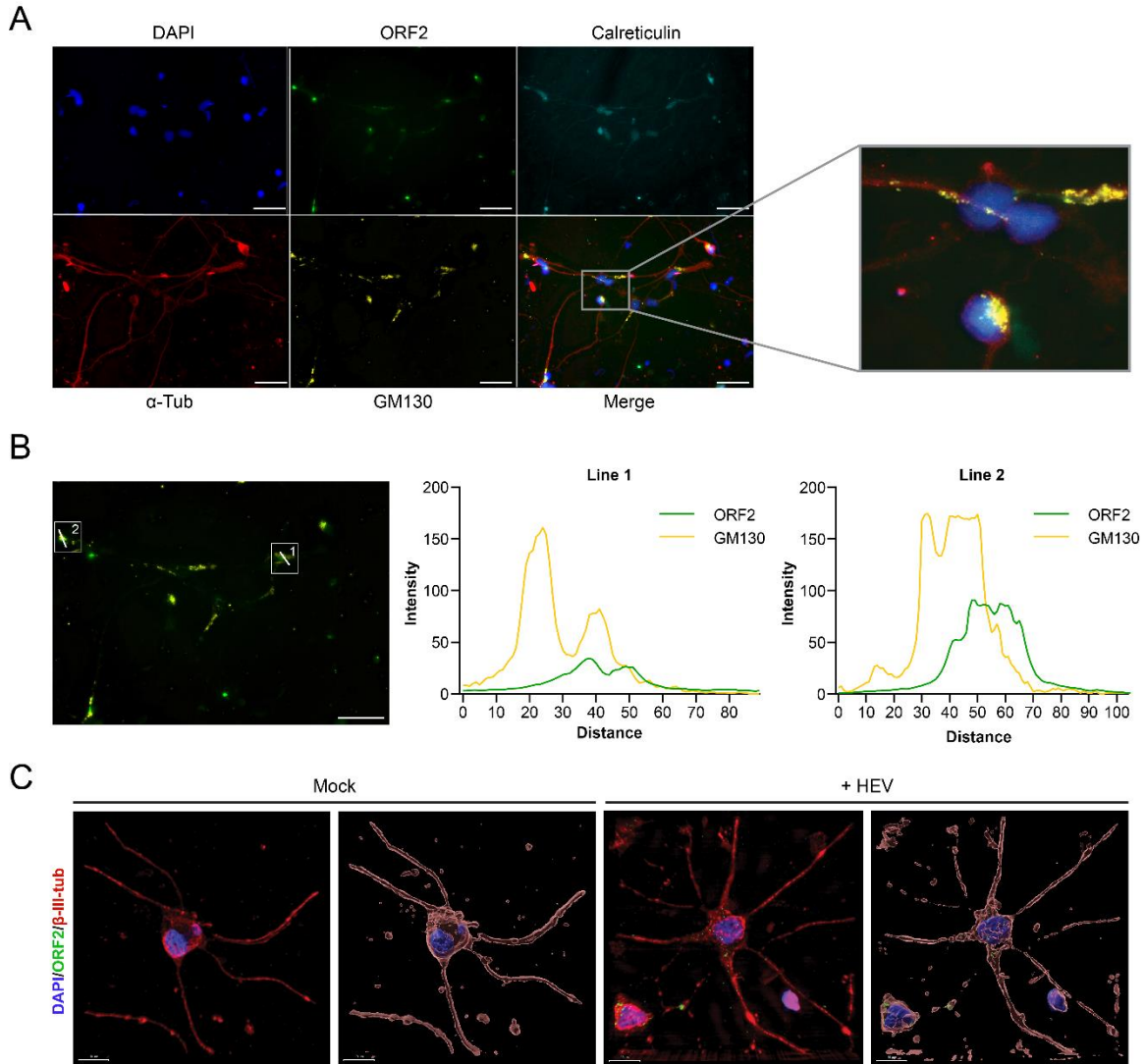


Fig. S3. The ORF2 encoded capsid protein of HEV is colocalized with the Golgi apparatus. **(A)** Virus localization was accessed by iterative indirect immunofluorescence imaging using multiple compartment markers. In different rounds of staining, antibodies of ORF2 (green), α -tubulin (α -Tub, red), GM130 (yellow) and calreticulin (cyan) were used to stain the HEV capsid protein, cytoskeleton, Golgi apparatus and the endoplasmic reticulum, respectively. The nucleus was stained by DAPI. White scalebar represents 50 μ m **(B)** Line graphs show the fluorescence intensities of ORF2 capsid protein (green) and Golgi apparatus (GM130, yellow) measured across the regions of interest indicated by the white lines in the immunofluorescence image. White scalebar represents 50 μ m. **(C)** Confocal microscope images of uninfected iPNS (Mock) and HEV-infected neurons were acquired. A 3D reconstruction of the surfaces was performed using the analysis tool Imaris 9.8.0 and a video was also generated. Cells were stained with DAPI (blue), with an antibody against ORF2-encoded capsid protein (green) and with β -III-tubulin (red). White scalebar represents 10 μ m.

A

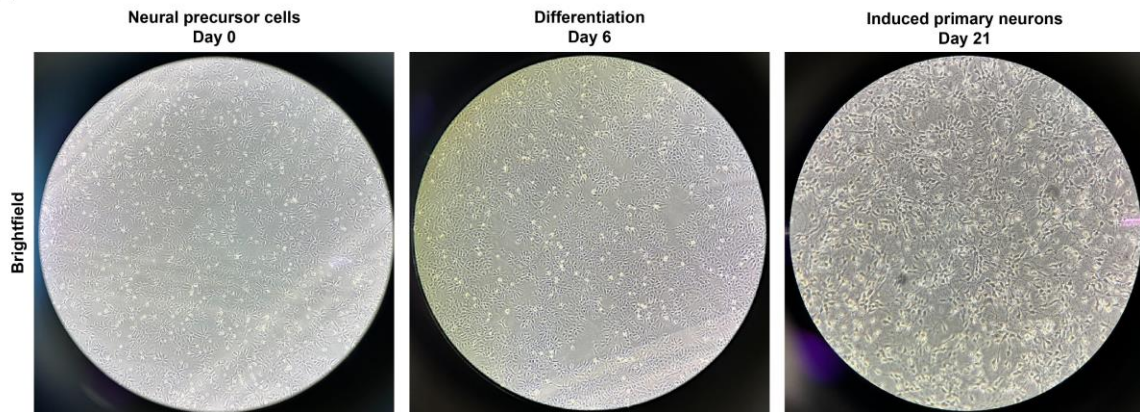


Fig. S4. Neurite development during differentiation into induced primary neurons. Brightfield images were taken at three differentiation timepoints: neural precursor cells (day 0), cells during differentiation (day 6), and induced primary neurons after 21 days of differentiation.

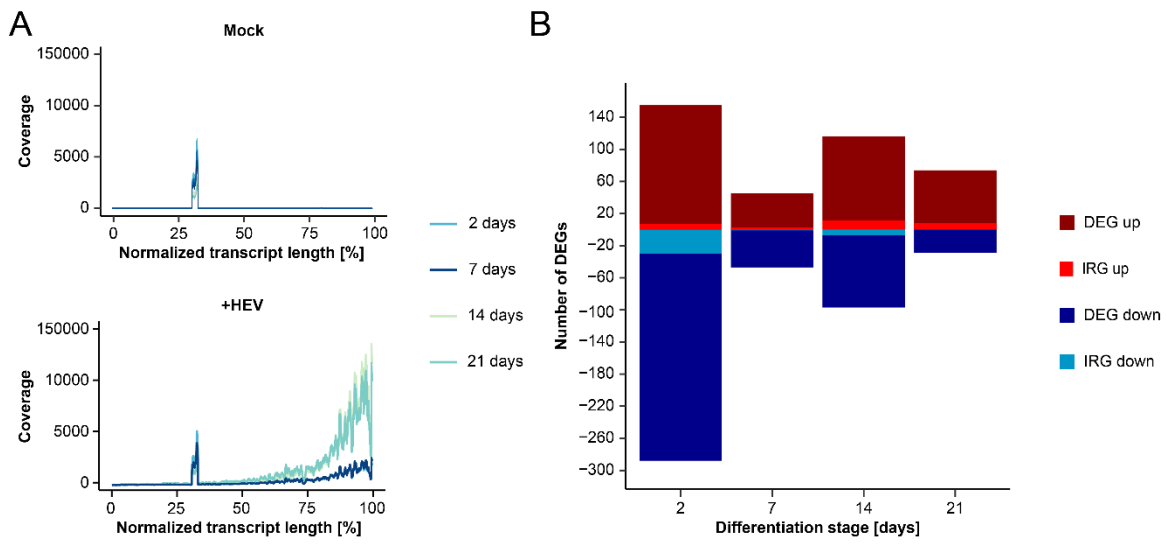


Fig. S5. General response of primary neurons to HEV infection. (A) Normalized coverage of mapped reads along the HEV genome in uninfected (Mock) and infected neurons (+HEV, five days post-infection) at the timepoints of 2, 7, 14 and 21 days of differentiation. **(B)** The number of differentially expressed genes (DEGs) by HEV infection compared to uninfected cells is shown over the course of differentiation. An online single-cell database of interferon (IFN)-regulated genes (IRGs) (<http://isg.data.cvr.ac.uk/>) was utilized, filtered for genes significantly up- or downregulated ($FDR \leq 0.05$ and \log_2 fold change of 2 or -2) in humans, and were compared to the identified DEGs. Lighter colors indicate the number of IRGs that were significantly upregulated (red) or downregulated (blue).

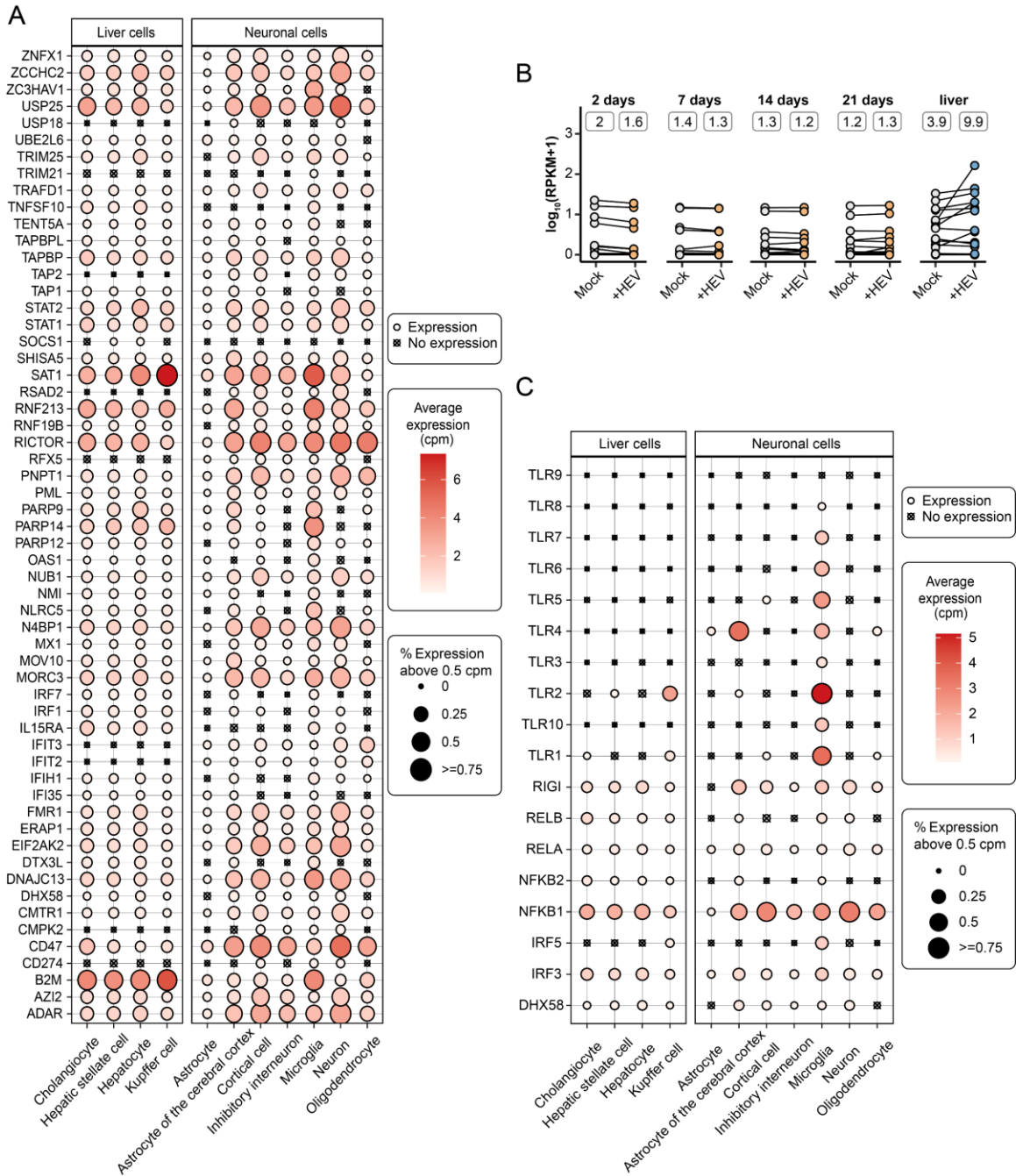


Fig. S6. Pathogen sensing in primary neurons and interferon-stimulated gene (ISG) expression across nervous system cell types. (A) Expression of ISGs in neuronal and liver cells in publicly available single-cell databases. **(B)** Overview and comparative analysis of the expression levels of pathogen-sensing genes measured in RPKM values, in both uninfected (Mock) and HEV-infected (+HEV) induced primary neurons and primary human hepatocytes (liver). The values above displayed represent the mean RPKM expression levels of all genes under each respective condition. **(C)** Expression of sensing genes in neuronal and liver cells in publicly

available single-cell databases. cpm: counts per million. RPKM: Reads per kilobase of transcript per million mapped reads.

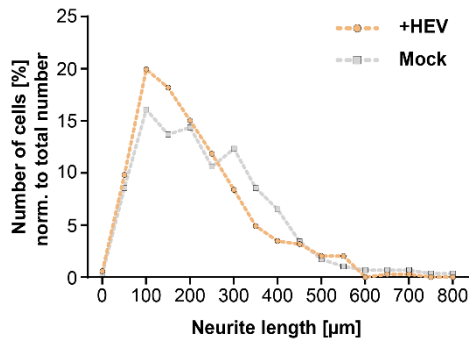


Fig. S7. Shift in neurite length after HEV inoculation. The neurite length of mock-inoculated (Mock) and HEV-inoculated (+HEV) iPNS (21-day-old cells) was examined through microscopic analysis five days post-inoculation. The frequency distribution is depicted as the percentage of cells (normalized to the total cell number) with a given neurite length.

DEG: ■ Downregulated ■ Unchanged ■ Upregulated



Fig. S8: Differential regulation of genes in 21-day-old neurons relative to various stages of differentiation. The genes presented display either upregulation or downregulation in 21-day-old primary neurons following HEV infection. These differentially regulated genes (DEGs) are depicted here in comparison with their expression patterns in 2, 7, and 14-day-old cells.

Table S1. Association between upregulated human genes at 21 days post-HEV infection and neurite length. Related genes include only genes of the same family. / indicating no correlation with neurite outgrowth.

Gene name (explanation)	Involved in neurite outgrowth regulation	Reported potential function of the encoded protein	References	Log ₂ fold change	False discovery rate (FDR)	Mock: Reads per kilobase of transcripts per million mapped reads	+HEV: Reads per kilobase of transcripts per million mapped reads
<i>MOV10</i> (MOV10 RISC complex RNA helicase)	yes	Knockout of <i>MOV10</i> gene in murine neuroblastoma (Neuro2a) cells lead to reduced neurite length, which could be rescued upon re-induction of <i>MOV10</i> gene	(1, 2)	1.340118	4.392000e-09	0.58869067	1.49156581
<i>ATF3</i> (Activating transcription factor 3)	yes	ATF3 promotes neurite outgrowth in rat dorsal root ganglion neurons	(3, 4)	1.055239	2.749000e-06	1.73958633	3.61697248
<i>TREX1</i> (Three prime repair exonuclease 1)	yes	A <i>TREX1</i> mutant (V91M) lead to a significant reduction in axonal outgrowth in mouse primary cortical neurons compared to the wildtype cells	(5)	1.654444	2.098385e-02	0.2436382	0.7727430
<i>TGFB1</i> (Transforming growth factor beta induced)	yes	TGFB increased neurite outgrowth in scratched primary midbrain	(6–9)	1.148161	7.208000e-14	4.5132269	10.0050609

		cultures of dopaminergic cells (Wistar rats); TGFBI rescued neurite outgrowth in inhibitory environment via canonical ALK5/SMAD3 signaling in mouse N1E-115 neuron-like cells; TGF- β 31 has a capability of promoting axonal regeneration in rat hippocampal neurons					
<i>ARAP3</i> (ArfGAP with RhoGAP domain, ankyrin repeat and PH domain 3)	yes	Neurites outgrowth of rat pheochromocytoma (PC12) cells was affected by <i>ARAP3</i> under NGF or bFGF treatment	(10, 11)	1.355979	2.495000e-13	1.078767	2.762672
<i>VEGFA</i> (Vascular endothelial growth factor A)	yes	VEGF enhanced axonal/neurite outgrowth of mouse dorsal root and superior cervical ganglia, rat retinal ganglion cells and rat cerebral cortical neurons	(12–16)	1.027099	2.351000e-06	1.310088	2.671190
<i>SERPINE1</i> (Serpin family E member 1)	yes	PAI-1 (plasminogen activator inhibitor-1, Serpine1), an endogenous inhibitor of tissue plasminogen activator (tPA), led to a decrease neurite length	(17, 18)	1.710986	1.403000e-09	0.54960125	1.80225088

		of rat primary neuronal progenitor cells					
<i>CAVI</i> (Caveolin 1)	yes	Caveolin 1 was required for axonal outgrowth of motor neurons (from <i>Xenopus</i>); synapsin-Calveolin-1 enhanced axonal growth in differentiated NPCs derived from human iPSCs	(19, 20)	1.406676	3.374000e-04	0.3073646	0.8167771
<i>ENG</i> (Endoglin)	yes	Endoglin is a co-receptor of transforming growth factor β (TGF- β), thus it participates in TGF- β signaling pathway	(21)	1.130649	1.031000e-12	3.97019850	8.69529148
<i>PLAU</i> (Plasminogen activator, urokinase)	yes	Inhibition of uPA (another abbreviation for PLAU) gene expression reduced axonal length in Neuro2 cells and mouse dorsal root ganglia	(22, 23)	2.015380	1.463000e-10	0.44035605	1.78463341
<i>FOSL1</i> (FOS like 1, AP-1 transcription factor subunit)	yes	FOSL1 siRNA injection in rats led to reduced axon growths in Schwann cells	(24)	4.547930	6.536000e-04	0.03115952	0.77415355
<i>NPAS4</i> (Neuronal PAS domain protein 4)	yes	<i>NPAS4</i> gene knockdown in Neuro2a cells inhibited neurite outgrowth; In <i>NPAS4</i> knockout hippocampal	(25)	1.423217	3.957000e-04	0.31114766	0.83648270

		neurons (mouse) neurite outgrowth was abolished					
<i>NNMT</i> (Nicotinamide N-methyltransferase)	yes	NNMT stable expression in SH-SY5Y cells reduced the neurite length compared to SH-SY5Y cells without NNMT, whereas the number of neurites and branches increased	(26)	2.016775	1.567549e-02	0.1095445	0.4487772
<i>ROBO3</i> (Roundabout guidance receptor 3)	yes	The lack of ROBO3 in mouse led to a decreased axon outgrowth in spinal cord explants	(27)	1.433759	4.375000e-30	17.05178	46.07125
<i>CSPG4</i> (Chondroitin sulfate proteoglycan 4)	yes	CSPG reduced neurite length in mouse cerebral cortical neurons and granule cells in a dose-dependent manner; led also in human-derived SH-SY5Y cells to reduction of neurite length; CSPG reduced neurite length in mouse cerebellar granule cells in a dose-dependent manner; NG2 (synonym for CSPG4) also inhibited neurite growth from embryonic rat	(28–32)	1.128576	3.951000e-06	0.40782702	0.89232606

		dorsal root ganglia neurons					
<i>ACSF2</i> (Acyl-CoA synthetase family member 2)	yes	ACSF2-deficient Neuro2a cells showed significantly blunted neurite outgrowth in response to retinoic acid; Acyl-CoA synthetase 2 overexpression enhanced neurite outgrowth in PC12 cells	(33, 34)	1.136887	2.484000e-03	0.5538275	1.2198368
<i>GADD45B</i> (Growth arrest and DNA damage inducible beta)	yes	Mutated GADD45B mice showed impaired activity-dependent neurogenesis and dendritic development; GADD45G/p38 MAPK/CDC25B signaling pathway enhanced neurite outgrowth by promoting microtubule polymerization	(35–37)	2.575122	1.250000e-14	0.5593153	3.3446735
<i>GDF15</i> (Growth differentiation factor 15)	yes	The neurites length of the primary culture of rat retinal ganglion cells were increased by GDF15 treatment	(38)	1.549360	2.145562e-02	0.20693118	0.60944003
<i>PLAUR</i> (Plasminogen activator, urokinase receptor)	yes	Exogenous uPA (PLAU) increased neurite length in mouse Neuro2a cells	(23, 39)	1.597618	8.919000e-05	0.2644716	0.8027418

		most likely via the interaction with uPAR (PLAUR); blocking of uPAR stimulated neurite formation					
<i>CCDC8</i> (Coiled-coil domain containing 8)	yes	CCDC8 increased the number of neurites per cell in rat PC12 cells	(40)	1.014808	3.547000e-07	1.96235660	3.96682179
<i>LIF</i> (LIF interleukin 6 family cytokine)	yes	LIF promoted neurite outgrowth of rat mammalian auditory neurons in vitro; LIF promoted neurite growth of P12-E2 cells	(41, 42)	1.708004	1.181000e-03	0.14429893	0.47372902
<i>HSPA6</i> (Heat shock protein family A (Hsp70) member 6)	Related genes	HSP70 alone and especially in combination with HSP40 increased neurite outgrowth in Neuro2a cells; Hsp27 is involved in GDNF-induced neurite outgrowth in rat PC12-GFRR1-RET cells; Neurite outgrowth was mediated by the heat shock protein HSP90a	(43–45)	5.420693	4.269000e-49	0.23718946	10.22849984
<i>CNPY1</i> (Canopy FGF signaling regulator 1)	Related genes	CNPY2 (MSAP) increased the number of neurites on mouse N2-A and rat PC12 cells	(46)	1.105491	1.345000e-06	0.9920355	2.1358922

<i>TNFRSF10D</i> (TNF receptor superfamily member 10d)	Related genes	Tumor necrosis factor (TNF) decreased the total neurite length per cell, the axonal length and the number of branch points of primary mice hippocampal neurons cocultured with astrocyte-enriched glial cells	(47)	1.009861	6.689000e-05	0.9730365	1.9607246
<i>CA2</i> (Carbonic anhydrase 2)	Related genes	CA8 induced neurite outgrowth in Neuro-2a cells but not in human SK-N-SH cells	(48)	1.173081	1.356000e-05	1.55652100	3.51307191
<i>PRDM12</i> (PR/SET domain 12)	Related genes	Disruption of the PRDM14 gene resulted in axon growth defects in zebrafish	(49)	1.605972	2.794000e-13	1.303463	3.971309
<i>PLD5P1</i> (Phospholipase D family member 5 pseudogene 1)	Related genes	Phospholipase D1 inhibited neurite outgrowth of rat neural stem cells via increase of synapsin 1 expression; Overexpression of PLD1 resulted in neurons to increase tissue plasminogen activator (tPA) release and therefore to tPA-dependent neurite extension	(50, 51)	1.077125	2.514724e-02	0.2453015	0.5186789

<i>ADAMTS14</i> (ADAM metalloproteinase with thrombospondin type 1 motif 14)	Related genes	ADAMTS4 and ADAMTS5 led to an increase in neurite length in rat primary neurons	(52)	1.414671	3.603000e-06	0.3116316	0.8320400
<i>SLC15A3</i> (Solute carrier family 15 member 3)	Related genes	SLC25A12 expression was associated with dendrites outgrowth in mouse embryonic cortical neurons	(53)	1.547206	7.039000e-03	0.1887240	0.5542340
<i>SLC38A4</i> (Solute carrier family 38 member 4)	Related genes	SLC25A12 expression was associated with dendrites outgrowth in mouse embryonic cortical neurons	(53)	2.649440	2.497000e-05	0.07092644	0.44991230
<i>RNF135</i> (Ring finger protein 135)	Related genes	Molluscan RING-finger protein L-TRIM is essential for neuronal outgrowth	(54)	1.374997	4.669022e-02	0.1963424	0.5120273
<i>VMAC</i> (Vimentin type intermediate filament associated coiled-coil protein)	Related genes	The length of astrocytic processes after entorhinal cortex lesion was shorter in GFAP ^{-/-} Vim ^{-/-} than wildtype mice (Vim=Vimentin)	(55)	1.107776	3.657000e-03	0.6299788	1.3598131
<i>DKK1</i> (Dickkopf like acrosomal protein 1)	Related genes	Dickkopf-1 protein (Dkk1) induced neurite outgrowth in TC-32 cells	(56)	1.540152	2.998727e-02	0.2884431	0.8444769
<i>SNAIL</i>	Related genes	SNAIL transcription factor in prostate cancer	(57)	4.665358	6.446000e-06	0.05172846	1.36721883

(Snail family transcriptional repressor 1)		cells promoted neurite outgrowth					
<i>RSC1A1</i> (regulator of solute carriers 1)	/	/	/	2.927757	3.623000e-04	0.07599916	0.58922164
<i>ID3</i> (Inhibitor Of DNA Binding 3)	/	/	/	1.748242	3.465000e-30	11.07878444	37.23229514
<i>ACY1</i> (Aminoacylase 1)	/	/	/	1.214575	5.344000e-03	0.4597700	1.0693794
<i>AHSG</i> (Alpha 2-HS glycoprotein)	/	/	/	1.638499	6.277000e-04	0.4287744	1.3402270
<i>CYTL1</i> (Cytokine like 1)	/	/	/	1.380700	1.976895e-02	0.5647939	1.4770756
<i>ALB</i> (albumin)	/	/	/	2.926996	1.716000e-22	0.45646804	3.48247791
<i>ARRDC3</i> (Arrestin domain containing 3)	/	/	/	1.051950	6.683000e-14	8.266954	17.142819
<i>PRDM13</i> (PR/SET domain 13)	/	/	/	1.489413	5.248000e-03	0.19102079	0.53853983
<i>NUDT18</i> (Nudix hydrolase 18)	/	/	/	1.008633	7.463000e-03	0.8883395	1.7897523
<i>COL13A1</i> (Collagen type XIII alpha 1 chain)	/	/	/	1.657751	1.174000e-11	0.6850415	2.1640083
<i>NUDT8</i> (Nudix hydrolase 8)	/	/	/	1.076710	3.940508e-02	0.8883395	1.7897523

<i>GSX1</i> (GS homeobox 1)	/	/	/	1.575707	5.544000e-03	0.2902809	0.8695302
<i>NPIA8</i> (Nuclear pore complex interacting protein family member A8)	/	/	/	1.261234	4.423000e-03	0.59556003	1.43100559
<i>SLX1B</i> (SLX1 homolog B, structure-specific endonuclease subunit)	/	/	/	5.257244	3.554000e-20	0.1814754	7.0508826
<i>TMEM100</i> (Transmembrane protein 100)	/	/	/	1.055878	2.705273e-02	0.37648448	0.78435276
<i>IDI</i> (Inhibitor Of DNA Binding 1)	/	/	/	2.002127	7.104000e-13	1.28754784	5.16763625
<i>PABPCIL</i> (Poly(A) binding protein cytoplasmic 1 like)	/	/	/	1.165967	2.460000e-04	0.60324994	1.35526644
<i>COL9A3</i> (Collagen type IX alpha 3 chain)	/	/	/	1.177167	3.118000e-03	0.4791457	1.0854985
<i>GGTI</i> (Gamma-glutamyltransferase 1)	/	/	/	1.093269	4.897000e-03	0.34972246	0.74733914
<i>HMOX1</i> (Heme oxygenase 1)	/	/	/	1.390076	7.152000e-06	0.8629264	2.2650443
<i>FAM156A</i>	/	/	/	1.007986	1.842000e-10	2.654675	5.340031

(Family with sequence similarity 156 member A)							
<i>RENBP</i> (Renin binding protein)	/	/	/	1.017142	3.818000e-03	1.112195	2.253698
gene: <i>ENSG00000271741</i>	/	/	/	2.823195	1.416000e-05	0.0853795	0.6117999
gene: <i>ENSG00000254706</i>	/	/	/	1.233511	1.727228e-02	0.6527534	1.5394736
<i>HSPE1-MOB4</i> Readthrough	/	/	/	2.099540	1.567000e-03	0.3633585	1.5715336
gene: <i>ENSG00000284820</i>	/	/	/	1.405860	6.634000e-05	0.2522317	0.6696288
<i>STIMATE-MUSTN1</i> Readthrough	/	/	/	1.252719	4.557356e-02	0.3728846	0.8921909
gene: <i>ENSG00000286001</i>	/	/	/	1.012625	2.275000e-03	0.5374444	1.0855183
gene: <i>ENSG00000289027</i>	/	/	/	5.189919	2.232000e-12	0.02737338	1.02447209
gene: <i>ENSG00000284292</i>	/	/	/	1.213351	2.323063e-02	0.2717099	0.6319702
gene: <i>ENSG00000288640</i>	/	/	/	1.230503	4.934000e-09	1.4441618	3.3907027
gene: <i>ENSG00000254536</i>	/	/	/	1.179328	2.286894e-02	0.3346430	0.7600015
<i>ZFP91-CNTF</i> Readthrough	/	/	/	1.918956	8.172000e-04	0.1824514	0.6944137
gene: <i>ENSG00000285901</i>	/	/	/	2.267598	7.375000e-15	0.3936985	1.9001755

<i>ATF7-NPFF</i> Readthrough	/	/	/	3.141335	1.104214e-02	0.05481481	0.50224828
gene: <i>ENSG00000258830</i>	/	/	/	1.653482	2.940749e-02	0.15348320	0.48679998
gene: <i>ENSG00000273167</i>	/	/	/	1.298390	8.305000e-03	0.1909486	0.4710442
<i>ARHGAP11A-SCG5</i> Readthrough	/	/	/	1.304814	6.584000e-04	0.3203187	0.7929361
gene: <i>ENSG00000173867</i>	/	/	/	1.029718	4.505890e-02	0.2787915	0.5704761
gene: <i>ENSG00000255439</i>	/	/	/	3.833119	9.724000e-09	0.07577064	1.09860765

Table S2. Association between downregulated human genes at 21 days post-HEV infection and neurite length. Related genes include only genes of the same family. / indicating no correlation with neurite outgrowth.

Gene name (explanation)	Involved in neurite outgrowth regulation	Reported potential function of the encoded protein	References	Log ₂ fold change	False discovery rate (FDR)	Mock: Reads per kilobase of transcripts per million mapped reads	+HEV: Reads per kilobase of transcripts per million mapped reads
<i>BCAN</i> (Brevican)	yes	Brevican inhibits neurite outgrowth from rat cerebellar granule neurons; Brevican is an inhibitor of neurite outgrowth in the proteoglycan-enriched myelin fraction in cerebellar granule cells	(58, 59)	-1.358337	1.465000e-21	13.3752915	5.2167392
<i>PCP4L1</i> (Purkinje cell protein 4 like 1)	yes	PCP4 enhance neurite outgrowth, while a knockdown reduces neurite outgrowth in PC12 cells	(60)	-1.344553	1.421000e-04	2.53007119	0.99478298
<i>RGS5</i> (Regulator of G protein signaling 5)	yes	RGS5 leads to an inhibitory effect of neurite outgrowth by sonic hedgehog in mouse primary cortical neurons; RGS5 switched astrocytes from neuroprotective to	(61, 62)	-1.426282	8.729000e-10	1.3988044	0.5201488

		pro-inflammatory property via binding to the receptor TNFR2					
<i>PITX2</i> (Paired like homeodomain 2)	yes	Loss of PITX2 disrupts axonal growth in the dorsal midbrain in mouse	(63)	-1.336801	3.957000e-04	0.60078189	0.23744417
<i>PENK</i> (Proenkephalin)	yes	Proenkephalin produces the peptide [Met]enkephalin and [Leu]enkephalin via proteolytic cleavage; [Met]enkephalin and [Leu]enkephalin both has a neurite growth-promoting effect in rat dorsal root ganglia	(64)	-1.529499	1.205000e-10	2.83594972	0.98163152
<i>CDC42EP2</i> (CDC42 effector protein 2)	yes	bind to, and negatively regulate the function of CDC42; CDC42 stimulates neurite outgrowth in primary chick spinal cord neurons ; Cdc42 plays a l role in neurite outgrowth of PC12 cells and cerebellar granule neurons	(65, 66)	-2.061407	8.766000e-05	0.9645701	0.2298073
<i>NTF3</i> (Neurotrophin 3)	yes	NTF3 promotes neurite outgrowth in rat hippocampal cells and sensory neurons	(67–69)	-1.199443	3.067719e-02	1.17344583	0.50953183

<i>ADORA2A</i> (Adenosine A2a receptor)	yes	An ADORA2A antagonist rescued the decreased neurite length in rat primary cortical neurons from an attention deficit and hyperactivity disorder model; the suppression of ADORA2A seems to correlate with neurite outgrowth promoted by ultrasound stimulation in PC12 cells	(70, 71)	-1.614159	8.454000e-06	1.0091193	0.3289512
<i>SNCB</i> (Synuclein beta)	Related genes	Alpha-Synuclein reversed the SPTBN1-induced neurite over-branching in SH-SY5Y cells; Beta-synuclein is discussed as a natural negative regulator of alpha-synuclein aggregation	(72, 73)	-1.033695	2.206000e-05	4.7782508	2.3330362
<i>PHOX2A</i> (Paired like homeobox 2A)	Related genes	PHOX2B knockdown reduced neurite length in human motor neurons and induced short spinal neurons in zebrafish	(74)	-2.067991	3.509352e-02	0.4672003	0.1097638
<i>SEZ6L</i> (Seizure related 6 homolog like)	Related genes	SEZ6 suppression leads to decreased neurite length in PC12 cells; SEZ6 family member are	(75, 76)	-1.166480	2.178000e-07	1.35654181	0.60410517

		known to affect neurite length (SEZ6 and SEZ6L2)					
<i>SYNPR</i> (Synaptoporin)	/	/	/	-1.002154	4.107603e-02	0.66902816	0.33344849
<i>MAFA</i> (MAF bZIP transcription factor A)	/	/	/	-1.084093	4.790000e-07	3.8772766	1.8283201
<i>ADRA2A</i> (Adrenoceptor alpha 2A)	/	/	/	-1.381024	2.780000e-08	1.8795963	0.7211471
<i>NKX1-2</i> (NK1 homeobox 2)	/	/	/	-1.445285	4.276000e-08	2.00510214	0.73565542
<i>NPIA3</i> (Nuclear pore complex interacting protein family member A3)	/	/	/	-1.304036	2.042396e-02	0.7830245	0.3160825
<i>SULT1A4</i> (Sulfotransferase family 1A member 4)	/	/	/	-2.107897	7.278000e-06	1.7434995	0.4025545
<i>BOLA2B</i> (BolA family member 2B)	/	/	/	-1.307911	1.149725e-02	1.4269165	0.5746954
<i>PRSS53</i> (Serine protease 53)	/	/	/	-1.794549	2.217000e-07	1.8605236	0.5351388
<i>DIPK1C (FAM69C)</i>	/	/	/	-1.007293	2.264083e-02	0.76087239	0.37797824

(Divergent protein kinase domain 1C)							
<i>FAM156B</i> (<i>TMEM29B</i>) (Family with sequence similarity 156 member B)	/	/	/	-1.189186	9.333000e-14	6.426085	2.817976
<i>P3R3URF-PIK3R3</i> Readthrough	/	/	/	-1.833480	1.452810e-02	0.7193576	0.2001381
gene: <i>ENSG00000258465</i>	/	/	/	-1.602100	1.054000e-05	1.09259278	0.35915470
gene: <i>ENSG00000255872</i>	/	/	/	-1.477783	3.862000e-03	0.7375282	0.2639103
gene: <i>ENSG00000284057</i>	/	/	/	-1.088332	1.605000e-07	3.8209871	1.7965551
<i>TBCEL-TECTA</i> Readthrough	/	/	/	-1.183985	4.005257e-02	1.0880918	0.4775082
gene: <i>ENSG00000272921</i>	/	/	/	-1.046384	3.162111e-02	2.515808	1.215900
gene: <i>ENSG00000262304</i>	/	/	/	-1.626821	2.406000e-05	0.66801872	0.21579793
<i>RPL36A-HNRNPH2</i> Readthrough	/	/	/	-3.273787	1.058000e-03	1.776631	0.178886

Movie S1 (separate file): The ORF2 encoded capsid protein of HEV is around the nucleus and in neurites Confocal microscope images of HEV-infected neurons were acquired. A 3D reconstruction of the surfaces was performed using the analysis tool Imaris 9.8.0. Cells were stained with DAPI (blue), with an antibody against ORF2-encoded capsid protein (green) and with β -III-tubulin (red).

SI References

1. P. J. Kenny, *et al.*, The FMRP–MOV10 complex: a translational regulatory switch modulated by G-Quadruplexes. *Nucleic Acids Res.* **48**, 862–878 (2020).
2. G. Skariah, *et al.*, Mov10 suppresses retroelements and regulates neuronal development and function in the developing brain. *BMC Biol.* **15**, 54 (2017).
3. H. R. Katz, A. A. Arcese, O. Bloom, J. R. Morgan, Activating Transcription Factor 3 (ATF3) is a Highly Conserved Pro-regenerative Transcription Factor in the Vertebrate Nervous System. *Front. Cell Dev. Biol.* **10** (2022).
4. R. Seiffers, A. J. Allchorne, C. J. Woolf, The transcription factor ATF-3 promotes neurite outgrowth. *Mol. Cell. Neurosci.* **32**, 143–154 (2006).
5. D. Halder, *et al.*, TREX1 Deficiency Induces ER Stress-Mediated Neuronal Cell Death by Disrupting Ca²⁺ Homeostasis. *Mol. Neurobiol.* **59**, 1398–1418 (2022).
6. K. Abe, P. Chu, A. Ishihara, H. Saito, Transforming growth factor- β 1 promotes re-elongation of injured axons of cultured rat hippocampal neurons. *Brain Res.* **723**, 206–209 (1996).
7. J. Kaiser, *et al.*, TGF β 1 Induces Axonal Outgrowth via ALK5/PKA/SMURF1-Mediated Degradation of RhoA and Stabilization of PAR6. *eNeuro* **7**, ENEURO.0104-20.2020 (2020).
8. J. Knöferle, *et al.*, TGF- β 1 enhances neurite outgrowth via regulation of proteasome function and EFABP. *Neurobiol. Dis.* **38**, 395–404 (2010).
9. S. Li, X. Gu, S. Yi, The Regulatory Effects of Transforming Growth Factor- β on Nerve Regeneration. *Cell Transplant.* **26**, 381–394 (2017).
10. C.-Y. Jeon, *et al.*, p190RhoGAP and Rap-dependent RhoGAP (ARAP3) inactivate RhoA in response to nerve growth factor leading to neurite outgrowth from PC12 cells. *Exp. Mol. Med.* **42**, 335–344 (2010).

11. C.-Y. Jeon, *et al.*, Neurite outgrowth from PC12 cells by basic fibroblast growth factor (bFGF) is mediated by RhoA inactivation through p190RhoGAP and ARAP3. *J. Cell. Physiol.* **224**, 786–794 (2010).
12. S. Böcker-Meffert, *et al.*, Erythropoietin and VEGF promote neural outgrowth from retinal explants in postnatal rats. *Invest. Ophthalmol. Vis. Sci.* **43**, 2021–2026 (2002).
13. K. Jin, X. O. Mao, D. A. Greenberg, Vascular endothelial growth factor stimulates neurite outgrowth from cerebral cortical neurons via Rho kinase signaling. *J. Neurobiol.* **66**, 236–242 (2006).
14. A. A. Khaibullina, J. M. Rosenstein, J. M. Krum, Vascular endothelial growth factor promotes neurite maturation in primary CNS neuronal cultures. *Brain Res. Dev. Brain Res.* **148**, 59–68 (2004).
15. J. M. Rosenstein, N. Mani, A. Khaibullina, J. M. Krum, Neurotrophic effects of vascular endothelial growth factor on organotypic cortical explants and primary cortical neurons. *J. Neurosci. Off. J. Soc. Neurosci.* **23**, 11036–11044 (2003).
16. M. Sondell, G. Lundborg, M. Kanje, Vascular endothelial growth factor has neurotrophic activity and stimulates axonal outgrowth, enhancing cell survival and Schwann cell proliferation in the peripheral nervous system. *J. Neurosci. Off. J. Soc. Neurosci.* **19**, 5731–5740 (1999).
17. S. H. Lee, *et al.*, tPA Regulates Neurite Outgrowth by Phosphorylation of LRP5/6 in Neural Progenitor Cells. *Mol. Neurobiol.* **49**, 199–215 (2014).
18. H. Xin, *et al.*, Increasing tPA activity in astrocytes induced by multipotent mesenchymal stromal cells facilitate neurite outgrowth after stroke in the mouse. *PloS One* **5**, e9027 (2010).
19. M. Breuer, H. Berger, A. Borchers, Caveolin 1 is required for axonal outgrowth of motor neurons and affects *Xenopus* neuromuscular development. *Sci. Rep.* **10**, 16446 (2020).
20. S. Wang, *et al.*, Caveolin-1 Phosphorylation Is Essential for Axonal Growth of Human Neurons Derived From iPSCs. *Front. Cell. Neurosci.* **13** (2019).
21. M. L. Mancini, *et al.*, Endoglin is required for myogenic differentiation potential of neural crest stem cells. *Dev. Biol.* **308**, 520–533 (2007).
22. M. A. Dent, Y. Sumi, R. J. Morris, P. J. Seeley, Urokinase-type plasminogen activator expression by neurons and oligodendrocytes during process outgrowth in developing rat brain. *Eur. J. Neurosci.* **5**, 633–647 (1993).

23. E. Semina, *et al.*, Urokinase and urokinase receptor participate in regulation of neuronal migration, axon growth and branching. *Eur. J. Cell Biol.* **95**, 295–310 (2016).
24. Q. Chen, L. Zhang, F. Zhang, S. Yi, FOSL1 modulates Schwann cell responses in the wound microenvironment and regulates peripheral nerve regeneration. *J. Biol. Chem.* **299** (2023).
25. J. Yun, *et al.*, Neuronal Per Arnt Sim (PAS) domain protein 4 (NPAS4) regulates neurite outgrowth and phosphorylation of synapsin I. *J. Biol. Chem.* **288**, 2655–2664 (2013).
26. M. G. Thomas, *et al.*, Nicotinamide N-methyltransferase expression in SH-SY5Y neuroblastoma and N27 mesencephalic neurones induces changes in cell morphology via ephrin-B2 and Akt signalling. *Cell Death Dis.* **4**, e669 (2013).
27. M. Barber, *et al.*, The Role of Robo3 in the Development of Cortical Interneurons. *Cereb. Cortex N. Y. NY* **19**, i22–i31 (2009).
28. C. L. Dou, J. M. Levine, Inhibition of neurite growth by the NG2 chondroitin sulfate proteoglycan. *J. Neurosci. Off. J. Soc. Neurosci.* **14**, 7616–7628 (1994).
29. D. L. Hynds, D. M. Snow, Neurite Outgrowth Inhibition by Chondroitin Sulfate Proteoglycan: Stalling/Stopping Exceeds Turning in Human Neuroblastoma Growth Cones. *Exp. Neurol.* **160**, 244–255 (1999).
30. J. Jin, *et al.*, Effect of chondroitin sulfate proteoglycans on neuronal cell adhesion, spreading and neurite growth in culture. *Neural Regen. Res.* **13**, 289–297 (2018).
31. G. Loers, *et al.*, Identification and characterization of synthetic chondroitin-4-sulfate binding peptides in neuronal functions. *Sci. Rep.* **9**, 1064 (2019).
32. Y. M. Ughrin, Z. J. Chen, J. M. Levine, Multiple Regions of the NG2 Proteoglycan Inhibit Neurite Growth and Induce Growth Cone Collapse. *J. Neurosci.* **23**, 175–186 (2003).
33. D. Maignel, M. Morita, Z. Pei, Z. Jia, P. A. Watkins, ACSF2: A MEDIUM-CHAIN ACYL-CoA SYNTHETASE WITH A POTENTIAL ROLE IN NEURONAL DIFFERENTIATION. [Preprint] (2022). Available at: <http://biorxiv.org/lookup/doi/10.1101/2022.03.28.486105> [Accessed 29 April 2024].
34. J. R. Marszalek, C. Kitidis, A. Dararutana, H. F. Lodish, Acyl-CoA synthetase 2 overexpression enhances fatty acid internalization and neurite outgrowth. *J. Biol. Chem.* **279**, 23882–23891 (2004).
35. Y. Kase, T. Sato, Y. Okano, H. Okano, The GADD45G/p38 MAPK/CDC25B signaling pathway enhances neurite outgrowth by promoting microtubule polymerization. *iScience* **25**, 104089 (2022).

36. D. K. Ma, *et al.*, Neuronal Activity–Induced Gadd45b Promotes Epigenetic DNA Demethylation and Adult Neurogenesis. *Science* **323**, 1074–1077 (2009).
37. E. Matsunaga, S. Nambu, M. Oka, A. Iriki, Comparative analysis of developmentally regulated expressions of *Gadd45a*, *Gadd45b*, and *Gadd45g* in the mouse and marmoset cerebral cortex. *Neuroscience* **284**, 566–580 (2015).
38. Y. Iwata, *et al.*, The effect of GDF15 on the neurite outgrowth in retinal ganglion cells. *Invest. Ophthalmol. Vis. Sci.* **60**, 624 (2019).
39. P. S. Klimovich, *et al.*, Urokinase receptor regulates nerve regeneration through its interaction with $\alpha 5\beta 1$ -integrin. *Biomed. Pharmacother.* **125**, 110008 (2020).
40. V. Laketa, J. C. Simpson, S. Bechtel, S. Wiemann, R. Pepperkok, High-Content Microscopy Identifies New Neurite Outgrowth Regulators. *Mol. Biol. Cell* **18**, 242–252 (2007).
41. L. N. Gillespie, G. M. Clark, P. F. Bartlett, P. L. Marzella, LIF is more potent than BDNF in promoting neurite outgrowth of mammalian auditory neurons in vitro. *Neuroreport* **12**, 275–279 (2001).
42. Y. Y. Wu, R. A. Bradshaw, Induction of Neurite Outgrowth by Interleukin-6 Is Accompanied by Activation of Stat3 Signaling Pathway in a Variant PC12 Cell (E2) Line*. *J. Biol. Chem.* **271**, 13023–13032 (1996).
43. Z. Hong, *et al.*, Phosphoproteome Study Reveals Hsp27 as a Novel Signaling Molecule Involved in GDNF-Induced Neurite Outgrowth. *J. Proteome Res.* **8**, 2768–2787 (2009).
44. T. Ishima, M. Iyo, K. Hashimoto, Neurite outgrowth mediated by the heat shock protein Hsp90 α : a novel target for the antipsychotic drug aripiprazole. *Transl. Psychiatry* **2**, e170–e170 (2012).
45. H. Takeuchi, *et al.*, Hsp70 and Hsp40 improve neurite outgrowth and suppress intracytoplasmic aggregate formation in cultured neuronal cells expressing mutant SOD1. *Brain Res.* **949**, 11–22 (2002).
46. B. C. Bornhauser, P.-A. Olsson, D. Lindholm, MSAP is a novel MIR-interacting protein that enhances neurite outgrowth and increases myosin regulatory light chain. *J. Biol. Chem.* **278**, 35412–35420 (2003).
47. H. Neumann, *et al.*, Tumor Necrosis Factor Inhibits Neurite Outgrowth and Branching of Hippocampal Neurons by a Rho-Dependent Mechanism. *J. Neurosci.* **22**, 854–862 (2002).
48. M.-S. Huang, *et al.*, Roles of carbonic anhydrase 8 in neuronal cells and zebrafish. *Biochim. Biophys. Acta BBA - Gen. Subj.* **1840**, 2829–2842 (2014).

49. C. Liu, W. Ma, W. Su, J. Zhang, Prdm14 acts upstream of islet2 transcription to regulate axon growth of primary motoneurons in zebrafish. *Dev. Camb. Engl.* **139**, 4591–4600 (2012).
50. M.-S. Yoon, *et al.*, Role of phospholipase D1 in neurite outgrowth of neural stem cells. *Biochem. Biophys. Res. Commun.* **329**, 804–811 (2005).
51. Y. Zhang, Y. Kanaho, M. A. Frohman, S. E. Tsirka, Phospholipase D1-promoted release of tissue plasminogen activator facilitates neurite outgrowth. *J. Neurosci. Off. J. Soc. Neurosci.* **25**, 1797–1805 (2005).
52. M. G. Hamel, *et al.*, Multimodal Signaling by the ADAMTSs (a disintegrin and metalloproteinase with thrombospondin motifs) Promote Neurite Extension. *Exp. Neurol.* **210**, 428–440 (2008).
53. A.-M. Lepagnol-Bestel, *et al.*, SLC25A12 expression is associated with neurite outgrowth and is upregulated in the prefrontal cortex of autistic subjects. *Mol. Psychiatry* **13**, 385–397 (2008).
54. M. T. van Diepen, *et al.*, The molluscan RING-finger protein L-TRIM is essential for neuronal outgrowth. *Mol. Cell. Neurosci.* **29**, 74–81 (2005).
55. U. Wilhelmsson, *et al.*, Absence of Glial Fibrillary Acidic Protein and Vimentin Prevents Hypertrophy of Astrocytic Processes and Improves Post-Traumatic Regeneration. *J. Neurosci.* **24**, 5016–5021 (2004).
56. Y. Endo, *et al.*, Wnt-3a and Dickkopf-1 Stimulate Neurite Outgrowth in Ewing Tumor Cells via a Frizzled3- and c-Jun N-Terminal Kinase-Dependent Mechanism. *Mol. Cell. Biol.* **28**, 2368–2379 (2008).
57. G. Edwards, *et al.*, SNAIL Transcription factor in prostate cancer cells promotes neurite outgrowth. *Biochimie* **180**, 1–9 (2021).
58. B. P. Niederöst, D. R. Zimmermann, M. E. Schwab, C. E. Bandtlow, Bovine CNS Myelin Contains Neurite Growth-Inhibitory Activity Associated with Chondroitin Sulfate Proteoglycans. *J. Neurosci.* **19**, 8979–8989 (1999).
59. H. Yamada, *et al.*, The brain chondroitin sulfate proteoglycan brevican associates with astrocytes ensheathing cerebellar glomeruli and inhibits neurite outgrowth from granule neurons. *J. Neurosci. Off. J. Soc. Neurosci.* **17**, 7784–7795 (1997).
60. S. Harashima, Y. Wang, T. Horiuchi, Y. Seino, N. Inagaki, Purkinje cell protein 4 positively regulates neurite outgrowth and neurotransmitter release. *J. Neurosci. Res.* **89**, 1519–1530 (2011).
61. C. Liu, *et al.*, Regulator of G protein signaling 5 (RGS5) inhibits sonic hedgehog function in mouse cortical neurons. *Mol. Cell. Neurosci.* **83**, 65–73 (2017).

62. S. Yin, *et al.*, RGS5 augments astrocyte activation and facilitates neuroinflammation via TNF signaling. *J. Neuroinflammation* **20**, 203 (2023).
63. D. M. Martin, *et al.*, *PITX2* is required for normal development of neurons in the mouse subthalamic nucleus and midbrain. *Dev. Biol.* **267**, 93–108 (2004).
64. O. B. Ilyinsky, *et al.*, Effects of opioid peptides and naloxone on nervous tissue in culture. *Neuroscience* **22**, 719–735 (1987).
65. I. Ahmed, Y. Calle, S. Iwashita, A. Nur-E-Kamal, Role of Cdc42 in neurite outgrowth of PC12 cells and cerebellar granule neurons. *Mol. Cell. Biochem.* **281**, 17–25 (2006).
66. M. D. Brown, B. J. Cornejo, T. B. Kuhn, J. R. Bamberg, Cdc42 stimulates neurite outgrowth and formation of growth cone filopodia and lamellipodia. *J. Neurobiol.* **43**, 352–364 (2000).
67. L. Mohiuddin, K. Fernandez, D. R. Tomlinson, P. Fernyhough, Nerve growth factor and neurotrophin-3 enhance neurite outgrowth and up-regulate the levels of messenger RNA for growth-associated protein GAP-43 and T alpha 1 alpha-tubulin in cultured adult rat sensory neurones. *Neurosci. Lett.* **185**, 20–23 (1995).
68. G. Morfini, M. C. DiTella, F. Feiguin, N. Carri, A. Cáceres, Neurotrophin-3 enhances neurite outgrowth in cultured hippocampal pyramidal neurons. *J. Neurosci. Res.* **39**, 219–232 (1994).
69. D. M. White, Contribution of neurotrophin-3 to the neuropeptide Y-induced increase in neurite outgrowth of rat dorsal root ganglion cells. *Neuroscience* **86**, 257–263 (1998).
70. C. B. Alves, *et al.*, Caffeine and adenosine A2A receptors rescue neuronal development *in vitro* of frontal cortical neurons in a rat model of attention deficit and hyperactivity disorder. *Neuropharmacology* **166**, 107782 (2020).
71. H. Maruyama, K. Fujiwara, M. Kumeta, D. Koyama, Ultrasonic control of neurite outgrowth direction. *Sci. Rep.* **11**, 20099 (2021).
72. M. Hashimoto, E. Rockenstein, M. Mante, M. Mallory, E. Masliah, β -Synuclein Inhibits α -Synuclein Aggregation: A Possible Role as an Anti-Parkinsonian Factor. *Neuron* **32**, 213–223 (2001).
73. H. J. Lee, K. Lee, H. Im, α -Synuclein modulates neurite outgrowth by interacting with SPTBN1. *Biochem. Biophys. Res. Commun.* **424**, 497–502 (2012).
74. S. Mitsuzawa, *et al.*, Reduced PHOX2B stability causes axonal growth impairment in motor neurons with TARDBP mutations. *Stem Cell Rep.* **16**, 1527–1541 (2021).

75. K. M. Munro, A. Nash, M. Piloni, S. F. Lichtenthaler, J. M. Gunnensen, Functions of the Alzheimer's Disease Protease BACE1 at the Synapse in the Central Nervous System. *J. Mol. Neurosci.* **60**, 305–315 (2016).
76. J. Zhang, *et al.*, Sez-6 may play an important role in neurite outgrowth through the PKCgamma signaling pathways. *Z. Naturforschung C J. Biosci.* **66**, 614–620 (2011).