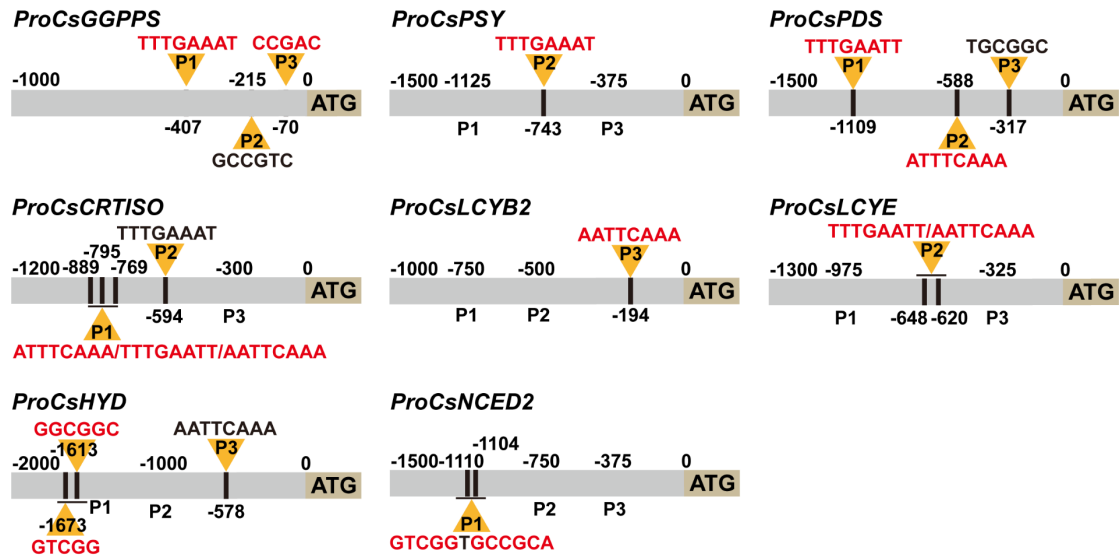


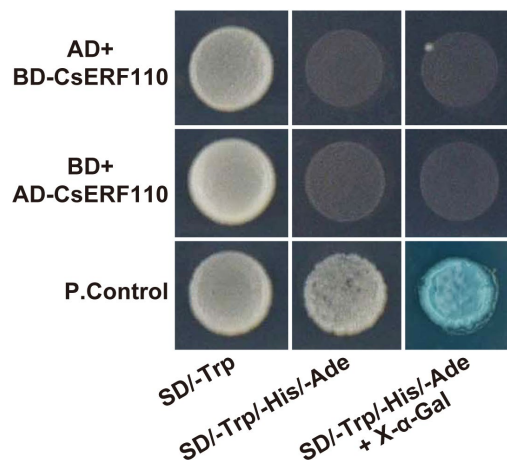
Supplemental Figure 2. *CsERF110* is involved in ABA-induced citrus carotenoid accumulation as a nucleus-localized transcriptional activator.

(A) Subcellular localization of *CsERF110* in *Nicotiana benthamiana* (*N. benthamiana*). *CsERF110*-GFP, GFP signal; NF-YA4-mCherry, RFP signal; Merged, combined GFP and RFP signals. Bright Field and white light were background colors. Bars = 75 μ m. **(B)** *CsERF110* transactivation activity assay. PGBKT7-53+PGADT7-RecT and empty vector PGBKT7 were used as positive control (P. Control) and negative control (N. Control), respectively. **(C)** and **(D)** Transcriptional activity analysis of *CsERF110* using a dual luciferase system. Empty pBD and pBD-VP16 served as the negative and positive controls, respectively. The luciferase activity was quantified as the transcriptional activity of *CsERF110*. The data were expressed mean \pm SD of at least three biological replicates. Statistically significant differences were determined by Student's t-test (*, $P < 0.05$; **, $P < 0.01$).



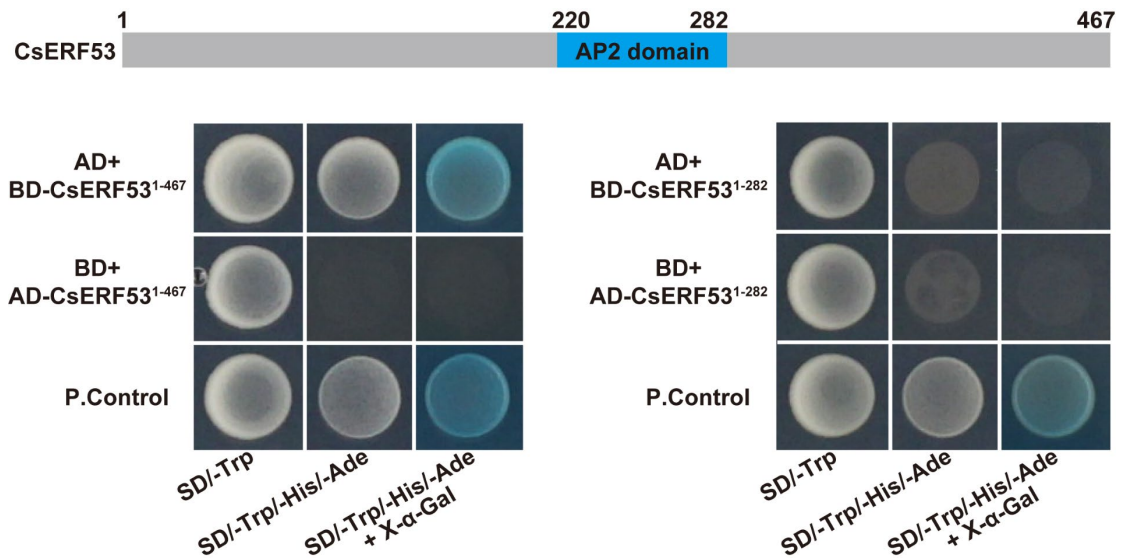
Supplemental Figure 3. Schematic diagram of target gene promoters.

Schematic diagram of target gene promoters (*CsGGPPS*, *CsPSY*, *CsPDS*, *CsCRTISO*, *CsLCYB2*, *CsLCYE*, *CsHYD*, and *CsNCED2*). These black lines represent putative *CsERF110* binding motif of these promoters.



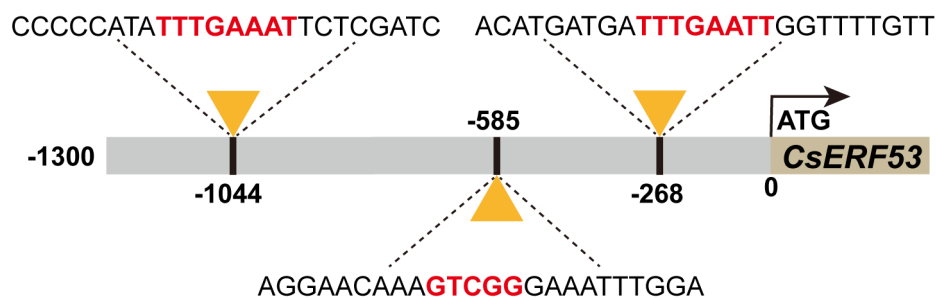
Supplemental Figure 4. *CsERF110* has no transcriptional self-activation in yeast cells.

Co-transformation of pGBKT7-p53 and pGADT7-RecT was used as a positive control (P. Control). Yeast grown in SD/-Trp/-Leu medium and SD/-Trp/-Leu/-His/-Ade medium is shown. The interaction is indicated by yeast growth and X- α -Gal staining.



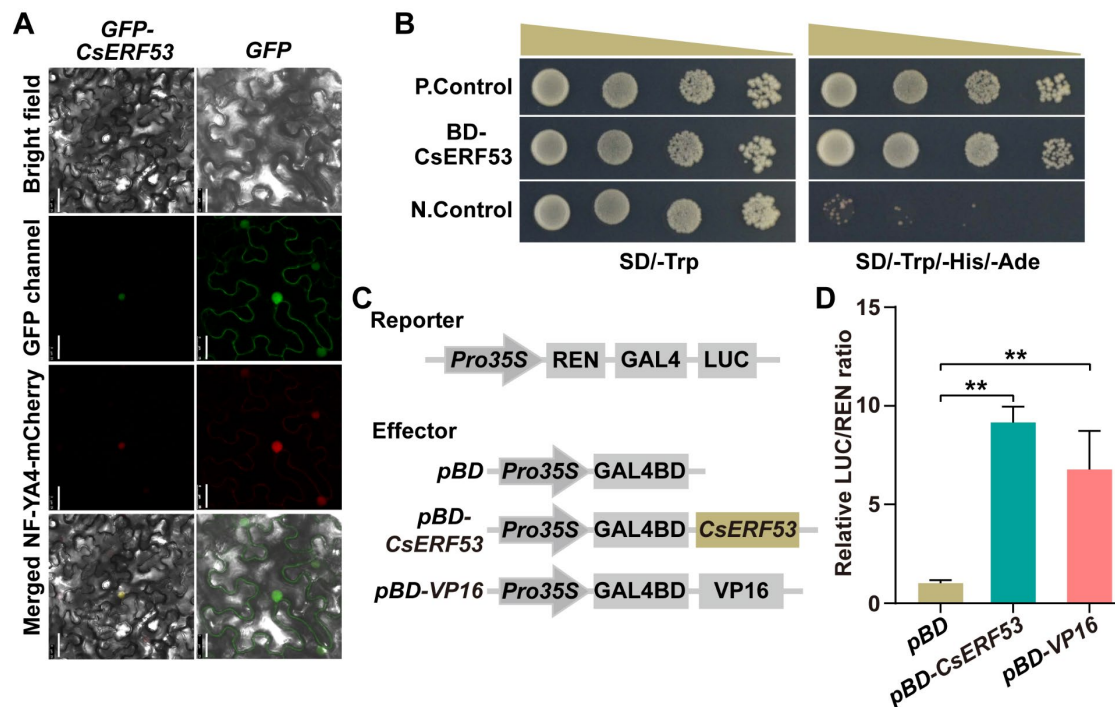
Supplemental Figure 6. Analysis of CsERF53 transcriptional self-activation in yeast cells.

Co-transformation of pGBKT7-p53 and pGADT7-RecT was used as a positive control (P. Control). Yeast grown in SD/-Trp/-Leu medium and SD/-Trp/-Leu/-His/-Ade medium is shown. The interaction is indicated by yeast growth and X- α -Gal staining.



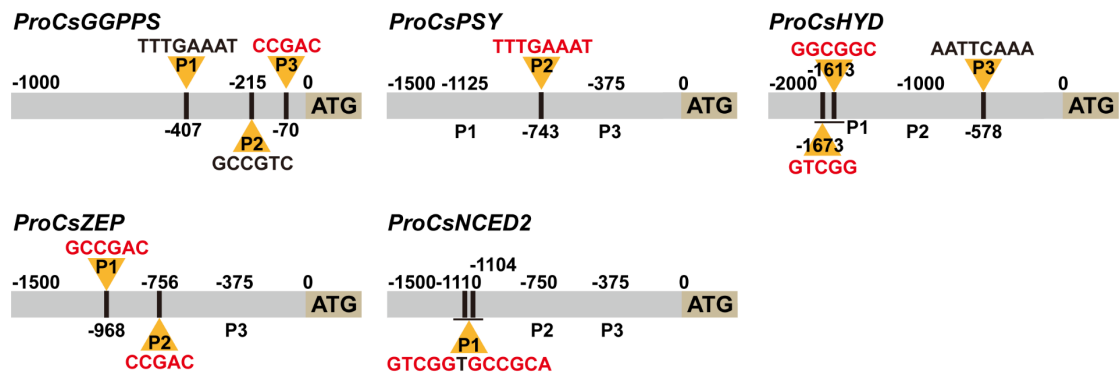
Supplemental Figure 7. Schematic representation of potential binding elements of CsERF110 in the CsERF53 promoter.

Yellow triangles represent the position of potential *cis*-acting elements of CsERF110 in the CsERF53 promoter, and red letters represent the sequences of the potential *cis*-elements of CsERF110.



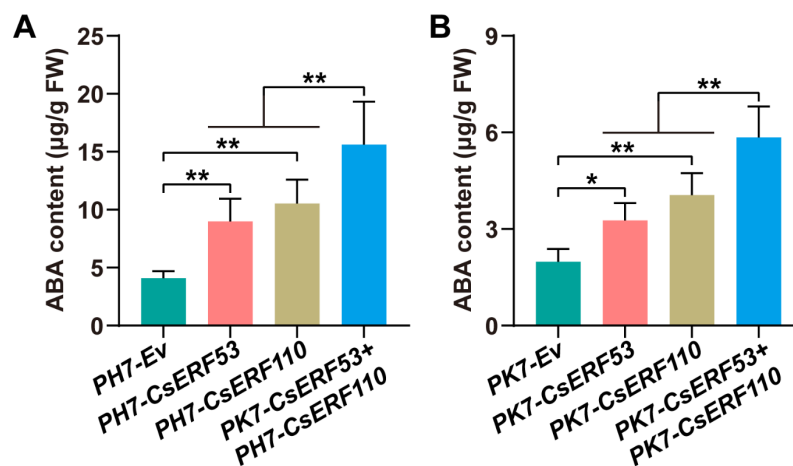
Supplemental Figure 8. *CsERF53* is involved in ABA-promoted citrus carotenoid accumulation as a nucleus-localized transcriptional activator.

(A) Subcellular localization of *CsERF53* in *N. benthamiana* leaves. *CsERF53*-GFP, GFP signal; NF-YA4-mCherry, RFP signal; Merged, combined GFP and RFP signals. Bright Field and white light were background colors. Images on the right, bars = 75 μm ; Images on the left, bars = 50 μm . **(B)** *CsERF53* transactivation activity assay. PGBKT7-53+PGADT7-RecT and empty vector PGBKT7 were used as positive control (P. Control) and negative control (N. Control), respectively. **(C)** and **(D)** Transcriptional activity analysis of *CsERF53* using a dual luciferase system. Empty pBD and pBD-VP16 served as the negative and positive controls, respectively. The luciferase activity was quantified as the transcriptional activity of *CsERF53*. The data were expressed mean \pm SD of at least three biological replicates. Statistically significant differences were determined by Student's t-test (*, $P < 0.05$; **, $P < 0.01$).



Supplemental Figure 9. Schematic diagram of target gene promoters.

Schematic diagram of target gene promoters (*CsGGPPS*, *CsPSY*, *CsHYD*, and *CsNCED2*). These black lines represent putative *CsERF53* binding motif of these promoters.



Supplemental Figure 10. Overexpression of *CsERF53* and *CsERF110* significantly promotes ABA accumulation.

The content of ABA in stably transformed citrus calli (**A**) and transiently injected citrus fruit (**B**). Data represent means \pm SD of three biological replicates. Asterisks indicate statistically significant differences determined by Student's *t*-test (*, $0.01 < P < 0.05$; **, $P < 0.01$; n.s., no significant difference).