Science Advances

Supplementary Materials for

A general temperature-guided language model to design proteins of enhanced stability and activity

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Sci. Adv. **10**, eadr2641 (2024) DOI: 10.1126/sciadv.adr2641

The PDF file includes:

Figs. S1 and S2 Tables S1 to S9 Legend for data S1 References

Other Supplementary Material for this manuscript includes the following:

Data S1

Supplementary Materials



Smoothed Training Loss During Pretraining

Fig. S1. Training and validation curves during the pre-training process. (A) The smoothed training loss during pre-training. From left to right are MLM Loss, OGT Loss, and Correlation Loss. The stopping criterion for pre-training was reaching the maximum allocated compute budget and we fixed the training steps at 200,000. (B) Validation performance of MLM and OGT predictions. During the pre-training process, we recorded the validation loss on the validation set every 5,000 steps.



Fig. S2. **Comparison of predicted and actual OGT.** Scatter plot of predicted OGT versus actual OGT on the held-out validation set.

Table S1. Ablation study results for the PRIME model. The numbers in parentheses represent the p-values comparing the baseline PRIME data with the data from the ablation experiments. We used Spearman correlation as our evaluation metric, where a higher value indicates better performance.

PRIME (MSE-Loss) refers to the use of MSE loss to align the MLM and OGT modules. We found that this MSE approach yielded inferior results compared to using Pearson correlation as the loss function.

	ProteinGym	Stability	Activit	ΔT_{m}
			У	
PRIME (-OGT)	0.447 (p=4e-2)	0.564	0.452	0.362 (p=3e-2)
PRIME (-Corr)	0.472 (p=4e-2)	0.588	0.479	0.429 (p=4e-2)
PRIME (-MLM)	0.024 (p=2e-4)	0.130	-0.089	0.150 (p=1e-4)
PRIME (MSE-Loss)	0.432 (p=1e-3)	0.498	0.426	0.363 (p=3e-2)
PRIME	0.486	0.609	0.502	0.437

Table S2. Results of benchmarking PRIME against various baseline models on the ProteinGym, Stability, and ΔT_m datasets. The "Stability" dataset refers to ProteinGym-Stability, and "Activity" refers to ProteinGym-Activity. We used Spearman correlation as our evaluation metric, where a higher value indicates better performance. The numbers in parentheses represent the p-values from the Wilcoxon Signed Rank Test, comparing the predictions of the original PRIME model to those of the subsequent models. "PRIME (homo)" denotes the fine-tuning of the PRIME model using the MLM loss on homologous sequences of the target proteins present in either ProteinGym or ΔT_m datasets.

	ProteinGym	Stability	Activity	ΔT_{m}
ESM-2	0.414 (p=2e-3)	0.523	0.425	0.330 (p=1e-3)
ESM-2 (Homo)	0.438 (p=3e-3)	0.574	0.462	0.385 (p=2e-3)
ESM-1v	0.407 (p=3e-4)	0.477	0.42	0.279 (p=2e-3)
MSA-Transformer	0.434 (p=1e-2)	0.495	0.457	0.327 (p=3e-2)
Tranception-EVE	0.456 (p=2e-3)	0.500	0.487	0.247 (p=8e-3)
SaProt	0.457 (p=1e-4)	0.592	0.458	0.311 (p=1.1e-2)
CARP	0.368 (p=2e-2)	0.412	0.395	0.272 (p=2e-3)
MIFST	0.400 (p=1e-4)	0.485	0.327	0.377 (p=3e-2)
Stability Oracle	0.386 (p=7e-4)	0.568	0.368	0.412 (p=9e-3)
Rosetta	0.154 (p=9.6e- 13)	0.154	0.154	0.198 (p=4e-13)
GEMME	0.455 (p=4.1e-2)	0.519	0.484	0.354 (p=2e-2)
PRIME	0.486	0.609	0.502	0.437
PRIME (Homo)	0.496 (p=0.039)	0.612	0.521	0.456 (p=8e-3)

Table S3. Supervised prediction of T_m (melting temperature) and T_{opt} (optimal enzymatic activity temperature). Four metrics are employed to gauge the models' accuracy and predictive ability: RMSE, Spearman correlation, Pearson correlation, and R^2 (coefficient of determination). The number in parentheses represents the p-values from the Wilcoxon Signed Rank Test, comparing the predictions of the original PRIME model to those of the subsequent models. We trained PRIME and ESM2 with three different random seeds, while the results for DeepET were obtained from Ref(82).

		RMSE	Pearson	R ²	Spearman
	ESM-2-	13.845(±0.1524)	0.693(±0.012)	0.552(±0.0198)	0.585(±0.0221)
т	650M(p=1e-12)				
I opt	DeepET(p=1e-9)	12.21	0.76	0.57	0.62
	PRIME	11.23 (±0.2200)	0.81 (±0.0092)	0.63 (±0.0142)	0.76 (±0.0139)
		RMSE	Pearson	R ²	Spearman
	ESM-2-	5.589(±0.1642)	0.774(±0.008)	0.592(±0.015)	0.548(±0.02)
т	650M(p=1e-8)				
1 m	DeepET(p=1e-7)	6.30	0.76	0.58	0.55
	PRIME	4.80 (±0.1739)	0.84 (±0.0064)	0.70 (±0.0115)	0.64 (±0.0134)

Table S4. Results from the FLIP benchmark: The results for CARP, ESM-1b and ESM-1v are sourced from the literature (*32*). Spearman correlation is used as the evaluation metric, where a higher value indicates better performance. We implemented a global fine-tuning approach to PRIME and ESM2, where all parameters participated in training.

AAV	mut-des	1-vs-many	2-vs-many	7-vs-many	low-vs-high
CARP-640M	0.750	0.310	0.510	0.580	0.250
ESM-1b	0.700	0.310	0.650	0.610	0.330
ESM-1v	0.790	0.100	0.700	0.700	0.340
ESM-2	0.385	0.440	0.569	0.594	0.310
PRIME	0.82(±0.012)	0.523(±0.01)	0.788(±0.0130	0.762(±0.021)	0.421(±0.013)
GB1	1-vs-many	2-vs-many	3-vs-many	low-vs-high	
CARP-640M	0.150	0.180	0.620	0.120	
ESM-1b	0.280	0.550	0.790	0.590	
ESM-1v	0.320	0.320	0.770	0.100	
ESM-2	0.180	0.470	0.790	0.530	
PRIME	0.32(±0.008)	0.598(±0.012)	0.824(±0.030)	0.613(±0.014)	
MELTOME	Mixed	Human-cell	Human		
CARP-640M	0.530	0.686	0.724		
ESM-1b	0.680	0.750	0.700		
ESM-1v	0.650	0.770	0.780		
ESM-2	0.490	0.627	0.654		
PRIME	0.724(±0.002)	0.825(±0.009)	0.804(±0.023)		

Table S5. The comparative results of four distinct strategies for selecting the top 15 single-point mutations within the ProteinGym dataset, specifically focusing on proteins with saturated single-point mutation data and wild-type sequence similarity below 30% to the PRIME pre-training dataset. The strategies compared are as follows:

- 1) The strategy method described in this paper, which involves fine-tuning the PRIME model using homologous sequences of the target protein.
- 2) Fine-tuning ESM2 on the same homologous sequences.
- 3) The ESM vote strategy from "Hie et al, Nature Biotechnology, 2023,"(74), which integrates five ESM-1v models with the ESM1b model, setting alpha to 1 and adjusting the value of k for each dataset to yield only 15 mutations.
- 4) Random mutations, which serve as a baseline for comparison.

The evaluation metrics include the number of positive single-point mutations, the fitness of the most advantageous mutant, and the median fitness of all 15 mutants. The checkpoints files of PRIME (homo) and ESM2 (homo) are available on our GitHub.

Datasets	Metric	PRIME	ESM2	ESM-	Random
		(homo)	(homo)	vote	
HSP82_YE	Fraction with	7	4	5	2
AST_Flynn_	improved fitness				
2019 (0)	Maximum improved	0.36	0.27	0.13	0.11
	fitness				
	Median fitness	0.02	-0.12	-0.13	-0.07
KKA2_KLE	Fraction with	14	13	12	2
PN_Melniko	improved fitness				
v_2014 (1.0)	Maximum improved	2.48	2.36	2.75	1.72
	fitness				
	Median fitness	1.62	0.64	1.09	0.75
NKX31_HU	Fraction with	15	13	5	2
MAN_Tsub	improved fitness				
oyama_2023	Maximum improved	2.14	0.76	0.42	0.12
_2L9R (0)	fitness				
	Median fitness	0.36	0.24	-0.16	0.22
PTEN_HU	Fraction with	2	0	0	0
MAN_Migh	improved fitness				
ell_2018 (0)	Maximum improved	0.48	0.00	0.00	0.00
	fitness				
	Median fitness	-0.84	-1.06	-0.95	-0.97
TAT_HV1B	Fraction with	7	5	8	2
R_Fernande	improved fitness				
s_2016 (0)	Maximum improved	0.65	0.24	0.37	0.03
	fitness				
	Median fitness	0.02	-0.02	0.01	-0.07

Table S6. The comparative result of the top 15 single-point mutations for T7 RNA polymerase and Tgo-D4K, as selected by four different model methods. The evaluation criteria include the number of positive single-point mutations, the fitness of the most advantageous mutant, and the median fitness of all 15 mutants. For T7 RNA polymerase, fitness is measured by the change in T_m relative to the wild type. For DNA polymerase, fitness is measured by the polymerization activity for 2'-fluoroarabino nucleic acid (a non-natural nucleic acid), with values representing the fold change in activity relative to the wild type. The checkpoints files for PRIME (homo) and ESM2 (homo) strategies are available on our GitHub repository. Rosetta scores protein-saturated single-point mutations by ranking based on predicted values of the unfolding free energy. The energy function used to calculate this unfolding free energy includes all energy terms referenced in the literature (75).

Protein	Metric	PRIME	ESM2	ESM-	Rosetta
		(homo)	(homo)	vote	
	Number	13	8	7	1
	with				
T7 RNA	improved T _m				
polymerase	Maximum	3.5	0.8	3.5	0.6
	improved T _m				
	Median	1.8	0.2	0	-0.7
	improved T _m				
	Number	10	7	6	5
	with				
	improved				
Tgo-D4K	fitness				
	Maximum	3.27	2.15	2.07	2.06
	improved				
	fold				
	Median fold	1.21	0.99	0.93	0.93

		Zero-Shot	Round 1	Round 2
	Random	0.43 (±0.0389)	0.2397(±0.1612)	0.2286(±0.1577)
	ftMLDE	0.44 (±0.0062)	0.55 (0.0302)	0.55 (0.0820)
Max Fitness achieved	ESM-2 (tiered)	0.45	0.64(±0.02568)	0.66(±0.03645)
	PRIME (tiered)	0.45	0.66(±0.04157)	0.84(±0.0362)
	ESM-2 (top)	0.44	0.66 (±0.0406)	0.66 (±0.0406)
	PRIME (top)	0.45	0.69 (±0.0587)	0.86 (±0.0295)
	Random	0.06 (±0.0086)	0.01 (±0.0064)	0.01 (±0.0063)
	ftMLDE	0.08 (±0.0034)	0.13 (±0.0018)	0.14 (±0.0017)
Mean Fitness achieved	ESM-2 (tiered)	0.07	0.05(±0.0024)	0.1(±0.0011)
	PRIME (tiered)	0.08	0.18(±0.0019)	0.29(±0.0013)
	ESM-2 (top)	0.09	0.07 (±0.0088)	0.10 (±0.0009)
	PRIME (top)	0.11	0.22 (±0.0053)	0.32 (±0.0032)

Table S7. Results of the in-silico benchmarking of iterative strategies based on PRIME and ftMLDE.

Table S8. Pseudocode for the alternating training strategy used to optimize MLM, OGT, and Correlation losses.

Input: Mini Batch, training step <i>i</i> .
Output: Loss of the mini-batch.
sequence_ids, ogt = batch['sequence_ids'], batch['ogt']
if i % 3 == 1:
noised_sequence_ids = add_noise(sequence_ids)
reconstruc_sequence_ids = model(noised_sequence_ids, task = 'MLM')
loss = compute_mlm_loss(sequence_ids, noised_sequence_ids)
return loss
elif i % 3 == 2:
<pre>pred_ogt = model(sequence_ids, task = 'OGT')</pre>
loss = mse_loss(pre_ogt, ogt)
return loss
elif i % 3 == 0:
reconstruction_logits = model(sequence_ids)
mutant_indices = random_generate_k_points(sequence_ids)
mutant_types = random_generate_k_types(sequence_ids)
<pre>mutant_sequence_ids = sequence_ids.clone()</pre>
mutant_sequence_ids.scatter_(1, mutant_indices, mutant_types)
mlm_scores = reconstruction_logits[:, mutant_indices, mutant_types].log_softmax(dim=-1)
pred_ogt = prime(mutant_sequence_ids, task = 'OGT')
loss = 1 - pearson(pred_ogt, mlm_scores)
return loss

MLM Loss	Corr Loss	OGT Loss	ProteinGym	ΔT_m	ProteinGym-
Weight	Weight	Weight			Stability
1	1	1	0.440	0.360	0.557
1	0.5	2	0.433	0.351	0.555
0.01	0.5	0.05	0.432	0.349	0.552
1	2	0.05	0.431	0.349	0.551
2	2	2	0.431	0.348	0.550
0.5	2	0.5	0.430	0.348	0.549
1	0.01	0.5	0.430	0.347	0.549
1	1	0.5	0.429	0.346	0.549
0.5	0.5	2	0.429	0.345	0.549
0.5	2	2	0.428	0.345	0.548
0.5	0.5	0.5	0.428	0.345	0.547
1	1	0.01	0.428	0.344	0.547
0.01	1	0.01	0.427	0.344	0.547
0.01	0.5	0.01	0.427	0.344	0.547
0.01	2	0.05	0.427	0.344	0.546
0.05	0.05	2	0.426	0.343	0.545
0.05	1	2	0.426	0.342	0.544
0.5	0.5	1	0.425	0.342	0.544
1	0.05	0.01	0.424	0.342	0.544
0.05	0.01	2	0.424	0.342	0.544
2	2	0.01	0.424	0.341	0.544
0.05	0.05	0.05	0.424	0.341	0.543
0.01	0.01	1	0.424	0.340	0.543
2	2	1	0.423	0.340	0.543
2	0.01	0.05	0.423	0.340	0.542
0.5	0.05	0.5	0.423	0.340	0.542
1	2	1	0.422	0.339	0.542
0.5	1	0.5	0.422	0.339	0.541
0.5	1	1	0.422	0.339	0.541
0.5	0.05	2	0.421	0.339	0.541
0.01	0.05	1	0.421	0.339	0.541
0.05	0.05	0.5	0.420	0.339	0.540
0.05	1	0.05	0.420	0.338	0.540
1	0.01	2	0.420	0.338	0.540
0.01	0.05	0.5	0.419	0.338	0.540
0.5	2	0.01	0.418	0.338	0.540

Table S9. Performance of zero-shot mutation prediction at a 1:1:1 weight ratio on the ProteinGym and ΔT_m datasets.

0.5	0.05	0.05	0.418	0.338	0.540
1	1	0.05	0.418	0.337	0.540
2	0.05	2	0.418	0.337	0.540
0.5	0.5	0.05	0.418	0.337	0.539
0.5	0.01	2	0.418	0.337	0.539
0.5	0.01	0.05	0.418	0.337	0.539
0.5	0.01	0.01	0.418	0.337	0.538
0.05	1	1	0.417	0.337	0.537
0.5	1	0.05	0.417	0.337	0.537
1	0.05	2	0.417	0.336	0.537
0.05	0.5	2	0.417	0.336	0.537
0.05	1	0.01	0.417	0.336	0.537
0.05	0.01	1	0.417	0.336	0.536
0.01	0.5	1	0.416	0.336	0.536
0.05	0.5	0.05	0.416	0.336	0.536
1	0.01	0.01	0.416	0.336	0.536
0.01	2	0.5	0.416	0.335	0.535
0.01	0.5	0.5	0.416	0.334	0.535
0.5	2	1	0.416	0.334	0.535
2	0.5	0.05	0.415	0.334	0.535
0.01	1	1	0.415	0.333	0.535
0.01	0.01	0.01	0.415	0.333	0.534
0.5	0.05	0.01	0.415	0.333	0.534
1	0.5	0.01	0.415	0.332	0.534
0.01	2	2	0.415	0.332	0.534
2	1	2	0.414	0.332	0.534
2	2	0.5	0.414	0.332	0.534
2	0.01	1	0.414	0.332	0.533
2	0.5	0.01	0.414	0.331	0.533
0.5	0.05	1	0.413	0.331	0.533
2	2	0.05	0.413	0.331	0.533
2	0.5	2	0.413	0.331	0.532
2	0.01	0.01	0.412	0.331	0.532
2	0.05	0.05	0.412	0.331	0.532
2	0.5	1	0.412	0.331	0.531
0.05	0.01	0.01	0.412	0.331	0.531
2	1	0.01	0.412	0.330	0.531
1	2	2	0.412	0.330	0.530
0.05	0.5	0.5	0.412	0.330	0.530
0.01	0.01	0.05	0.412	0.330	0.530
1	0.05	1	0.412	0.329	0.530
1	2	0.5	0.411	0.329	0.530

2	1	1	0.411	0.329	0.530
0.01	0.01	2	0.411	0.329	0.529
0.05	2	0.01	0.411	0.329	0.529
0.05	0.05	1	0.411	0.328	0.529
0.05	0.05	0.01	0.411	0.328	0.529
0.05	0.5	1	0.410	0.328	0.529
1	0.01	1	0.410	0.328	0.528
2	0.5	0.5	0.410	0.328	0.528
0.5	0.01	1	0.410	0.327	0.528
0.01	2	0.01	0.410	0.327	0.528
0.05	0.01	0.05	0.410	0.327	0.527
0.5	0.01	0.5	0.410	0.327	0.527
0.05	0.5	0.01	0.410	0.326	0.527
1	0.5	0.5	0.409	0.326	0.527
2	0.05	0.01	0.409	0.326	0.527
0.01	0.05	0.05	0.409	0.325	0.527
1	2	0.01	0.409	0.325	0.526
0.5	2	0.05	0.409	0.324	0.526
1	0.05	0.05	0.409	0.324	0.526
1	0.05	0.5	0.409	0.324	0.526
0.01	1	2	0.408	0.323	0.525
2	0.05	0.5	0.408	0.323	0.525
0.01	0.5	2	0.408	0.323	0.525
0.05	2	1	0.408	0.323	0.525
1	0.5	0.05	0.408	0.323	0.525
0.05	2	0.05	0.407	0.322	0.524
0.05	2	2	0.407	0.322	0.523
0.5	0.5	0.01	0.407	0.322	0.523
0.01	0.01	0.5	0.407	0.322	0.523
0.01	2	1	0.406	0.321	0.523
1	0.5	1	0.406	0.321	0.522
0.05	0.01	0.5	0.406	0.321	0.522
2	1	0.05	0.406	0.320	0.521
0.01	0.05	2	0.406	0.320	0.521
2	1	0.5	0.405	0.320	0.521
0.5	1	2	0.405	0.320	0.521
0.01	1	0.05	0.405	0.319	0.520
2	0.01	0.5	0.405	0.319	0.520
0.01	0.05	0.01	0.402	0.319	0.520
2	0.05	1	0.402	0.318	0.519
0.05	1	0.5	0.401	0.317	0.518
2	0.01	2	0.400	0.316	0.516

0.01	1	0.5	0.399	0.316	0.516
0.05	2	0.5	0.399	0.315	0.514
1	0.01	0.05	0.398	0.312	0.514
0.5	1	0.01	0.397	0.311	0.511
1	1	2	0.390	0.300	0.510

Other Supplementary Material for this manuscript includes the

following:

Supplementary Data File: Sheets S1 to S5.

Sheet S1: LbCas12a saturation single-point mutation ranking, featuring single-point mutation T_m results, along with two rounds of multipoint experimental results for T_m and activity.

Sheet S2: T7 RNA polymerase saturation single-point mutation ranking, including single-point mutation T_m results, two rounds of multipoint experimental results for T_m and activity, and the top fifteen single-point mutation types and Tm results predicted by Rosetta, ESM-vote, and ESM2 (homo).

Sheet S3: Creatinase saturation single-point mutation ranking with single-point mutation T_m results.

Sheet S4: Tgo-D4K saturation single-point mutation ranking, including single-point mutation activity results and the top fifteen single-point mutation types and activity results predicted by Rosetta, ESM-vote, and ESM2 (homo).

Sheet S5: VHH saturation single-point mutation ranking, with single-point mutation EC50 results.

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