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Supplemental information

**An IL-2 mutein increases regulatory T cell
suppression of dendritic cells via IL-10
and CTLA-4 to promote T cell anergy**

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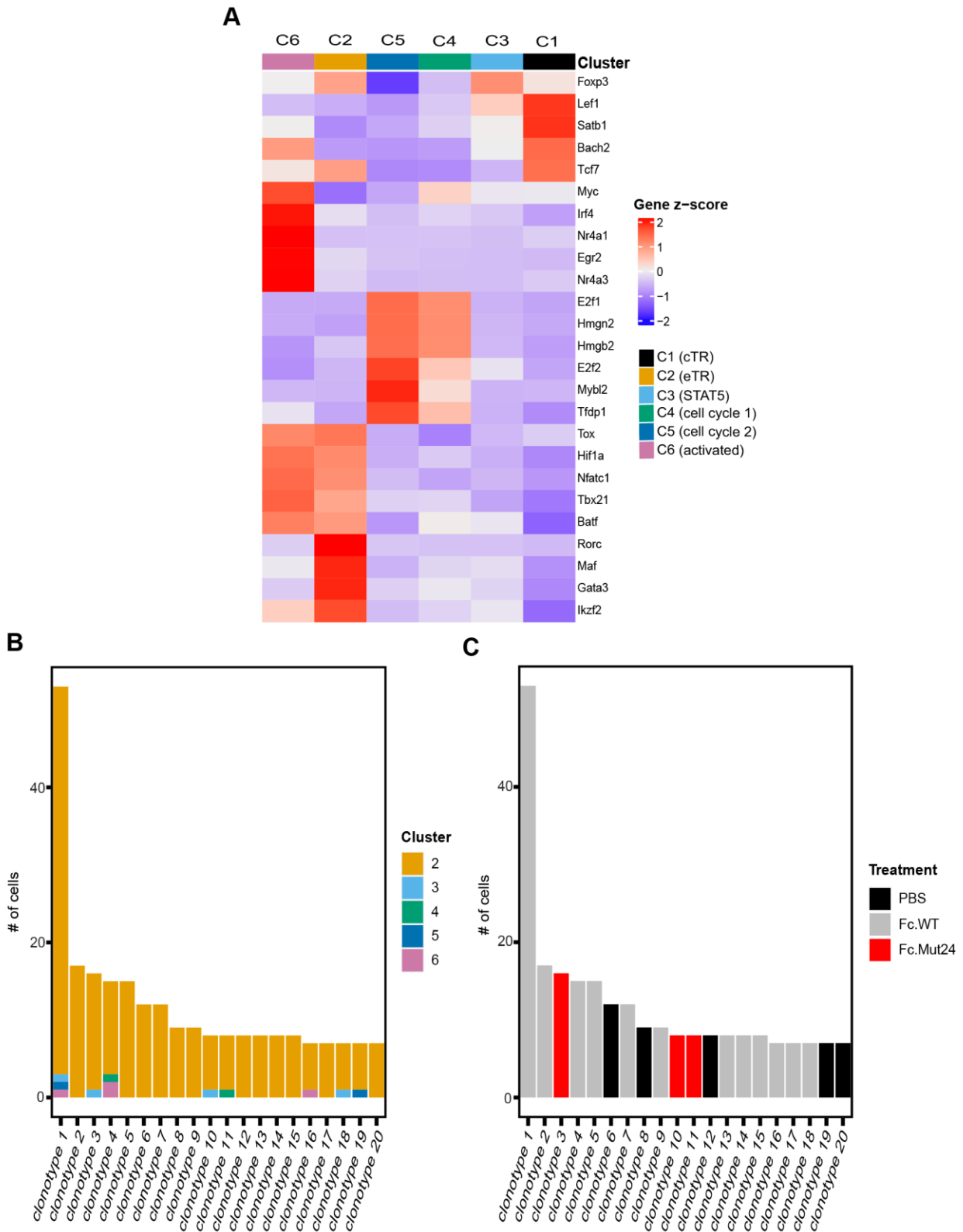


Figure S1. Fc.Mut24 does not impact Fc μ 3⁺ T_R cell clonal expansion, related to Figure 1. Splenocytes from C57BL/6 Fc μ 3-mRFP treated with PBS, Fc.WT, or Fc.Mut24 were harvested 3 days later and sorted to obtain CD4⁺ Fc μ 3⁺ regulatory T (T_R) cells for single-cell RNA-seq (scRNA-seq). **A.** Heat map showing the mean expression (z-score) of representative transcription factor genes (rows) for each cluster (columns). **B&C.** Bar graphs showing the number of cells identified for each of the top twenty T_R clonotypes colored by cluster (**B**) or treatment group (**C**).

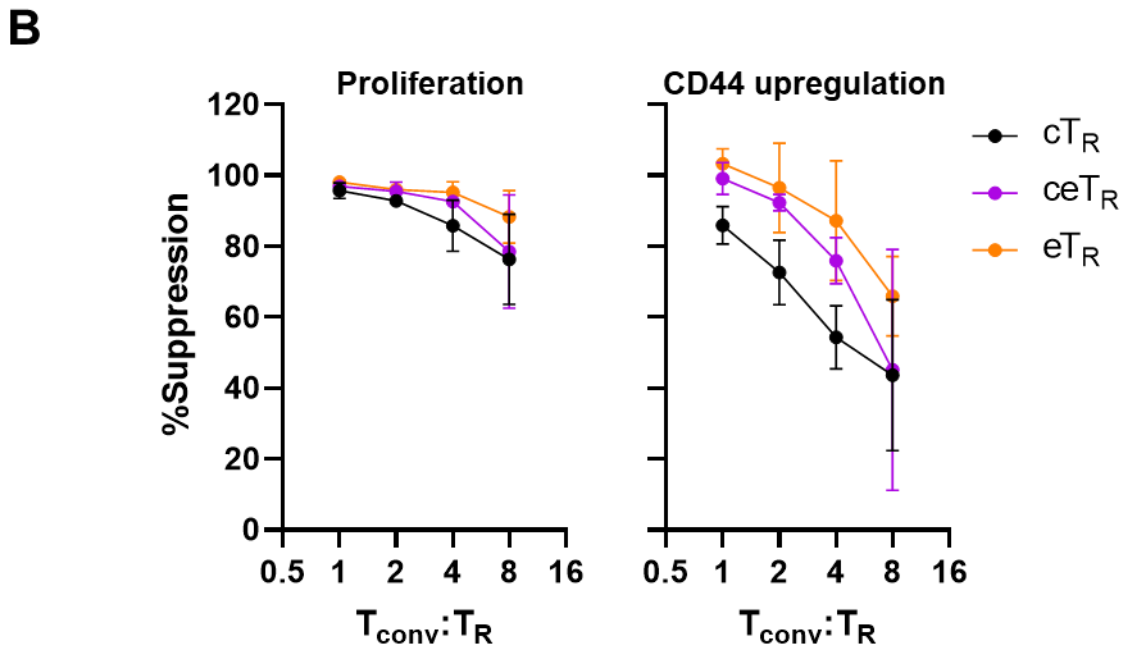
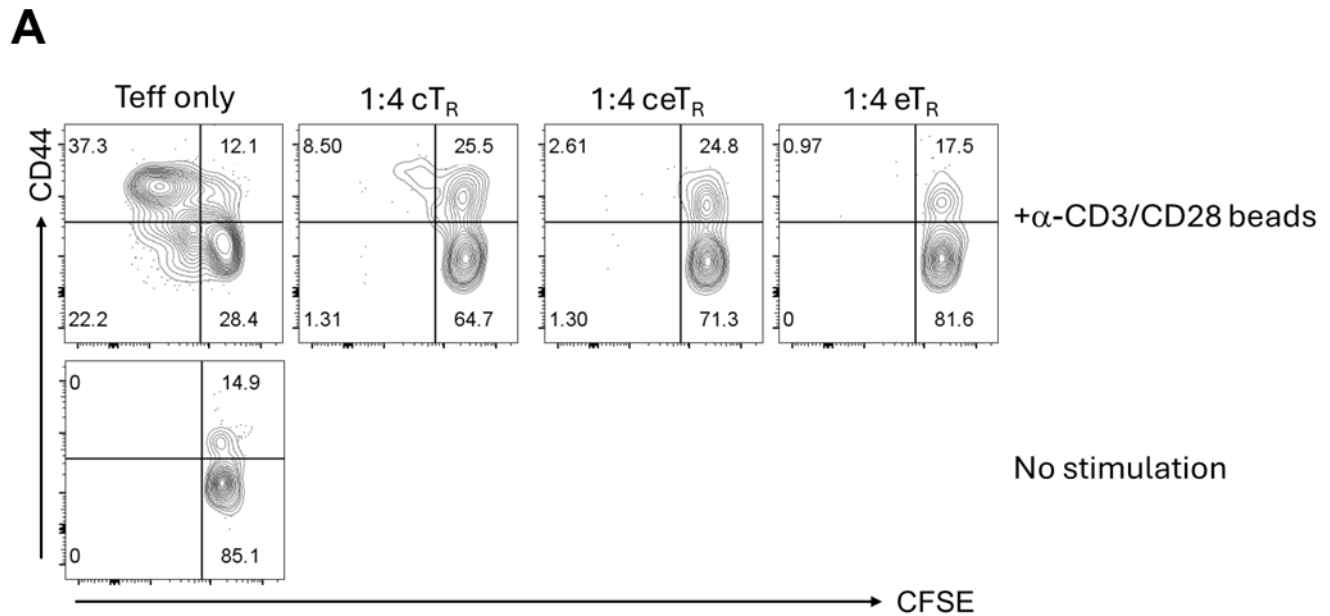


Figure S2. Fc.Mut24-expanded c_{TR}, ce_{TR} and e_{TR} are highly suppressive, related to Figure 2. c_{TR}, ce_{TR} or e_{TR} were sorted from the spleens of B6.Foxp3-mRPF mice treated with Fc.Mut24 3 days previously (gated as in Figure 2), and their ability to suppress activation-induced proliferation and upregulation of CD44 by CFSE-labeled and sorted Foxp3-mRFP-negative T_{conv} was assessed. **A.** Representative flow cytometric analysis of cell proliferation (based on CFSE-dilution) and CD44 expression by gated Teff cells activated alone or mixed in a 1:4 T_R:T_{conv} ratio with the indicated population of sorted T_R. **B.** Analysis of %Suppression of T_{conv} proliferation and CD44 upregulation by each of the sorted T_R populations as indicated. No statistical significance was observed in the ability of any of the T_R populations to suppress either proliferation or CD44 upregulation by one-way ANOVA with Tukey's multiple comparison test.

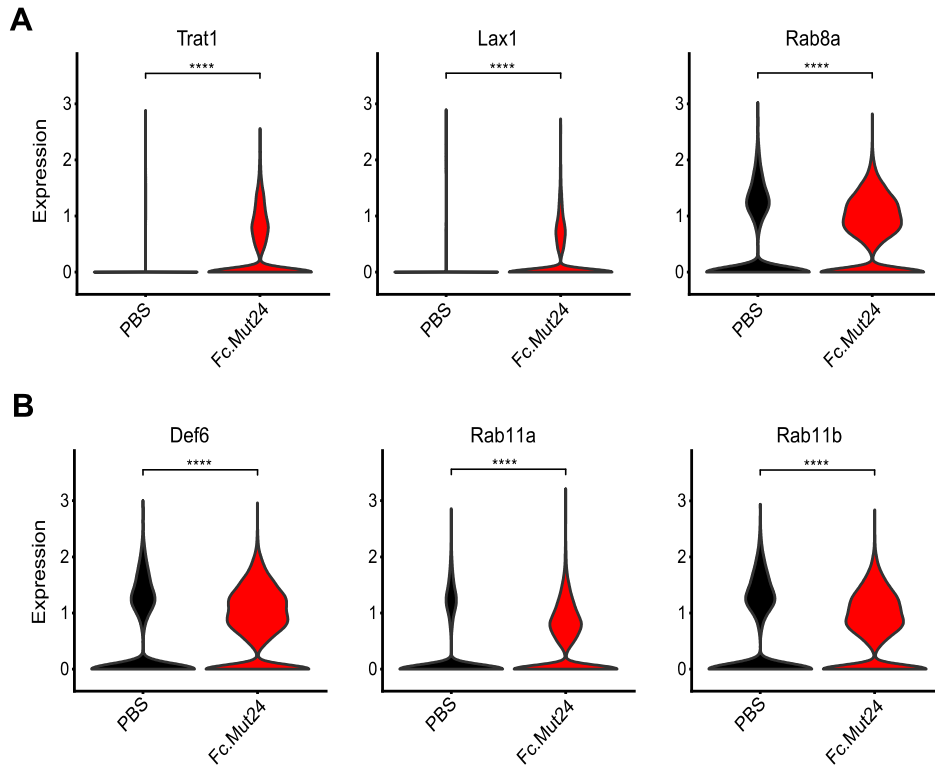


Figure S3. Fc.Mut24 increases the expression of genes involved in CTLA-4 trafficking and recycling in Foxp3⁺ T_R cells, related to Figure 3. Splenocytes from C57BL/6 Foxp3-mRFP treated with PBS, Fc.WT, or Fc.Mut24 were harvested 3 days later and sorted to obtain CD4⁺ Foxp3⁺ regulatory T (T_R) cells for single-cell RNA-seq (scRNA-seq). Violin plots showing normalized expression for the indicated genes involved in CTLA-4 trafficking to the cell surface **(A)** or endosomal recycling of CTLA-4 **(B)**.

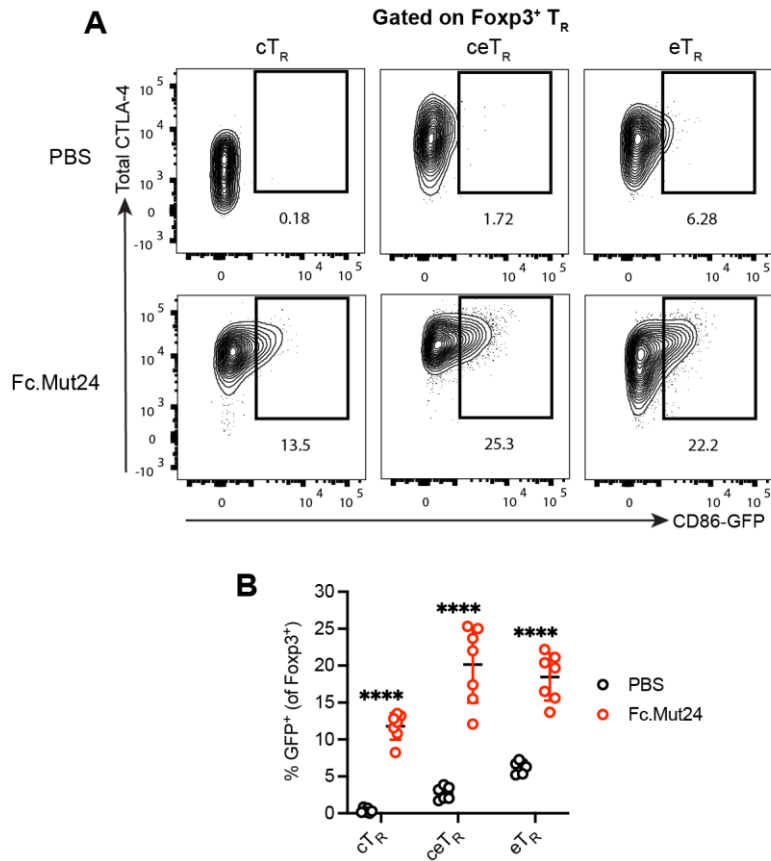


Figure S4. Fc.Mut24 increases CTLA-4-dependent transendocytosis by different Foxp3⁺ T_R cell populations, related to Figure 4. A. Capture of CD86-GFP by CD44^{lo} CD62L^{hi} central T_R (cT_R), CD44^{hi} CD62L^{hi} central effector T_R (ceT_R), and CD44^{hi} CD62L^{lo} effector T_R (eT_R) cells between different treatment groups. See Figure 4A for the experimental design schematic. Representative flow cytometry plots are shown. **B.** Graph shows mean \pm SD with individual data points ($n = 7$); **** $P \leq 0.0001$, multiple unpaired t -tests. Data are representative of 2-3 independent experiments.

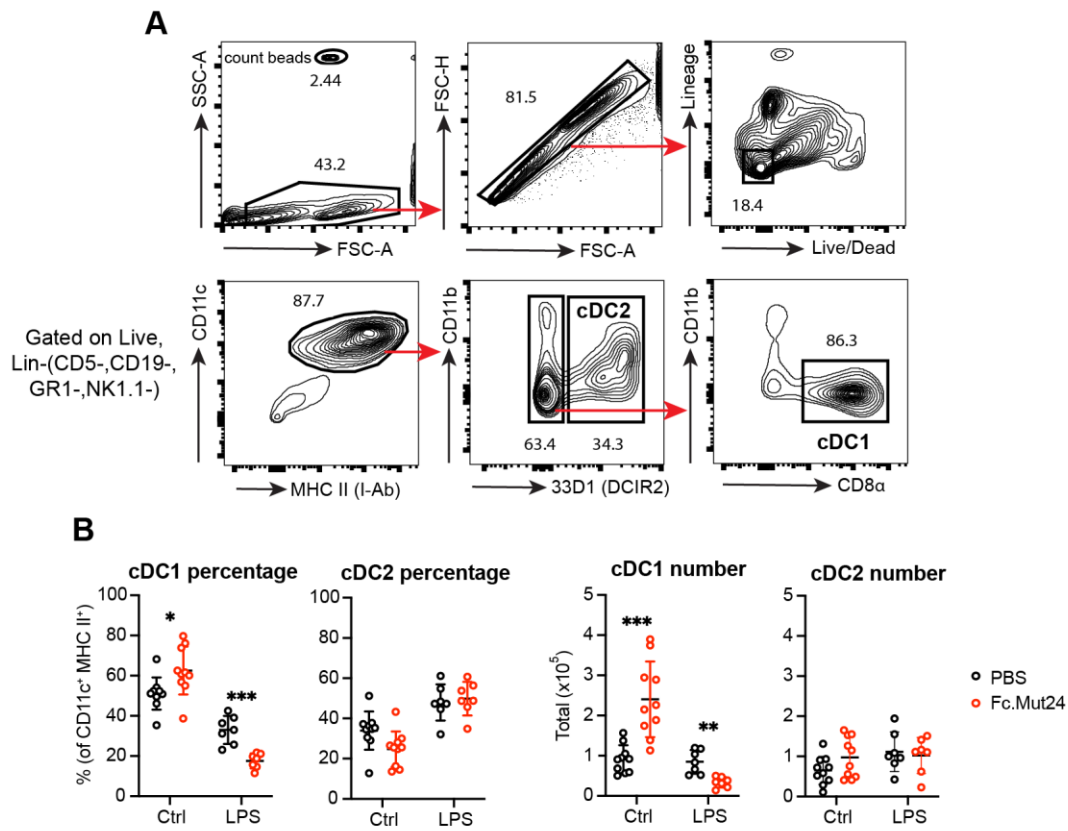


Figure S5. Fc.Mut24 expands cDC1s in C57BL/6 mice, related to Figure 5. A. Representative gating strategy for CD11c⁺MHC II⁺33D1⁻ CD8α⁺ cDC1 and CD11c⁺MHC II⁺33D1⁺ cDC2. **B.** Percentage and number of cDC1 and cDC2 between different treatment groups. See Figure 5A for the experimental design schematic. Graphs show mean ± SD with individual data points (n = 7 to 10); * $P \leq 0.05$, ** $P \leq 0.01$, *** $P \leq 0.001$, multiple unpaired t -tests. Ctrl, Control. Data are representative of 2-3 independent experiments.

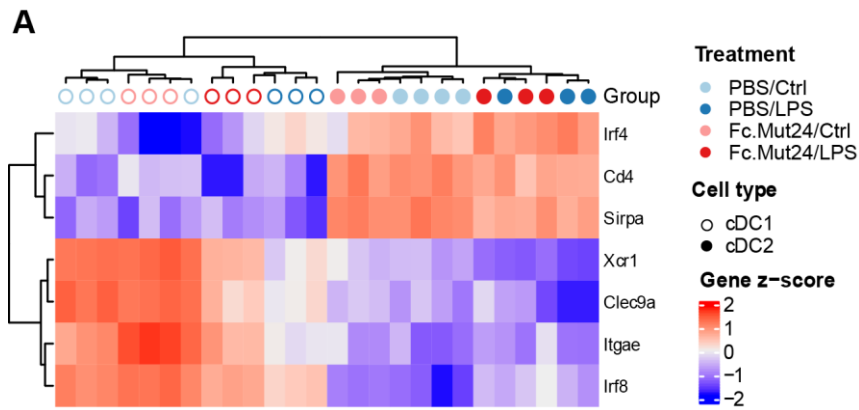


Figure S6. Expression of known identity markers in dendritic cells after Fc.Mut24 treatment, related to Figure 5. Unstimulated (Ctrl) or LPS-stimulated splenocytes from C57BL/6 mice treated with PBS or Fc.Mut24 were harvested 4 days post-treatment and sorted to obtain cDC1 or cDC2 populations for bulk RNA-seq. See Figure 5A for the experimental design schematic. **A.** Heatmap showing the mean expression (z-score) of select genes differentially expressed between cDC1 and cDC2 for all treatment groups.

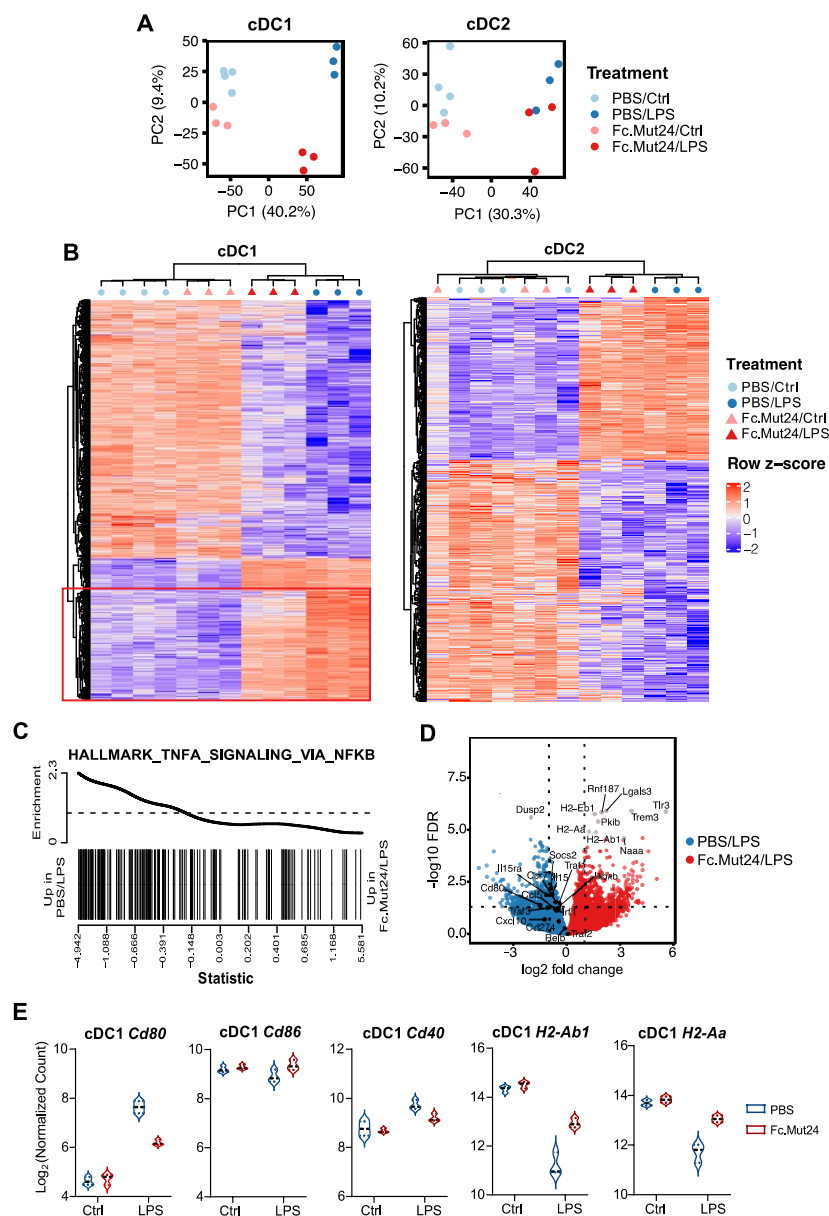


Figure S7. Impact of Fc.Mut24 treatment of the LPS-mediated transcriptional response in dendritic cells, related to Figure 5. Unstimulated (Ctrl) or LPS-stimulated splenocytes from C57BL/6 mice treated with PBS or Fc.Mut24 were harvested 4 days post-treatment and sorted to obtain cDC1 or cDC2 populations for bulk RNA-seq. See Figure 5A for the experimental design schematic. **A.** Principal component analysis of global transcriptional signatures from indicated cell types and treatment groups. **B.** Heatmap showing the mean expression (z-score) of all genes differentially expressed between LPS and Ctrl samples in cDC1 and cDC2 (FDR < 0.01, \log_2 fold-change > 1). In cDC1s, a subset of genes (red rectangle) have diminished LPS-mediated upregulation in the presence of Fc.Mut24. **C.** Gene set enrichment analysis of cDC1s showing that the Hallmark TNFA Signaling via NFKB pathway is enriched in PBS/LPS relative to Fc.Mut24/LPS samples (FDR = 0.005). **D.** Volcano plot of cDC1s comparing Fc.Mut24 treatment versus PBS with the top ten differentially expressed genes by FDR (gray) and select NF κ B-regulated genes (black) highlighted. **E.** Violin plots of cDC1s showing \log_2 normalized counts for select genes: *Cd80*, *Cd86*, *CD40*, *H2-Ab1* (I-A^b beta chain), and *H2-Aa* (I-A^b alpha chain).

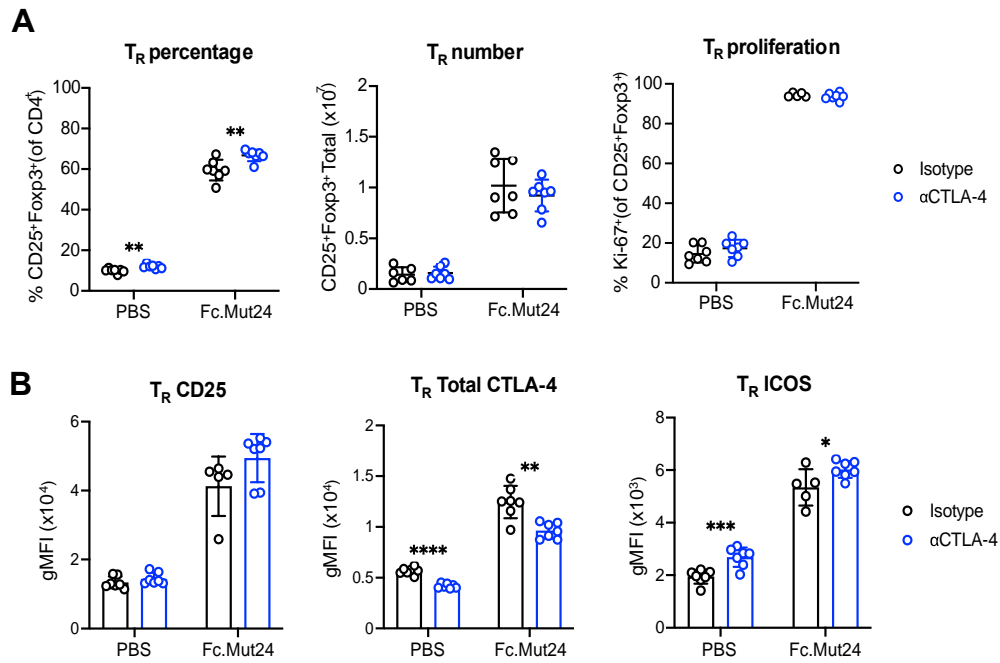


Figure S8. The anti-CTLA-4 antibody 4F10 does not cause Foxp3⁺ T_R cell depletion during Fc.Mut24 treatment, related to Figure 5. Splenocytes from C57BL/6 mice treated with PBS or Fc.Mut24 followed by isotype or anti-CTLA-4 antibody and LPS stimulation were analyzed by flow cytometry. Gates were set on live, CD4⁺ cells. **A&B.** Graphs show mean ± SD with individual data points (n = 7); **P* ≤ 0.05, ***P* ≤ 0.01, ****P* ≤ 0.001, *****P* ≤ 0.0001, multiple unpaired *t*-tests. Data are representative of 2-3 independent experiments. **B.** Gates were set on CD25⁺ Foxp3⁺ T_R for analysis of the expression of indicated T_R activation markers.

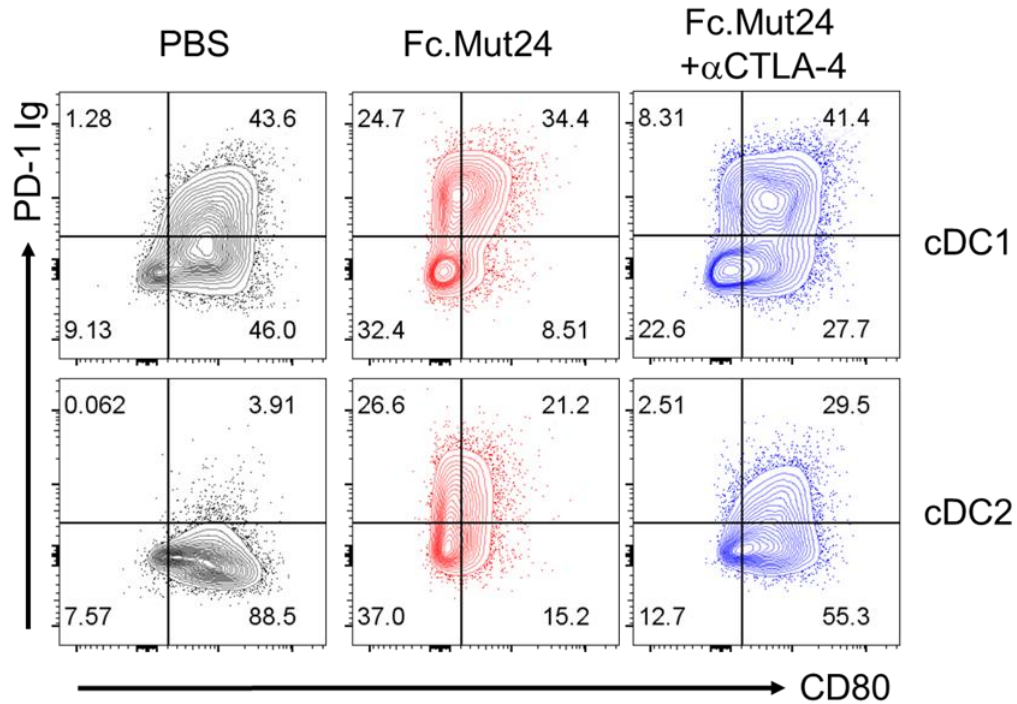
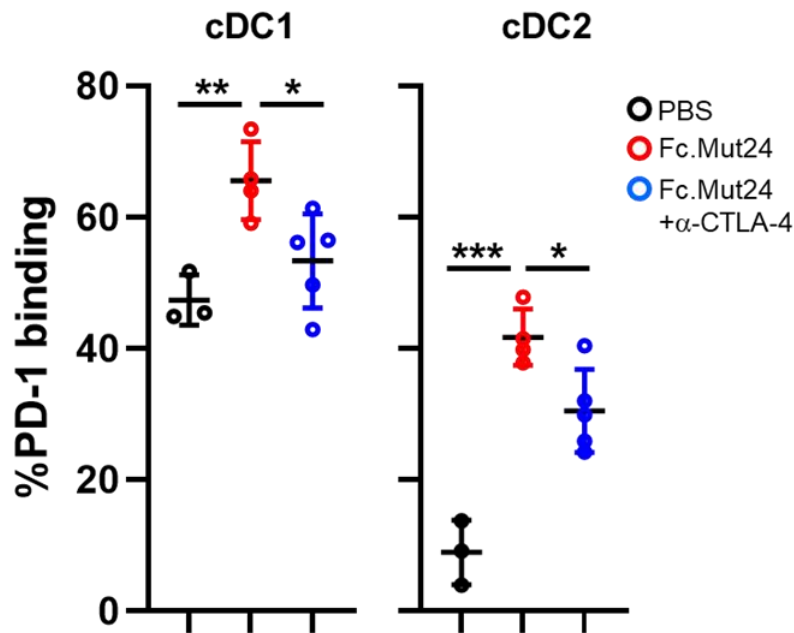
A**B**

Figure S9. Fc.Mut24 treatment enhances PD-1 binding to splenic DCs, related to Figure 5. Splenic DCs (gated as in Fig. S5) from C57BL/6 mice treated with PBS, Fc.Mut24, or Fc.Mut24 + anti-CTLA-4 antibody followed by LPS were analyzed by flow cytometry. **A.** Representative flow cytometry plots showing CD80 expression and PD-1 binding by gated cDC1 and cDC2. **B.** Graphs show mean \pm SD with individual data points in each group as indicated ($n = 3-5$ mice/group); $*P \leq 0.05$, $**P \leq 0.01$, $***P \leq 0.001$, by one-way ANOVA with Tukey's multiple comparison test. Data are representative of 2 independent experiments.

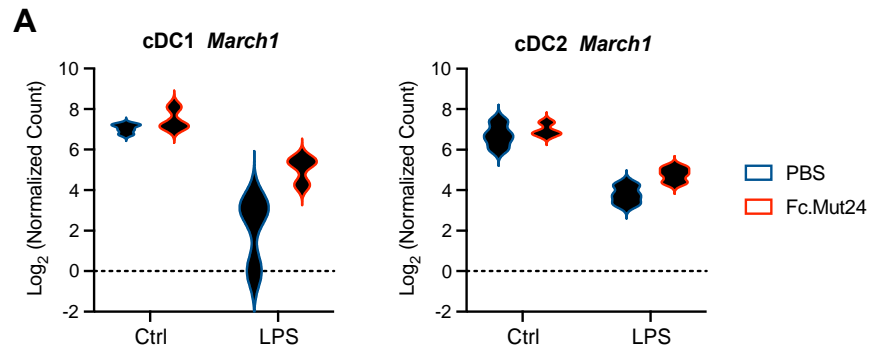


Figure S10. Fc.Mut24 increases *March1* gene expression in cDC1s from C57BL/6 mice, related to Figure 6. Unstimulated (Ctrl) or LPS-stimulated splenocytes from C57BL/6 mice treated with PBS or Fc.Mut24 were harvested 4 days post-treatment and sorted to obtain cDC1 or cDC2 populations for bulk RNA-seq. See Figure 5A for the experimental design schematic **A**. Violin plots showing Log₂ normalized counts for *March1* gene expression between treatment groups.

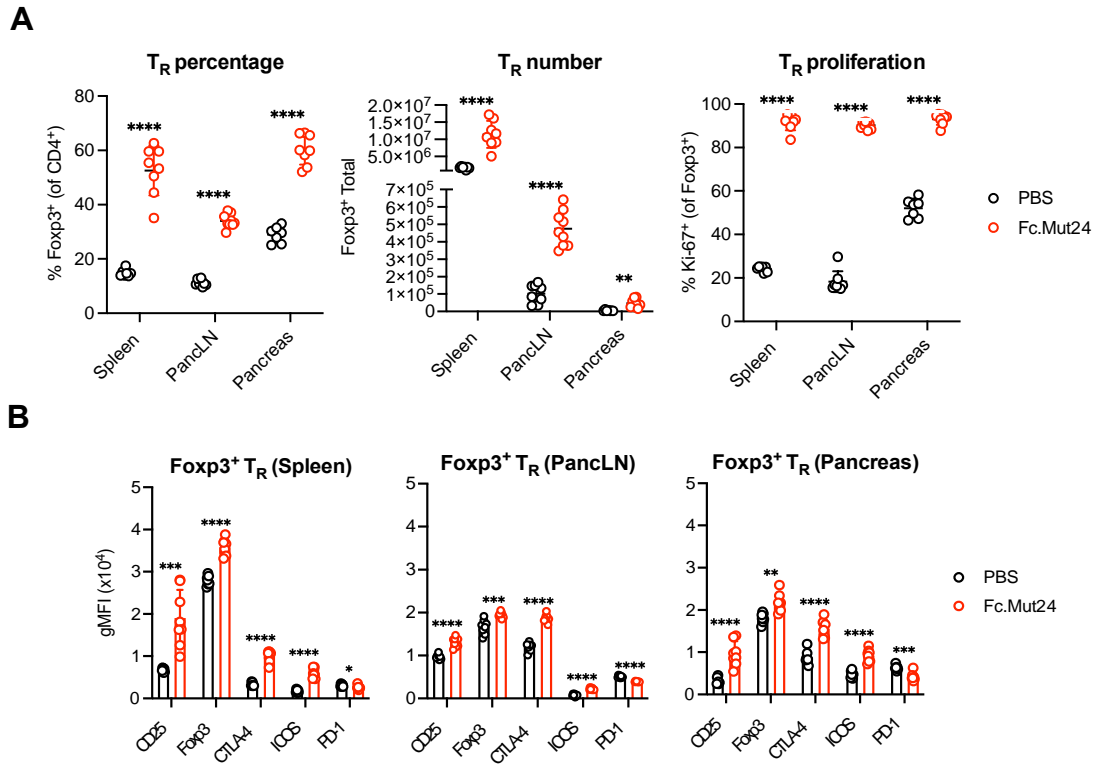


Figure S11. Fc.Mut24 promotes Foxp3⁺ T_R cell activation in NOD mice, related to Figure 7. The spleen, pancreatic lymph node (PancLN), and pancreas of 8-10-week-old female non-obese diabetic (NOD) mice treated with PBS or Fc.Mut24 were harvested 4 days later and analyzed by flow cytometry. Gates were set on live, CD45⁺, CD4⁺ cells. Percentage, number, and proliferation (Ki-67⁺) of Foxp3⁺ T_R cells **(A)** and expression of indicated T_R activation markers **(B)**. **A&B.** All graphs show mean \pm SD with individual data points ($n = 6$ to 9); * $P \leq 0.05$, ** $P \leq 0.01$, *** $P \leq 0.001$, **** $P \leq 0.0001$, multiple unpaired t -tests. Data are representative of 2-3 independent experiments.

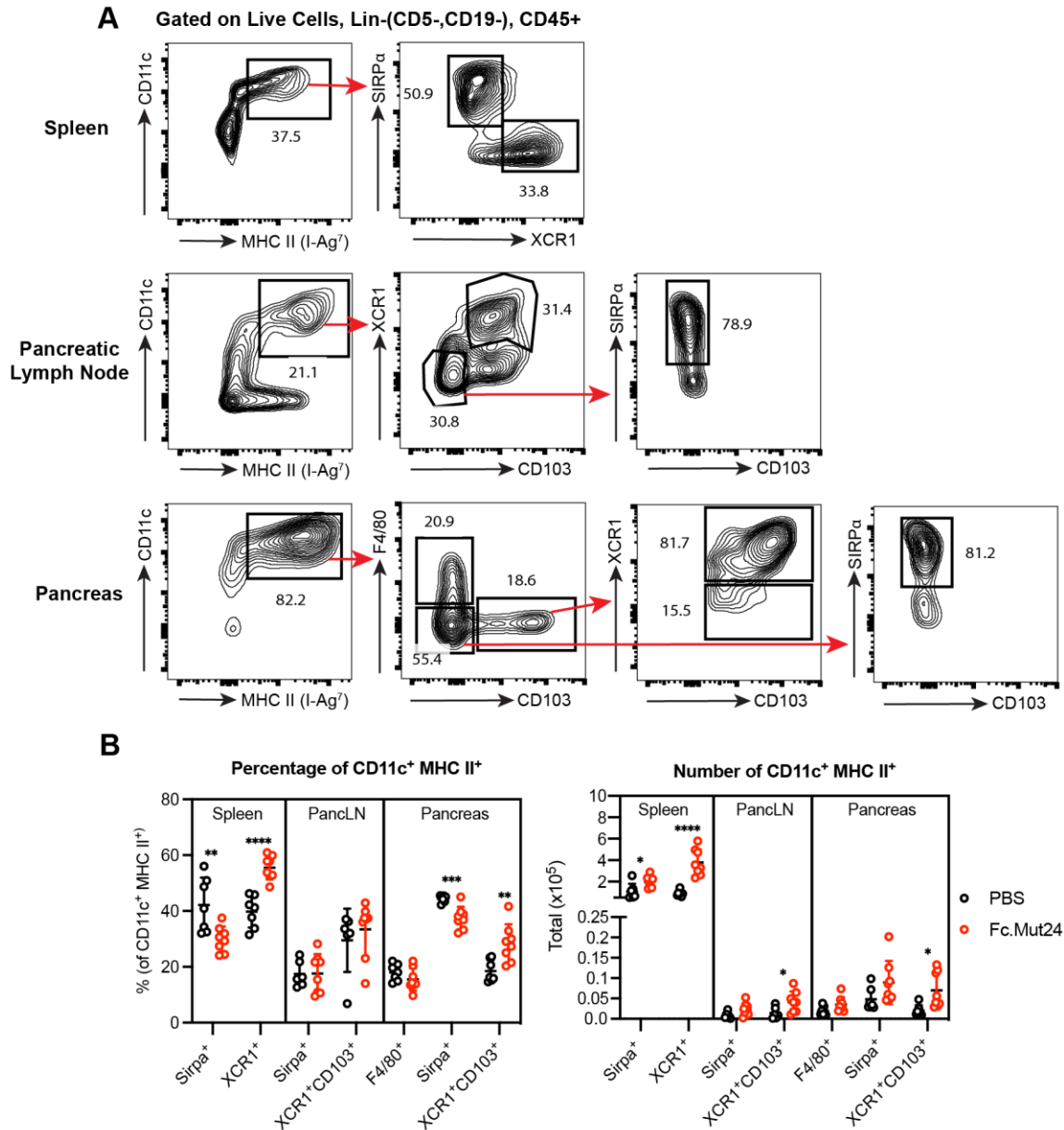


Figure S12. Fc.Mut24 expands cDC1s in NOD mice, related to Figure 7. The spleen, pancreas, and pancreatic lymph node (PancLN) of 8-10-week-old female non-obese diabetic (NOD) mice treated with PBS or Fc.Mut24 were harvested 4 days later and analyzed by flow cytometry. **A.** Representative gating strategy for the CD11c⁺ MHC II⁺ antigen-presenting cell subsets in different tissues. **B.** Percentage and number of different subsets of CD11c⁺ MHC II⁺ antigen-presenting cells. Graphs show mean \pm SD with individual data points (n = 6 to 8); * $P \leq 0.05$, ** $P \leq 0.01$, *** $P \leq 0.001$, **** $P \leq 0.0001$, multiple unpaired *t*-tests. Data are representative of 2-3 independent experiments.

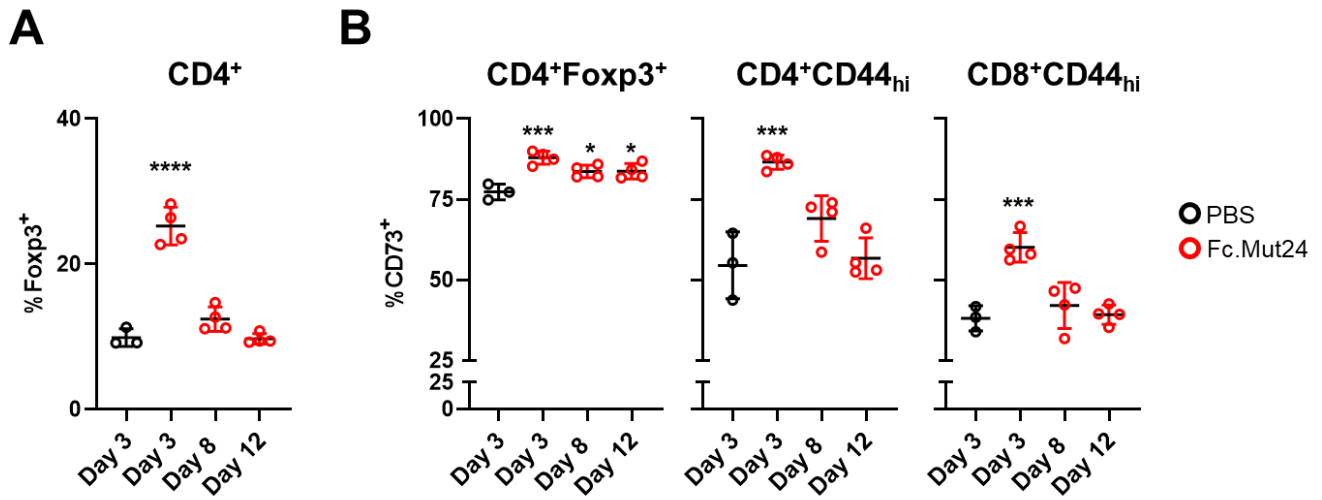


Figure S13. CD73 expression is transiently upregulated by CD4⁺ and CD8⁺ effector T cells after Fc.Mut24 treatment of NOD mice, related to Figure 7. T cells from the pancLN of NOD mice treated with either PBS (black symbols) or Fc.Mut24 (red symbols) were analyzed by flow cytometry. **A.** Frequency of Foxp3⁺ T_R among total CD4⁺ T cells at the indicated times after treatment. **B.** Frequency of CD73⁺ cells among CD4⁺Foxp3⁺, CD4⁺Foxp3⁻CD44^{hi}, and CD8⁺CD44^{hi} populations at the indicated times after treatment. * $P \leq 0.05$, ** $P \leq 0.01$, *** $P \leq 0.001$, **** $P \leq 0.0001$, one-way ANOVA with Tukey's multiple comparison test. Data are from 1 experiment.