

Supplementary Information

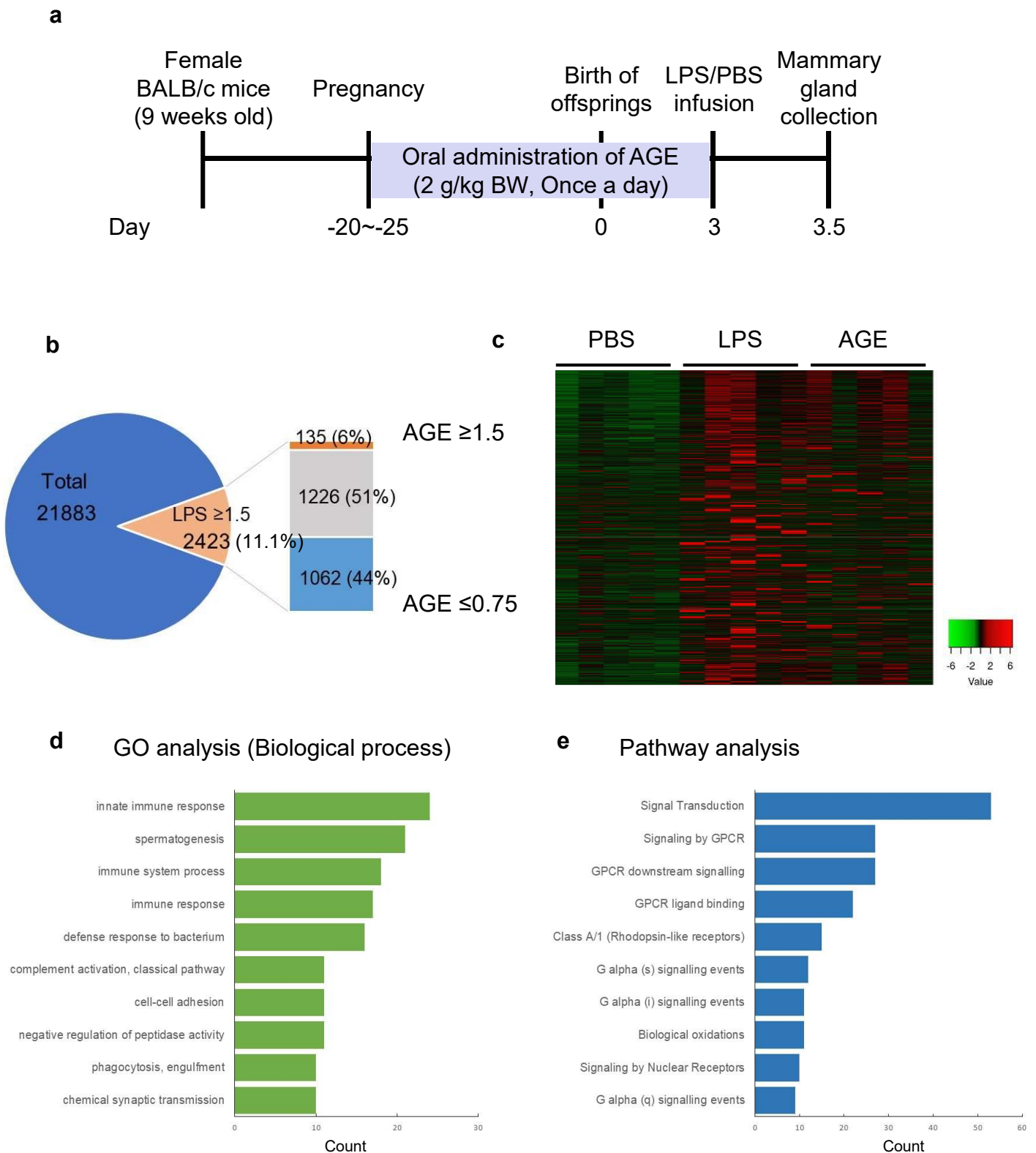
***S*-Allylmercaptocysteine inhibits TLR4-mediated inflammation through enhanced formation of inhibitory MyD88 splice variant in mammary epithelial cells**

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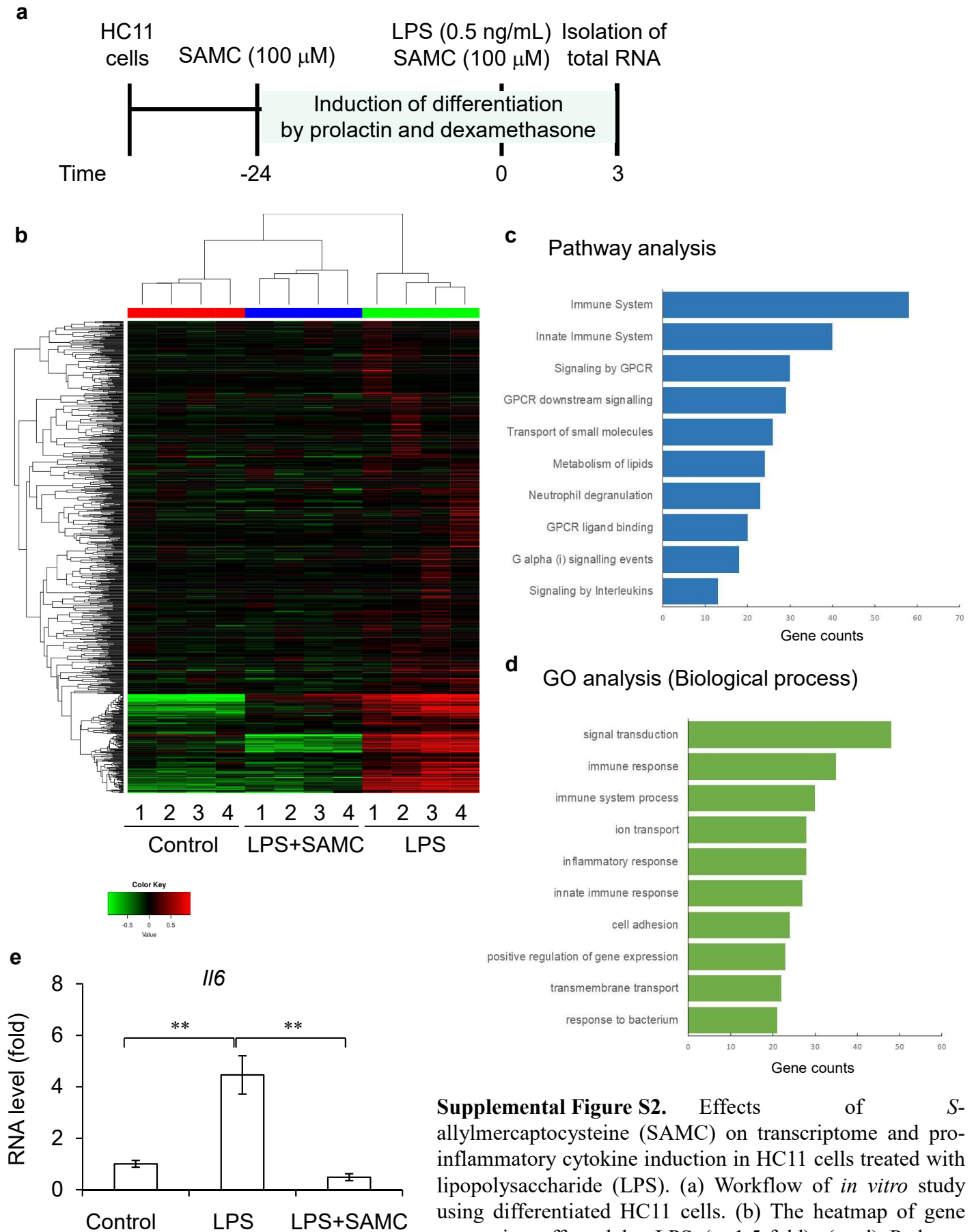
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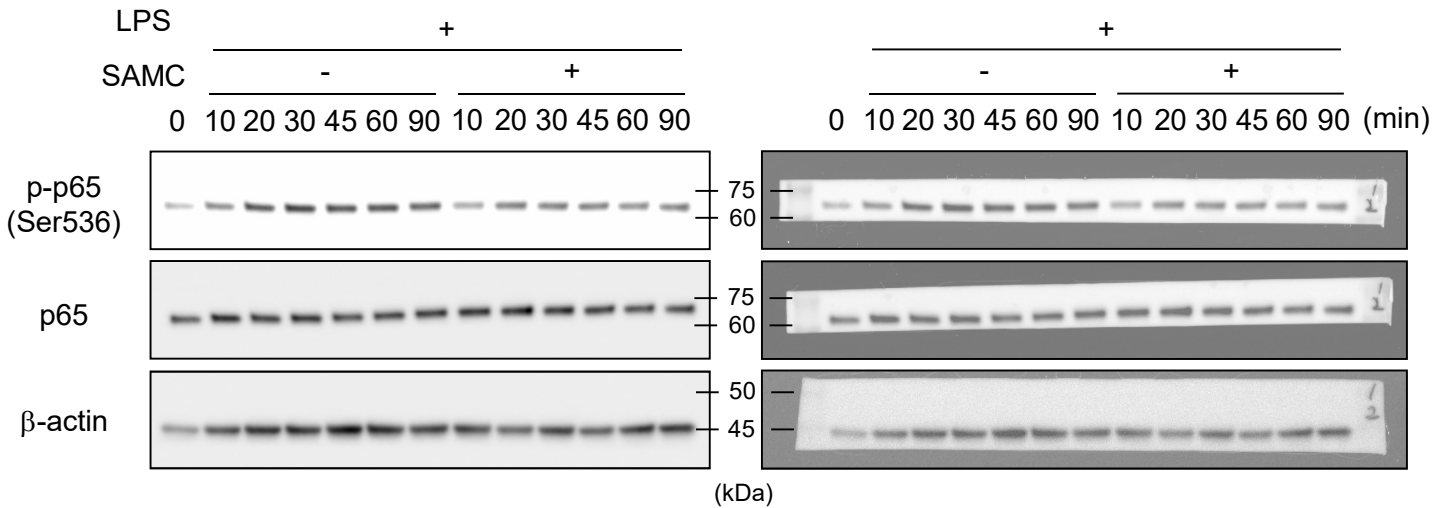


Supplemental Figure S1. Effects of aged garlic extract (AGE) treatment on transcriptome of the mammary glands treated with lipopolysaccharide (LPS) in mice ($n=5$). (a) Workflow of animal treatment. (b) The proportion of genes increased by intramammary LPS injection (≥ 1.5 -fold vs PBS-injected side, 11.1%) and the proportions of these genes increased (≥ 1.5 -fold vs LPS-injected tissues of control mice, 6.0%) or decreased (≤ 0.75 -fold, 44.0%) by AGE administration. (c) The heatmap of gene expression increased by LPS (≥ 1.5 -fold). PBS and LPS columns show the gene expression in the mammary glands of PBS- and LPS-injected side of control mice, respectively. AGE column shows the gene expression in the LPS-injected mammary glands of mice administered with AGE. (d, e) Gene ontology (GO) term annotation for biological process (d) and pathway analysis (e) of the genes increased by LPS injection (≥ 1.5 -fold) and decreased by AGE administration (≤ 0.75 -fold).

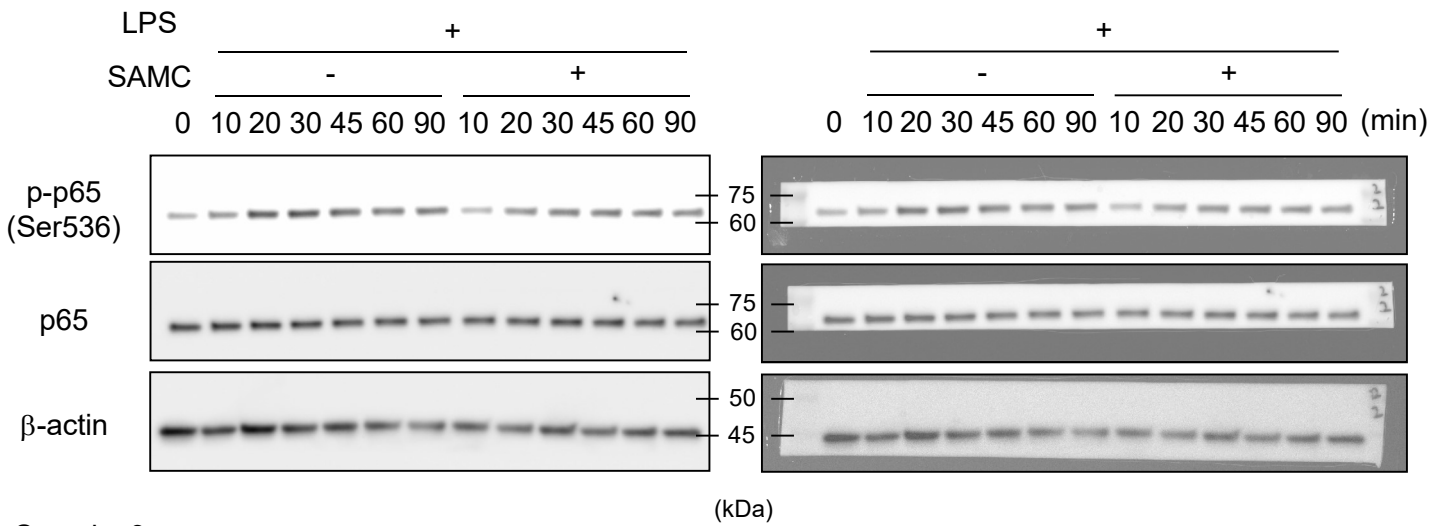


Supplemental Figure S2. Effects of *S*-allylmercaptocysteine (SAMC) on transcriptome and pro-inflammatory cytokine induction in HC11 cells treated with lipopolysaccharide (LPS). (a) Workflow of *in vitro* study using differentiated HC11 cells. (b) The heatmap of gene expression affected by LPS (≥ 1.5 -fold). (c, d) Pathway analysis (c) and GO term annotation for biological process (d) of the genes increased by LPS (≥ 1.5 -fold) and decreased by SAMC treatment (≤ 0.75 -fold). (e) mRNA expression of *Il6* in HC11 cells was examined by real-time quantitative PCR. Data are shown as mean \pm SD, $n = 4-5$. ** denotes significant difference ($p < 0.01$).

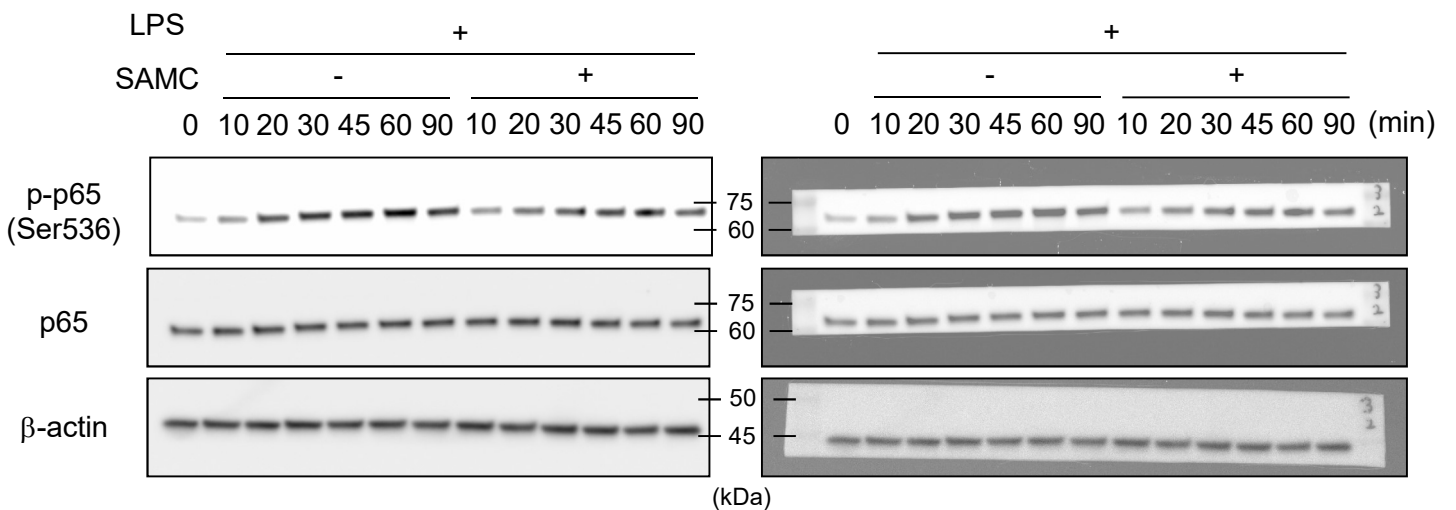
Sample: 1



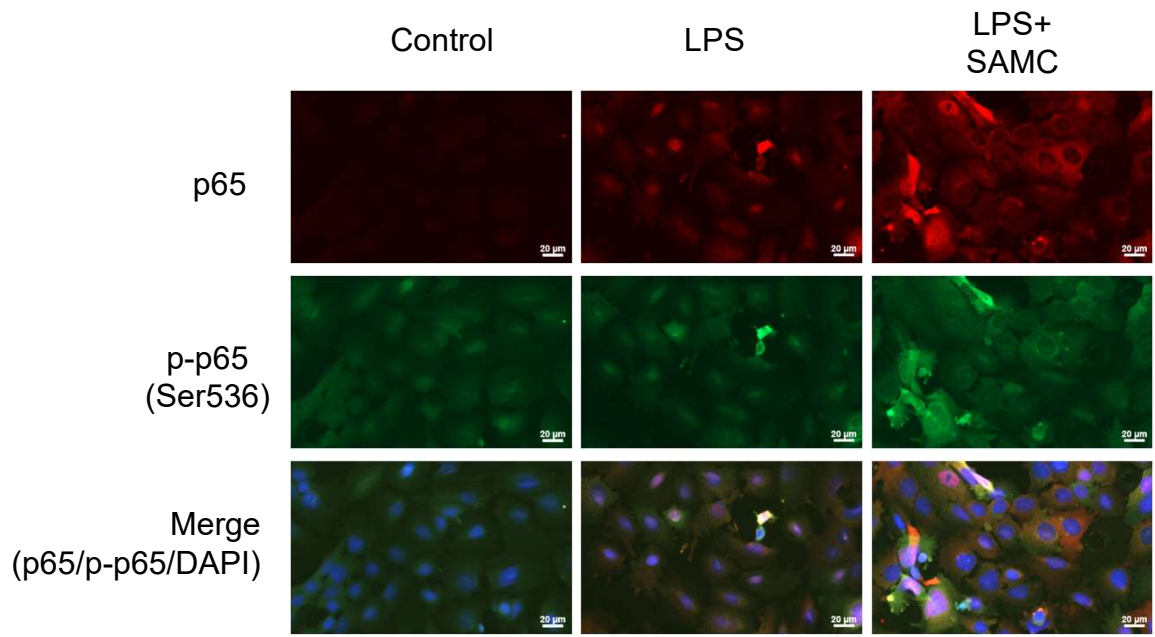
Sample: 2



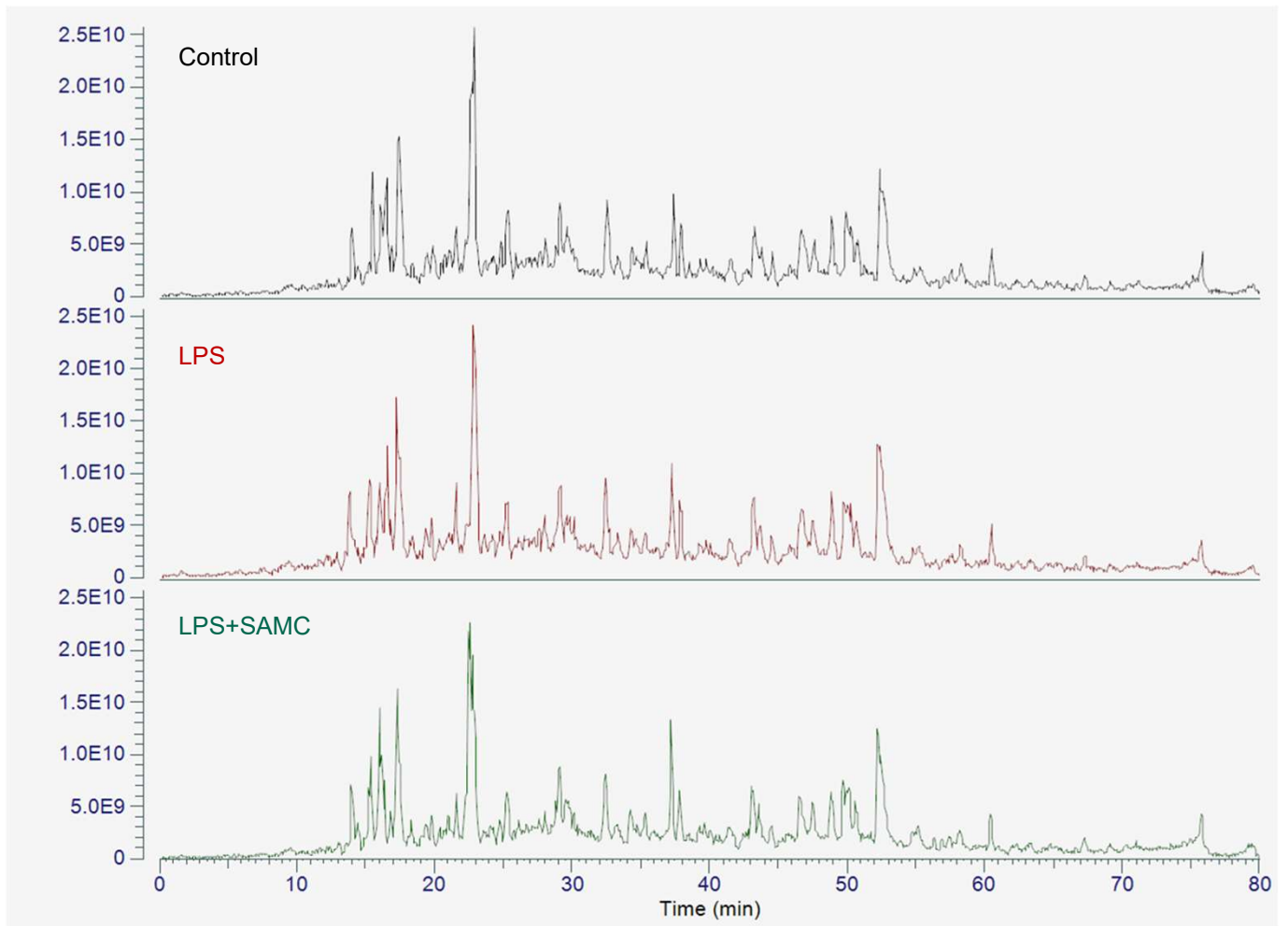
Sample: 3



Supplemental Figure S3. Western blots of NF-κB p65 phosphorylation in HC11 cells treated with or without lipopolysaccharide (LPS) and *S*-allylmercaptocysteine (SAMC). Molecular size markers are indicated between the blots used in Fig. 1c. Kda = kilodalton.

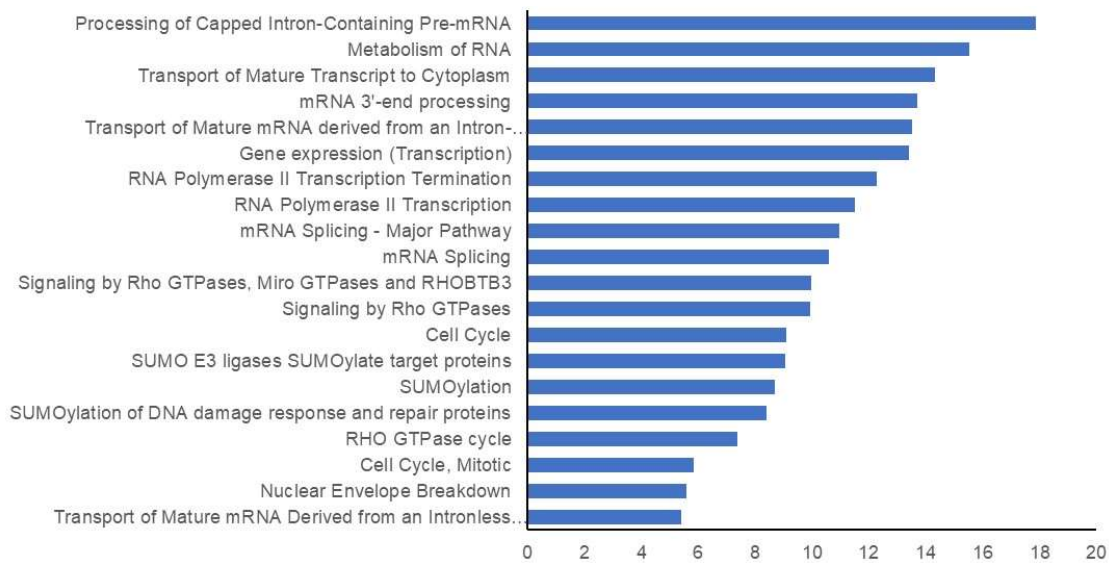


Supplementary Figure S4. Effect of *S*-allylmercaptocysteine (SAMC) on lipopolysaccharide (LPS)-induced activation of NF- κ B p65. HC11 cells were immunostained with anti-p65 (red), anti-p-p65 (Ser536) (green) and DAPI (nucleus) (blue) after stimulation with LPS (100 ng/mL) in the presence or absence of SAMC (300 μ M) for 1 h. Scale bar, 50 μ m.

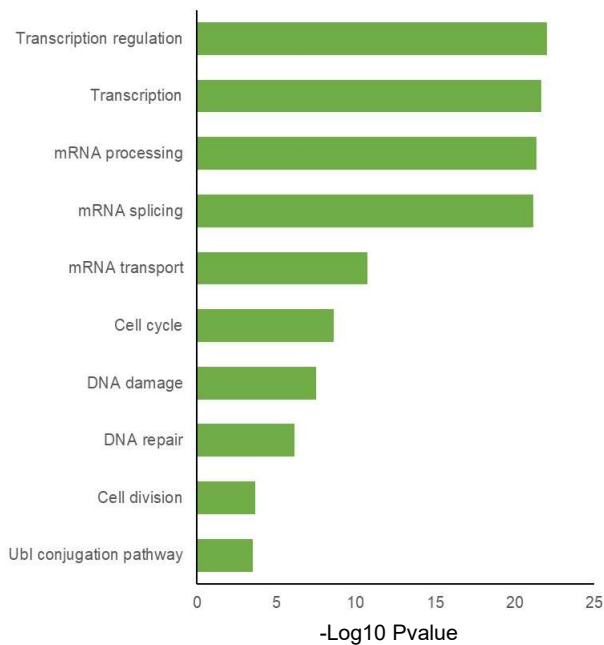


Supplementary Figure S5. Total ion current chromatograms obtained from data-independent acquisition-MS analysis of phospho-peptide extracts.

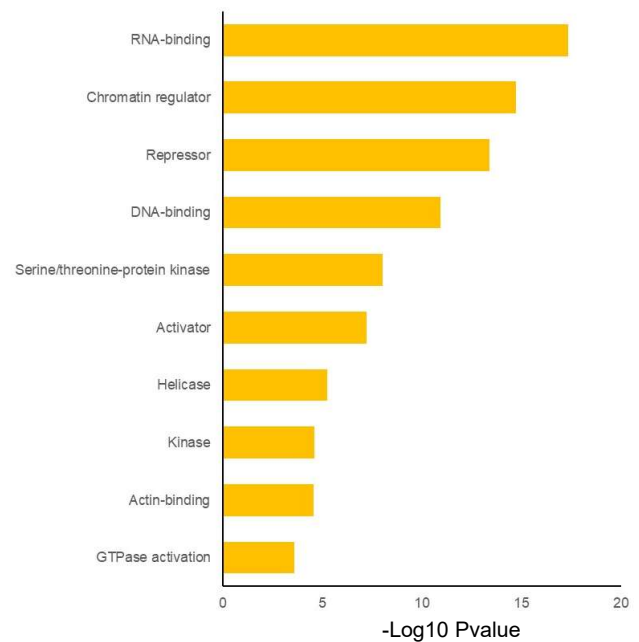
a Pathway analysis



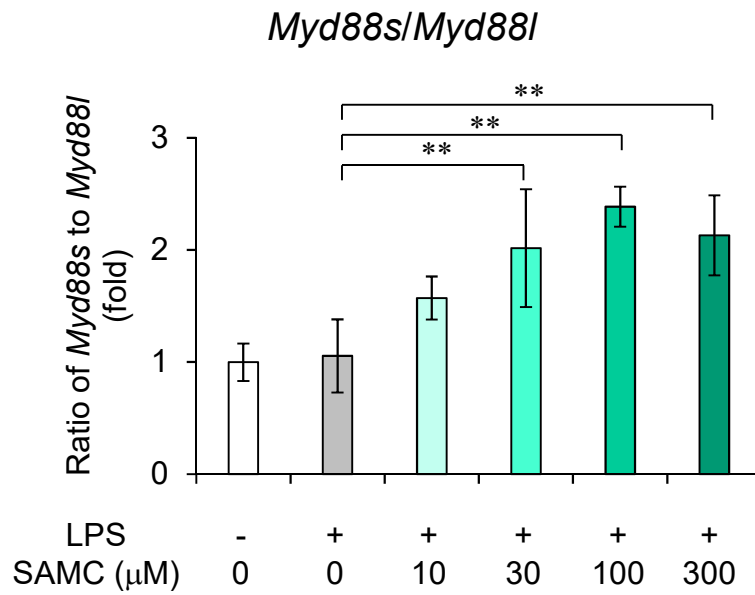
b Biological process



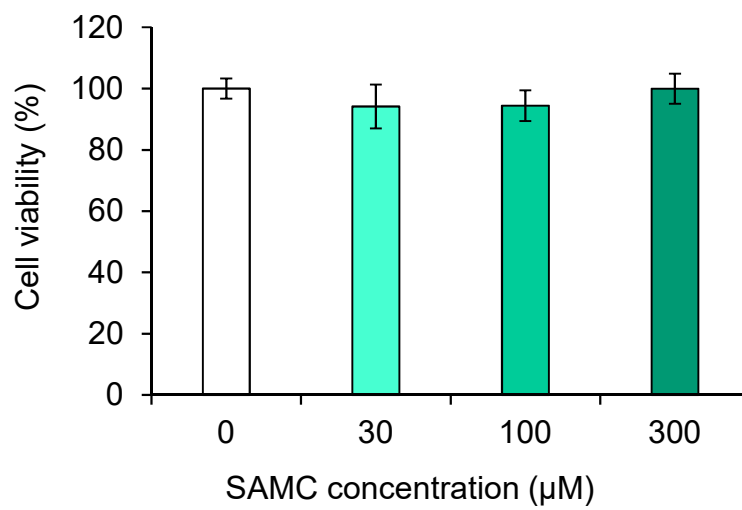
c Molecular function



Supplementary Figure S7. Effect of lipopolysaccharide (LPS) on phosphorylation of mRNA splicing-related proteins. Pathway analysis (a), gene ontology term annotation for biological process (b) and molecular function (c) of the genes increased by LPS (≥ 1.5 -fold vs. Control).



Supplementary Figure S8. *S*-Allylmercaptocysteine (SAMC) increases the mRNA expression ratio of *Myd88s* to canonical *Myd88l*. Real-time qPCR analysis of the concentration-dependent effect of SAMC (10-300 μM) on the mRNA expression ratio of *Myd88s* to canonical *Myd88l* in HC11 cells treated with lipopolysaccharide (LPS) stimulation for 2 h. Data are shown as mean ± SD, n = 5. ** denotes significant difference ($p < 0.01$).



Supplementary Figure S9. *S*-Allylmercaptocysteine (SAMC) has no effect on the viability of HC11 cells. After treatment with SAMC (30-300 µM) for 24 h, the HC11 cell viability was evaluated by WST-8 assay. Data are shown as mean \pm SD, n = 5.

Supplementary Table S1. Primer sequences used in this study.

Genes		Primer sequences
Mouse <i>Il6</i>	forward	5'-ATGATGCTGGTGACAACCACGG-3'
	reverse	5'-CAGGTCTGTTGGGAGTGGTATCC-3'
Mouse <i>Tnf</i>	forward	5'-CTGAACTTCGGGGTGATCGG-3'
	reverse	5'-GGCTTGTCACTCGAATTTTGAGA-3'
Mouse <i>Cxcl1</i>	forward	5'-CACTGCACCCAAACCGAAGTC-3'
	reverse	5'-GGGAGCTTCAGGGTCAAGGC-3'
Mouse <i>Ccl2</i>	forward	5'-CATCCACGTGTTGGCTCA-3'
	reverse	5'-GATCATCTTGCTGGTGAATGAGT-3'
Mouse <i>Myd88s</i>	forward	5'-GGAGCTGAAGTCGCGCATCGGACAAAC-3'
	reverse	5'-GTCTGTTCTAGTTGCCGGATCATCTCCTGCAC-3'
Mouse <i>MyD88l</i>	forward	5'-ACCACCCTTGATGACCCCCT-3'
	reverse	5'-GTCACGGTCGGACACACACA-3'
Mouse <i>Hprt</i>	forward	5'-GCAGTACAGCCCCAAAATGG-3'
	reverse	5'-TCCAACAAAGTCTGGCCTGT-3'