

Supplementary Information for

Late-Stage (Radio)Fluorination of Alkyl Phosphonates via Electrophilic Activation

Kaiqiang Zhang, *et al.*

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1 Supplementary Notes

1.1 General reagent information

Commercially available reagents were procured from Energy Chemical (China) and Sigma-Aldrich (Switzerland) and utilized as received unless specified otherwise. Anhydrous *N,N*-dimethylformamide (DMF), anhydrous tetrahydrofuran (THF), anhydrous dimethyl sulfoxide (DMSO), all stored over molecular sieves, were purchased from Energy Chemical. Additional common solvents were procured from Sinopharm (China) and employed without further treatment. Deuterated solvents including CDCl₃, D₂O, CD₃OD, CH₃OD, and DMSO-*d*₆ were obtained from Energy Chemical; CD₂Cl₂ was procured from Cambridge Isotope Laboratories (United States). Ultra-pure water was obtained from an ELGA PURELAB flex system (UK).

The compounds (**S16–S36**, E[c(RGDyK)]₂) were procured from commercial suppliers and employed as reagents in the direct fluorination of alkyl phosphonates *via* electrophilic activation.

1.2 General analytical information

All fluorination reactions were carried out with magnetic agitation in oven-dried glass-bottomed vessels. Subsequent concentration of organic solutions was performed under reduced pressure using a Heidolph rotary evaporator (Hei-VAP Value, Germany). The reaction progress was tracked *via* thin-layer chromatography (TLC) on aluminum plates (2.5 cm × 5 cm) coated with silicone GF254, supplied by Energy Chemical, or on polyamide film plates sourced from Lu Qiao Si-Jia Biochemistry Plastics Company (China). Chromatographic visualization utilized a 254 nm ultraviolet fluorescence quenching technique, alkaline potassium permanganate staining, or iodine staining.

¹H, ¹³C, ³¹P and ¹⁹F nuclear magnetic resonance (NMR) spectra were recorded on a Zhongke-Niujin AS 400 MHz or Bruker AVANCE 600 MHz NMR spectrometer (China) at room temperature. All the chemical shifts (δ) for protons were reported in ppm downfield from tetramethylsilane (TMS) and were referenced to residual protium in the deuterated solvents (¹H NMR: CD₃OD at 3.31 ppm, CDCl₃ at 7.26 ppm). Chemical shifts for carbon signals were reported in ppm and were referenced to the carbon resonances of the solvent peak (¹³C NMR: CD₃OD at 49.00 ppm, CDCl₃ at 77.00 ppm). Multiplicity was described with singlet (s), doublet (d) and multiplet (m). Coupling constants (*J*) were given in Hertz (Hz). High resolution mass spectrometry (HRMS)

data is recorded on a Waters Xevo G2-XS ToF mass Spectrometer (USA) or Thermo Q-Exactive Mass Spectrometer (USA). High-performance liquid chromatography (HPLC) analysis was performed using a NanoChrom C18 column (5 μm , 4.6 \times 250 mm, China). HPLC purification was carried out on a SEP Basic-C18 semi-preparative column (120A 5 μm 10 \times 250 mm China).

1.3 General radiochemistry information

Activation of the Sep-Pak® Plus Short C18 cartridge (Waters, Part No. WAT020515): Using a 10 mL syringe, extract 5 mL of methanol and slowly wash the C18 column. Then, push 10 mL of air through the column to remove any remaining methanol. Next, wash the C18 column with 10 mL of pure water. Finally, push 10 mL of air through the column to complete the activation process.

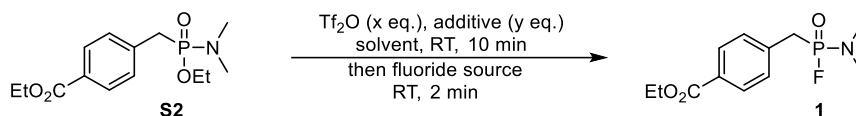
Activation of the Sep-Pak® light QMA cartridge (Waters, USA): The QMA column was activated using a 10 mL syringe to draw 5 mL of a pre-prepared 0.5 M K_2CO_3 solution, which was slowly washed through the column. Subsequently, 10 mL of air flushed the column to remove the liquid. Next, a wash with 10 mL of pure water was performed on the QMA column. Finally, an additional 10 mL of air completed the activation process.

HPLC analysis was performed using two Thermo Fisher Dionex ultra-mate 3000 instruments (United States) denoted as HPLC 1# and HPLC 2#, both equipped with an SPD-20A UV detector and an Elysia Raytest Gabi Star γ -radiation detector from Hungary, interconnected in series. In both HPLC 1# and 2#, the retention times of the radio peaks and the corresponding UV peaks were observed to shift by 0.10 to 0.20 min and 0.50 to 0.80 min, respectively, due to the distance between the UV detector (254 nm) and the radiation detector. Radio-TLC were acquired using an Eckert & Ziegler MS-1000F scanner from the United States, which was equipped with Mini-Scan and Flow-Count functionalities. All phosphonate fluorination precursors are now numbered as **S1–S57**, and the resulting fluorinated phosphonates are numbered **1–46**, with [^{18}F] added before the corresponding Arabic numerals to indicate the ^{18}F -products. Intermediates in the synthesis of phosphonate precursors are labeled as **i-1** to **i-10**, and unstable intermediates in mechanistic studies are represented as **INT-1** to **INT-5**. The bases in the additive screening are represented by the letter 'a' and superscript Arabic numerals from 1 to 11 (**a¹-a¹¹**).

2. Supplementary Methods

2.1 Condition optimization for fluorination of alkyl phosphonates

Table S1: Selected optimization table for alkyl phosphonates fluorination^[a]



Entry	$\text{Tf}_2\text{O}/x$ eq.	Additive/ y eq.	Solvnet ^[b]	Fluoride source	Conversion (%) ^[c]
1	1.0	Pyridine/1.0	CH_2Cl_2	TBAF	11 ± 6
2	1.0	Pyridine/1.5	CH_2Cl_2	TBAF	36 ± 3
3	1.5	Pyridine/1.0	CH_2Cl_2	TBAF	6 ± 2
4	2.0	Pyridine/1.5	CH_2Cl_2	TBAF	19 ± 3
5	2.0	Pyridine/2.0	CH_2Cl_2	TBAF	27 ± 5
6	1.5	Pyridine/2.0	CH_2Cl_2	TBAF	47 ± 1
7	1.5	Diphenylsulfane/2.0	CH_2Cl_2	TBAF	0
8	1.5	Pyridine/2.0	CH_2Cl_2	$\text{Et}_3\text{N} \cdot 3\text{HF}$	93 ± 3
9 ^[d]	TFAA, 1.5 eq.	Pyridine/2.0	CH_2Cl_2	$\text{Et}_3\text{N} \cdot 3\text{HF}$	trace
10	1.5	Pyridine/2.0	THF	$\text{Et}_3\text{N} \cdot 3\text{HF}$	trace
11	1.5	Pyridine/2.0	Toluene	$\text{Et}_3\text{N} \cdot 3\text{HF}$	11 ± 2
12	1.5	Pyridine/2.0	CH_3CN	$\text{Et}_3\text{N} \cdot 3\text{HF}$	35 ± 2
13	1.5	Pyridine/2.0	1,4-Dioxane	$\text{Et}_3\text{N} \cdot 3\text{HF}$	3 ± 1

^[a] Reactions were performed using 0.2 mmol **16**, tetrabutylammonium fluoride (TBAF 1.2 eq.)/ $\text{Et}_3\text{N} \cdot 3\text{HF}$ (0.5 eq.) in solvent (0.2 M).

^[b] CH_2Cl_2 : dichloromethane; THF: tetrahydrofuran; CH_3CN : acetonitrile.

^[c] Conversions determined by ^{31}P NMR.

^[d] Replace Tf_2O with trifluoroacetic anhydride (TFAA).

2.2 General synthetic procedures and characterization

General compounds information

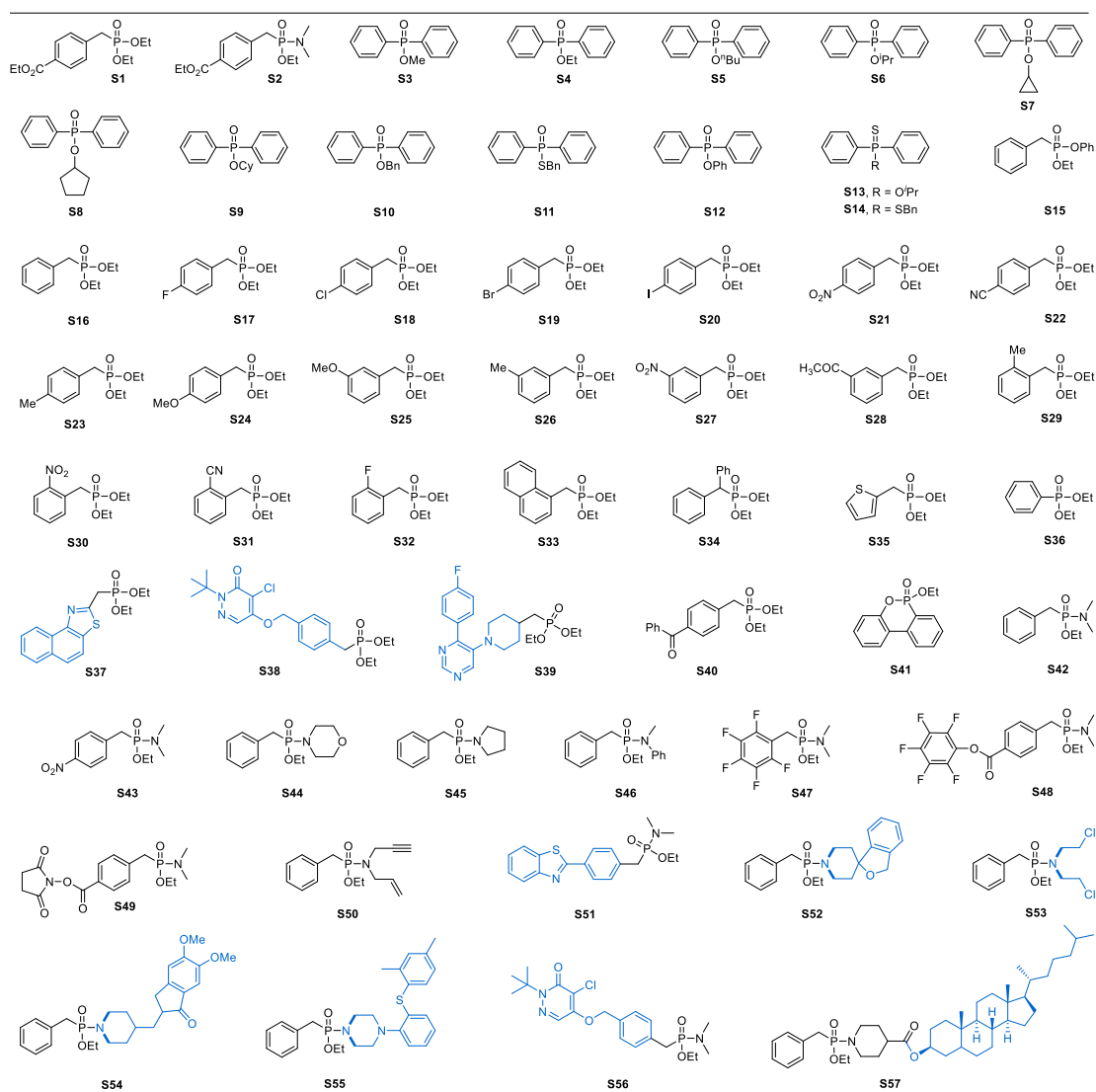


Fig. S1: An Overview of ethyl phosphonates (S1–S57) was discussed in this work.

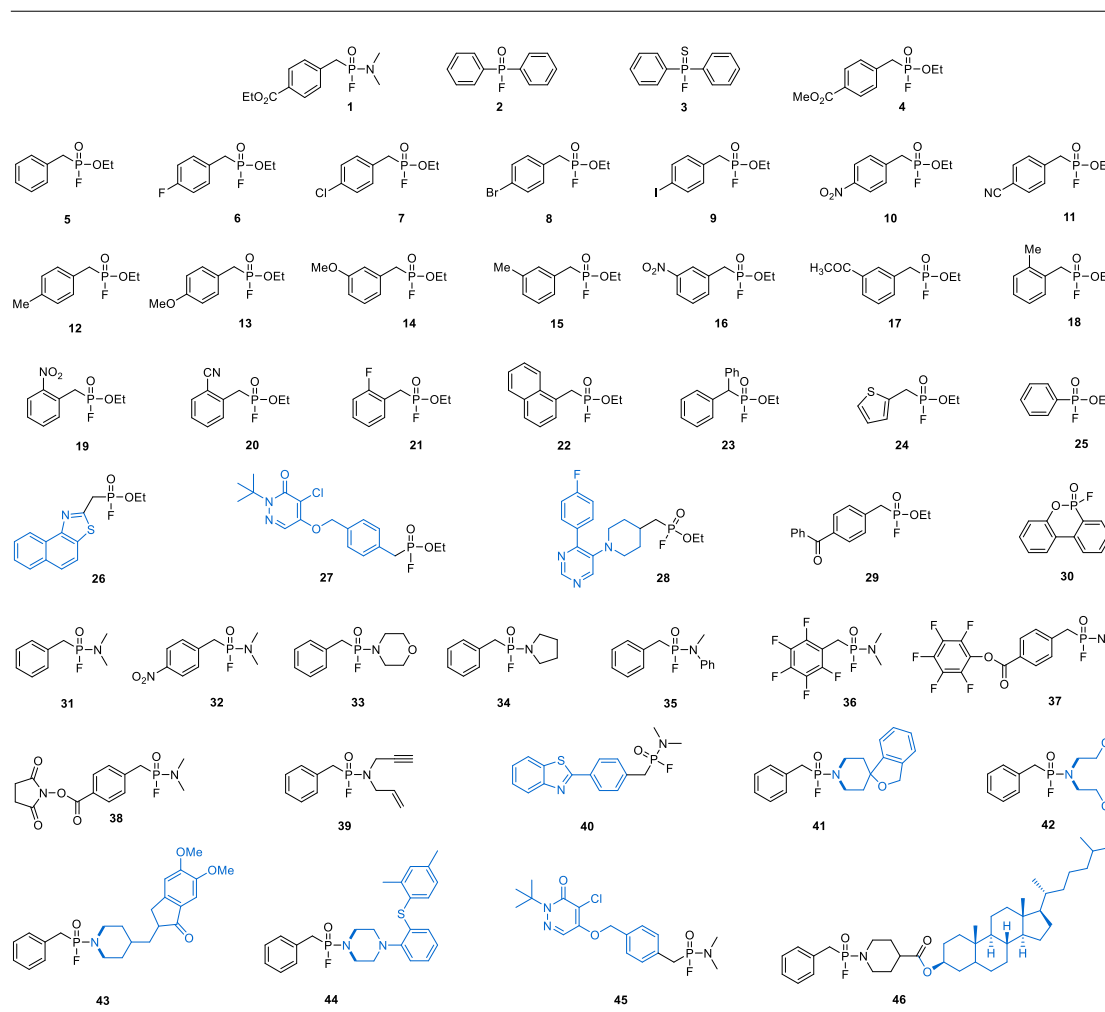
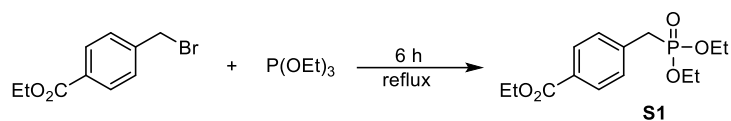


Fig. S2: An Overview of fluorophosphines (1–46) was discussed in this work.

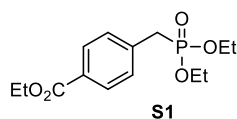
2.2.1 General synthetic procedures for substrates

Procedure A for the formation of S1



4-(Bromomethyl)benzoic acid ethyl ester (1.1 eq.) and triethyl phosphite (1.0 eq.) were initially stirred at room temperature without additional solvent and subsequently heated to 150 °C for refluxing over a period of 3–6 h¹. The progress of the reaction was carefully monitored using thin-layer chromatography (TLC) or ³¹P NMR. Once the reaction reached completion, any residual unreacted triethyl phosphite was removed through vacuum distillation. The resulting crude product was then subjected to purification by silica gel column chromatography with a mixture of petroleum ether and ethyl acetate as the eluent in a ratio of 3:1, effectively removing the solvent and yielding the purified compound S1.

Ethyl 4-((diethoxyphosphoryl)methyl)benzoate (S1)



Colorless oil, yield: 73%.

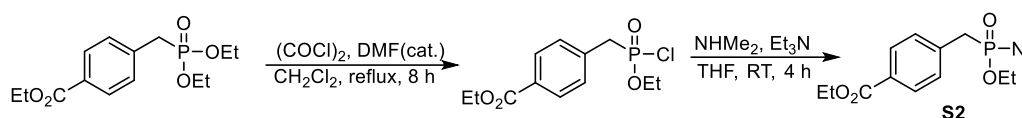
¹H NMR (400 MHz, CDCl₃): δ 7.95 (d, *J* = 7.6 Hz, 2H), 7.3 (d, *J* = 7.6 Hz, 2H), 4.33 (q, *J* = 6.8 Hz, 2H), 3.98 (t, *J* = 7.2 Hz, 4H), 3.17 (d, *J* = 22.4 Hz, 2H), 1.35 (t, *J* = 7.2 Hz, 3H), 1.21 (t, *J* = 6.8 Hz, 6H).

¹³C NMR (100 MHz, CDCl₃): δ 166.3, 136.9 (d, *J* = 9.2 Hz), 129.7, 129.6, 129.0 (d, *J* = 3.5 Hz), 62.2 (d, *J* = 6.7 Hz), 60.8, 33.9 (d, *J* = 136.8 Hz), 16.3 (d, *J* = 5.8 Hz), 14.2.

³¹P NMR (162 MHz, CDCl₃): δ 25.2 (s, 1P).

HRMS (m/z) calculated for C₁₄H₂₂O₅P⁺ ([M+H]⁺): 301.1199, found: 301.1201.

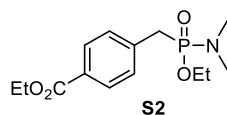
Procedure B for the formation of S2



Step 1: Under ambient conditions, a 2 M solution of oxalyl chloride (2 eq.) was gradually added dropwise to the ethyl 4-((diethoxyphosphoryl)methyl)benzoate (**S1**) (1.0 eq.). Next, 2 drops of DMF were introduced as a catalyst. The reaction mixture was then heated to 60 °C and refluxed for 8 h². The reaction progress was monitored using ³¹P NMR, as the P^v-Cl compound typically exhibits a chemical shift around 35–45 ppm. While TLC is ineffective in detecting the reaction process of unstable intermediates, ³¹P NMR enables clear product identification and quantification of conversion. Upon completion of the reaction, the solvent and excess oxalyl chloride were removed under reduced pressure. The resulting product can be utilized directly in the subsequent step without requiring further purification.

Step 2: The initial product was dissolved in tetrahydrofuran, followed by the slow addition of a mixture of NHMe₂ and Et₃N, both dissolved in THF, under ice bath conditions. Subsequently, the reaction mixture was allowed to warm up to RT and react for 2 h, with the progress of the reaction monitored by TLC. After the reaction was complete, the precipitate was filtered, and the solvent was removed under reduced pressure. The crude product was then purified using silica gel column chromatography to eliminate the solvent and obtain the purified compounds **S2**.

Ethyl 4-(((dimethylamino)(ethoxy)phosphoryl)methyl)benzoate (**S2**)



White solid, yield: 65%.

¹H NMR (400 MHz, CDCl₃): δ 7.91 (d, *J* = 8.0 Hz, 2H), 7.30 (d, *J* = 6.4 Hz, 2H), 4.30 (q, *J* = 7.2 Hz, 2H), 3.79–4.01 (m, 2H), 3.04–3.19 (m, 2H), 2.48 (t, *J* = 8 Hz, 6H), 1.32 (t, *J* = 7.2 Hz, 3H), 1.32 (t, *J* = 7.2 Hz, 3H).

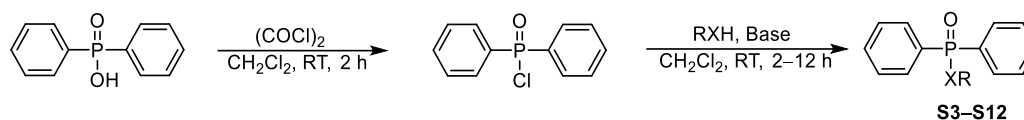
¹³C NMR (100 MHz, CDCl₃): δ 166.3, 137.9 (d, *J* = 8.1 Hz), 129.6, 129.5, 128.7 (d, *J* = 3.2 Hz), 60.7, 59.6 (d, *J* = 6.8 Hz), 36.2 (d, *J* = 3.9 Hz), 33.9 (d, *J* = 123.8 Hz), 16.1

(d, $J = 6.5$ Hz), 14.2.

^{31}P NMR (162 MHz, CDCl_3): δ 29.9 (s, 1P).

HRMS (m/z) calculated for $\text{C}_{14}\text{H}_{23}\text{NO}_4\text{P}^+$ ($[\text{M}+\text{H}]^+$): 300.1359; found: 300.1351.

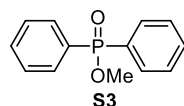
General procedure C for the formation of diphenylphosphinate (S3–S12).



Step 1: The diphenylphosphinic acid was dissolved in a small amount of dichloromethane, and then oxalyl chloride (1.5 eq. 2 M in CH_2Cl_2) was slowly added dropwise with vigorous stirring. The reaction was conducted at room temperature for 2 h³, and its progress was monitored using ^{31}P NMR. After the reaction was completed, the solvent and residual oxalyl chloride were removed by vacuum evaporation without the need for purification.

Step 2: The reaction mixture from the previous step was then dissolved in CH_2Cl_2 . A mixture of RXH and base (base = 2.0 eq. Et_3N for $\text{R} = \text{Ar}/\text{Bn}$; base = 1.2 eq. NaH for $\text{R} = \text{Alkyl}$) was gradually added dropwise under an ice bath, and the reaction was continued at room temperature for 2–12 h³. The advancement of the reaction was monitored by TLC. Upon completion of the reaction, the organic layer was extracted, and the collected organic phase was dried and concentrated. Subsequently, the crude product was purified using silica gel column chromatography with a petroleum ether: ethyl acetate eluent in a ratio of 3:2 to 1:1, to remove the solvent and obtain the purified compounds **S3–S12**.

Methyl diphenylphosphinate (S3)



Colorless oil, yield: 87%.

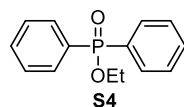
^1H NMR (400 MHz, CDCl_3): δ 3.74 (d, $J = 11.2$ Hz, 3H), 7.40–7.44 (m, 4H), 7.47–7.51 (m, 2H), 7.76–7.81 (m, 4H).

^{13}C NMR (100 MHz, CDCl_3): δ 132.2 (d, $J = 2.6$ Hz), 131.6 (d, $J = 10$ Hz), 130.3, 128.6 (d, $J = 13.1$ Hz), 51.53 (d, $J = 6$ Hz).

^{31}P NMR (162 MHz, CDCl_3): δ 33.3 (s, 1P).

HRMS (m/z) calculated for $C_{13}H_{14}O_2P^+$ ($[M+H]^+$): 233.0726; found: 233.0733.

Ethyl diphenylphosphinate (S4)



Colorless oil, yield: 64%.

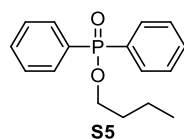
1H NMR (400 MHz, $CDCl_3$): δ 7.79 (dd, $J = 12$ Hz, 8 Hz, 4H), 7.45–7.49 (m, 2H), 7.38–7.43 (m, 4H), 4.04–4.11 (m, 2H), 1.33 (t, $J = 6.8$ Hz, 3H).

^{13}C NMR (100 MHz, $CDCl_3$): δ 131.9 (d, $J = 10.4$ Hz), 131.5 (d, $J = 40.4$ Hz), 131.4 (d, $J = 137$ Hz), 128.4 (d, $J = 52$ Hz), 61.1 (d, $J = 23.6$ Hz), 16.3 (d, $J = 26$ Hz).

^{31}P NMR (162 MHz, $CDCl_3$): δ 31.5 (s, 1P).

HRMS (m/z) calculated for $C_{14}H_{16}O_2P^+$ ($[M+H]^+$): 247.0882; found: 247.0890.

Butyl diphenylphosphinate (S5)



White solid, yield: 77%.

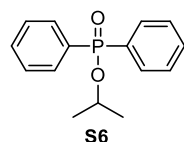
1H NMR (400 MHz, $CDCl_3$): δ 7.78–7.83 (m, 4H), 7.48–7.52 (m, 2H), 7.41–7.46 (m, 4H), 4.02 (q, $J = 6.8$ Hz, 2H), 1.67–1.74 (m, 2H), 1.38–1.48 (m, 2H), 0.91 (t, $J = 7.6$ Hz, 3H).

^{13}C NMR (100 MHz, $CDCl_3$): δ 132.0 (d, $J = 2.6$ Hz), 133.7 (d, $J = 136$ Hz), 131.6 (d, $J = 10$ Hz), 128.5 (d, $J = 13$ Hz), 64.7 (d, $J = 6$ Hz), 32.6 (d, $J = 6.5$ Hz), 18.8, 13.6.

^{31}P NMR (162 MHz, $CDCl_3$): δ 31.1 (s, 1P).

HRMS (m/z) calculated for $C_{16}H_{20}O_2P^+$ ($[M+H]^+$): 275.1195; found: 275.1208.

Isopropyl diphenylphosphinate (S6)



White solid, yield: 52%.

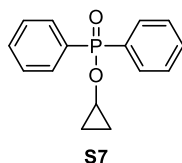
1H NMR (400 MHz, $CDCl_3$): δ 7.78–7.83 (m, 4H), 7.47–7.50 (m, 2H), 7.40–7.44 (m, 4H), 4.63–4.70 (m, 1H), 1.33 (d, $J = 6.4$ Hz, 6H).

¹³C NMR (100 MHz, CDCl₃): δ 24.3 (d, *J* = 4 Hz), 70.2 (d, *J* = 6 Hz), 128.4 (d, *J* = 13 Hz), 131.6 (d, *J* = 10 Hz), 131.9 (d, *J* = 2.6 Hz), 132.4 (d, *J* = 136 Hz).

³¹P NMR (162 MHz, CDCl₃): δ 29.7 (s, 1P).

HRMS (m/z) calculated for C₁₅H₁₈O₂P⁺ ([M+H]⁺): 261.1039; found: 261.1044.

Cyclopropyl diphenylphosphinate (S7)



Colorless oil, yield: 37%.

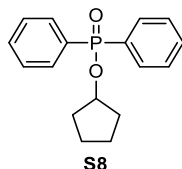
¹H NMR (400 MHz, CDCl₃): δ 7.75–7.80 (m, 4H), 7.45–7.49 (m, 2H), 7.38–7.43 (m, 4H), 3.86–3.91 (m, 1H), 0.82–0.83 (m, 2H), 0.51–0.55 (m, 2H).

¹³C NMR (100 MHz, CDCl₃): δ 132.1, 131.3 (d, *J* = 137 Hz), 131.5 (d, *J* = 10.2 Hz), 128.4 (d, *J* = 13.1 Hz), 49.8 (d, *J* = 6 Hz), 5.7 (d, *J* = 4.1 Hz).

³¹P NMR (162 MHz, CDCl₃): δ 32.4 (s, 1P).

HRMS (m/z) calculated for C₁₅H₁₆O₂P⁺ ([M+H]⁺): 259.0882; found: 259.0886.

Cyclopentyl diphenylphosphinate (S8)



White solid, yield: 75%.

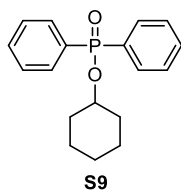
¹H NMR (400 MHz, CDCl₃): δ 7.76–7.81 (m, 4H), 7.45–7.50 (m, 2H), 7.41–7.43 (m, 4H), 4.88 (m, 1H), 1.79–1.89 (m, 6H), 1.56 (m, 2H).

¹³C NMR (100 MHz, CDCl₃): δ 132.4 (d, *J* = 137 Hz), 131.8 (d, *J* = 2.4 Hz), 131.6 (d, *J* = 10 Hz), 128.4 (d, *J* = 13 Hz), 78.7 (d, *J* = 6.3 Hz), 34.28 (d, *J* = 4.1 Hz), 23.0.

³¹P NMR (162 MHz, CDCl₃): δ 29.9 (s, 1P).

HRMS (m/z) calculated for C₁₇H₂₀O₂P⁺ ([M+H]⁺): 287.1195; found: 287.1201.

Cyclohexyl diphenylphosphinate (S9)



White solid, yield: 69%.

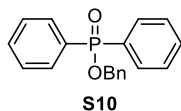
¹H NMR (400 MHz, CDCl₃): δ 7.78–7.83 (m, 4H), 7.48–7.50 (m, 2H), 7.42–7.44 (m, 4H), 4.40–4.42 (m, 1H), 1.87–1.88 (m, 2H), 1.70–1.72 (m, 2H), 1.59–1.61 (m, 2H), 1.37–1.47 (m, 1H), 1.26–1.36 (m, 3H).

¹³C NMR (100 MHz, CDCl₃): δ 132.6 (d, *J* = 136 Hz), 131.8 (d, *J* = 2.4 Hz), 131.5 (d, *J* = 10 Hz), 128.6 (d, *J* = 13 Hz), 74.9 (d, *J* = 6.1 Hz), 33.9 (d, *J* = 3.5 Hz), 25.1, 23.5.

³¹P NMR (162 MHz, CDCl₃): δ 29.7 (s, 1P).

HRMS (m/z) calculated for C₁₈H₂₂O₂P⁺ ([M+H]⁺): 301.1352; found: 301.1359.

Benzyl diphenylphosphinate (S10)



White solid, yield: 81%.

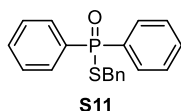
¹H NMR (400 MHz, CDCl₃): δ 7.81–7.86 (m, 4H), 7.51–7.53 (m, 2H), 7.41–7.46 (m, 4H), 7.29–7.37 (m, 5H), 5.07 (d, *J* = 6.8 Hz, 2H).

¹³C NMR (100 MHz, CDCl₃): δ 136.3 (d, *J* = 7.4 Hz), 132.1 (d, *J* = 2.6 Hz), 131.6 (d, *J* = 10.2 Hz), 131.9, 130.6, 128.5 (d, *J* = 7.9 Hz), 128.1 (d, *J* = 63.9 Hz), 128.2, 66.2 (d, *J* = 5.5 Hz).

³¹P NMR (162 MHz, CDCl₃): δ 32.3 (s, 1P).

HRMS (m/z) calculated for C₁₉H₁₈O₂P⁺ ([M+H]⁺): 309.1039; found: 309.1042.

S-benzyl diphenylphosphinothioate (S11)



White solid, yield: 78%.

¹H NMR (400 MHz, CDCl₃): δ 7.84–7.89 (m, 4H), 7.45–7.52 (m, 6H), 7.39–7.45 (m, 4H), 7.20–7.23 (m, 5H), 4.02 (d, *J* = 2.3 Hz, 2H).

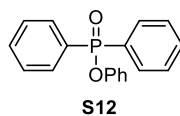
¹³C NMR (100 MHz, CDCl₃): δ 136.8 (d, *J* = 5.3 Hz), 133.1 (d, *J* = 106 Hz), 132.3 (d, *J* = 2.8 Hz), 131.5 (d, *J* = 10.4 Hz), 128.9, 128.7, 128.53 (d, *J* = 2.4 Hz), 127.4, 33.1 (d,

$J = 1.7$ Hz).

^{31}P NMR (162 MHz, CDCl_3): δ 42.7 (s, 1P).

HRMS (m/z) calculated for $\text{C}_{19}\text{H}_{18}\text{OPS}^+$ ($[\text{M}+\text{H}]^+$): 325.0810; found: 325.0816.

Phenyl diphenylphosphinate (S12)



White solid, yield: 65%.

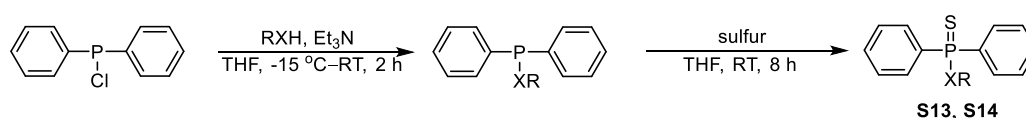
^1H NMR (400 MHz, CDCl_3): δ 7.11 (t, $J = 3.4$ Hz, 1H), 7.24–7.31 (m, 4H), 7.48–7.52 (m, 4H), 7.55–7.59 (m, 2H), 7.91–7.97 (m, 4H).

^{13}C NMR (100 MHz, CDCl_3): δ 150.9 (d, $J = 8.3$ Hz), 132.4 (d, $J = 2.7$ Hz), 131.8 (d, $J = 10.3$ Hz), 130.9 (d, $J = 137$ Hz), 129.6, 128.5 (d, $J = 13.4$ Hz), 124.5, 120.7 (d, $J = 4.7$ Hz).

^{31}P NMR (162 MHz, CDCl_3): δ 30.4 (s, 1P).

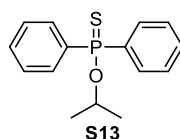
HRMS (m/z) calculated for $\text{C}_{18}\text{H}_{16}\text{O}_2\text{P}^+$ ($[\text{M}+\text{H}]^+$): 295.0882; found: 295.0894.

General procedure D for the formation of diphenylphosphinothioate (S13, S14)



Chlorodiphenylphosphane (1.1 equiv.) was dissolved in THF, and the temperature was lowered to $-15\text{ }^{\circ}\text{C}$. Then, a mixture of 1.0 eq. RXH (propane-2-alcohol/phenyl methylmercaptan) and 1.2 eq. Et_3N was slowly added dropwise. The reaction mixture was then allowed to warm up to room temperature (RT) and react for 2 h⁴. After the completion of the reaction, sulfur (5.0 eq.) was added to the reaction system, and the reaction was continued at RT for 8 h. The progress of the reaction was monitored by TLC. Upon completion of the reaction, the reaction mixture was extracted, and the organic phase was collected, dried, and concentrated under reduced pressure. Subsequently, the crude product was purified using silica gel column chromatography to remove the solvent and obtain the purified compounds **S13**, **S14**.

O-Isopropyl diphenylphosphinothioate (**S13**)



White solid, yield: 43%.

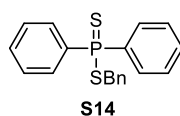
¹H NMR (400 MHz, CDCl₃): δ 7.86–7.92 (m, 4H), 7.38–7.44 (m, 6H), 4.86–4.95 (m, 1H), 1.30 (d, $J = 6.4$ Hz, 6H).

¹³C NMR (100 MHz, CDCl₃): δ 133.1, 131.9 (d, $J = 2.6$ Hz), 131.6 (d, $J = 10$ Hz), 128.4 (d, $J = 13$ Hz), 70.3 (d, $J = 6$ Hz), 24.3 (d, $J = 4$ Hz).

³¹P NMR (162 MHz, CDCl₃): δ 42.1 (s, 1P).

HRMS (m/z) calculated for $\text{C}_{15}\text{H}_{18}\text{OPS}^+$ ($[\text{M}+\text{H}]^+$): 277.0810; found: 277.0816.

Benzyl diphenylphosphinodithioate (**S14**)



White solid, yield: 21%.

¹H NMR (400 MHz, CDCl₃): δ 7.96–7.81 (m, 4H), 7.51–7.54 (m, 6H), 7.30–7.32 (m,

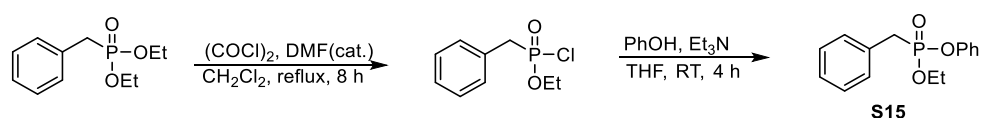
2H), 7.19–7.27 (m, 3H), 4.22 (d, $J = 3$ Hz, 2H).

^{13}C NMR (100 MHz, CDCl_3): δ 136.7 (d, $J = 5.7$ Hz), 134.1 (d, $J = 84$ Hz), 133.8 (d, $J = 3$ Hz), 131.5 (d, $J = 11.3$ Hz), 129.3, 128.6, 128.5 (d, $J = 4.1$ Hz), 127.44, 35.9 (d, $J = 1.4$ Hz).

^{31}P NMR (162 MHz, CDCl_3): δ 64.1 (s, 1P).

HRMS (m/z) calculated for $\text{C}_{19}\text{H}_{18}\text{PS}_2^+$ ($[\text{M}+\text{H}]^+$): 341.0582; found: 341.0588.

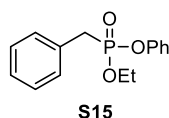
General procedure E for the formation of S15



Step 1: The synthesis method employs **Step 1 of Procedure B**.

Step 2: The initial product was dissolved in tetrahydrofuran, followed by the slow addition of a mixture of PhOH and Et_3N , both dissolved in THF, under ice bath conditions. Subsequently, the reaction mixture was allowed to warm up to RT and react for 2 h, with the progress of the reaction monitored by TLC. After the reaction was complete, the precipitate was filtered, and the solvent was removed under reduced pressure. The crude product was then purified using silica gel column chromatography to eliminate the solvent and obtain the purified compounds **S15**.

Phenyl benzylphosphonate (S15)



Colorless oil, yield: 54%.

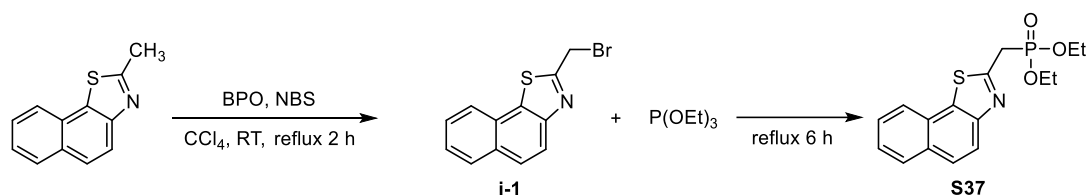
^1H NMR (400 MHz, CDCl_3): δ 7.33–7.21 (m, 7H), 7.15–7.06 (m, 3H), 4.13–3.98 (m, 2H), 3.31 (dd, $J = 21.7, 2.6$ Hz, 2H), 1.19 (t, $J = 7.1$ Hz, 3H).

^{13}C NMR (100 MHz, CDCl_3): δ 150.5 (d, $J = 8.8$ Hz), 130.9 (d, $J = 9.4$ Hz), 129.9 (d, $J = 6.8$ Hz), 129.6, 128.6 (d, $J = 3.2$ Hz), 127.0 (d, $J = 3.7$ Hz), 124.8, 120.4 (d, $J = 4.3$ Hz), 63.0 (d, $J = 7.2$ Hz), 33.7 (d, $J = 138.9$ Hz), 16.2 (d, $J = 5.9$ Hz).

^{31}P NMR (162 MHz, CDCl_3): δ 23.3 (s, 1P).

HRMS (m/z) calculated for $\text{C}_{15}\text{H}_{18}\text{O}_3\text{P}^+$ ($[\text{M}+\text{H}]^+$): 277.0988; found: 277.0982.

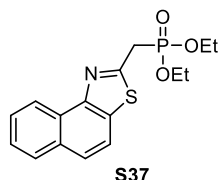
Procedure F for the formation of S37



Step 1: 2-methylnaphtho[2,1-*d*]thiazole (4 mmol, 1.0 eq.) and *N*-bromosuccinimide (NBS, 1.0 eq.) were introduced into the reaction flask, succeeded by the addition of the dibenzoyl peroxide (BPO, 0.05 eq.) in 50 mL CCl₄, while maintaining constant stirring. Elevate the mixture's temperature to 80 °C and maintain reflux for a duration of 2 h⁵. The reaction's progression was monitored using TLC. After the completion of the reaction, allow it to cool to RT, filter the mixture, and remove CCl₄ under reduced pressure. For column chromatography purification of the residue, a mixture of petroleum ether and ethyl acetate (3:2–1:1) was employed as the elution, resulting in the isolation of the purified bromine compound **i-1**.

Step 2: The synthesis method employs **Procedure A**.

Ethyl (naphtho[1,2-*d*]thiazol-2-ylmethyl)phosphonofluoridate (S37)



Colorless oil, yield: 74%.

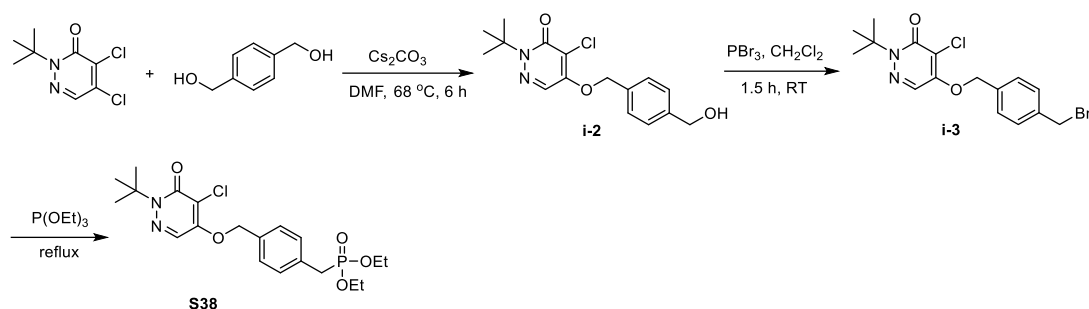
¹H NMR (600 MHz, CDCl₃): δ 8.76 (d, *J* = 8.3 Hz, 1H), 7.92 (d, *J* = 8.1 Hz, 1H), 7.84 (d, *J* = 7.4 Hz, 1H), 7.78 (d, *J* = 8.8 Hz, 1H), 7.65 (t, *J* = 7.6 Hz, 1H), 7.56 (t, *J* = 7.2 Hz, 1H), 4.22–4.13 (m, 4H), 3.84 (d, *J* = 20.0 Hz, 2H), 1.31 (t, *J* = 7.2 Hz, 6H).

¹³C NMR (151 MHz, CDCl₃): δ 159.8 (d, *J* = 9.8 Hz), 149.2, 132.6, 131.9, 128.5, 128.1, 127.0, 126.1, 125.9, 123.8, 118.8, 63.0 (d, *J* = 6.6 Hz), 33.5, 16.4 (d, *J* = 5.7 Hz).

³¹P NMR (243 MHz, CDCl₃): δ 21.2 (s, 1P).

HRMS (m/z) calculated for C₁₆H₁₈NNaO₃PS⁺ ([M+Na]⁺): 358.0637; found: 358.0630.

Procedure G for the formation of S38

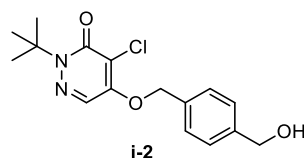


Step 1: 2-(*tert*-Butyl)-4,5-dichloropyridazin-3(2H)-one (4.53 mmol) was dissolved in 15 mL dry DMF. 1,4-phenylene dimethanol (3.2 g, 23.16 mmol) and Cs₂CO₃ (6.0 g, 18.41 mmol) were added, and the mixture was stirred at 68 °C for 6 h, then cooled. The product was extracted with EtOAc, dried under vacuum, and purified by column chromatography to yield compound **i-2** as a white solid⁶.

Step 2: Compound **i-2** (0.91 g, alcoholic) was dissolved in 15 ml of dry dichloromethane. PBr₃ (0.14 ml) was then introduced dropwise into the solution. The reaction was performed at room temperature for approximately 1.5 h. The crude product was isolated by extraction with water (30 mL) and dried under vacuum. The resulting white solid product **i-3** was obtained in a quantitative yield, ready for the subsequent step without further purification.

Step 3: The synthesis of compound **S38** was completed according to the synthesis **Procedure A** of this study.

2-(*Tert*-butyl)-4-chloro-5-((4-(hydroxymethyl)benzyl)oxy)pyridazin-3(2H)-one (**i-2**)



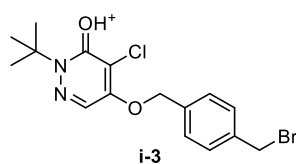
White solid, yield: 76%.

¹H NMR (400 MHz, CDCl₃): δ 7.71 (s, 1H), 7.40 (d, *J* = 1.6 Hz, 4H), 5.30 (s, 2H), 4.71 (s, 2H), 1.61 (s, 9H).

¹³C NMR (100 MHz, CDCl₃): δ 159.0, 153.7, 141.7, 134.0, 127.4, 127.3, 125.1, 118.2, 71.6, 66.4, 64.7, 27.8.

HRMS (*m/z*) calculated for C₁₆H₂₀ClN₂O₃⁺ ([M+H]⁺): 323.1157; found: 323.1151.

5-((4-(Bromomethyl)benzyl)oxy)-2-(tert-butyl)-4-chloropyridazin-3(2H)-one (i-3)



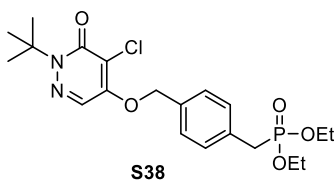
White solid, yield: 96%.

¹H NMR (400 MHz, CDCl₃): δ 7.71 (s, 1H), 7.43 (d, *J* = 7.6 Hz, 2H), 7.39 (d, *J* = 7.6 Hz, 2H), 5.30 (s, 2H), 4.49 (s, 2H), 1.62 (s, 9H).

¹³C NMR (100 MHz, CDCl₃): δ 158.9, 153.5, 138.4, 135.0, 129.6, 127.4, 124.9, 118.2, 71.3, 66.4, 32.7, 27.8.

HRMS (m/z) calculated for C₁₆H₁₉BrClN₂O₂⁺ ([M+H]⁺): 385.0313; found: 385.0318.

Diethyl 4-(((1-(tert-butyl)-5-chloro-6-oxo-1,6-dihydropyridazin-4-yl)oxy)methyl)benzyl)phosphonate (S38)



White solid, yield: 69%.

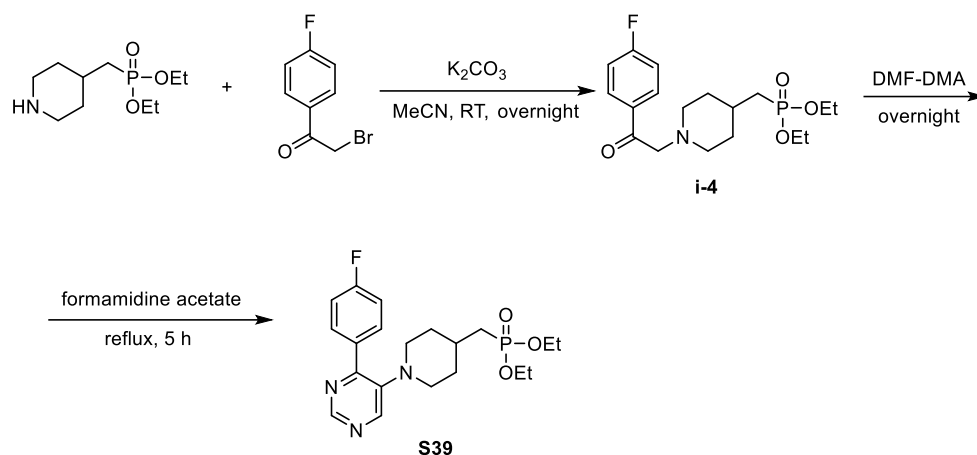
¹H NMR (400 MHz, CDCl₃): δ 7.69 (s, 1H), 7.29–7.32 (s, 4H), 5.26 (d, *J* = 1.6 Hz, 2H), 3.96–3.99 (m, 4H), 3.12 (d, *J* = 21.6 Hz, 2H), 1.58 (s, 9H), 1.19, (t, *J* = 7.2 Hz, 6H).

¹³C NMR (100 MHz, CDCl₃): δ 158.9, 153.5, 133.4 (d, *J* = 3.9 Hz), 132.3 (d, *J* = 9.0 Hz), 130.2 (d, *J* = 6.5 Hz), 127.2 (d, *J* = 2.9 Hz), 125.0, 118.1, 71.4, 66.3, 62.1 (d, *J* = 6.7 Hz), 33.4 (d, *J* = 137.5 Hz), 27.7, 16.2 (d, *J* = 5.9 Hz).

³¹P NMR (162 MHz, DMSO-*d*₆): δ 25.9 (1P).

HRMS (m/z) calculated for C₂₀H₂₉ClN₂O₅P⁺ ([M+H]⁺): 443.1497; found: 443.1492.

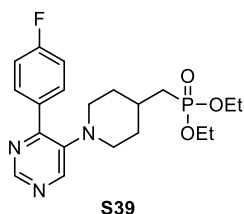
Procedure H for the formation of S39



To a solution of diethyl (piperidin-4-ylmethyl)phosphonate (5.8 g, 25.0 mmol) and K_2CO_3 (4.33 g, 31.3 mmol) in acetonitrile (50 mL), 2-bromo-1-(4-fluorophenyl)ethanone (4.91 g, 22.7 mmol) dissolved in acetonitrile (30 mL) was added dropwise at room temperature. The reaction mixture was stirred overnight at the same temperature. Following this, the mixture was concentrated under reduced pressure, diluted with water, and extracted with ethyl acetate. The organic phase was washed with brine, dried over sodium sulfate, filtered, and then concentrated in vacuo. The resulting pale yellow solid, **i-4** (7.42 g, 88%), was used directly in the subsequent reaction without additional purification.

A solution of **i-4** (3.71 g, 10 mmol) and DMF-DMA (18.2 mL, 130 mmol) was heated to reflux overnight. After cooling, the mixture was concentrated under reduced pressure. The residue was then dissolved in *n*-butanol (10 mL) and diisopropylethylamine (DIPEA, 10 mL). Formamidine acetate (3.6 g, 35 mmol) was added, and the resulting mixture was stirred at 100 °C overnight. The reaction mixture was then diluted with water and extracted with ethyl acetate. The organic layer was washed with water and brine, dried over magnesium sulfate, filtered, and concentrated under reduced pressure. The crude product was purified by column chromatography on silica gel to afford **S39** (1.75 g, 43%) as a dark brown oil.

Diethyl ((1-(4-(4-fluorophenyl)pyrimidin-5-yl)piperidin-4-yl)methyl)phosphonate (S39)



Dark brown oil, yield: 43%.

¹H NMR (400 MHz, CDCl₃): δ 8.88 (s, 1H), 8.40 (s, 1H), 8.06–8.19 (m, 2H), 7.13–7.19 (m, 2H), 4.06–4.13 (m, 4H), 3.18 (d, *J* = 10.1 Hz, 2H), 2.62–2.68 (m, 2H), 2.04–2.07 (m, 2H), 1.82–1.90 (m, 3H), 1.69–1.78 (m, 2H), 1.32 (t, *J* = 7.2 Hz, 6H).

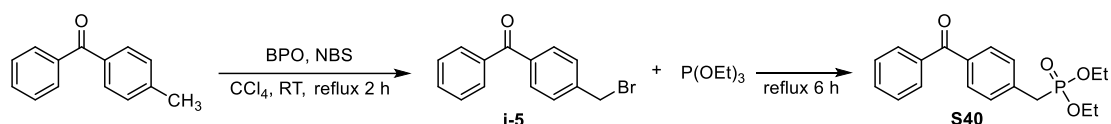
¹³C NMR (100 MHz, CDCl₃): δ 164.3, 162.7, 156.5, 152.4, 147.1, 144.0, 134.0, 130.2 (d, *J* = 8.3 Hz), 115.4 (d, *J* = 21.2 Hz), 61.5 (d, *J* = 6.6 Hz), 50.7, 33.1 (d, *J* = 11.0 Hz), 32.7, 31.8, 30.3 (d, *J* = 4.3 Hz), 16.4 (d, *J* = 5.7 Hz).

³¹P NMR (162 MHz, CDCl₃): δ 30.5 (s, 1P).

¹⁹F NMR (376 MHz, CDCl₃): δ -110.5 (s, 1F).

HRMS (m/z) calculated for C₂₀H₂₇FN₃NaO₃P⁺ ([M+Na]⁺): 430.1666; found: 430.1660.

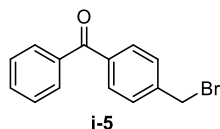
Procedure I for the formation of S40



Step 1: Phenyl(*p*-tolyl)methanone (4 mmol, 1.0 eq.) and *N*-bromosuccinimide (NBS, 1.0 eq.) were introduced into the reaction flask, succeeded by the addition of the dibenzoyl peroxide (BPO, 0.05 eq.) in 50 mL CCl₄, while maintaining constant stirring. Elevate the mixture's temperature to 80 °C and maintain reflux for a duration of 2 h⁷. The reaction's progression was monitored using TLC. After the completion of the reaction, allow it to cool to RT, filter the mixture, and remove CCl₄ under reduced pressure. For column chromatography purification of the residue, a mixture of petroleum ether and ethyl acetate (3:2–1:1) was employed as the elution, resulting in the isolation of the purified compound **i-5**.

Step 2: The synthesis method employs **Procedure A**.

Diethyl (4-benzoylbenzyl)phosphonate (**i-5**)



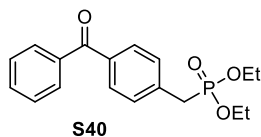
White solid, yield: 93%.

¹H NMR (400 MHz, CDCl₃): δ 7.77–7.80 (m, 4H), 7.60 (t, *J* = 7.6 Hz, 1H), 7.47–7.51 (m, 4H), 4.53 (s, 2H).

¹³C NMR (100 MHz, CDCl₃): δ 195.9, 142.1, 137.4, 132.5, 130.5, 129.9, 128.9, 128.3, 32.2.

HRMS (*m/z*) calculated for C₁₄H₁₂BrO⁺ ([*M*+*H*]⁺): 275.0066; found: 275.0061.

Diethyl (4-benzoylbenzyl)phosphonate (**S40**)



White solid, yield: 77%.

¹H NMR (400 MHz, CDCl₃): δ 7.77 (t, *J* = 6.0 Hz, 4H), 7.58 (t, *J* = 6.8 Hz, 1H), 7.47

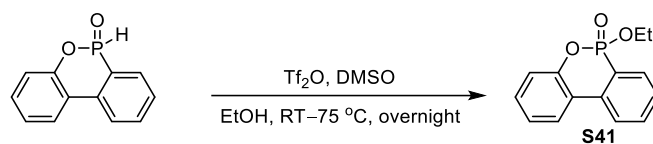
(t, $J = 6.8$ Hz, 2H), 7.41 (d, $J = 8.0$ Hz, 2H), 4.01–4.09 (m, 4H), 3.22 (d, $J = 22.4$ Hz, 2H), 1.26 (t, $J = 7.2$ Hz, 6H).

^{13}C NMR (100 MHz, CDCl_3): δ 196.2, 137.6, 136.7 (d, $J = 9.1$ Hz), 136.1 (d, $J = 3.4$ Hz), 132.3, 130.3 (d, $J = 2.8$ Hz), 129.9, 129.7 (d, $J = 6.3$ Hz), 128.3, 62.3 (d, $J = 6.7$ Hz), 33.9 (d, $J = 137$ Hz), 16.4 (d, $J = 5.8$ Hz).

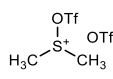
^{31}P NMR (162 MHz, CDCl_3): δ 25.2 (s, 1P).

HRMS (m/z) calculated for $\text{C}_{18}\text{H}_{22}\text{O}_4\text{P}^+$ ($[\text{M}+\text{H}]^+$): 333.1250; found: 333.1256.

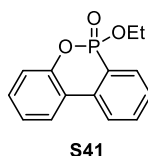
Procedure J for the formation of S41



The mixture of the dibenzo[*c,e*][1,2]oxaphosphinine 6-oxide (0.2 mmol, 40.4 mg, 1.0 eq.), Tf₂O (0.6 mmol, 169.3 mg, 3.0 eq.), and DMSO (0.4 mmol, 31.2 mg, 2.0 eq.) was dissolved in 1.0 mL of EtOH under a N₂ atmosphere. (Tf₂O and DMSO form an *in situ*

 active intermediate to initiate the reaction.) Subsequently, the reaction mixture was heated in a round-bottom flask in a 75 °C oil bath overnight⁸. The progress of the reaction was monitored using TLC. After the completion of the reaction, water was added to quench it, and then the organic phase was extracted and dried under reduced pressure. Finally, the crude product was purified using silica gel column chromatography with petroleum ether/ethyl acetate (4:1–3:1) as the eluent to obtain the purified compound S41.

6-Ethoxydibenzo[*c,e*][1,2]oxaphosphinine 6-oxide (S41)



Colorless oil, yield: 77%.

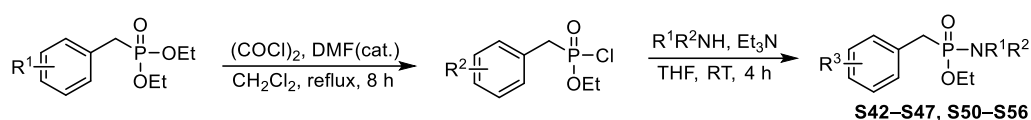
¹H NMR (400 MHz, CDCl₃): δ 7.94–8.02 (m, 3H), 7.72 (t, *J* = 7.6 Hz, 1H), 7.51–7.55 (m, 1H), 7.40 (t, *J* = 15.2 Hz, 1H), 7.24–7.30 (m, 2H), 4.19–4.27 (m, 2H), 1.29 (t, *J* = 7.2 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃): δ 149.8 (d, *J* = 7.7 Hz), 136.9 (d, *J* = 7 Hz), 133.4 (d, *J* = 2.2 Hz), 130.4, 130.0 (d, *J* = 9.1 Hz), 128.2 (d, *J* = 15.3 Hz), 125.2, 124.6, 123.9 (d, *J* = 12 Hz), 123.3 (d, *J* = 181 Hz), 122.5 (d, *J* = 12 Hz), 120.1, (d, *J* = 6.6 Hz), 62.9 (d, *J* = 6.6 Hz), 16.2 (d, *J* = 5.7 Hz).

³¹P NMR (162 MHz, CDCl₃): δ 10.2 (1P).

HRMS (m/z) calculated for C₁₄H₁₄O₃P⁺ ([M+H]⁺): 261.0675; found: 261.0673.

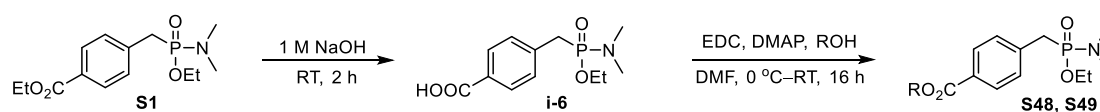
General procedure K for the formation of S42–S47, S50–S56,



Step 1: Under ambient conditions, a 2 M solution of oxalyl chloride (2.0 eq.) was gradually added dropwise to the desired benzyl phosphonate diethyl compound (1.0 eq.). Next, 2 drops of DMF were introduced as a catalyst. The reaction mixture was then heated to 60 °C and refluxed for 8 h⁹. The reaction progress was monitored using ³¹P-NMR (the P–Cl compound typically exhibits a chemical shift around 35–45 ppm). Upon completion of the reaction, the solvent and excess oxalyl chloride were removed under reduced pressure. The resulting product can be utilized directly in the subsequent step without requiring further purification.

Step 2: The initial product was dissolved in tetrahydrofuran, followed by the slow addition of a mixture of R¹R²NH and Et₃N, both dissolved in THF, under ice bath conditions. Subsequently, the reaction mixture was allowed to warm up to RT and react for 2 h, with the progress of the reaction monitored by TLC. After the reaction was complete, the precipitate was filtered, and the solvent was removed under reduced pressure. The crude product was then purified using silica gel column chromatography to eliminate the solvent and obtain the purified compounds S42–S47, S50–S56.

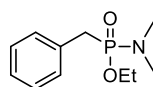
General procedure L for the formation of S48, S49



Step 1: Compound ethyl 4-(((ethoxyphosphoryl)oxy)methyl)benzoate **S1** (1.0 eq.) was dissolved in a 1 M NaOH aqueous solution (3.0 eq.) and stirred at RT for 2 h. The progress of the reaction was monitored by TLC until the reaction mixture became clear. After the reaction was complete, the mixture was extracted once with CH₂Cl₂, and the aqueous phase was acidified with 1 M HCl. Subsequently, CH₂Cl₂ was added for three extractions, and the organic layer was collected, dried, and concentrated to obtain the carboxylated compound **i-6** resulting from the hydrolysis, which was directly used in the next reaction without purification.

Step 2: The carboxylated compound **i-6** (1.0 eq.) was dissolved in CH₂Cl₂, and 1-(3-dimethylaminopropyl)-3-ethylcarbodiimide hydrochloride (EDCI, 1.5 eq.) and 4-dimethylaminopyridine (DMAP, 2.0 eq.) were added at 0 °C. Subsequently, the hydroxyl-containing compound (*N*-hydroxysuccinimide or pentafluorophenol) dissolved in CH₂Cl₂ was slowly added dropwise to the reaction mixture. After the addition was complete, the reaction was allowed to proceed at room temperature for 16 h¹⁰, and the progress of the reaction was monitored by TLC. Upon completion of the reaction, water was added to quench the reaction, and then the mixture was extracted with CH₂Cl₂. The organic layer was collected, dried, and concentrated to remove the organic solvent. Finally, the crude product was purified using silica gel column chromatography (petroleum ether: ethyl acetate = 1:1–2:3) to obtain the purified compounds **S48** and **S49**.

Ethyl *P*-benzyl-*N,N*-dimethylphosphonamidate (S42**)**



S42

Yellowish solid, yield: 79%.

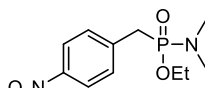
¹H NMR (400 MHz, CDCl₃): δ 7.27–7.31 (m, 5H), 3.85–4.07 (m, 2H), 3.04–3.19 (m, 2H), 2.52 (d, *J* = 8.8 Hz, 6H), 1.28 (t, *J* = 6.8 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃): δ 132.2 (d, *J* = 8.0 Hz), 129.5 (d, *J* = 6.2 Hz), 128.2 (d, *J* = 2.7 Hz), 126.3 (d, *J* = 3.3 Hz), 59.5 (d, *J* = 7.0 Hz), 36.2 (d, *J* = 3.9 Hz), 33.6 (d, *J* = 124.9 Hz), 16.0 (d, *J* = 6.5 Hz).

³¹P NMR (162 MHz, CDCl₃): δ 31.3 (1P).

HRMS (m/z) calculated for C₁₁H₁₉NO₂P⁺ ([M+H]⁺): 228.1148; found: 228.1141.

Ethyl *N,N*-dimethyl-*P*-(4-nitrobenzyl)phosphonamidate (S43**)**



S43

Yellowish solid, yield: 87%.

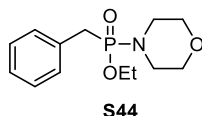
¹H NMR (400 MHz, CDCl₃): δ 1.23 (t, *J* = 7.2 Hz, 6H), 2.55 (d, *J* = 8.8 Hz, 2H), 3.82–4.02 (m, 2H), 7.39–7.42 (m, 2H), 8.10 (d, *J* = 8.4 Hz, 2H).

¹³C NMR (100 MHz, CDCl₃): δ 16.0 (d, *J* = 6.5 Hz), 33.7 (d, *J* = 123.7 Hz), 36.2 (d, *J* = 4 Hz), 123.5 (d, *J* = 2.7 Hz), 59.9 (d, *J* = 6.9 Hz), 130.4 (d, *J* = 6.0 Hz), 140.6 (d, *J* = 8.2 Hz), 146.7 (d, *J* = 3.6 Hz).

³¹P NMR (162 MHz, CDCl₃): δ 29.0 (1P).

HRMS (m/z) calculated for C₁₁H₁₈N₂O₄P⁺ ([M+H]⁺): 273.0999; found: 273.1006.

Ethyl benzyl(morpholino)phosphinate (S44)



White solid, yield: 72%.

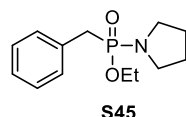
¹H NMR (400 MHz, CDCl₃): δ 1.34 (t, *J* = 14 Hz, 3H), 2.85–2.96 (m, 4H), 3.08–3.27 (m, 2H), 3.48–3.55 (m, 4H), 4.12–4.18 (m, 2H), 7.26–7.37 (m, 5H).

¹³C NMR (100 MHz, CDCl₃): δ 131.9 (d, *J* = 7.7 Hz), 129.8 (d, *J* = 6.4 Hz), 128.5 (d, *J* = 2.4 Hz), 126.8 (d, *J* = 3.8 Hz), 67.1 (d, *J* = 4.8 Hz), 59.9 (d, *J* = 7.3 Hz), 44.2, 34.1 (d, *J* = 126 Hz), 16.3 (d, *J* = 6.7 Hz).

³¹P NMR (162 MHz, CDCl₃): δ 28.5 (1P).

HRMS (m/z) calculated for C₁₃H₂₁NO₃P⁺ ([M+H]⁺): 270.1254; found: 270.1250.

Ethyl benzyl(pyrrolidin-1-yl)phosphinate (S45)



Colorless oil, yield: 70%.

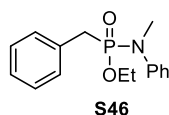
¹H NMR (400 MHz, CDCl₃): δ 1.29 (t, *J* = 6.8 Hz, 3H), 1.69–1.75 (m, 4H), 3.07–3.09 (m, 2H), 2.95–2.97 (m, 2H), 3.15 (dd, *J* = 28 Hz, 8.8 Hz, 2H), 3.88–4.09 (m, 2H), 7.21–7.24 (m, 1H), 7.29 (d, *J* = 4.4 Hz, 4H).

¹³C NMR (100 MHz, CDCl₃): δ 16.3 (d, *J* = 6.5 Hz), 26.2 (d, *J* = 7.7 Hz), 33.7 (d, *J* = 125 Hz), 46.5 (d, *J* = 4.2 Hz), 59.7 (d, *J* = 6.8 Hz), 126.5 (d, *J* = 3.2 Hz), 128.3 (d, *J* = 2.5 Hz), 129.6 (d, *J* = 6.2 Hz), 132.5 (d, *J* = 8 Hz).

³¹P NMR (162 MHz, CDCl₃): δ 28.5 (1P).

HRMS (m/z) calculated for C₁₃H₂₁NO₂P⁺ ([M+H]⁺): 254.1304; found: 254.1308.

Ethyl *P*-benzyl-*N*-methyl-*N*-phenylphosphonamidate (S46)



Yellow oil, yield: 26%.

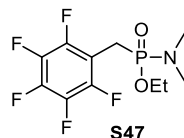
¹H NMR (400 MHz, CDCl₃): δ 7.21–7.30 (m, 5H), 7.11–7.16 (m, 4H), 7.05–7.08 (m, 1H), 3.94–4.19 (m, 2H), 3.24 (d, *J* = 20 Hz, 2H), 3.0 (d, *J* = 7.6 Hz, 3H), 1.29 (t, *J* = 7.2 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃): δ 144.3 (d, *J* = 4.6 Hz), 131.5 (d, *J* = 8.2 Hz), 129.8 (d, *J* = 6.4 Hz), 128.9, 128.3 (d, *J* = 2.9 Hz), 126.7 (d, *J* = 3.5 Hz), 123.4, 121.8 (d, *J* = 3.4 Hz), 60.4 (d, *J* = 7.0 Hz), 36.2 (d, *J* = 3.8 Hz), 33.7 (d, *J* = 125.6 Hz), 16.1 (d, *J* = 6.7 Hz).

³¹P NMR (162 MHz, CDCl₃): δ 28.4 (1P).

HRMS (m/z) calculated for C₁₆H₂₁NO₂P⁺ ([M+H]⁺): 290.1304; found: 290.1309.

Ethyl *N,N*-dimethyl-*P*-((perfluorophenyl)methyl)phosphonamidate (S47)



Colorless oil, yield: 36%.

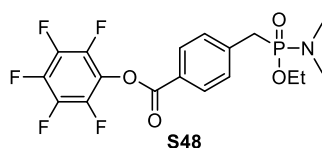
¹H NMR (400 MHz, CDCl₃): δ 1.29 (t, *J* = 6.8 Hz, 3H), 2.64 (d, *J* = 9.2 Hz, 6H), 3.10–3.17 (m, 2H), 3.88–4.09 (m, 2H).

¹³C NMR (100 MHz, CDCl₃): δ 145.1 (d, *J* = 245.7 Hz), 143.8 (m), 141.2 (m), 138.7 (m), 136.2 (m), 107.5 (m), 60.2 (d, *J* = 6.7 Hz), 35.9 (d, *J* = 4.2 Hz), 20.7 (d, *J* = 128.6 Hz), 15.9 (d, *J* = 6.7 Hz).

³¹P NMR (162 MHz, CDCl₃): δ 26.1 (1P). **¹⁹F NMR (376 MHz, CDCl₃):** δ -141.8 (2F), -156.3 (1F), -162.3 (2F).

HRMS (m/z) calculated for C₁₁H₁₄F₅NO₂P⁺ ([M+H]⁺): 318.0677; found: 318.0671.

Perfluorophenyl 4-(((dimethylamino)(ethoxy)phosphoryl)methyl)benzoate (S48)



White solid, yield: 43%.

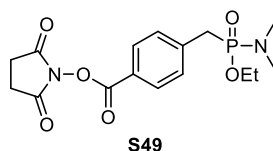
¹H NMR (400 MHz, CDCl₃): δ 8.13 (d, *J* = 7.6 Hz, 2H), 7.47 (d, *J* = 8.0 Hz, 2H), 3.88–4.09 (m, 2H), 3.12–3.29 (m, 2H), 2.60 (d, *J* = 8.8 Hz, 6H), 1.30 (t, *J* = 7.2 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃): δ 165.6, 162.4, 140.5 (d, *J* = 8.1 Hz), 139.2, 138.2, 136.7, 130.8 (d, *J* = 2.5 Hz), 130.3 (d, *J* = 6 Hz), 125.2 (d, *J* = 3.4 Hz), 59.9 (d, *J* = 6.9 Hz), 36.4 (d, *J* = 4 Hz), 34.1 (d, *J* = 123.6 Hz), 16.2 (d, *J* = 6.6 Hz).

³¹P NMR (162 MHz, CDCl₃): δ 24.1 (1P).

HRMS (m/z) calculated for C₁₈H₁₈F₅NO₄P⁺ ([M+H]⁺): 438.0888; found: 438.0893.

2,5-Dioxopyrrolidin-1-yl 4-(((dimethylamino)(ethoxy)phosphoryl)methyl)benzoate (S49)



White solid, yield: 55%.

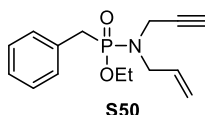
¹H NMR (400 MHz, CDCl₃): δ 1.14 (t, *J* = 7.2 Hz, 3H), 2.43 (d, *J* = 8.8 Hz, 6H) 2.75 (s, 4H), 3.07 (dd, *J* = 22 Hz, 8.8 Hz, 2H), 3.73–3.93 (m, 2H), 7.31 (d, *J* = 7.2 Hz, 2H), 7.92 (d, *J* = 8.0 Hz, 2H).

¹³C NMR (100 MHz, CDCl₃): δ 169.1, 161.4, 140.5 (d, *J* = 8.1 Hz), 130.2 (d, *J* = 2.4 Hz), 129.9 (d, *J* = 5.9 Hz), 122.9 (d, *J* = 3.1 Hz), 59.5 (d, *J* = 6.8 Hz), 35.9 (d, *J* = 3.9 Hz), 33.8 (d, *J* = 123.3 Hz), 25.3, 15.8 (d, *J* = 6.5 Hz).

³¹P NMR (162 MHz, CDCl₃): δ 29.3 (1P).

HRMS (m/z) calculated for C₁₆H₂₂N₂O₆P⁺ ([M+H]⁺): 369.1210; found: 369.1213.

Ethyl *N*-allyl-*P*-benzyl-*N*-(prop-2-yn-1-yl)phosphonamidate (S50)



White solid, yield: 56%.

¹H NMR (400 MHz, CDCl₃): δ 7.27–7.32 (m, 4H), 7.21–7.23 (m, 1H), 5.45–5.55 (m, 1H), 5.16 (dd, *J* = 22.8 Hz, 17.2 Hz, 2H), 3.89–4.10 (m, 2H), 3.51–3.80 (m, 4H), 3.08–3.23 (m, 2H), 1.27 (t, *J* = 7.2 Hz, 3H), 2.20 (s, 1H).

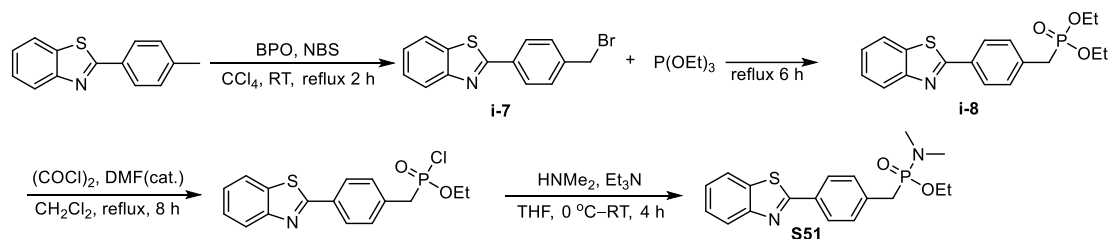
¹³C NMR (100 MHz, CDCl₃): δ 133.5 (d, *J* = 3.1 Hz), 132.0 (d, *J* = 8.7 Hz), 129.9 (d, *J* = 6.3 Hz), 128.4 (d, *J* = 2.8 Hz), 126.6 (d, *J* = 3.4 Hz), 118.5, 79.7, 71.9, 60.14 (d, *J*

= 7 Hz), 47.5 (d, $J = 3.6$ Hz), 34.8 (d, $J = 126$ Hz), 34.0, (d, $J = 5.3$ Hz), 16.1 (d, $J = 6.9$ Hz).

^{31}P NMR (162 MHz, CDCl_3): δ 30.3 (1P).

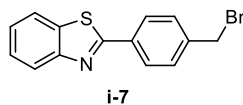
HRMS (m/z) calculated for $\text{C}_{15}\text{H}_{21}\text{NO}_2\text{P}^+$ ($[\text{M}+\text{H}]^+$): 278.1304; found: 278.1307.

Procedure M for the formation of S51



The synthesis methods for preparing **i-7** and **i-8** in **Steps 1** and **2**, respectively, utilize **Procedure A**. The synthesis method for preparing compound **S51** in **Steps 3** and **4** employs **Procedure E**.

2-(4-(Bromomethyl)phenyl)benzo[d]thiazole (i-7)



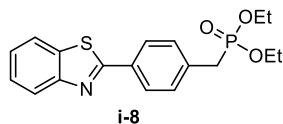
White solid, yield: 97%.

^1H NMR (400 MHz, CDCl_3): δ 8.07–8.09 (m, 3H), 7.91 (d, $J = 8.0$ Hz, 1H), 7.52 (d, $J = 8.4$ Hz, 3H), 7.40 (t, $J = 7.2$ Hz, 1H), 4.54 (s, 2H).

^{13}C NMR (100 MHz, CDCl_3): δ 167.2, 154.0, 140.6, 135.0, 133.6, 129.7, 127.9, 126.4, 125.4, 123.3, 121.6, 32.5.

HRMS (m/z) calculated for $\text{C}_{14}\text{H}_{11}\text{BrNS}^+$ ($[\text{M}+\text{H}]^+$): 303.9790; found: 303.9793.

Diethyl (benzo[d]thiazol-2-ylmethyl)phosphonate (i-8)



White solid, yield: 77%.

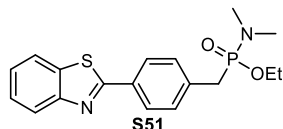
^1H NMR (400 MHz, CDCl_3): δ 8.05 (t, $J = 7.6$ Hz, 3H), 7.90 (d, $J = 8.0$ Hz, 1H), 7.49 (t, $J = 7.2$ Hz, 1H), 7.40–7.44 (m, 2H), 7.38 (t, $J = 7.6$ Hz, 1H), 4.00–4.08 (m, 4H), 3.22 (d, $J = 22.0$ Hz, 2H), 1.26 (t, $J = 6.8$ Hz, 6H).

¹³C NMR (100 MHz, CDCl₃): δ 167.7, 154.0, 134.9 (d, *J* = 2.4 Hz), 134.8, 132.3 (d, *J* = 3.5 Hz), 130.4 (d, *J* = 6.5 Hz), 127.7 (d, *J* = 3.0 Hz), 126.3, 125.2, 123.1, 121.6, 62.3 (d, *J* = 6.7 Hz), 33.8 (d, *J* = 137.0 Hz), 16.4 (d, *J* = 6.0 Hz).

³¹P NMR (162 MHz, CDCl₃): δ 25.5 (1P).

HRMS (m/z) calculated for C₁₈H₂₁NO₃PS⁺ ([M+H]⁺): 362.0974; found: 362.0970.

Ethyl *P*-benzyl-*N,N*-bis(2-chloroethyl)phosphoramidate (S51)



White solid, yield: 64%.

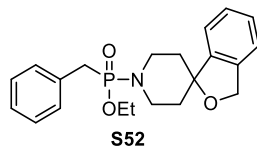
¹H NMR (400 MHz, CDCl₃): δ 8.02–8.07 (m, 3H), 7.88 (t, *J* = 8.0 Hz, 1H), 7.47 (t, *J* = 7.2 Hz, 1H), 7.35–7.42 (m, 3H), 3.87–4.09 (m, 2H), 3.11–3.25 (m, 2H), 2.57 (d, *J* = 8.8 Hz, 6H), 1.29 (t, *J* = 6.9 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃): δ 167.8, 153.9, 136.0 (d, *J* = 8.2 Hz), 134.9, 131.9 (d, *J* = 3.5 Hz), 130.3 (d, *J* = 6.0 Hz), 127.6 (d, *J* = 2.4 Hz), 126.3, 125.1, 123.0, 121.6, 59.8 (d, *J* = 6.8 Hz), 36.4 (d, *J* = 3.7 Hz), 33.9 (d, *J* = 124.3 Hz), 16.2 (d, *J* = 6.1 Hz).

³¹P NMR (162 MHz, CDCl₃): δ 30.2 (1P).

HRMS (m/z) calculated for C₁₈H₂₂N₂O₂PS⁺ ([M+H]⁺): 361.1134; found: 361.1139.

Ethyl benzyl(3*H*-spiro[isobenzofuran-1,4'-piperidin]-1'-yl)phosphinate (S52)



White solid, yield: 78%.

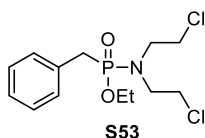
¹H NMR (400 MHz, CDCl₃): δ 7.30–7.36 (m, 4H), 7.22–7.25 (m, 3H), 7.15–7.18 (m, 1H), 6.93–6.95 (m, 1H), 5.0 (s, 2H), 3.92–4.12 (m, 2H), 3.37–3.43 (m, 2H), 2.93–3.09 (m, 4H), 2.51 (d, *J* = 8.8 Hz, 1H), 1.56–1.64 (m, 3H), 1.31 (t, *J* = 7.2 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃): δ 145.2, 138.7, 132.5 (d, *J* = 7.7 Hz), 129.8, 128.5, 127.7, 127.3, 126.7, 120.8 (d, *J* = 51.3 Hz), 84.5, 70.7, 59.7 (d, *J* = 6.8 Hz), 40.8 (d, *J* = 20.9 Hz), 36.5 (d, *J* = 8.9 Hz), 34.2 (d, *J* = 126.5 Hz), 16.3 (d, *J* = 6.5 Hz).

³¹P NMR (162 MHz, CDCl₃): δ 29.7 (1P).

HRMS (m/z) calculated for C₂₁H₂₇NO₃P⁺ ([M+H]⁺): 372.1723; found: 372.1728.

Ethyl *P*-benzyl-*N,N*-bis(2-chloroethyl)phosphonamidate (S53)



White solid, yield: 54%.

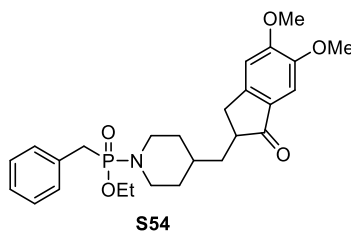
¹H NMR (400 MHz, CDCl₃): δ 7.25–7.33 (m, 5H), 3.91–4.15 (m, 2H), 3.40 (d, *J* = 6.8 Hz, 4H), 3.08–3.32 (m, 6H), 1.28 (t, *J* = 6.8 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃): 16.2 (d, *J* = 6.6 Hz), 34.3 (d, *J* = 125.4 Hz), 42.17, 48.7 (d, *J* = 4 Hz), 60.5 (d, *J* = 6.8 Hz), 126.9 (d, *J* = 3.4 Hz), 128.6 (d, *J* = 2.7 Hz), 129.7 (d, *J* = 6.4 Hz), δ 131.7 (d, *J* = 8 Hz).

³¹P NMR (162 MHz, CDCl₃): δ 31.0 (1P).

HRMS (m/z) calculated for C₁₃H₂₁Cl₂NO₂P⁺ ([M+H]⁺): 324.0681; found: 324.0688.

Ethyl benzyl(4-((5,6-dimethoxy-1-oxo-2,3-dihydro-1*H*-inden-2-yl)methyl)piperidin-1-yl)phosphinate (S54)



White solid, yield: 69%.

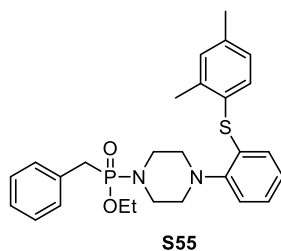
¹H NMR (400 MHz, CDCl₃): δ 7.27–7.33 (m, 5H), 7.18 (s, 1H), 6.87 (s, 1H), 3.98 (s, 3H), 3.92 (s, 3H), 3.48–3.51 (m, 2H), 3.20–3.26 (m, 2H), 3.05–3.15 (m, 1H), 2.67 (d, *J* = 14.4 Hz, 2H), 2.59–2.61 (m, 1H), 2.38–2.48 (m, 2H), 1.84–1.87 (m, 1H), 1.59–1.70 (m, 3H), 1.28–1.34 (m, 5H), 0.96–1.12 (m, 2H).

¹³C NMR (100 MHz, CDCl₃): δ 207.4, 155.5, 149.4, 148.6, 132.4 (d, *J* = 7.8 Hz), 129.8, (d, *J* = 6.1 Hz), 129.1, 128.3, 126.5, 107.3, 104.4, 59.6 (d, *J* = 6.8 Hz), 56.1 (d, *J* = 12.3 Hz), 44.9 (d, *J* = 5.4 Hz), 44.3 (d, *J* = 10.4 Hz), 38.8 (d, *J* = 10.5 Hz), 34.3, (d, *J* = 126 Hz), 34.4, (d, *J* = 7.8 Hz), 33.2, (d, *J* = 7.1 Hz), 32.1, 16.2 (d, *J* = 6.4 Hz).

³¹P NMR (162 MHz, CDCl₃): δ 29.2 (1P).

HRMS (m/z) calculated for C₂₆H₃₅NO₅P⁺ ([M+H]⁺): 472.2247; found: 472.2245.

Ethyl benzyl(4-(2-((2,4-dimethylphenyl)thio)phenyl)piperazin-1-yl)phosphinate (S55)



White solid, yield: 51%.

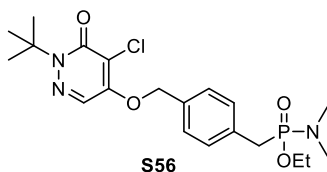
¹H NMR (400 MHz, CDCl₃): δ 7.29–7.39 (m, 6H), 7.18 (d, *J* = 7.2 Hz, 1H), 7.04–7.128 (m, 2H), 6.98 (d, *J* = 7.6 Hz, 1H), 6.87–6.92 (m, 1H), 6.54 (t, *J* = 7.2 Hz, 1H), 3.99–4.21 (m, 2H), 3.11–3.26 (m, 6H), 2.88–2.93 (m, 4H), 2.36 (dd, *J*₁ = 24.4 Hz, *J*₂ = 7.6 Hz, 6H), 1.34–1.39 (m, 3H).

¹³C NMR (100 MHz, CDCl₃): δ 149.0, 142.3, 139.2, 136.1, 134.6, 131.3 (d, *J* = 8.1 Hz), 131.6, 129.9 (d, *J* = 6.0 Hz), 128.5, 127.8, 126.7, 126.2, 125.4, 124.5, 119.9, 59.8 (d, *J* = 6.6 Hz), 53.4, 52.1, 44.5, 34.3 (d, *J* = 125.8 Hz), 20.8 (d, *J* = 61.1 Hz), 16.3 (d, *J* = 6.3 Hz).

³¹P NMR (162 MHz, CDCl₃): δ 29.9 (1P).

HRMS (m/z) calculated for C₂₇H₃₄N₂O₂PS⁺ ([M+H]⁺): 481.2073; found: 481.2073.

Ethyl *P*-(4-(((1-(tert-butyl)-5-chloro-6-oxo-1,6-dihydropyridazin-4-yl)oxy)methyl)benzyl)-*N,N*-dimethylphosphonamidate (S56)



White solid, yield: 49%.

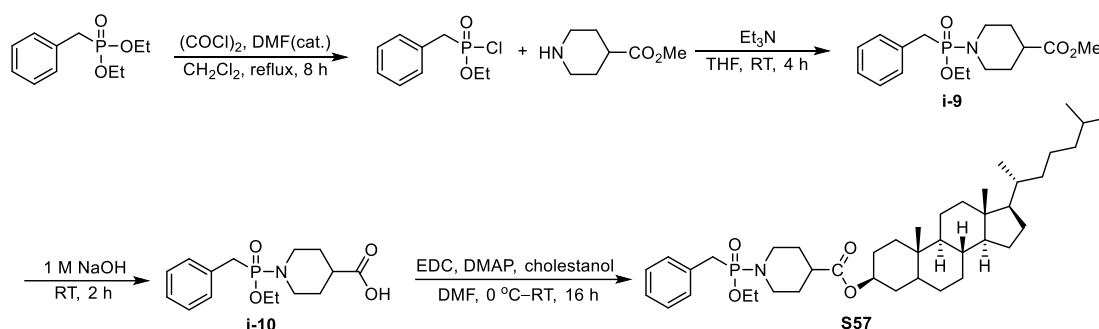
¹H NMR (400 MHz, CDCl₃): δ 7.70 (s, 1H), 7.31–7.36 (m, 4H), 5.29 (s, 2H), 3.84–4.03 (m, 2H), 3.08–3.17 (m, 2H), 2.55 (d, *J* = 8.8 Hz, 6H), 1.62 (s, 9H), 1.28, (t, *J* = 7.2 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃): δ 159.0, 153.6, 133.3 (d, *J* = 8.1 Hz), 133.2 (d, *J* = 3.7 Hz), 130.3 (d, *J* = 6.1 Hz), 127.3 (d, *J* = 2.8 Hz), 125.1, 118.3, 71.6, 66.4, 59.7 (d, *J* = 7.0 Hz), 36.4 (d, *J* = 3.9 Hz), 33.4 (d, *J* = 124.9 Hz), 27.8, 16.2 (d, *J* = 6.6 Hz).

³¹P NMR (162 MHz, DMSO-*d*₆): δ 30.8 (1P).

HRMS (m/z) calculated for C₂₀H₃₀ClN₃O₄P⁺ ([M+H]⁺): 442.1657; found: 442.1655.

Procedure N for the formation of S57

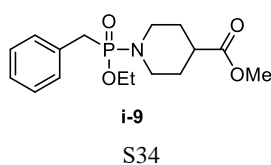


The synthetic methods for preparing **i-9** in **steps 1** and **2** are consistent with the aforementioned "General **procedure B** for the formation of **S2**."

Step 3: Compound **i-9** (1.0 eq.), which is 1-(benzyloxycarbonyl) piperidine-4-carboxylic acid methyl ester, was dissolved in a 1 M NaOH aqueous solution (3.0 eq.) and stirred at RT for 2 h. The progress of the reaction was monitored using TLC until the reaction mixture became clear. Upon completion of the reaction, the mixture was subjected to a single extraction with dichloromethane, and the resulting aqueous phase was acidified using 1 M HCl. Then, CH_2Cl_2 was added for three extractions, and the organic layer was collected, dried, and concentrated to obtain the carboxylated compound **i-10** through hydrolysis.

Step 4: Compound **i-10** (1.0 eq.) carboxylate was dissolved in DMF, and 1-(3-dimethylaminopropyl)-3-ethylcarbodiimide hydrochloride (EDCI 1.5 eq.) and DMAP (2.0 eq.) were added at 0°C . Then, the hydroxyl-containing compound cholestanol was slowly added to the reaction mixture dissolved in DMF. After the addition was complete, the reaction was allowed to proceed at room temperature for 16 h, and the progress of the reaction was monitored by TLC. Upon completion of the reaction, the organic solvent was removed under reduced pressure, and the residue was purified using silica gel column chromatography with a mixture of petroleum ether and ethyl acetate (1:1–2:3) as the eluent to remove the solvent and obtain the purified compound **S57**.

Methyl 1-(benzyl(ethoxy)phosphoryl)piperidine-4-carboxylate (**i-9**)



White solid, yield: 86%.

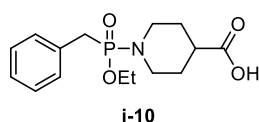
¹H NMR (400 MHz, CDCl₃): δ 1.30 (t, *J* = 7.2 Hz, 3H), 1.39–1.54 (m, 2H), 1.78 (t, *J* = 11.6 Hz, 2H), 2.31–2.65 (m, 3H), 3.03–3.23 (m, 2H), 3.37–3.47 (m, 2H), 3.67 (s, 3H), 4.05–4.11 (m, 2H), 7.24–7.30 (m, 5H).

¹³C NMR (100 MHz, CDCl₃): δ 174.8, 132.2 (d, *J* = 7.9 Hz), 129.7 (d, *J* = 6.4 Hz), 128.3 (d, *J* = 2.4 Hz), 126.6 (d, *J* = 3.2 Hz), 59.7 (d, *J* = 6.9 Hz), 51.6, 43.5, 40.9, 34.2 (d, *J* = 125.6 Hz), 28.4 (dd, *J* = 8.3 Hz, 3.9 Hz), 16.2 (d, *J* = 6.5 Hz).

³¹P NMR (162 MHz, CDCl₃): δ 29.1 (1P).

HRMS (m/z) calculated for C₁₆H₂₅NO₄P⁺ ([M+H]⁺): 326.1516; found: 326.1511.

1-(Benzyl(ethoxy)phosphoryl)piperidine-4-carboxylic acid (i-10)



White solid, yield: 82%.

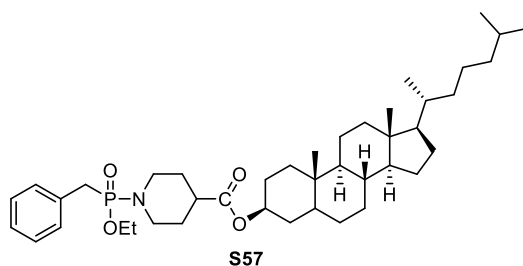
¹H NMR (400 MHz, MeOD): δ 7.22–7.27 (m, 5H), 6.04 (s, 1H), 3.88–4.11 (m, 2H), 3.34–3.45 (m, 2H), 3.04–3.23 (m, 2H), 2.29–2.64 (m, 3H), 1.79 (t, *J* = 11.2 Hz, 2H), 1.40–1.52 (m, 2H), 1.28 (t, *J* = 7.2 Hz, 3H).

¹³C NMR (100 MHz, MeOD): δ 177.8, 131.9 (d, *J* = 8.1 Hz), 129.8 (d, *J* = 6.4 Hz), 128.4 (d, *J* = 2.6 Hz), 126.7 (d, *J* = 3.2 Hz), 60.2 (d, *J* = 7.0 Hz), 43.6, 40.8, 34.3 (d, *J* = 125.9 Hz), 28.3 (dd, *J* = 10.4 Hz, 3.7 Hz), 16.2 (d, *J* = 6.5 Hz).

³¹P NMR (162 MHz, MeOD): δ 29.7 (1P).

HRMS (m/z) calculated for C₁₅H₂₃NO₄P⁺ ([M+H]⁺): 312.1359; found: 312.1361.

(3*S*,8*R*,9*S*,10*S*,13*R*,14*S*,17*R*)-10,13-Dimethyl-17-((*R*)-6-methylheptan-2-yl)hexadecahydro-1*H*-cyclopenta[*a*]phenanthren-3-yl 1-(benzyl(ethoxy)phosphoryl)piperidine-4-carboxylate (S57)



White solid, yield: 40%.

¹H NMR (400 MHz, DMSO-*d*₆): δ 7.27–7.29 (m, 4H), 7.22–7.23 (m, 1H), 4.04–4.09

(m, 1H), 3.34–3.50 (m, 2H), 3.01–3.21 (m, 1H), 2.24–2.64 (m, 2H), 1.91–2.04 (m, 2H), 1.61–1.83 (m, 8H), 1.46–1.53 (m, 6H), 1.21–1.42 (m, 15H), 0.97–1.04 (m, 11H), 0.90 (d, $J = 6.4$ Hz, 3H), 0.86 (d, $J = 6.4$ Hz, 6H), 0.81 (s, 3H), 0.64 (s, 3H).

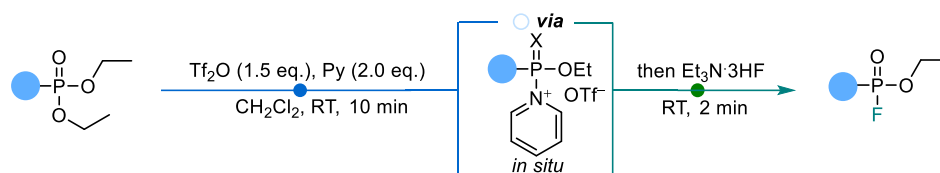
^{13}C NMR (100 MHz, $\text{DMSO-}d_6$): δ 174.1, 132.3 (d, $J = 7.8$ Hz), 129.8 (d, $J = 6.3$ Hz), 128.4 (d, $J = 2.7$ Hz), 126.7 (d, $J = 3.3$ Hz), 73.7, 59.7 (d, $J = 6.9$ Hz), 56.3 (d, $J = 13.6$ Hz), 54.2, 48.9, 44.6, 43.6, 42.6, 41.3, 39.9, 39.5, 36.7, 36.1, 35.8, 35.4 (d, $J = 2.1$ Hz), 34.3 (d, $J = 125.3$ Hz), 33.9, 31.9, 28.6, 28.5, 28.4 (d, $J = 3.5$ Hz), 28.2, 27.9, 27.4, 25.6, 24.9, 24.2, 23.8, 22.6 (d, $J = 25.1$ Hz), 21.2, 18.6, 16.3 (d, $J = 6.6$ Hz), 12.1 (d, $J = 16.1$ Hz).

^{31}P NMR (162 MHz, $\text{DMSO-}d_6$): δ 29.0 (1P).

HRMS (m/z) calculated for $\text{C}_{42}\text{H}_{69}\text{NO}_4\text{P}^+$ ($[\text{M}+\text{H}]^+$): 682.4959; found: 682.4953.

2.2.2 Representative procedure for the fluorination of alkyl phosphonates

General procedure for the synthesis of ethyl phosphorofluoridate



The diethyl phosphonate compound (0.2 mmol) was dissolved in 1 mL of CH_2Cl_2 at RT, and then Tf_2O (0.3 mmol, 50.5 μL , 1.5 eq.) was added to the reaction mixture for 5 min, followed by Py (0.4 mmol, 32 μL , 2.0 eq.) for an additional 5 min. Following this, the fluorinating reagent $\text{Et}_3\text{N}\cdot 3\text{HF}$ (0.5 eq.) was introduced into the system. The progress of the reaction was monitored using TLC or ^{31}P NMR, and after 2 min, the reaction was completed. The reaction mixture, with a volume of approximately 1 mL, underwent direct purification through silica gel column chromatography to afford the desired products **4–30**.

General procedure for the synthesis of fluorophosphoramidate

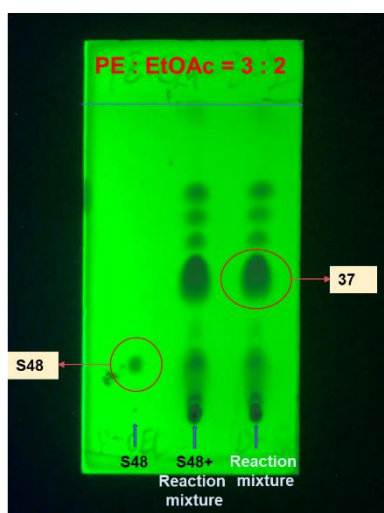
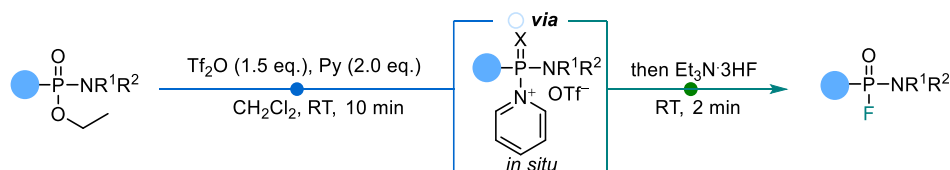
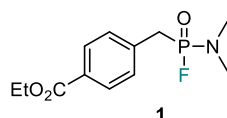


Fig. S3: Thin Layer Chromatography (TLC) analysis of the synthesis of fluorophosphoramidate (**37**) from phosphoramidate ethyl ester (**S43**).

The phosphonic acid monoethyl ester compound (0.2 mmol) was dissolved in 1 mL of CH_2Cl_2 at RT. Subsequently, Tf_2O (0.3 mmol, 50.5 μL , 1.5 eq.) was added dropwise,

and the reaction proceeded for 5 min. Following this, Py (0.4 mmol, 32 μ L, 2.0 eq.) was slowly introduced to the reaction mixture, and the reaction was allowed to continue for an additional 5 min. Then, the fluorinating reagent Et₃N·3HF (0.5 eq.) was incorporated into the system. The progress of the reaction was monitored using TLC or ³¹P NMR, and after 2 min, the reaction reached completion. The reaction mixture, with an approximate volume of 1 mL, underwent direct purification through silica gel column chromatography to eliminate the solvent and obtain the purified compounds **1**, **31–46**.

Ethyl 4-(((dimethylamino)fluorophosphoryl)methyl)benzoate (**1**)



White solid, yield: 92%.

¹H NMR (400 MHz, CDCl₃): δ 7.97 (d, J = 7.6 Hz, 2H), 7.34 (d, J = 2.0 Hz, 2H), 7.32 (d, J = 2.4 Hz, 2H), 4.31–4.36 (m, 2H), 3.23–3.38 (m, 2H), 2.60 (dd, J = 2.4 Hz, 10.0 Hz), 1.35 (t, J = 7.2 Hz, 3H).

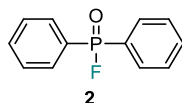
¹³C NMR (100 MHz, CDCl₃): δ 166.1, 135.7 (dd, J = 2.4 Hz, 8.6 Hz), 129.9 (d, J = 2.9 Hz), 129.5, 129.4, 60.9, 35.8, 32.7 (dd, J = 32.9 Hz, 129.0 Hz), 14.2.

³¹P NMR (162 MHz, CDCl₃): δ 32.64 (d, J = 1050.9 Hz, 1P).

¹⁹F NMR (376 MHz, CDCl₃): δ -66.37 (d, J = 1049.8 Hz, 1F).

HRMS (m/z) calculated for C₁₂H₁₈NFO₃P⁺ ([M+H]⁺): 274.1003, found 274.1007.

Diphenylphosphinic fluoride (**2**)



Colorless oil, yield: 68%.

¹H NMR (400 MHz, CDCl₃): δ 7.26–7.58 (m, 6H), 7.78–7.83 (m, 4H).

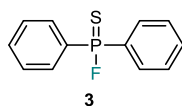
¹³C NMR (100 MHz, CDCl₃): δ 133.2 (d, J = 7 Hz), 131.3 (d, J = 11.2 Hz), 128.7 (d, J = 14 Hz), 128.6 (dd, J = 140.4 Hz, 22.1 Hz).

³¹P NMR (162 MHz, CDCl₃): δ 40.8 (d, J = 1020 Hz, 1P).

¹⁹F NMR (376 MHz, CDCl₃): δ -75.1 (d, J = 1020 Hz, 1F).

HRMS (m/z) calculated for C₁₂H₁₀FN₀OP⁺ ([M+Na]⁺): 243.0346; found: 243.0340.

Diphenylphosphinothioic fluoride (3)



Colorless oil, yield: 83%.

$^1\text{H NMR}$ (400 MHz, CDCl_3): δ 7.84–7.89 (m, 4H), 7.56–7.59 (m, 2H), 7.47–7.52 (m, 4H).

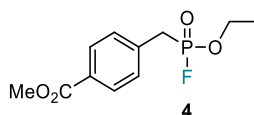
$^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 132.8 (d, $J = 2.6$ Hz), 131.0 (d, $J = 12.3$ Hz), 128.7 (d, $J = 14.1$ Hz), 53.4.

$^{31}\text{P NMR}$ (162 MHz, CDCl_3): δ 102.2 (d, $J = 1022$ Hz, 1P).

$^{19}\text{F NMR}$ (376 MHz, CDCl_3): δ -82.6 (d, $J = 1022$ Hz, 1F).

HRMS (m/z) calculated for $\text{C}_{12}\text{H}_{10}\text{FNaPS}^+$ ($[\text{M}+\text{Na}]^+$): 259.0117; found: 259.0113.

Methyl 4-((ethoxyfluorophosphoryl)methyl)benzoate (4)



Colorless oil, yield: 42%.

$^1\text{H NMR}$ (400 MHz, CDCl_3): δ 8.02 (d, $J = 8.0$ Hz, 2H), 7.38 (d, $J = 6.8$ Hz, 2H), 4.17–4.24 (m, 2H), 3.92 (s, 3H), 3.36 (d, $J = 23.2$ Hz, 2H), 1.29 (t, $J = 7.2$ Hz, 3H).

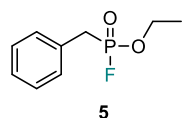
$^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 166.6, 134.7 (d, $J = 10.6$ Hz), 130.1 (d, $J = 2.7$ Hz), 129.8, 129.7, 64.2 (d, $J = 7.4$ Hz), 52.1 (d, $J = 20.5$ Hz), 32.4 (dd, $J = 141.7$ Hz, 25 Hz), 16.2 (d, $J = 5.4$ Hz).

$^{31}\text{P NMR}$ (162 MHz, CDCl_3): δ 23.7 (d, $J = 1078$ Hz, 1P).

$^{19}\text{F NMR}$ (376 MHz, CDCl_3): δ -65.4 (d, $J = 1077$ Hz, 1F).

HRMS (m/z) calculated for $\text{C}_{11}\text{H}_{15}\text{FO}_4\text{P}^+$ ($[\text{M}+\text{H}]^+$): 261.0687; found: 261.0683.

Ethyl benzylphosphonofluoridate (5)



Colorless oil, yield: 73%.

$^1\text{H NMR}$ (400 MHz, CDCl_3): δ 1.28 (t, $J = 7.2$ Hz, 3H), 3.30 (d, $J = 22.4$ Hz, 2H), 4.15–4.23 (m, 2H), 7.29–7.36 (m, 5H).

$^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 129.7 (d, $J = 7$ Hz), 129.4 (d, $J = 9.8$ Hz), 128.8 (d, J

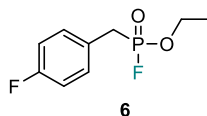
= 2.9 Hz), 127.5 (d, $J = 3.6$ Hz), 63.9 (d, $J = 7.3$ Hz), 32.2 (d, $J = 141.7$ Hz, 24.6 Hz), 16.2 (d, $J = 5.5$ Hz).

^{31}P NMR (162 MHz, CDCl_3): δ 25.1 (d, $J = 1077$ Hz, 1P).

^{19}F NMR (376 MHz, CDCl_3): δ -64.3 (d, $J = 1076$ Hz, 1F).

HRMS (m/z) calculated for $\text{C}_9\text{H}_{13}\text{FO}_2\text{P}^+$ ($[\text{M}+\text{H}]^+$): 203.0632; found: 203.0638.

Ethyl (4-fluorobenzyl)phosphonofluoridate (6)



Colorless oil, yield: 91%.

^1H NMR (400 MHz, CDCl_3): δ 1.30 (t, $J = 7.2$ Hz, 3H), 3.28 (d, $J = 22.4$ Hz, 2H), 4.17–4.24 (m, 2H), 7.03 (t, $J = 8.4$ Hz, 2H), 7.26–7.27 (m, 2H).

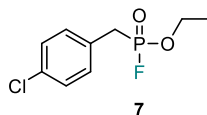
^{13}C NMR (100 MHz, CDCl_3): δ 162.3 (dd, $J = 245$ Hz, 4 Hz), 131.2 (t, $J = 7.7$ Hz), 125.1 (d, $J = 35.1$ Hz), 115.8 (dd, $J = 21.6$ Hz, 2.9 Hz), 63.9 (d, $J = 7.3$ Hz), 31.3 (dd, $J = 142.7$ Hz, 24.8 Hz), 16.2 (d, $J = 5.2$ Hz).

^{19}F NMR (376 MHz, CDCl_3): δ -64.5 (d, $J = 1078$ Hz, 1F), -114.7 (d, $J = 5.3$ Hz, 1F).

^{31}P NMR (162 MHz, CDCl_3): δ 24.5 (d, $J = 1078$ Hz, 1P).

HRMS (m/z) calculated for $\text{C}_9\text{H}_{12}\text{F}_2\text{O}_2\text{P}^+$ ($[\text{M}+\text{H}]^+$): 221.0537; found: 221.0535.

Ethyl (4-chlorobenzyl)phosphonofluoridate (7)



Colorless oil, yield: 77%.

^1H NMR (400 MHz, CDCl_3): δ 1.29 (t, $J = 6.8$ Hz, 3H), 3.26 (d, $J = 22.8$ Hz, 2H), 4.16–4.24 (m, 2H), 7.21–7.24 (m, 2H), 7.31 (d, $J = 8.4$ Hz, 2H).

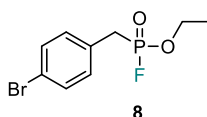
^{13}C NMR (100 MHz, CDCl_3): δ 16.2 (d, $J = 5.3$ Hz), 31.6 (dd, $J = 142.5$ Hz, 24.9 Hz), 64.1 (d, $J = 7.4$ Hz), 127.9 (d, $J = 9.9$ Hz), 129.0 (d, $J = 3$ Hz), 130.9 (d, $J = 7.1$ Hz), 133.6 (d, $J = 4.5$ Hz).

^{31}P NMR (162 MHz, CDCl_3): δ 24.2 (d, $J = 1078$ Hz, 1P).

^{19}F NMR (376 MHz, CDCl_3): δ -64.1 (d, $J = 1077$ Hz, 1F).

HRMS (m/z) calculated for $\text{C}_9\text{H}_{12}\text{ClFO}_2\text{P}^+$ ($[\text{M}+\text{H}]^+$): 237.0242; found: 237.0245.

Ethyl (4-bromobenzyl)phosphonofluoridate (8)



Colorless oil, yield: 87%.

¹H NMR (400 MHz, CDCl₃): δ 1.29 (t, *J* = 7.2 Hz, 3H), 3.25 (d, *J* = 22.8 Hz, 2H), 4.16–4.24 (m, 2H), 7.16–7.18 (m, 2H), 7.46 (d, *J* = 8.4 Hz, 2H).

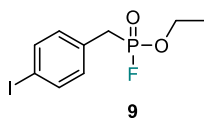
¹³C NMR (100 MHz, CDCl₃): δ 132.0 (d, *J* = 2.8 Hz), 131.3 (d, *J* = 7.1 Hz), 128.5 (d, *J* = 9.8 Hz), 121.7 (d, *J* = 4.8 Hz), 64.1 (d, *J* = 7.3 Hz), 31.6 (dd, *J* = 142.3 Hz, 24.8 Hz), 16.2 (d, *J* = 5.5 Hz).

¹⁹F NMR (376 MHz, CDCl₃): δ -64.1 (d, *J* = 1077 Hz, 1F).

³¹P NMR (162 MHz, CDCl₃): δ 24.2 (d, *J* = 1078 Hz, 1P).

HRMS (m/z) calculated for C₉H₁₂BrFO₂P⁺ ([M+H]⁺): 280.9737; found: 280.9731.

Ethyl (4-iodobenzyl)phosphonofluoridate (9)



Colorless oil, yield: 76%.

¹H NMR (400 MHz, CDCl₃): δ 1.30 (t, *J* = 6.8 Hz, 3H), 3.23 (d, *J* = 21.6 Hz, 2H), 4.16–4.24 (m, 2H), 7.04 (d, *J* = 8 Hz, 2H), 7.66 (d, *J* = 7.2 Hz, 2H).

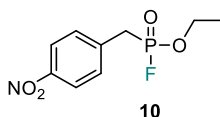
¹³C NMR (100 MHz, CDCl₃): δ 137.9 (d, *J* = 2.7 Hz), 131.5 (d, *J* = 6.9 Hz), 129.2 (d, *J* = 9.5 Hz), 93.1 (d, *J* = 5 Hz), 64.0 (d, *J* = 7.4 Hz), 31.8 (dd, *J* = 142.2 Hz, 24.7 Hz), 16.2 (d, *J* = 5.3 Hz).

³¹P NMR (162 MHz, CDCl₃): δ 23.9 (d, *J* = 1078 Hz, 1P).

¹⁹F NMR (376 MHz, CDCl₃): δ -63.9 (d, *J* = 1078 Hz, 1F).

HRMS (m/z) calculated for C₉H₁₂FIO₂P⁺ ([M+H]⁺): 328.9598; found: 328.9593.

Ethyl (4-nitrobenzyl)phosphonofluoridate (10)



Colorless oil, yield: 55%.

¹H NMR (400 MHz, CDCl₃): δ 1.31 (t, *J* = 6.8 Hz, 3H), 3.41 (d, *J* = 23.6 Hz, 2H), 4.19–4.27 (m, 2H), 7.48 (dd, *J* = 8.8 Hz, 2.4 Hz, 2H), 8.20 (d, *J* = 8.4 Hz, 2H).

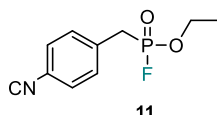
¹³C NMR (100 MHz, CDCl₃): δ 16.2 (d, *J* = 5.4 Hz), 32.2 (dd, *J* = 141.7 Hz, 25.4 Hz), 64.4 (d, *J* = 7.3 Hz), 123.9 (d, *J* = 2.7 Hz), 130.6 (d, *J* = 6.8 Hz), 137.2 (d, *J* = 9.7 Hz), 147.4 (d, *J* = 3.9 Hz).

³¹P NMR (162 MHz, CDCl₃): δ 22.5 (d, *J* = 1078 Hz, 1P).

¹⁹F NMR (376 MHz, CDCl₃): δ -62.9 (d, *J* = 1077 Hz, 1F).

HRMS (m/z) calculated for C₉H₁₂FNO₄P⁺ ([M+H]⁺): 248.0482; found: 248.0486.

Ethyl (4-cyanobenzyl)phosphonofluoridate (11)



Colorless oil, yield: 74%.

¹H NMR (400 MHz, CDCl₃): δ 1.30 (t, *J* = 7.2 Hz, 3H), 3.36 (d, *J* = 23.6 Hz, 2H), 4.19–4.27 (m, 2H), 7.42 (d, *J* = 7.6 Hz, 2H), 7.64 (d, *J* = 7.6 Hz, 2H).

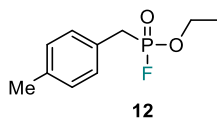
¹³C NMR (100 MHz, CDCl₃): δ 16.2 (d, *J* = 5.4 Hz), 32.5 (dd, *J* = 142 Hz, 25.3 Hz), 64.3 (d, *J* = 7.2 Hz), 111.7 (d, *J* = 3.9 Hz), 118.3, 130.5 (d, *J* = 7 Hz), 132.6 (d, *J* = 2.9 Hz), 135.1 (d, *J* = 9.6 Hz).

³¹P NMR (162 MHz, CDCl₃): δ 22.8 (d, *J* = 1078 Hz, 1P).

¹⁹F NMR (376 MHz, CDCl₃): δ -65.0 (d, *J* = 1078 Hz, 1F).

HRMS (m/z) calculated for C₁₀H₁₂FNO₂P⁺ ([M+H]⁺): 228.0584; found: 228.0587.

Ethyl (4-methylbenzyl)phosphonofluoridate (12)



Colorless oil, yield: 89%.

¹H NMR (400 MHz, CDCl₃): δ 1.29 (t, *J* = 7.2 Hz, 3H), 2.33 (s, 3H), 3.26 (d, *J* = 22.4 Hz, 2H), 4.15–4.22 (m, 2H), 7.13–7.19 (m, 4H).

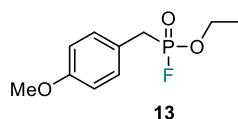
¹³C NMR (100 MHz, CDCl₃): δ 137.2 (d, *J* = 3.9 Hz), 129.5, 129.4, 126.2 (d, *J* = 9.9 Hz), 63.8 (d, *J* = 7.3 Hz), 31.7 (dd, *J* = 141.7 Hz, 24.3 Hz), 21.0, 16.2 (d, *J* = 5.4 Hz).

³¹P NMR (162 MHz, CDCl₃): δ 25.4 (d, *J* = 1078 Hz, 1P).

¹⁹F NMR (376 MHz, CDCl₃): δ -64.7 (d, *J* = 1076 Hz, 1F).

HRMS (m/z) calculated for C₁₀H₁₅FO₂P⁺ ([M+H]⁺): 217.0788; found: 217.0782.

Ethyl (4-methoxybenzyl)phosphonofluoridate (13)



Colorless oil, yield: 43%.

¹H NMR (400 MHz, CDCl₃): δ 7.24–7.31 (m, 2H), 6.89–6.92 (m, 2H), 4.20–4.22 (m, 2H), 3.80 (s, 3H), 3.23–3.30 (m, 2H), 1.30–1.34 (m, 3H).

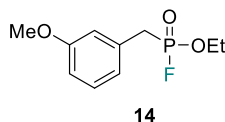
¹³C NMR (100 MHz, CDCl₃): δ 159.0, 130.7, 121.1, 114.3, 63.8, 55.2, 31.2 (dd, *J* = 143.1 Hz, 25.1 Hz), 16.2.

³¹P NMR (162 MHz, CDCl₃): δ 25.4 (d, *J* = 1078 Hz, 1P).

¹⁹F NMR (376 MHz, CDCl₃): δ -65.0 (d, *J* = 1078 Hz, 1F).

HRMS (m/z) calculated for C₁₀H₁₅FO₃P⁺ ([M+H]⁺): 233.0737; found: 233.0733.

Ethyl (3-methoxybenzyl)phosphonofluoridate (14)



Colorless oil, yield: 76%.

¹H NMR (400 MHz, CDCl₃): δ 7.22–7.26 (m, 1H), 6.86–6.89 (m, 1H), 6.81–6.84 (m, 2H), 4.15–4.23 (m, 2H), 3.8 (s, 3H), 3.23–3.30 (m, 2H), 1.3 (t, *J* = 7.1 Hz, 3H).

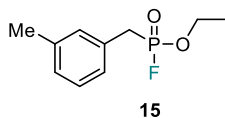
¹³C NMR (100 MHz, CDCl₃): δ 159.8 (d, *J* = 3.2 Hz), 130.7 (d, *J* = 9.8 Hz), 129.8 (d, *J* = 3.4 Hz), 122.0 (d, *J* = 7.2 Hz), 115.3 (d, *J* = 7.1 Hz), 113.0 (d, *J* = 3.8 Hz), 63.9 (d, *J* = 7.4 Hz), 55.2, 32.2 (dd, *J* = 142.4, 24.4 Hz), 16.2 (d, *J* = 5.6 Hz).

³¹P NMR (162 MHz, CDCl₃): δ 24.9 (d, *J* = 1076.6 Hz, 1P).

¹⁹F NMR (376 MHz, CDCl₃): δ -64.2 (d, *J* = 1076.1 Hz, 1F).

HRMS (m/z) calculated for C₁₀H₁₅FO₃P⁺ ([M+H]⁺): 233.0737; found: 233.0728.

Ethyl (3-methylbenzyl)phosphonofluoridate (15)



Colorless oil, yield: 74%.

¹H NMR (400 MHz, CDCl₃): δ 7.20–7.26 (m, 1H), 7.11 (s, 3H), 4.15–4.23 (m, 2H), 3.26 (dd, *J* = 22.4 Hz, 2 Hz, 2H), 2.34 (s, 3H), 1.27–1.29 (m, 3H).

¹³C NMR (100 MHz, CDCl₃): δ 138.5 (d, *J* = 3 Hz), 130.4 (d, *J* = 7 Hz), 129.2 (d, *J* =

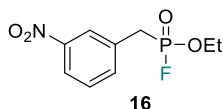
9.8 Hz), 128.5 (dd, $J = 40.4$ Hz, 2.7 Hz), 126.7 (d, $J = 7$ Hz), 63.9 (d, $J = 7.2$ Hz), 32.1 (dd, $J = 141.6$ Hz, 24.2 Hz), 21.3, 16.2 (d, $J = 5.3$ Hz).

^{31}P NMR (162 MHz, CDCl_3): δ 25.3 (d, $J = 1077$ Hz, 1P).

^{19}F NMR (376 MHz, CDCl_3): δ -64.3 (d, $J = 1077$ Hz, 1F).

HRMS (m/z) calculated for $\text{C}_{10}\text{H}_{15}\text{FO}_2\text{P}^+$ ($[\text{M}+\text{H}]^+$): 217.0788; found: 217.0781.

Ethyl (3-nitrobenzyl)phosphonofluoridate (16)



Colorless oil, yield: 77%.

^1H NMR (600 MHz, CDCl_3): δ 8.19–8.15 (m, 1H), 7.66–7.67 (m, 1H), 7.54 (t, $J = 8.0$ Hz, 1H), 4.22–4.28 (m, 1H), 3.41 (d, $J = 23.2$ Hz, 1H), 1.32 (t, $J = 7.1$ Hz, 2H).

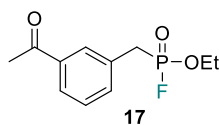
^{13}C NMR (151 MHz, CDCl_3): δ 148.5, 135.7 (d, $J = 6.6$ Hz), 131.7 (d, $J = 9.5$ Hz), 129.9 (d, $J = 3.3$ Hz), 124.6 (d, $J = 7.6$ Hz), 122.7 (d, $J = 3.3$ Hz), 64.4 (d, $J = 7.6$ Hz), 31.9 (dd, $J = 143.7$, 25.6 Hz), 16.2 (d, $J = 5.6$ Hz).

^{31}P NMR (243 MHz, CDCl_3): δ 22.9 (d, $J = 1077.1$ Hz, 1P).

^{19}F NMR (565 MHz, CDCl_3): δ -63.3 (d, $J = 1078.6$ Hz, 1F).

HRMS (m/z) calculated for $\text{C}_9\text{H}_{12}\text{FNO}_4\text{P}^+$ ($[\text{M}+\text{H}]^+$): 248.0482; found: 248.0487.

Ethyl (3-acetylbenzyl)phosphonofluoridate (17)



Colorless oil, yield: 47%.

^1H NMR (400 MHz, CDCl_3): δ 7.9 (s, 2H), 7.5 (d, $J = 8.0$ Hz, 1H), 7.4 (t, $J = 7.8$ Hz, 1H), 4.17–4.25 (m, 2H), 3.4 (d, $J = 22.9$ Hz, 2H), 2.6 (s, 3H), 1.3 (t, $J = 7.6$ Hz, 3H).

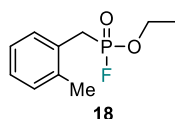
^{13}C NMR (100 MHz, CDCl_3): δ 191.1, 133.0 (d, $J = 7.8$ Hz), 130.7 (d, $J = 10.7$ Hz), 129.4 (d, $J = 3.9$ Hz), 128.4 (d, $J = 8.2$ Hz), 126.4 (d, $J = 4.5$ Hz), 124.9, 64.2 (d, $J = 8.3$ Hz), 32.1 (dd, $J = 143.1$, 25.7 Hz), 29.6, 16.2 (d, $J = 6.4$ Hz).

^{31}P NMR (162 MHz, CDCl_3): δ 23.8 (d, $J = 1077.3$ Hz, 1P).

^{19}F NMR (376 MHz, CDCl_3): δ -63.8 (d, $J = 1077.3$ Hz, 1F).

HRMS (m/z) calculated for $\text{C}_{11}\text{H}_{15}\text{FO}_3\text{P}^+$ ($[\text{M}+\text{H}]^+$): 245.0737; found: 245.0731.

Ethyl (2-methylbenzyl)phosphonofluoridate (18)



Colorless oil, yield: 66%.

¹H NMR (400 MHz, CDCl₃): δ 1.21 (t, *J* = 6.8 Hz, 3H), 2.31 (s, 3H), 3.23 (d, *J* = 22.8 Hz, 2H), 4.07–4.15 (m, 2H), 7.11–7.12 (m, 3H), 7.16–7.19 (m, 2H).

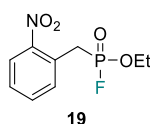
¹³C NMR (100 MHz, CDCl₃): δ 16.2 (d, *J* = 5.5 Hz), 19.8, 29.5 (dd, *J* = 141.4 Hz, 24.2 Hz), 63.8 (d, *J* = 7.4 Hz), 126.3 (d, *J* = 3.4 Hz), 127.7 (d, *J* = 3.9 Hz), 130.5 (d, *J* = 5.9 Hz), 130.7 (d, *J* = 3.2 Hz), 137.0 (d, *J* = 2.7 Hz).

³¹P NMR (162 MHz, CDCl₃): δ 25.3 (d, *J* = 1079 Hz, 1P).

¹⁹F NMR (376 MHz, CDCl₃): δ -63.5 (d, *J* = 1079 Hz, 1F).

HRMS (m/z) calculated for C₁₀H₁₅FO₂P⁺ ([M+H]⁺): 217.0788; found: 217.0791.

Ethyl (2-nitrobenzyl)phosphonofluoridate (19)



Colorless oil, yield: 32%.

¹H NMR (400 MHz, CDCl₃): δ 8.04 (d, *J* = 8.4 Hz, 1H), 7.58–7.62 (m, 1H), 7.47–7.50 (m, 2H), 4.18–4.25 (m, 2H), 3.77–3.97 (m, 2H), 1.29 (t, *J* = 7.2 Hz, 3H).

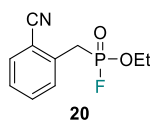
¹³C NMR (100 MHz, CDCl₃): δ 149.0, 133.5 (d, *J* = 4.2 Hz), 133.1 (d, *J* = 7.3 Hz), 128.8 (d, *J* = 4.5 Hz), 125.6 (d, *J* = 3.9 Hz), 125.3 (d, *J* = 11.0 Hz), 64.4 (d, *J* = 8.1 Hz), 29.4 (dd, *J* = 143.5, 26.2 Hz), 16.1 (d, *J* = 6.4 Hz).

³¹P NMR (162 MHz, CDCl₃): δ 22.3 (d, *J* = 1079.1, 1P).

¹⁹F NMR (376 MHz, CDCl₃): δ -62.7 (d, *J* = 1077.7 Hz, 1F).

HRMS (m/z) calculated for C₉H₁₂FNO₄P⁺ ([M+H]⁺): 248.0482; found: 248.0485.

Ethyl (2-cyanobenzyl)phosphonofluoridate (20)



Colorless oil, yield: 43%.

¹H NMR (400 MHz, CDCl₃): δ 7.66–7.68 (m, 1H), 7.52–7.61 (m, 2H), 7.39–7.42 (m, 1H), 4.24–4.28 (m, 2H), 3.56 (d, *J* = 23.6 Hz, 2H), 1.31 (t, *J* = 7.2 Hz, 3H).

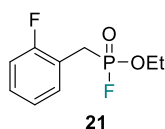
¹³C NMR (100 MHz, CDCl₃): δ 133.6 (d, *J* = 10.2 Hz), 133.1 (d, *J* = 6.1 Hz), 133.0 (d, *J* = 6.5 Hz), 130.9 (d, *J* = 6.5 Hz), 128.2 (d, *J* = 4.5 Hz), 117.2 (d, *J* = 3.1 Hz), 113.5 (d, *J* = 8.6 Hz), 64.5 (d, *J* = 8.0 Hz), 30.8 (dd, *J* = 143.7, 26.6 Hz), 16.1 (d, *J* = 6.5 Hz).

³¹P NMR (162 MHz, CDCl₃): δ 21.8 (d, *J* = 1078.5 Hz, 1P).

¹⁹F NMR (376 MHz, CDCl₃): δ -62.9 (d, *J* = 1078.1 Hz, 1F).

HRMS (m/z) calculated for C₁₀H₁₂FNO₂P⁺ ([M+H]⁺): 228.0584; found: 228.0567.

Ethyl (2-fluorobenzyl)phosphonofluoridate (21)



Colorless oil, yield: 81%.

¹H NMR (400 MHz, CDCl₃): δ 7.27–7.38 (m, 1H), 7.27–7.31 (m, 1H), 7.06–7.15 (m, 2H), 4.19–4.26 (m, 2H), 3.36 (dd, *J* = 22.8, 3.2 Hz, 2H), 1.31 (t, *J* = 7.2 Hz, 3H).

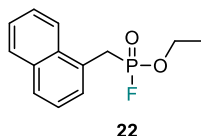
¹³C NMR (100 MHz, CDCl₃): δ 160.8 (dd, *J* = 247.4, 7.8 Hz), 131.6 (dd, *J* = 5.9, 3.2 Hz), 129.5 (dd, *J* = 8.0, 3.9 Hz), 124.4 (t, *J* = 3.7 Hz), 117.0 (dd, *J* = 15.5, 10.1 Hz), 115.7 (dd, *J* = 21.8, 3.2 Hz), 64.1 (d, *J* = 7.2 Hz), 25.1 (ddd, *J* = 145.6, 26.5, 3.5 Hz), 16.2 (d, *J* = 5.6 Hz).

³¹P NMR (162 MHz, CDCl₃): δ 23.7 (d, *J* = 1077.9, 5.18 Hz, 1P).

¹⁹F NMR (376 MHz, CDCl₃): δ -64.0 (d, *J* = 1076.1 Hz, 1F), -116.8 (s, 1F).

HRMS (m/z) calculated for C₉H₁₂F₂O₂P⁺ ([M+H]⁺): 221.0537; found: 221.0531.

Ethyl (naphthalen-1-ylmethyl)phosphonofluoridate (22)



Colorless oil, yield: 70%.

¹H NMR (400 MHz, CDCl₃): δ 1.19 (t, *J* = 7.2 Hz, 3H), 3.78 (dd, *J* = 23.2 Hz, 2.4 Hz, 2H), 4.09–4.16 (m, 2H), 7.43–7.53 (m, 3H), 7.57 (t, *J* = 7.2 Hz, 1H), 7.82 (d, *J* = 7.6 Hz, 1H), 7.87 (d, *J* = 8 Hz, 1H), 8.03 (d, *J* = 8.4 Hz, 1H).

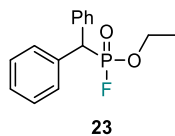
¹³C NMR (100 MHz, CDCl₃): δ 133.9 (d, *J* = 2.8 Hz), 131.7 (d, *J* = 5.5 Hz), 128.8, 128.6 (d, *J* = 8.1 Hz), 128.5 (d, *J* = 4.4 Hz), 126.5, 125.9, 125.8 (d, *J* = 10.8 Hz), 125.3 (d, *J* = 4.2 Hz), 123.8, 64.0 (d, *J* = 7.3 Hz), 29.2 (dd, *J* = 142.6 Hz, 24.9 Hz), 16.1 (d, *J* = 4.3 Hz).

³¹P NMR (162 MHz, CDCl₃): δ 24.8 (d, *J* = 1081 Hz, 1P).

¹⁹F NMR (376 MHz, CDCl₃): δ -62.9 (d, *J* = 1080 Hz, 1F).

HRMS (m/z) calculated for C₁₃H₁₅FO₂P⁺ ([M+H]⁺): 253.0788; found: 253.0781.

Ethyl benzhydrylphosphonofluoridate (23)



Colorless oil, yield: 42%.

¹H NMR (400 MHz, CDCl₃): δ 1.19 (t, *J* = 6.8 Hz, 3H), 4.06–4.21 (m, 2H), 4.60 (d, *J* = 27.2 Hz, 2H), 7.30 (d, *J* = 6.4 Hz, 2H), 7.35 (t, *J* = 7.2 Hz, 4H), 7.50 (d, *J* = 7.2 Hz, 4H).

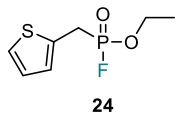
¹³C NMR (100 MHz, CDCl₃): δ 16.0 (d, *J* = 4.3 Hz), 49.9 (dd, *J* = 141.3 Hz, 22.9 Hz), 64.6 (d, *J* = 6.9 Hz), 127.7, 128.9, 129.2 (dd, *J* = 18 Hz, 8.5 Hz), 134.9 (d, *J* = 3.7 Hz).

³¹P NMR (162 MHz, CDCl₃): δ 23.4 (d, *J* = 1094 Hz, 1P).

¹⁹F NMR (376 MHz, CDCl₃): δ -64.2 (d, *J* = 1094 Hz, 1F).

HRMS (m/z) calculated for C₁₅H₁₇FO₂P⁺ ([M+H]⁺): 279.0945; found: 279.0947.

Ethyl (thiophen-2-ylmethyl)phosphonofluoridate (24)



Colorless oil, yield: 51%.

¹H NMR (400 MHz, CDCl₃): δ 7.15–7.19 (m, 1H), 6.90–6.95 (m, 2H), 4.14–4.22 (m, 2H), 3.45 (dd, *J* = 21.2 Hz, 6.4 Hz, 2H), 1.24 (t, *J* = 7.2 Hz, 3H).

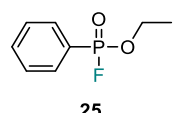
¹³C NMR (100 MHz, CDCl₃): δ 129.6 (d, *J* = 11.3 Hz), 127.9 (d, *J* = 8.8 Hz), 127.3, 125.5, 64.3 (d, *J* = 7.1 Hz), 26.6 (dd, *J* = 148 Hz, 27.5 Hz), 16.2 (d, *J* = 4.3 Hz).

³¹P NMR (162 MHz, CDCl₃): δ 22.6 (d, *J* = 1080 Hz, 1P).

¹⁹F NMR (376 MHz, CDCl₃): δ -64.8 (d, *J* = 1080 Hz, 1F).

HRMS (m/z) calculated for C₇H₁₁FO₂PS⁺ ([M+H]⁺): 209.0196; found: 209.0197.

Ethyl phenylphosphonofluoridate (25)



Colorless oil, yield: 59%.

¹H NMR (400 MHz, CDCl₃): δ 1.42 (t, *J* = 7.2 Hz, 3H), 4.30–4.38 (m, 2H), 7.52 (dd, *J* = 12.4 Hz, 7.2 Hz, 2H), 7.64 (t, *J* = 7.6 Hz, 1H), 7.85 (dd, *J* = 14.4 Hz, 8 Hz, 2H).

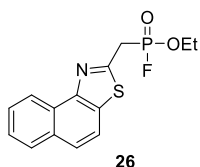
¹³C NMR (100 MHz, CDCl₃): δ 133.6 (d, *J* = 2.7 Hz), 131.7 (d, *J* = 10.8 Hz), 128.7 (d, *J* = 16.3 Hz), 123.8 (d, *J* = 29.9 Hz), 63.9 (d, *J* = 6.1 Hz), 16.3 (d, *J* = 5.7 Hz).

³¹P NMR (162 MHz, CDCl₃): δ 17.1 (d, *J* = 1043 Hz, 1P).

¹⁹F NMR (376 MHz, CDCl₃): δ -63.9 (d, *J* = 1042 Hz, 1F).

HRMS (m/z) calculated for C₈H₁₁FO₂P⁺ ([M+H]⁺): 189.0475; found: 189.0471.

Ethyl (naphtho[1,2-*d*]thiazol-2-ylmethyl)phosphonofluoridate (26)



Colorless oil, yield: 45%.

¹H NMR (600 MHz, CDCl₃): δ 8.78 (d, *J* = 8.2 Hz, 1H), 7.95 (d, *J* = 8.1 Hz, 1H), 7.87 (d, *J* = 8.8 Hz, 1H), 7.83 (d, *J* = 8.8 Hz, 1H), 7.68 (t, *J* = 7.2 Hz, 1H), 7.59 (t, *J* = 7.2 Hz, 1H), 4.33–4.40 (m, 2H), 4.00 (dd, *J* = 22.9, 2.3 Hz, 2H), 1.38 (t, *J* = 7.1 Hz, 3H).

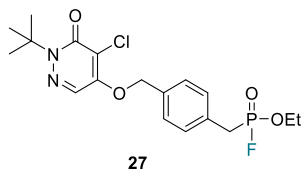
¹³C NMR (151 MHz, CDCl₃): δ 156.9 (dd, *J* = 9.7, 1.9 Hz), 149.3 (d, *J* = 1.9 Hz), 132.6 (d, *J* = 1.9 Hz), 132.0, 128.5, 128.1, 127.2, 126.4, 126.3, 123.8, 118.6, 65.0 (d, *J* = 7.4 Hz), 31.6 (dd, *J* = 146.0, 27.7 Hz), 16.3 (d, *J* = 5.9 Hz).

³¹P NMR (243 MHz, CDCl₃): δ 19.5 (d, *J* = 1073.7 Hz, 1P).

¹⁹F NMR (565 MHz, CDCl₃): δ -61.6 (d, *J* = 1075.0 Hz, 1F).

HRMS (m/z) calculated for C₁₄H₁₄FNO₂PS⁺ ([M+H]⁺): 310.0461; found: 310.0455.

Ethyl (4-(((1-(tert-butyl)-5-chloro-6-oxo-1,6-dihydropyridazin-4-yl)oxy)methyl)benzyl)phosphonofluoridate (27)



Colorless oil, yield: 57%.

¹H NMR (600 MHz, CDCl₃): δ 7.71 (s, 1H), 7.38 (d, *J* = 7.9 Hz, 2H), 7.33–7.35 (m, 2H), 5.29 (s, 2H), 4.17–4.22 (m, 2H), 3.28–3.30 (m, 2H), 1.61 (s, 9H), 1.28 (t, *J* = 7.1 Hz, 3H).

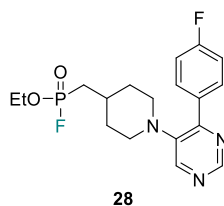
¹³C NMR (151 MHz, CDCl₃): δ 159.0, 153.6, 134.3 (d, *J* = 4.3 Hz), 130.2 (d, *J* = 6.8 Hz), 130.1 (d, *J* = 10.0 Hz), 127.5 (d, *J* = 3.8 Hz), 125.0, 118.3, 71.4, 66.4, 64.0 (d, *J* = 7.6 Hz), 31.9 (dd, *J* = 142.7, 24.4 Hz), 27.8, 16.2 (d, *J* = 5.5 Hz).

³¹P NMR (243 MHz, CDCl₃): δ 24.5 (d, *J* = 1077.4 Hz, 1P).

¹⁹F NMR (565 MHz, CDCl₃): δ -64.0 (d, *J* = 1077.9 Hz, 1F).

HRMS (m/z) calculated for C₁₈H₂₄ClFN₂O₄P⁺ ([M+H]⁺): 417.1141; found: 417.1146.

Ethyl ((1-(4-(4-fluorophenyl)pyrimidin-5-yl)piperidin-4-yl)methyl)phosphonofluoridate (28)



Colorless oil, yield: 50%.

¹H NMR (600 MHz, CDCl₃): δ 8.89 (s, 1H), 8.40 (s, 1H), 8.09–8.11 (m, 2H), 7.15 (t, *J* = 8.7 Hz, 2H), 4.23–4.31 (m, 2H), 3.16–3.21 (m, 2H), 2.67 (t, *J* = 11.4 Hz, 2H), 1.91–1.93 (m, 1H), 1.87–1.90 (m, 3H), 1.67 (s, 3H), 1.38 (t, *J* = 7.0 Hz, 3H).

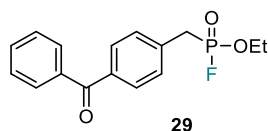
¹³C NMR (151 MHz, CDCl₃): δ 164.4, 162.8, 156.6, 152.7, 147.3, 143.9, 134.0, 130.3 (d, *J* = 7.9 Hz), 115.5 (d, *J* = 22.0 Hz), 63.2 (d, *J* = 7.6 Hz), 50.7, 46.7, 32.9 (d, *J* = 11.5 Hz), 31.1 (dd, *J* = 142.4, 21.3 Hz), 29.7, 16.3 (d, *J* = 5.5 Hz).

³¹P NMR (243 MHz, CDCl₃): δ 29.7 (d, *J* = 1072.0 Hz, 1P).

¹⁹F NMR (376 MHz, CDCl₃): δ -60.3 (d, *J* = 1072.9 Hz, 1F), -110.4 (s, 1F).

HRMS (m/z) calculated for C₁₈H₂₃F₂N₃O₂P⁺ ([M+H]⁺): 382.1490; found: 382.1483.

Ethyl (4-benzoylbenzyl)phosphonofluoridate (29)



Colorless oil, yield: 78%.

¹H NMR (400 MHz, DMSO-*d*₆): δ 7.78 (d, *J* = 8.0 Hz, 4H), 7.59 (t, *J* = 7.2 Hz, 1H), 7.48 (t, *J* = 7.2 Hz, 2H), 7.42 (d, *J* = 6.8 Hz, 2H), 4.20–4.28 (m, 2H), 3.39 (d, *J* = 23.6 Hz, 2H), 1.32 (t, *J* = 6.8 Hz, 3H).

¹³C NMR (100 MHz, DMSO-*d*₆): δ 196.0, 137.3, 136.8, 134.2 (d, *J* = 9.7 Hz), 132.5, 130.6 (d, *J* = 2.7 Hz), 129.9, 129.6 (d, *J* = 20.0 Hz), 128.3 (d, *J* = 14.5 Hz), 64.2 (d, *J* =

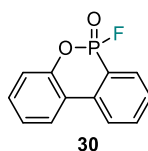
7.5 Hz), 32.44 (dd, $J_1 = 141.5$ Hz, $J_2 = 25.0$ Hz), 16.2.

^{31}P NMR (162 MHz, $\text{DMSO-}d_6$): δ 23.8 (d, $J = 1077$ Hz, 1P).

^{19}F NMR (376 MHz, CDCl_3): δ -63.43 (d, $J = 1077$ Hz, 1F).

HRMS (m/z) calculated for $\text{C}_{16}\text{H}_{17}\text{FO}_3\text{P}^+$ ($[\text{M}+\text{H}]^+$): 307.0894; found: 307.0899.

6-Fluorodibenzo[*c,e*][1,2]oxaphosphinine 6-oxide (30)



Colorless oil, yield: 67%.

^1H NMR (400 MHz, CDCl_3): δ 7.29–7.36 (m, 2H), 7.44 (t, $J = 3.9$ Hz, 1H), 7.56–7.61 (m, 1H), 7.82 (t, $J = 8$ Hz, 1H), 7.97–8.12 (m, 3H).

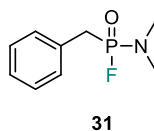
^{13}C NMR (100 MHz, CDCl_3): δ 149.3 (d, $J = 8.3$ Hz), 137.5 (d, $J = 7.9$ Hz), 134.9, 131.0 (t, $J = 4.6$ Hz), 128.6 (d, $J = 16.4$ Hz), 125.6, 125.3, 124.3 (d, $J = 13$ Hz), 121.7 (d, $J = 12.7$ Hz), 120.3 (d, $J = 7.3$ Hz), 119.5 (d, $J = 28.9$ Hz), 117.7, (d, $J = 29$ Hz).

^{31}P NMR (162 MHz, CDCl_3): δ 7.3 (d, $J = 1074$ Hz, 1P).

^{19}F NMR (376 MHz, CDCl_3): δ -55.4 (d, $J = 1073$ Hz, 1F).

HRMS (m/z) calculated for $\text{C}_{12}\text{H}_9\text{FO}_2\text{P}^+$ ($[\text{M}+\text{H}]^+$): 235.0319; found: 235.0325.

P-benzyl-*N,N*-dimethylphosphonamidic fluoride (31)



Colorless oil, yield: 67%.

^1H NMR (400 MHz, CDCl_3): δ 7.32–7.37 (m, 5H), 3.24–3.38 (m, 2H), 2.65 (d, $J = 9.6$, 6H).

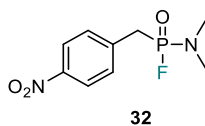
^{13}C NMR (100 MHz, CDCl_3): δ 130.4 (dd, $J = 8.6, 2.2$ Hz), 129.4 (d, $J = 6.6$ Hz), 128.7, 127.2, 35.74 (d, $J = 4.1$ Hz), 32.5 (dd, $J = 32.0, 129.4$ Hz).

^{31}P NMR (162 MHz, CDCl_3): δ 34.1 (d, $J = 1049$ Hz, 1P).

^{19}F NMR (376 MHz, CDCl_3): δ -67.3 (d, $J = 1049$ Hz, 1F).

HRMS (m/z) calculated for $\text{C}_9\text{H}_{14}\text{FNOP}^+$ ($[\text{M}+\text{H}]^+$): 202.0792; found: 202.0797.

N,N-Dimethyl-*P*-(4-nitrobenzyl)phosphonamidic fluoride (32)



Yellowish solid, yield: 65%.

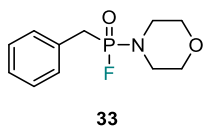
¹H NMR (400 MHz, CDCl₃): δ 8.18 (d, *J* = 8 Hz, 2H), 7.47 (d, *J* = 7.6 Hz, 2H), 3.29–3.45 (m, 2H), 2.68 (d, *J* = 9.6 Hz, 6H).

¹³C NMR (100 MHz, CDCl₃): δ 147.2 (d, *J* = 3.5 Hz), 138.4 (d, *J* = 8.5 Hz), 130.4 (d, *J* = 6.5 Hz), 123.9 (d, *J* = 2.4 Hz), 35.8 (d, *J* = 3.8 Hz), 32.5 (dd, *J* = 129.2 Hz, 33.7 Hz). **³¹P NMR (162 MHz, CDCl₃):** δ 31.5 (d, *J* = 1053 Hz, 1P).

¹⁹F NMR (376 MHz, CDCl₃): δ -68.1 (d, *J* = 1050 Hz, 1F).

HRMS (m/z) calculated for C₁₂H₉FO₂P⁺ ([M+H]⁺): 247.0642; found: 247.0644.

Benzyl(morpholino)phosphinic fluoride (33)



White solid, yield: 61%.

¹H NMR (400 MHz, CDCl₃): δ 7.33–7.40 (m, 5H), 3.56–3.60 (m, 4H), 3.25–3.43 (m, 2H), 3.07–3.16 (m, 4H).

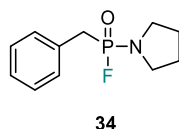
¹³C NMR (100 MHz, CDCl₃): δ 130.1 (d, *J* = 8.6 Hz), 129.6 (d, *J* = 6.8 Hz), 128.9 (d, *J* = 2.7 Hz), 127.5 (d, *J* = 3.5 Hz), 66.9 (d, *J* = 4.1 Hz), 43.9, 38.9 (dd, *J* = 129.9 Hz, 31.3 Hz).

³¹P NMR (162 MHz, CDCl₃): δ 31.0 (d, *J* = 1055 Hz, 1P).

¹⁹F NMR (376 MHz, CDCl₃): δ -65.5 (d, *J* = 1055 Hz, 1F).

HRMS (m/z) calculated for C₁₂H₉FO₂P⁺ ([M+H]⁺): 244.0897; found: 244.0906.

Benzyl(pyrrolidin-1-yl)phosphinic fluoride (34)



White solid, yield: 51%.

¹H NMR (400 MHz, CDCl₃): δ 7.27–7.33 (m, 5H), 3.13–3.39 (m, 6H), 1.74–1.79 (m, 4H).

¹³C NMR (100 MHz, CDCl₃): δ 130.6 (d, *J* = 8.2 Hz), 129.5 (d, *J* = 6.5 Hz), 128.7 (d,

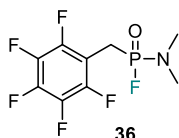
$J = 2$ Hz), 127.1 (d, $J = 1$ Hz), 46.4 (d, $J = 4$ Hz), 33.1 (dd, $J = 128.6$ Hz, 31.1 Hz), 26.1 (d, $J = 8.1$ Hz).

^{31}P NMR (162 MHz, CDCl_3): δ 31.3 (d, $J = 1044$ Hz, 1P).

^{19}F NMR (376 MHz, CDCl_3): δ -64.4 (d, $J = 1045$ Hz, 1F).

HRMS (m/z) calculated for $\text{C}_{12}\text{H}_9\text{FO}_2\text{P}^+$ $[\text{M}+\text{H}]^+$: 228.0948; found: 228.0941.

N,N-Dimethyl-*P*-((perfluorophenyl)methyl)phosphonamidic fluoride (36)



Light yellow oil, yield: 43%.

^1H NMR (400 MHz, CDCl_3): δ 3.25–3.41 (m, 2H), 2.64 (d, $J = 10.0$ Hz, 6H).

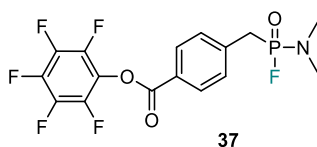
^{13}C NMR (100 MHz, CDCl_3): δ 179.9, 144.9 (d, $J = 250$ Hz), 140.0 (d, $J = 278$ Hz), 62.5 (d, $J = 6.5$ Hz), 20.9 (d, $J = 142$ Hz), 16.2 (d, $J = 6$ Hz).

^{31}P NMR (162 MHz, CDCl_3): δ 28.1 (d, $J = 1054$ Hz, 1P).

^{19}F NMR (376 MHz, CDCl_3): δ -65.3 (d, $J = 1053$ Hz, 2F), -141.5 (2F), -154.4 (1F), -161.2 (2F).

HRMS (m/z) calculated for $\text{C}_9\text{H}_9\text{F}_6\text{NOP}^+$ $[\text{M}+\text{H}]^+$: 292.0320; found: 292.0324.

Perfluorophenyl 4-(((dimethylamino)fluorophosphoryl)methyl)benzoate (37)



White solid, yield: 66%.

^1H NMR (400 MHz, CDCl_3): δ 2.70 (dd, $J = 10.0$ Hz, 2.0 Hz, 6H), 3.33–3.49 (m, 2H), 7.48 (d, $J = 8.0$ Hz, 2H), 8.17 (d, $J = 8.0$ Hz, 2H).

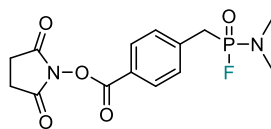
^{13}C NMR (100 MHz, CDCl_3): δ 162.2, 142.6 (d, $J = 8.5$ Hz), 140.1 (m), 139.2 (m), 137.9 (m), 136.7 (m), 131.2 (d, $J = 2.5$ Hz), 130.1 (d, $J = 6.5$ Hz), 126.1 (d, $J = 3.4$ Hz), 35.8 (d, $J = 4.2$ Hz), 32.8 (dd, $J = 32.7$ Hz, 129.3 Hz).

^{31}P NMR (162 MHz, CDCl_3): δ 32.0 ($J = 1052$ Hz, 1P).

^{19}F NMR (376 MHz, CDCl_3): δ -65.8 (d, $J = 1052$ Hz, 1F), -152.4 (d, $J = 17.7$ Hz, 2F), -157.8 (t, $J = 21.1$ Hz, 1F), -162.2 (t, $J = 21.8$ Hz, 2F).

HRMS (m/z) calculated for $\text{C}_{16}\text{H}_{13}\text{F}_6\text{NO}_3\text{P}^+$ $[\text{M}+\text{H}]^+$: 412.0532; found: 412.0537.

2,5-Dioxopyrrolidin-1-yl 4-(((dimethylamino)fluorophosphoryl)methyl)benzoate (38)



38

White solid, yield: 45%.

¹H NMR (400 MHz, CDCl₃): δ 2.65 (dd, *J* = 9.6 Hz, 1.6 Hz, 6H), 2.90 (s, 4H), 3.30–3.46 (m, 2H), 7.46 (d, *J* = 6.4 Hz, 2H), 8.10 (d, *J* = 8.0 Hz, 2H).

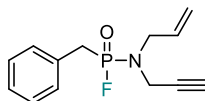
¹³C NMR (100 MHz, CDCl₃): δ 169.2, 161.5, 138.5 (d, *J* = 6.2 Hz), 131.0 (d, *J* = 2.8 Hz), 130.1 (d, *J* = 6.6 Hz), 124.2 (d, *J* = 3.4 Hz), 35.9 (d, *J* = 4.1 Hz), 33.0 (dd, *J* = 129.1 Hz, 33.7 Hz), 25.6.

³¹P NMR (162 MHz, CDCl₃): δ 31.9 (d, *J* = 1052 Hz, 1P).

¹⁹F NMR (376 MHz, CDCl₃): δ -65.7 (d, *J* = 1051 Hz, 1F).

HRMS (m/z) calculated for C₁₄H₁₇FN₂O₅P⁺ ([M+H]⁺): 343.0854; found: 343.0856.

***N*-Allyl-*P*-benzyl-*N*-(prop-2-yn-1-yl)phosphonamidic fluoride (39)**



39

White solid, yield: 63%.

¹H NMR (400 MHz, CDCl₃): δ 7.26–7.33 (m, 5H), 5.52–5.59 (m, 1H), 5.19–5.25 (m, 2H), 3.68–3.91 (m, 2H), 3.61–3.64 (m, 2H), 3.27–3.42 (m, 2H), 2.23 (s, 1H).

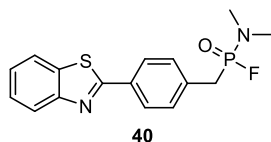
¹³C NMR (100 MHz, CDCl₃): δ 133.5 (d, *J* = 2 Hz), 129.9 (d, *J* = 8.6 Hz), 129.8 (d, *J* = 6.7 Hz), 128.8 (d, *J* = 2 Hz), 127.4 (d, *J* = 3.2 Hz), 119.4, 78.6, 72.6, 47.6 (d, *J* = 3.2 Hz), 34.0 (d, *J* = 5.4 Hz), 33.7 (d, *J* = 128.5 Hz).

³¹P NMR (162 MHz, CDCl₃): δ 33.1 (d, *J* = 1058 Hz, 1P).

¹⁹F NMR (376 MHz, CDCl₃): δ -65.4 (d, *J* = 1057 Hz, 1F).

HRMS (m/z) calculated for C₁₃H₁₆FNOP⁺ ([M+H]⁺): 252.0948; found: 252.0940.

***P*-(4-(benzo[*d*]thiazol-2-yl)benzyl)-*N,N*-dimethylphosphonamidic fluoride (40)**



White solid, yield: 47%.

¹H NMR (400 MHz, CDCl₃): δ 8.07 (d, *J* = 8.0 Hz, 3H), 7.91 (d, *J* = 7.7 Hz, 1H), 7.48–7.52 (m, 1H), 7.38–7.45 (m, 3H), 3.29–3.45 (m, 2H), 2.68 (dd, *J* = 9.7, 2.2 Hz, 6H).

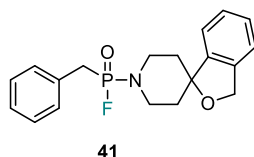
¹³C NMR (100 MHz, CDCl₃): δ 154.1, 135.0, 132.8, 130.2 (d, *J* = 6.8 Hz), 127.9 (d, *J* = 3.0 Hz), 127.8, 127.5, 126.4, 125.3, 123.2, 121.6, 35.9 (d, *J* = 4.2 Hz, 4H), 30.8 (dd, *J* = 43.9 Hz, 125.4 Hz).

³¹P NMR (162 MHz, CDCl₃): δ 32.9 (d, *J* = 1052 Hz, 1P).

¹⁹F NMR (376 MHz, CDCl₃): δ -66.4 (d, *J* = 1051 Hz, 1F).

HRMS (m/z) calculated for C₁₆H₁₇FN₂OPS⁺ ([M+H]⁺): 335.0778; found: 335.0772.

Benzyl(3*H*-spiro[isobenzofuran-1,4'-piperidin]-1'-yl)phosphinic fluoride (41)



White solid, yield: 63%.

¹H NMR (400 MHz, CDCl₃): δ 7.23–7.32 (m, 5H), 7.18–7.19 (m, 2H), 7.10–7.12 (m, 1H), 6.88–6.90 (m, 1H), 4.95 (s, 2H), 3.35–3.45 (m, 2H), 3.17–3.54 (m, 2H), 3.03–3.12 (m, 2H), 1.51–1.61 (m, 4H).

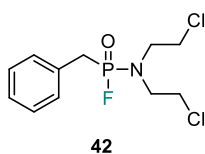
¹³C NMR (100 MHz, CDCl₃): δ 144.9, 138.6, 130.7 (d, *J* = 2.4 Hz), 130.6 (d, *J* = 2.3 Hz), 129.6 (d, *J* = 6.8 Hz), 128.9, 127.8, 127.4, 127.3, 120.9 (d, *J* = 54.5 Hz), 84.3, 70.9, 40.8 (d, *J* = 3.1 Hz), 36.4, 32.9 (dd, *J* = 131.0 Hz, 32.6 Hz).

³¹P NMR (162 MHz, CDCl₃): δ 32.1 (d, *J* = 1052 Hz, 1P).

¹⁹F NMR (376 MHz, CDCl₃): δ -64.6 (dd, *J* = 5.64 Hz, 1051 Hz, 1F).

HRMS (m/z) calculated for C₁₉H₂₂FNO₂P⁺ ([M+H]⁺): 346.1367; found: 346.1368.

P-benzyl-*N,N*-bis(2-chloroethyl)phosphonamidic fluoride (42)



White solid, yield: 71%.

$^1\text{H NMR}$ (400 MHz, CDCl_3): δ 7.34–7.38 (m, 5H), 3.35–3.47 (m, 8H), 3.23–3.31 (m, 2H).

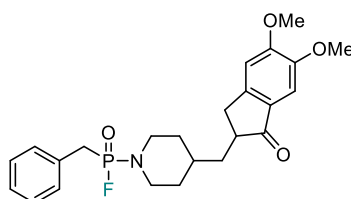
$^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 129.7, 129.6, 129.1, 127.7 (d, $J = 15.4$ Hz), 48.0 (d, $J = 3.9$ Hz), 41.6, 33.0 (dd, $J = 128.7$ Hz, 30.1 Hz).

$^{31}\text{P NMR}$ (162 MHz, CDCl_3): δ 34.0 (d, $J = 1062$ Hz, 1P).

$^{19}\text{F NMR}$ (376 MHz, CDCl_3): δ -62.9 (d, $J = 1061$ Hz, 1F).

HRMS (m/z) calculated for $\text{C}_{11}\text{H}_{16}\text{Cl}_2\text{FNOP}^+$ ($[\text{M}+\text{H}]^+$): 298.0325; found: 298.0331.

Benzyl(4-((5,6-dimethoxy-1-oxo-2,3-dihydro-1*H*-inden-2-yl)methyl)piperidin-1-yl)phosphinic fluoride (43)



43

White solid, yield: 76%.

$^1\text{H NMR}$ (400 MHz, CDCl_3): δ 7.38–7.46 (m, 5H), 7.36 (s, 1H), 6.95 (s, 1H), 4.06 (s, 3H), 4.00 (s, 3H), 3.58–3.66 (m, 2H), 3.42–3.48 (m, 1H), 3.29–3.83 (m, 2H), 2.74–2.77 (m, 3H), 1.88–1.93 (m, 3H), 1.70–1.75 (m, 3H), 1.11–1.22 (m, 2H).

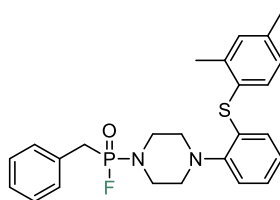
$^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 206.9, 155.3, 149.2, 148.3, 130.3 (d, $J = 8.3$ Hz), 129.3, 128.8, 128.4, 126.9, 107.1 (d, $J = 6.1$ Hz), 104.1 (d, $J = 10.5$ Hz), 55.8 (d, $J = 15.9$ Hz), 44.5, 43.8 (d, $J = 6.7$ Hz), 38.4, 33.8, 33.3, 32.4 (dd, $J = 131.7$ Hz, 38.5 Hz).

$^{31}\text{P NMR}$ (162 MHz, CDCl_3): δ 31.9 (d, $J = 1052$ Hz, 1P).

$^{19}\text{F NMR}$ (376 MHz, CDCl_3): δ -65.1 (dd, $J = 1052$ Hz, 25.5 Hz, 1F).

HRMS (m/z) calculated for $\text{C}_{24}\text{H}_{30}\text{FNO}_4\text{P}^+$ ($[\text{M}+\text{H}]^+$): 446.1891; found: 446.1897.

Benzyl(4-(2-((2,4-dimethylphenyl)thio)phenyl)piperazin-1-yl)phosphinic fluoride (44)



44

White solid, yield: 56%.

¹H NMR (400 MHz, CDCl₃): δ 7.30–7.39 (m, 6H), 7.15 (s, 1H), 7.02–7.08 (m, 2H), 6.95 (d, *J* = 8.0 Hz, 1H), 6.88 (t, *J* = 7.6 Hz, 1H), 3.24–3.43 (m, 6H), 2.92 (s, 4H), 2.33 (d, *J* = 25.6 Hz, 6H).

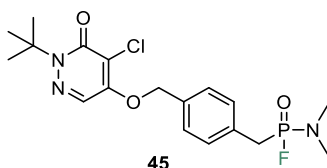
¹³C NMR (100 MHz, CDCl₃): δ 148.7, 142.3, 139.3, 136.1, 134.7, 131.7, 130.4 (d, *J* = 8.7 Hz), 129.7 (d, *J* = 6.8 Hz), 128.9 (d, *J* = 2.5 Hz), 127.8, 127.6, 127.4 (d, *J* = 3.2 Hz), 126.3, 125.5, 124.8, 120.0, 51.9 (d, *J* = 3.8 Hz), 44.3, 33.0 (dd, *J* = 130.4 Hz, 32 Hz), 21.2, 20.5.

³¹P NMR (162 MHz, CDCl₃): δ 31.6 (d, *J* = 1055 Hz, 1P).

¹⁹F NMR (376 MHz, CDCl₃): δ -64.6 (d, *J* = 1054 Hz, 1F).

HRMS (m/z) calculated for C₂₅H₂₉FN₂OPS⁺ ([M+H]⁺): 455.1717; found: 455.1713.

***P*-(4-(((1-(*tert*-butyl)-5-chloro-6-oxo-1,6-dihydropyridazin-4-yl)oxy)methyl)benzyl)-*N,N*-dimethylphosphonamidic fluoride (45)**



White solid, yield: 51%.

¹H NMR (400 MHz, CDCl₃): δ 7.71 (d, *J* = 4.5 Hz, 1H), 7.36 (dd, *J* = 3.3 Hz, 13.1 Hz, 4H), 5.30 (d, *J* = 2.4 Hz, 2H), 3.23–3.36 (m, 2H), 2.64–2.68 (m, 6H), 1.63 (d, *J* = 4.5 Hz, 9H).

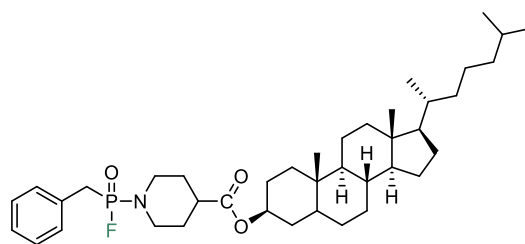
¹³C NMR (100 MHz, CDCl₃): δ 158.9, 153.6, 133.9 (d, *J* = 3.6 Hz), 131.3 (d, *J* = 8.8 Hz), 130.1 (d, *J* = 6.6 Hz), 127.6 (d, *J* = 2.8 Hz), 125.0, 118.3, 71.4, 66.4, 35.8 (d, *J* = 4.1 Hz), 32.2 (dd, *J* = 32.5 Hz, 130.5 Hz), 27.8 (s, 21H).

³¹P NMR (162 MHz, CDCl₃): δ 33.5 (d, *J* = 1051 Hz, 1P).

¹⁹F NMR (376 MHz, CDCl₃): δ -66.7 (d, *J* = 1051 Hz, 1F).

HRMS (m/z) calculated for C₁₈H₂₅ClFN₃O₃P⁺ ([M+H]⁺): 416.1301; found: 416.1306.

(3*S*,8*R*,9*S*,10*S*,13*R*,14*S*,17*R*)-10,13-Dimethyl-17-((*R*)-6-methylheptan-2-yl)hexadecahydro-1*H*-cyclopenta[*a*]phenanthren-3-yl 1-(benzylfluorophosphoryl)-piperidine-4-carboxylate (46)



46

White solid, yield: 50%.

¹H NMR (400 MHz, CDCl₃): δ 7.30–7.35 (m, 5H), 3.38–3.43 (m, 1H), 3.19–3.31 (m, 1H), 2.72–2.77 (m, 1H), 2.32–2.37 (m, 1H), 1.96 (d, *J* = 12 Hz, 1H), 1.71–1.81 (m, 4H), 1.64–1.67 (m, 2H), 1.43–1.58 (m, 8H), 1.22–1.38 (m, 14H), 1.04–1.15 (m, 6H), 0.94–0.99 (m, 3H), 0.85–0.90 (m, 10H), 0.81 (s, 3H), 0.64 (s, 3H).

¹³C NMR (100 MHz, CDCl₃): δ 173.6, 130.4 (d, *J* = 10.7 Hz), 129.6 (d, *J* = 6.8 Hz), 128.8 (d, *J* = 2.7 Hz), 127.4 (d, *J* = 3.4 Hz), 73.9, 56.3 (d, *J* = 13.5 Hz), 54.2, 44.6, 43.3, 42.6, 40.9, 39.9, 39.5, 36.7, 36.2, 35.8, 35.5, 34.0, 33.8, 33.5, 32.6, 32.2, 30.0, 29.7, 31.9, 28.6, 28.2 (d, *J* = 6.6 Hz), 28.0, 27.4, 24.2, 23.8, 22.7 (d, *J* = 25.3 Hz), 21.2, 18.7, 12.1 (d, *J* = 16.3 Hz).

³¹P NMR (162 MHz, CDCl₃): δ 31.7 (d, *J* = 1055 Hz, 1P).

¹⁹F NMR (376 MHz, CDCl₃): δ -65.4 (d, *J* = 1054 Hz, 1F).

HRMS (m/z) calculated for C₄₀H₆₄FNO₃P⁺ ([M+H]⁺): 656.4602; found: 656.4606.

2.3 Experimental Procedures for Radiochemistry

2.3.1 General procedure for the preparation of [¹⁸F]TBAF, [¹⁸F]CsF, [¹⁸F]KF/K₂₂₂, [¹⁸F]KF/18-cr-6

[¹⁸F]F⁻ was produced through the ¹⁸O(p, n)¹⁸F reaction using the IBA 18/9 cyclotron (Belgium) and was delivered in [¹⁸O]H₂O. The [¹⁸F] fluoride ([¹⁸F]F⁻) was extracted from the [¹⁸O]-enriched water using QMA and then introduced into a glass vial reactor with a pre-configured eluent (**Table S2**). The resulting solution was co-evaporated and dried at 110 °C, and this process was repeated three times using clean glass vials (300 μL acetonitrile × 3) for subsequent utilization.

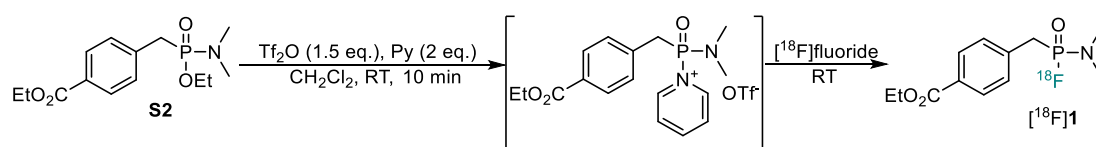
Table S2: Detailed formulas of eluent

Entry	[¹⁸ F]fluoride	Eluent
1	[¹⁸ F]TBAF	6.0 mg <i>n</i> Bu ₄ N ⁺ OH ⁻ in 100 μL CH ₃ CN
2	[¹⁸ F]CsF	7.5 mg Cs ₂ CO ₃ in 100 μL CH ₃ CN
3	[¹⁸ F]KF/K ₂₂₂	8.0 mg K ₂₂₂ , 1.0 mg K ₂ CO ₃ in CH ₃ CN/H ₂ O (4/1, v/v, 0.5 mL)
4	[¹⁸ F]KF/18-cr-6	6.0 mg 18-crown-6, 1.0 mg K ₂ CO ₃ in CH ₃ CN/H ₂ O (4/1, v/v, 0.5 mL)

2.3.2 Optimization studies for alkyl phosphonates ¹⁸F-fluorination

The reaction conditions for ¹⁸F-fluorination were reevaluated in comparison to fluorination. Initially, the conditions were kept consistent with those used for non-radioactive in order to screen suitable [¹⁸F]fluoride. The precursor **S2** underwent conversion into a pyridinium salt intermediate within a 10 min period. Subsequently, the intermediate was introduced into a glass vial reactor, where a previously prepared dry 37–55.5 MBq [¹⁸F]fluoride was present, and the mixture was incubated at RT with continuous shaking for 5–10 min. After the completion of the reaction, 9900 μL of water was added to quench the reaction and radiochemical conversion (RCC) was assessed using radio-TLC by dividing the area under the curve (AUC) of the radioactive peak of interest by the total AUC of all radioactive peaks. The radiochemical yield (RCY) is based on the ratio of the final isolated product to that of the initial radioactivity, with all amounts decay corrected to the same time¹¹. In all ¹⁸F-labeling reaction systems, due to the presence of acidic and water-soluble impurities in the reaction mixture, a simple purification step using a C18 cartridge is typically performed prior to HPLC analysis to remove these impurities.

Table S3: Optimization for alkyl phosphonates ¹⁸F-fluorination



Entry	Precursor load /μmol	[¹⁸ F]fluoride	Reaction time /min	RCC _{TLC} (%) ^[a]
1	20	[¹⁸ F]TBAF	5	10 ± 5
2	20	[¹⁸ F]CsF	5	37 ± 1
3	20	[¹⁸ F]KF/K ₂₂₂	5	58 ± 5
4	20	[¹⁸ F]KF/18-cr-6	5	5 ± 1
5	20	[¹⁸ F]KF/K ₂₂₂	10	78 ± 5
6	2	[¹⁸ F]KF/K ₂₂₂	10	67 ± 5
7	1	[¹⁸ F]KF/K ₂₂₂	10	62 ± 3
8	0.2	[¹⁸ F]KF/K ₂₂₂	10	16 ± 6
9	0.02	[¹⁸ F]KF/K ₂₂₂	10	5 ± 2

[a] RCC_{TLC}s determined by radio-TLC.

2.3.3 RadioTLC and radio-HPLC analysis of ^{18}F -labeled compounds

All ^{18}F -labeling reactions were performed following the specified protocol: 3 μmol of precursor were dissolved in 300 μL CH_2Cl_2 . Then, 2 eq. of Tf_2O were added for 5 min, followed by the addition of 1.5 eq. of pyridine for another 5 min. The resulting intermediate solution was subsequently divided into three equal portions and introduced into separate glass vial reactors, each containing pre-prepared 37–55.5 MBq dried $^{18}\text{F}[\text{Kf}/\text{K}_{222}]$. The reactors were continuously oscillated at room temperature for 10 minutes. Upon completion of the reaction, 9900 μL of water was added to quench the reaction. The RCCs of the reaction were determined using radio-TLC with methanol as the developing agent ($n = 3$). Additionally, the RCY and co-injection were measured using a radio-HPLC.

S15, as a precursor, undergoes highly selective ^{18}F -fluorination to yield an unstable phenyl benzylphosphonofluoridate, which, as reported in the literature¹², is prone to hydrolysis and cannot be isolated due to its instability, spontaneously converting into benzylphosphonofluoridic acid. The results demonstrated a high selectivity for the alkyl phosphonate esters, aligning with our expectations.

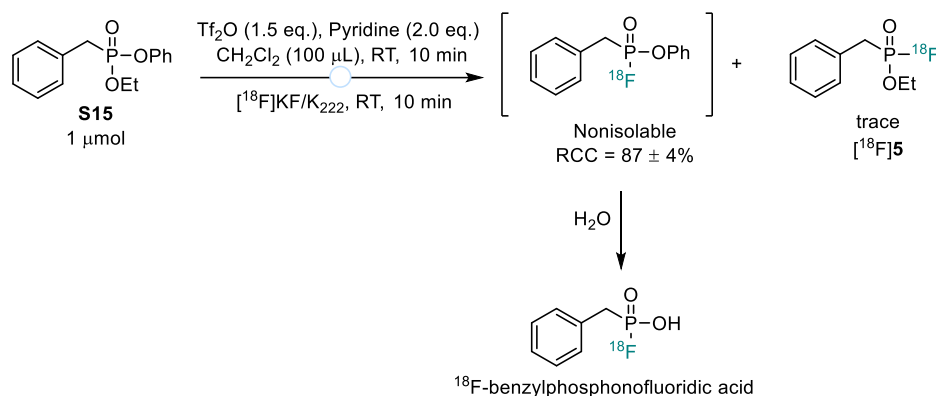


Fig. S4: The selectivity of the reaction was verified by ^{18}F -fluorination.

Table S4: Average radiochemical conversion for ^{18}F -benzylphosphonofluoridic acid

Run	1	2	3	Average
RCC _{TLC} (%)	77	76	80	78 ± 2

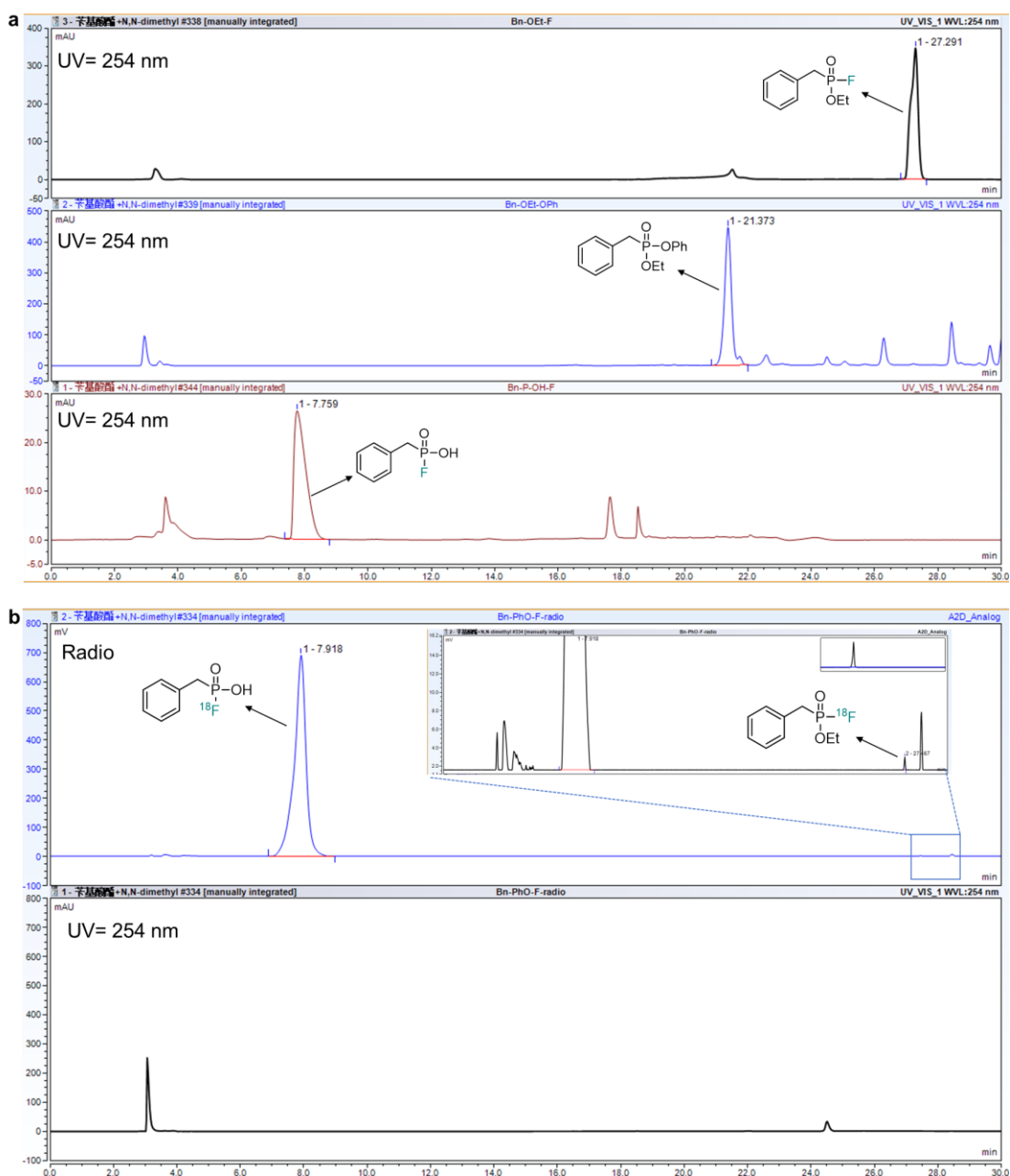


Fig. S5: **a.** HPLC traces of precursor and reference compounds; **b.** HPLC traces of crude ^{18}F -labeled products. 1# HPLC conditions; NanoChrom C18 column ($5\ \mu\text{m}$, $4.6 \times 250\ \text{mm}$); isocratic elution, 0–10 min, 10% of CH_3CN and 90% of water containing 0.1%TFA, 10–30 min, from 10% of CH_3CN and 90% of water containing 0.1%TFA to 70% of CH_3CN and 30% of water containing 0.1%TFA; flow rate: 1.0 mL/min.

Preparation of ^{18}F 1

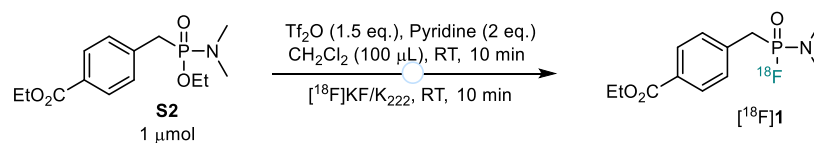


Table S5: Average radiochemical conversion for [^{18}F]1

Run	1	2	3	Average
RCC _{TLC} (%)	63	66	59	62 ± 3

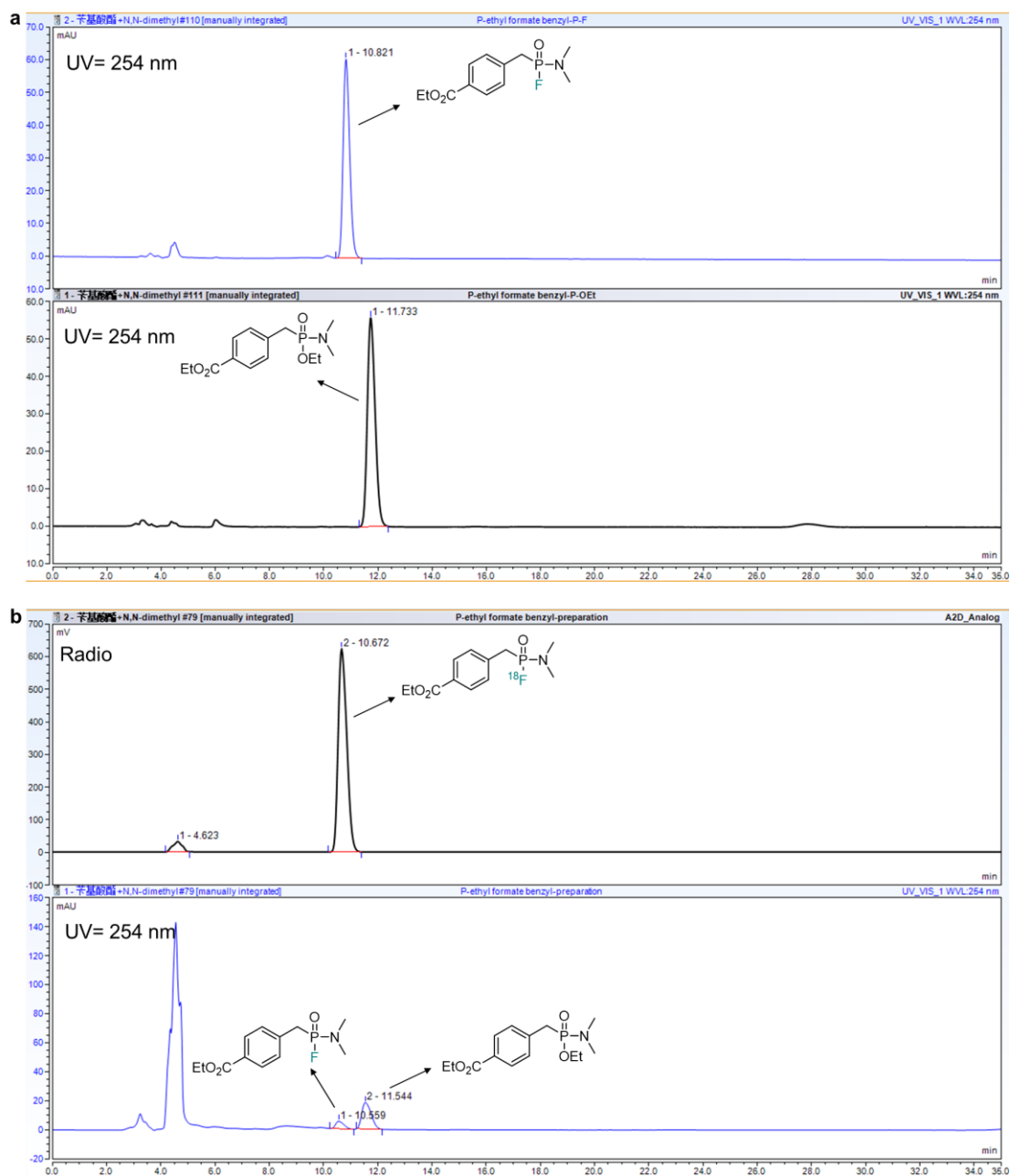


Fig. S6: **a.** HPLC traces of precursor and reference compounds; **b.** HPLC traces of crude ^{18}F -labeled products. 1# HPLC conditions; SEP Basic-C18 semi-preparative column (120A $5\mu\text{m}$ 10×250 mm); isocratic elution, 0–10 min, from 50% of CH_3CN and 50% of water containing 0.1%TFA to 40% of CH_3CN and 60% of water containing 0.1%TFA; 10–25 min, from 40% of CH_3CN and 60% of water containing 0.1%TFA to 35% of CH_3CN and 65%

of water containing 0.1%TFA; 25–35 min, 35% of CH₃CN and 65% of water containing 0.1%TFA; flow rate: 3.0 mL/min.

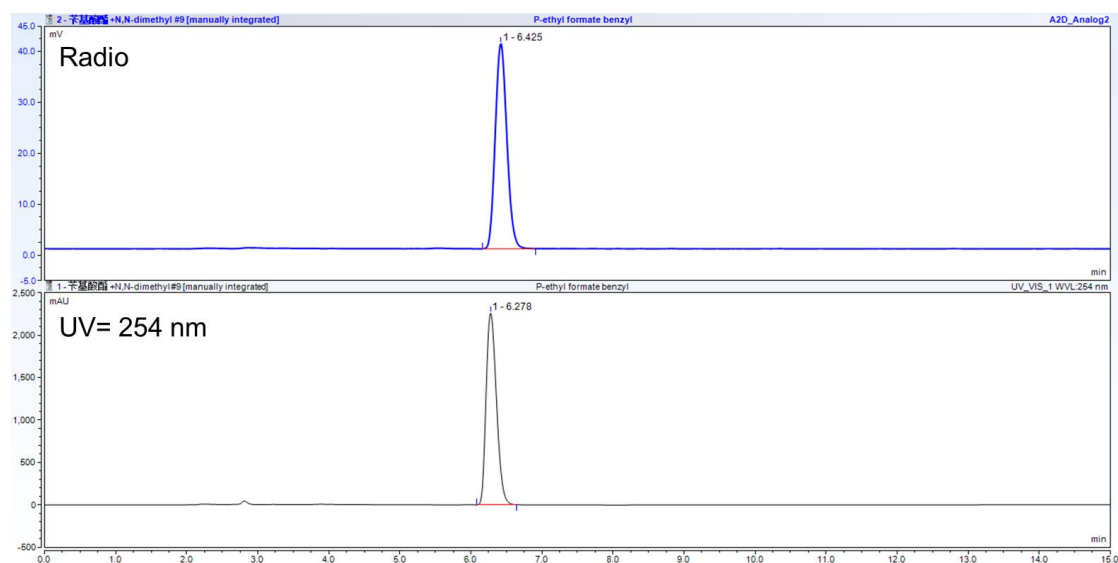


Fig. S7: Co-injection for [¹⁸F]**1**. 1# HPLC conditions; NanoChrom C18 column (5 μm, 4.6 × 250 mm); isocratic elution, H₂O/CH₃CN = 50/50, and the flow rate was 1.0 mL/min for 15 min. The RCP was > 99%. (The HPLC trace of the co-injection represents the results of the purified ¹⁸F-product mixed with the corresponding reference compound, allowing confirmation of radiochemical purity (RCP) from the co-injection HPLC trace.)

Preparation of [¹⁸F]**36**

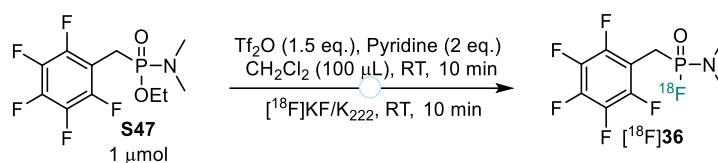


Table S6: Average radiochemical conversion for [¹⁸F]**36**

Run	1	2	3	Average
RCC _{TLC} (%)	75	84	69	76 ± 7

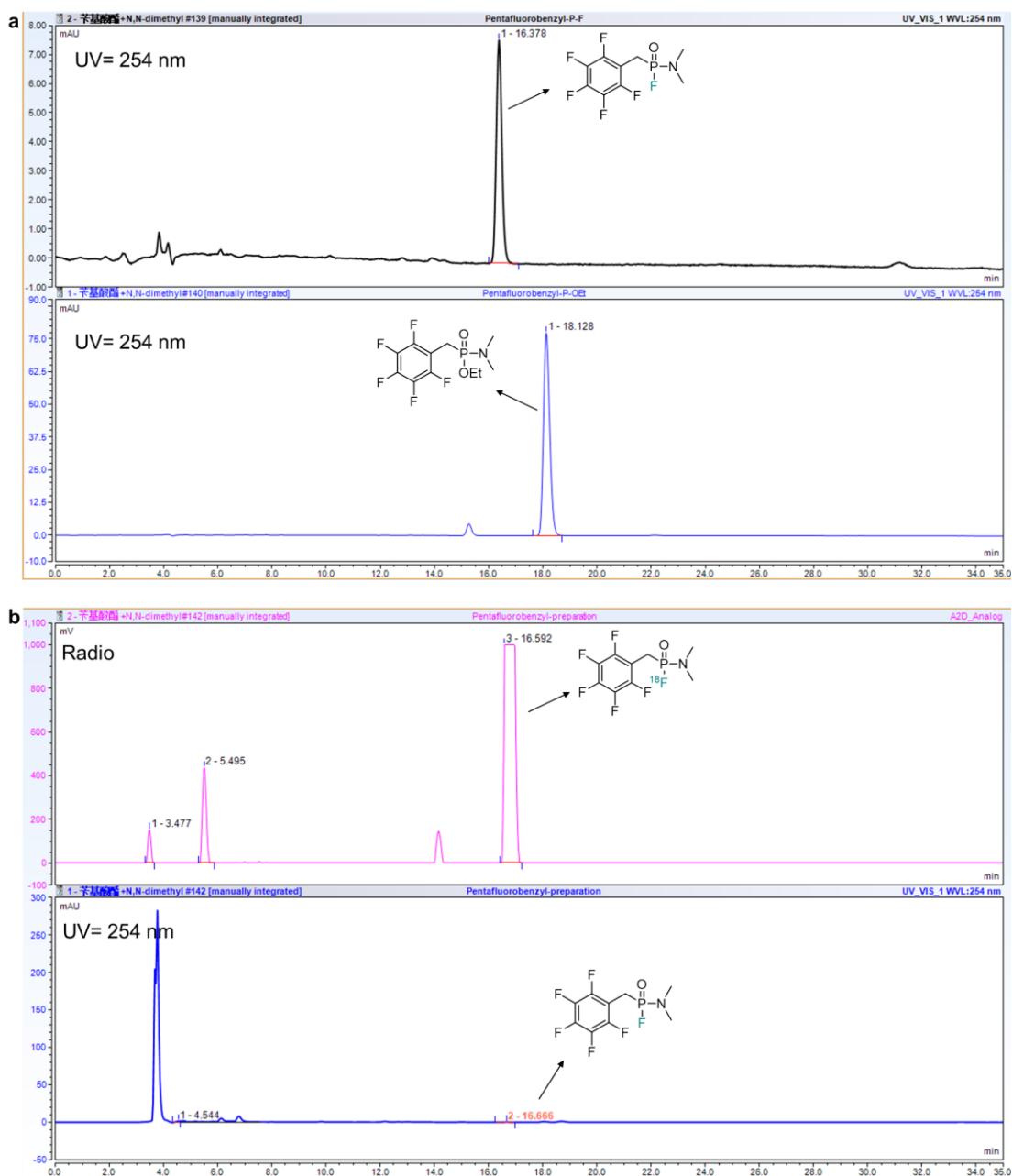


Fig. S8: **a.** HPLC traces of precursor and reference compounds; **b.** HPLC traces of crude ¹⁸F-labeled products. 1# HPLC conditions; SEP Basic-C18 semi-preparative column (120A 5 μ m 10 \times 250mm); isocratic elution, 0–10 min, from 40% of CH₃CN and 60% of water containing 0.1%TFA to 50% of CH₃CN and 50% of water containing 0.1%TFA; 10–20 min, 50% of CH₃CN and 50% of water containing 0.1%TFA; 20–35 min, from 50% of CH₃CN and 50% of water containing 0.1%TFA to 55% of CH₃CN and 45% of water containing 0.1%TFA; flow rate: 3.0 mL/min.

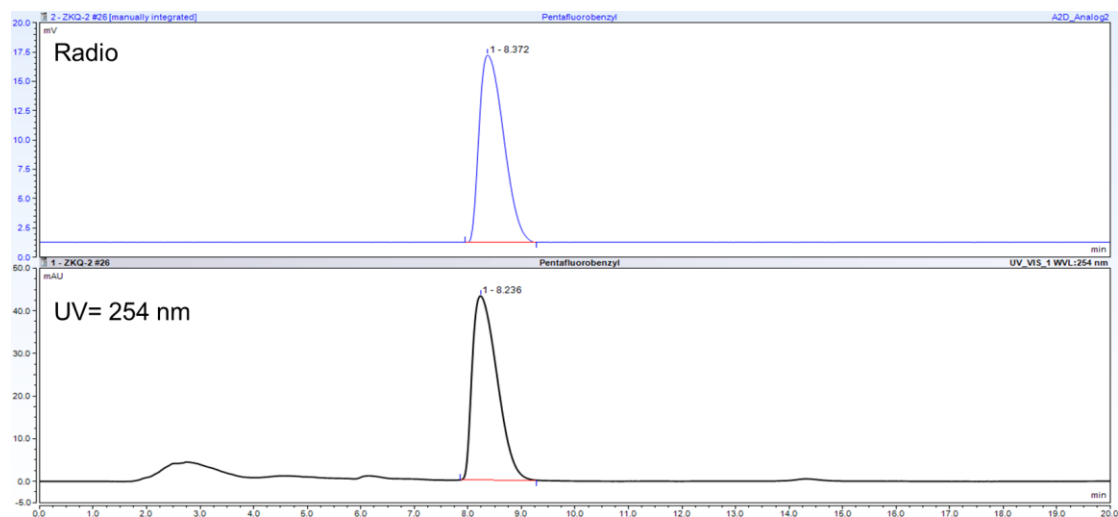


Fig. S9: Co-injection for [^{18}F]36. 1# HPLC condition; NanoChrom C18 column (5 μm , 4.6 \times 250 mm); isocratic elution, $\text{H}_2\text{O}/\text{CH}_3\text{CN} = 50/50$, and the flow rate was 1.0 mL/min for 20 min. The RCP was > 99%.

Preparation of [^{18}F]37

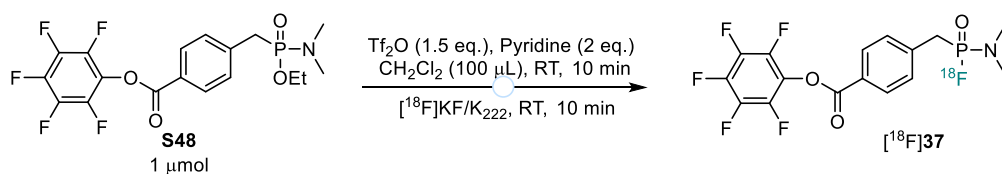


Table S7: Average radiochemical conversion for [^{18}F]37

Run	1	2	3	Average
RCC _{TLC} (%)	70	67	77	71 \pm 5

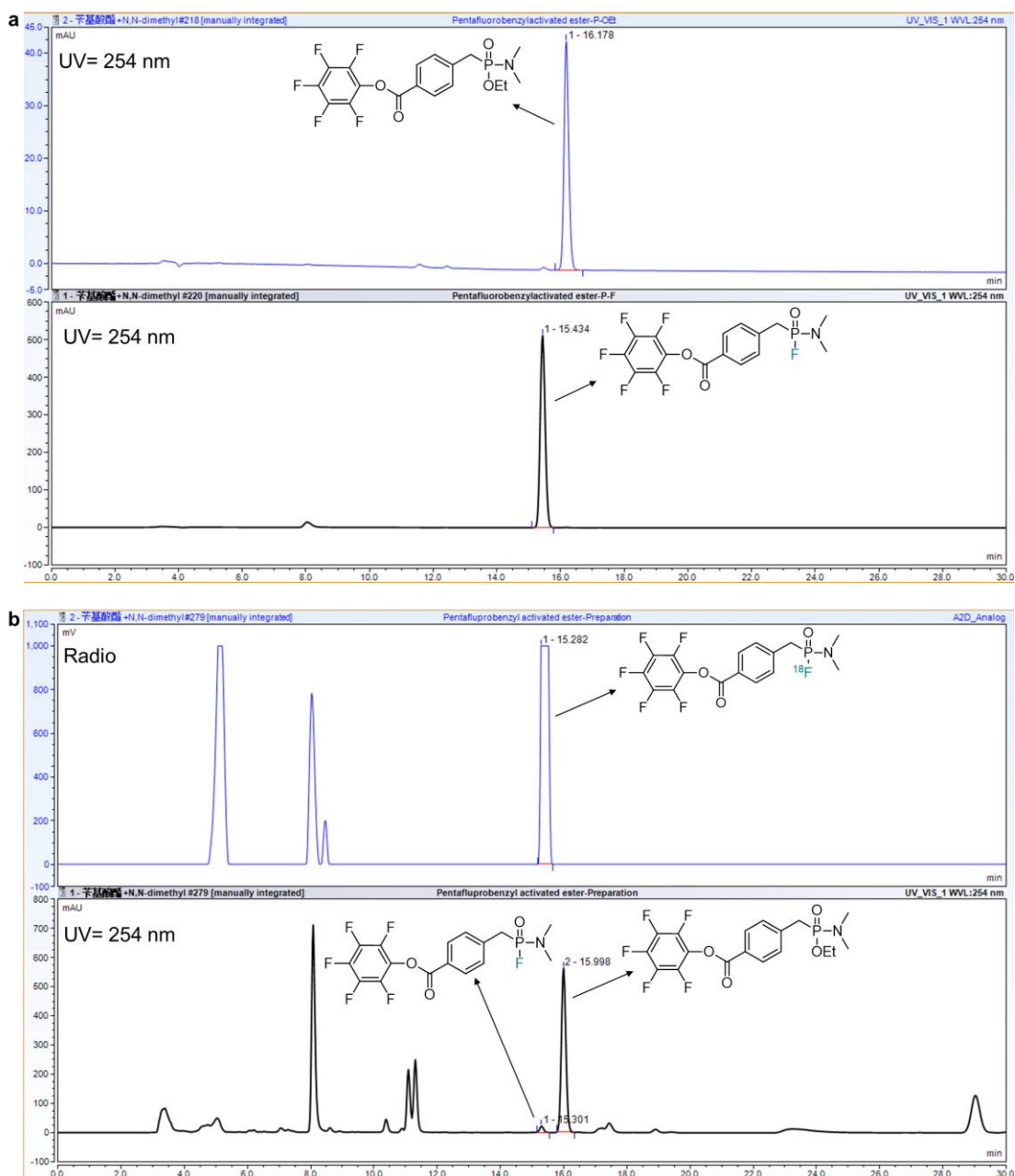


Fig. S10: **a.** HPLC traces of precursor and reference compounds; **b.** HPLC traces of crude ^{18}F -labeled products. 1# HPLC conditions; SEP Basic-C18 semi-preparative column (120A $5\mu\text{m}$ $10 \times 250\text{mm}$); isocratic elution, 0–10 min, from 50% of CH_3CN and 50% of water containing 0.1%TFA to 70% of CH_3CN and 30% of water containing 0.1%TFA; 10–30 min, from 70% of CH_3CN and 30% of water containing 0.1%TFA to 75% of CH_3CN and 25% of water containing 0.1%TFA; flow rate: 3.0 mL/min.

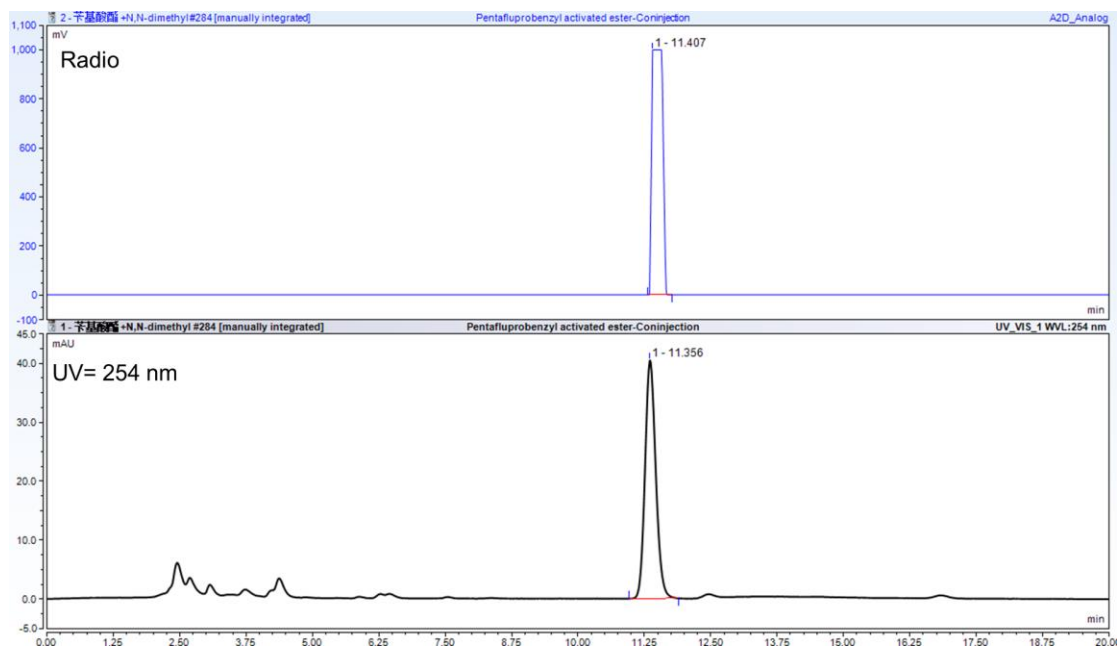


Fig. S11: Co-injection for $[^{18}\text{F}]\mathbf{37}$. 2# HPLC condition; NanoChrom C18 column (5 μm , 4.6×250 mm), isocratic elution, water containing 0.1%TFA/ $\text{CH}_3\text{CN} = 40/60$, and the flow rate was 1.0 mL/min for 20 min. The RCP was > 99%.

Preparation of $[^{18}\text{F}]\mathbf{38}$

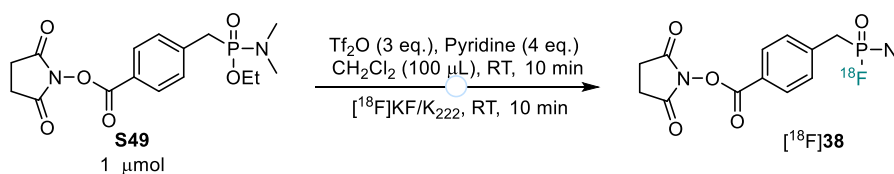


Table S8: Average radiochemical conversion for $[^{18}\text{F}]\mathbf{38}$

Run	1	2	3	Average
RCC _{TLC} (%)	90	88	85	88 ± 3

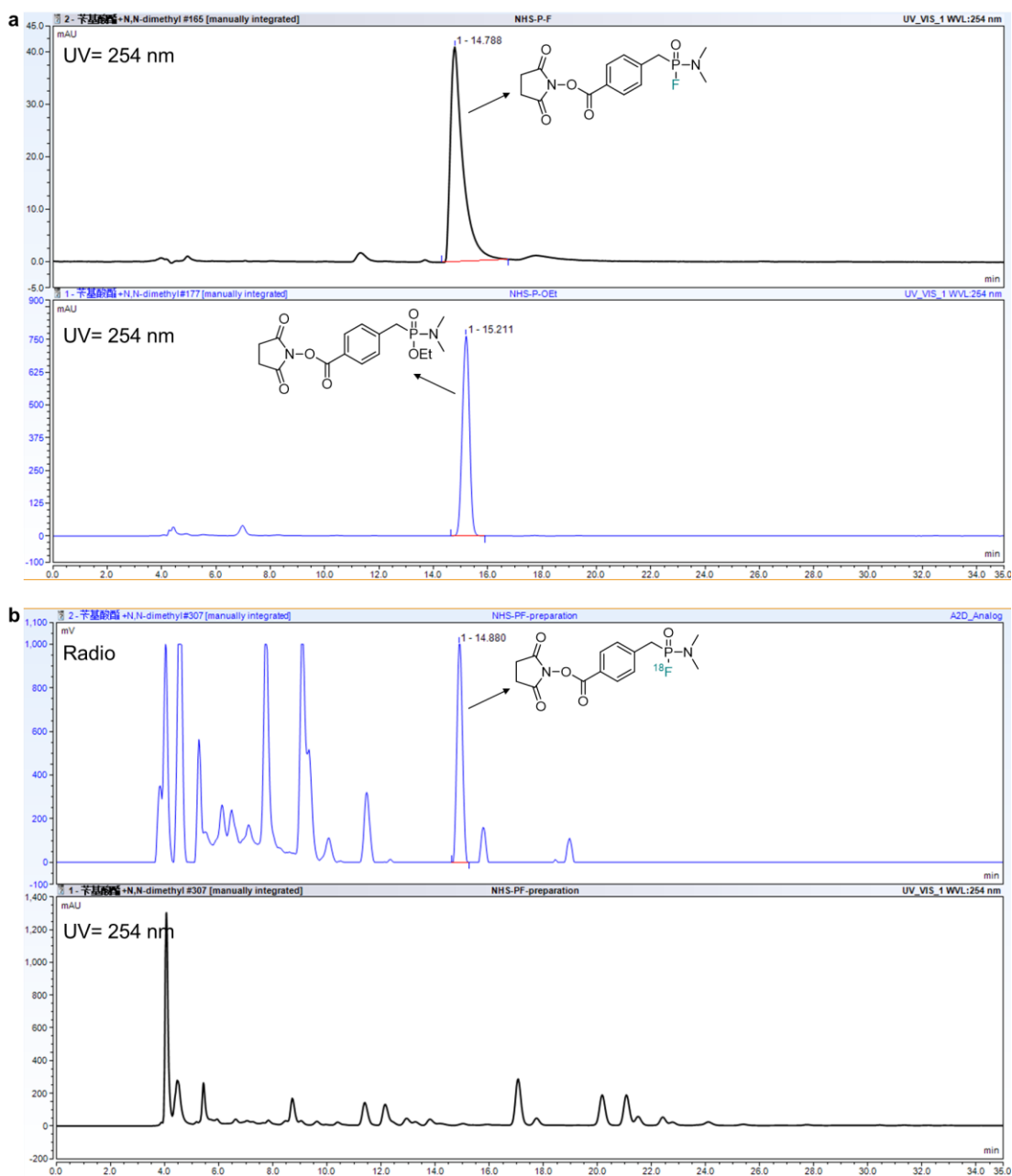


Fig. S12: a. HPLC traces of precursor and reference compounds; **b.** HPLC traces of crude ^{18}F -labeled products. 1# HPLC conditions; SEP Basic-C18 semi-preparative column (120A $5\mu\text{m}$ $10 \times 250\text{mm}$); isocratic elution, 0–20 min, from 30% of CH_3CN and 70% of water containing 0.1%TFA to 40% of CH_3CN and 60% of water containing 0.1%TFA; from 40% of CH_3CN and 60% of water containing 0.1%TFA to 50% of CH_3CN and 50% of water containing 0.1%TFA; flow rate: 3.0 mL/min.

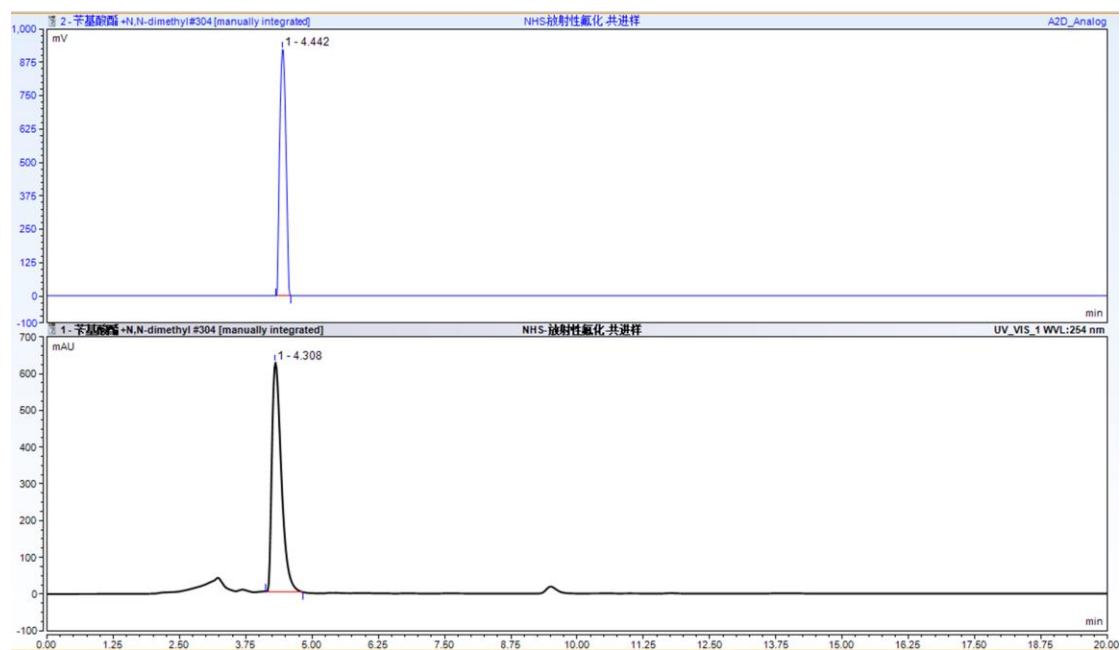


Fig. S13: Co-injection for $[^{18}\text{F}]\mathbf{38}$. 1# HPLC condition; NanoChrom C18 column ($5\ \mu\text{m}$, $4.6 \times 250\ \text{mm}$), isocratic elution, water containing 0.1%TFA/ $\text{CH}_3\text{CN} = 45/55$, and the flow rate was 1 mL/min for 20 min. The RCP was $> 99\%$.

Preparation of $[^{18}\text{F}]\mathbf{39}$

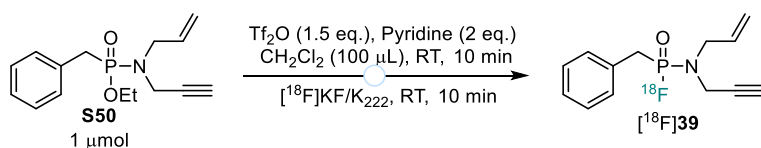


Table S9. Average radiochemical conversion for $[^{18}\text{F}]\mathbf{39}$

Run	1	2	3	Average
RCC _{TLC} (%)	73	85	72	77 ± 7

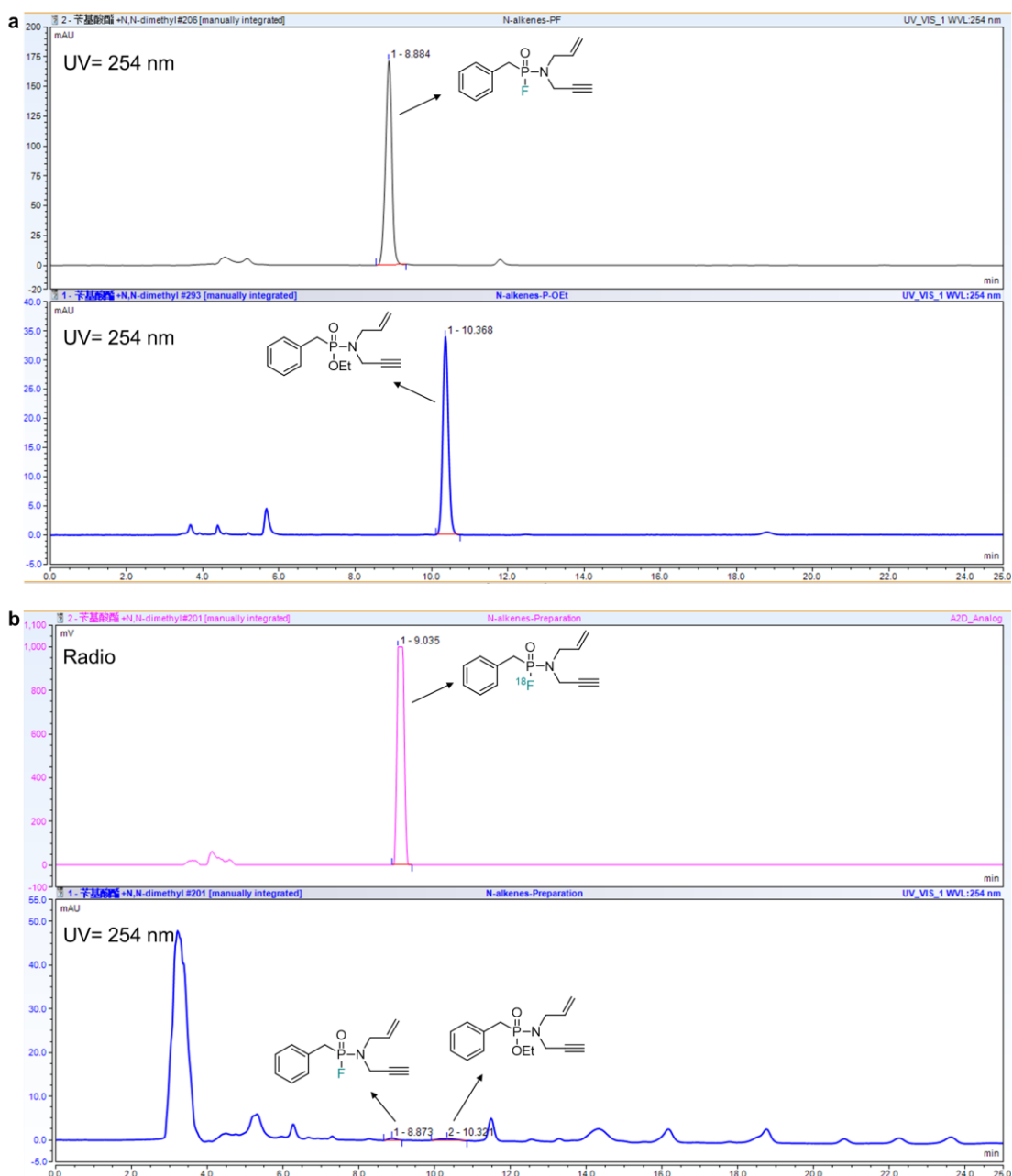


Fig. S14: **a.** HPLC traces of precursor and reference compounds; **b.** HPLC traces of crude ^{18}F -labeled products. 1# HPLC conditions; SEP Basic-C18 semi-preparative column (120A $5\mu\text{m}$ $10 \times 250\text{mm}$); isocratic elution, 0–10 min, from 50% of CH_3CN and 50% of water containing 0.1%TFA to 65% of CH_3CN and 35% of water containing 0.1%TFA; from 65% of CH_3CN and 35% of water containing 0.1%TFA to 70% of CH_3CN and 30% of water containing 0.1%TFA; flow rate: 3.0 mL/min.

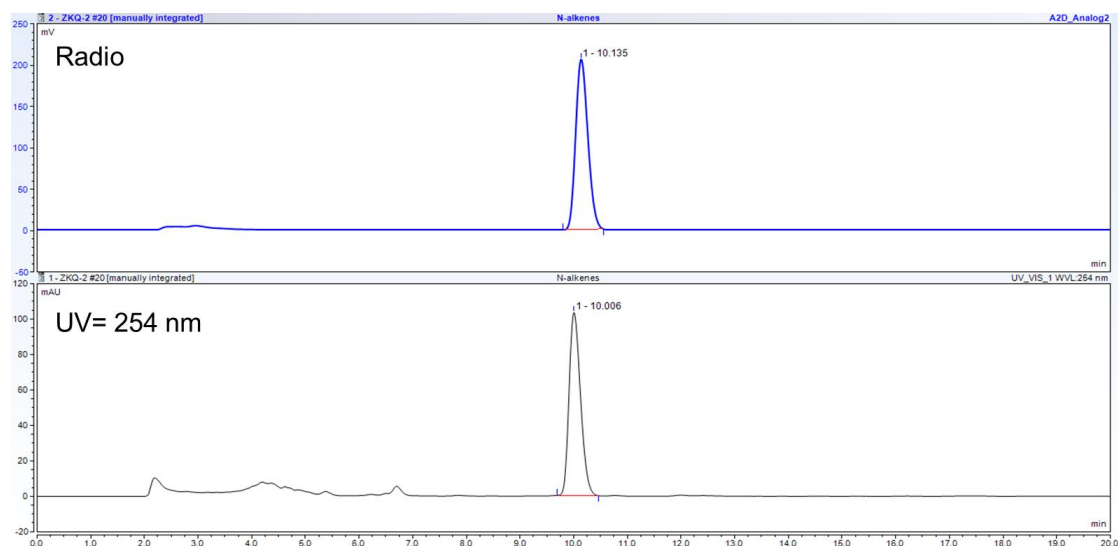


Fig. S15: Co-injection for $[^{18}\text{F}]\mathbf{39}$. 1# HPLC condition; NanoChrom C18 column (5 μm , 4.6×250 mm), isocratic elution, water containing 0.1%TFA/ $\text{CH}_3\text{CN} = 50/50$, and the flow rate was 1 mL/min for 20 min. The RCP was $>97\%$.

Preparation of $[^{18}\text{F}]\mathbf{40}$

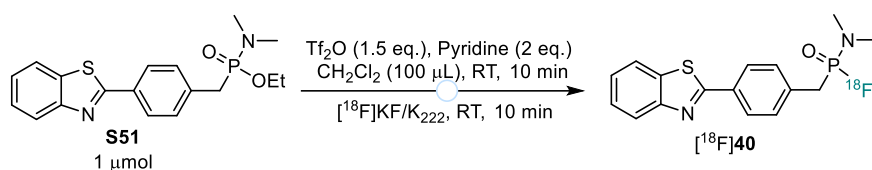


Table S10. Average radiochemical conversion for $[^{18}\text{F}]\mathbf{40}$

Run	1	2	3	Average
RCC _{TLC} (%)	61	59	55	58 ± 3

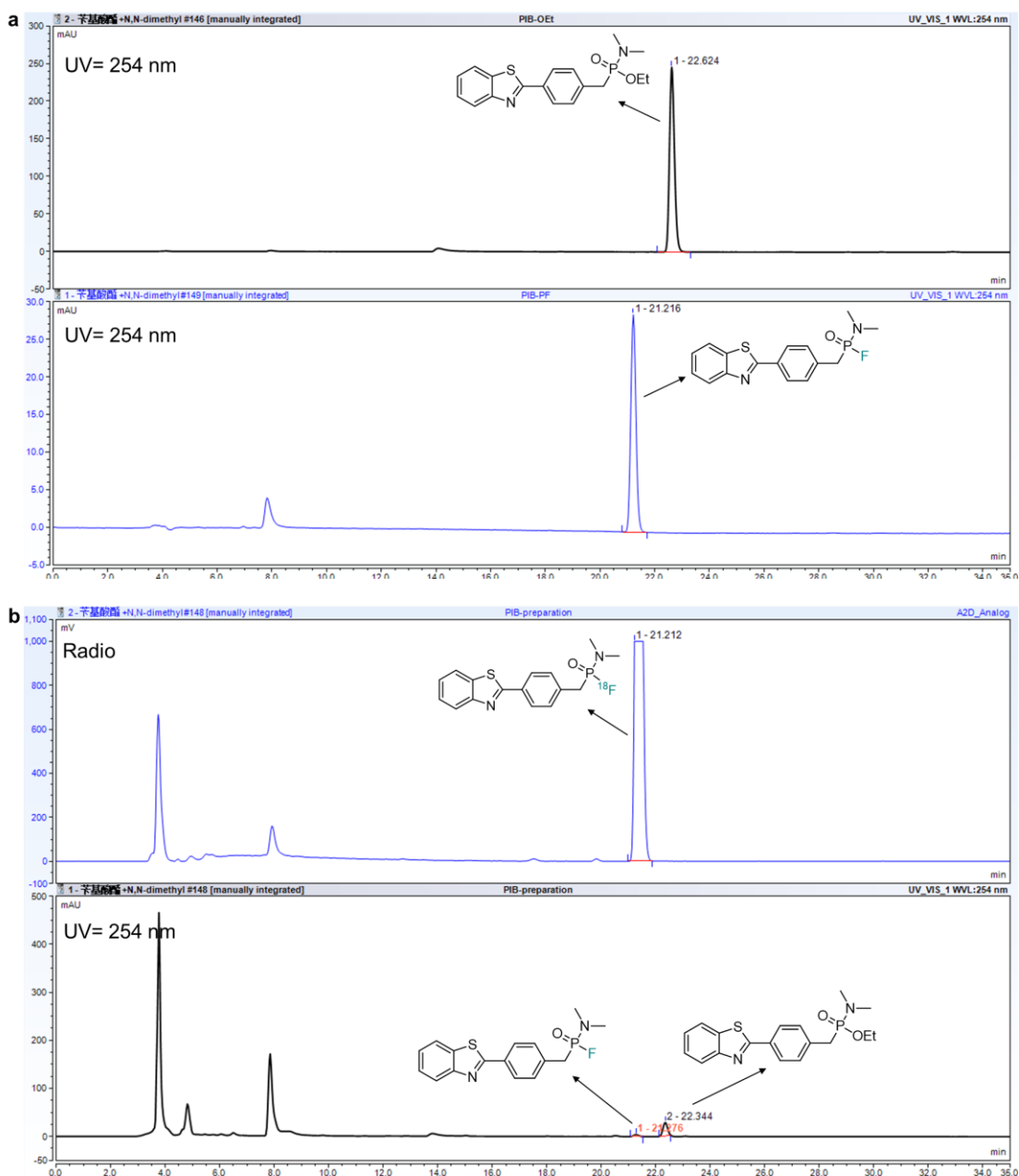


Fig. S16: **a.** HPLC traces of precursor and reference compounds; **b.** HPLC traces of crude ^{18}F -labeled products. 1# HPLC conditions; SEP Basic-C18 semi-preparative column (120A $5\mu\text{m}$ $10 \times 250\text{mm}$); isocratic elution, 0–10 min, from 40% of CH_3CN and 60% of water containing 0.1%TFA to 50% of CH_3CN and 50% of water containing 0.1%TFA; 10–15 min, from 50% of CH_3CN and 50% of water containing 0.1%TFA to 55% of CH_3CN and 45% of water containing 0.1%TFA; 15–18 min, from 55% of CH_3CN and 45% of water containing 0.1%TFA to 60% of CH_3CN and 40% of water containing 0.1%TFA; 18–35 min, 60% of CH_3CN and 40% of water containing 0.1%TFA; flow rate: 3.0 mL/min.

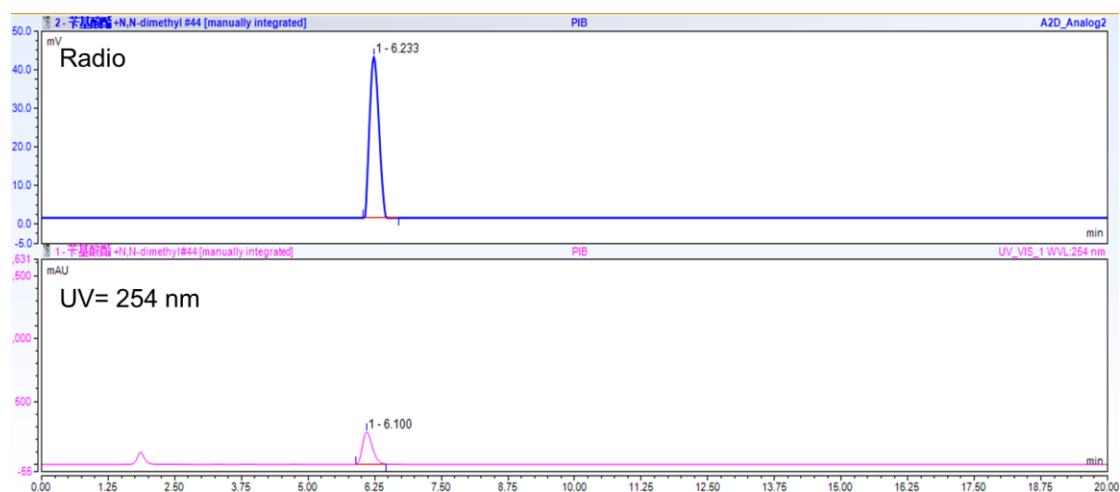


Fig. S17: Co-injection for [^{18}F]40. 1# HPLC condition; NanoChrom C18 column (5 μm , 4.6 \times 250 mm), isocratic elution, $\text{H}_2\text{O}/\text{CH}_3\text{CN} = 50/50$, and the flow rate was 1 mL/min for 20 min. The RCP was >99%.

Preparation of [^{18}F]41

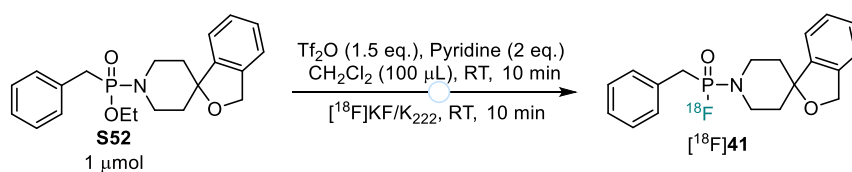


Table S11: Average radiochemical conversion for [^{18}F]41

Run	1	2	3	Average
RCC _{TLC} (%)	38	58	57	51 \pm 11

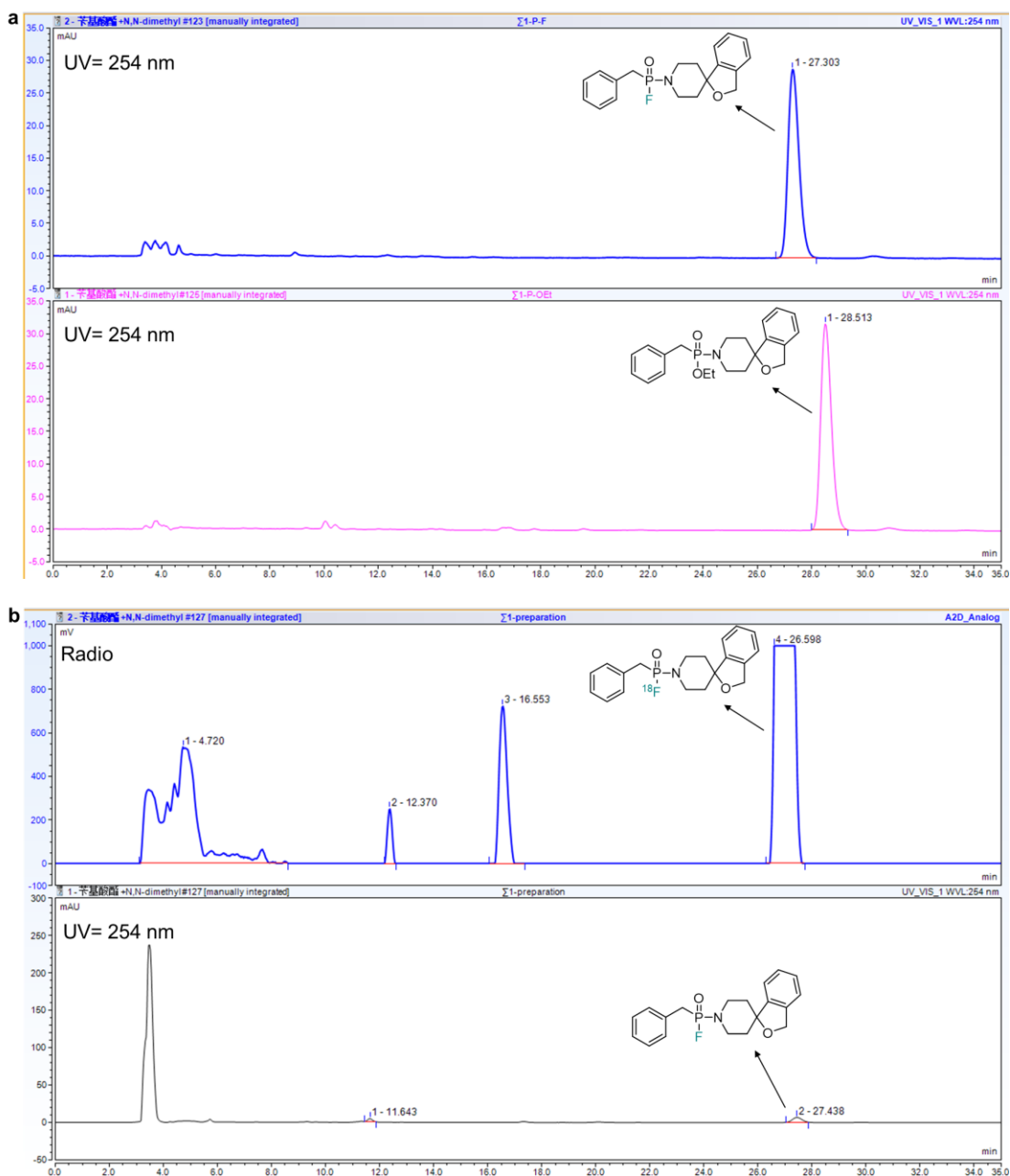


Fig. S18: **a.** HPLC traces of precursor and reference compounds; **b.** HPLC traces of crude ^{18}F -labeled products. 1# HPLC conditions; SEP Basic-C18 semi-preparative column (120A $5\mu\text{m}$ $10 \times 250\text{mm}$); isocratic elution, 0–10 min, from 40% of CH_3CN and 60% of water containing 0.1%TFA to 50% of CH_3CN and 50% of water containing 0.1%TFA; 10–20 min, 50% of CH_3CN and 50% of water containing 0.1%TFA for 10 min; 20–35 min, from 50% of CH_3CN and 50% of water containing 0.1%TFA to 55% of CH_3CN and 45% of water containing 0.1%TFA; flow rate: 3.0 mL/min.

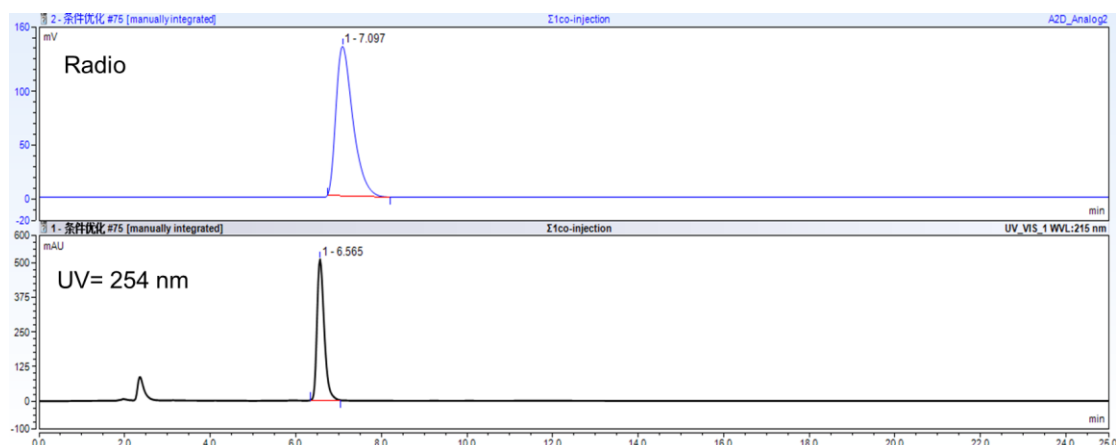


Fig. S19: Co-injection for $[^{18}\text{F}]\mathbf{41}$. 2# HPLC condition; NanoChrom C18 column (5 μm , 4.6×250 mm), isocratic elution, $\text{H}_2\text{O}/\text{CH}_3\text{CN} = 35/65$, and the flow rate was 1 mL/min for 25 min. The RCP was $> 99\%$.

Preparation of $[^{18}\text{F}]\mathbf{42}$

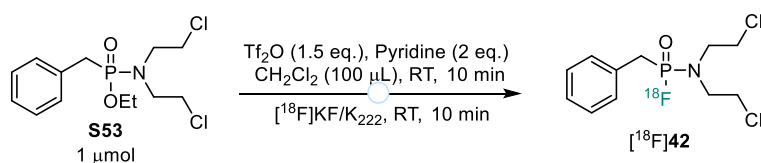
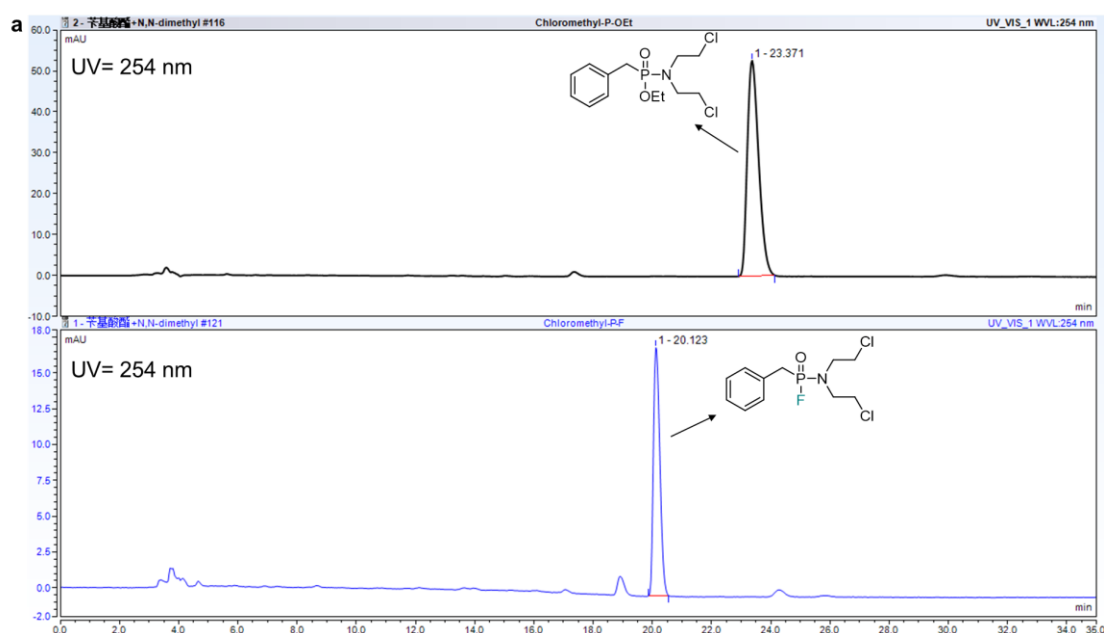


Table S12: Average radiochemical conversion for $[^{18}\text{F}]\mathbf{42}$

Run	1	2	3	Average
RCC _{TLC} (%)	86	62	77	75 ± 12



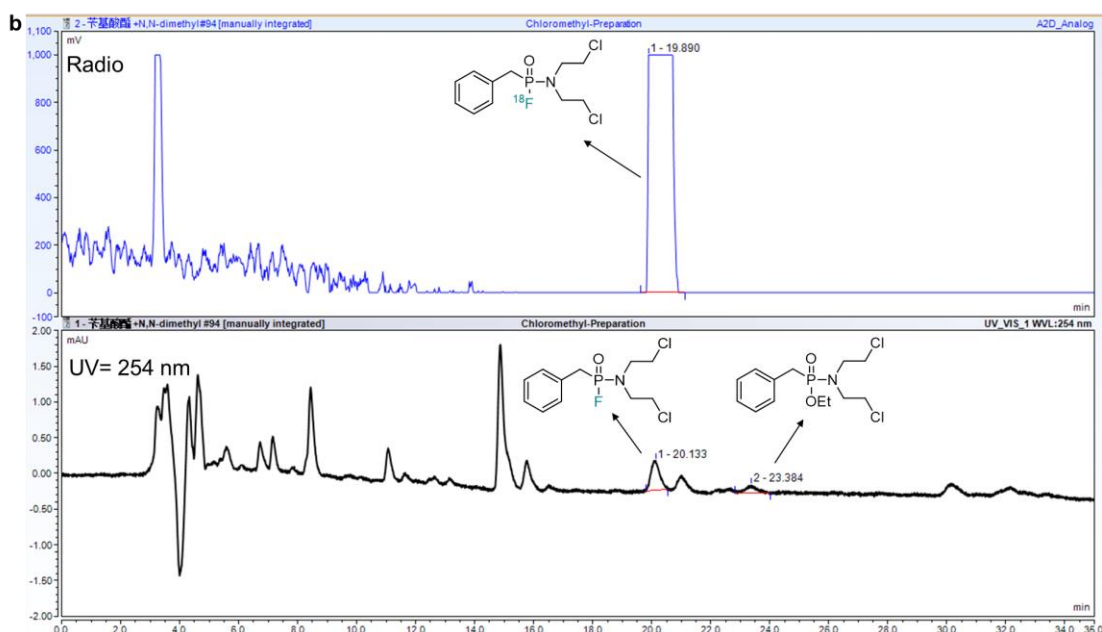


Fig. S20: a. HPLC traces of precursor and reference compounds; b. HPLC traces of crude ^{18}F -labeled products. 1# HPLC conditions; SEP Basic-C18 semi-preparative column (120A $5\mu\text{m}$ $10 \times 250\text{mm}$); isocratic elution, 0–10 min, from 40% of CH_3CN and 60% of water containing 0.1%TFA to 50% of CH_3CN and 50% of water containing 0.1%TFA; 10–20 min, 50% of CH_3CN and 50% of water containing 0.1%TFA; 20–35 min, from 50% of CH_3CN and 50% of water containing 0.1%TFA to 55% of CH_3CN and 45% of water containing 0.1%TFA; flow rate: 3.0 mL/min.

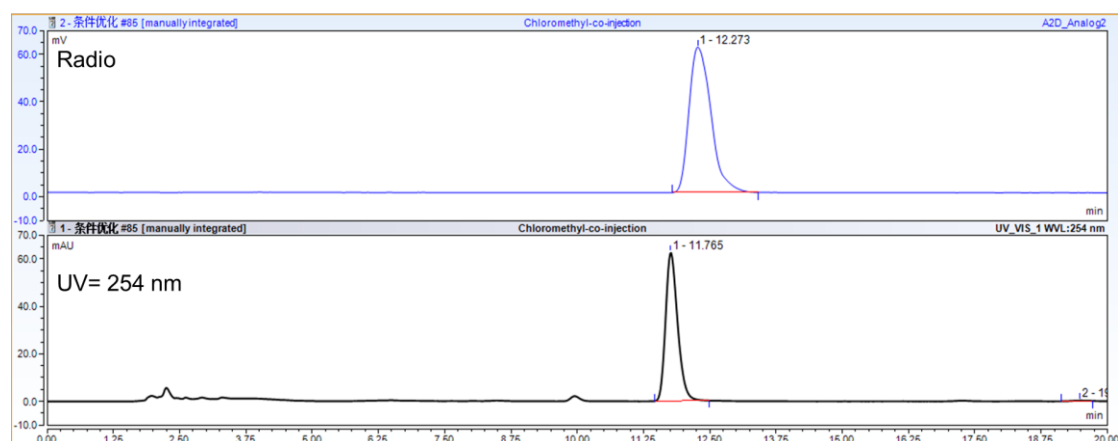


Fig. S21: Co-injection for ^{18}F 42. 2# HPLC condition: NanoChrom C18 column ($5\mu\text{m}$, $4.6 \times 250\text{mm}$), isocratic elution, $\text{H}_2\text{O}/\text{CH}_3\text{CN} = 50/50$, and the flow rate was 1 mL/min for 20 min. The RCP was $> 99\%$.

Preparation of [¹⁸F]43

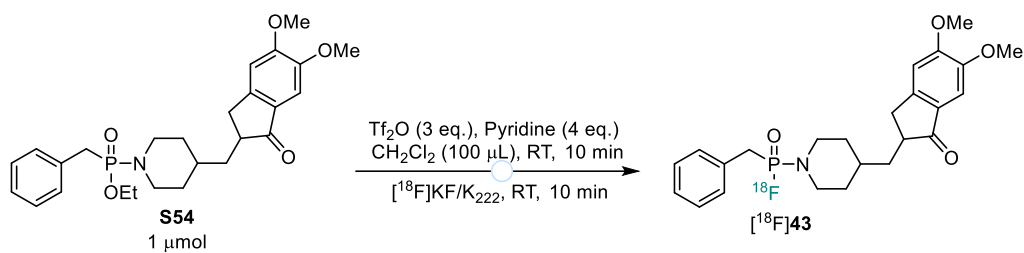
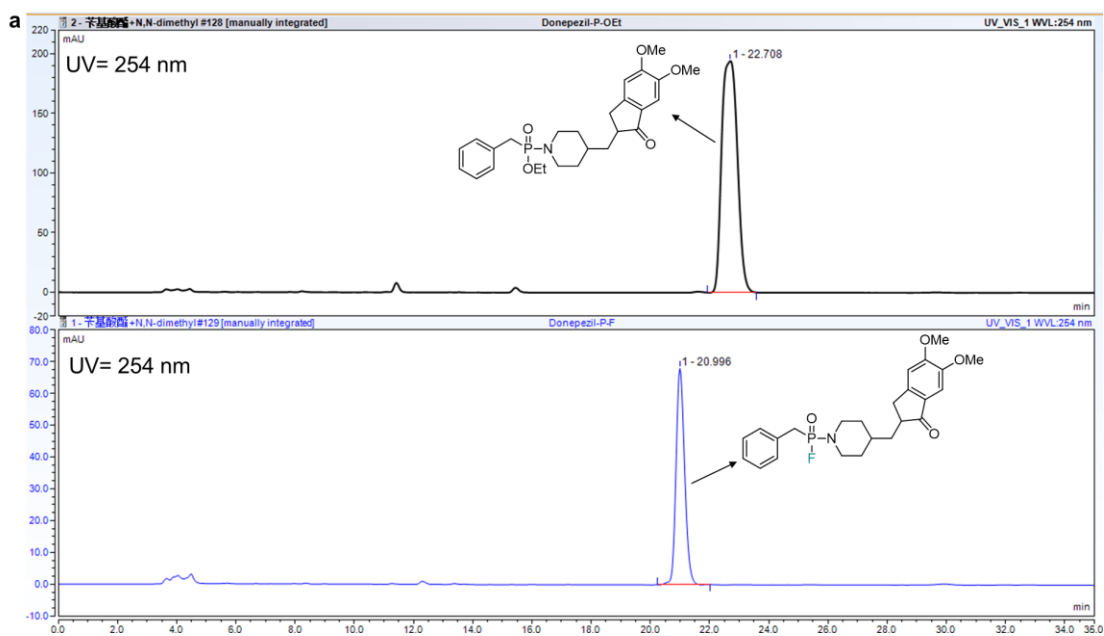


Table S13: Average radiochemical conversion for [¹⁸F]43

Run	1	2	3	Average
RCC _{TLC} (%)	63	57	64	61 ± 4



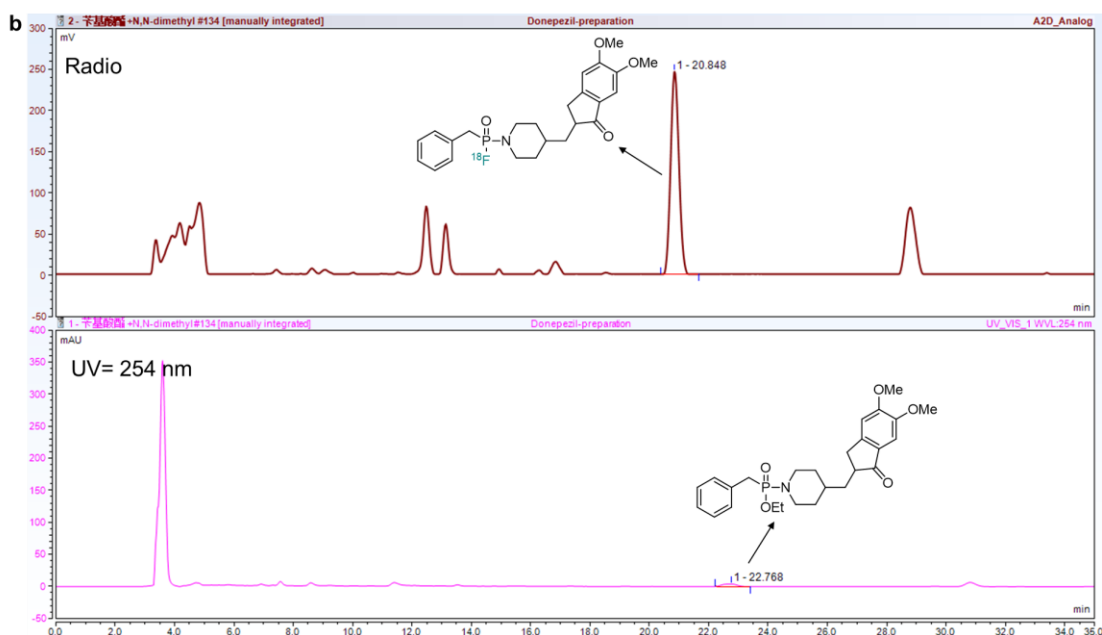


Fig. S22: **a.** HPLC traces of precursor and reference compounds; **b.** HPLC traces of crude ^{18}F -labeled products. 1# HPLC conditions; SEP Basic-C18 semi-preparative column (120A $5\mu\text{m}$ $10 \times 250\text{mm}$); isocratic elution, 0–10 min, from 40% of CH_3CN and 60% of water containing 0.1%TFA to 50% of CH_3CN and 50% of water containing 0.1%TFA; 10–20 min, 50% of CH_3CN and 50% of water containing 0.1%TFA; 20–35 min, from 50% of CH_3CN and 50% of water containing 0.1%TFA to 55% of CH_3CN and 45% of water containing 0.1%TFA; flow rate: 3.0 mL/min.

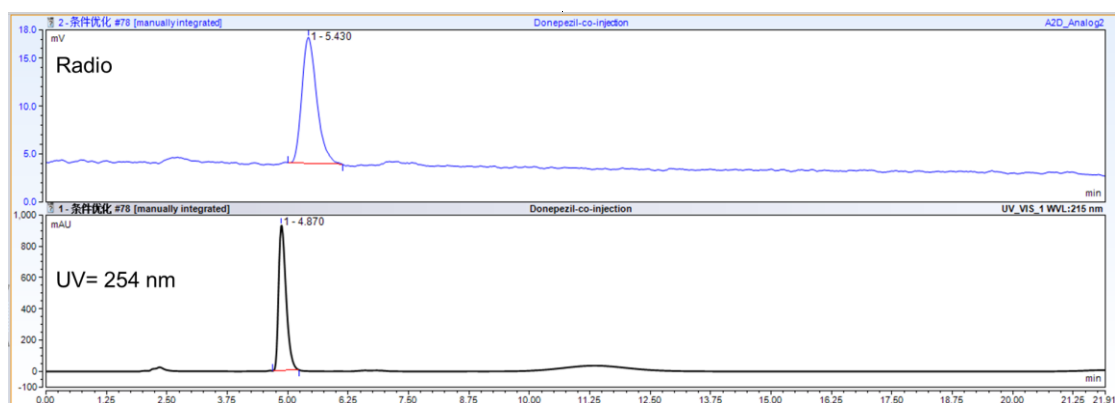


Fig. S23: Co-injection for $[^{18}\text{F}]\mathbf{43}$. 2# HPLC condition; NanoChrom C18 column ($5\mu\text{m}$, $4.6 \times 250\text{mm}$), isocratic elution, $\text{H}_2\text{O}/\text{CH}_3\text{CN} = 35/65$, and the flow rate was 1 mL/min for 20 min. The RCP was $> 97\%$.

Preparation of [¹⁸F]44

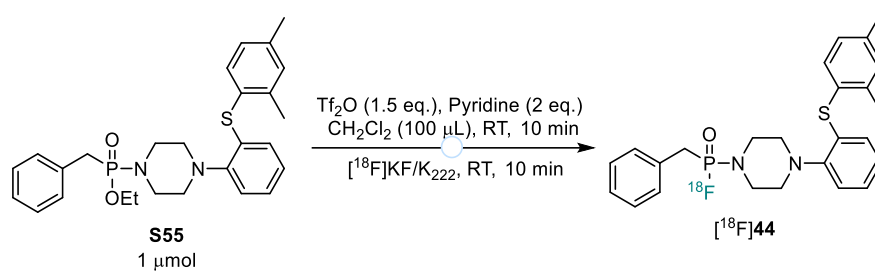
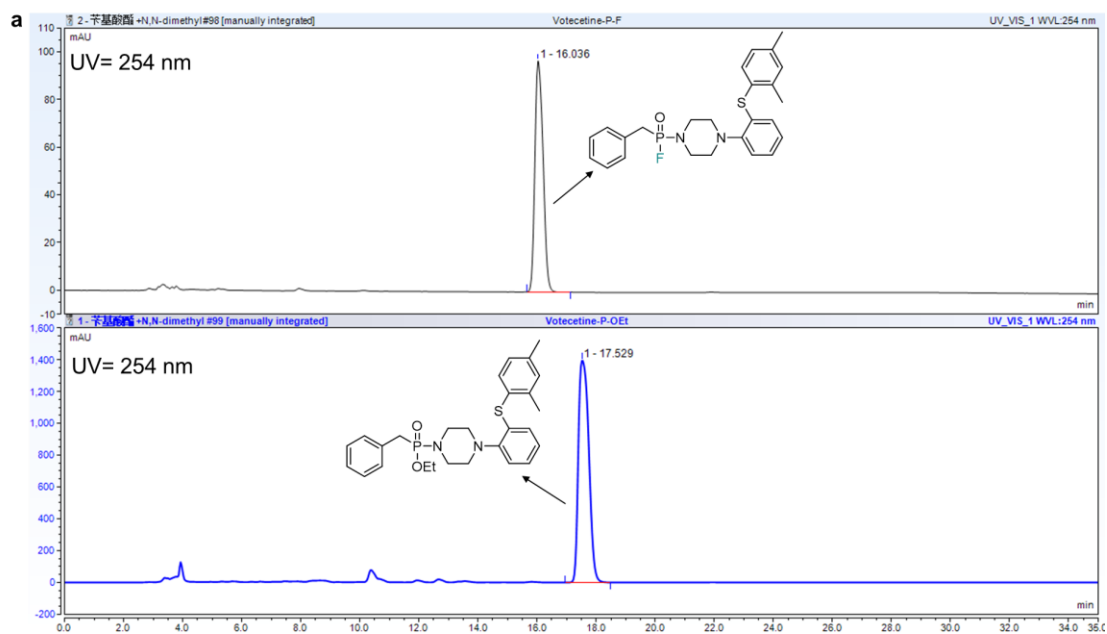


Table S14: Average radiochemical conversion for [¹⁸F]44

Run	1	2	3	Average
RCC _{TLC} (%)	67	66	71	68 ± 2



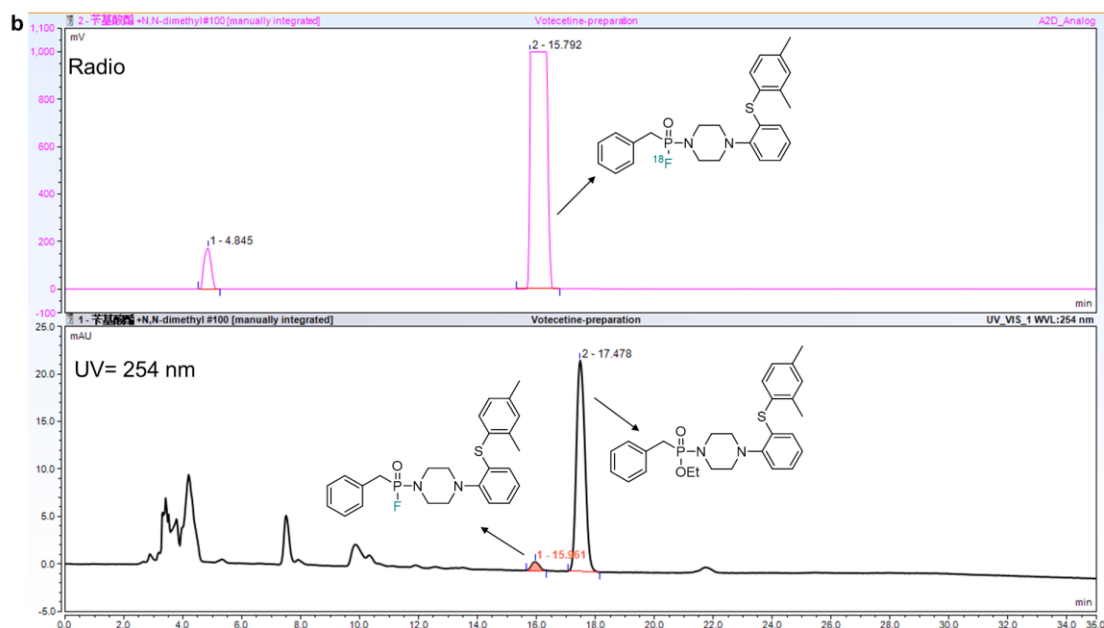


Fig. S24: a. HPLC traces of precursor and reference compounds; b. HPLC traces of crude ^{18}F -labeled products. 1# HPLC conditions; SEP Basic-C18 semi-preparative column (120A $5\mu\text{m}$ $10 \times 250\text{mm}$); isocratic elution, 0–15 min, from 80% of CH_3CN and 20% of water containing 0.1%TFA to 85% of CH_3CN and 15% of water containing 0.1%TFA; 15–25 min, 85% of CH_3CN and 15% of water containing 0.1%TFA; 25–35 min, from 85% of CH_3CN and 15% of water containing 0.1%TFA to 90% of CH_3CN and 10% of water containing 0.1%TFA; flow rate: 3.0 mL/min.

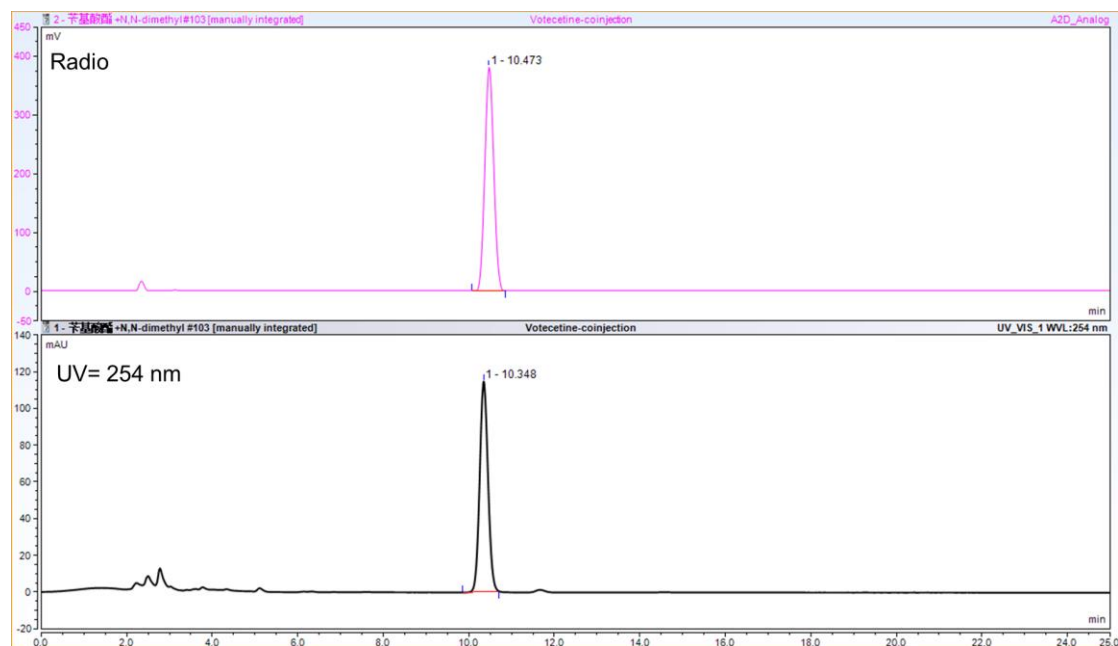


Fig. S25: Co-injection for ^{18}F 44. 1# HPLC condition: NanoChrom C18 column ($5\mu\text{m}$, $4.6 \times 250\text{mm}$), isocratic elution, water containing 0.1%TFA/ CH_3CN = 50/50, and the flow

rate was 1 mL/min for 20 min. The RCP was > 98%.

Preparation of [¹⁸F]45

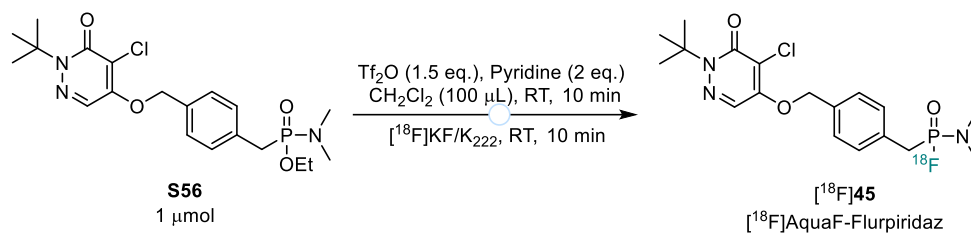
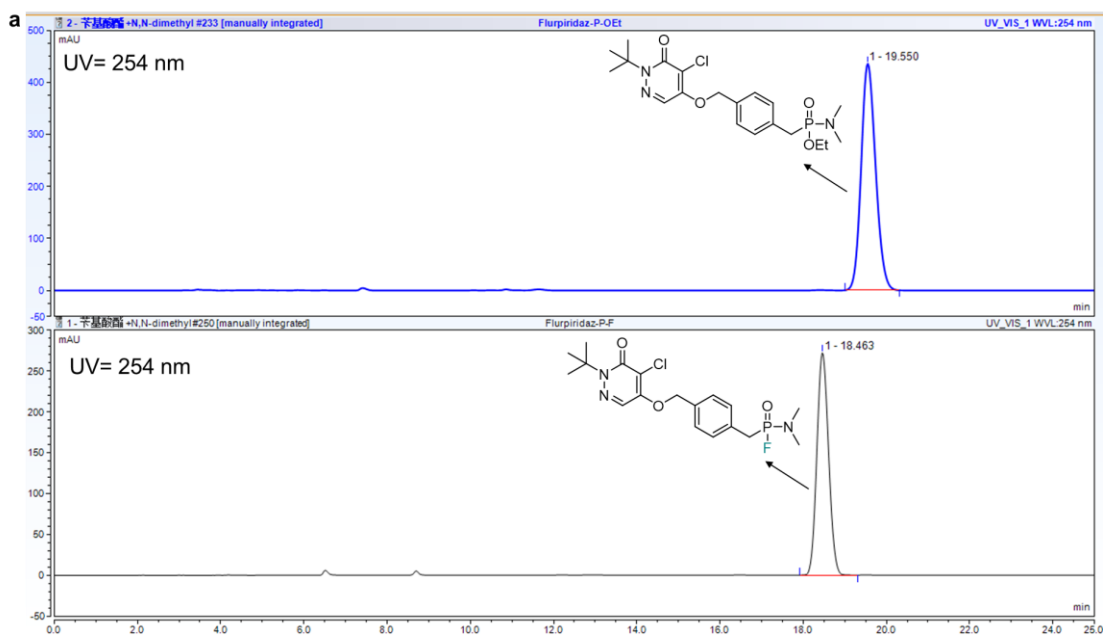


Table S15. Average radiochemical conversion for [¹⁸F]45

Run	1	2	3	Average
RCC _{TLC} (%)	65	59	47	57 ± 9



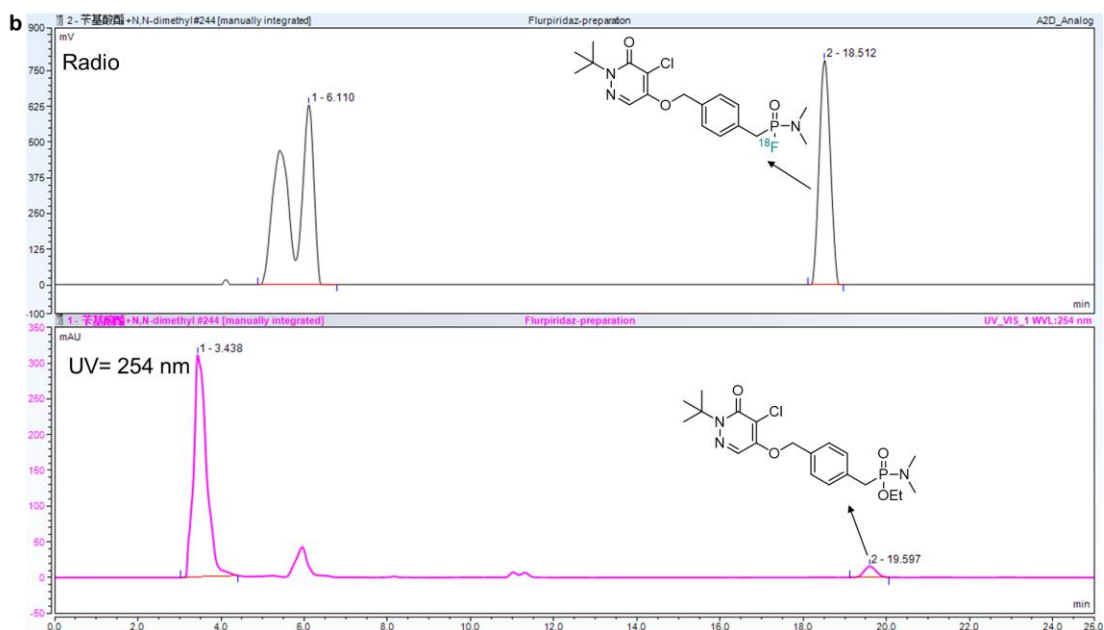


Fig. S26: a. HPLC traces of precursor and reference compounds; b. HPLC traces of crude ^{18}F -labeled products. 1# HPLC conditions; SEP Basic-C18 semi-preparative column (120A $5\mu\text{m}$ $10 \times 250\text{mm}$); elution: isocratic elution, water containing 0.1%TFA/ CH_3CN = 50/50, and the flow rate was 3 mL/min for 25 min.

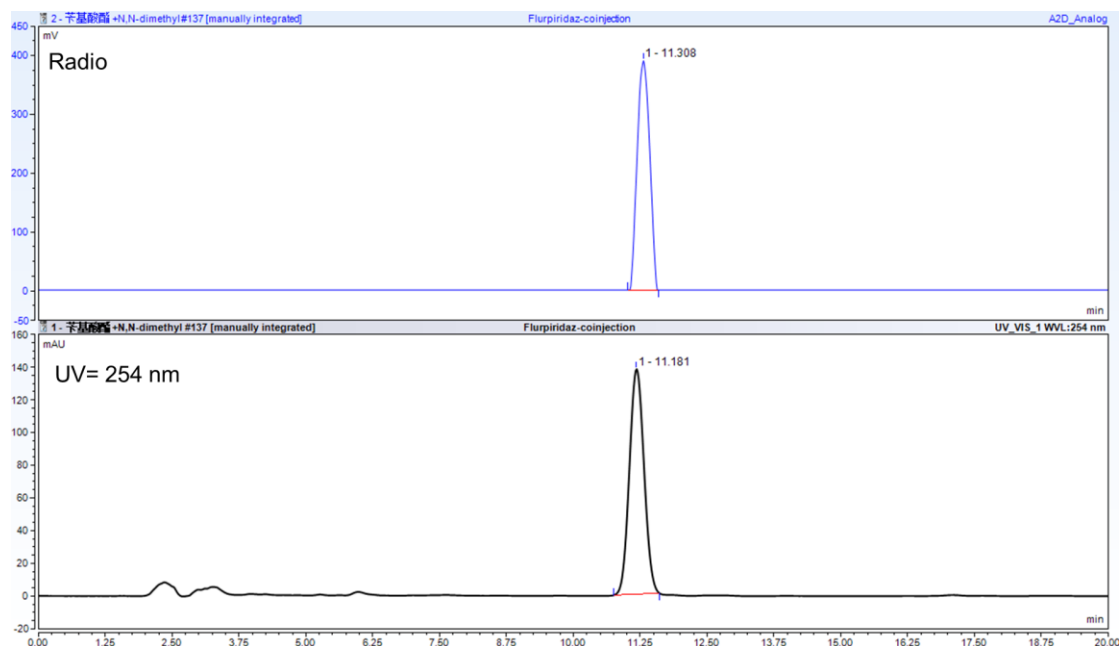


Fig. S27: Co-injection for ^{18}F 45. 1# HPLC condition: NanoChrom C18 column ($5\mu\text{m}$, $4.6 \times 250\text{mm}$), isocratic elution, $\text{H}_2\text{O}/\text{CH}_3\text{CN}$ = 50/50, and the flow rate was 1 mL/min for 20 min. The RCP was >98%.

Preparation of BFPA-E[c(RGDyK)]₂

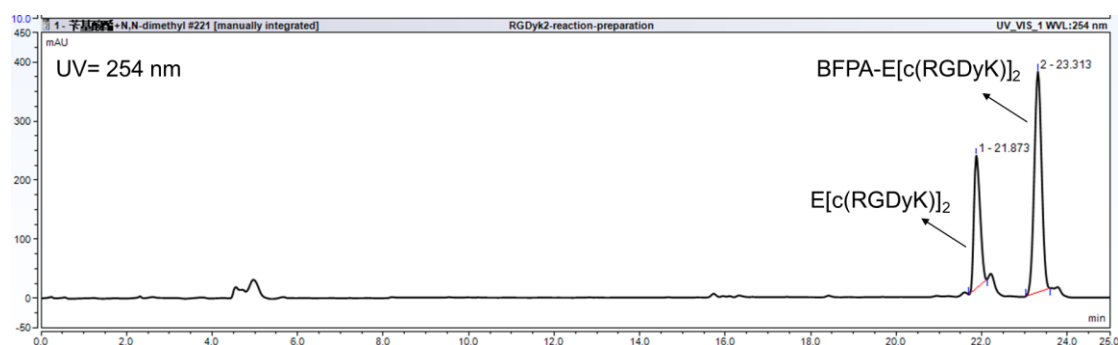
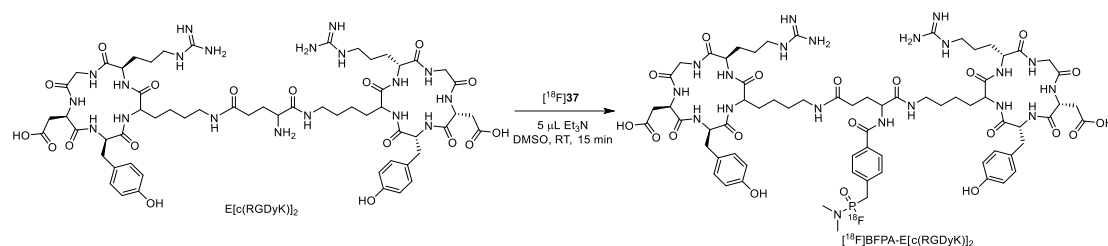


Fig. S28: HPLC traces of crude products. 1# HPLC conditions: SEP Basic-C18 semi-preparative column (120A 5 μ m 10 \times 250mm); gradient elution, 0–10 min, from 10% of CH₃CN and 90% of water containing 0.1%TFA to 20% of CH₃CN and 80% of water containing 0.1%TFA; 10–25 min, from 20% of CH₃CN and 80% of water containing 0.1%TFA to 30% of CH₃CN and 70% of water containing 0.1%TFA; flow rate: 3.0 mL/min.

Preparation of [¹⁸F]BFPA-E[c(RGDyK)]₂



T: FTMS + p ESI Full ms [200.0000-2000.0000]

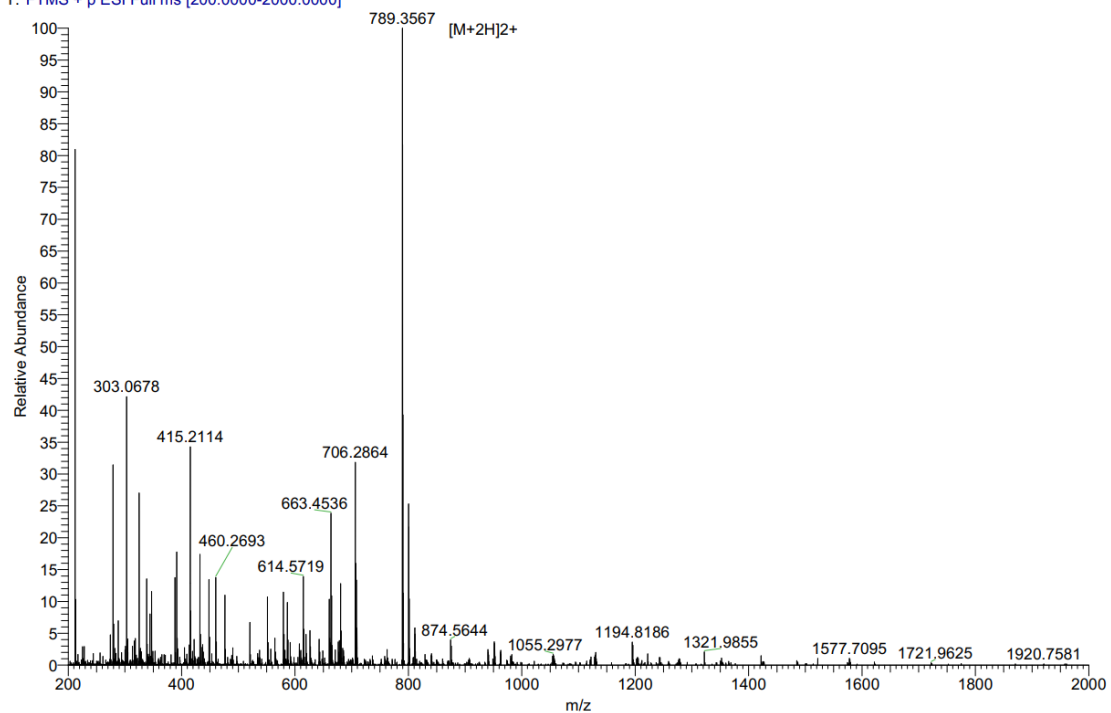


Fig. S29: Mass spectrometry identification of standards. **HRMS (m/z)** calculated for $C_{69}H_{99}FN_{20}O_{20}P^{2+}$ ($[M+2H]^{2+}$): 789.3567 found: 789.3567.

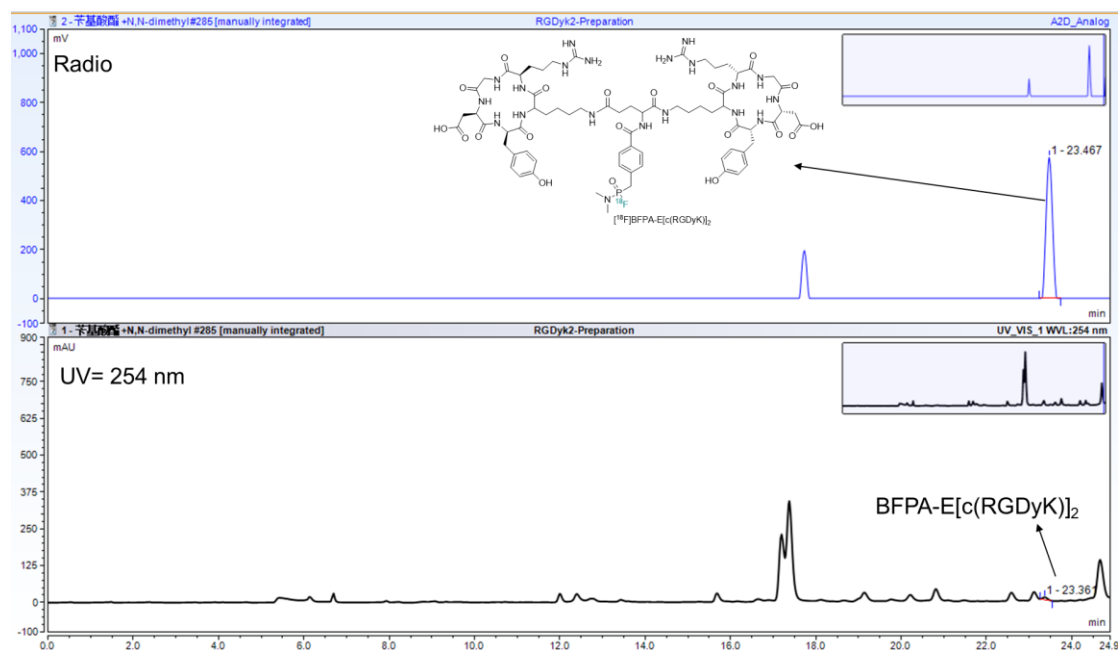


Fig. S30: HPLC traces of crude ^{18}F -labeled products. 1# HPLC conditions: SEP Basic-C18 semi-preparative column (120A $5\mu\text{m}$ $10 \times 250\text{mm}$); gradient elution, 0–10 min, from 10% of CH_3CN and 90% of water containing 0.1%TFA to 20% of CH_3CN and 80% of water containing 0.1%TFA; 10–25 min, from 20% of CH_3CN and 80% of water containing 0.1%TFA to 30% of CH_3CN and 70% of water containing 0.1%TFA; flow rate: 3.0 mL/min.

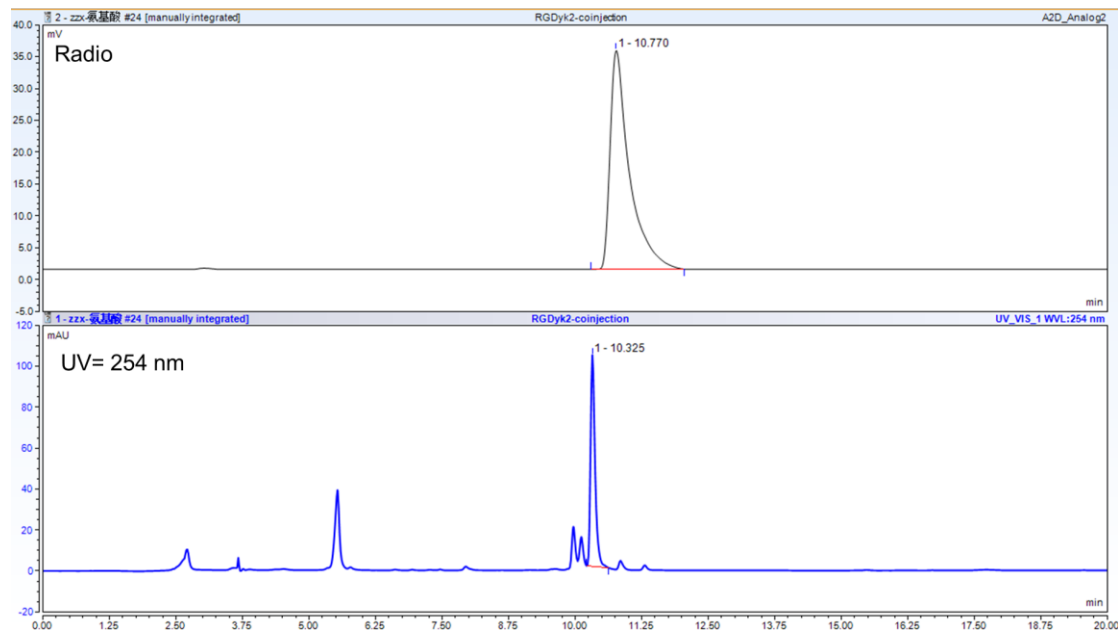
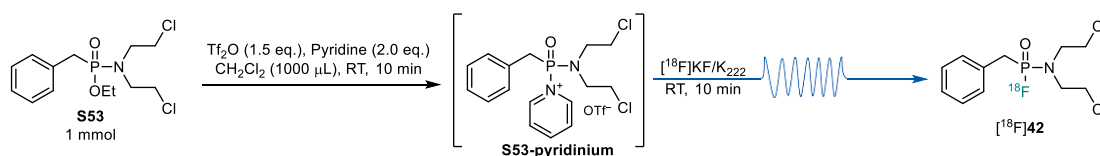


Fig. S31: Co-injection for ^{18}F -BFPA-E[c(RGDyK)]₂. 2# HPLC condition: NanoChrom C18 column ($5\mu\text{m}$, $4.6 \times 250\text{mm}$), gradient elution, from 10% of CH_3CN and 90% of water containing 0.1%TFA to 50% of CH_3CN and 50% of water containing 0.1%TFA, and the flow rate was 1 mL/min for 20 min. The RCP was $> 99\%$.

2.4 Automated radiosynthesis

2.4.1 Synthesizing [¹⁸F]42 on an Allinone module



The automated synthesis of [¹⁸F]42 was performed on an AllinOne module (Trasis, Ans, Belgium) following this stepwise procedure, and the resultant module program was edited in Trasis-AllinOne 2.3.4.

The Sep-Pak[®] Plus Short C18 cartridge and Sep-Pak[®] Light QMA cartridge utilized in the experiment were pre-activated and subsequently loaded into the cartridge as detailed in section 1.3 of this report. Slot 2 was filled with a solution containing 8.0 mg of K₂₂₂, 1.0 mg of K₂CO₃ in CH₃CN/H₂O (4/1, v/v, 0.5 mL) as the eluent. Slot 3 was filled with an anhydrous CH₂Cl₂ (1.0 mL) solution of the precursor 1-(benzyl(bis(2-chloroethyl)amino)phosphoryl)pyridin-1-ium. The solvent reservoirs (slot 14 and 15) were filled with anhydrous EtOH (~10 mL) and CH₃CN (~20 mL), respectively, and a water bag was placed in slot 16. During liquid transmission, the dead volume of the corresponding pipeline should be cleared based on the actual situation to ensure the precision of the liquid dosage.

The [¹⁸F]fluoride obtained from the [¹⁸O]H₂O supplied by the cyclotron is captured on the QMA located in slot 5. Subsequently, the QMA is purged with air, followed by the elution of [¹⁸F]fluoride from the QMA into the reactor using the eluent from slot 2. 1 mL of CH₃CN is added and then dried under an airflow at 110 °C. Once the drying process is completed and the reactor temperature returns to room temperature, the precursor solution from slot 3 is transferred into the reactor. The resulting mixture is agitated and thoroughly mixed using a syringe in slot 3, with this step repeated twice for homogenization. Subsequently, the loop is purged with 3 mL of air, and the reaction mixture is allowed to proceed at room temperature for 10 min. After the completion of the reaction, 9 mL of water is added to the reactor to dilute the reaction mixture. Subsequently, the organic compounds in the mixture are trapped on the C18 in slot 9. Using a syringe, 10 mL of water is slowly injected into the C18 for washing, followed by air blowing to remove fluoride ions and acidic impurities. Finally, the crude product is eluted with 0.5 mL of CH₃CN into a transfer vial, and then injected into the HPLC

system for separation. The crude reaction mixture is purified using reverse-phase semi-preparative HPLC (120A 5 μ m 10 \times 250mm). Receive the CH₃CN/H₂O mixed solution separated by HPLC in another transfer vial, dilute it with water (10 mL), then concentrate the product on the C18 in slot 13. Dry the captured product with air blowing, then elute the desired [¹⁸F]**42** into a product vial using 0.5 mL of EtOH. Subsequently, take an appropriate amount for the determination of A_m.

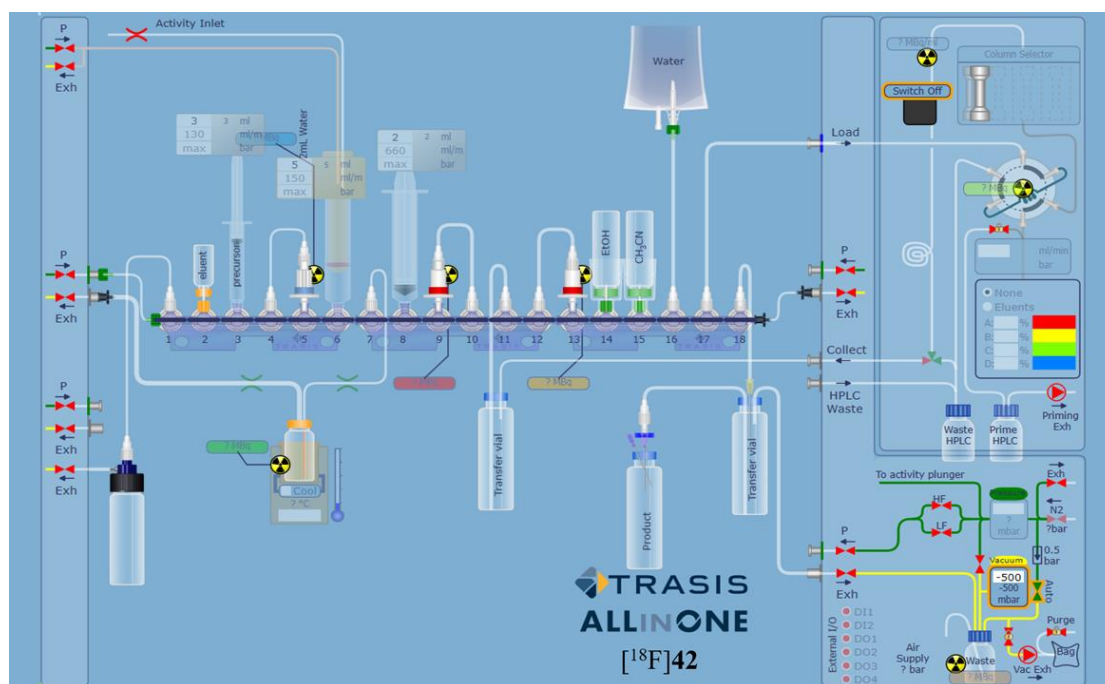


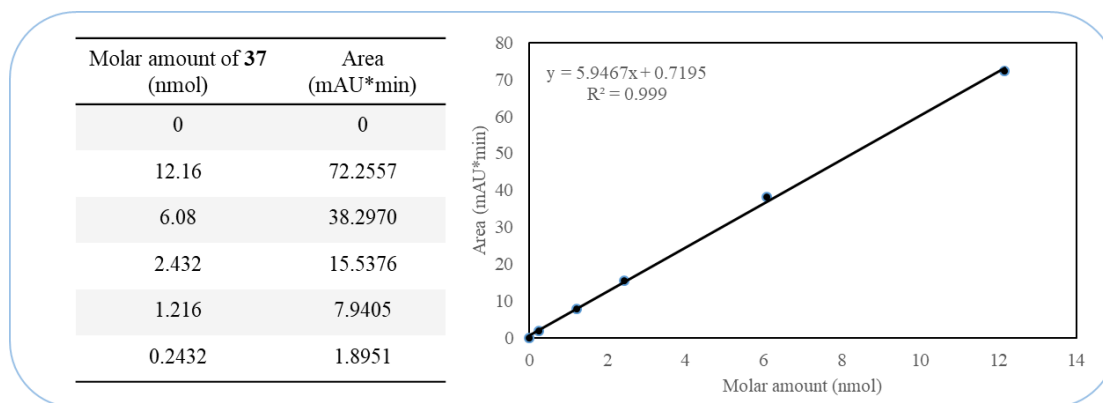
Fig. S32: Graphical representation of the cassette designed for the automated synthesis of [¹⁸F]**42** on a Trasis AllinOne synthesizer.

2.4.2 Molar activity calculations

A_m values were determined with the separated chemically and radiochemically pure samples > 99% purity samples. The sample was injected into HPLC to record the injection activity and injection time, respectively. Subsequently, the UV signal corresponding to the desired radioactive fluorination product was integrated, the chemical amount of the sample is calculated by the standard curve, and then A_m is calculated.

Molar activity calculation of [¹⁸F]**37**

The standard curve (**Fig. S33**) of HPLC peak area (UV, 254 nm) versus molar amount of **37** was plotted.



Standard curve equation: $y = 5.9467x + 0.7195$

Fig. S33: Standard curve acquired with **37** at 254 nm

Table S16: Molar activity calculation of [¹⁸F]**37**

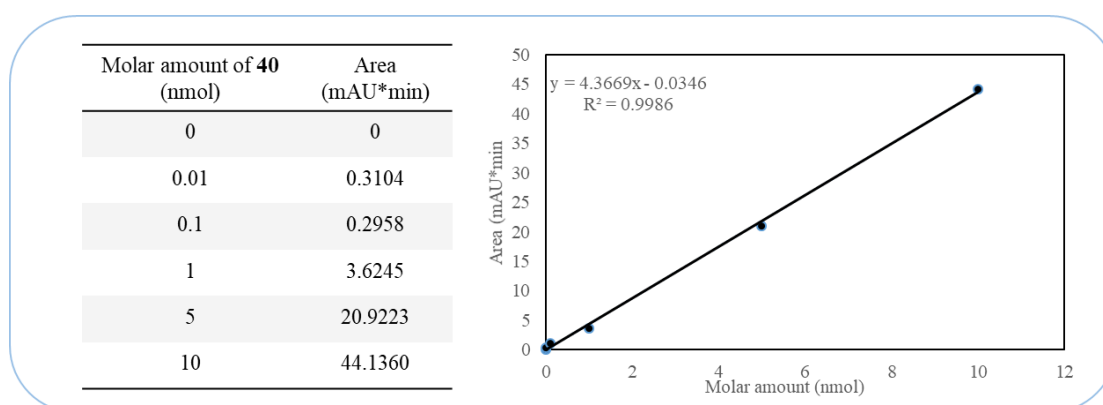
Measurement	Activity injected (MBq, d.c.)	Peak area (mAU*min)	Molar amount (μmol)	A_m (GBq/ μmol)
1	0.402	0.785	1.10×10^{-5}	36.5
2	2.459	1.028	5.19×10^{-5}	47.4

"d.c." means decay correction.

The isolated [¹⁸F]**37** has an average A_m of 42 ± 7 GBq/ μmol ($n = 2$).

Molar activity calculation of [¹⁸F]**40**

The standard curve (**Fig. S34**) of HPLC peak area (UV, 254 nm) versus molar amount of **40** was plotted.



Standard curve equation: $y = 4.33669x - 0.0346$

Fig. S34: Standard curve acquired with [¹⁸F]**40** at 254 nm

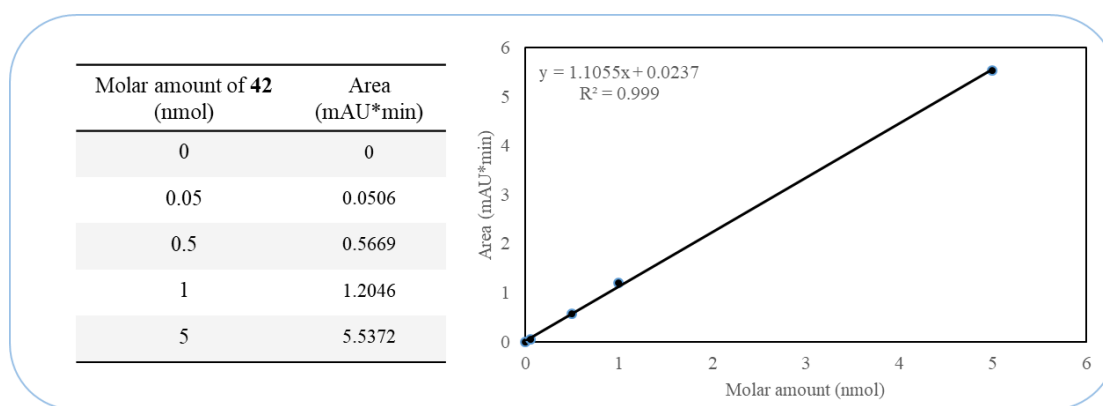
Table S17: Molar activity calculation of [¹⁸F]40

Measurement	Activity injected (MBq, d.c.)	Peak area (mAU*min)	Molar amount (μmol)	A_m (GBq/ μmol)
1	2.64	0.0852	2.74×10^{-5}	95
2	3.88	0.1118	3.35×10^{-5}	115.8

The isolated [¹⁸F]40 has an average A_m of 105 ± 14 GBq/ μmol ($n = 2$).

Molar activity calculation of [¹⁸F]42

The standard curve (Fig. S35) of HPLC peak area (UV, 254 nm) versus molar amount of 42 was plotted.



Standard curve equation: $y = 1.1055x + 0.0237$

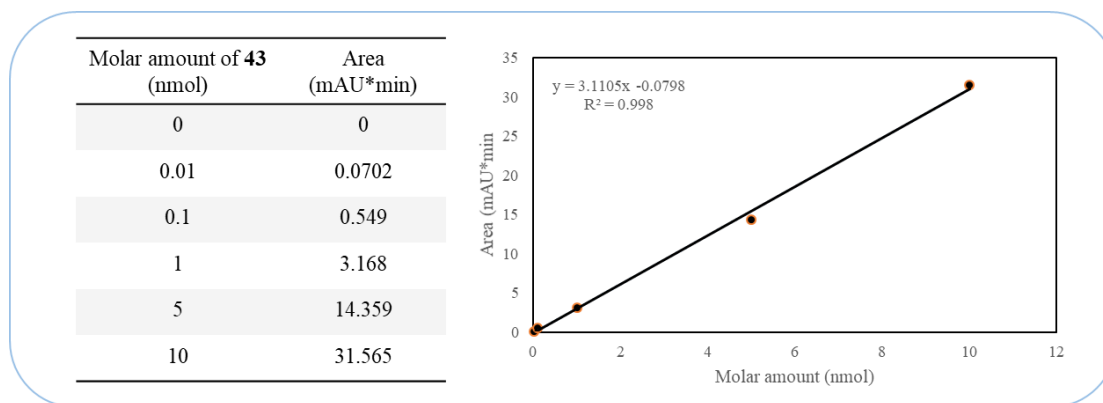
Fig. S35: Standard curve acquired with 42 at 254 nm**Table S18:** Molar activity calculation of [¹⁸F]42

Measurement	Activity injected (MBq, d.c.)	Peak area (mAU*min)	Molar amount (μmol)	A_m (GBq/ μmol)
1	7.08	0.0544	2.778×10^{-5}	254.9
2	8.03	0.0271	3.076×10^{-5}	261.1
3	7.41	0.0582	3.121×10^{-5}	237.4

The isolated [¹⁸F]42 has an average A_m of 251 ± 12 GBq/ μmol ($n = 3$).

Molar activity calculation of [¹⁸F]43

The standard curve (Fig. S36) of HPLC peak area (UV, 254 nm) versus molar amount of 43 was plotted.



Standard curve equation: $y = 3.1105x - 0.0798$

Fig. S36: Standard curve acquired with **43** at 254 nm

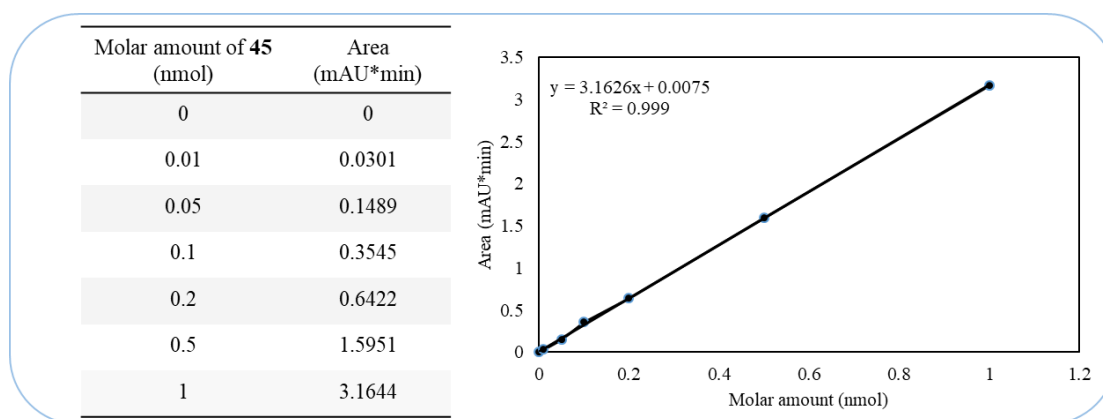
Table S19: Molar activity calculation of [¹⁸F]**43**

Measurement	Activity injected (MBq, d.c.)	Peak area (mAU*min)	Molar amount (μmol)	A_m (GBq/ μmol)
1	2.98	0.0982	5.72×10^{-5}	52.1
2	7.322	0.2550	10.76×10^{-5}	68

The isolated [¹⁸F]**37** has an average A_m of 60 ± 11 GBq/ μmol ($n = 2$).

Molar activity calculation of [¹⁸F]**45**

The standard curve (**Fig. S37**) of HPLC peak area (UV, 254 nm) versus molar amount of **45** was plotted.



Standard curve equation: $y = 3.1626x + 0.0075$

Fig. S37: Standard curve acquired with **45** at 254 nm

Table S20: Molar activity calculation of [¹⁸F]45

Measurement	Activity injected (MBq, d.c.)	Peak area (mAU*min)	Molar amount (μmol)	A _m (GBq/μmol)
1	2.891	0.0863	2.492 × 10 ⁻⁵	116

The isolated [¹⁸F]45 has an average A_m of 116 GBq/μmol.

2.5 From Stability to Interaction

2.5.1 Stability evaluation

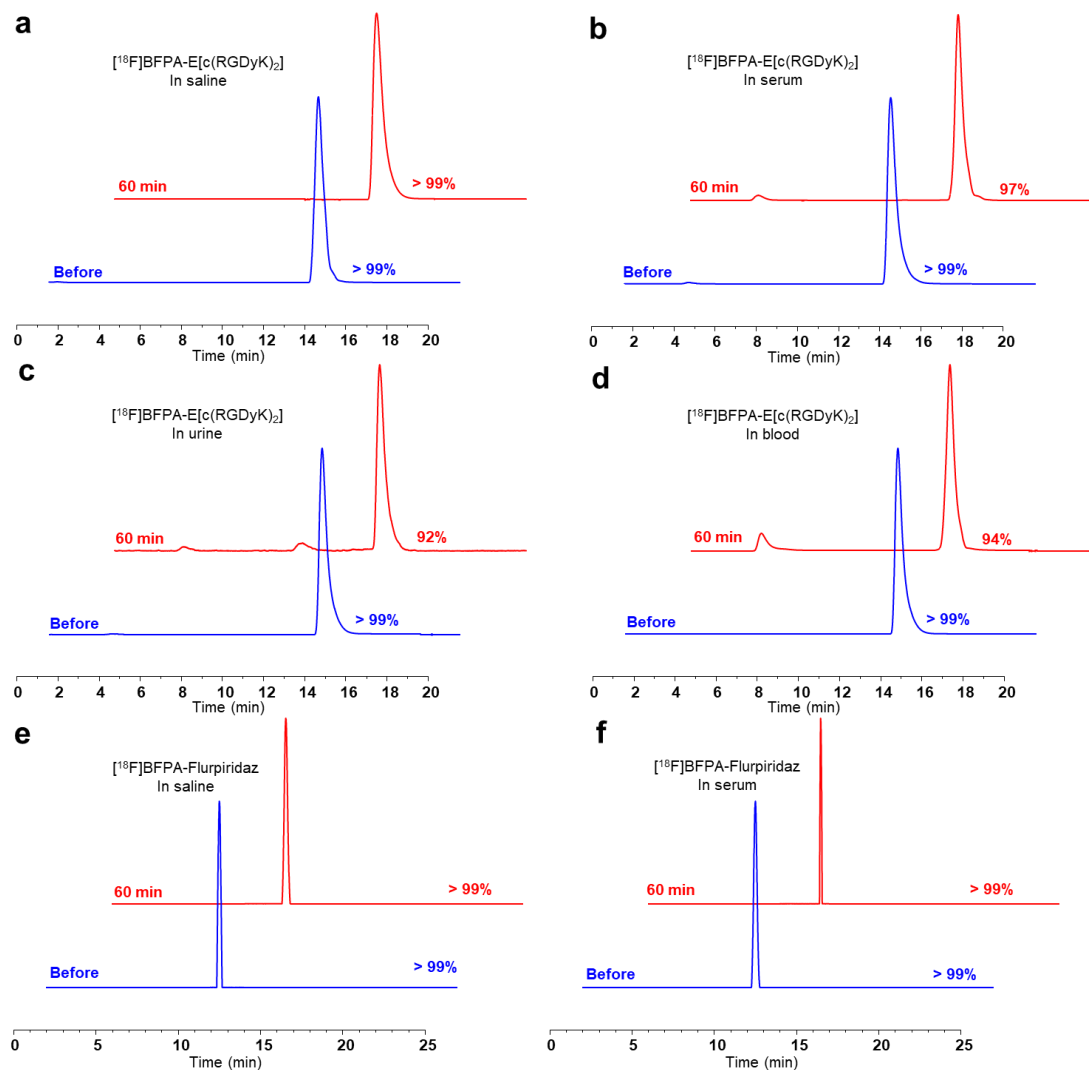


Fig. S38: Stability of radiotracer in *vivo* and *vitro* for 60 min. (a, b) Radio-HPLC analysis

demonstrated that the stability of [^{18}F]BFPA-E[c(RGDyK)]₂ in saline > 99% and serum > 97% *in vitro*. 1# HPLC conditions: NanoChrom C18 column (5 μm , 4.6 \times 250 mm), gradient elution, from 10% of CH₃CN and 90% of water containing 0.1%TFA to 50% of CH₃CN and 55% of water containing 0.1%TFA, and the flow rate was 1 mL/min for 20 min. (c, d) Radio-HPLC analysis demonstrated that [^{18}F]BFPA-E[c(RGDyK)]₂ remains stable at over 92% in urine and over 94% in blood *in vivo*. (e, f) Radio-HPLC analysis also confirmed that [^{18}F]BFPA-Flurpiridaz exhibited stability of over 99% in both saline and serum *in vitro*. HPLC conditions were consistent with Fig. S27.

2.5.2 IC₅₀ determination *via* cell competitive binding assay

The IC₅₀ value for [^{18}F]BFPA-Flurpiridaz was found to be 148.0 nM, which is significantly lower than the IC₅₀ of 248.2 nM for the parent compound Flurpiridaz. This suggests that the modified with fluorophosphine moiety tracer exhibits enhanced binding affinity.

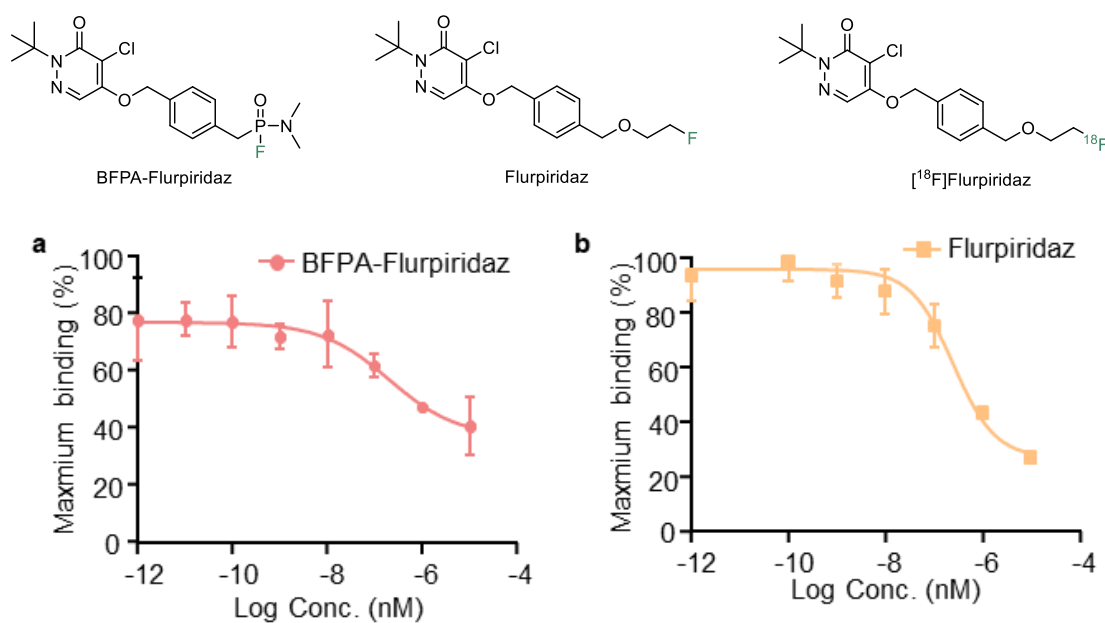
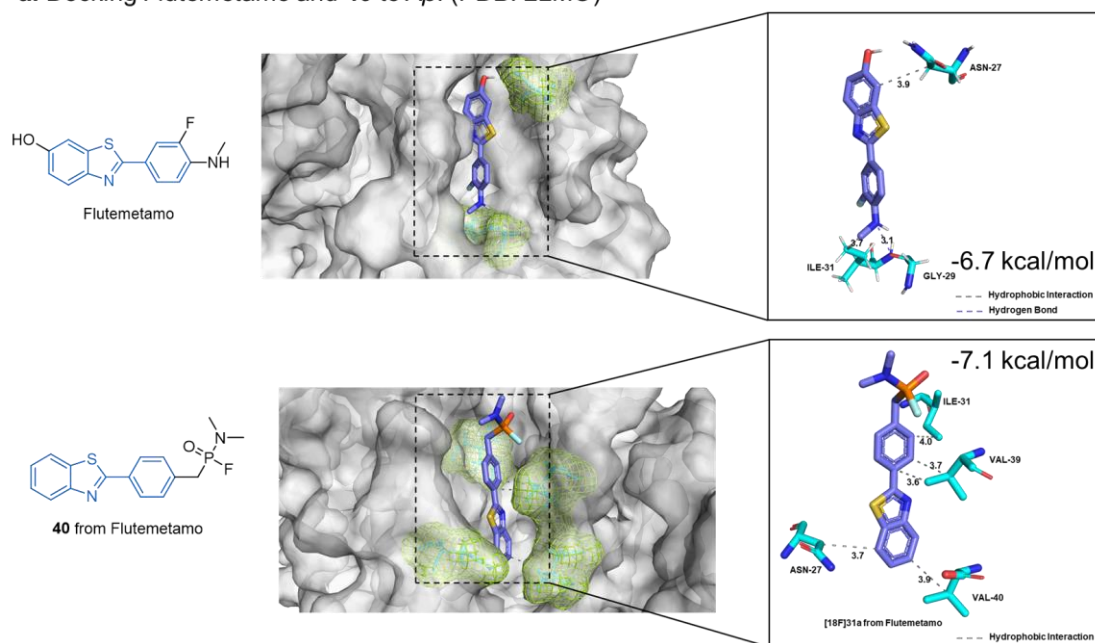


Fig. S39: IC₅₀ curves of BFPA-Flurpiridaz (a) and Flurpiridaz (b) inhibiting [^{18}F]Flurpiridaz.

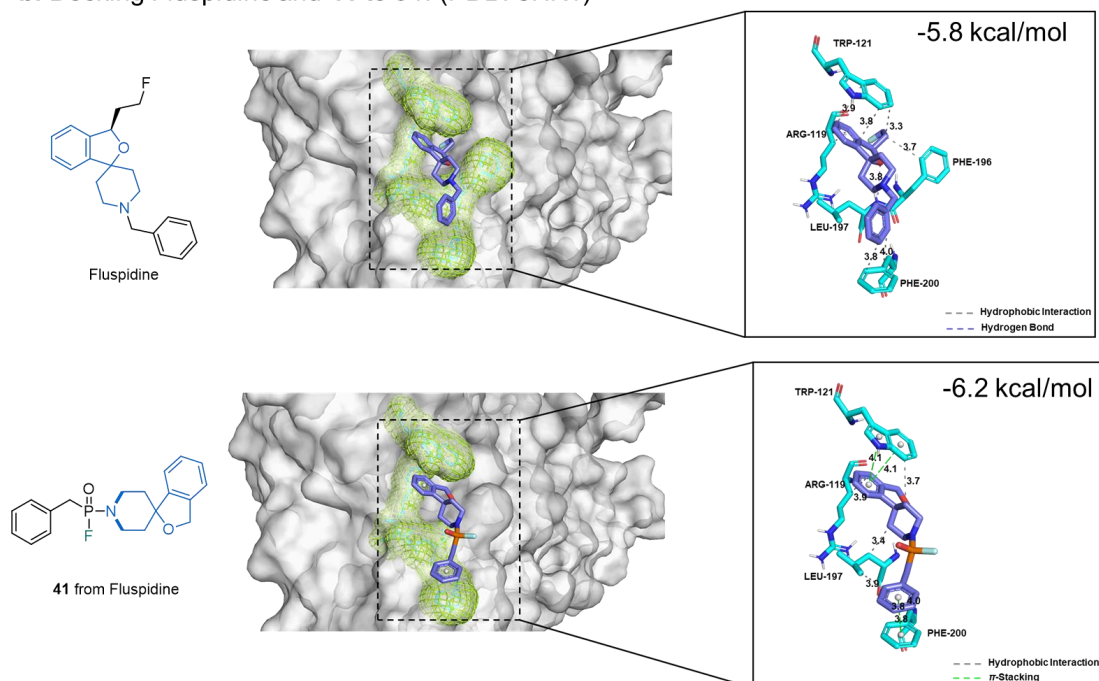
2.5.3 Molecular Docking

The docking results revealed that the binding modes of the parent drug and the BFPA-modified derivative were nearly identical, indicating that the modification did not impair the drug's binding affinity or interaction with the protein. Notably, in certain cases, the BFPA-modified derivatives were able to form an additional hydrogen bond (the additional hydrogen bond donor is from the P=O), which may enhance the overall binding interaction with the target protein.¹³⁻¹⁵

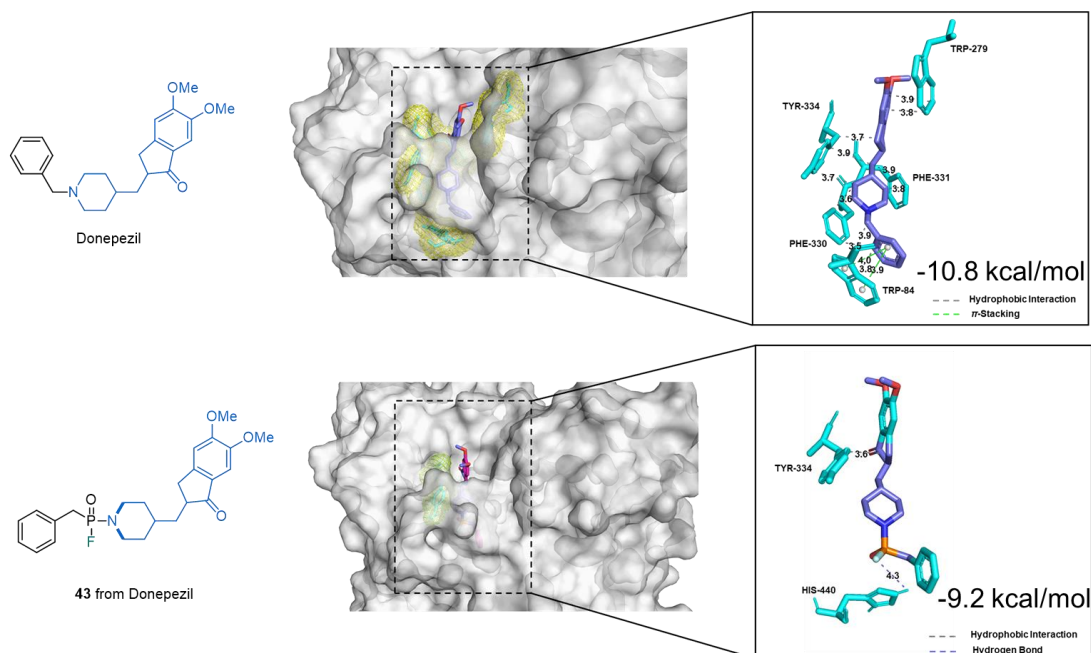
a. Docking Flutemetamo and **40** to A β . (PDB: 2LMO)



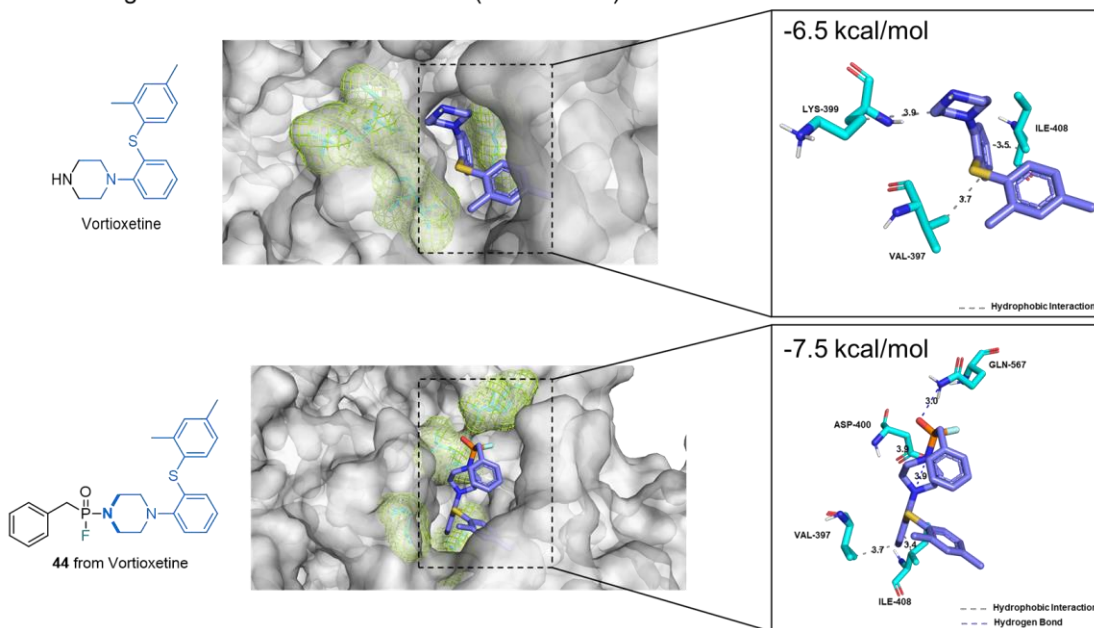
b. Docking Fluspidine and **41** to σ 1. (PDB: 5HK1)



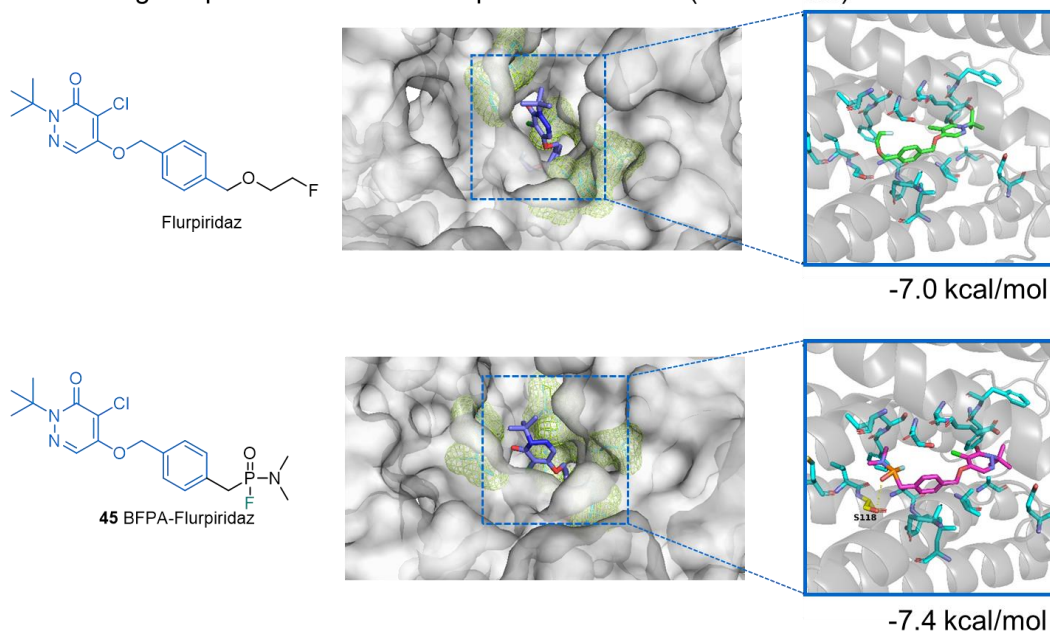
c. Docking Donepezil and **43** to AChE. (PDB: 1EVE)



d. Docking Vortioxetine and **44** to SERT. (PDB: 6ZDV)



e. Docking Flurpiridaz and BFPA-Flurpiridaz to MC I . (PDB: 7ZM8)



f. Docking E[c(RGDyK)]₂ and BFPA-E[c(RGDyK)]₂ to α_vβ₃. (PDB: 4G1E)

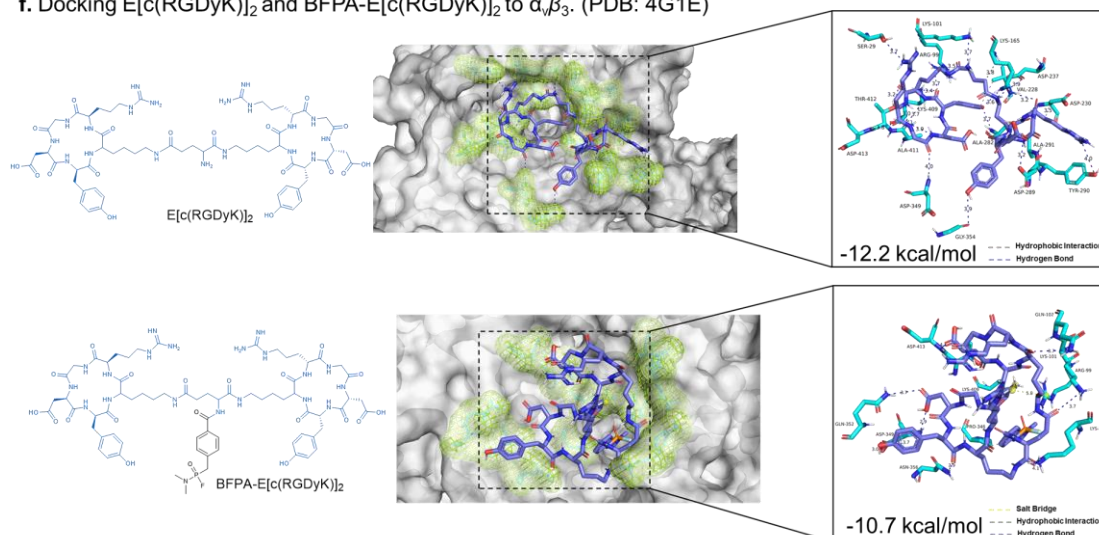
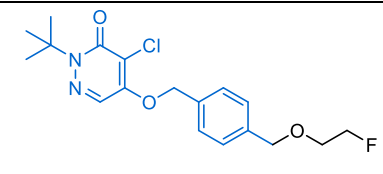

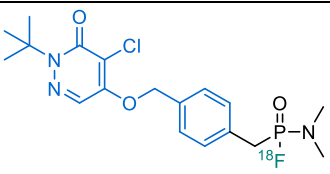
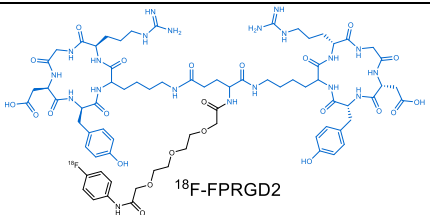

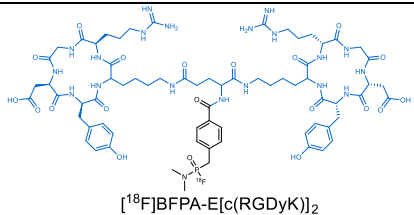
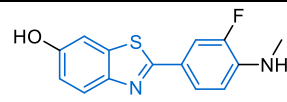

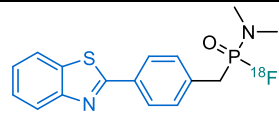
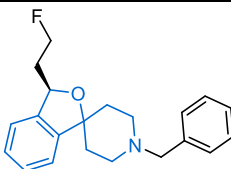

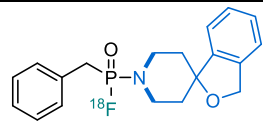
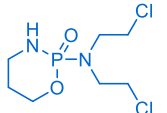

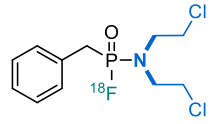
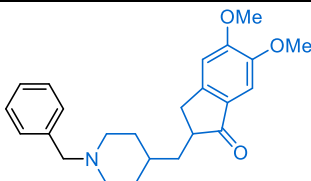

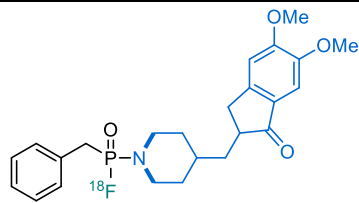
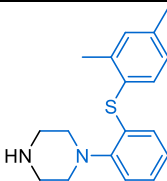

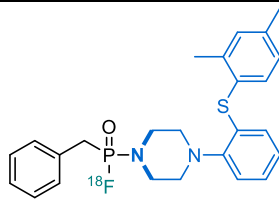


Fig. S40: Ligand–protein docking. All protein structures are sourced from the Protein Data Bank (PDB, <http://www.rcsb.org/>).

Table S21: Physicochemical Properties Analysis: cLog *P* and Log *D*

Log <i>D</i> _{7.4} of original molecule	Trend	Log <i>D</i> _{7.4} of BFPA-modified
 Flurpiridaz Log <i>P</i> = 2.73 ¹⁶		 [18F]BFPA-Flurpiridaz ([18F]45) Log <i>D</i> _{7.4} = 2.06 ± 0.01

 <p>^{18}F-FPRGD2</p> <p>$\text{Log } P = -2.28 \pm 0.05^{17}$</p>		 <p>$[^{18}\text{F}]\text{BFPA-E}[\text{c}(\text{RGDyK})]_2$</p> <p>$\text{Log } D_{7.4} = -2.05 \pm 0.04$</p>
cLog P of original molecule	Trend	cLog P of BFPA-modified
 <p>Flutemetamo</p> <p>$\text{cLog } P = 4.16$</p>		 <p>$[^{18}\text{F}]\mathbf{40}$ from Flutemetamo</p> <p>$\text{cLog } P = 3.09$</p>
 <p>Fluspidine</p> <p>$\text{cLog } P = 4.08$</p>		 <p>$[^{18}\text{F}]\mathbf{41}$ from Fluspidine</p> <p>$\text{cLog } P = 2.89$</p>
 <p>Cyclophosphamide</p> <p>$\text{cLog } P = 0.76$</p>		 <p>$[^{18}\text{F}]\mathbf{42}$ from cyclophosphamide</p> <p>$\text{cLog } P = 2.64$</p>
 <p>Donepezil</p> <p>$\text{cLog } P = 4.14$</p>		 <p>$[^{18}\text{F}]\mathbf{43}$ from Donepezil</p> <p>$\text{cLog } P = 3.93$</p>
 <p>Vortioxetine</p> <p>$\text{cLog } P = 4.51$</p>		 <p>$[^{18}\text{F}]\mathbf{44}$ from Vortioxetine</p> <p>$\text{cLog } P = 5.40$</p>

2.6 Positron emission tomography imaging

2.6.1 Animal models. All animal procedures were conducted in accordance with the Guide for the Care and Use of Laboratory Animals and approved by the Animal Care and Use Committee of Xiamen University (240506 XMULAC20240099).

2.6.2 *In vivo* MicroPET imaging of [¹⁸F]BFPA-E[c(RGDyK)]₂ was conducted in U87MG glioma xenografts in male nude mice.

All MicroPET/CT images were acquired using an Inveon MicroPET/CT scanner (Siemens) and subsequently analyzed with the Inveon Research Workplace Software (Siemens).

For the MicroPET imaging study, an U87MG tumor-bearing male nude mice received a bolus tail-vein injection of a solution of [¹⁸F]BFPA-E[c(RGDyK)]₂ (~3.7 MBq) dissolved in 0.9% saline (100 μL). The mouse was anesthetized using inhaled 2% isoflurane 25 min after injection and then placed on the MicroPET/CT scanner bed. A 5 min PET scan was performed at 30 min after injection.

2.6.3 Blocking study

A male nude mice with an U87MG tumor was administered a bolus tail-vein injection of a solution containing [¹⁸F]BFPA-E[c(RGDyK)]₂ (~3.7 MBq) and E[c(RGDyK)]₂ (200 μg), dissolved in 0.9% saline (100 μL). The mice was anesthetized with 2% inhaled isoflurane 25 min after injection and then carefully positioned on the MicroPET/CT scanner bed. Subsequently, a 5 min MicroPET scan was performed at 30 min post-injection to assess the imaging results.

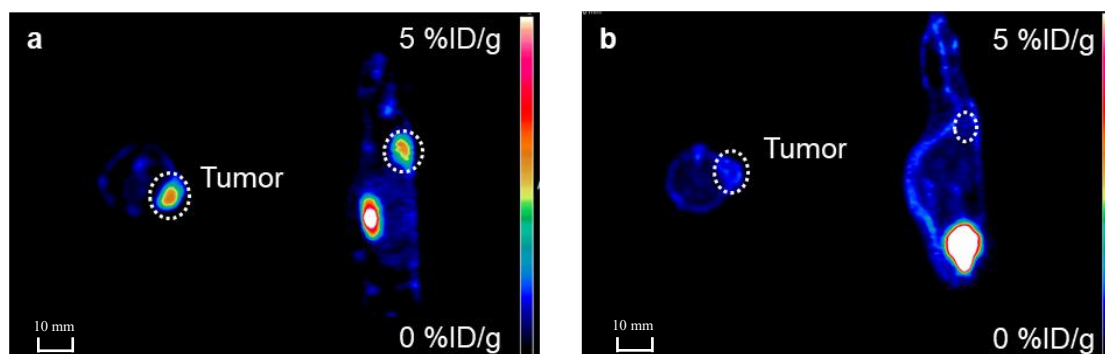


Fig. S41: **a** MicroPET images of [¹⁸F]BFPA-E[c(RGDyK)]₂ in U87MG xenograft mice at 30 min after tail vein injection. **b** MicroPET images of U87MG xenograft mice at 30 min after simultaneous injection of [¹⁸F]BFPA-E[c(RGDyK)]₂ and E[c(RGDyK)]₂ (200 μg).

2.7 Mechanism proposal and computational study

2.7.1 Research mechanisms: NMR and MS

The possible compounds in the reaction system were estimated by ^1H NMR, ^{19}F NMR and ^{31}P NMR *in situ*.

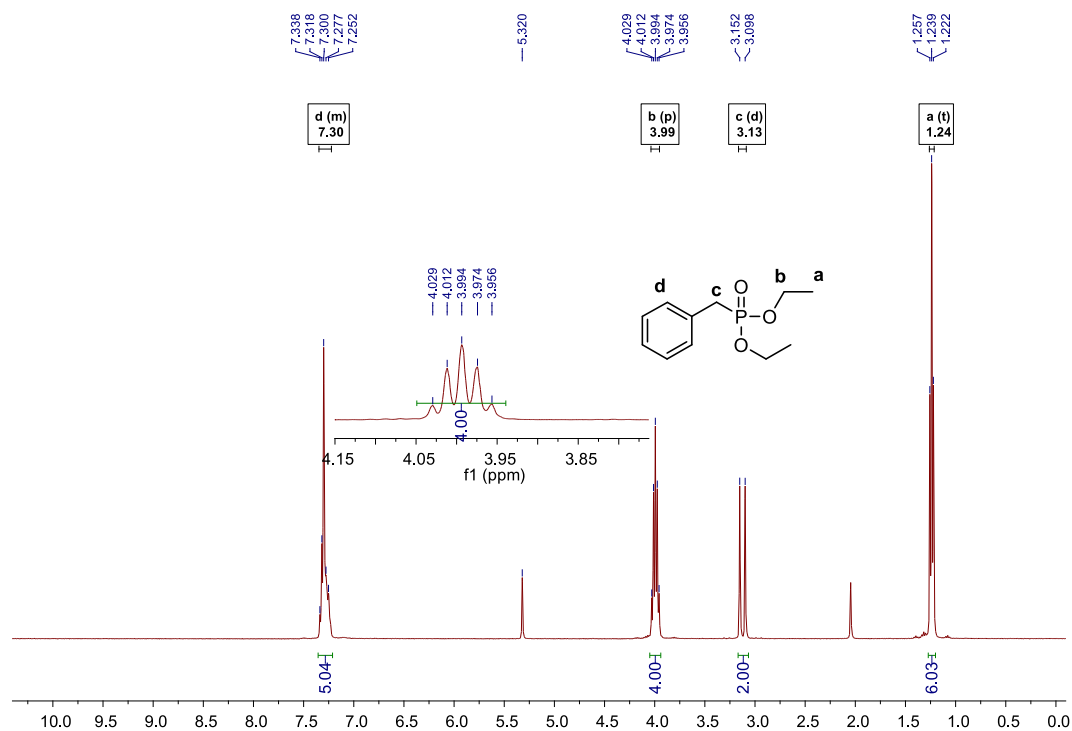


Fig. S42: ^1H NMR of CCOP(=O)(OCC)c1ccccc1 in CD_2Cl_2 , 400 MHz.

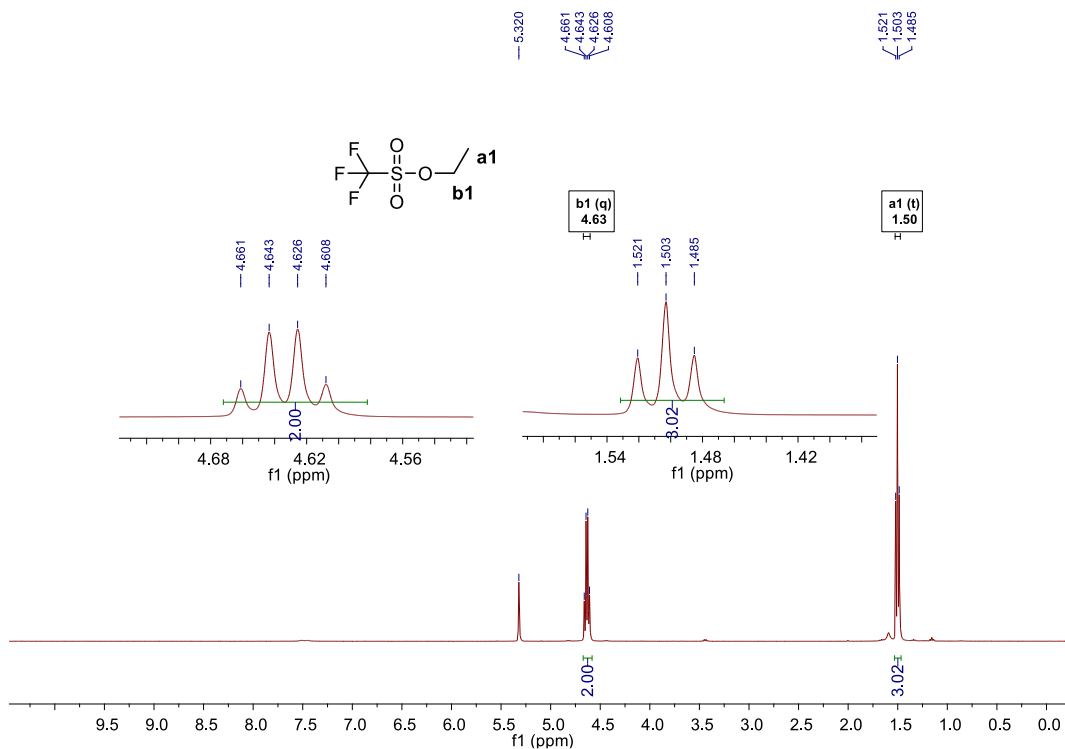


Fig. S43: ^1H NMR of TfOEt in CD_2Cl_2 , 400 MHz.

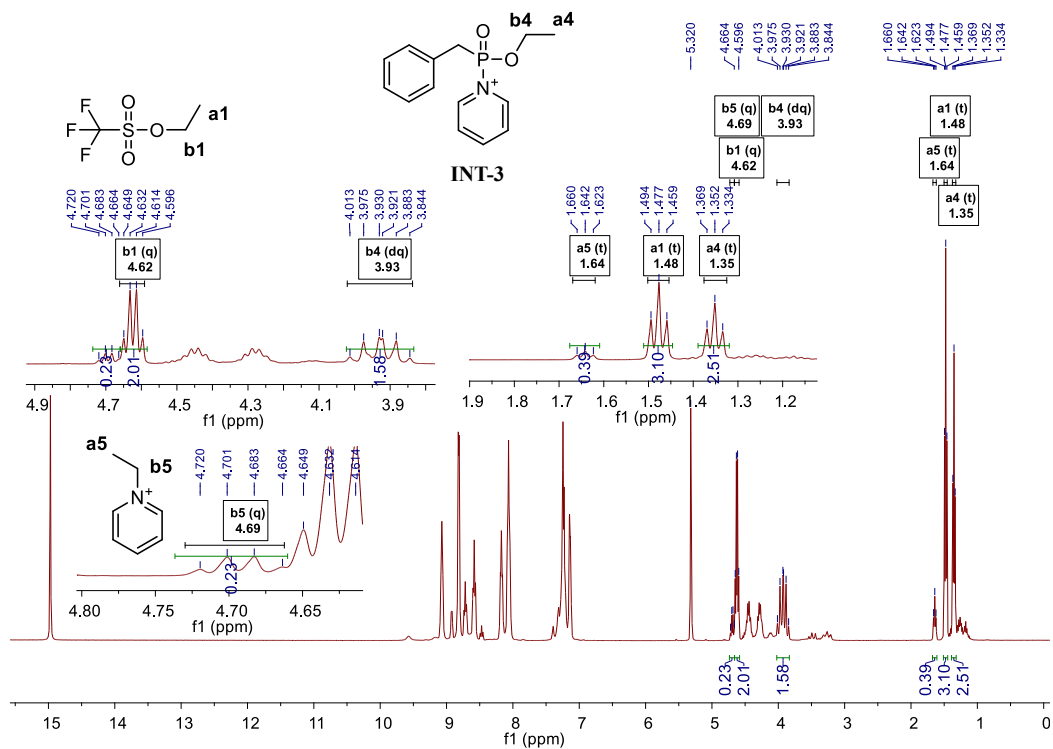


Fig. S44: Crude ^1H NMR of CCOP(=O)(F)F + Tf_2O + C1=CC=CC=C1N in CD_2Cl_2 , 400 MHz.

In the reaction system, 1-ethylpyridin-1-ium detected may be the product of the reaction between pyridine and TfOEt.

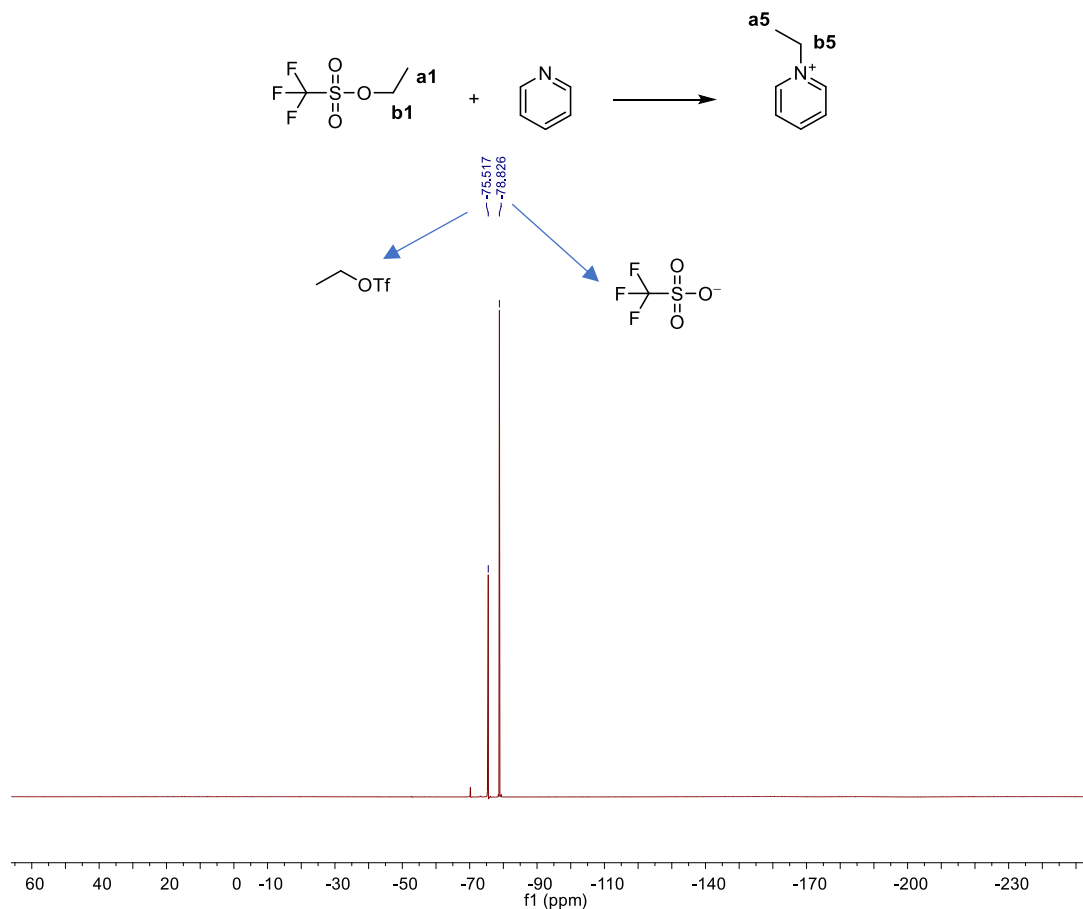
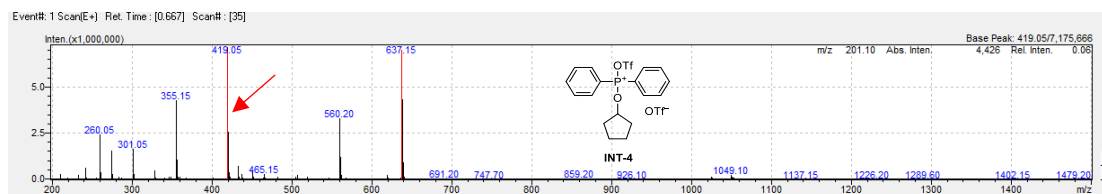


Fig. S45: Crude ¹⁹F NMR of CCOP(=O)(OC)c1ccccc1 + Tf₂O + C1=CC=NC=C1 in CD₂Cl₂, 162 MHz.

INT-4: MS (m/z) calculated for C₁₈H₁₉F₃O₄PS⁺ [M+H]⁺: 419.0688; found: 419.05.



INT-5: MS (m/z) calculated for C₂₃H₂₄F₃NO₄PS⁺ [M+H]⁺: 498.1110; found: 498.10.

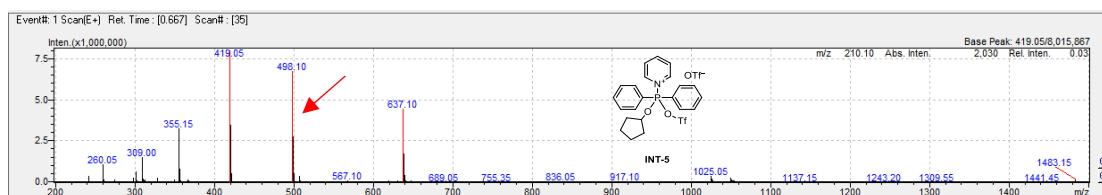


Fig. S46: MS determination of fluorination of compound **S8** *in situ*.

2.7.2 Reaction pathways and free-energy profiles

Computational Details:

Calculations were carried out using the Gaussian 09 programme. Geometries were optimized using the M06 density functional method¹⁸ and 6-31G* basis set. Solvation effect was treated by the default PCM method¹⁹. Then, frequency analyses were performed at the same level of theory. Single-point energy calculations were performed using the 6-311++G** basis set with the same density functional method and solvation model.

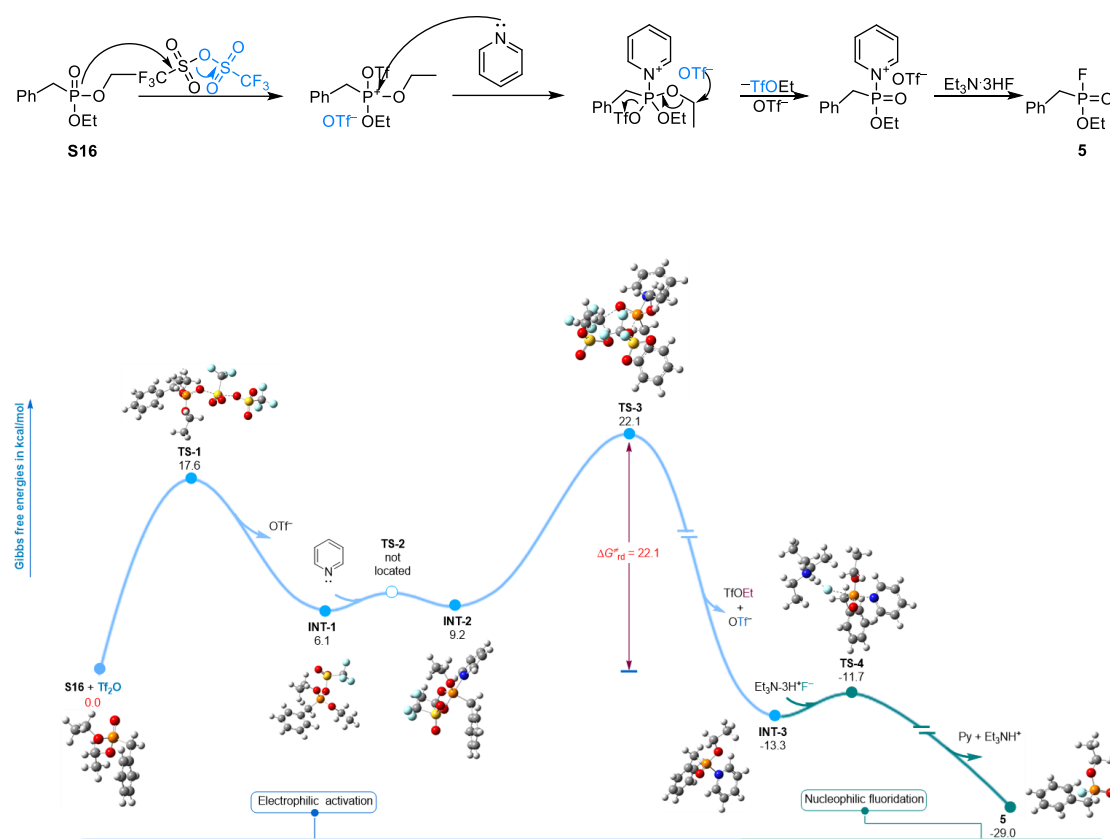


Fig.. S47: Using the Gaussian 09 program, DFT calculations at the m06+GD3/6-311++G** level of theory were employed to predict activation energy barriers for the mechanistic proposal for fluorination of alkyl phosphonates, along with reaction pathways and free-energy profiles. Gaussview 5.0 is used for data analysis.

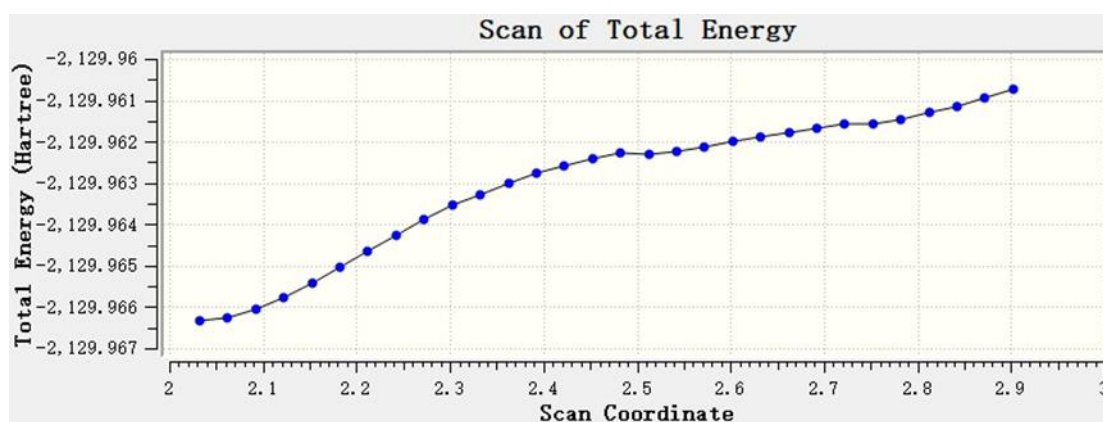
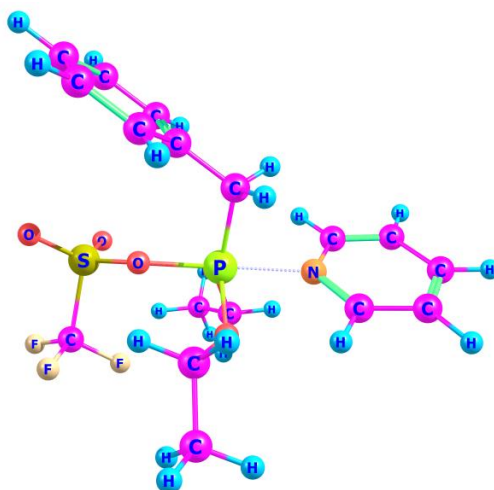


Fig. S48: Scan of total energy with respect to the P...N distance from 2.03 to 2.95 Å indicates that transition state between B and C may not exist.

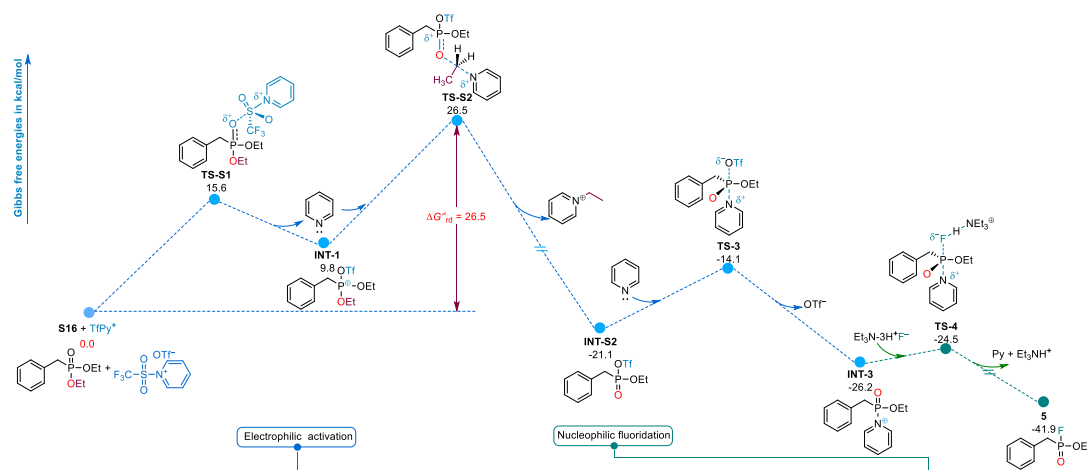


Fig. S49: The energy distribution of alkyl phosphonate (S16) activated by Tf₂O for fluorination at a less likely alternative pathway.

3. Supplementary Figures

3.1 Radio-TLC traces of RCCs

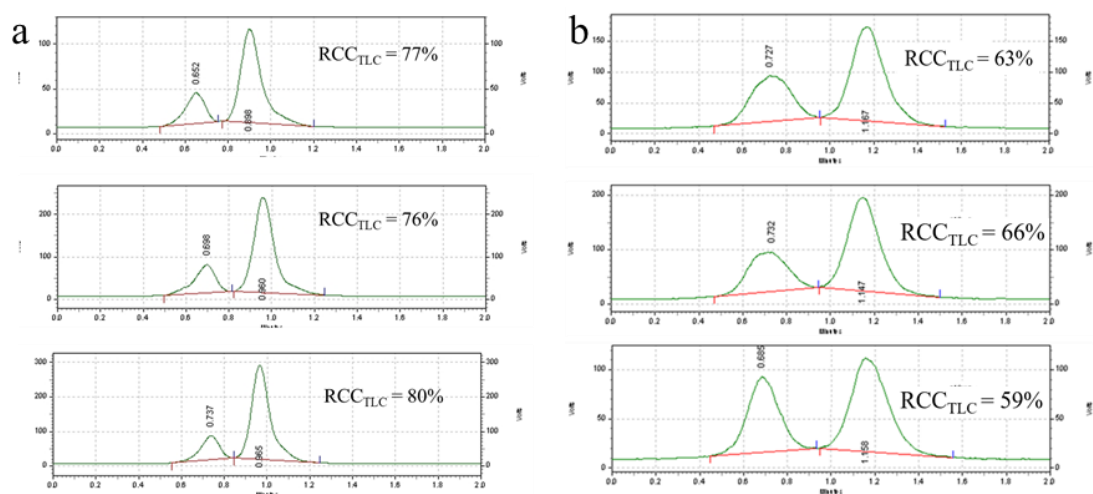


Fig. A1: Radio-TLC traces of (a) ^{18}F -benzylphosphonofluoridic acid ($\text{RCC} = 78 \pm 2\%$) and (b) ^{18}F]1 ($\text{RCC} = 62 \pm 3\%$). Developing solvent: MeOH.

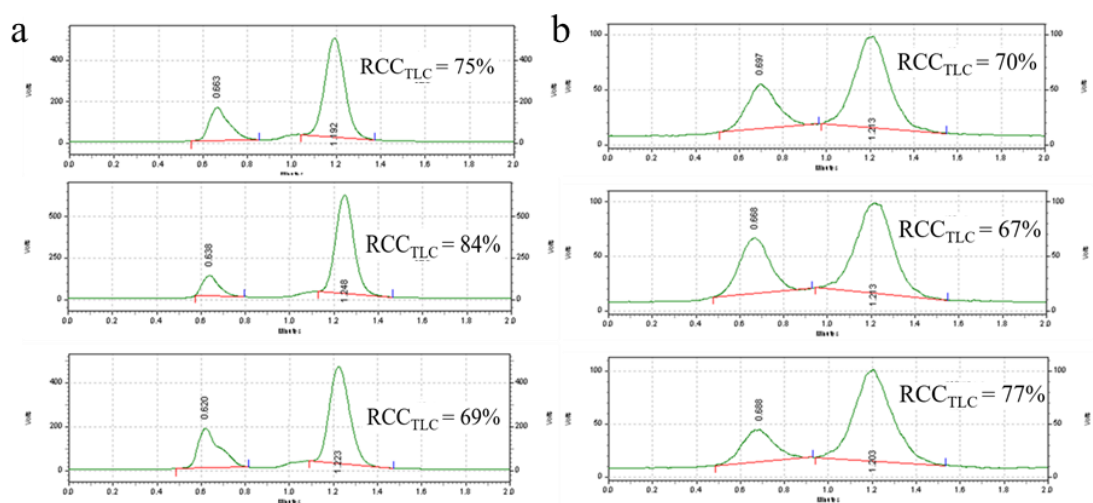


Fig. A2: Radio-TLC traces of (a) ^{18}F]36 ($\text{RCC} = 76 \pm 7\%$) and (b) ^{18}F]37 ($\text{RCC} = 71 \pm 5\%$). Developing solvent: MeOH.

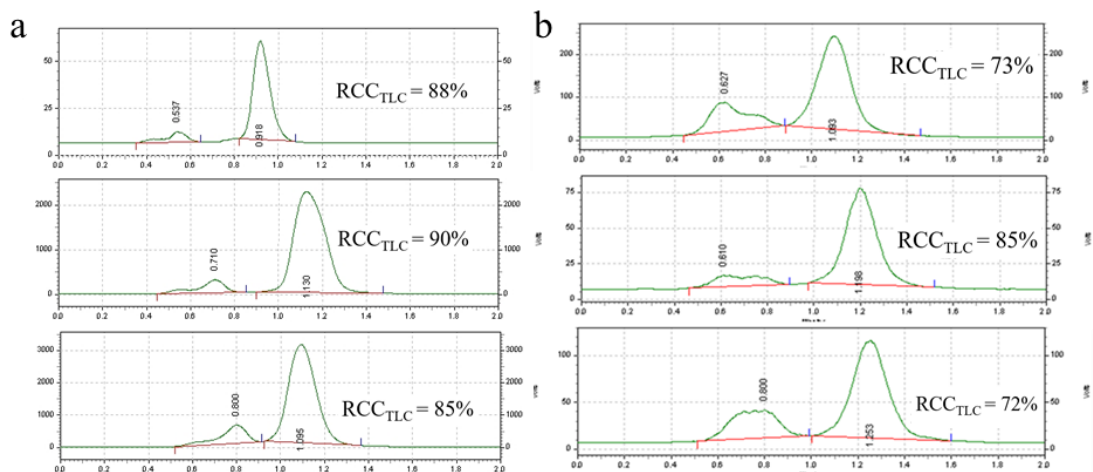


Fig. A3: Radio-TLC traces of (a) $[^{18}\text{F}]\mathbf{38}$ ($\text{RCC} = 88 \pm 3\%$) and (b) $[^{18}\text{F}]\mathbf{39}$ ($\text{RCC} = 77 \pm 7\%$). Developing solvent: MeOH.

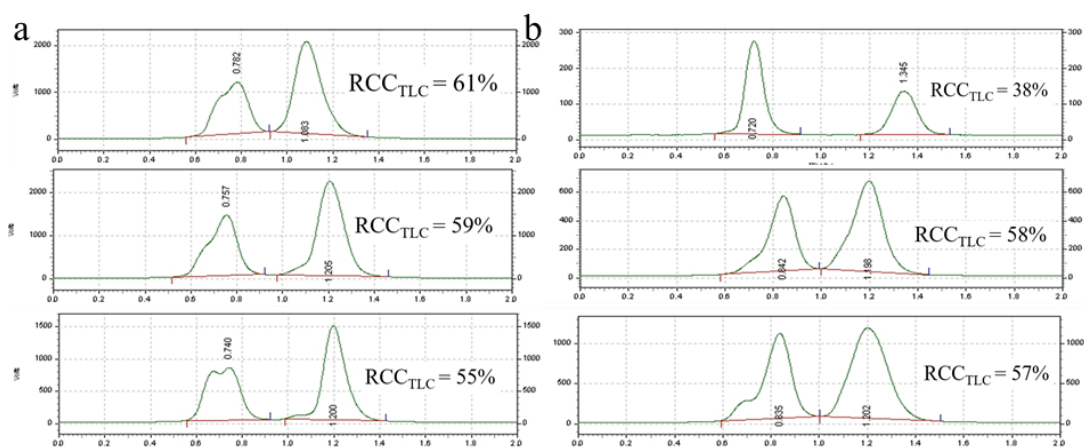


Fig. A4: Radio-TLC traces of (a) $[^{18}\text{F}]\mathbf{40}$ ($\text{RCC} = 58 \pm 3\%$) and (b) $[^{18}\text{F}]\mathbf{41}$ ($\text{RCC} = 51 \pm 11\%$). Developing solvent: MeOH.

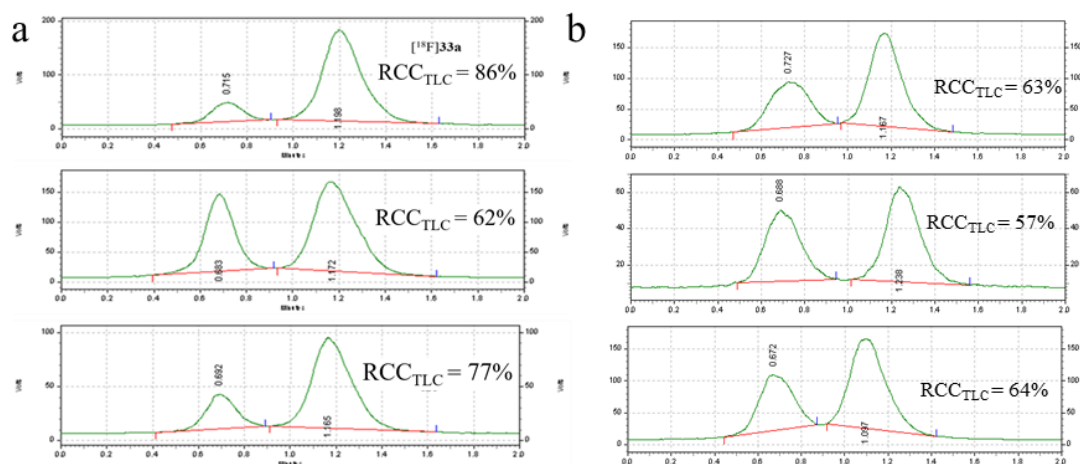


Fig. A5: Radio-TLC traces of (a) $[^{18}\text{F}]\mathbf{42}$ ($\text{RCC} = 75 \pm 12\%$) and (b) $[^{18}\text{F}]\mathbf{43}$ ($\text{RCC} = 61 \pm 4\%$). Developing solvent: MeOH.

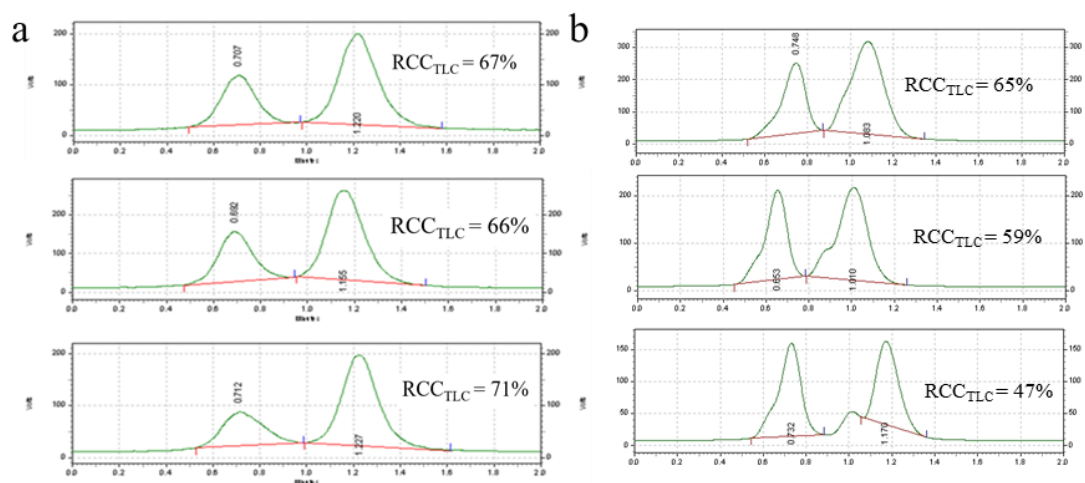
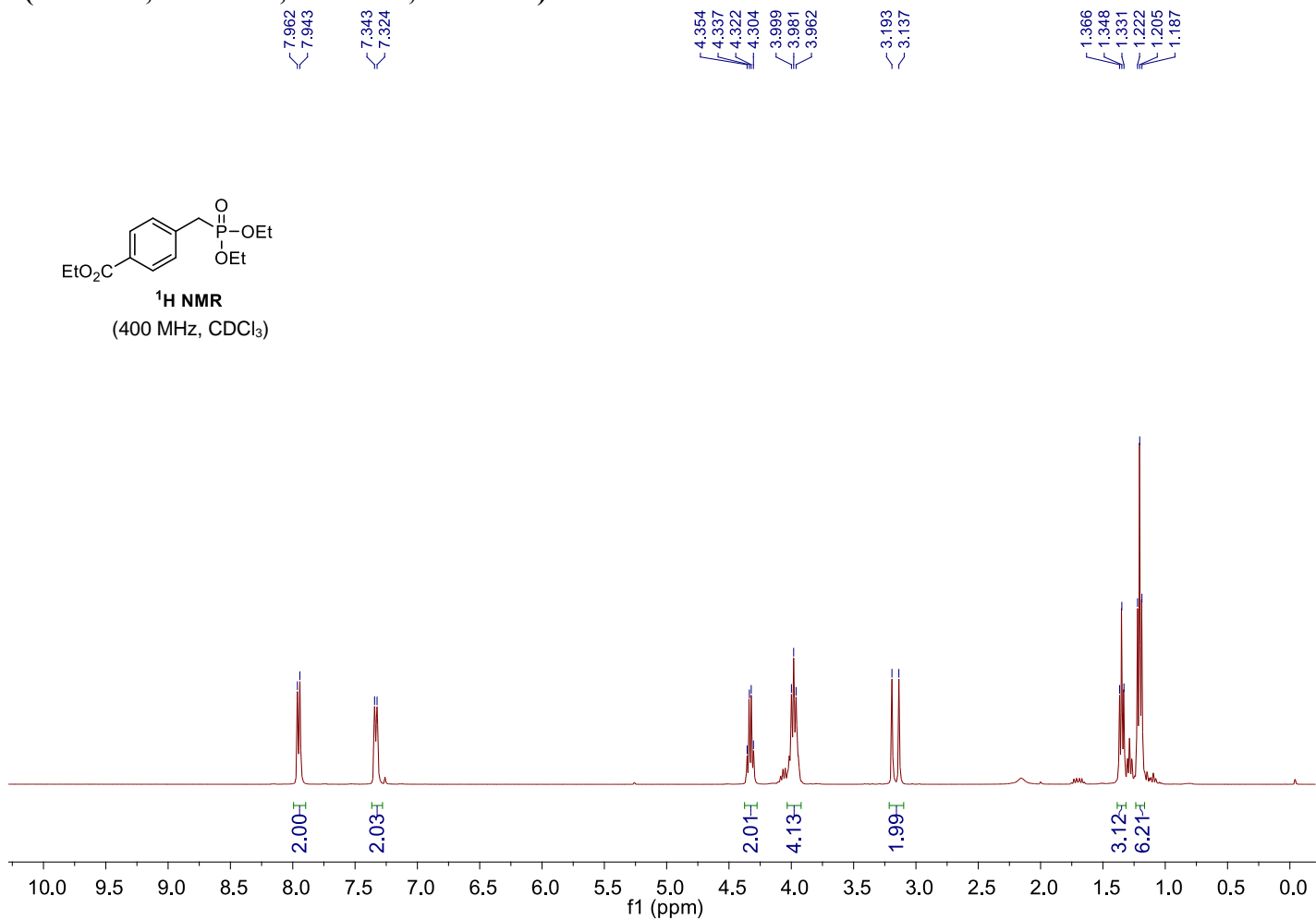
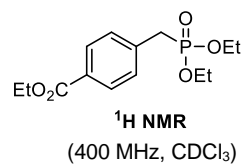
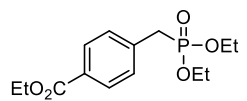


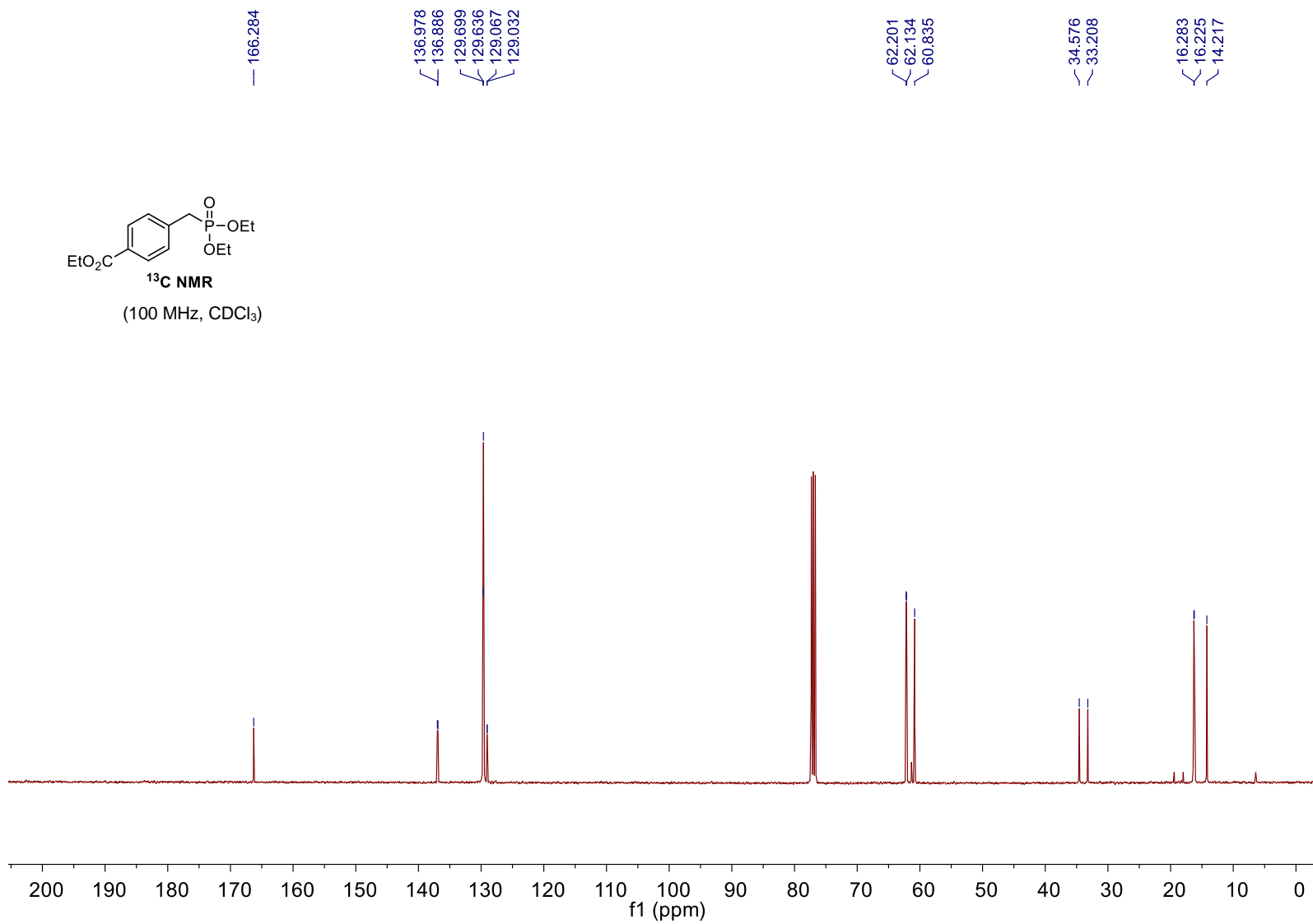
Fig. A6: Radio-TLC traces of (a) $[^{18}\text{F}]\mathbf{44}$ ($\text{RCC} = 68 \pm 2\%$) and (b) $[^{18}\text{F}]\mathbf{45}$ ($\text{RCC} = 57 \pm 9\%$). Developing solvent: MeOH.

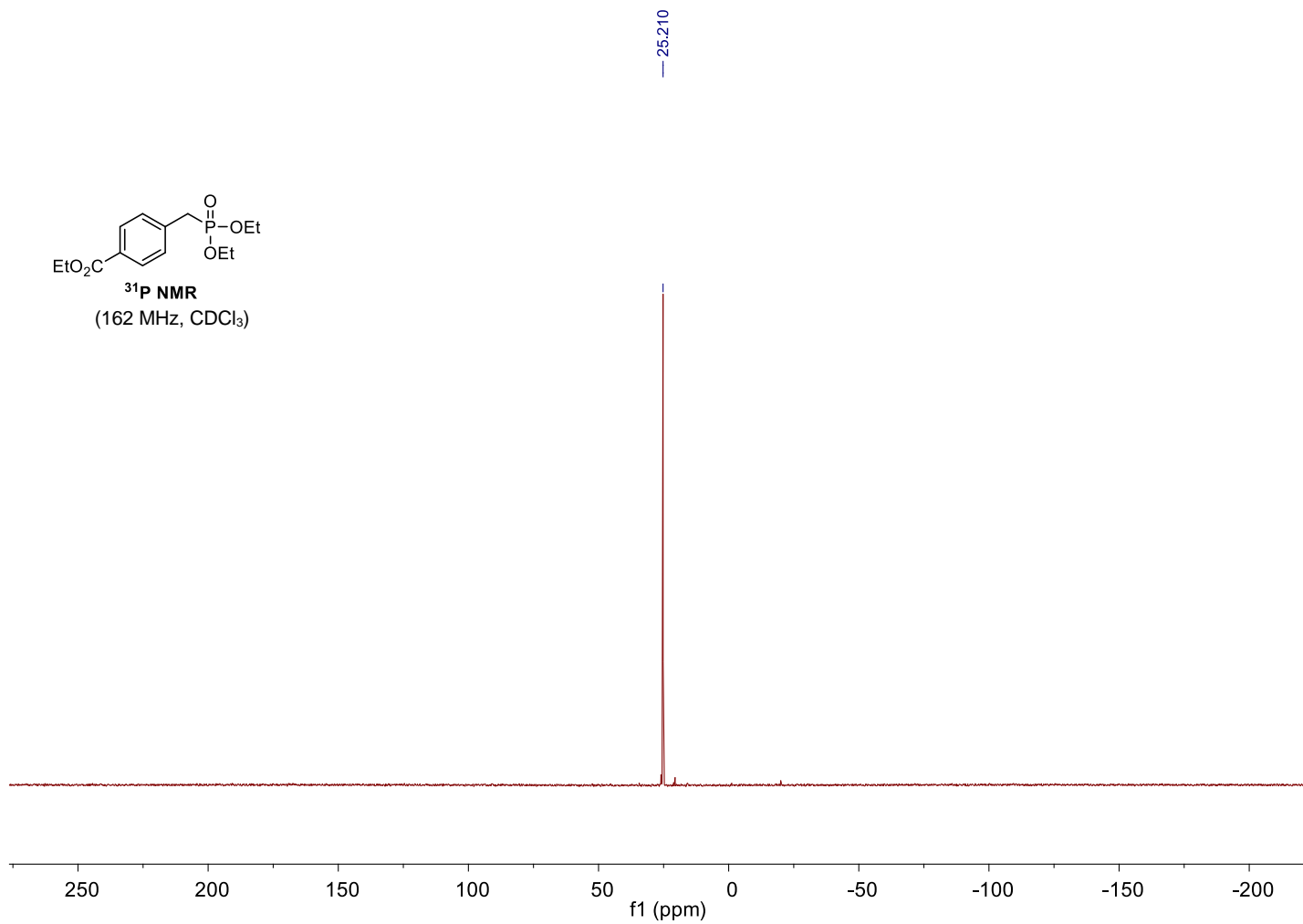
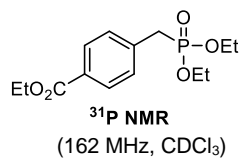
3.2 NMR spectra (^1H NMR, ^{13}C NMR, ^{31}P NMR, ^{19}F NMR)



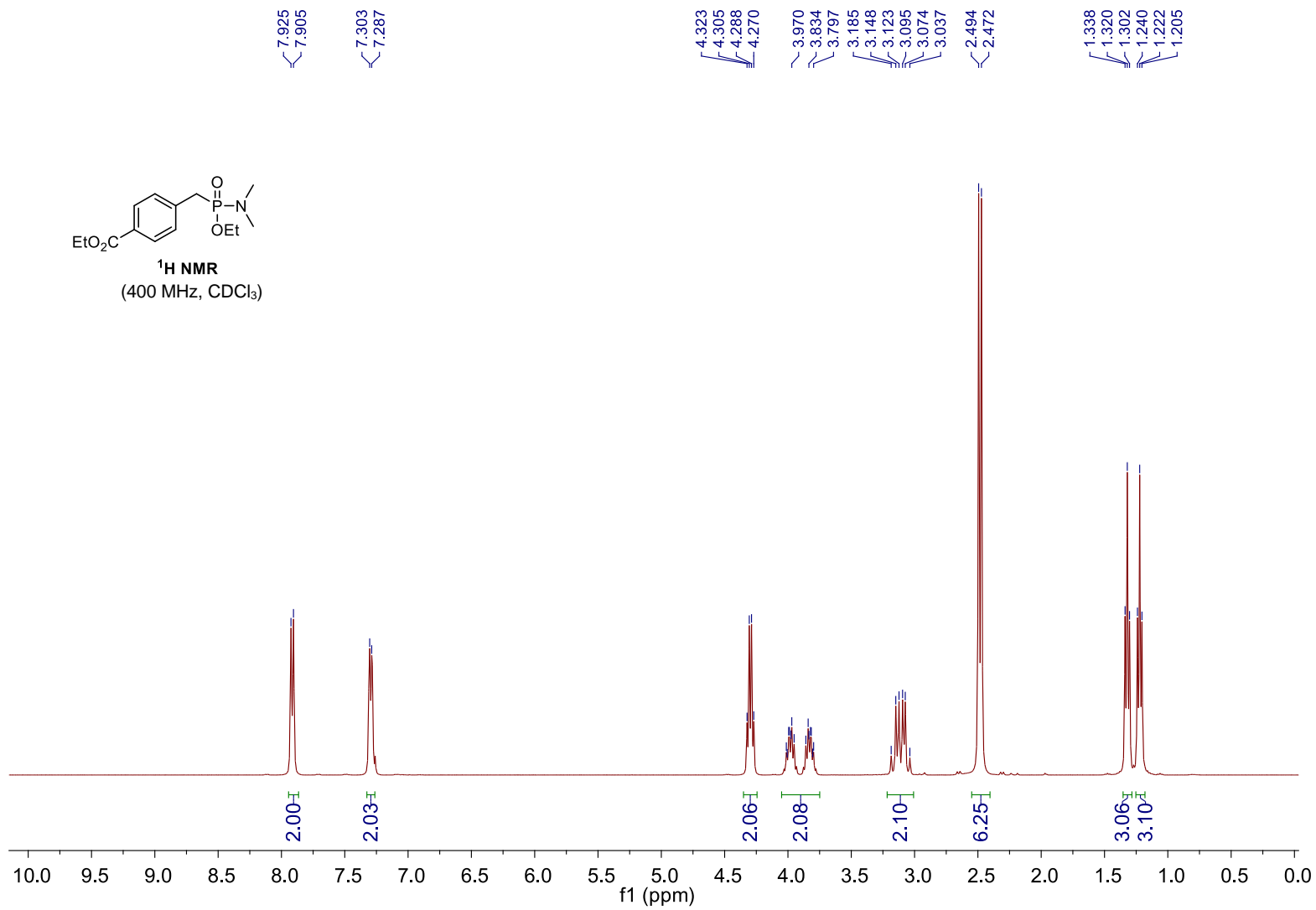
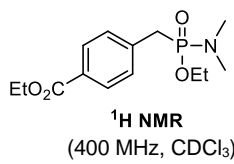


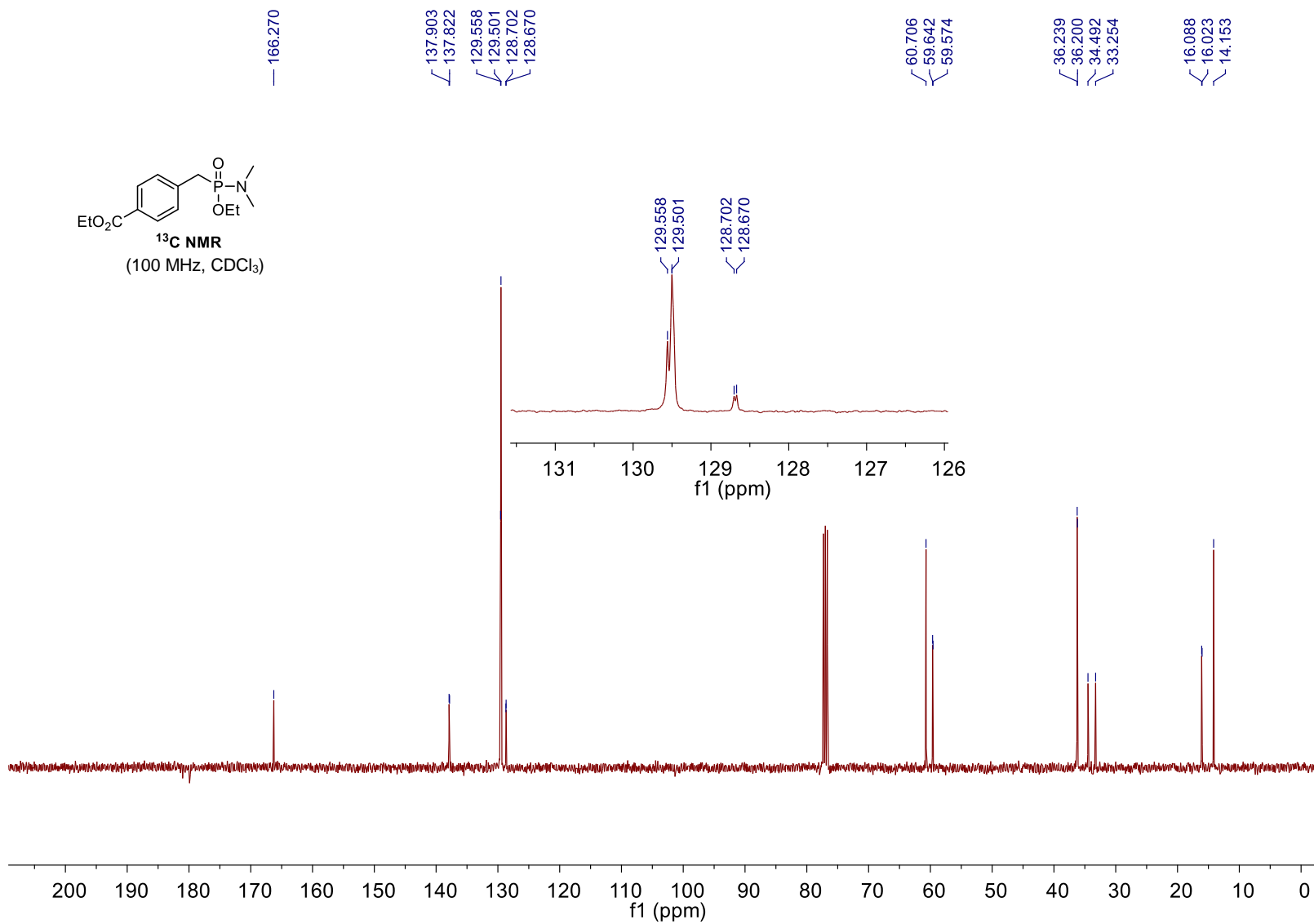
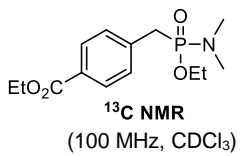
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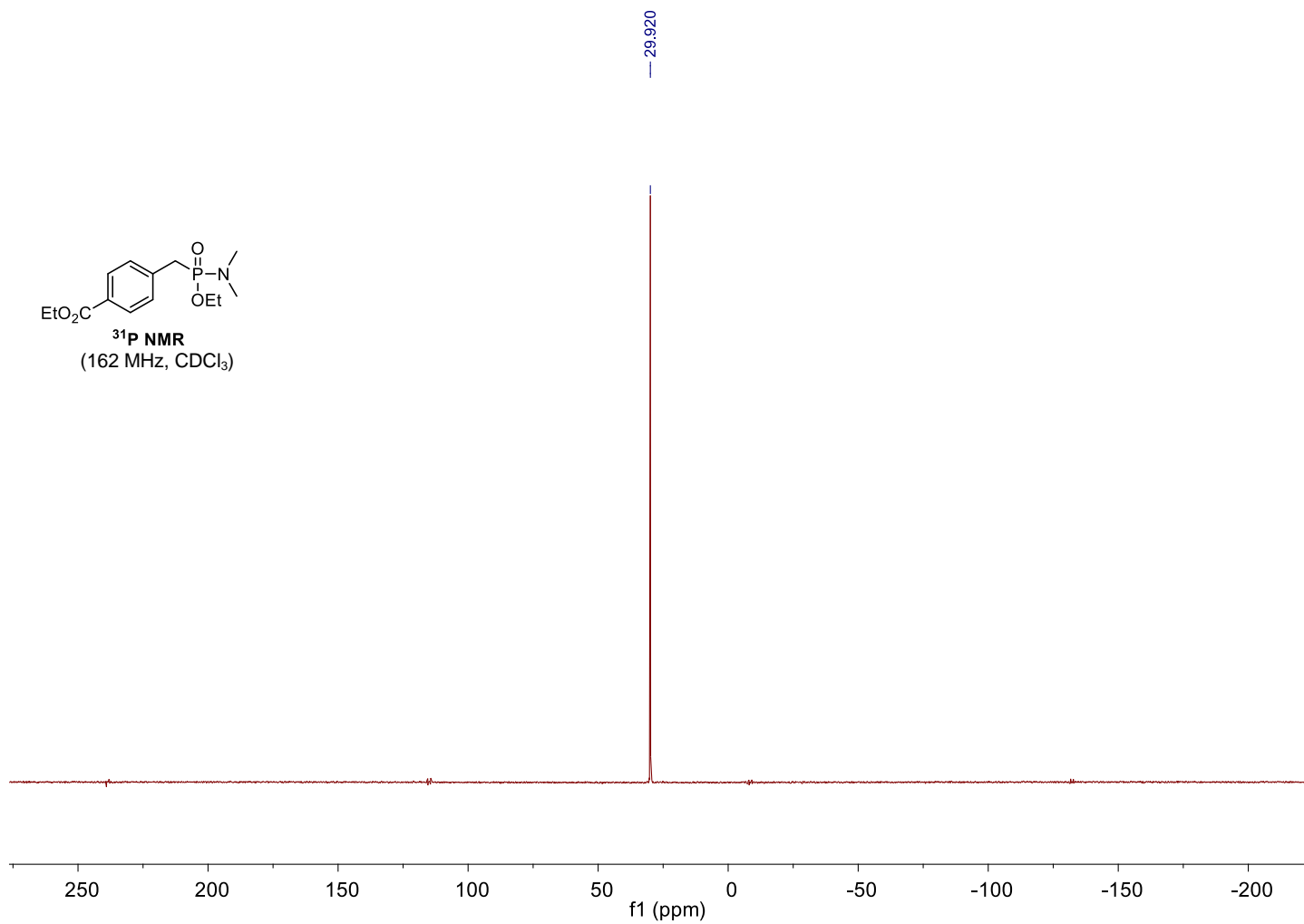
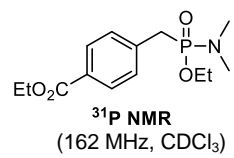




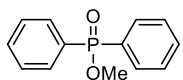
S108



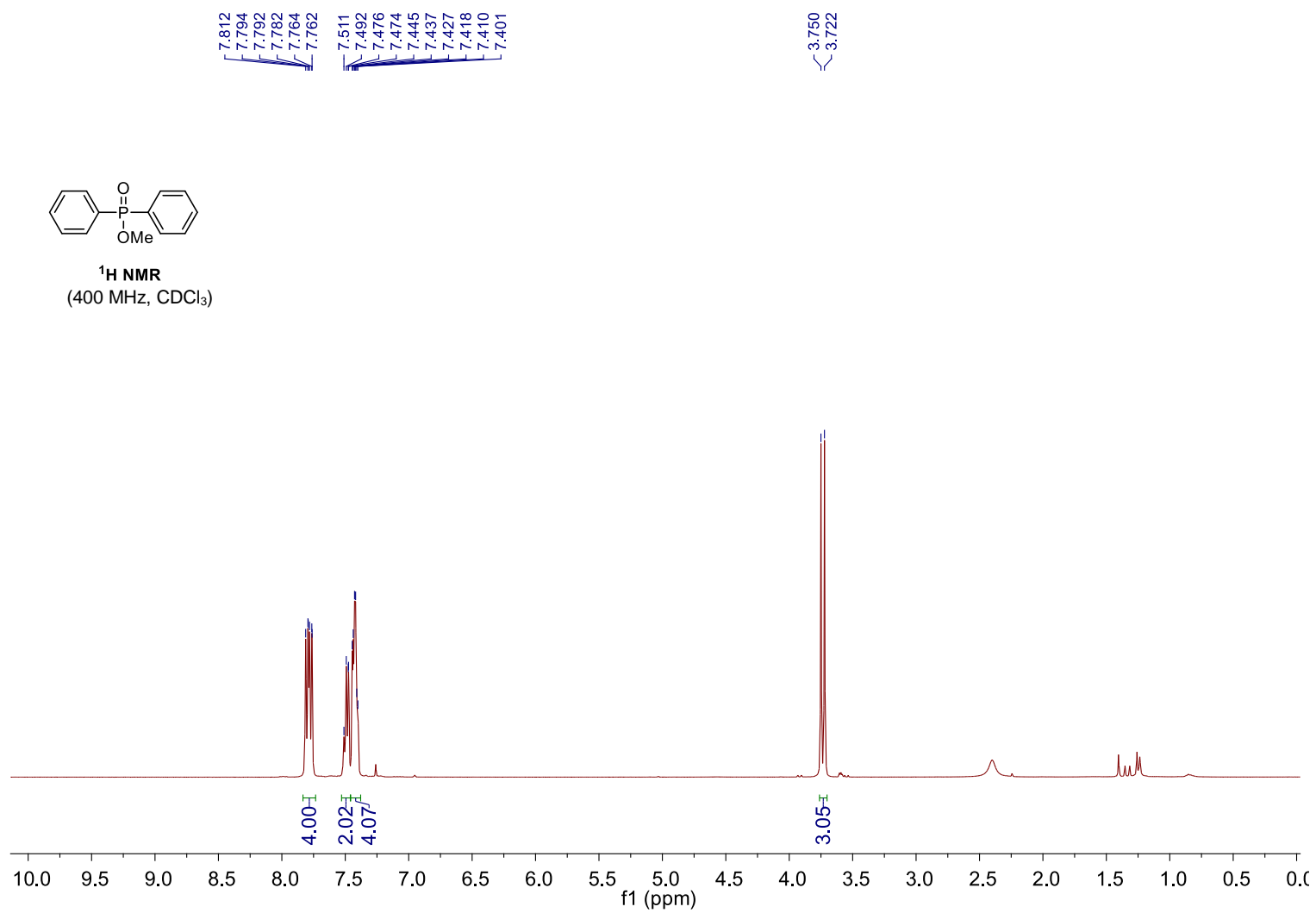


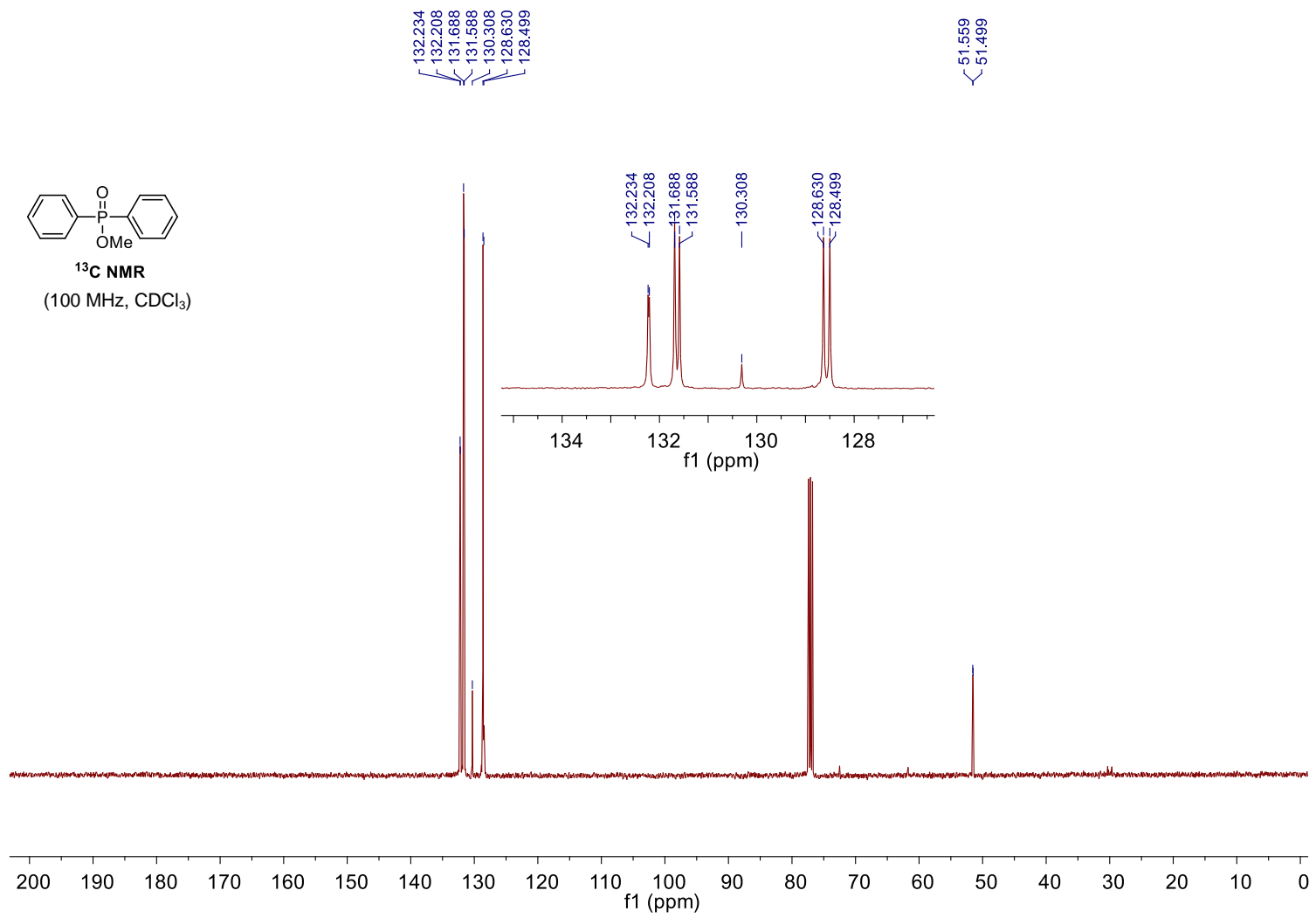
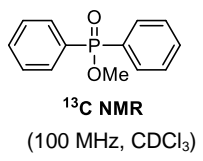


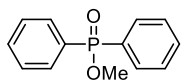
S111



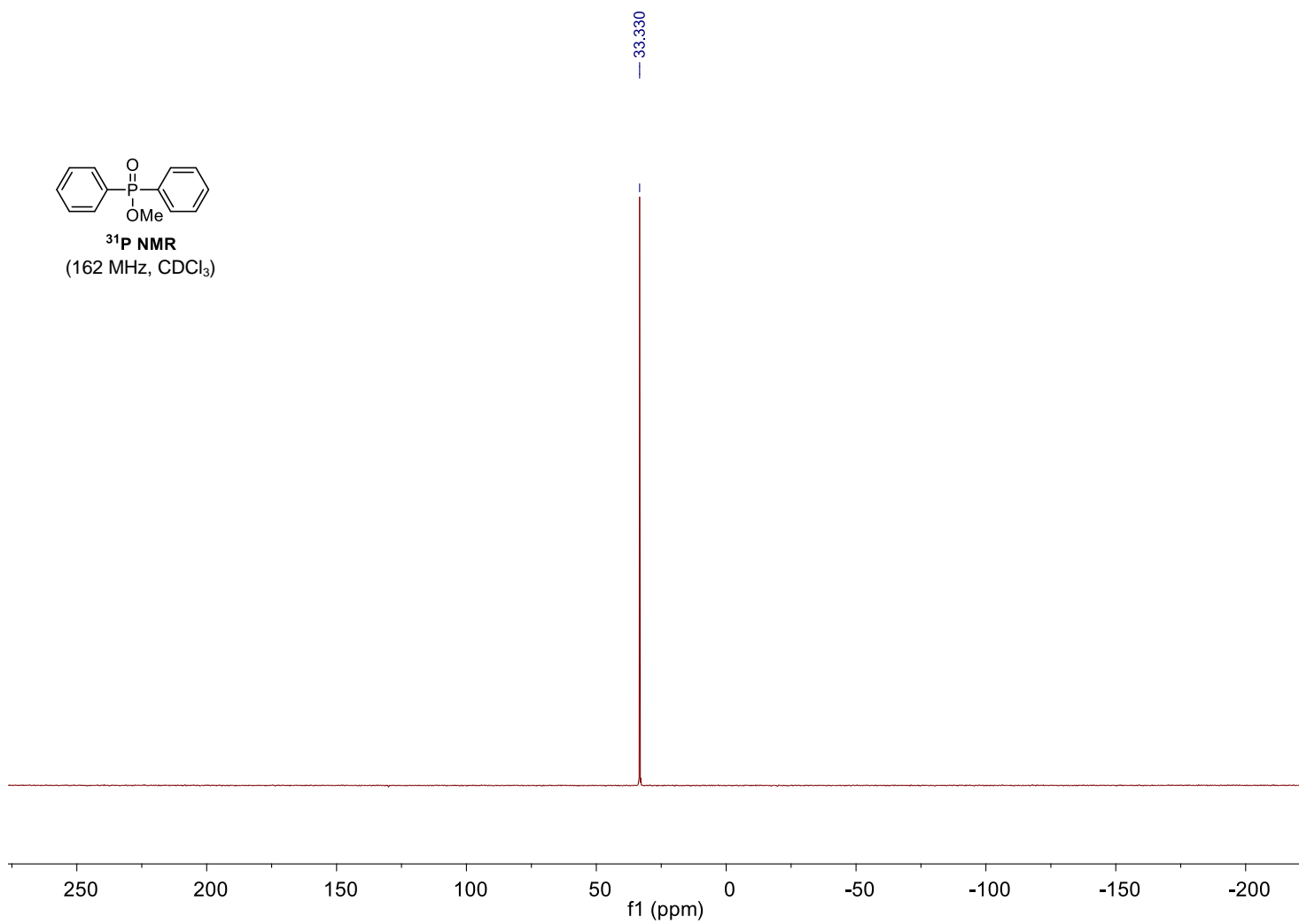
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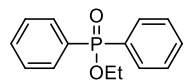




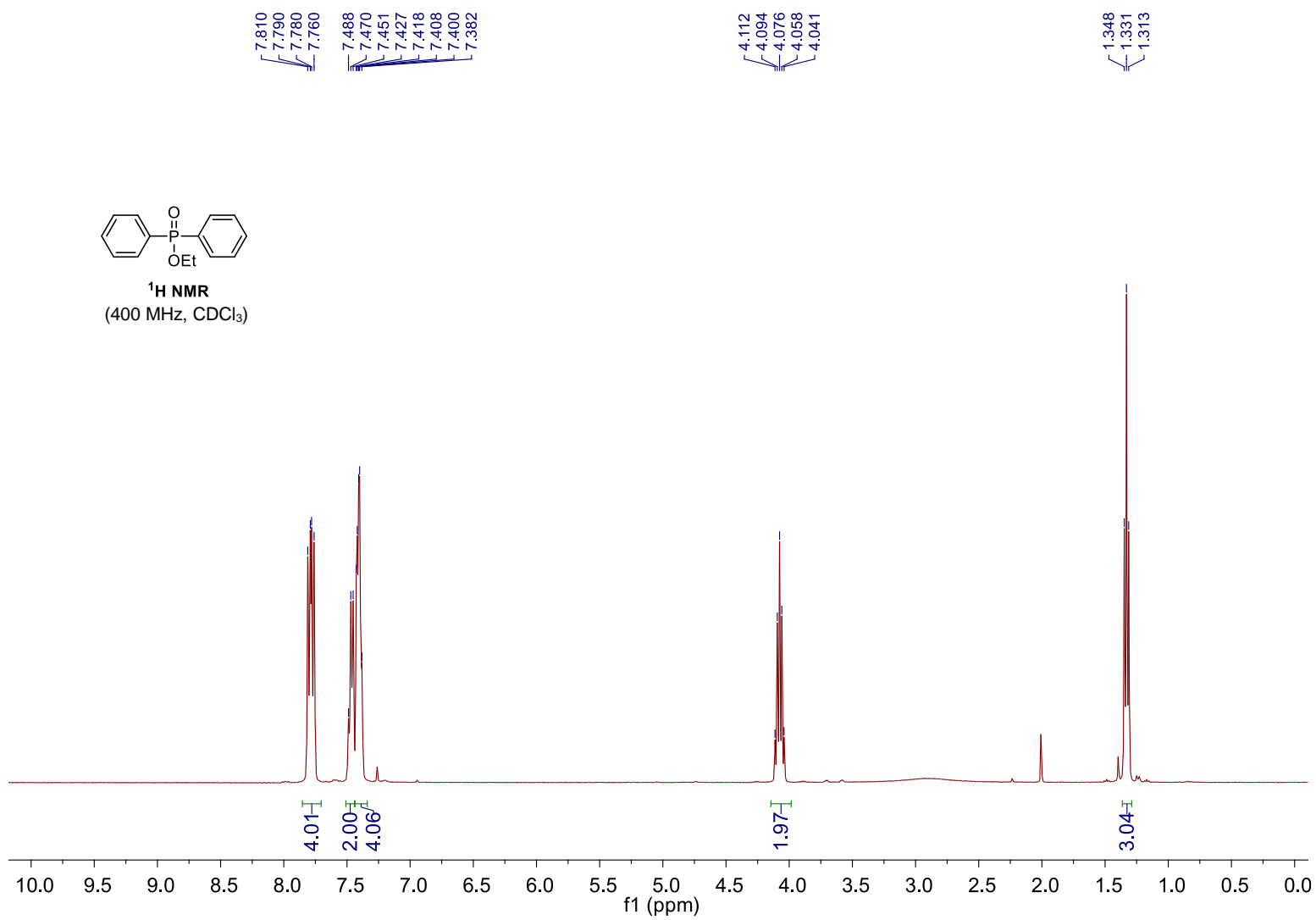
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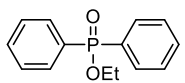


S114

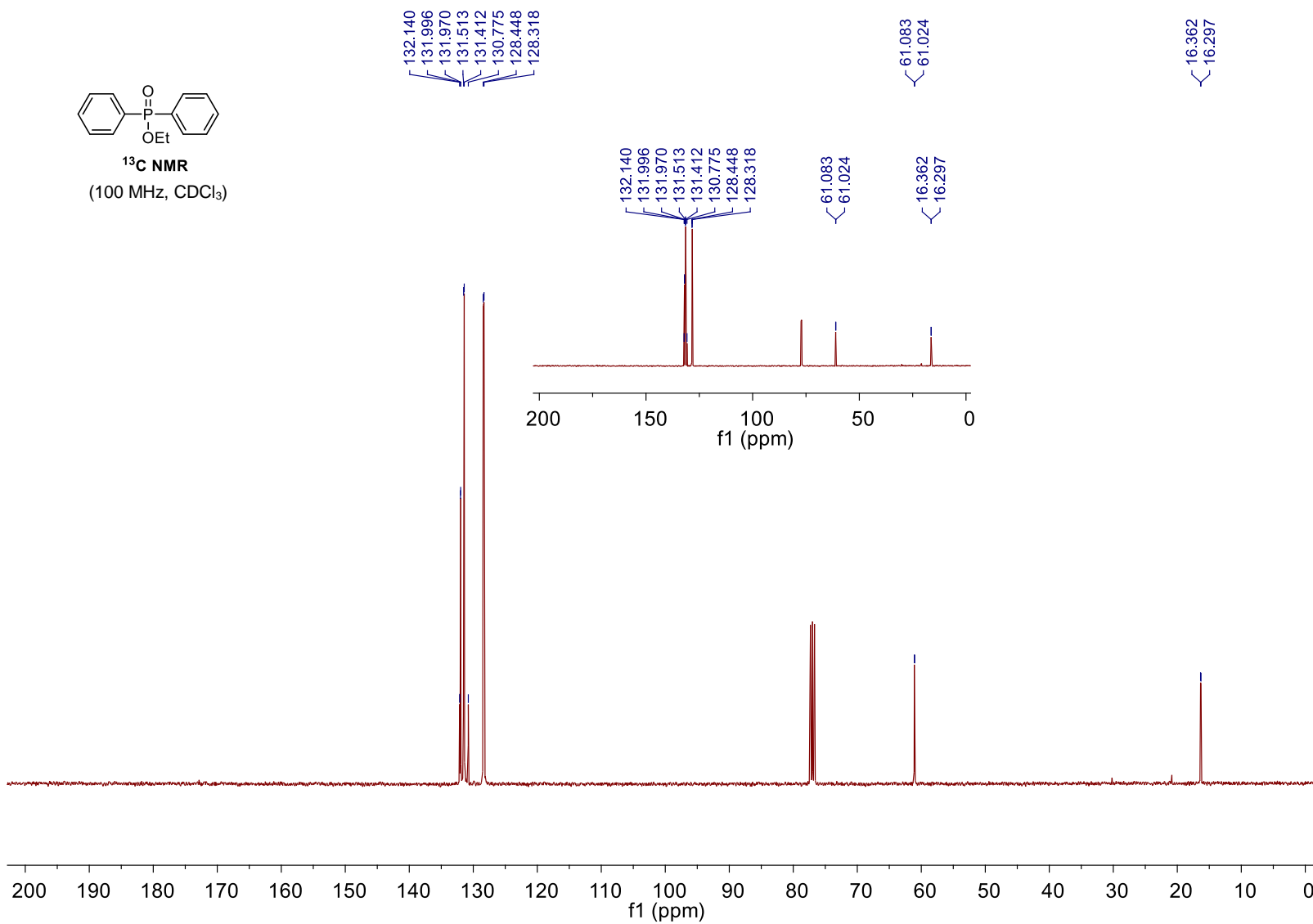


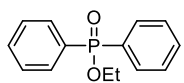
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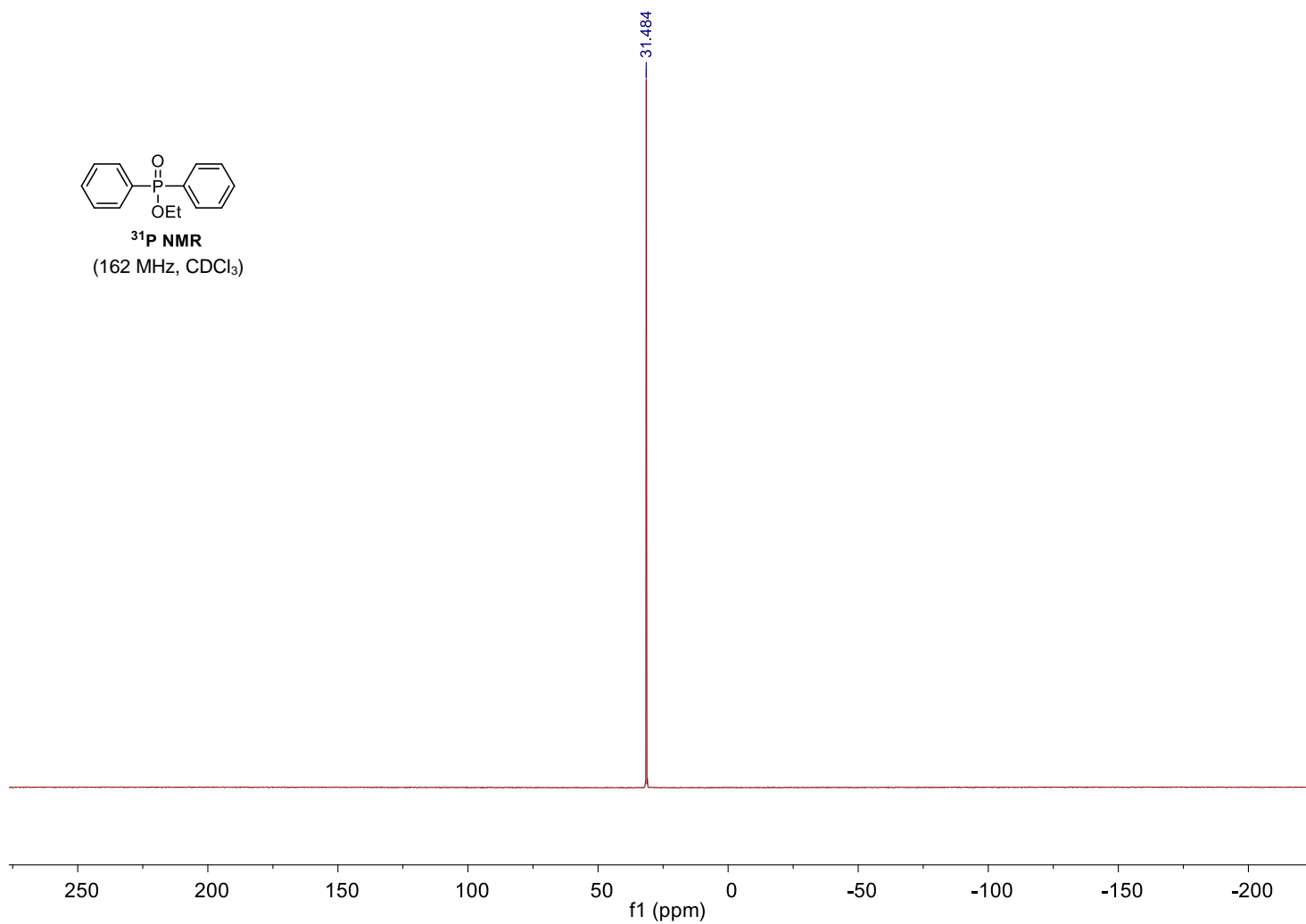


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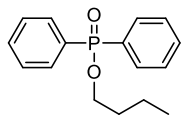




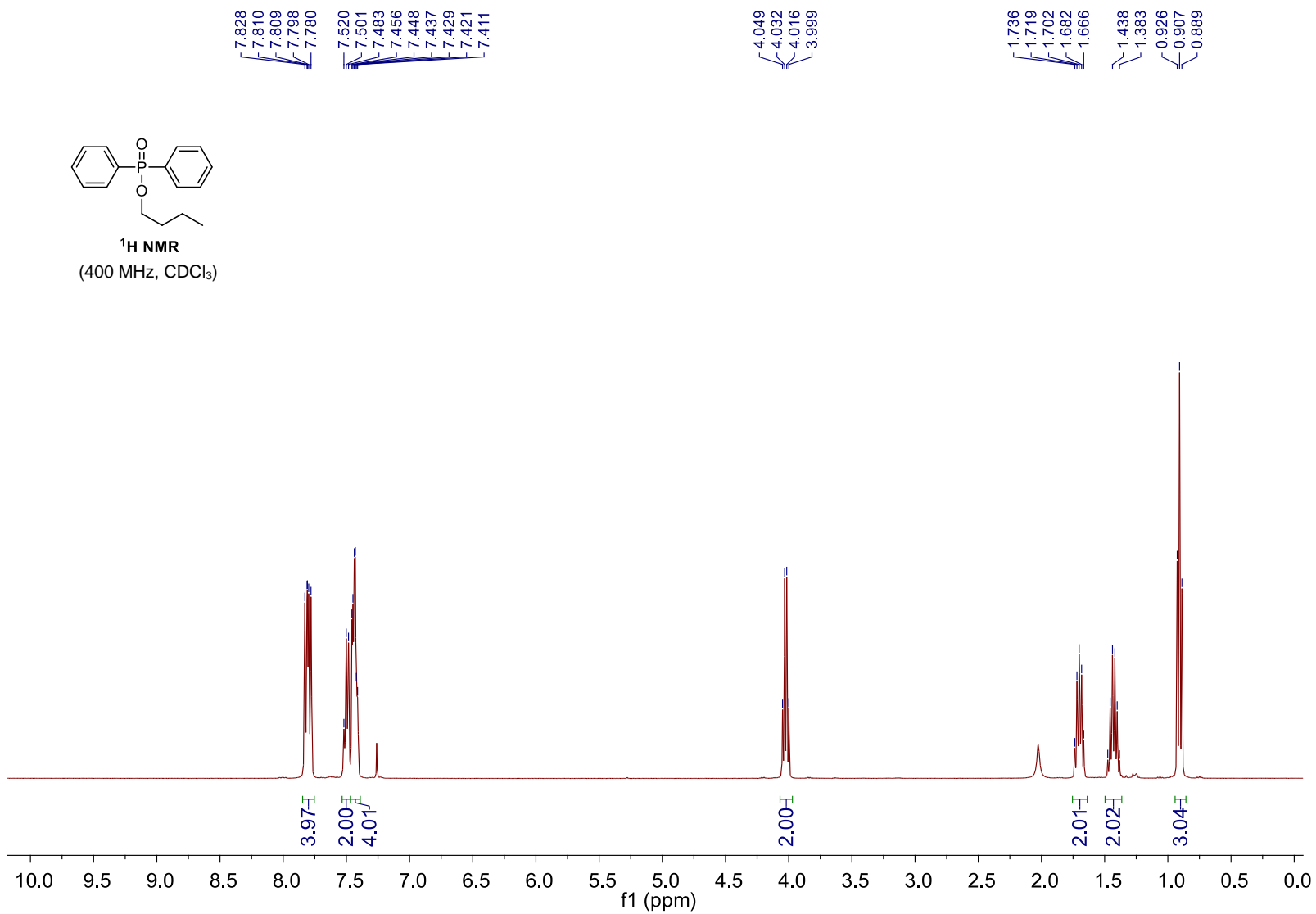
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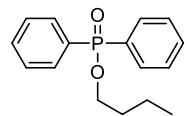


S117

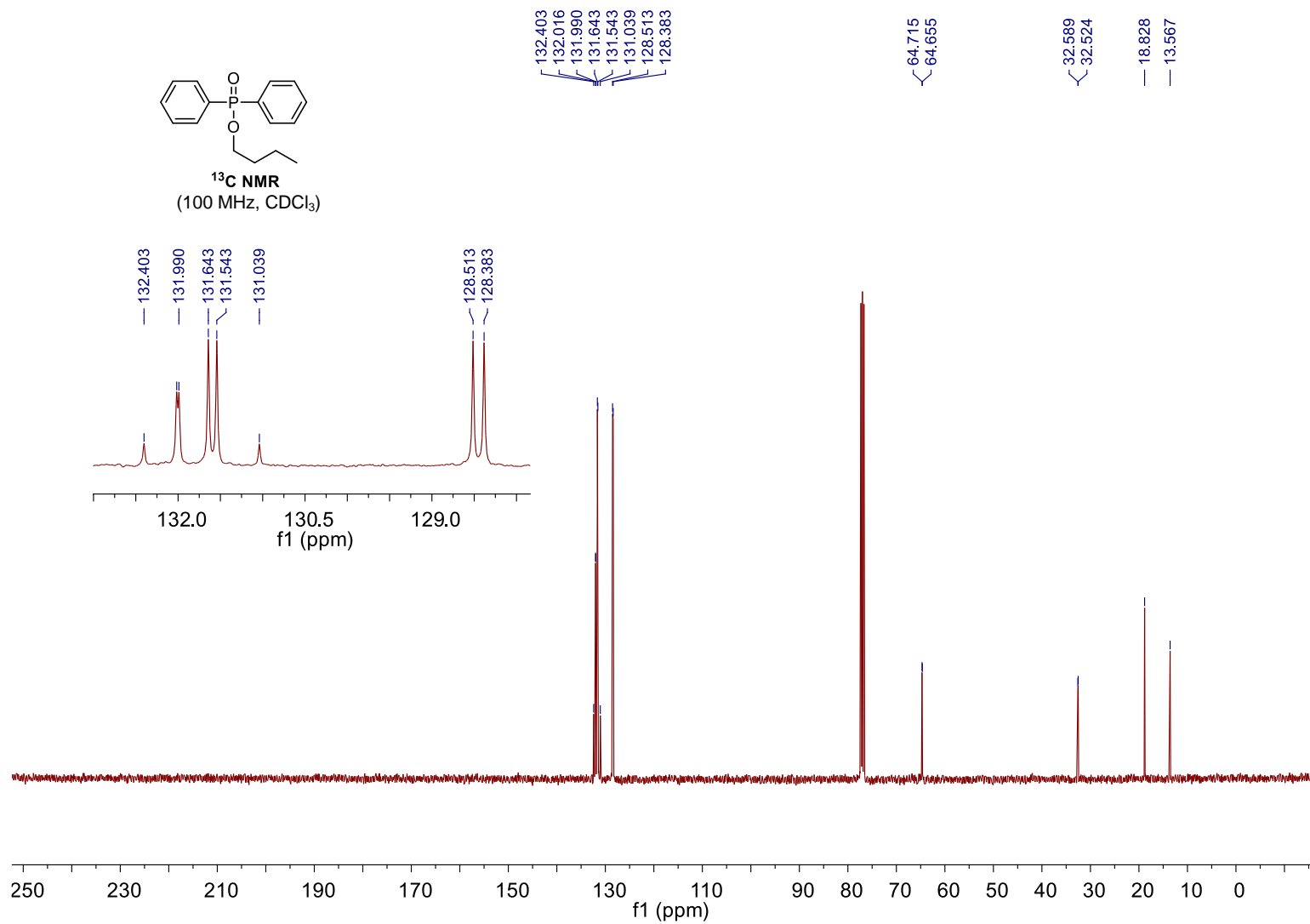


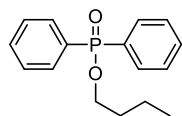
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(400 MHz, CDCl₃)



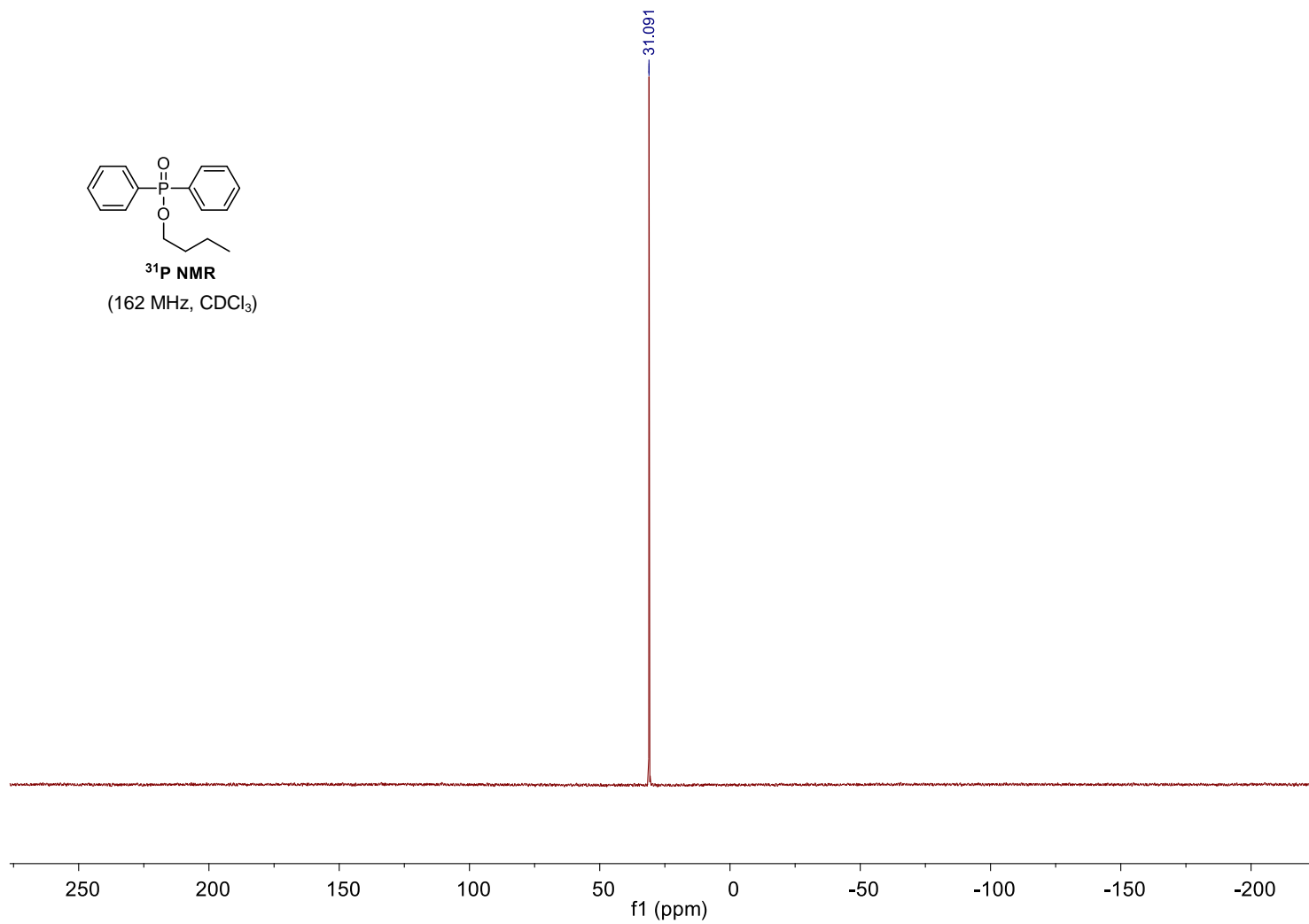


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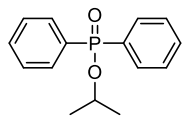




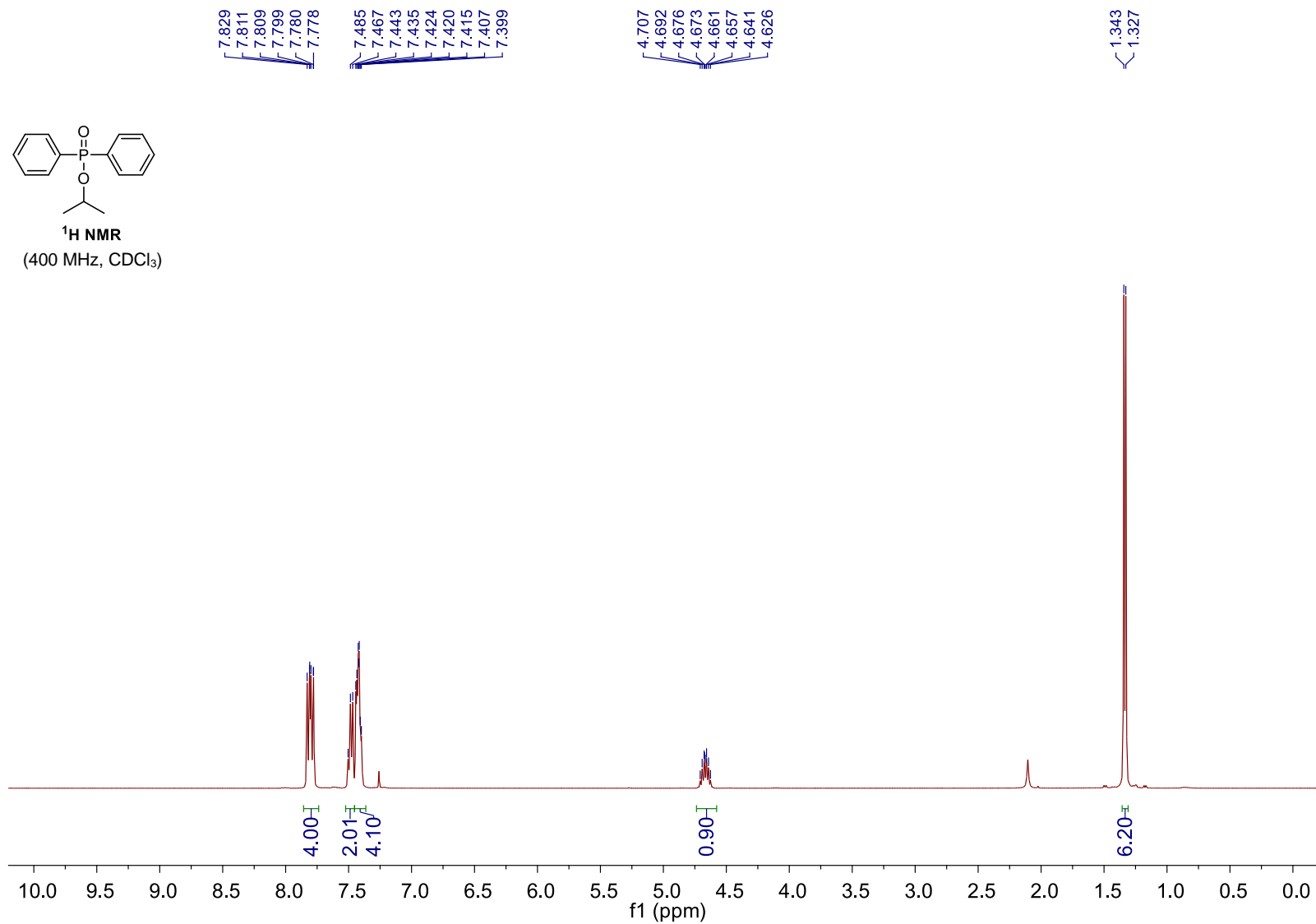
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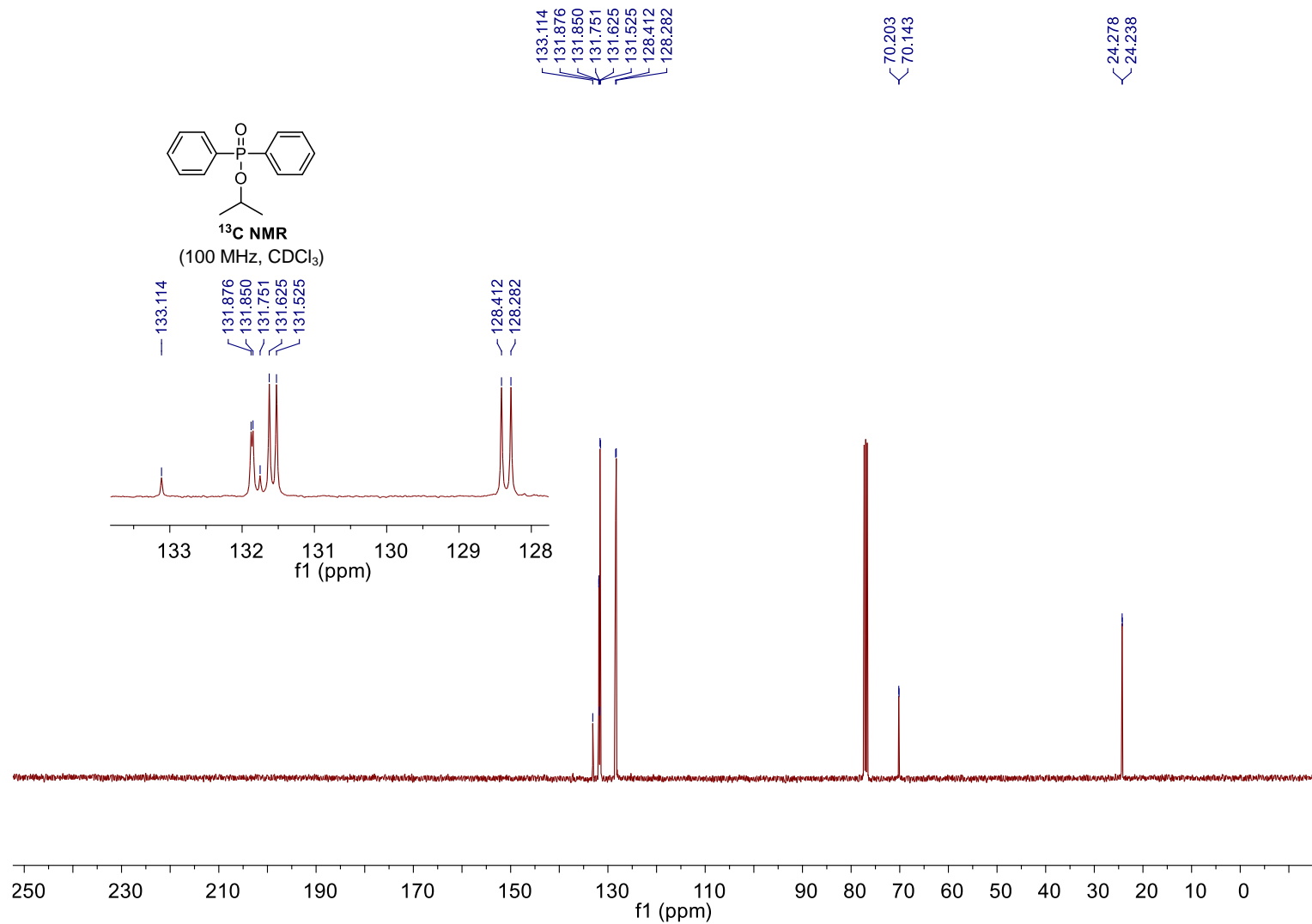


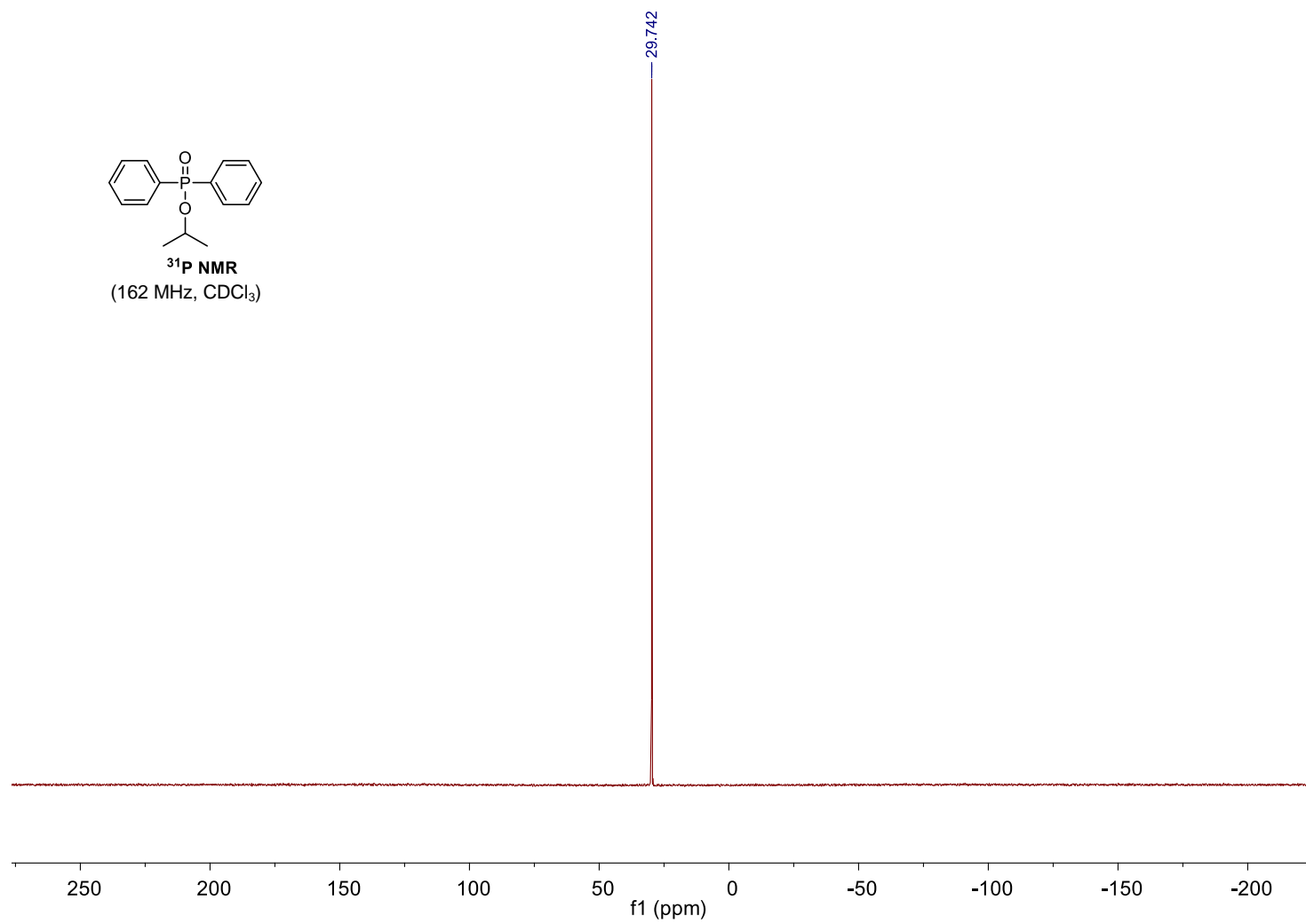
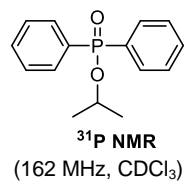
S120

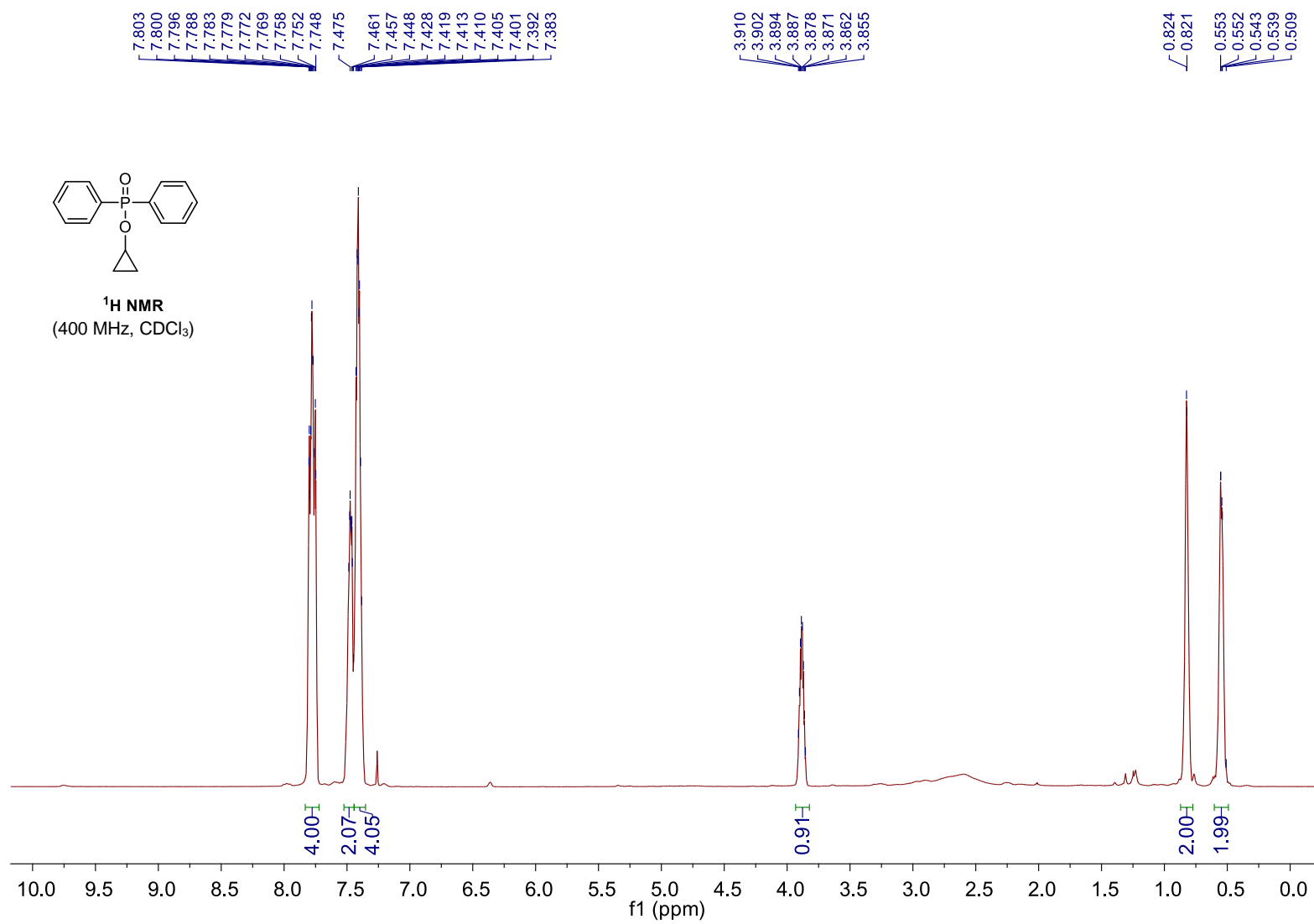


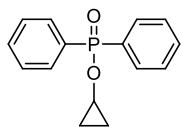
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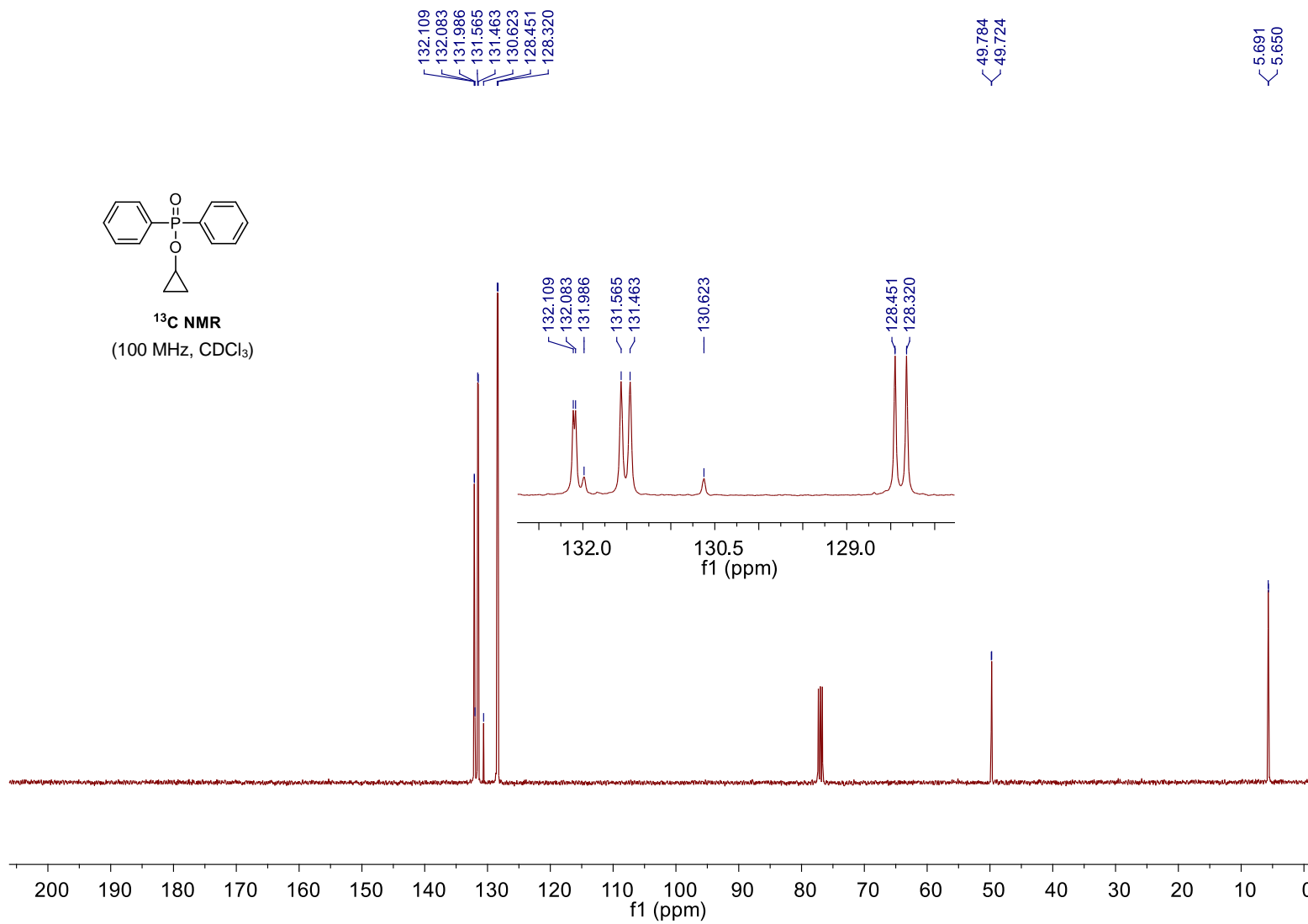


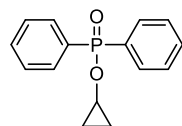




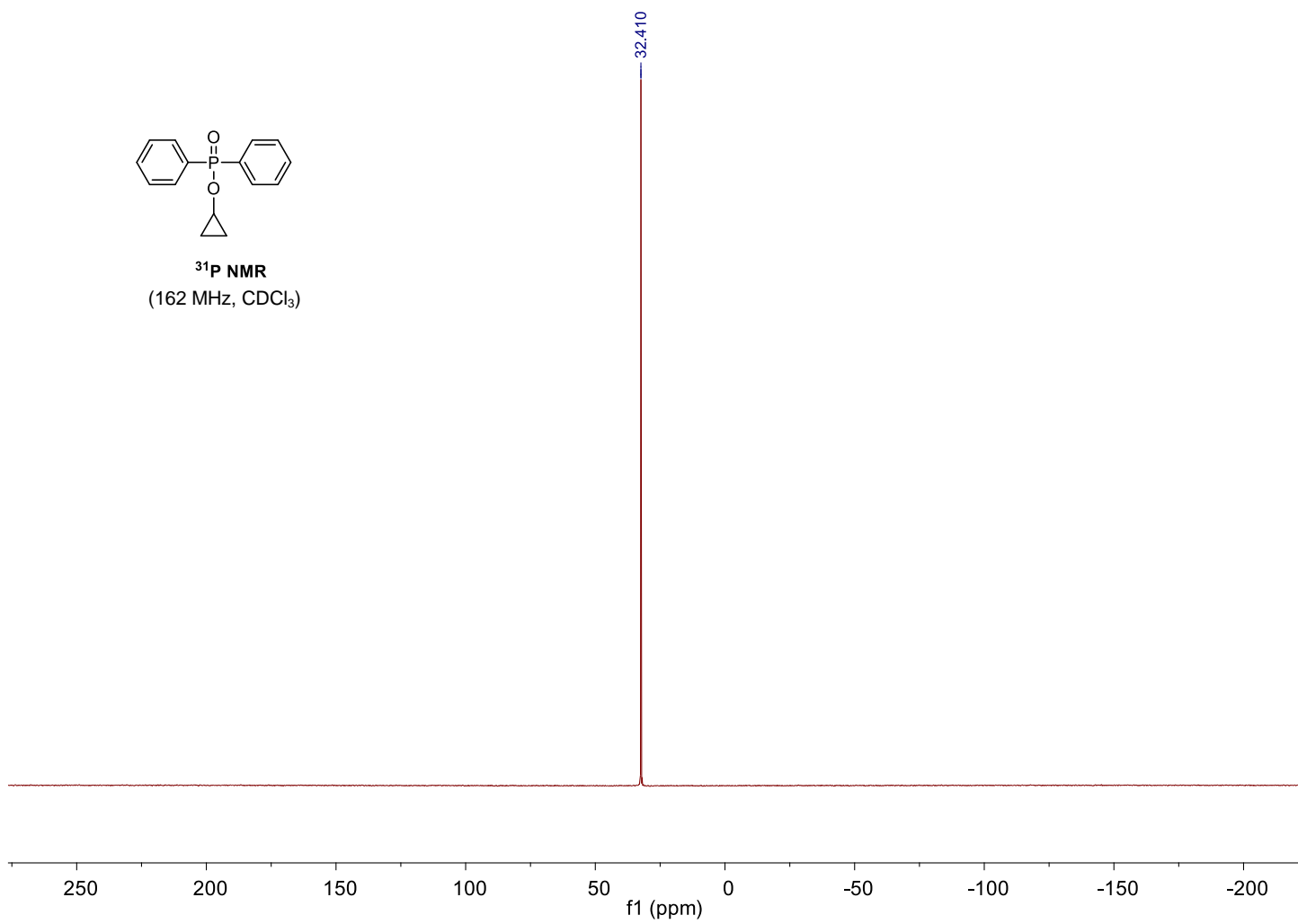


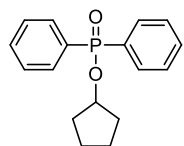
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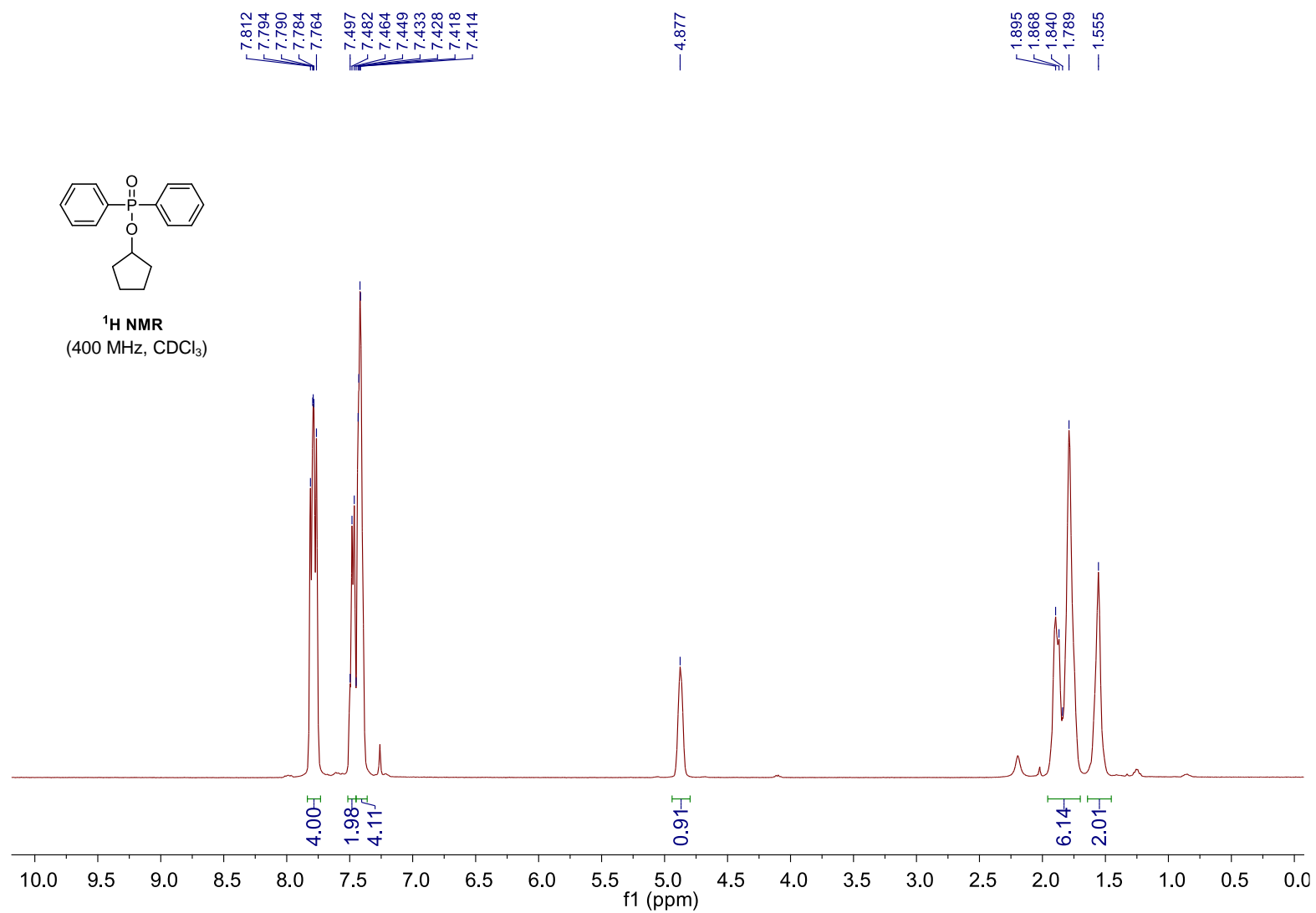


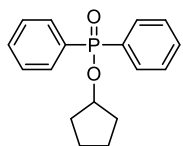
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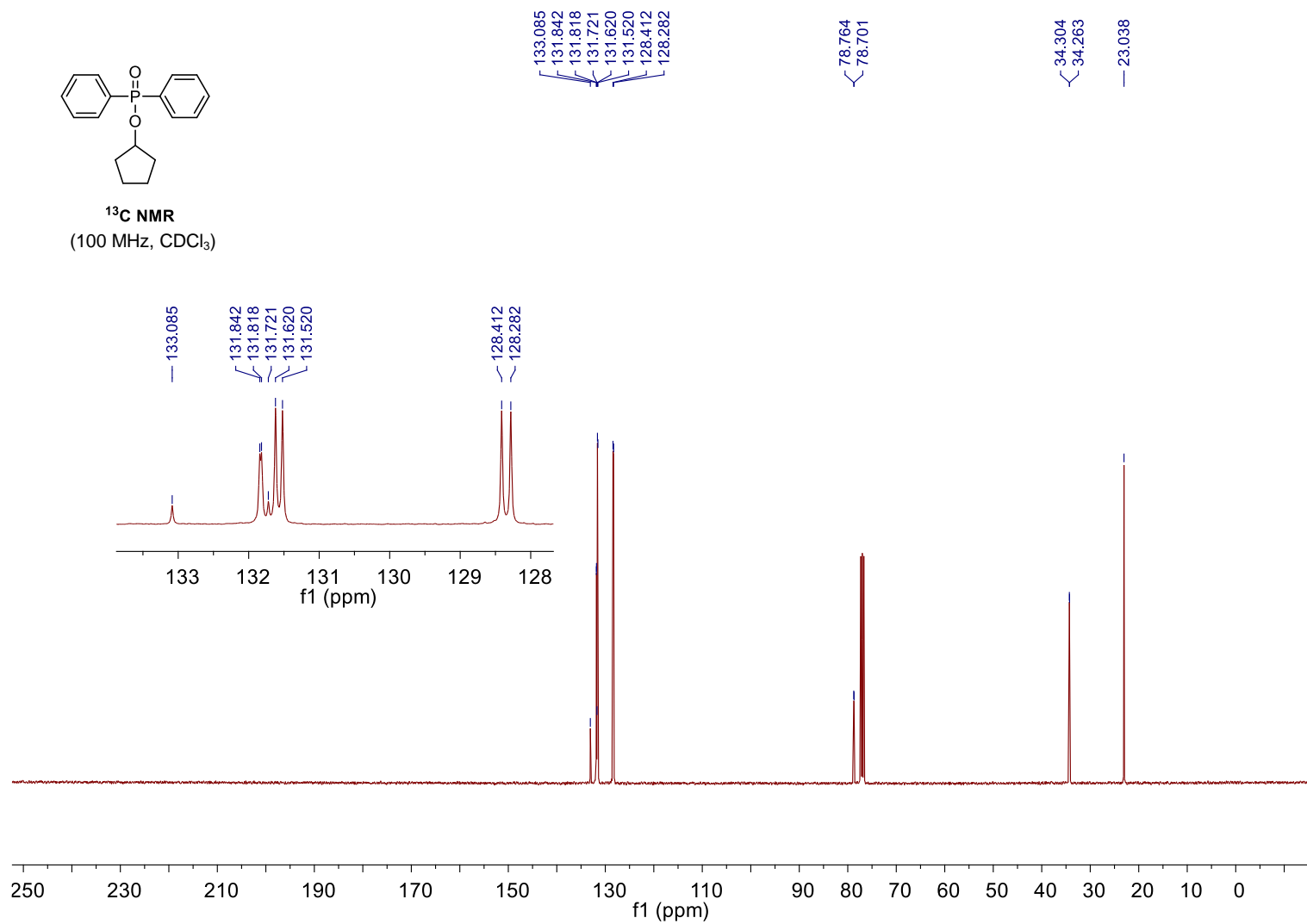


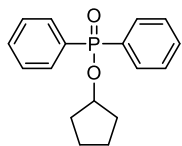
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(400 MHz, CDCl₃)



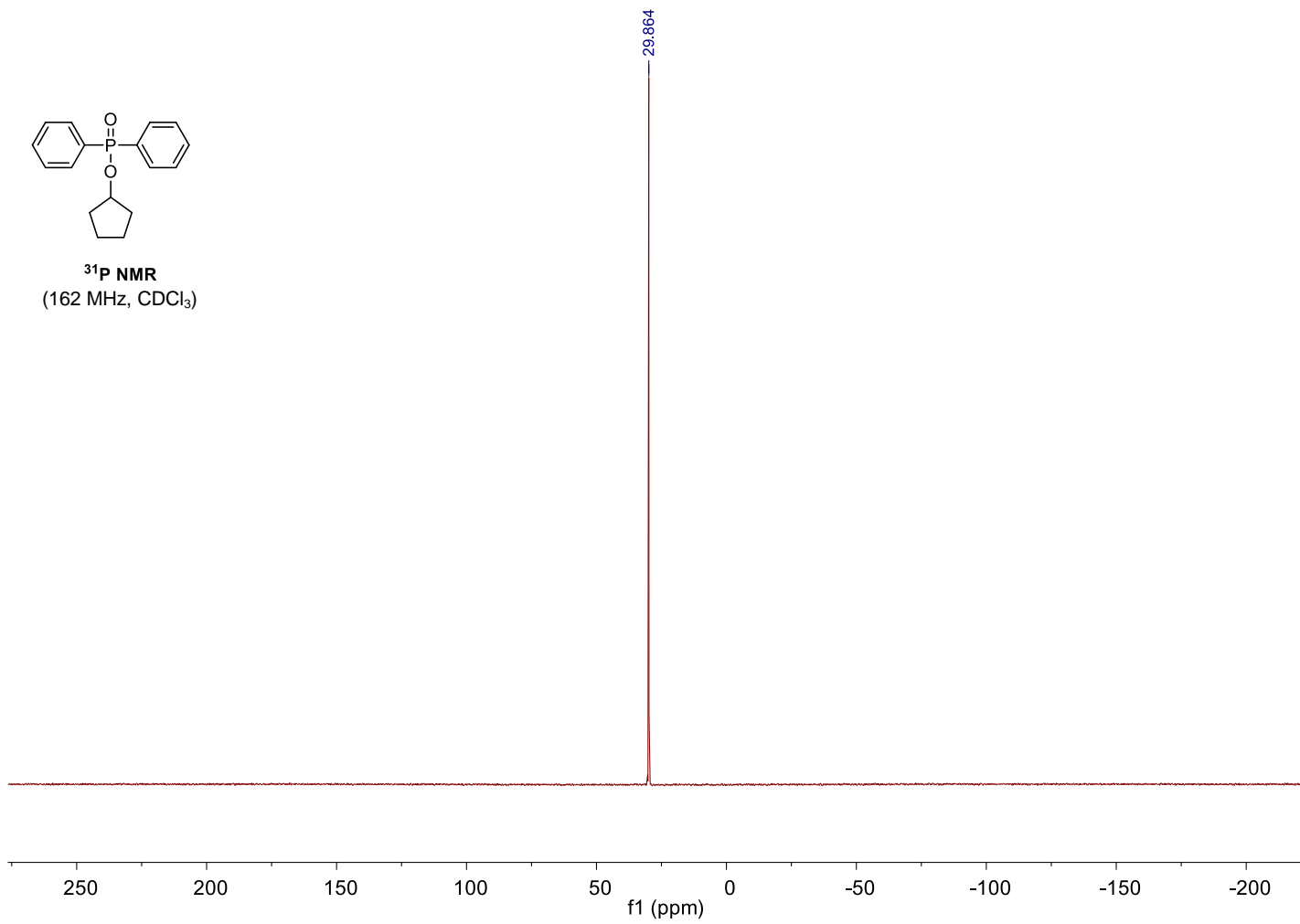


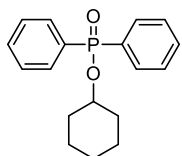
¹³C NMR
(100 MHz, CDCl₃)



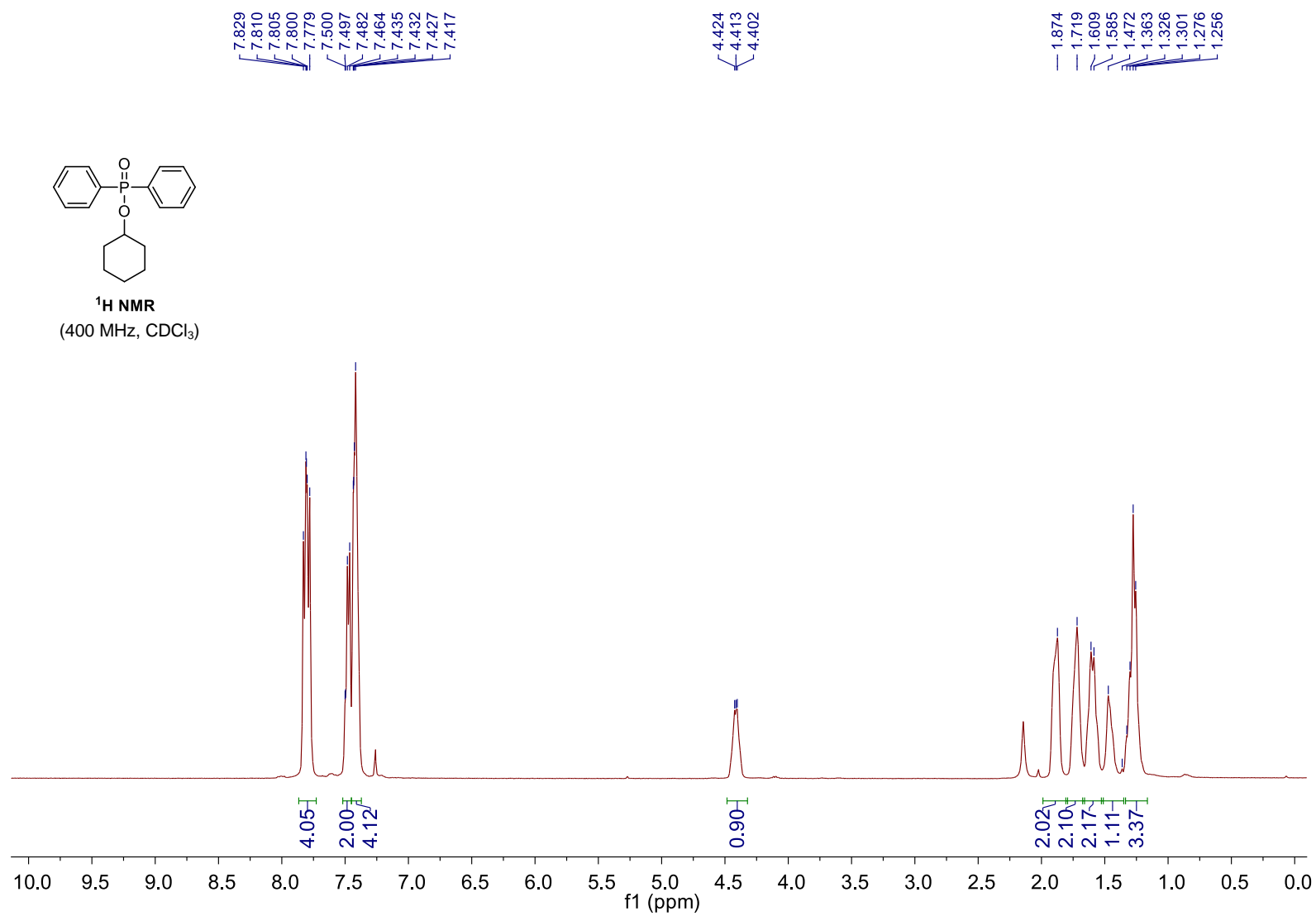


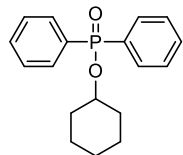
³¹P NMR
(162 MHz, CDCl₃)





¹H NMR
(400 MHz, CDCl₃)





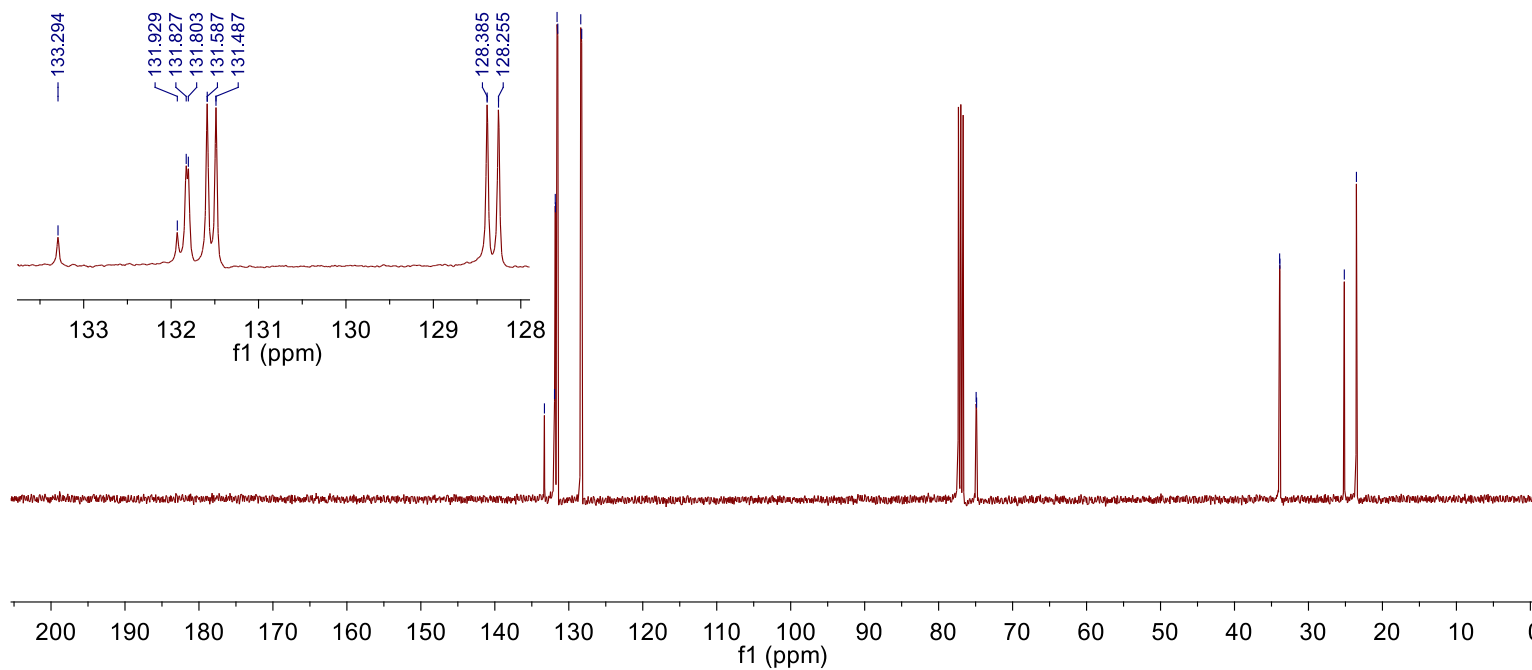
¹³C NMR
(100 MHz, CDCl₃)

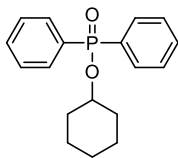
133.294
131.929
131.827
131.803
131.587
131.487
128.385
128.255

74.930
74.869

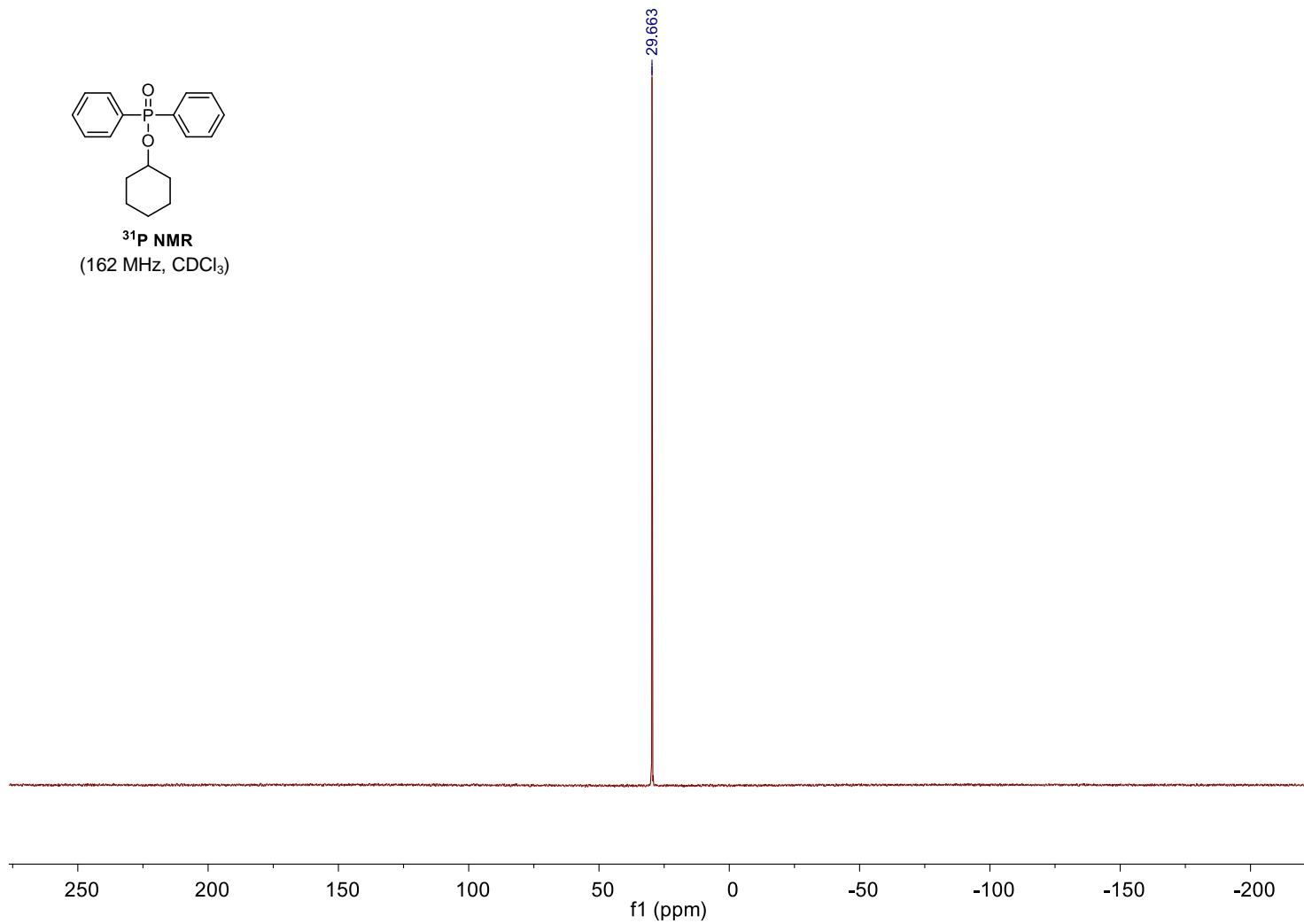
33.900
33.865

25.154
23.517

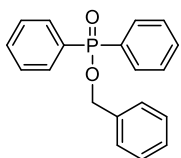




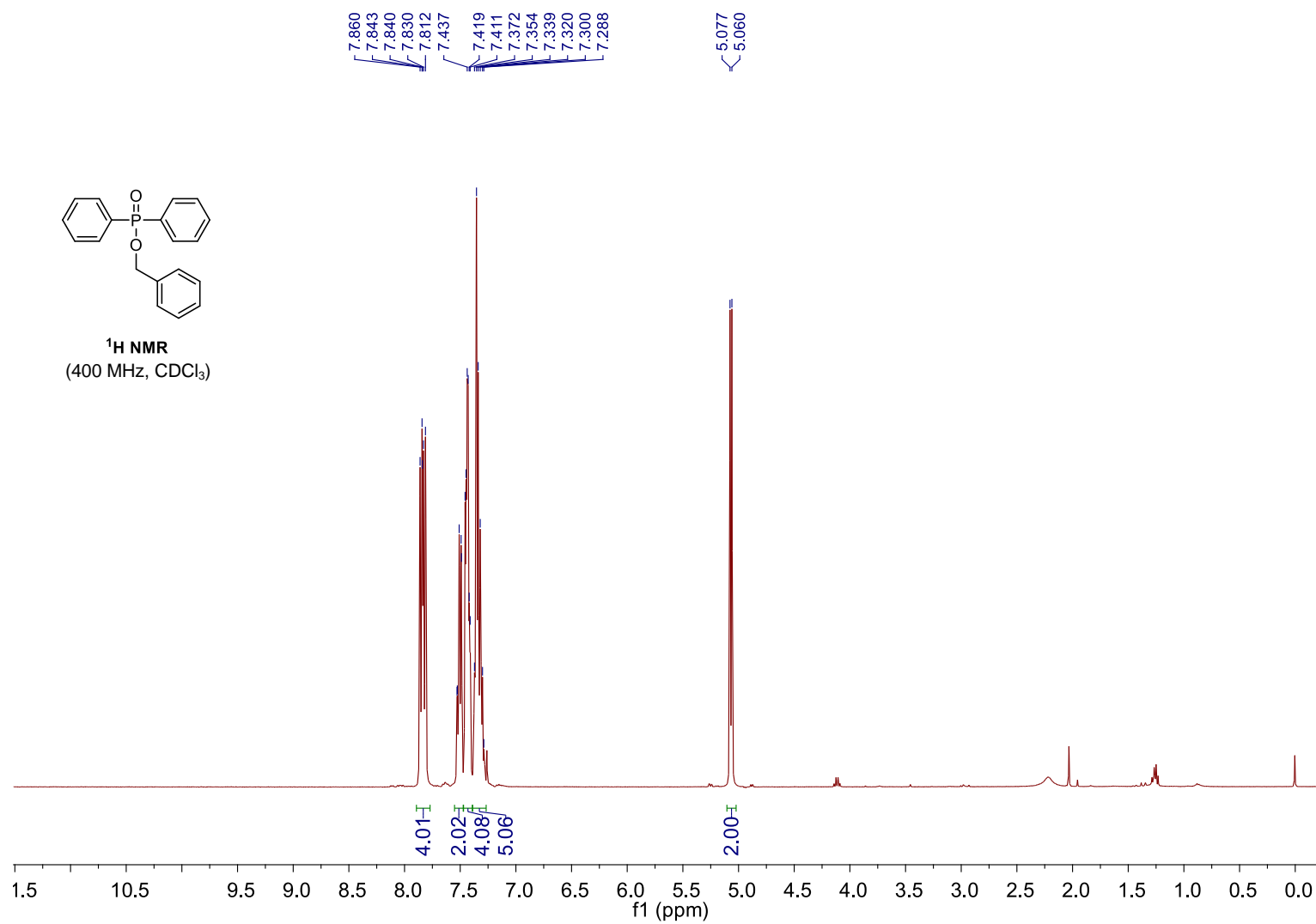
³¹P NMR
(162 MHz, CDCl₃)

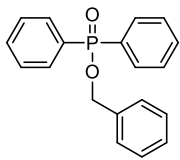


S132

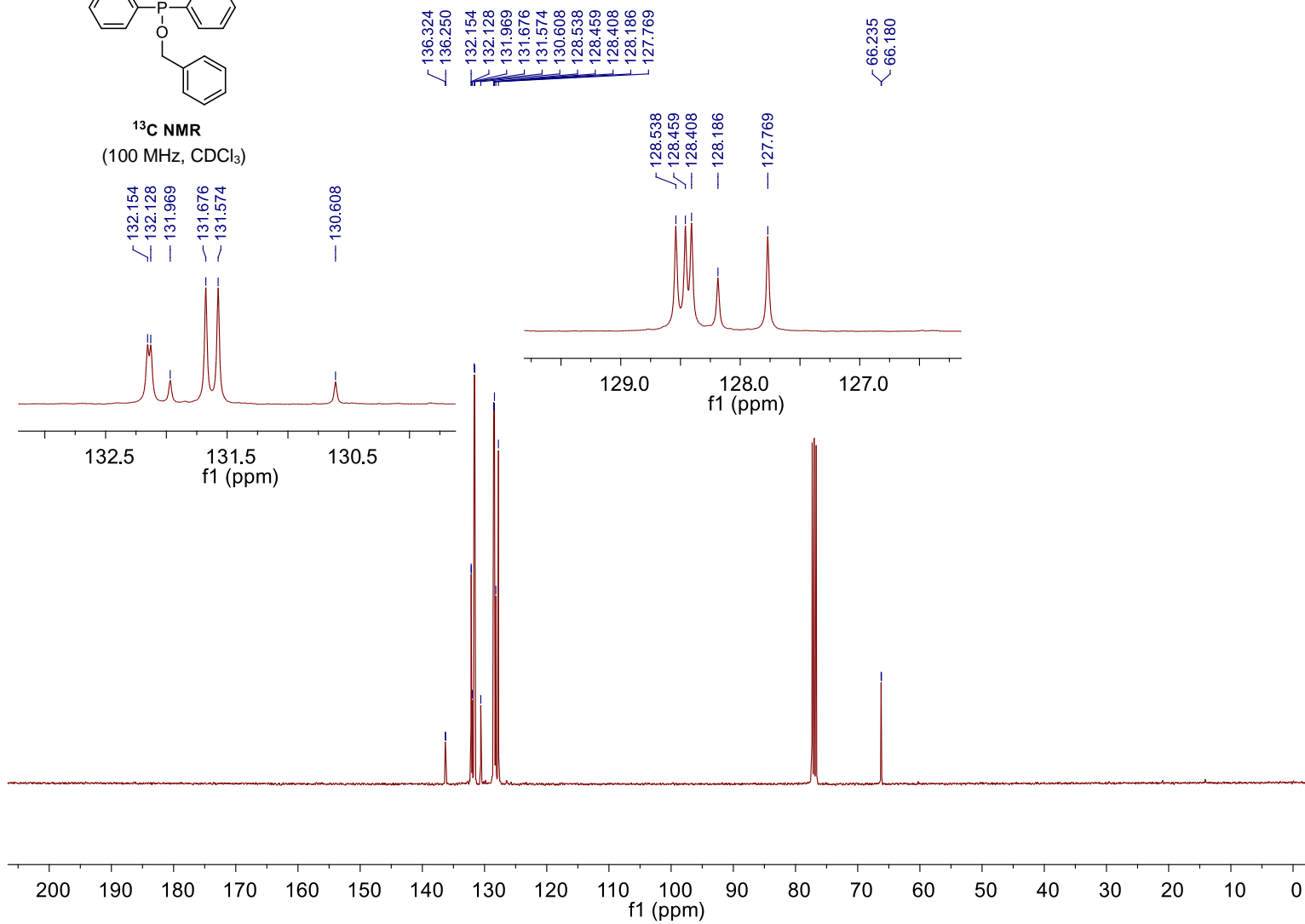


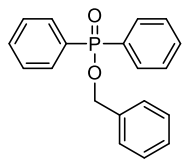
¹H NMR
(400 MHz, CDCl₃)



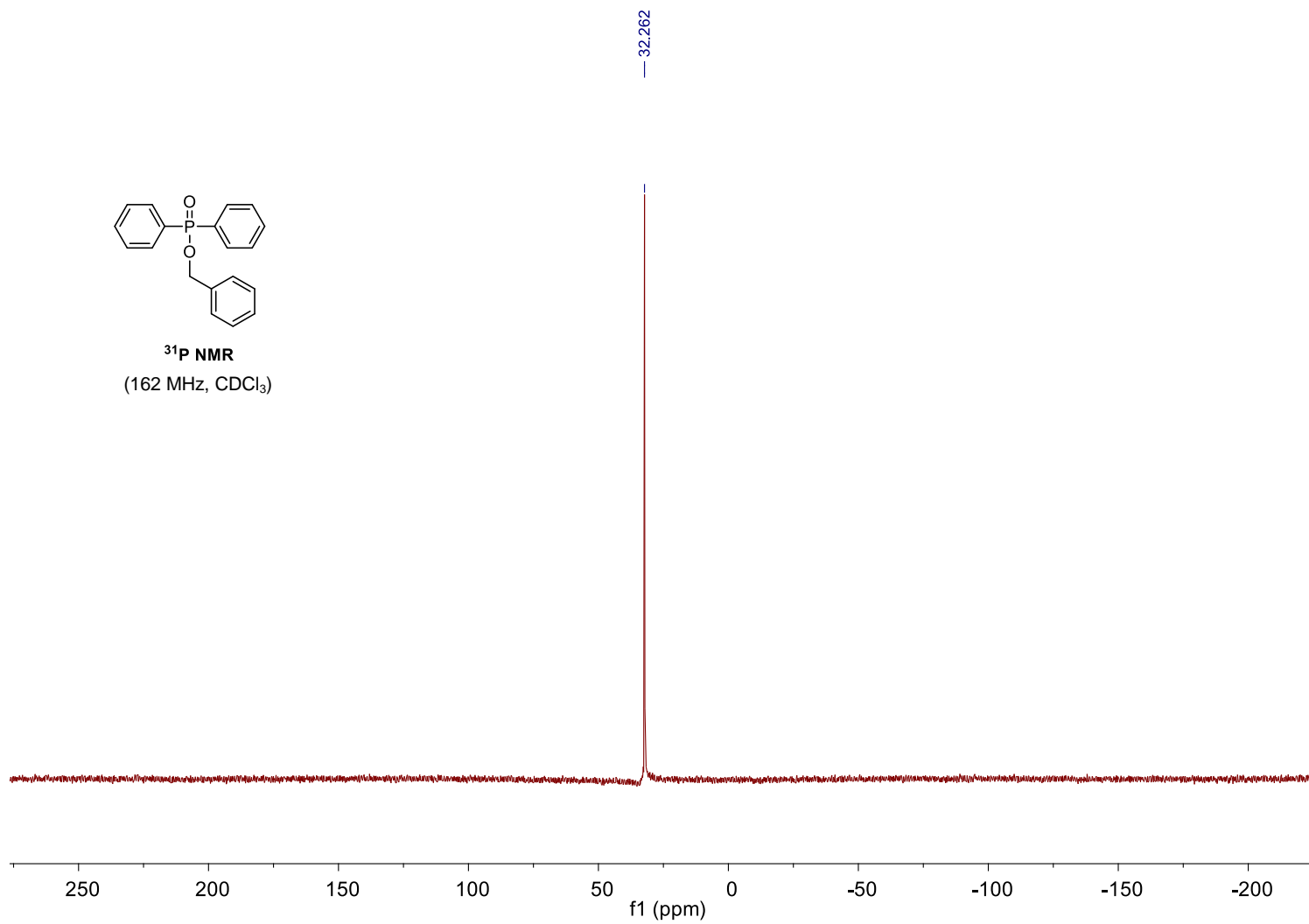


¹³C NMR
(100 MHz, CDCl₃)

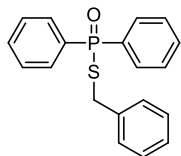




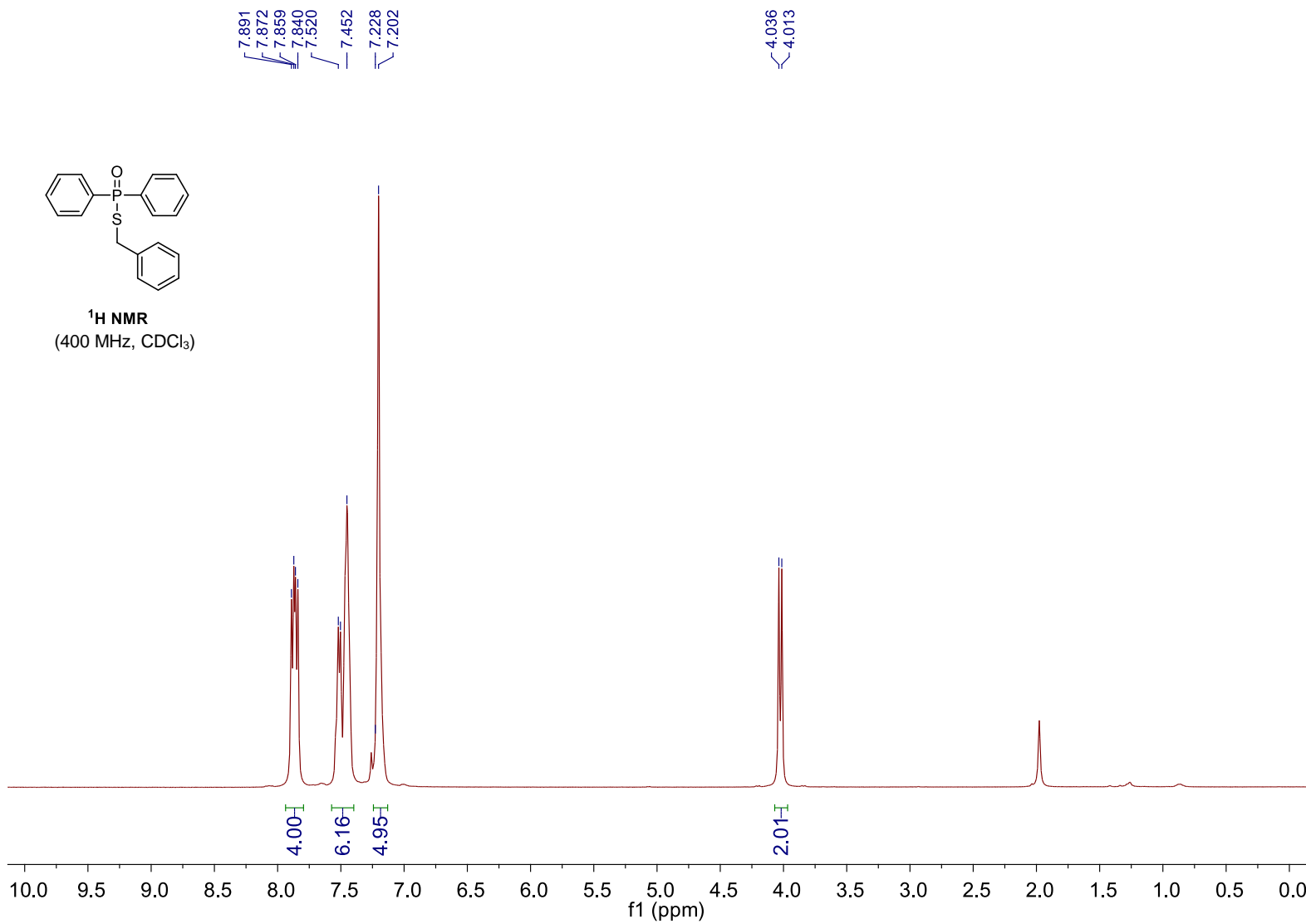
³¹P NMR
(162 MHz, CDCl₃)

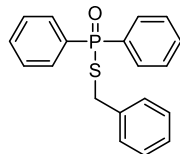


S135

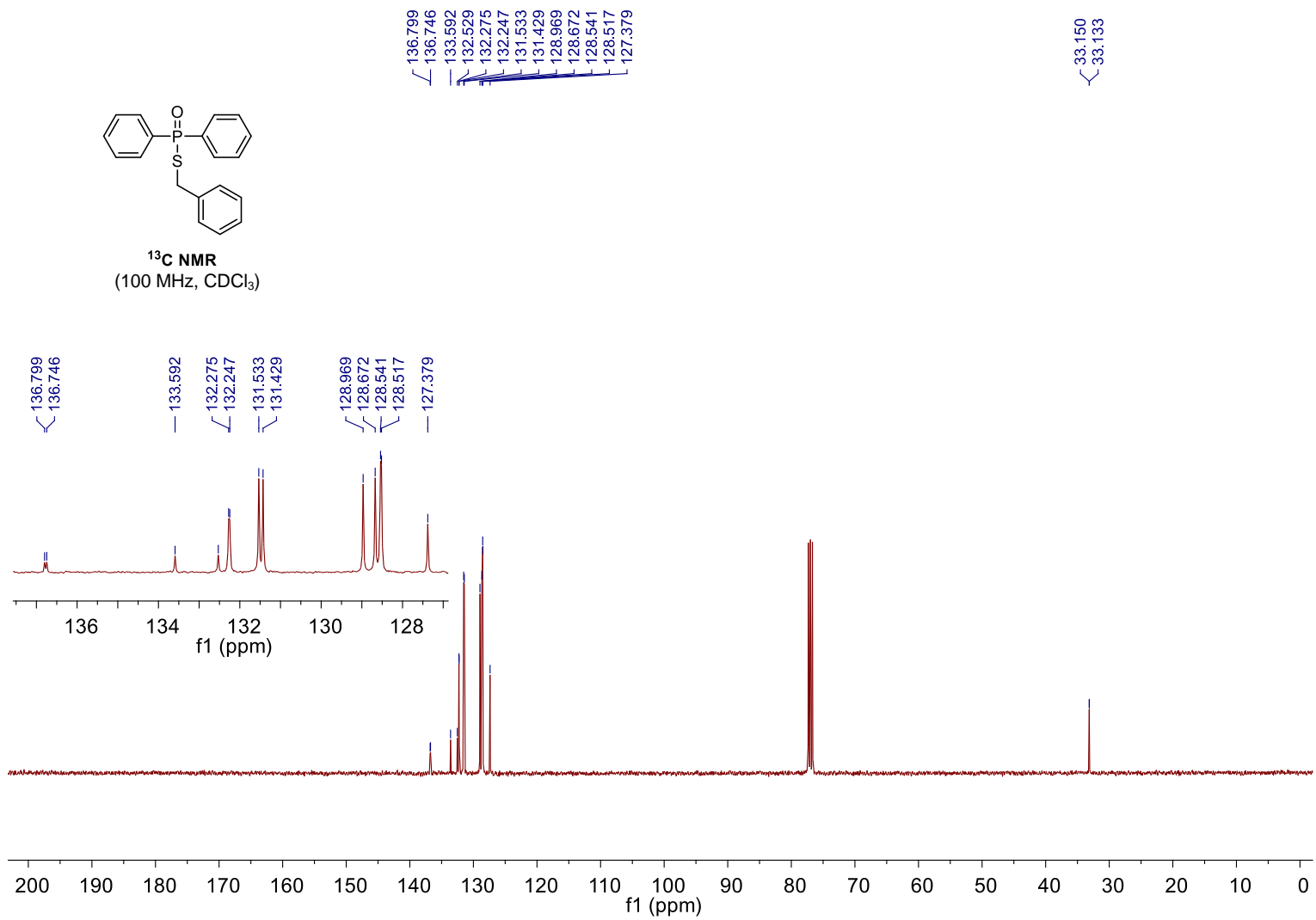


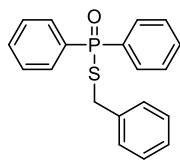
¹H NMR
(400 MHz, CDCl₃)



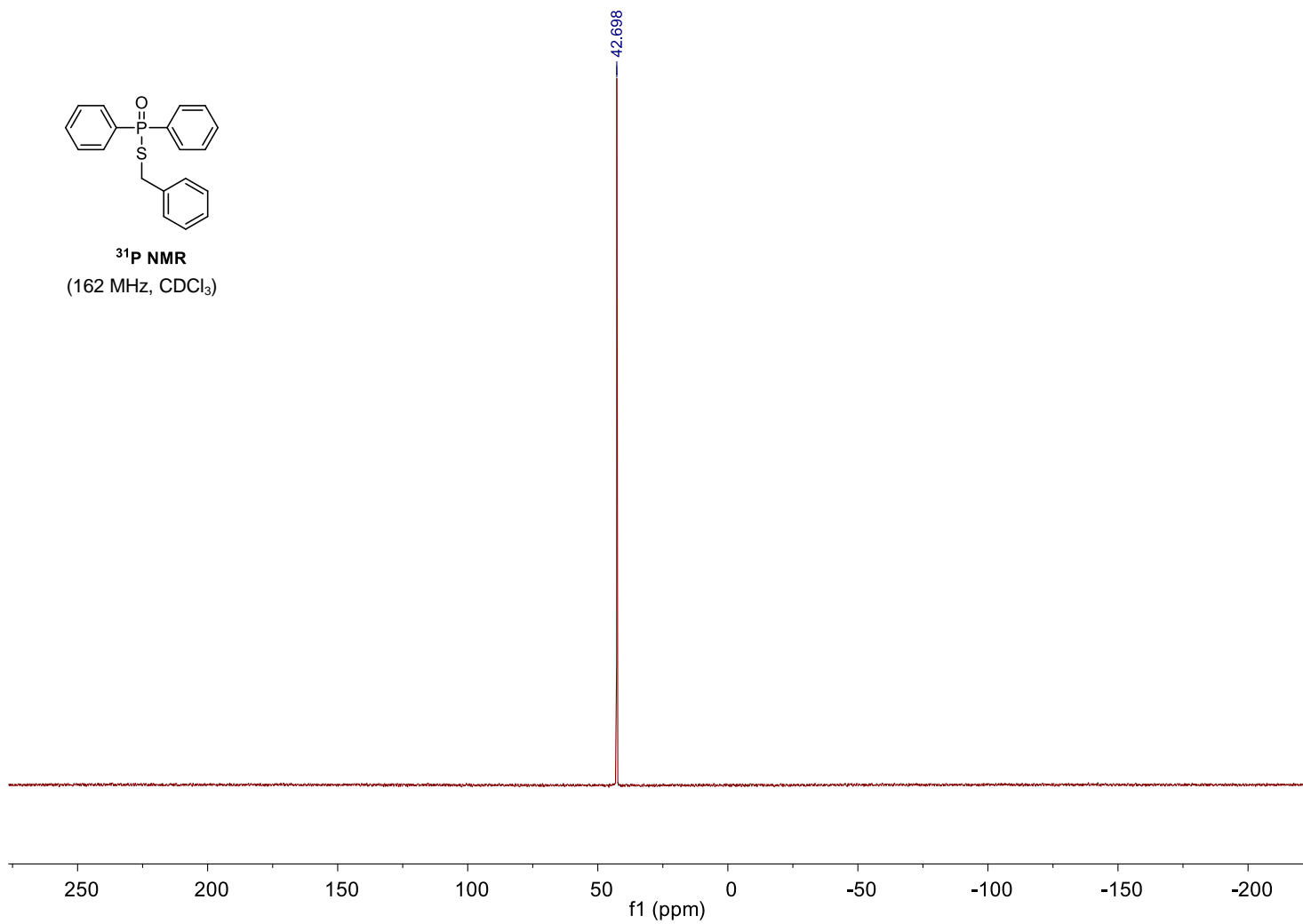


¹³C NMR
(100 MHz, CDCl₃)

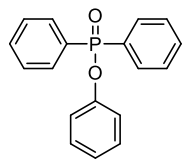




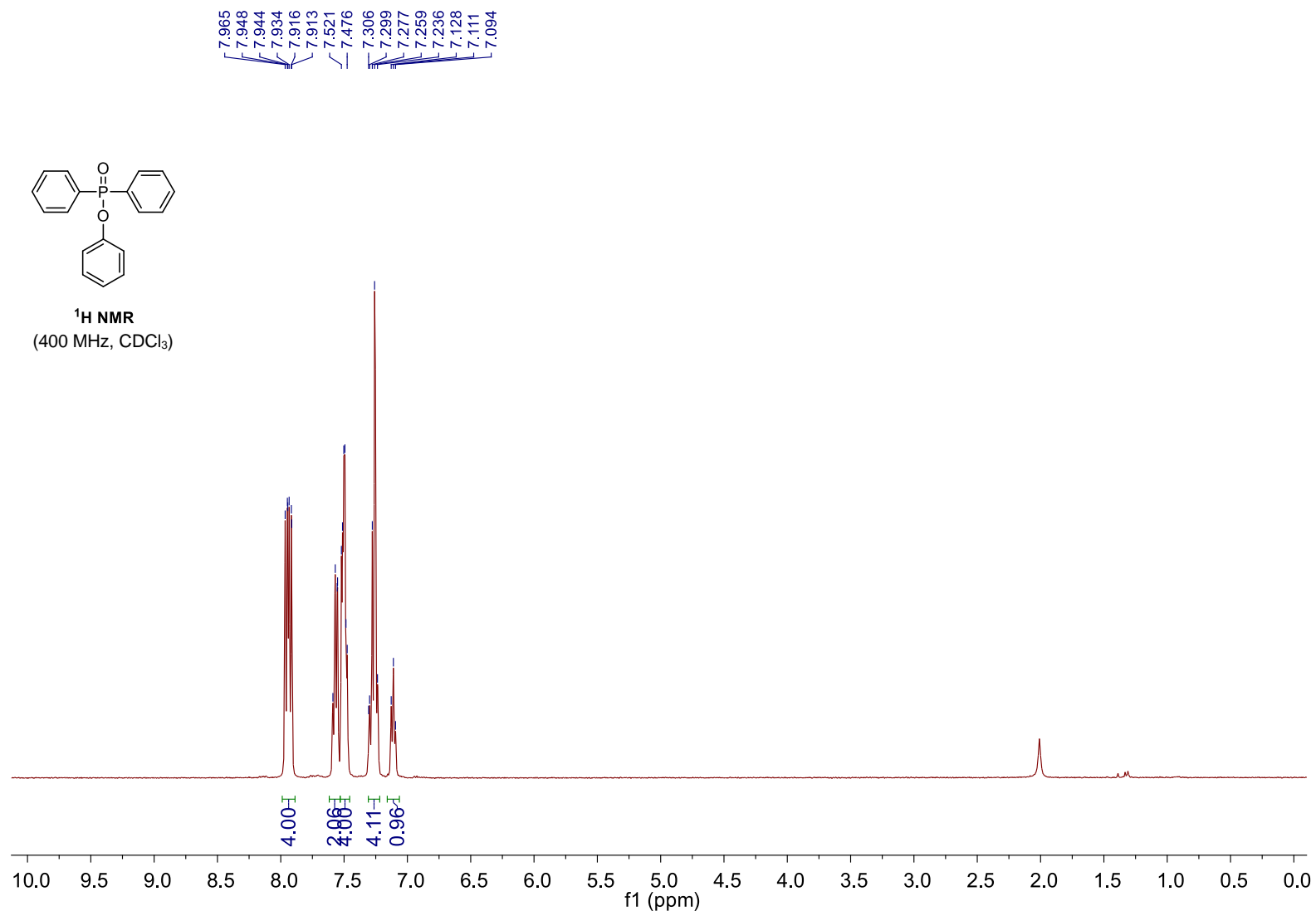
³¹P NMR
(162 MHz, CDCl₃)

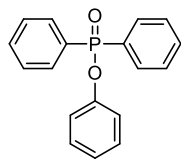


S138

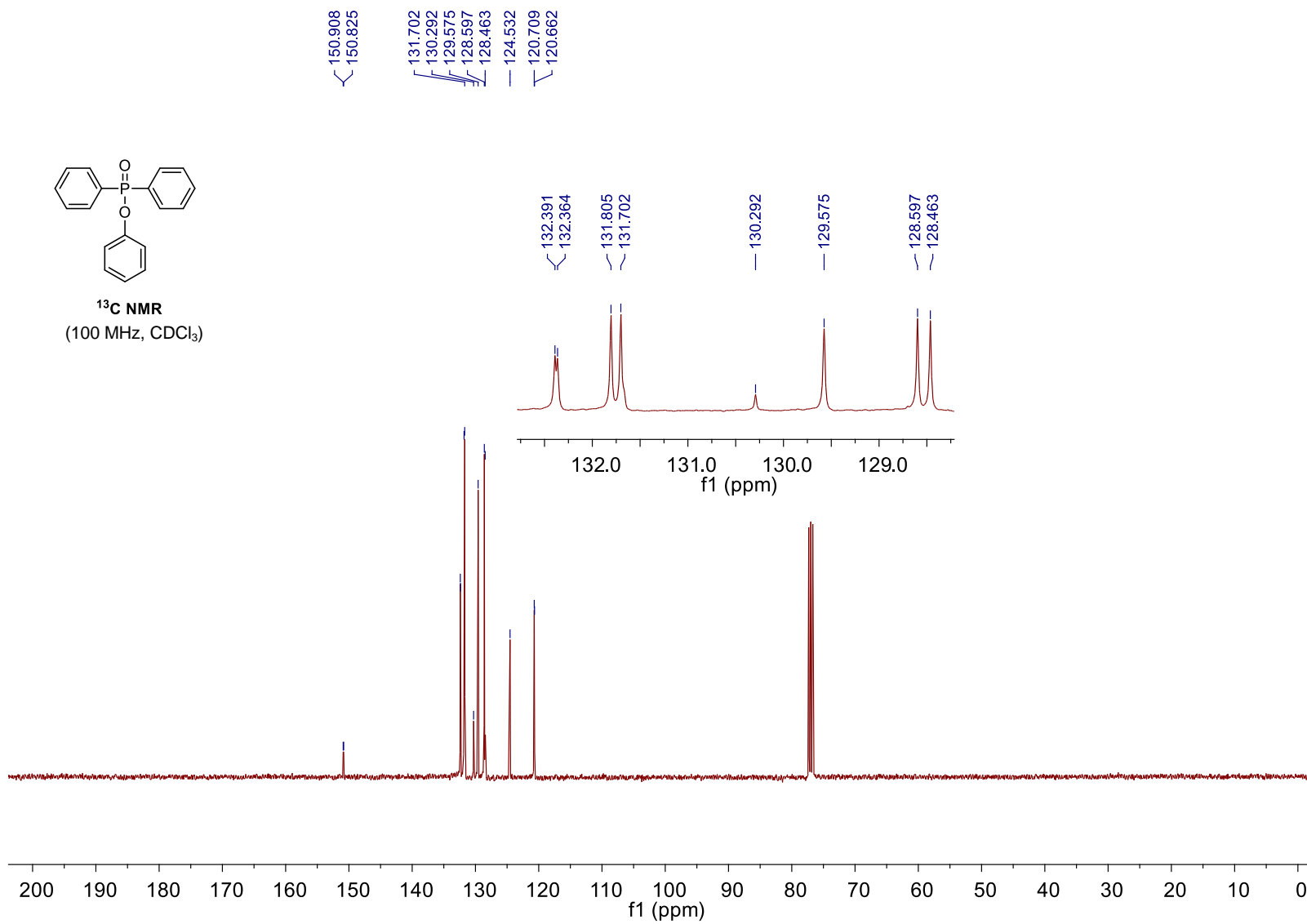


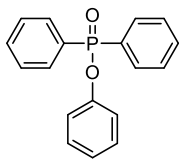
¹H NMR
(400 MHz, CDCl₃)



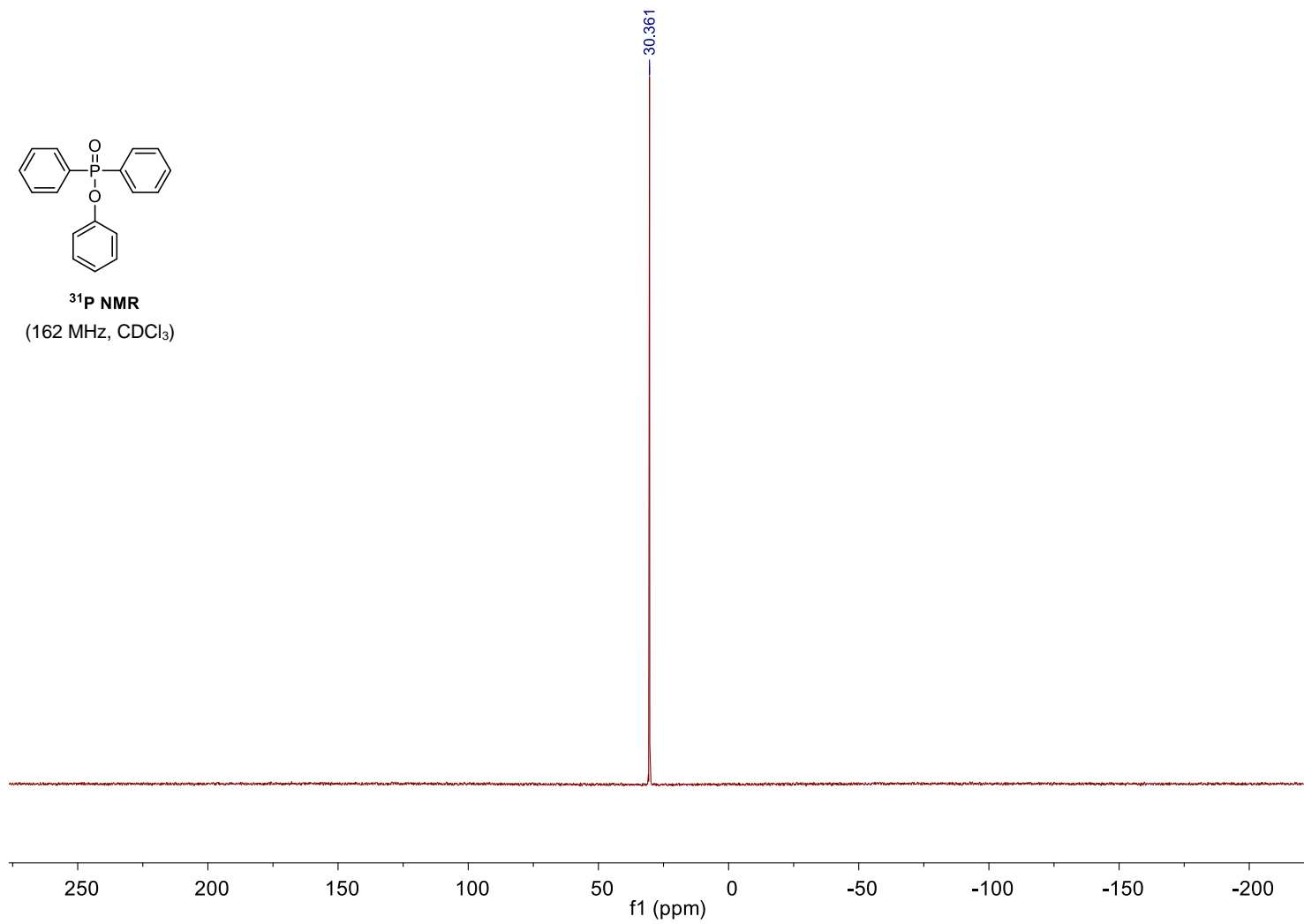


¹³C NMR
(100 MHz, CDCl₃)

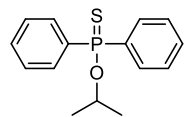




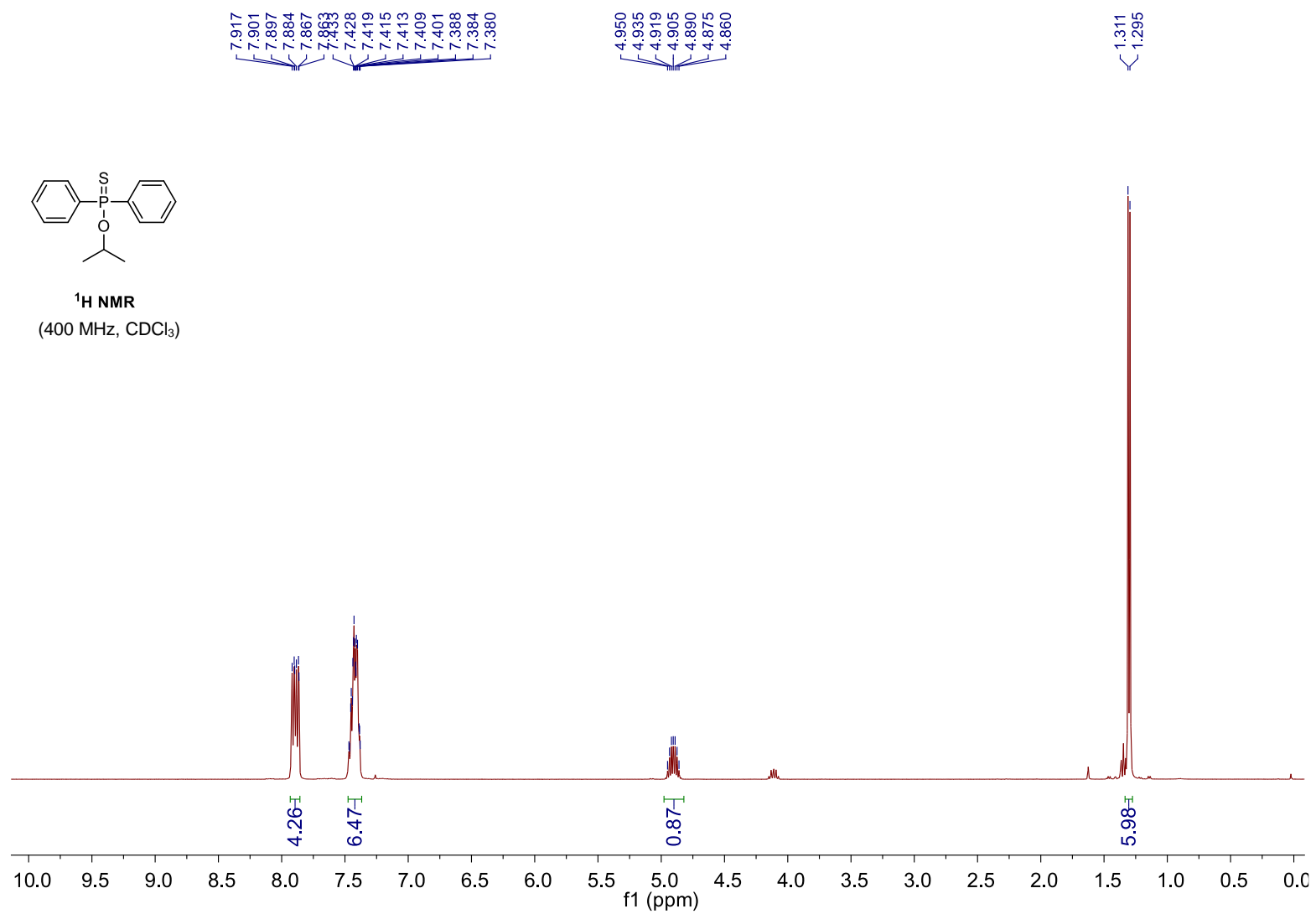
³¹P NMR
(162 MHz, CDCl₃)

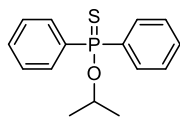


S141

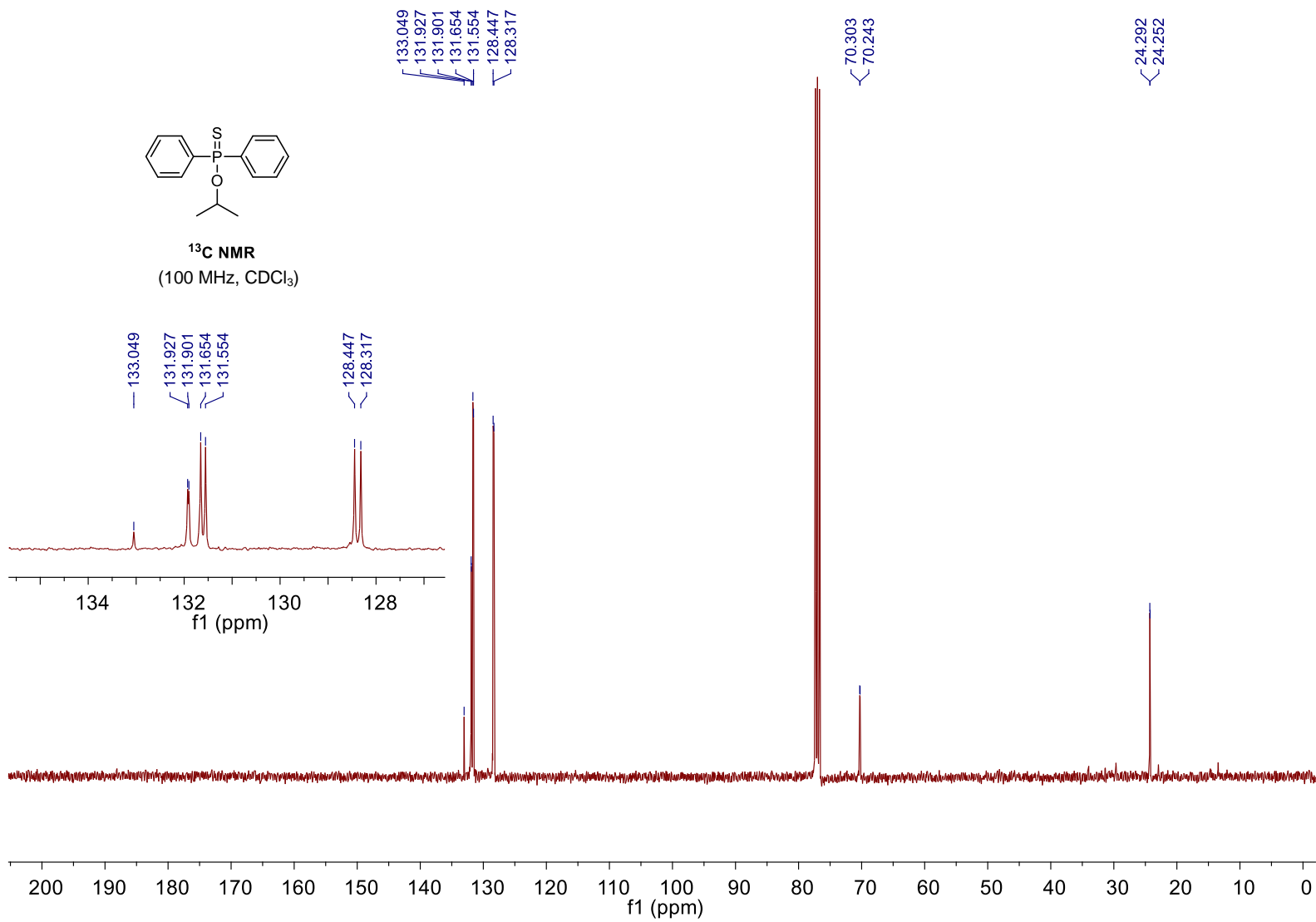


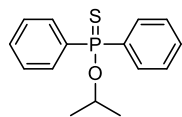
¹H NMR
(400 MHz, CDCl₃)



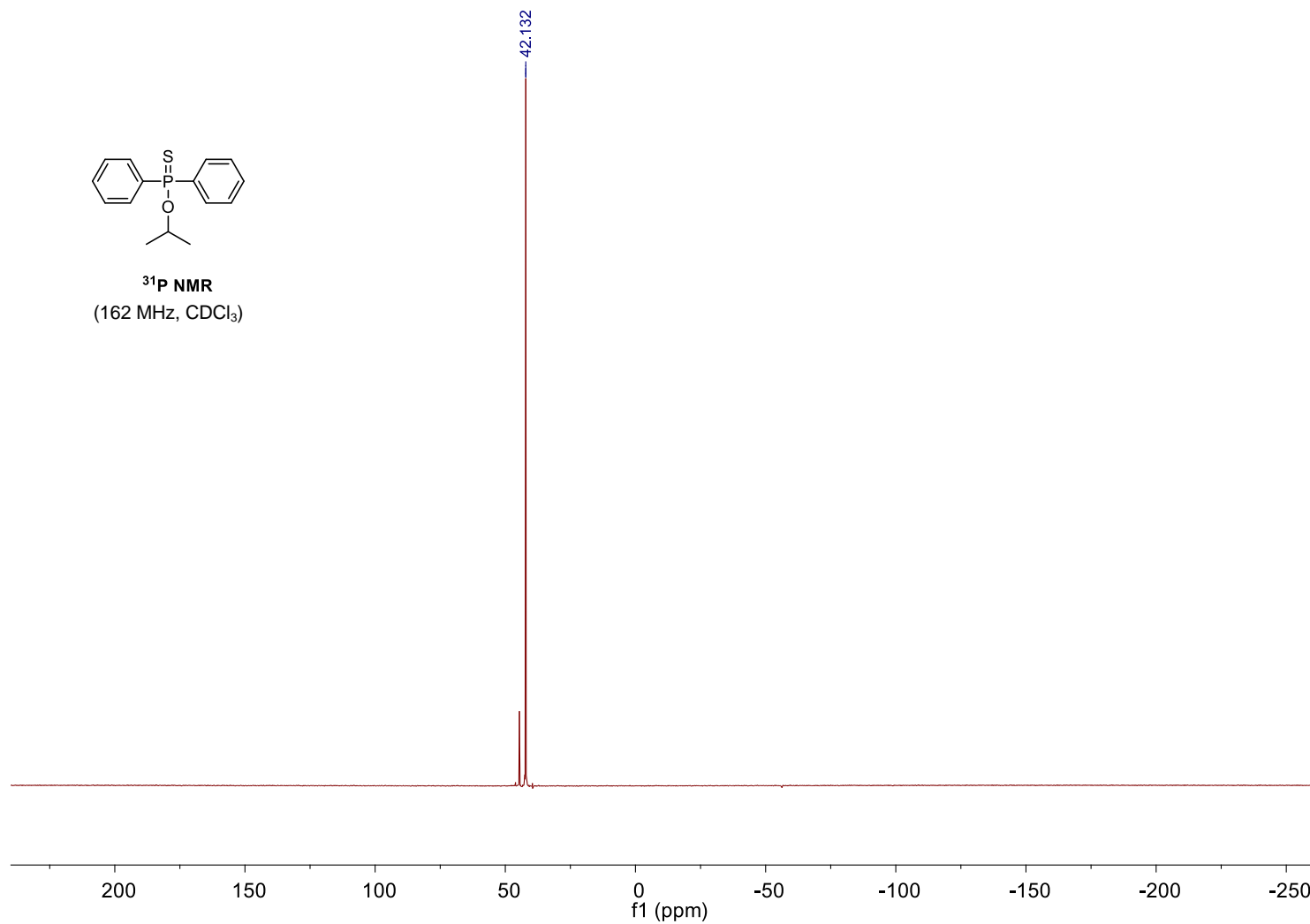


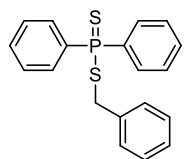
¹³C NMR
(100 MHz, CDCl₃)



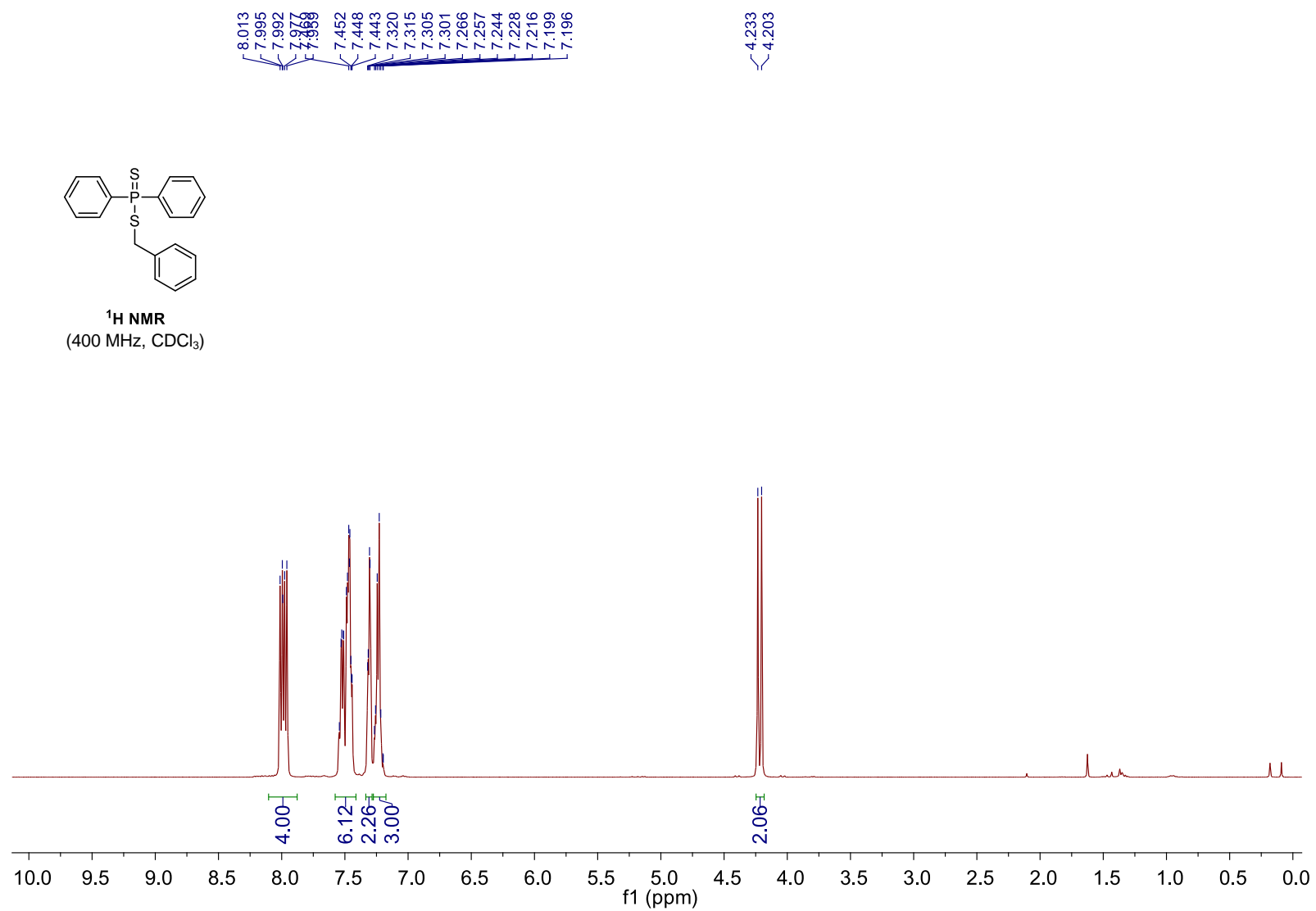


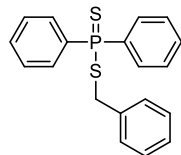
³¹P NMR
(162 MHz, CDCl₃)





¹H NMR
(400 MHz, CDCl₃)

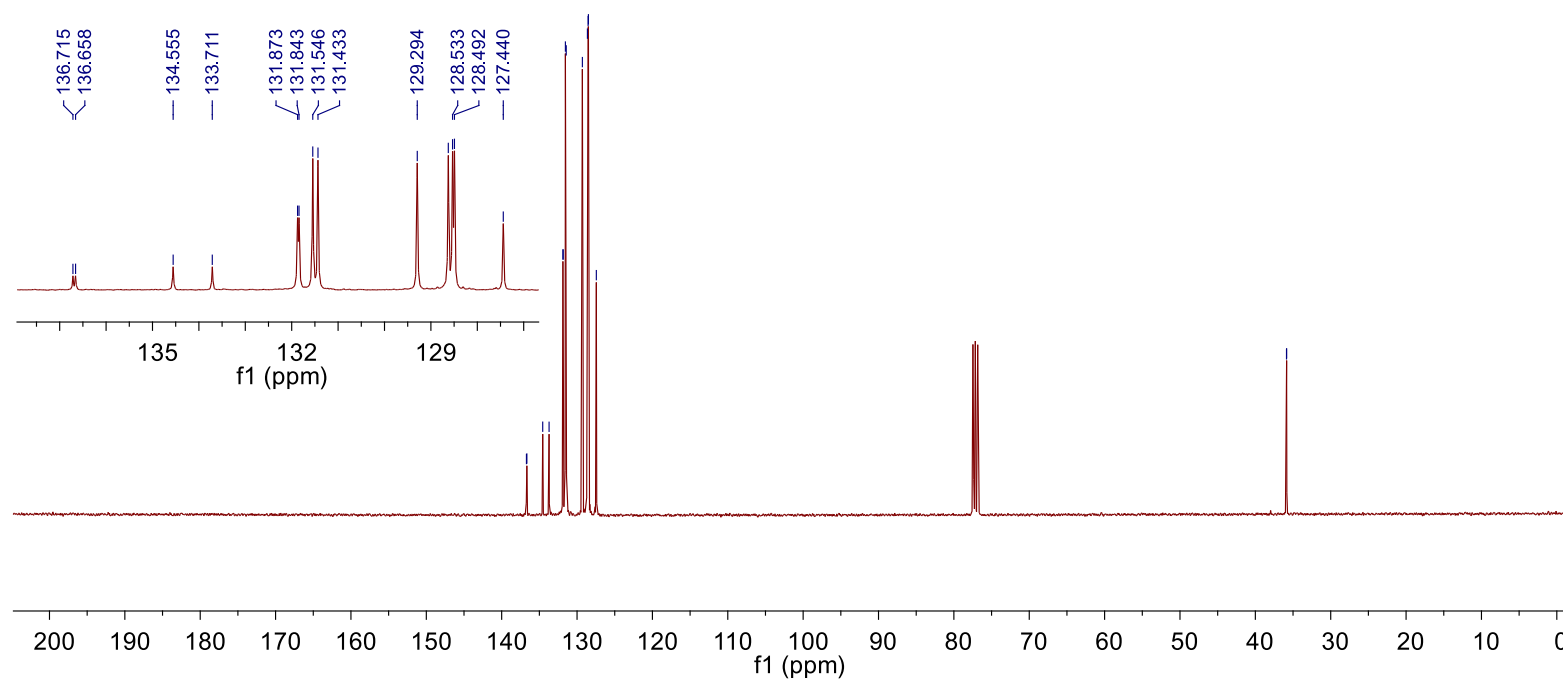


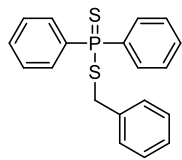


¹³C NMR
(100 MHz, CDCl₃)

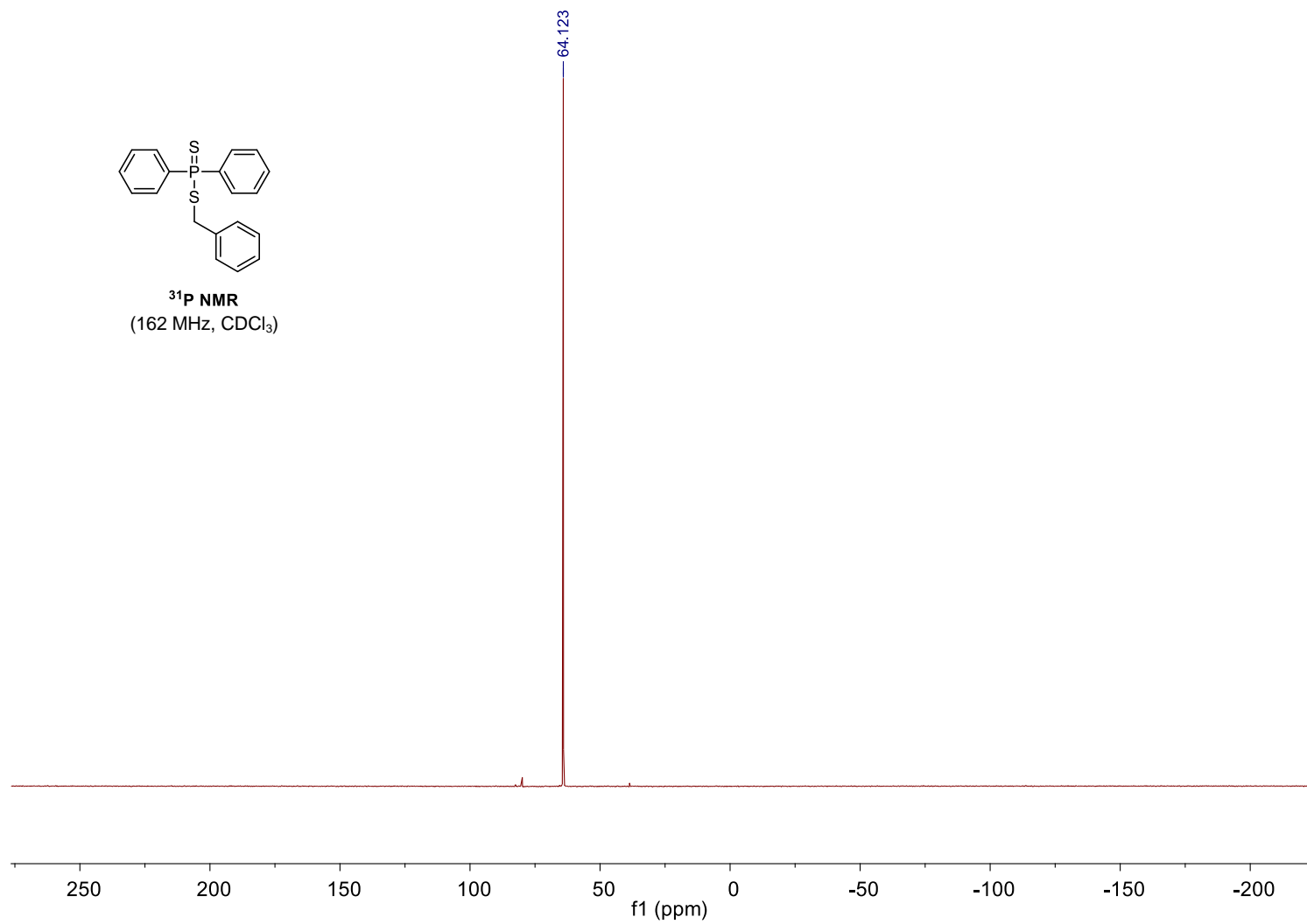
136.715
136.658
134.555
133.711
131.873
131.843
131.546
131.433
129.294
128.625
128.533
128.492
127.440

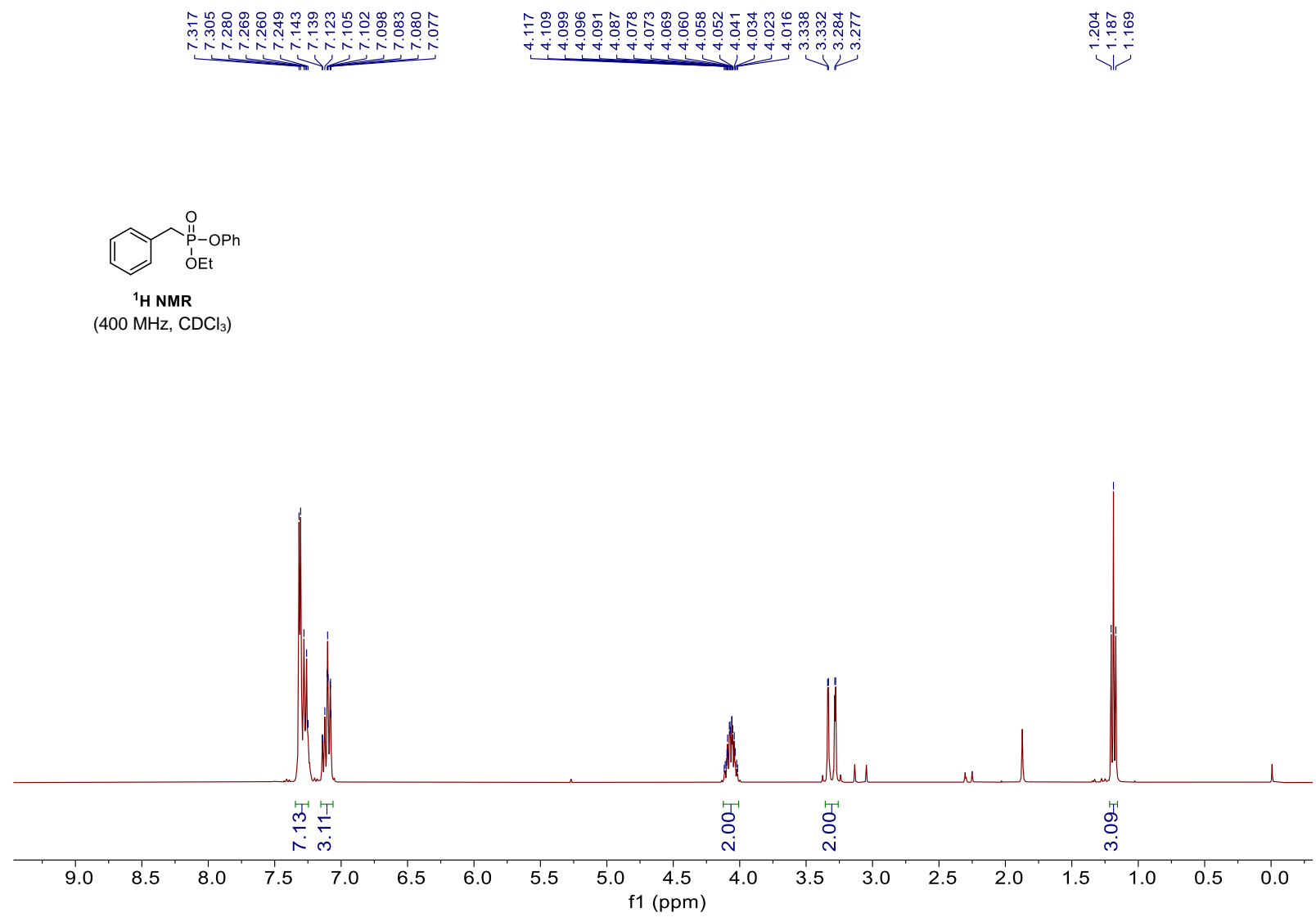
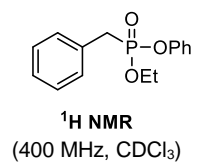
35.866
35.852

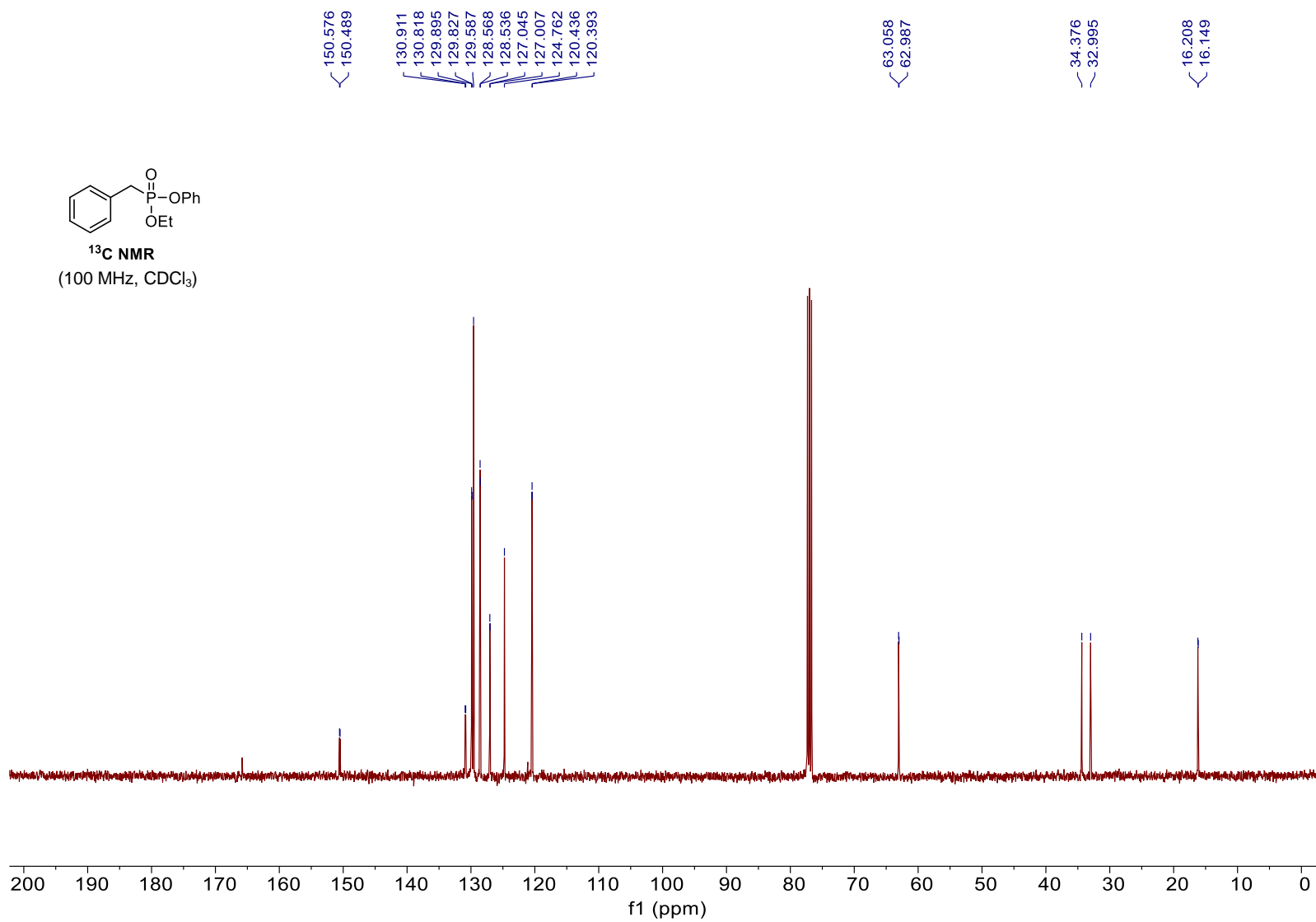
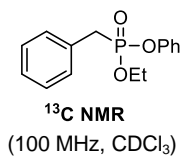


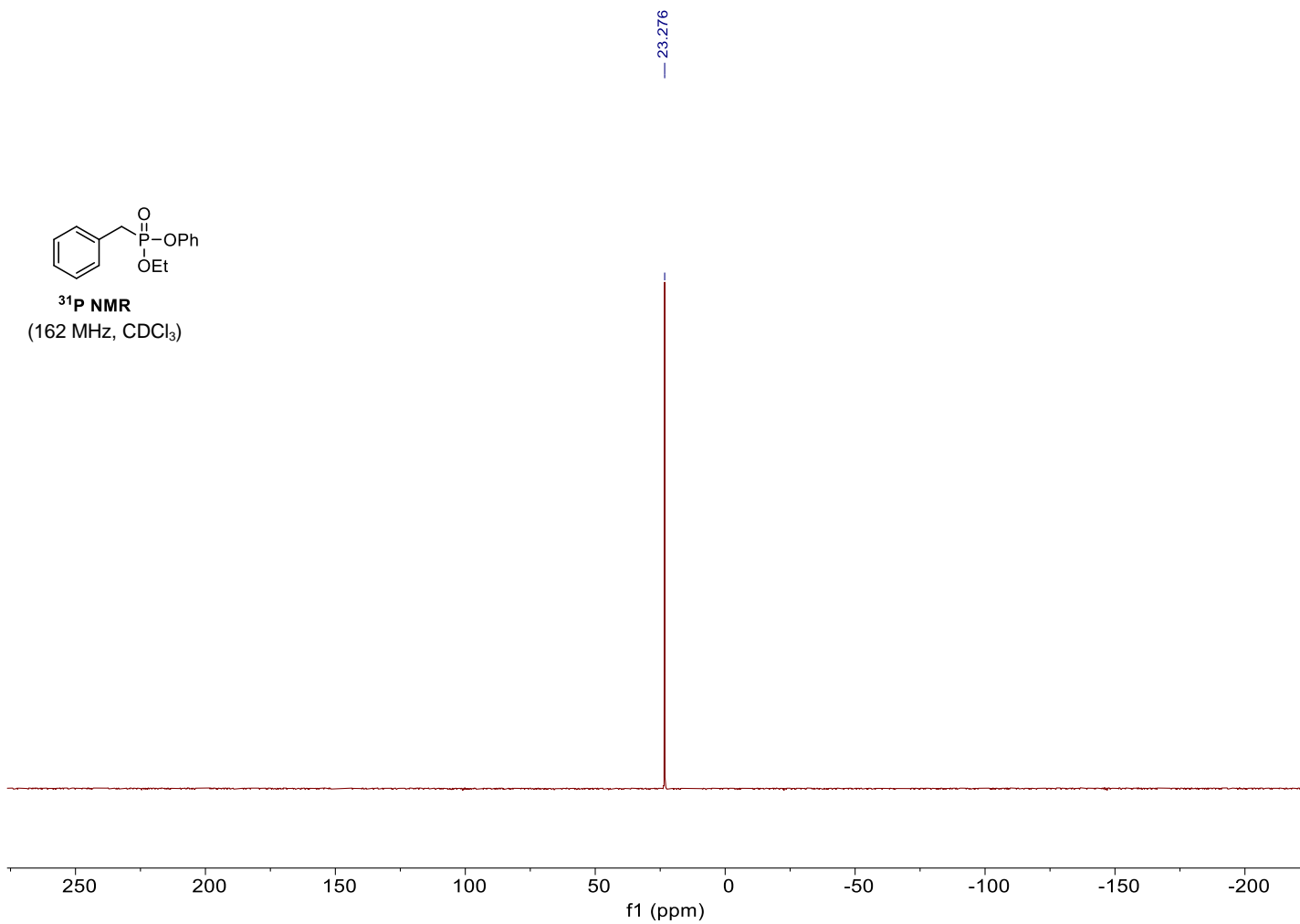
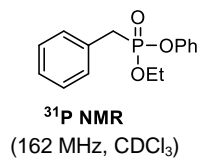


³¹P NMR
(162 MHz, CDCl₃)







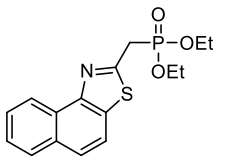


S150

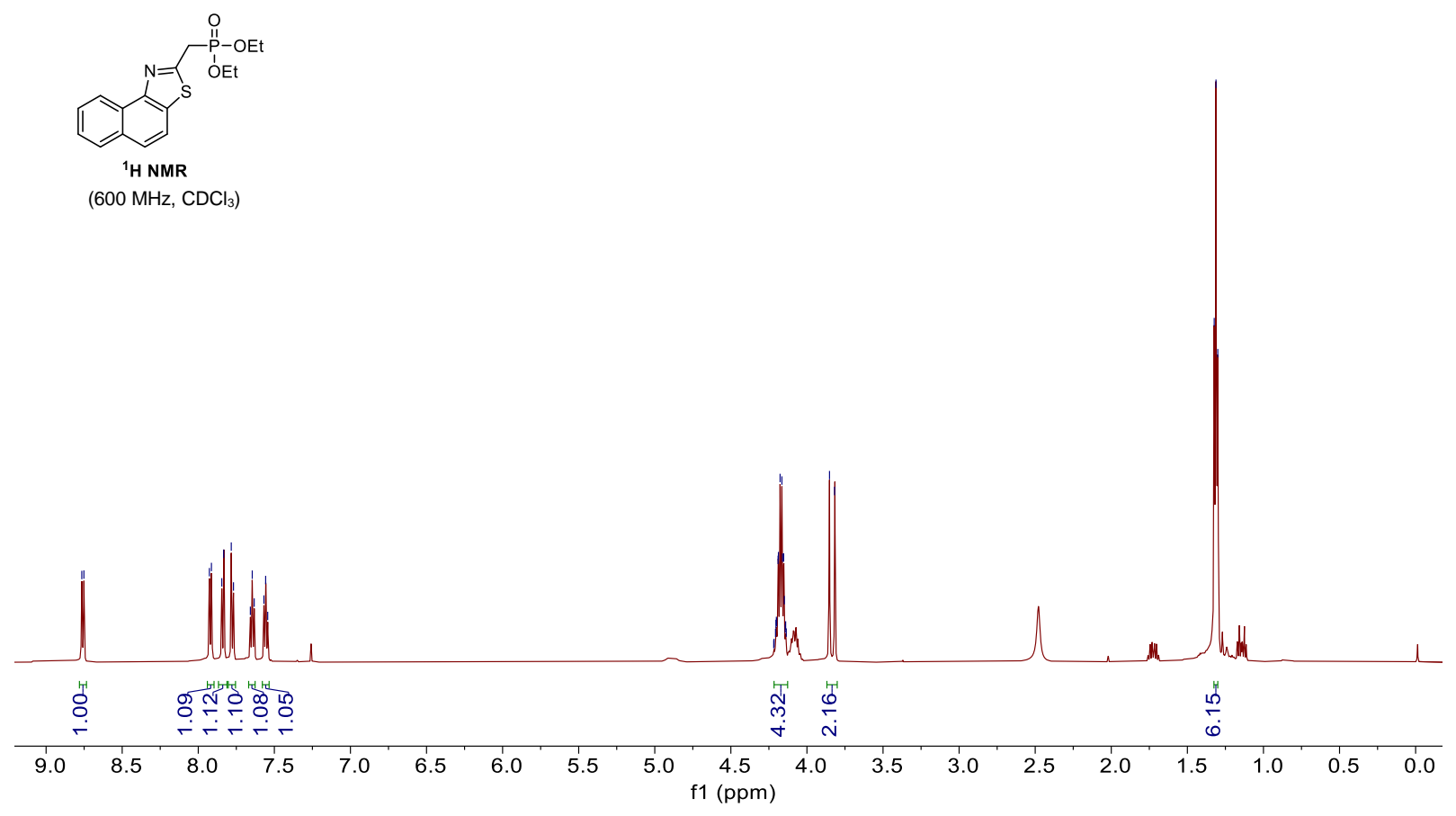
8.765
8.752
7.927
7.914
7.845
7.833
7.783
7.769
7.658
7.645
7.632
7.569
7.557
7.544

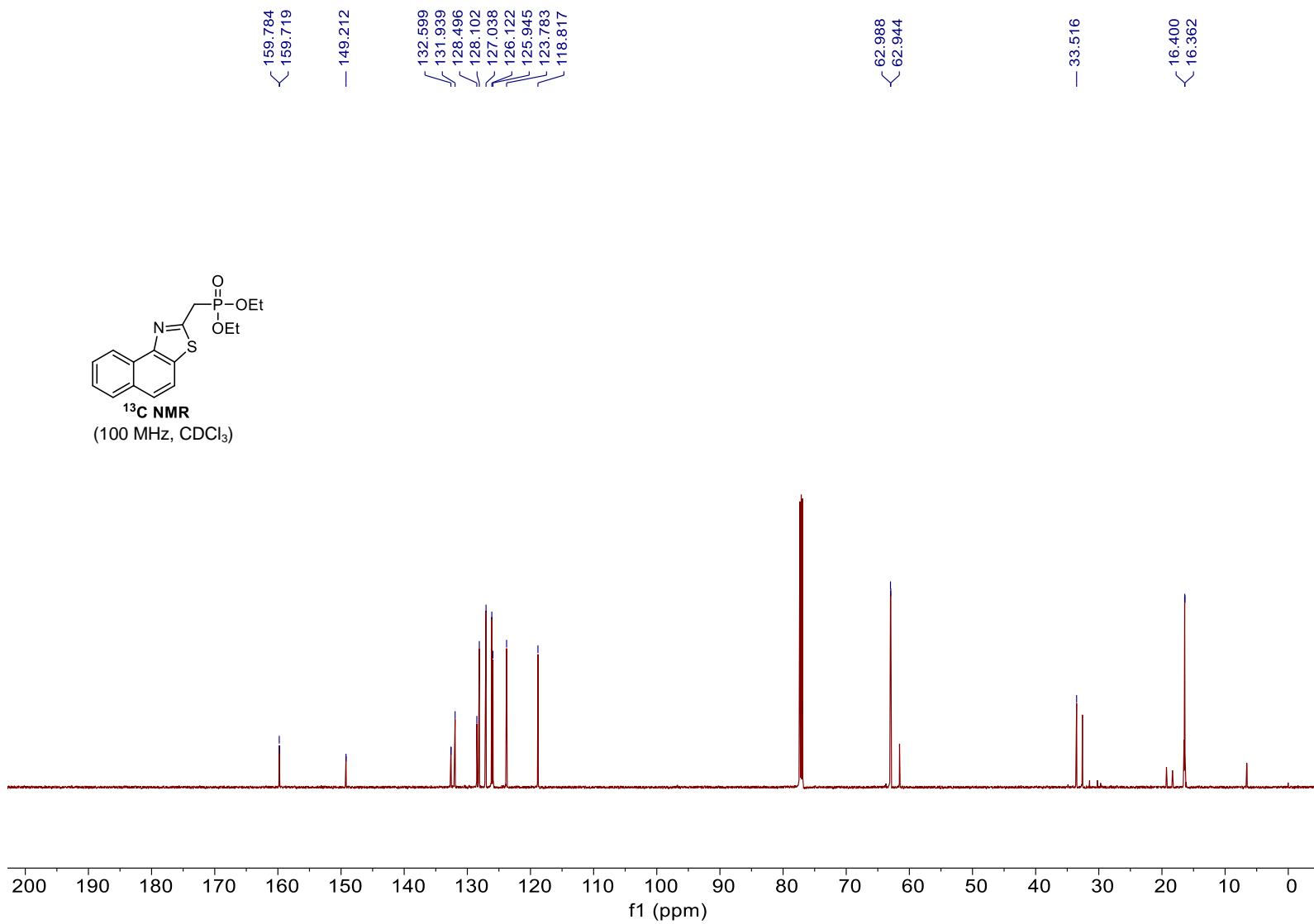
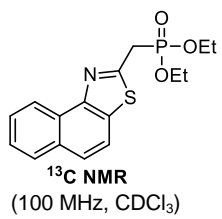
4.218
4.216
4.207
4.204
4.201
4.199
4.190
4.187
4.176
4.165
4.161
4.153
4.148
4.143
4.140
4.136
3.852
3.818

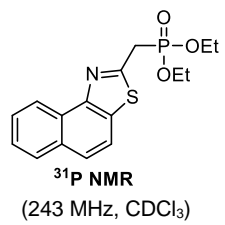
1.324
1.312
1.300



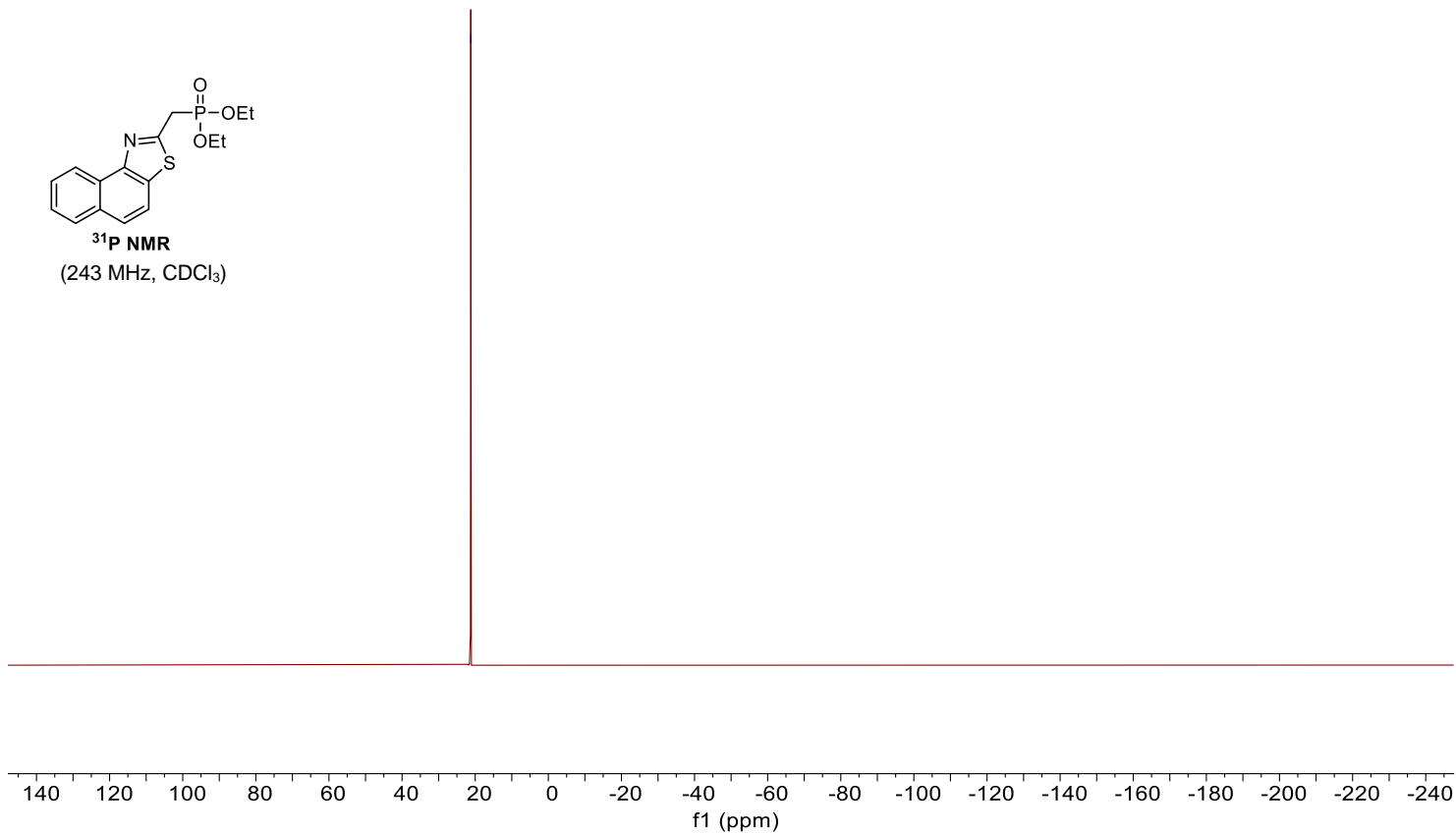
¹H NMR
(600 MHz, CDCl₃)

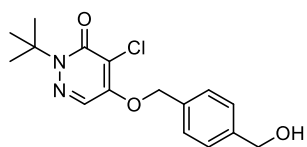




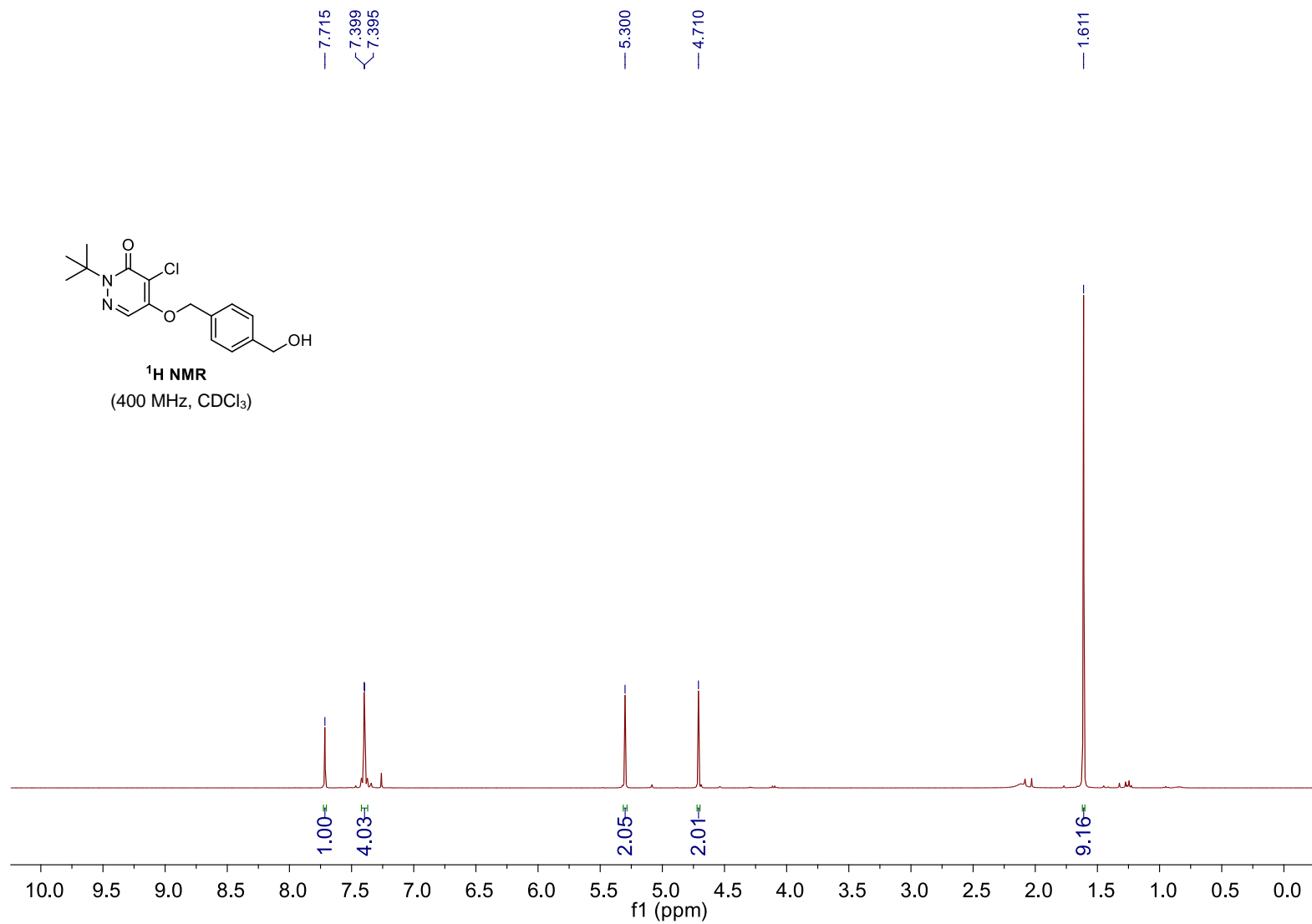


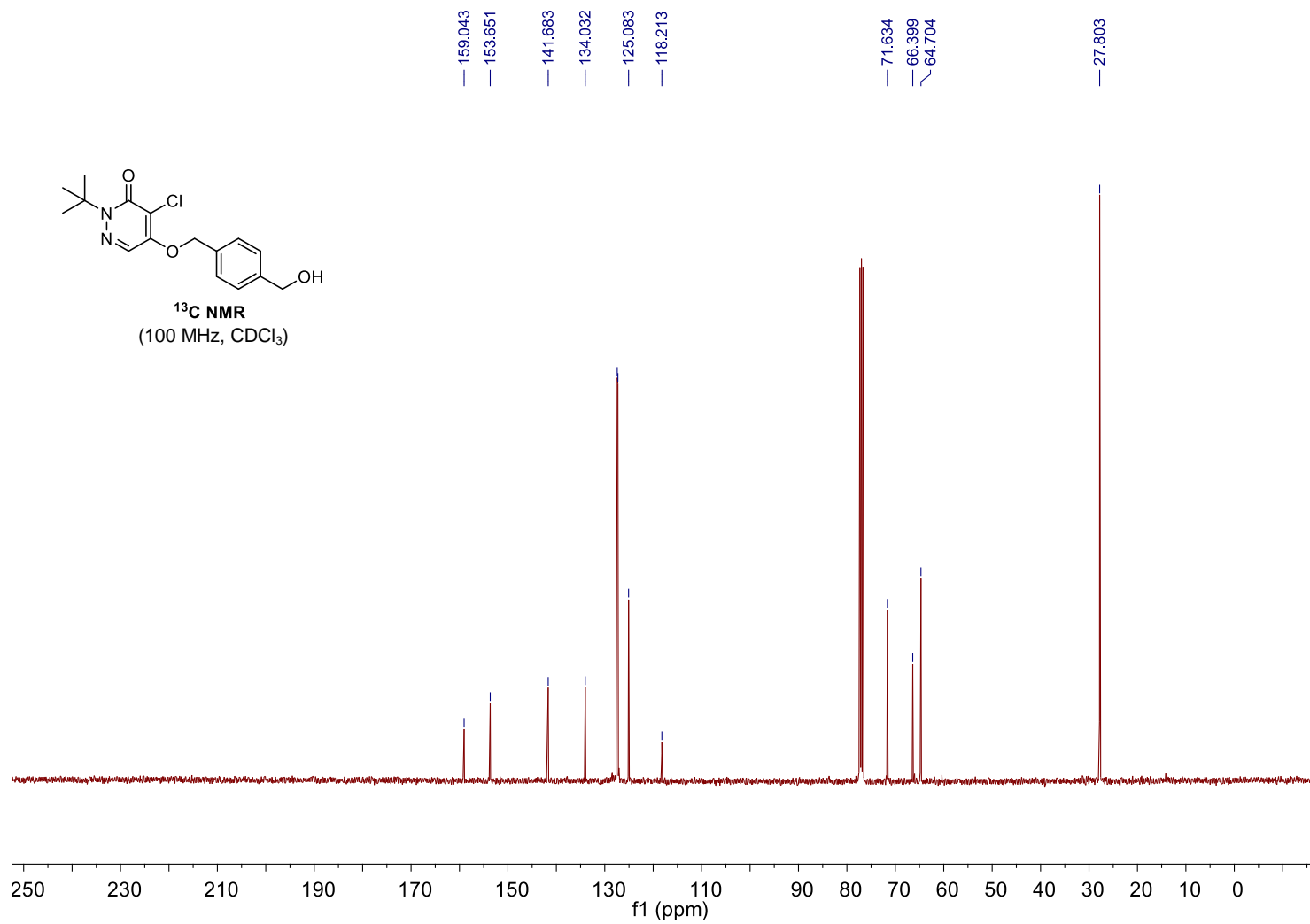
— 21.206

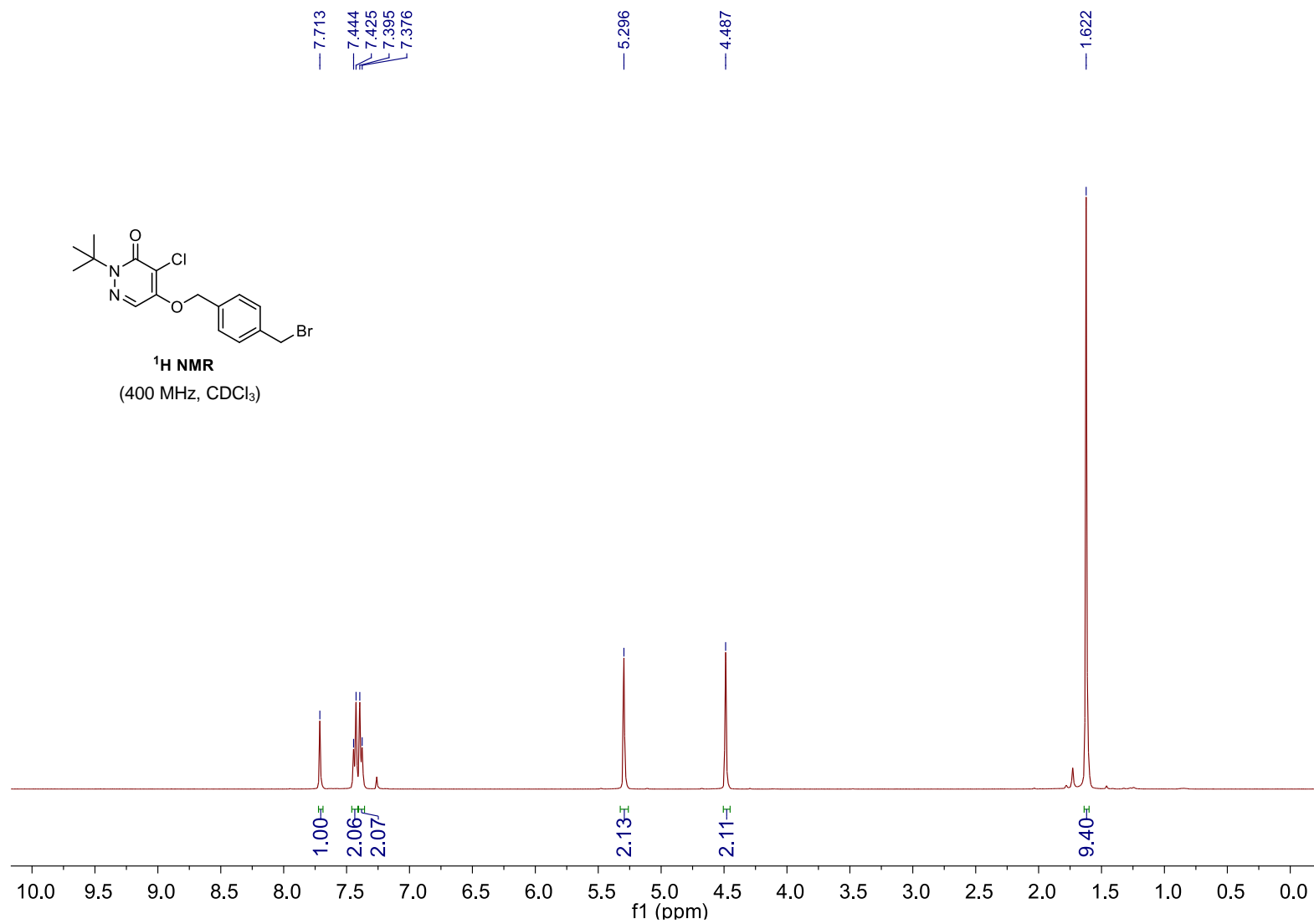


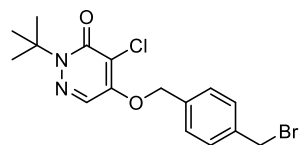


¹H NMR
(400 MHz, CDCl₃)

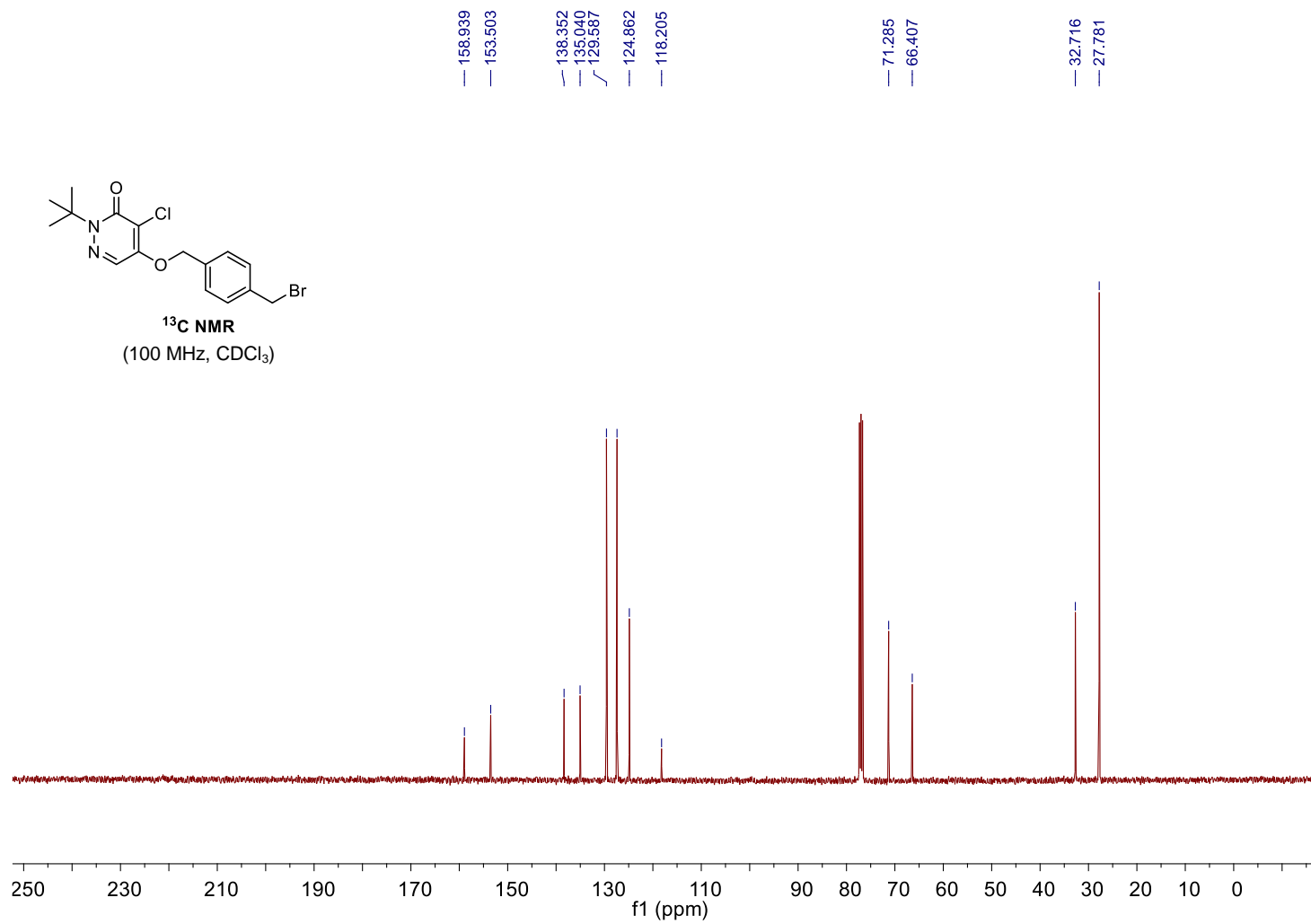


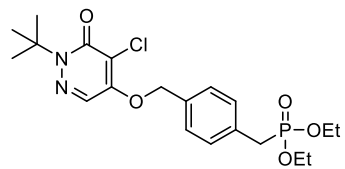




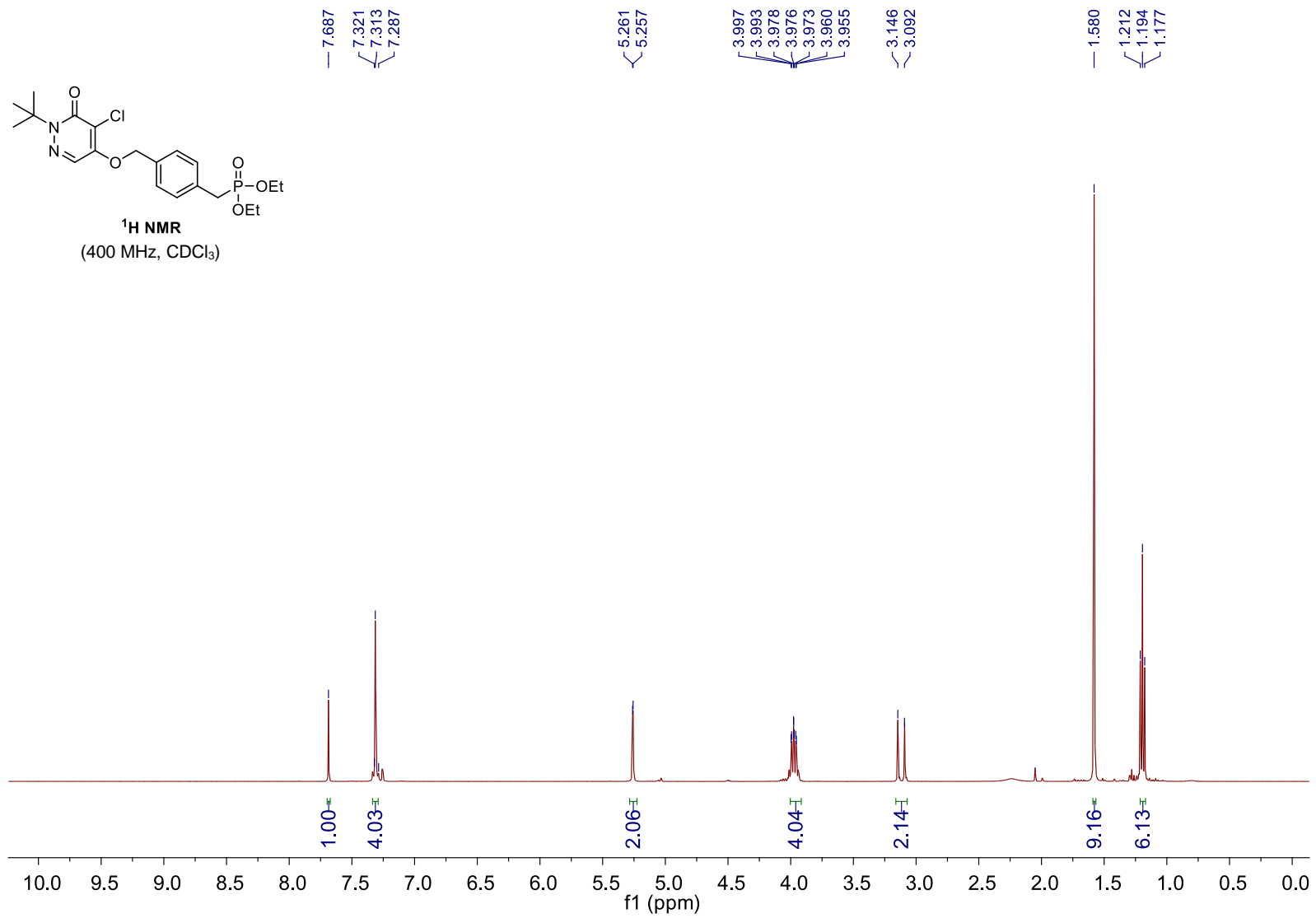


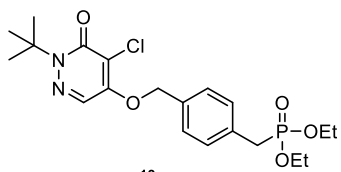
¹³C NMR
(100 MHz, CDCl₃)



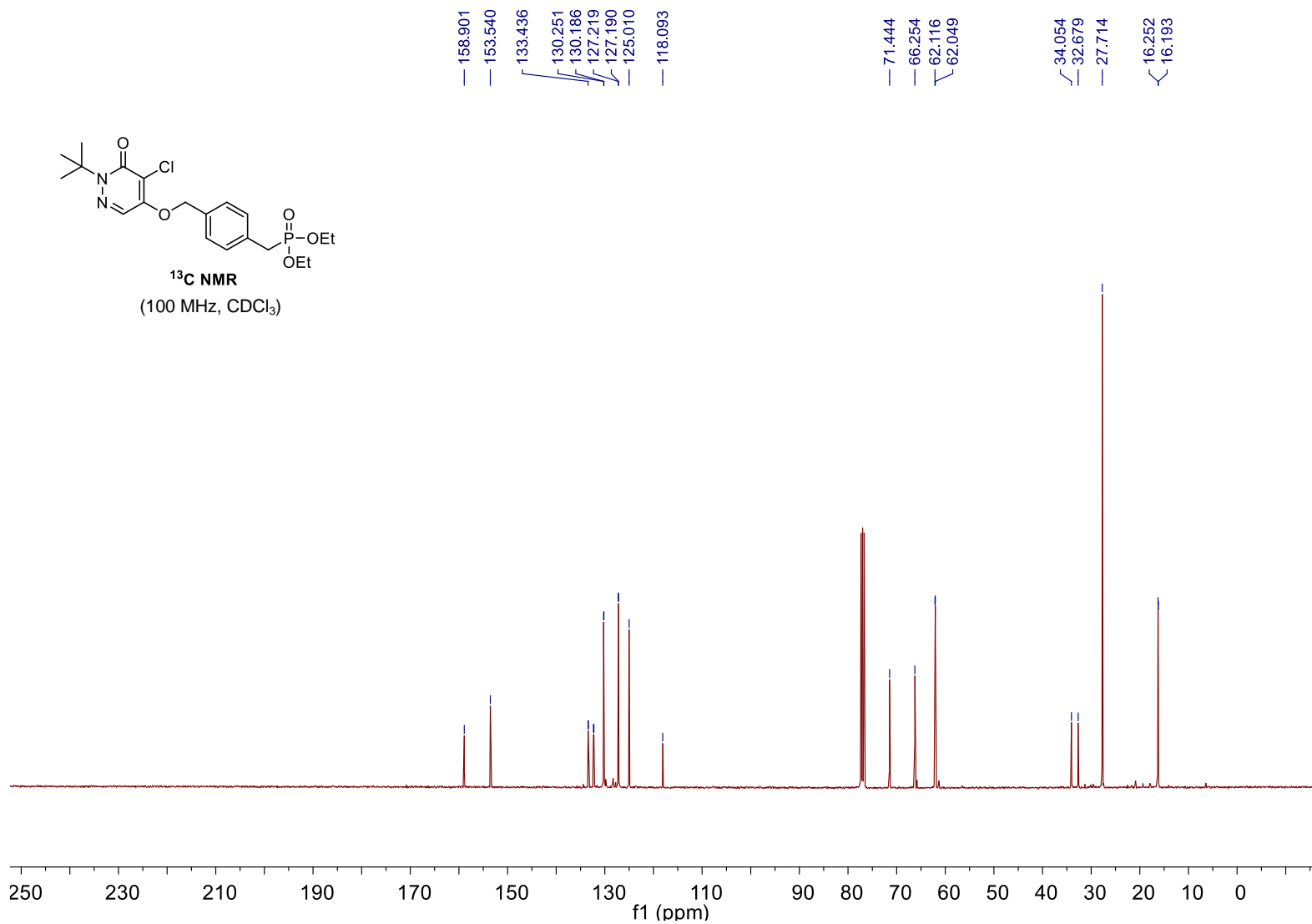


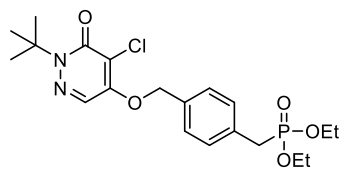
¹H NMR
(400 MHz, CDCl₃)



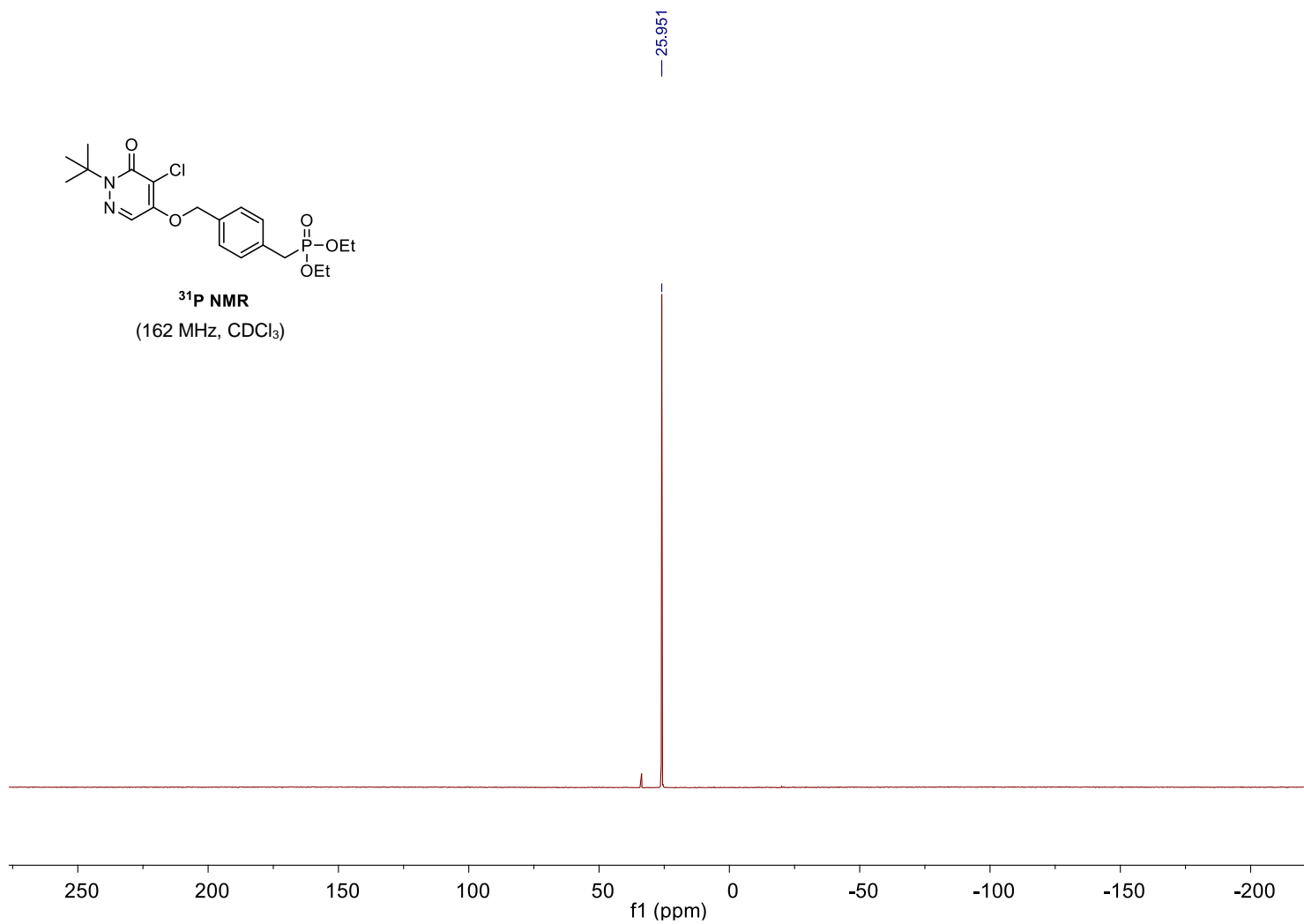


¹³C NMR
(100 MHz, CDCl₃)

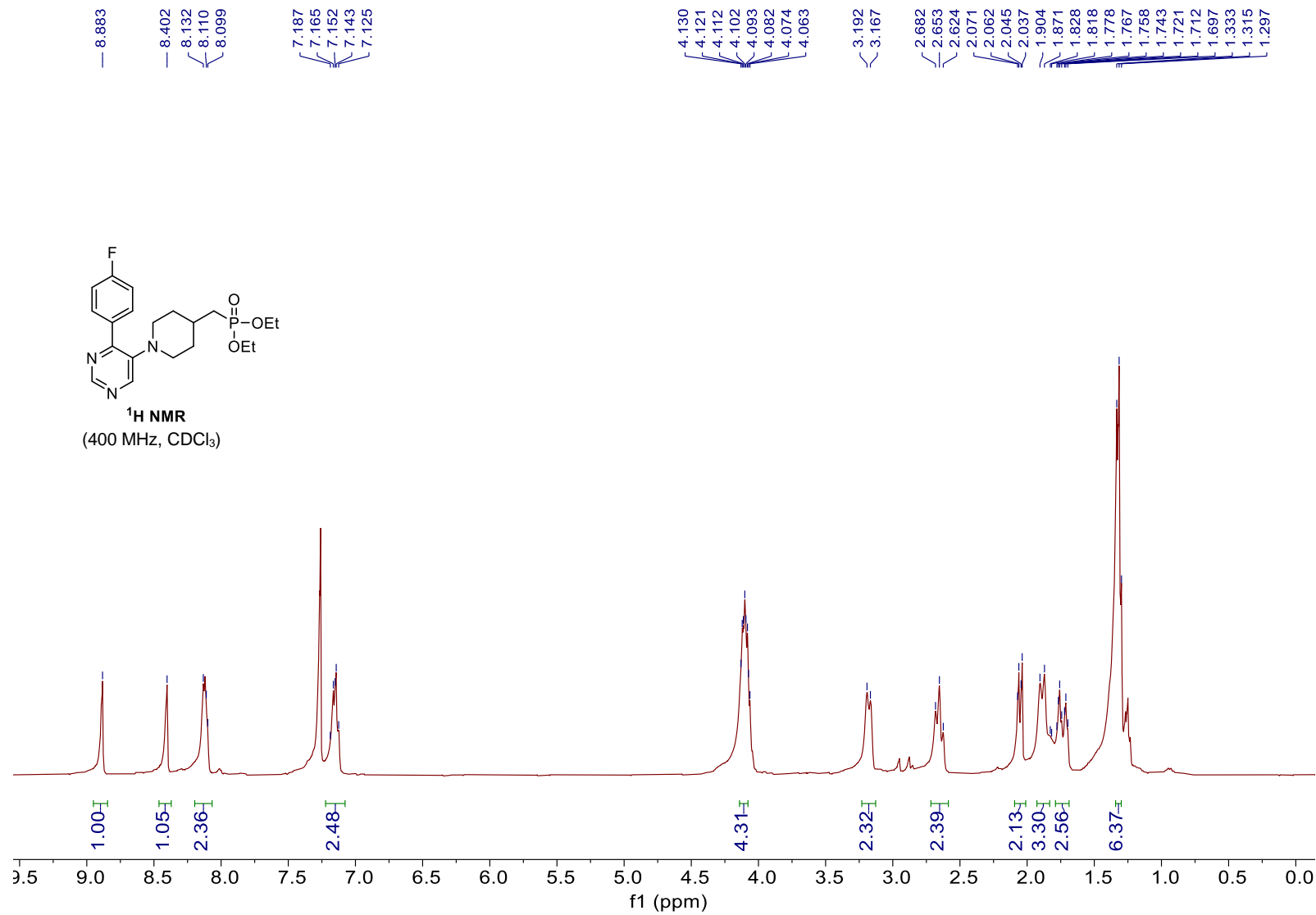
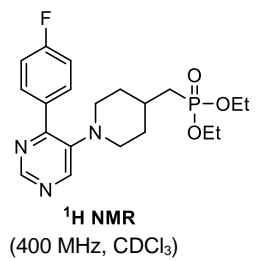


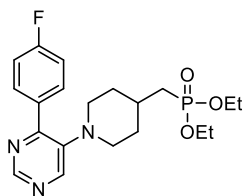


³¹P NMR
(162 MHz, CDCl₃)

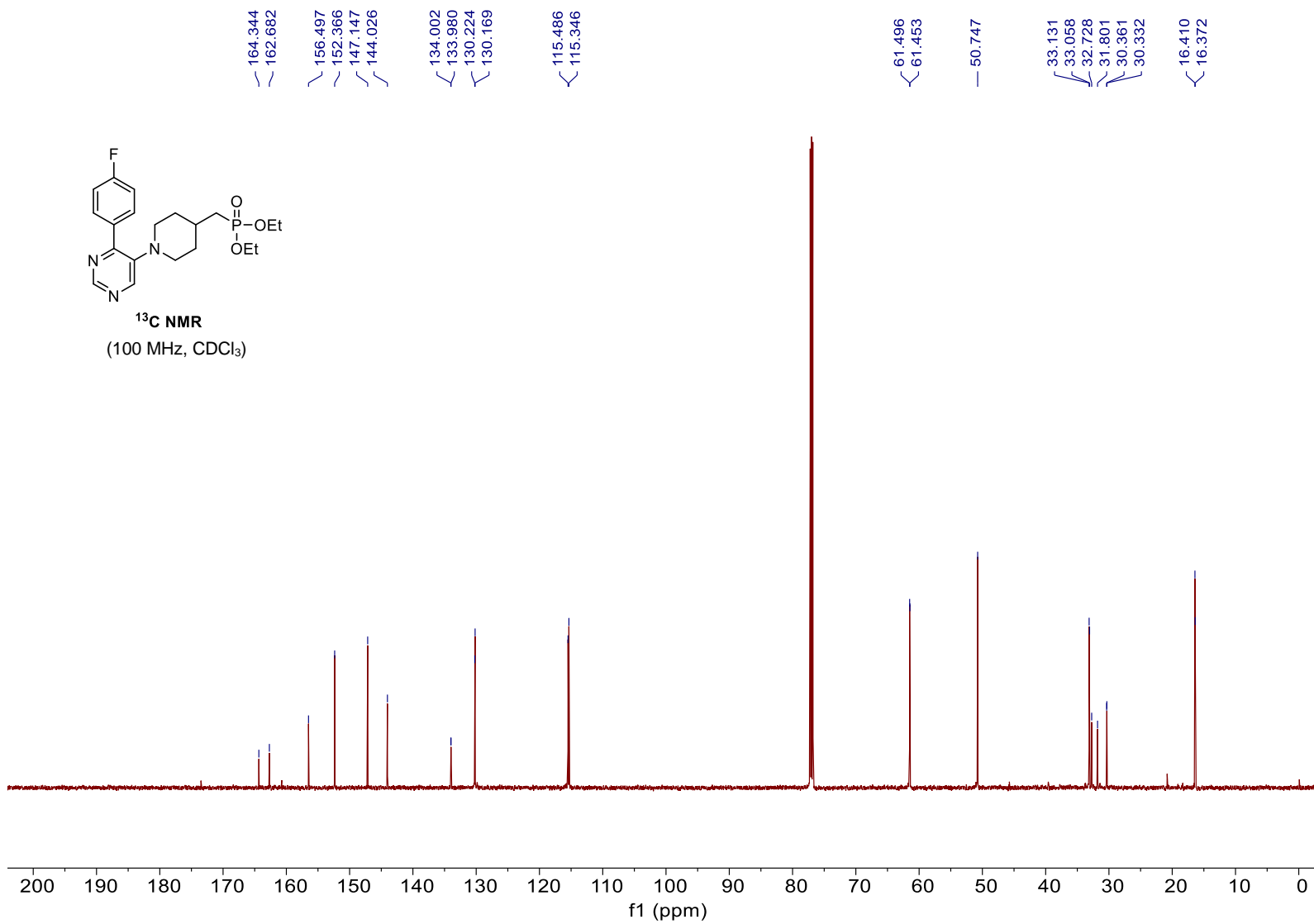


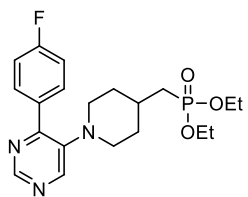
S160



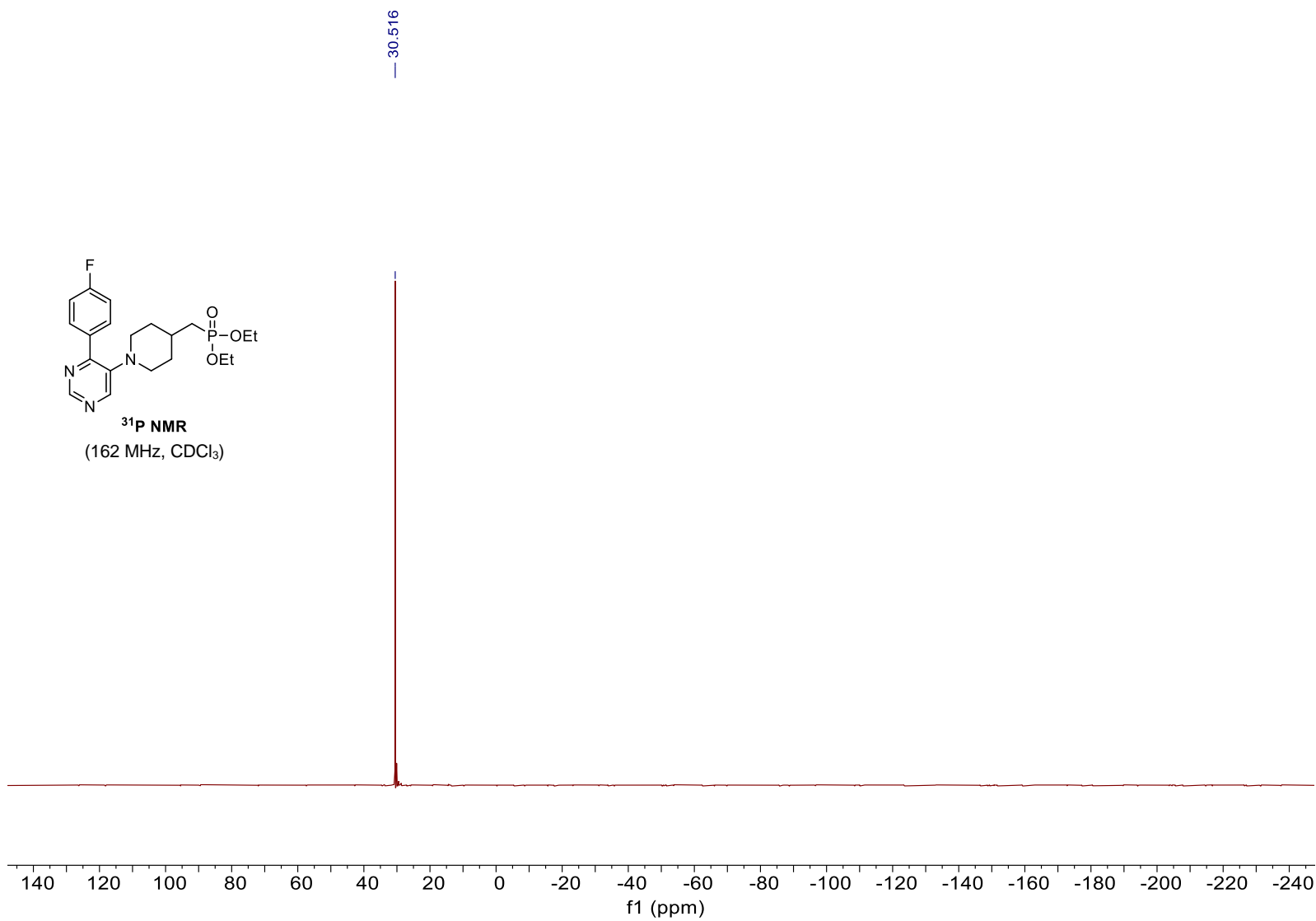


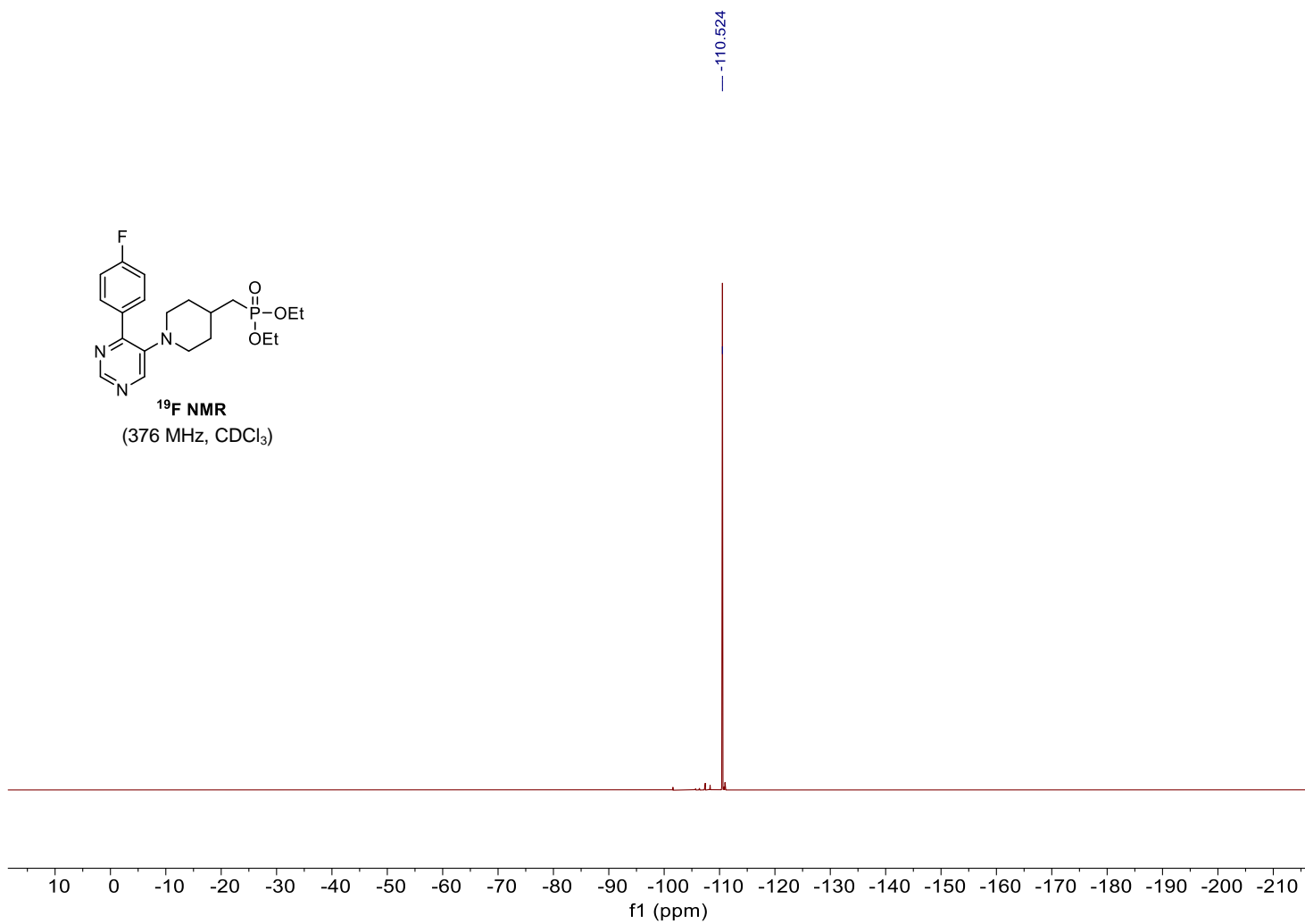
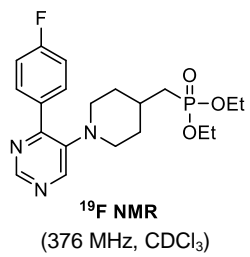
¹³C NMR
(100 MHz, CDCl₃)

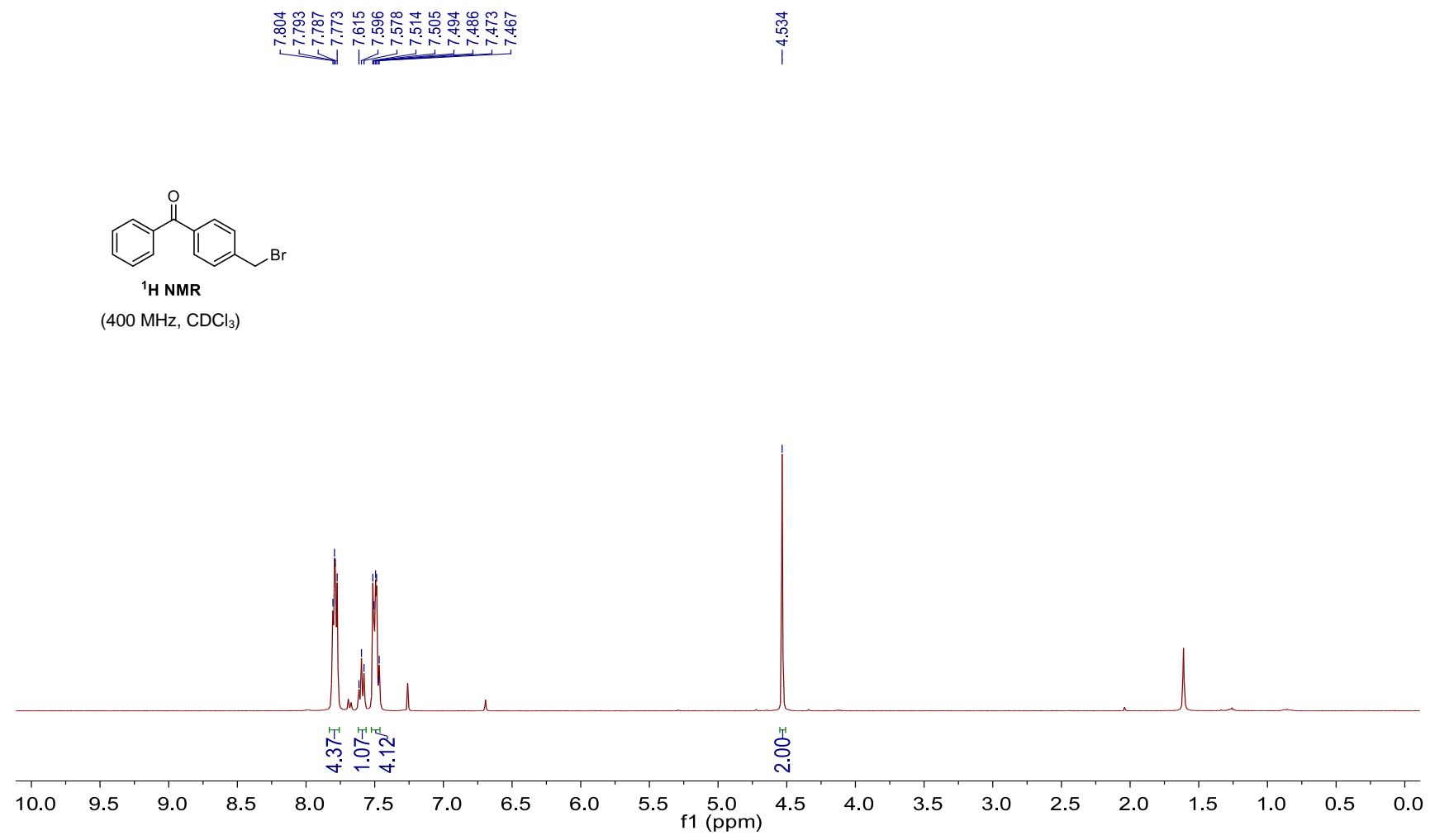
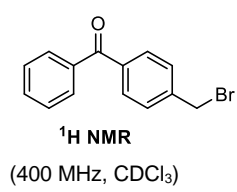


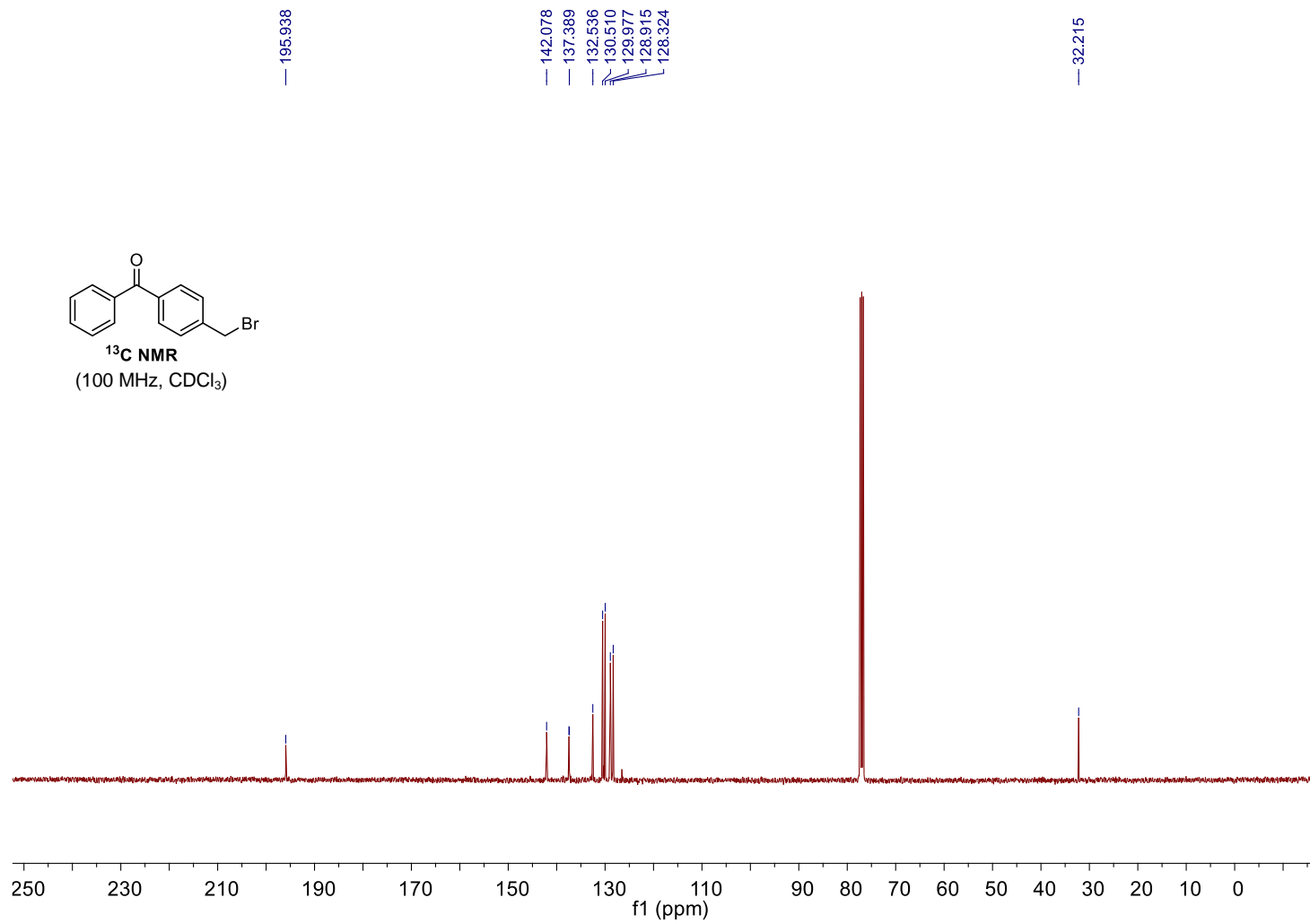


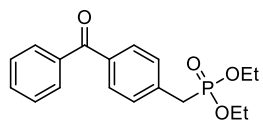
³¹P NMR
(162 MHz, CDCl₃)



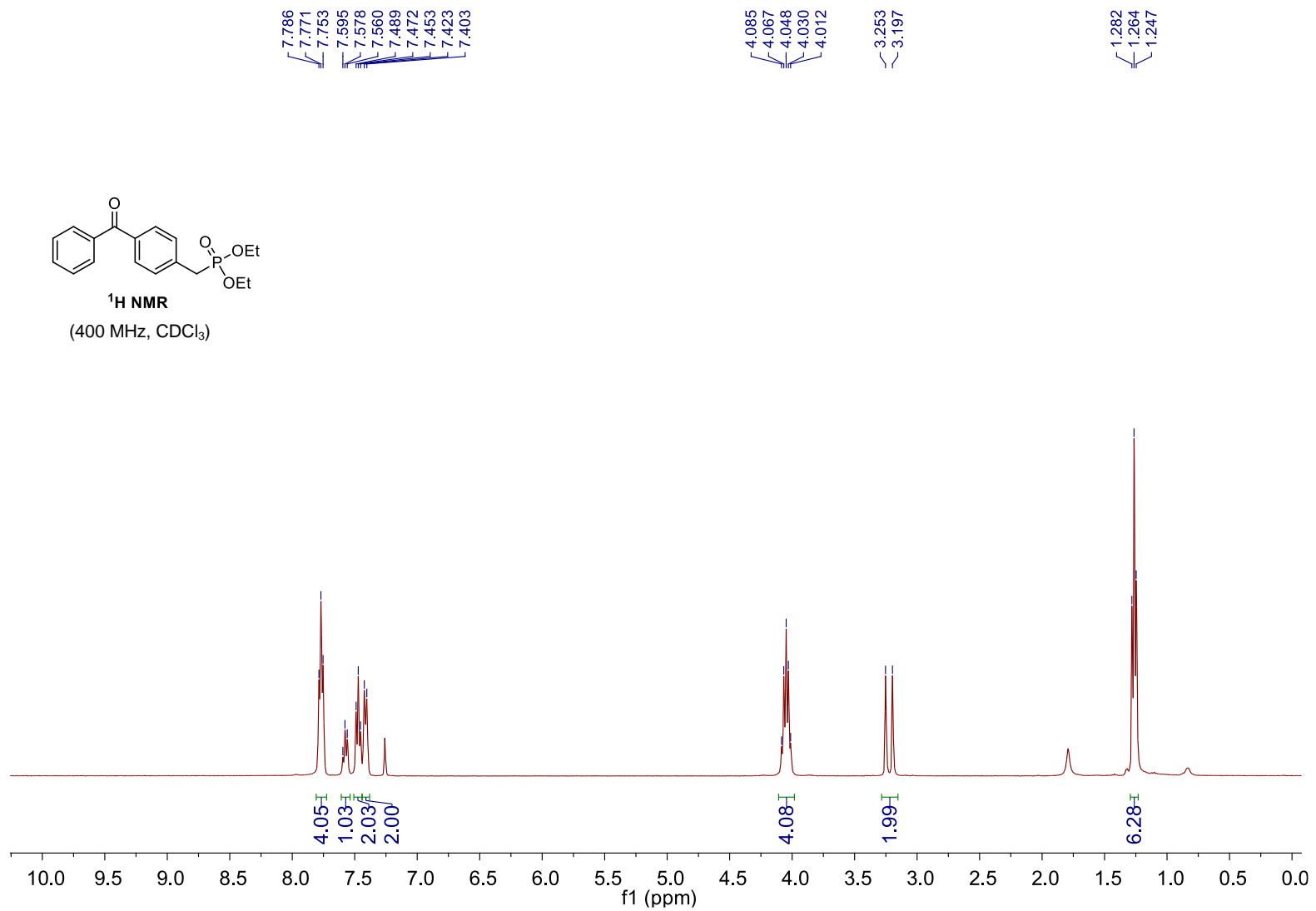


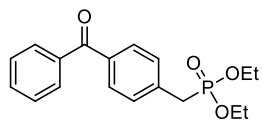




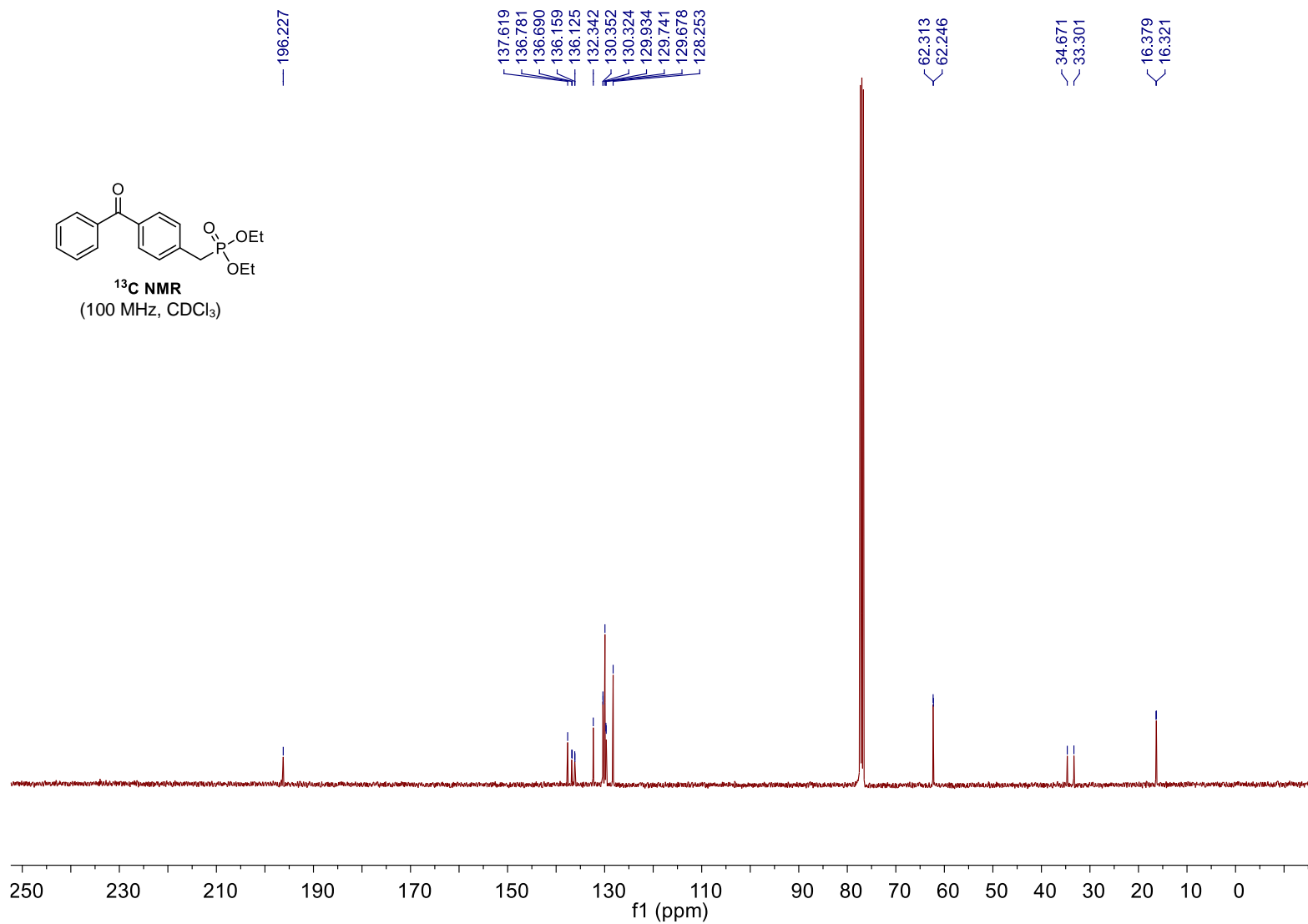


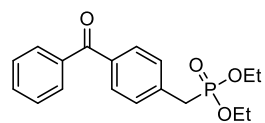
¹H NMR
(400 MHz, CDCl₃)



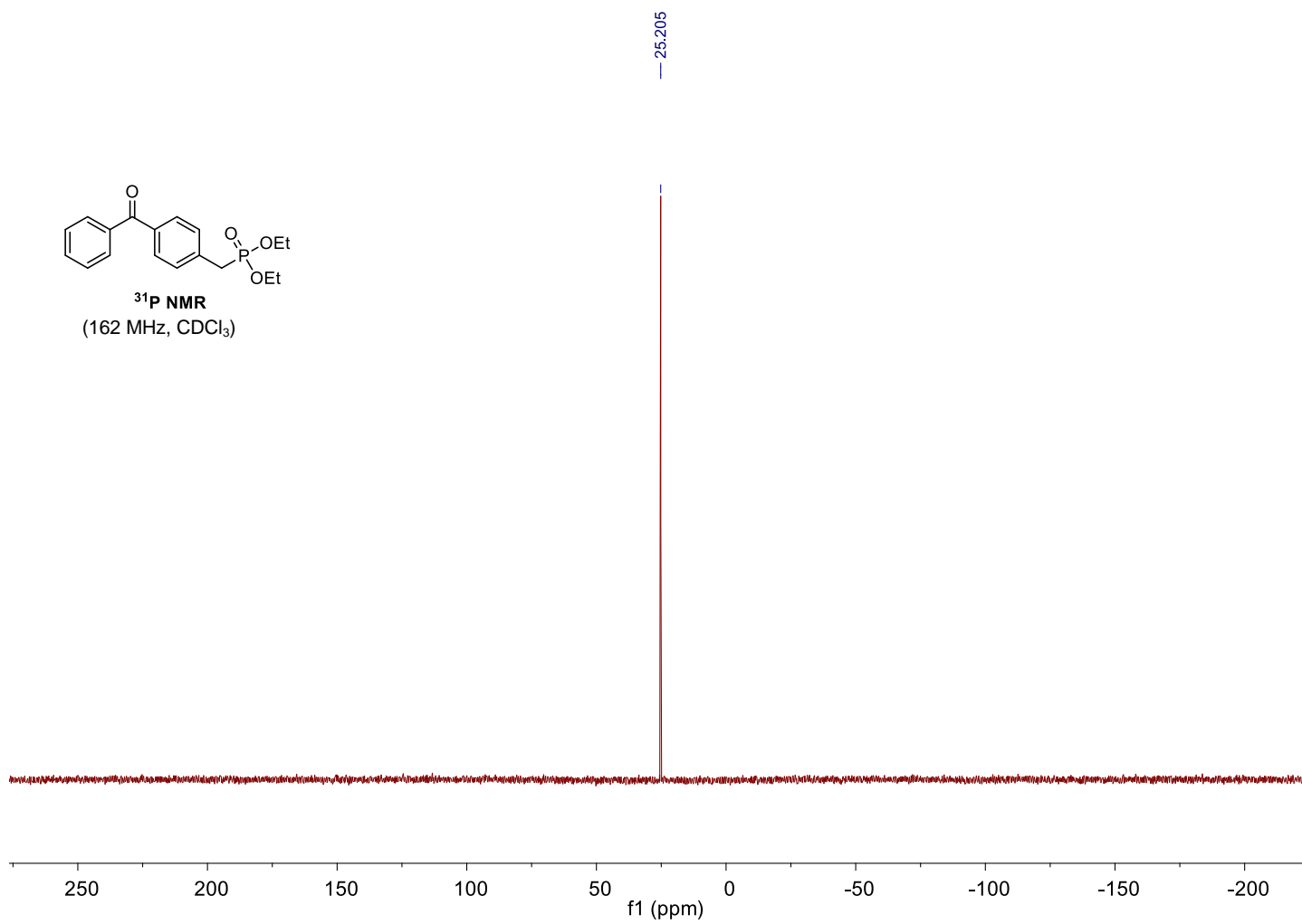


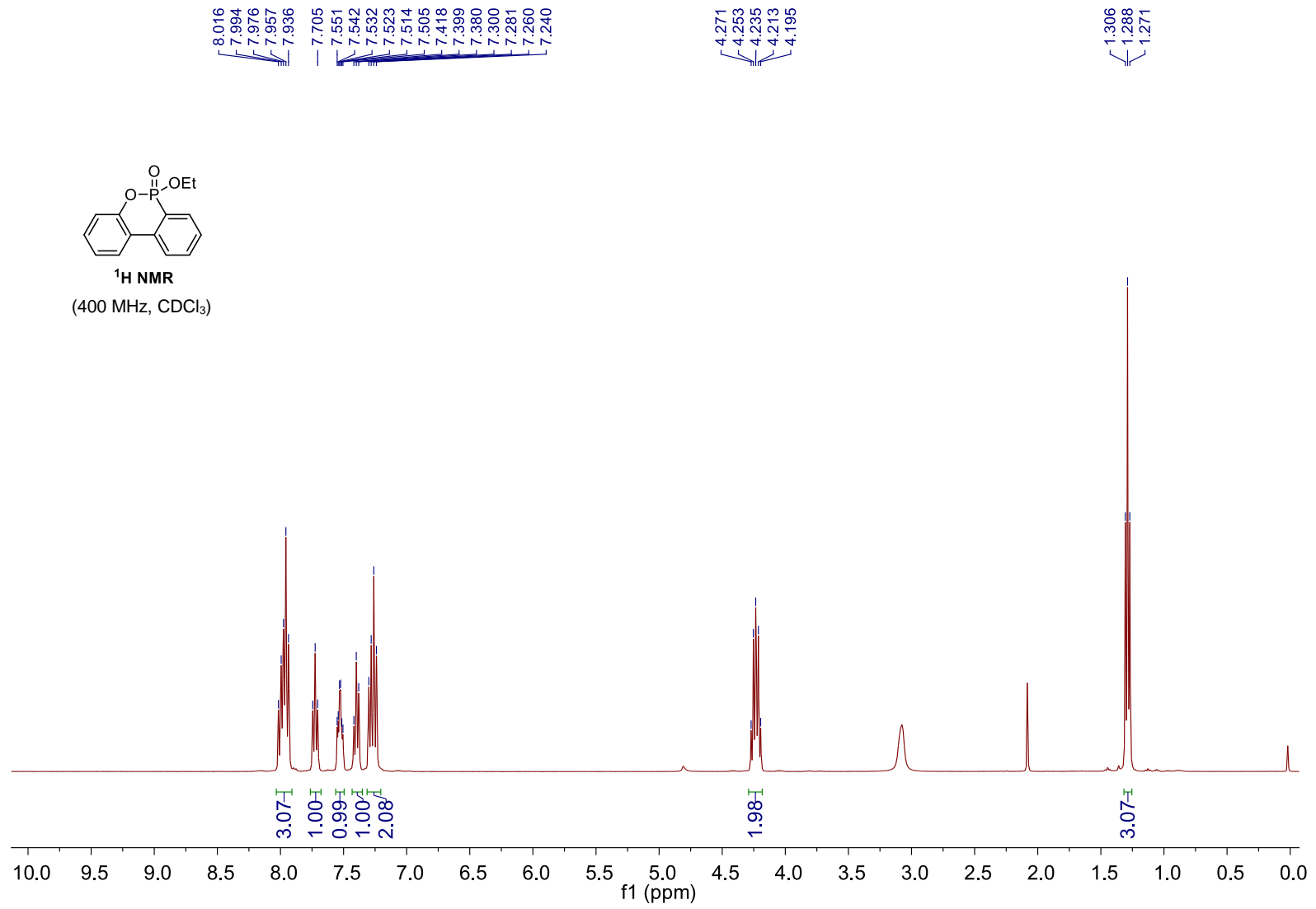
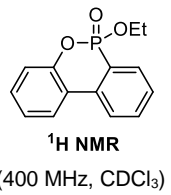
¹³C NMR
(100 MHz, CDCl₃)

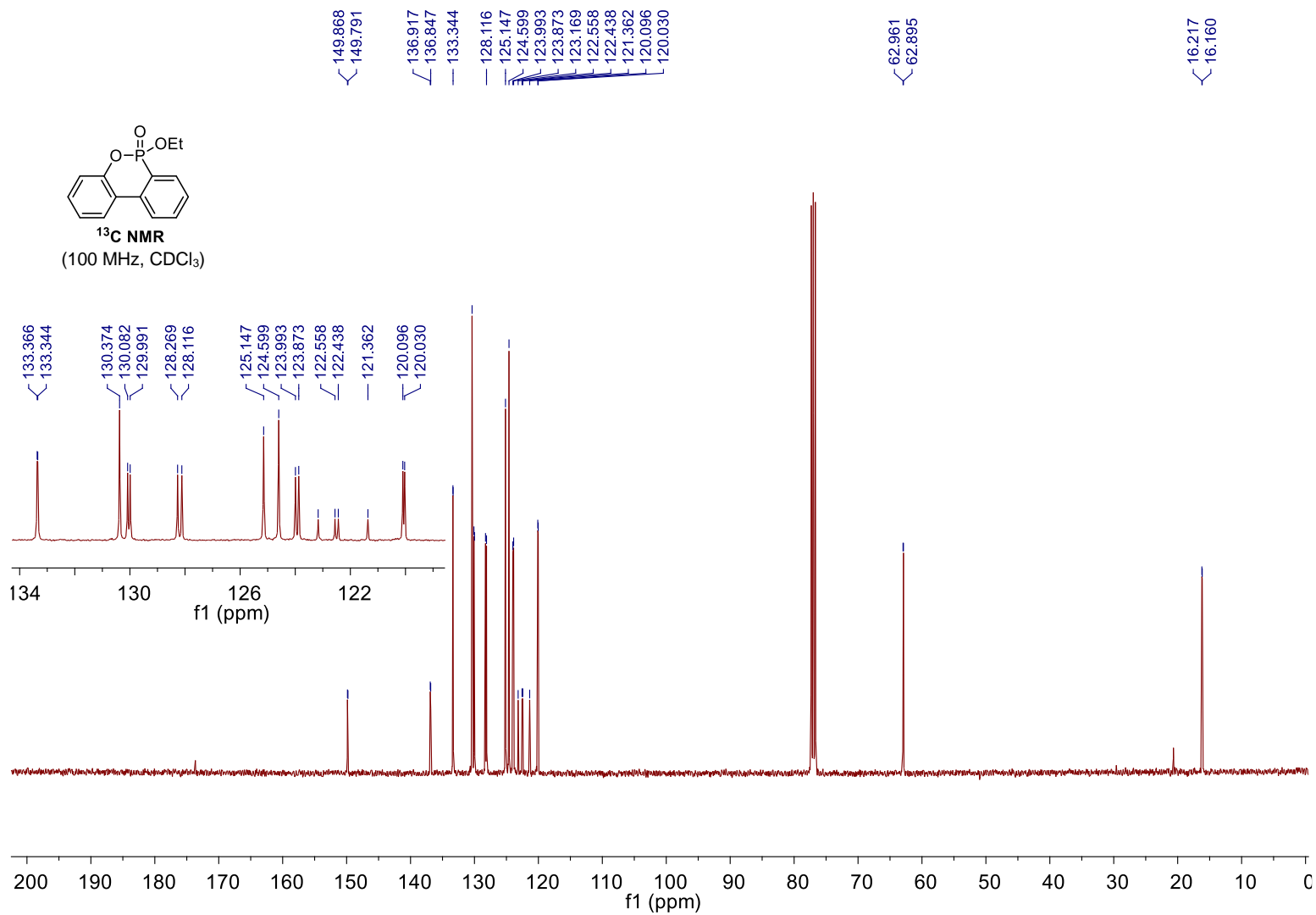
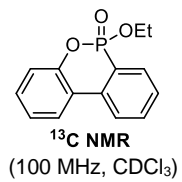


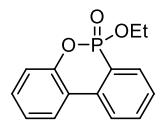


³¹P NMR
(162 MHz, CDCl₃)

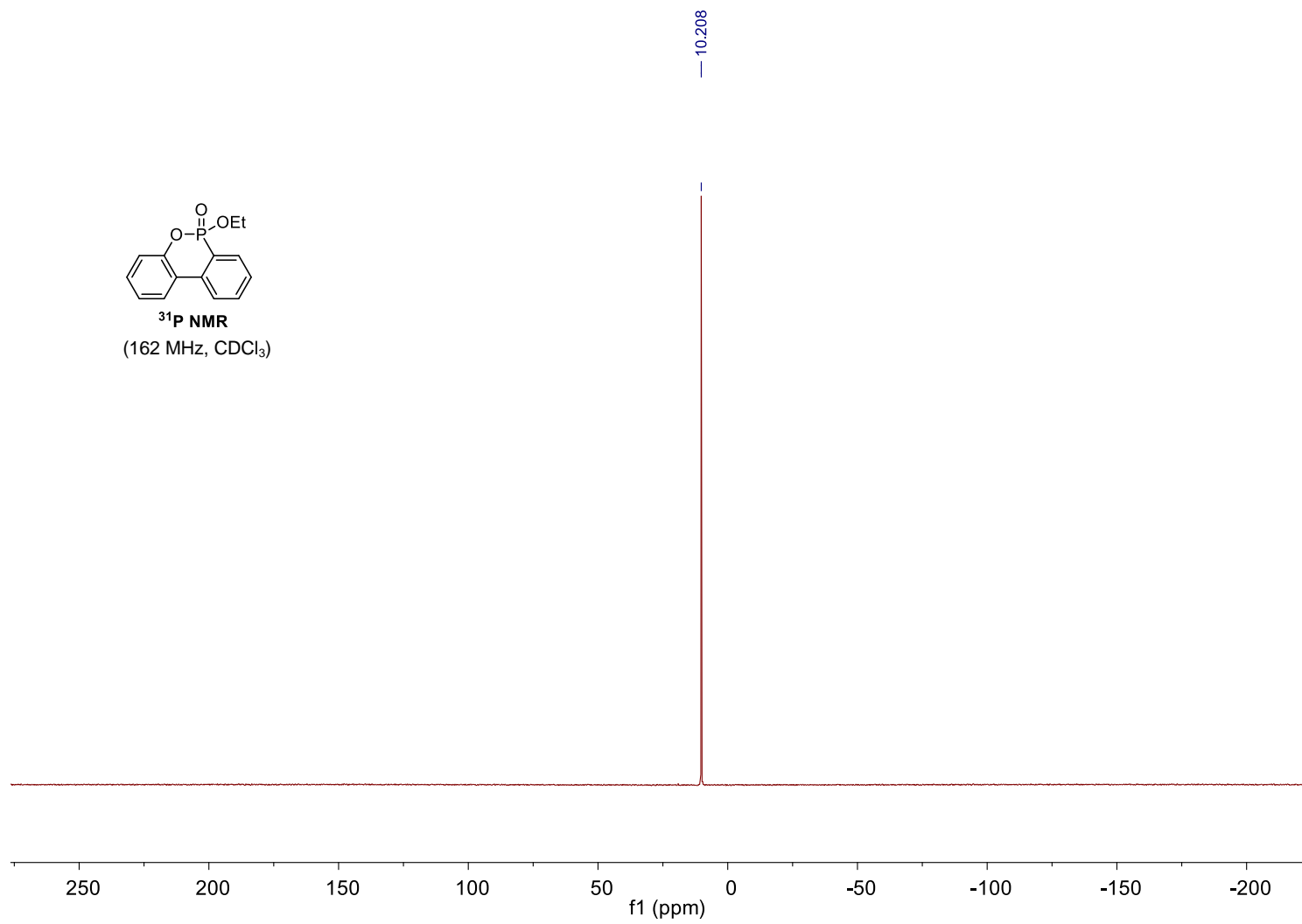


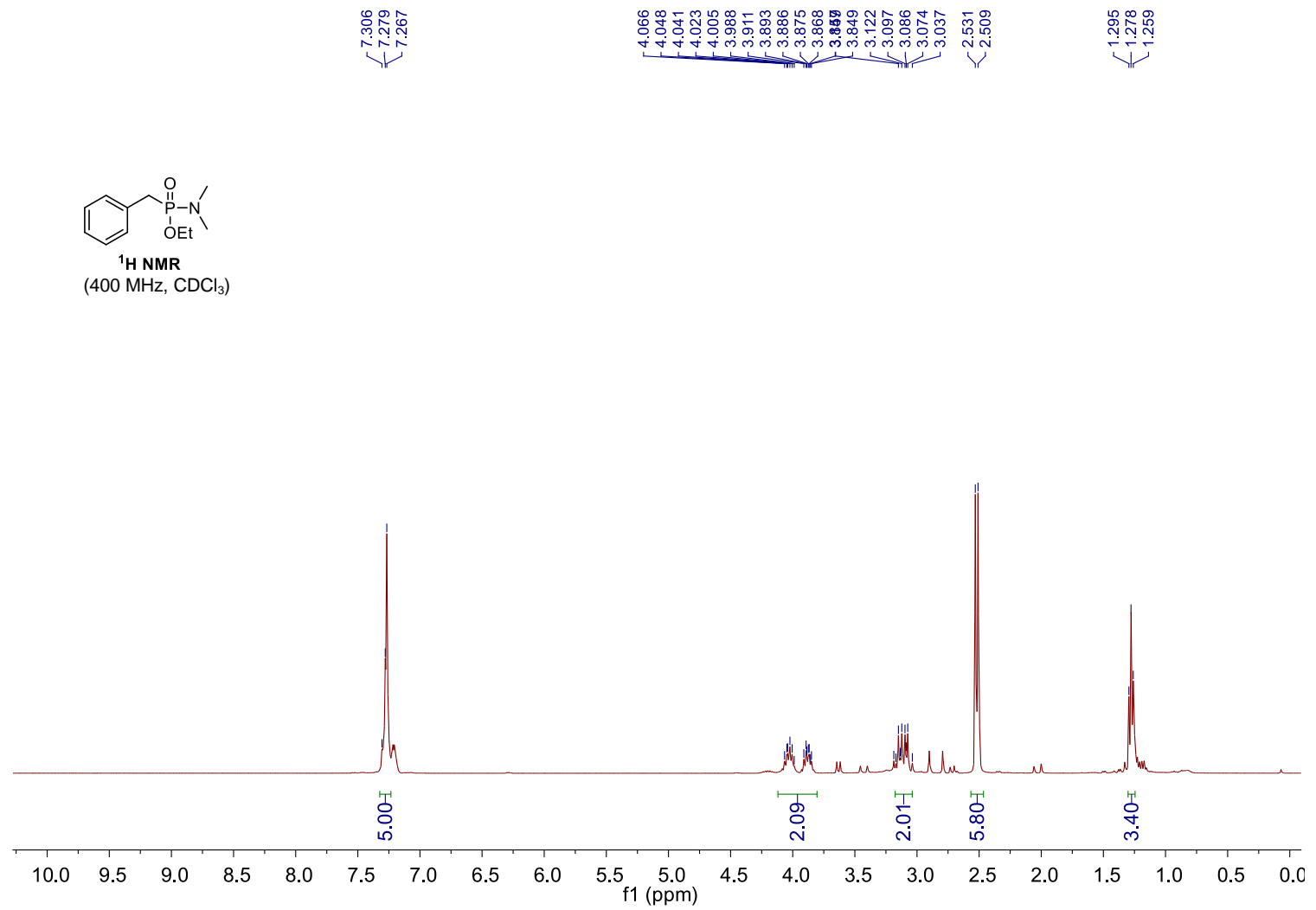


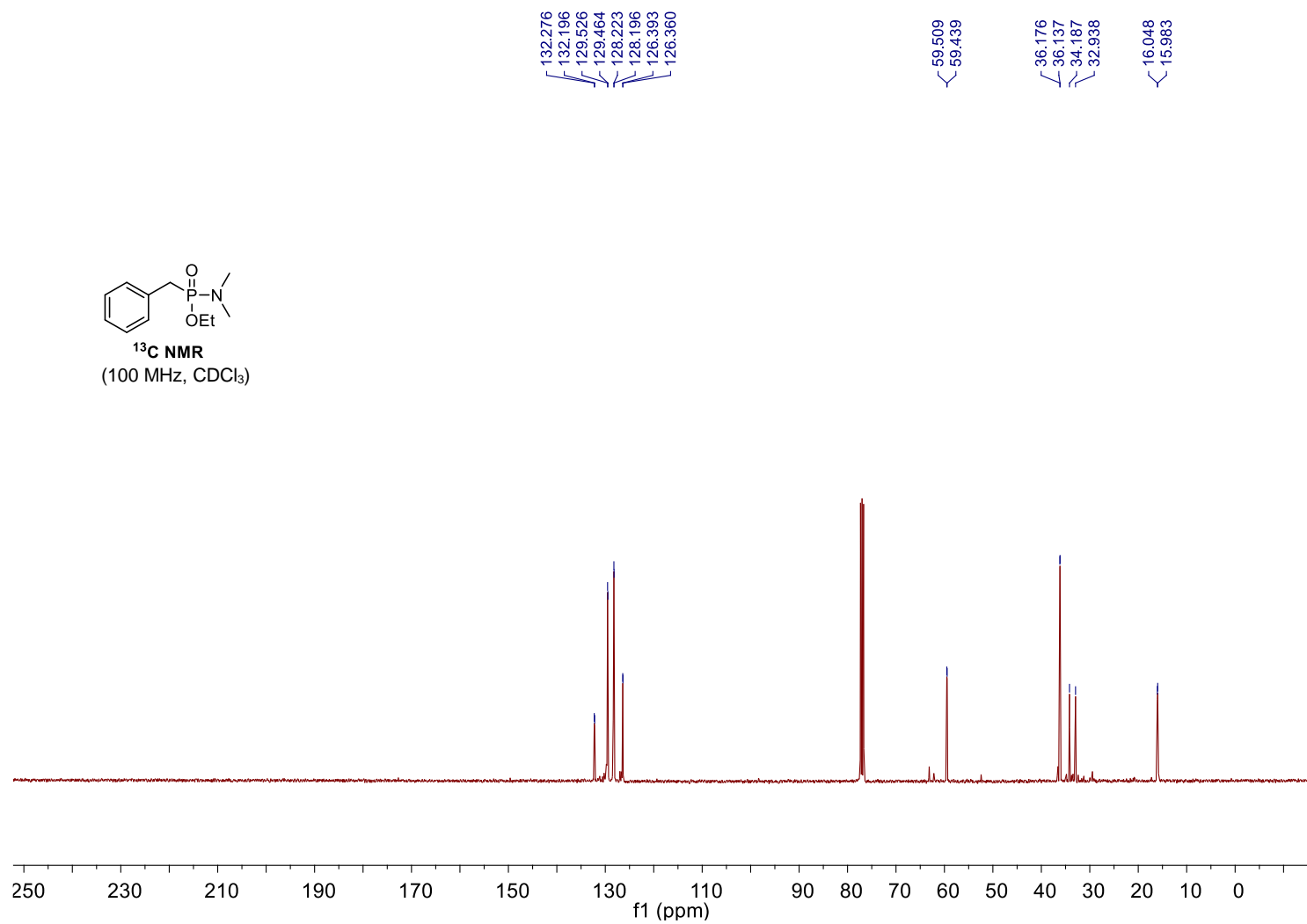
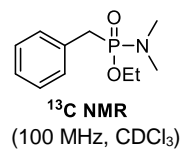


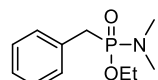


³¹P NMR
(162 MHz, CDCl₃)

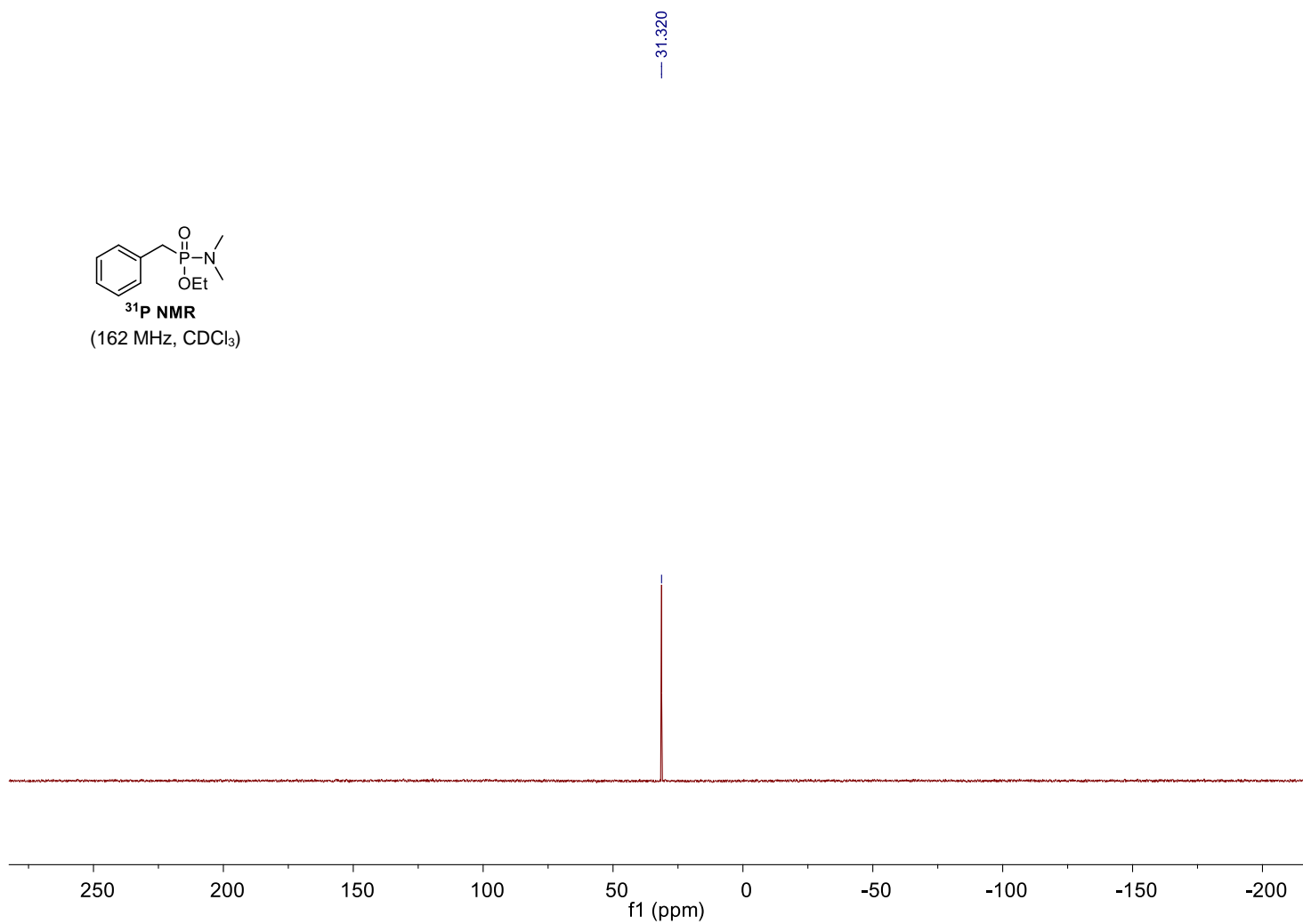


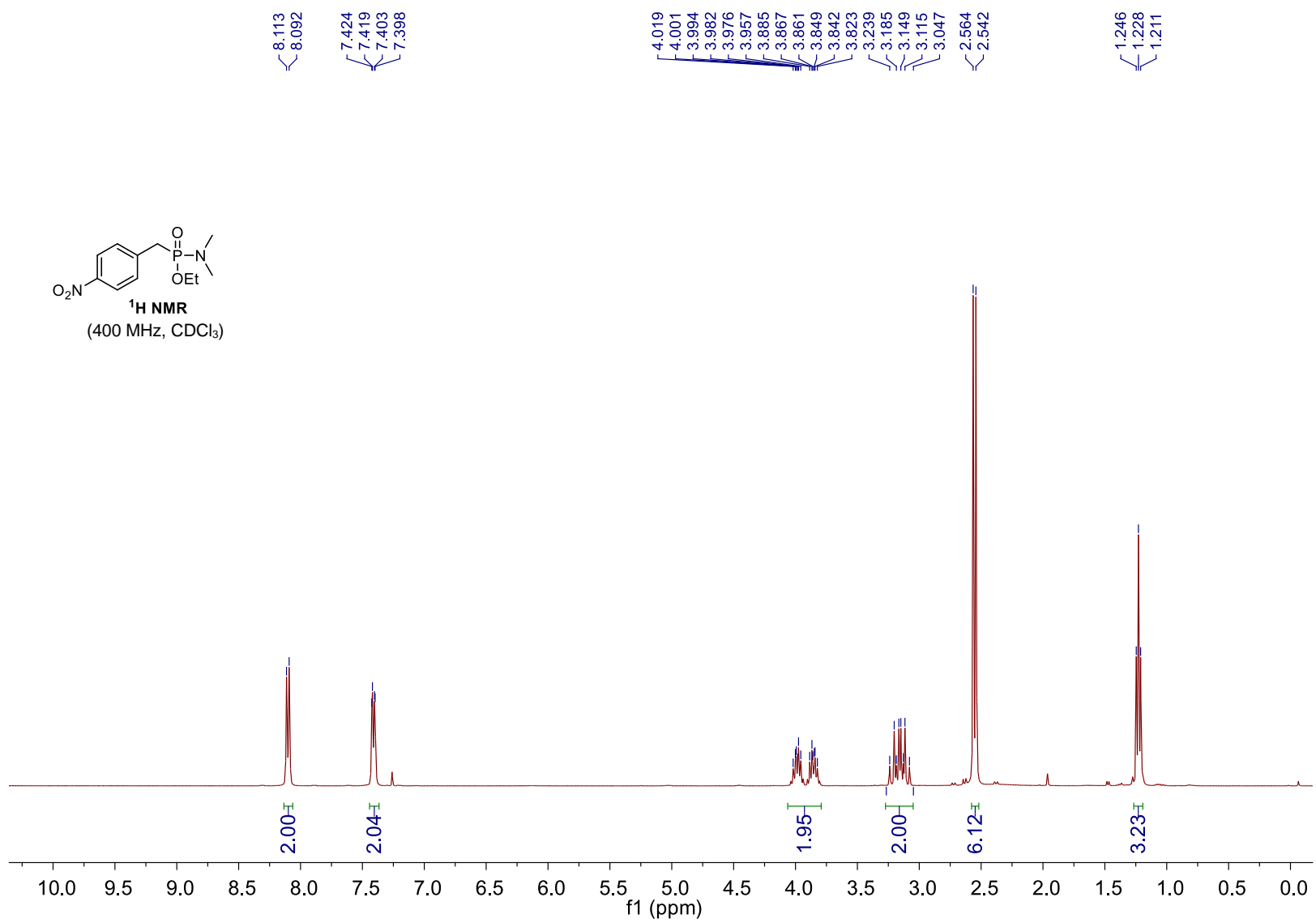
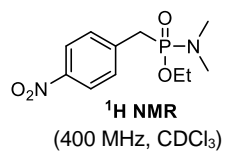


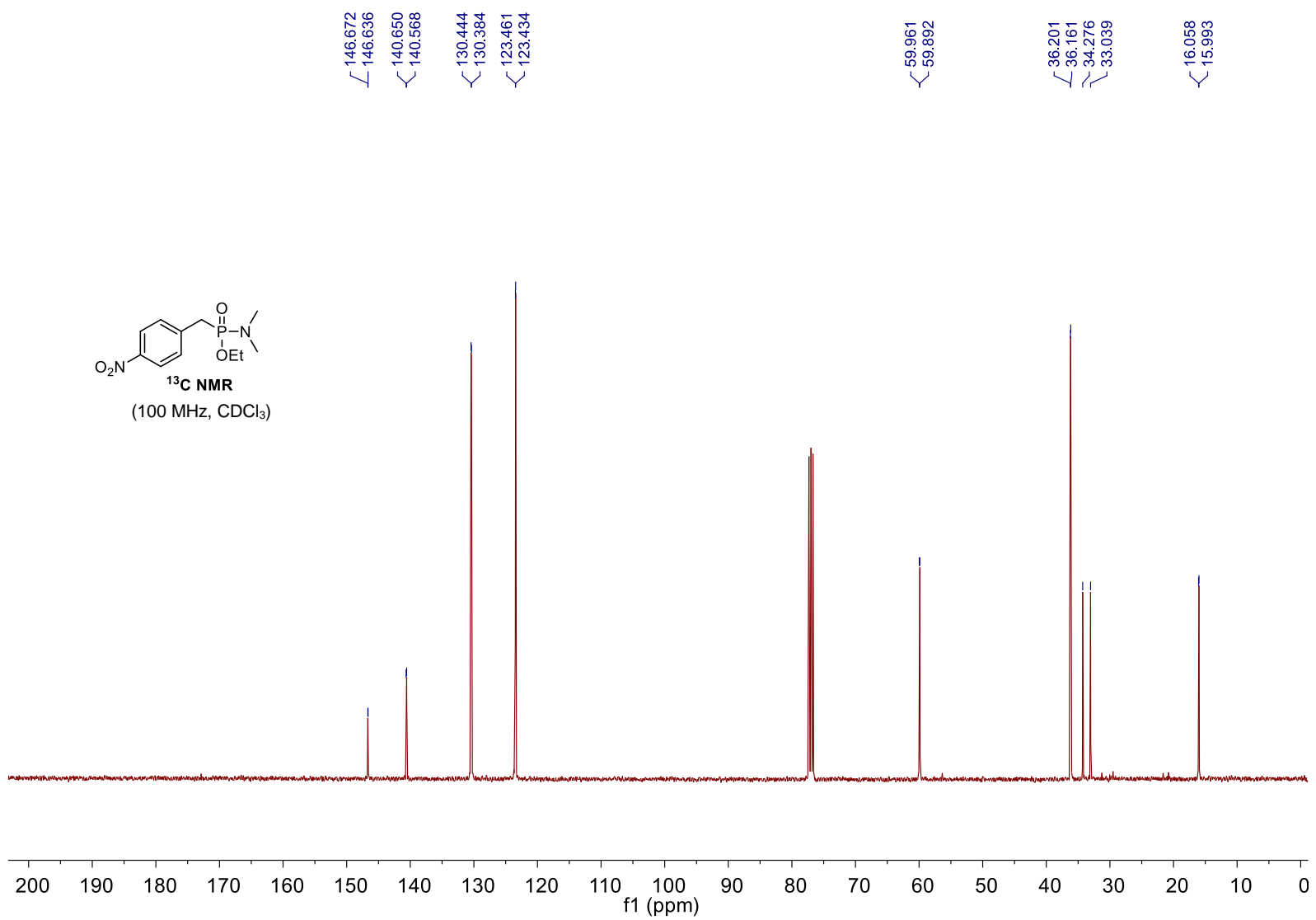


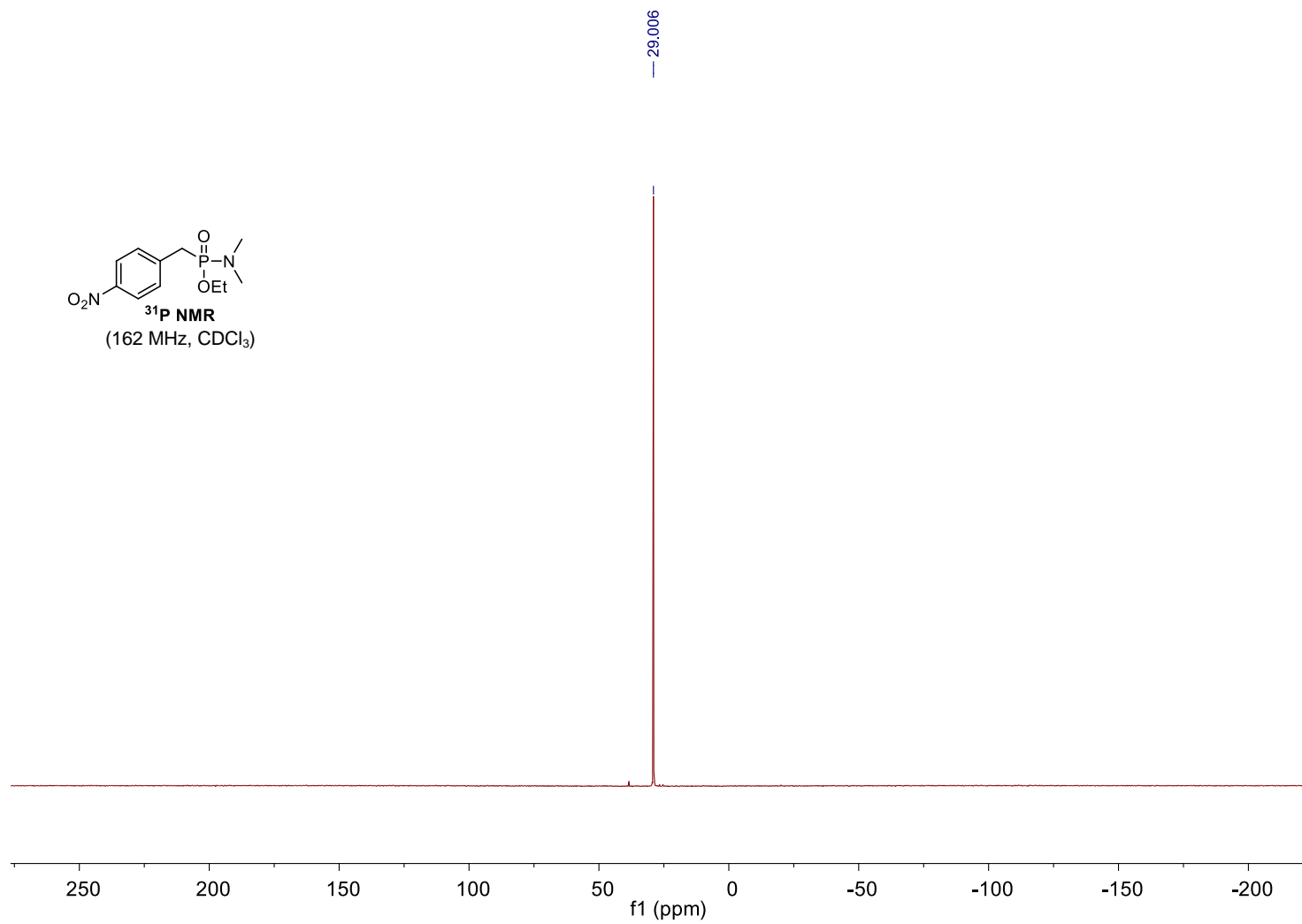
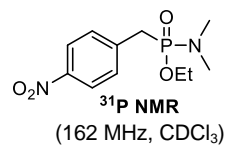


³¹P NMR
(162 MHz, CDCl₃)

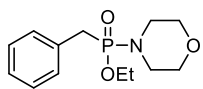




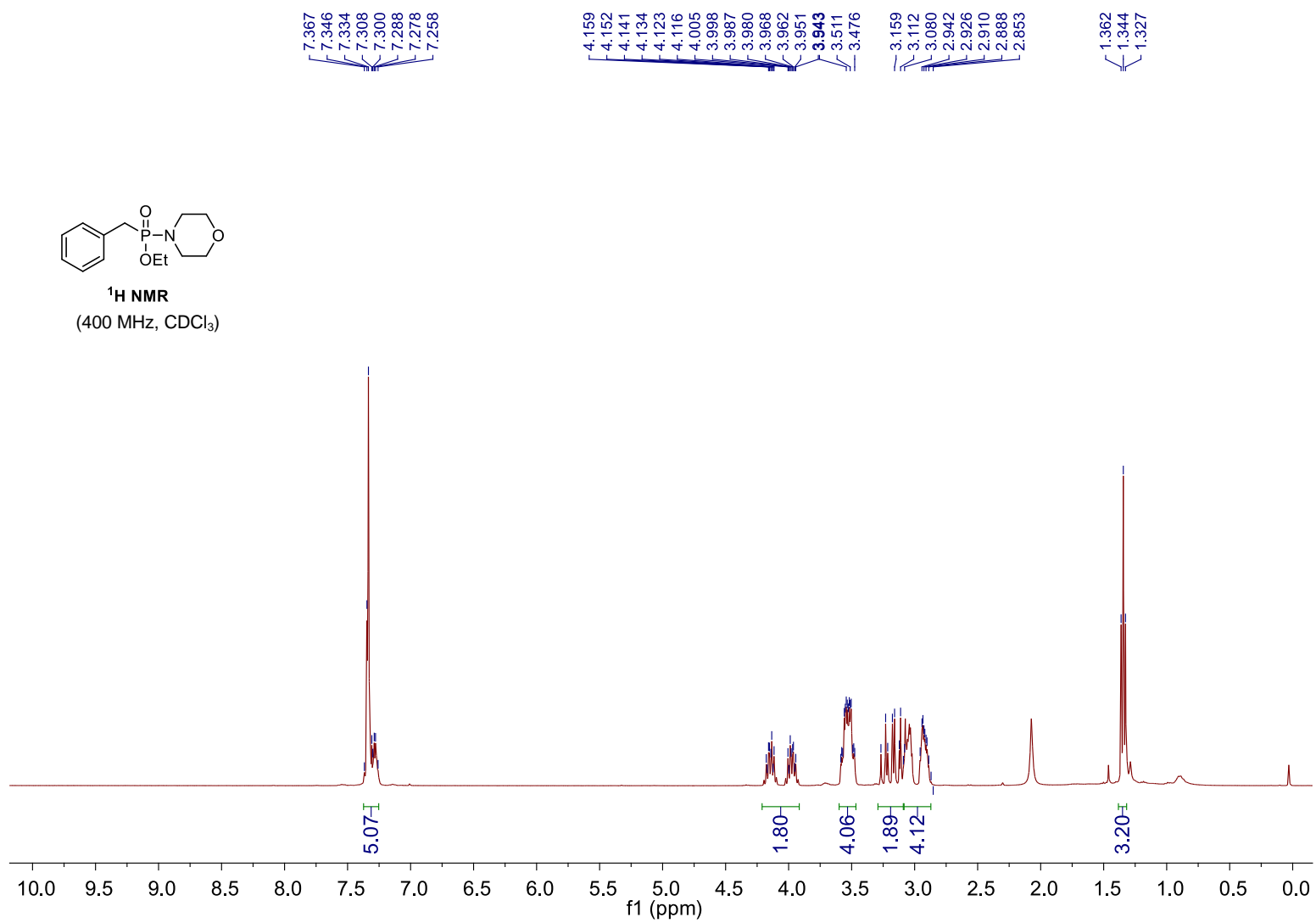


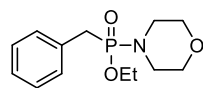


S178

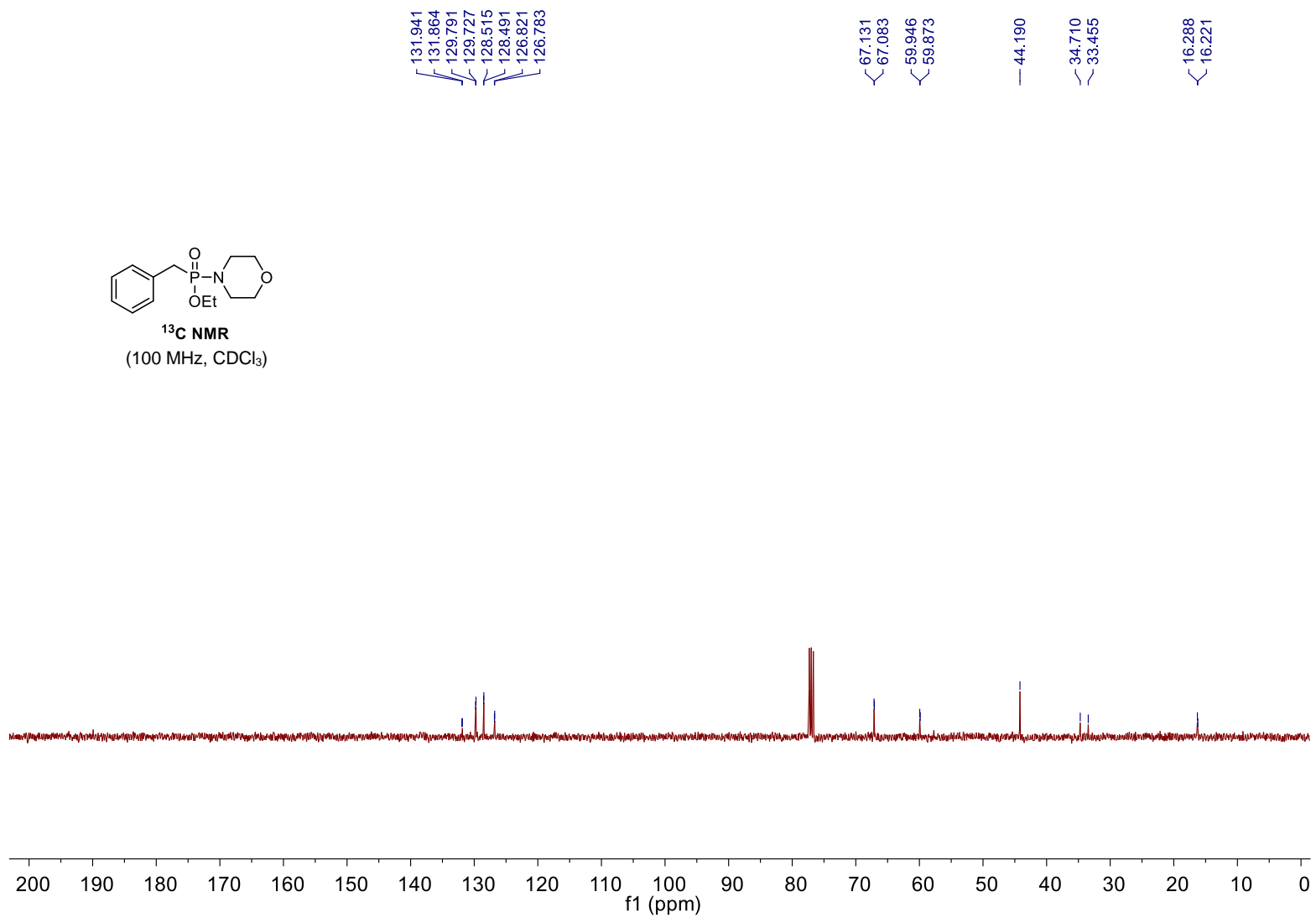


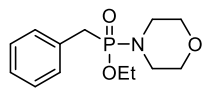
¹H NMR
(400 MHz, CDCl₃)



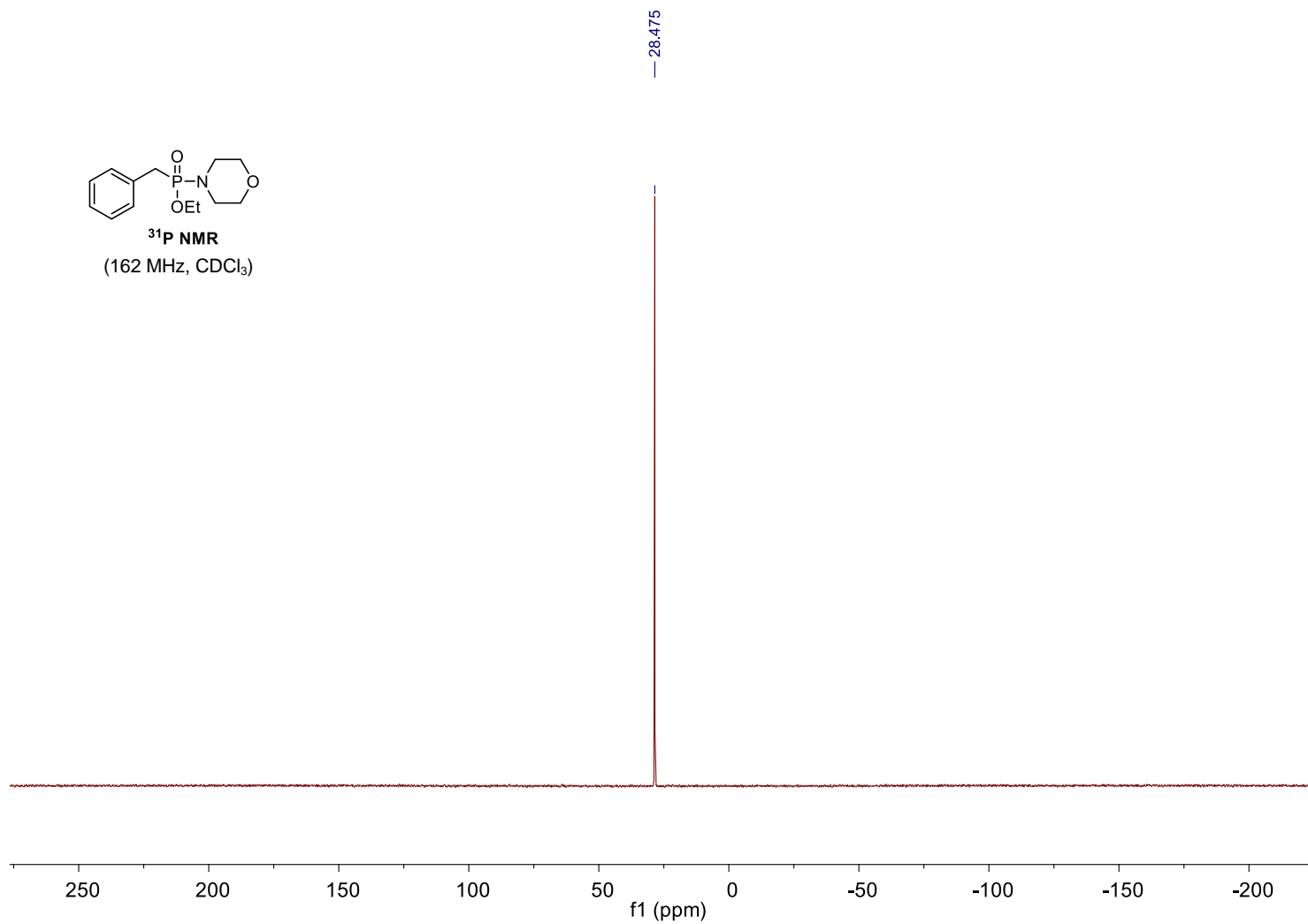


¹³C NMR
(100 MHz, CDCl₃)

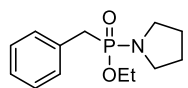




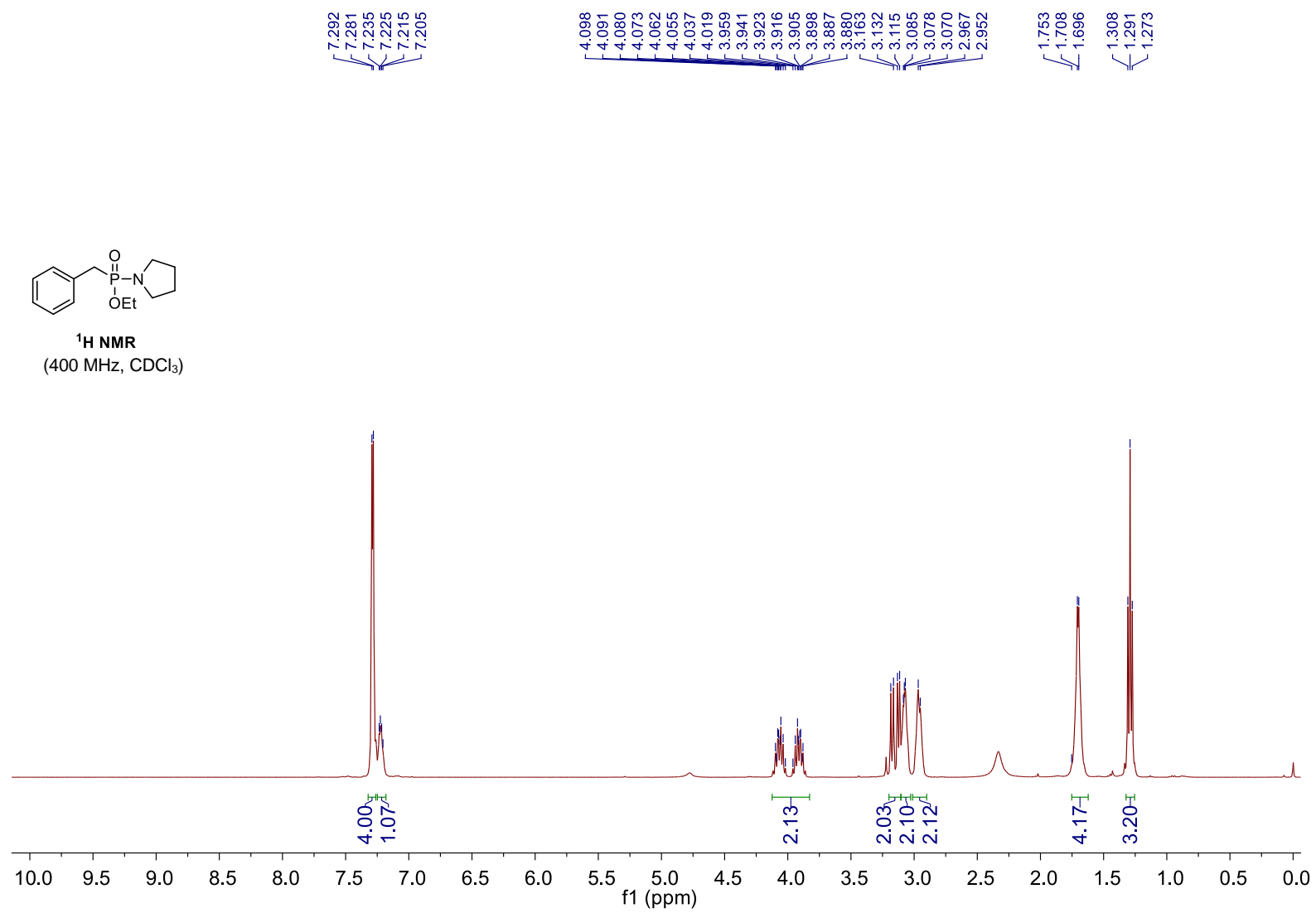
³¹P NMR
(162 MHz, CDCl₃)

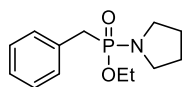


S181

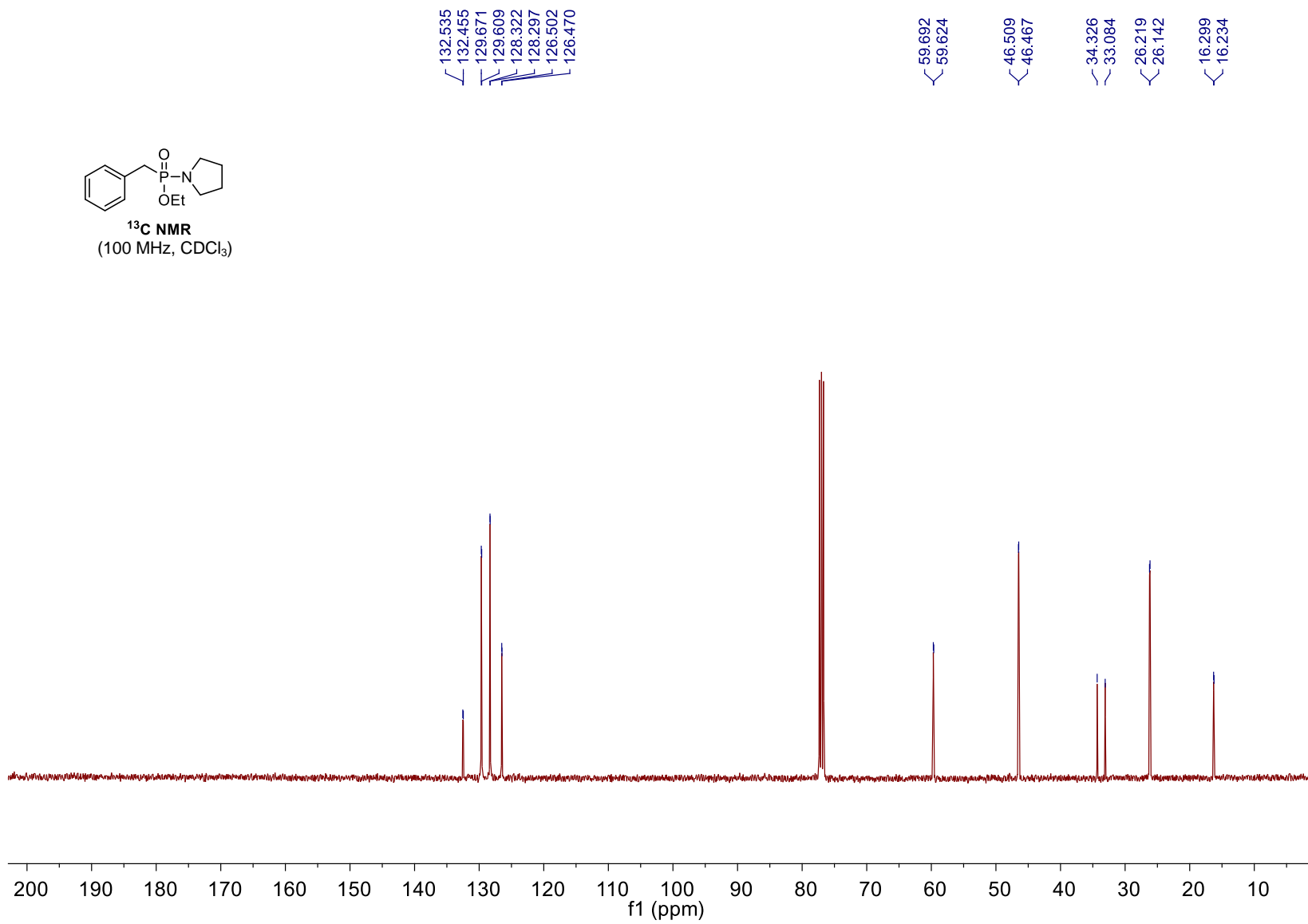


¹H NMR
(400 MHz, CDCl₃)

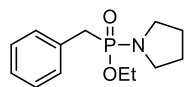




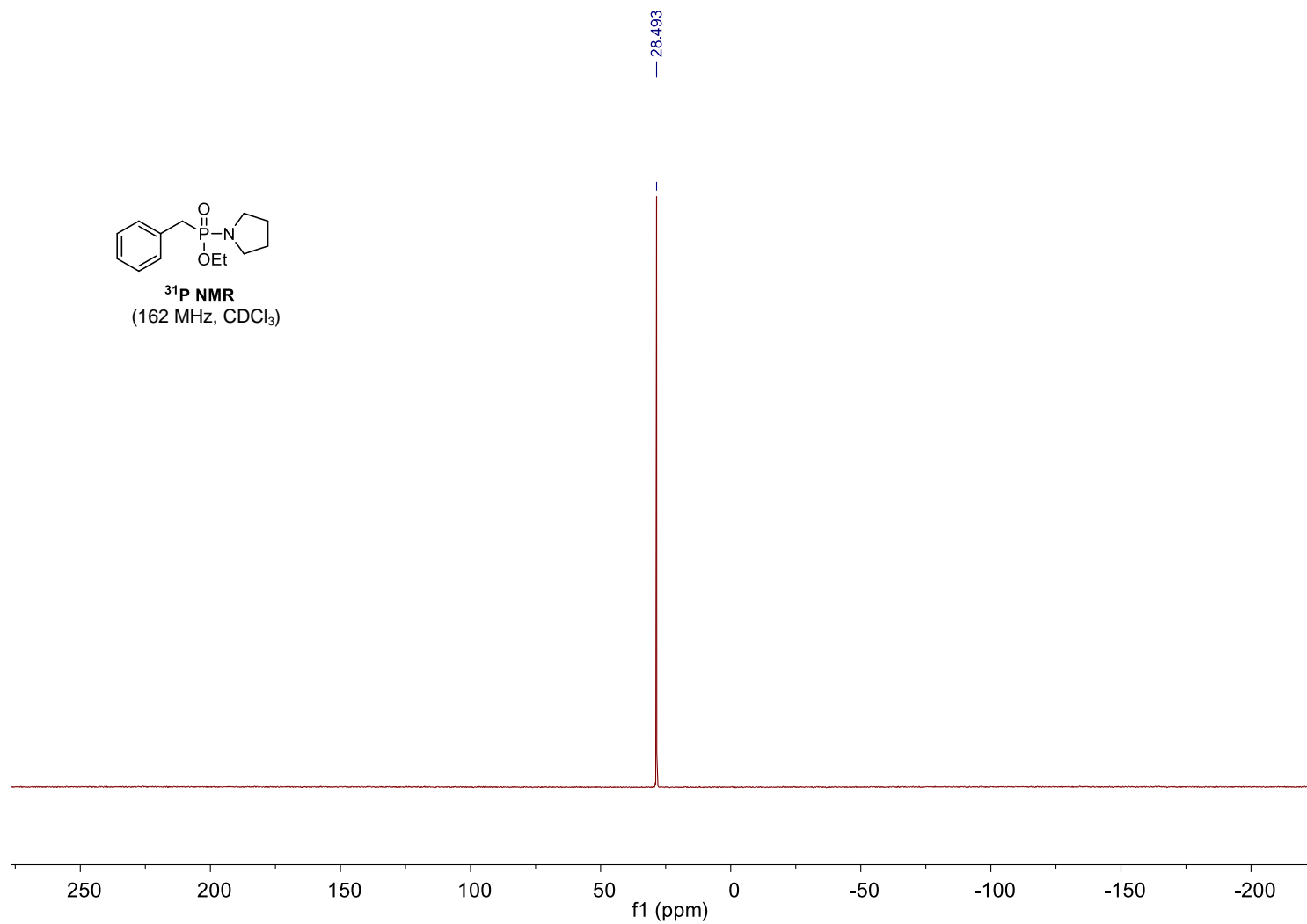
¹³C NMR
(100 MHz, CDCl₃)

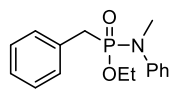


S183

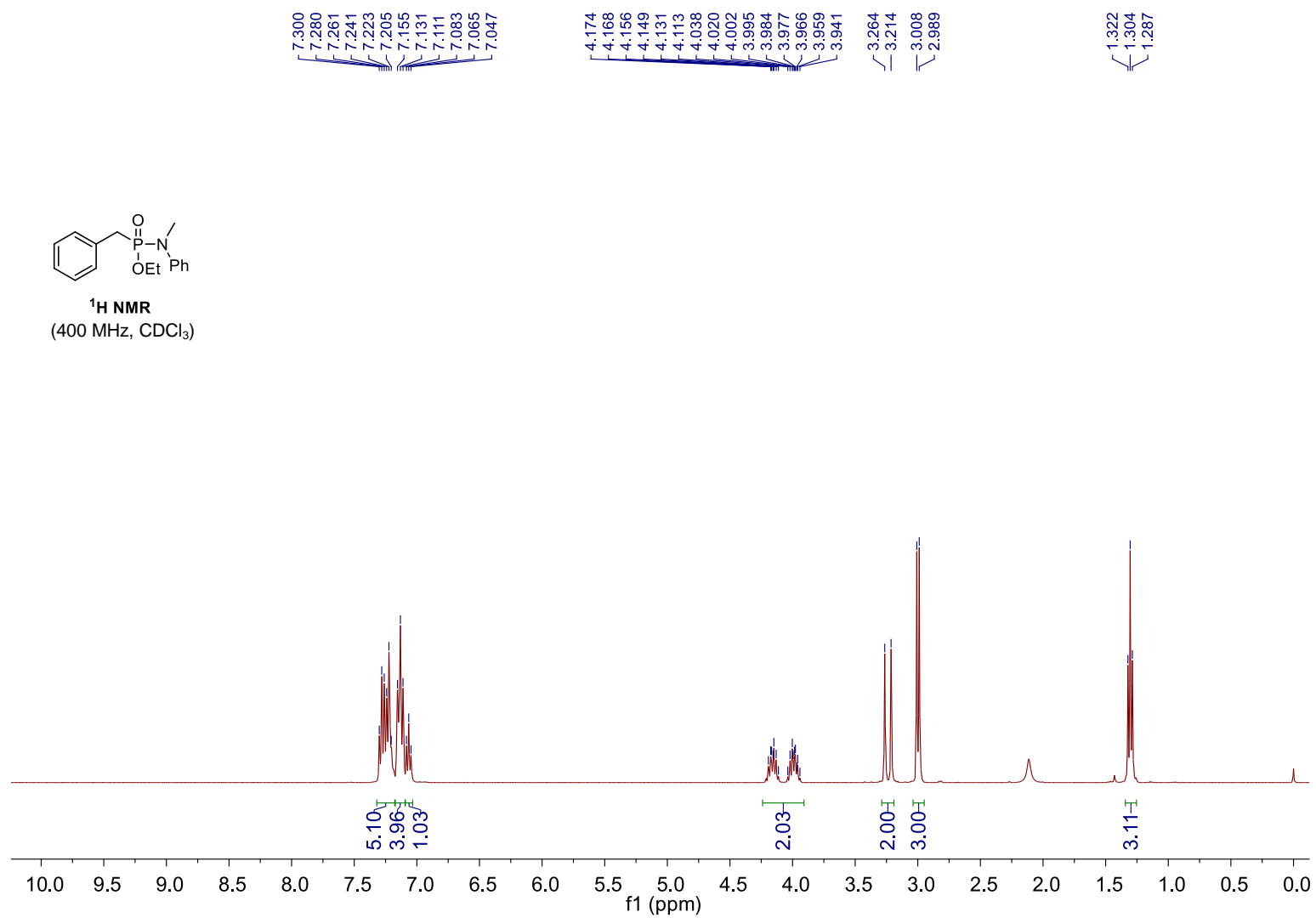


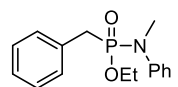
³¹P NMR
(162 MHz, CDCl₃)



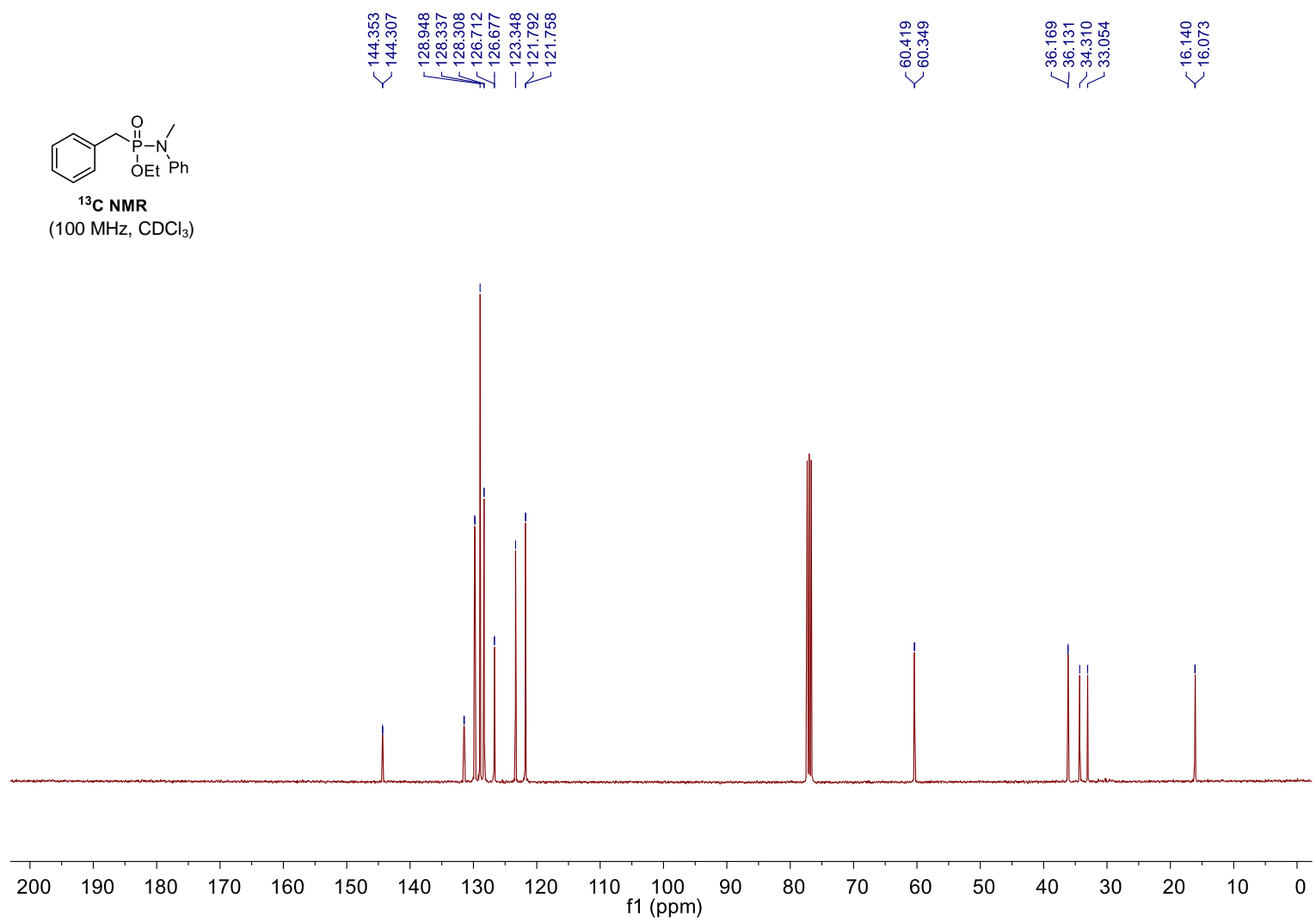


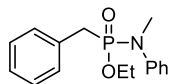
¹H NMR
(400 MHz, CDCl₃)



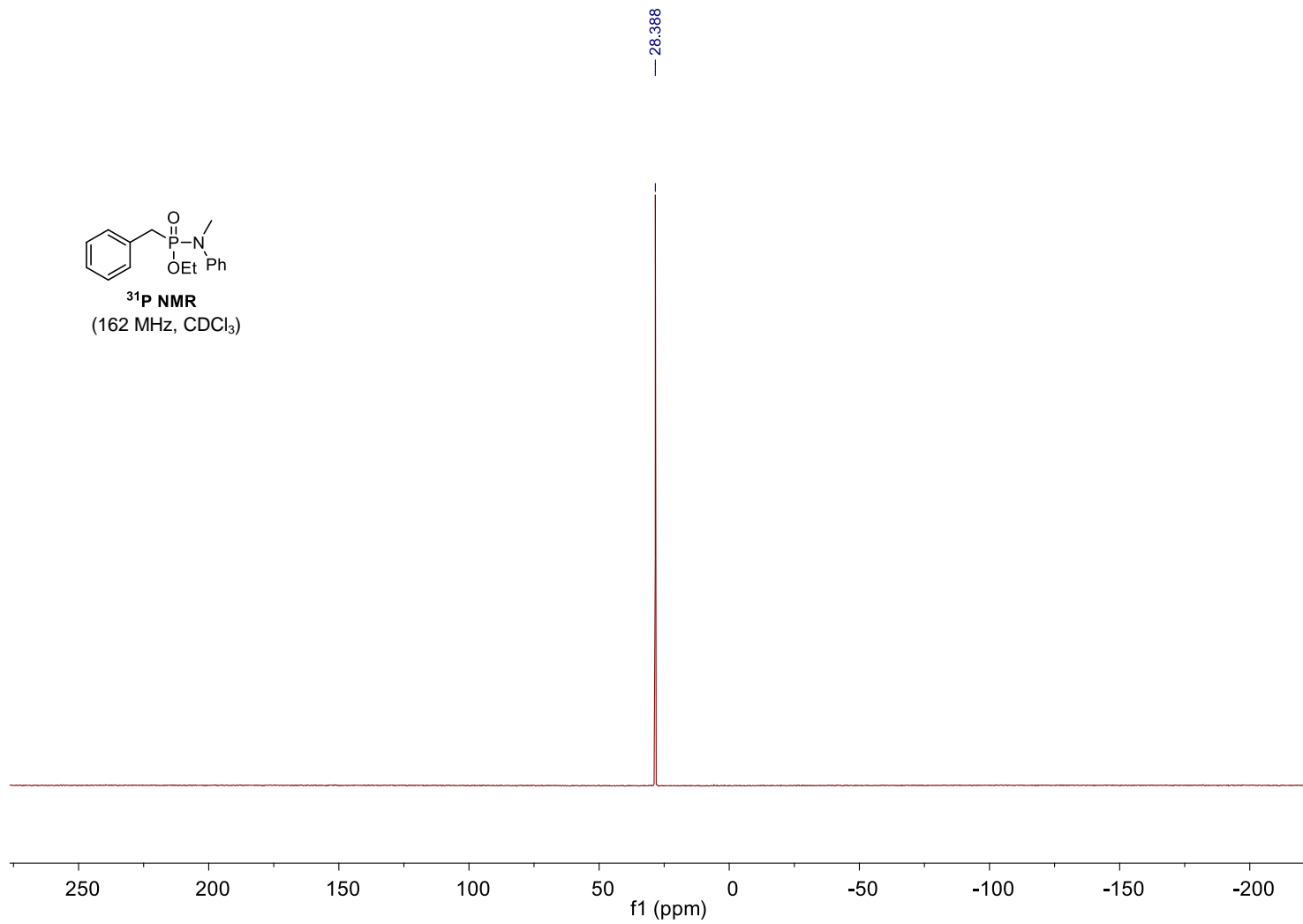


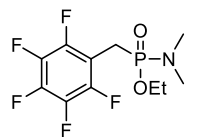
¹³C NMR
(100 MHz, CDCl₃)



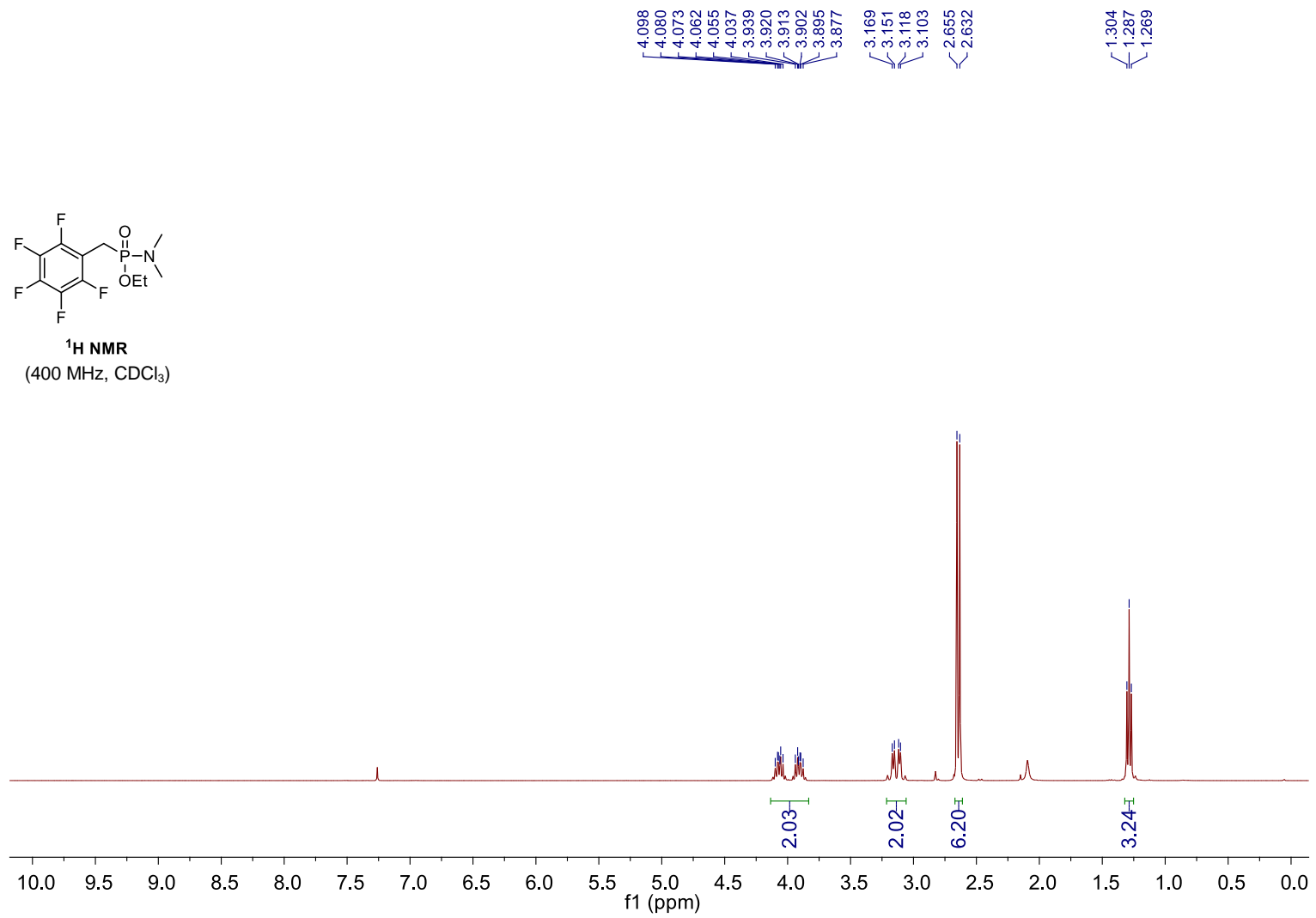


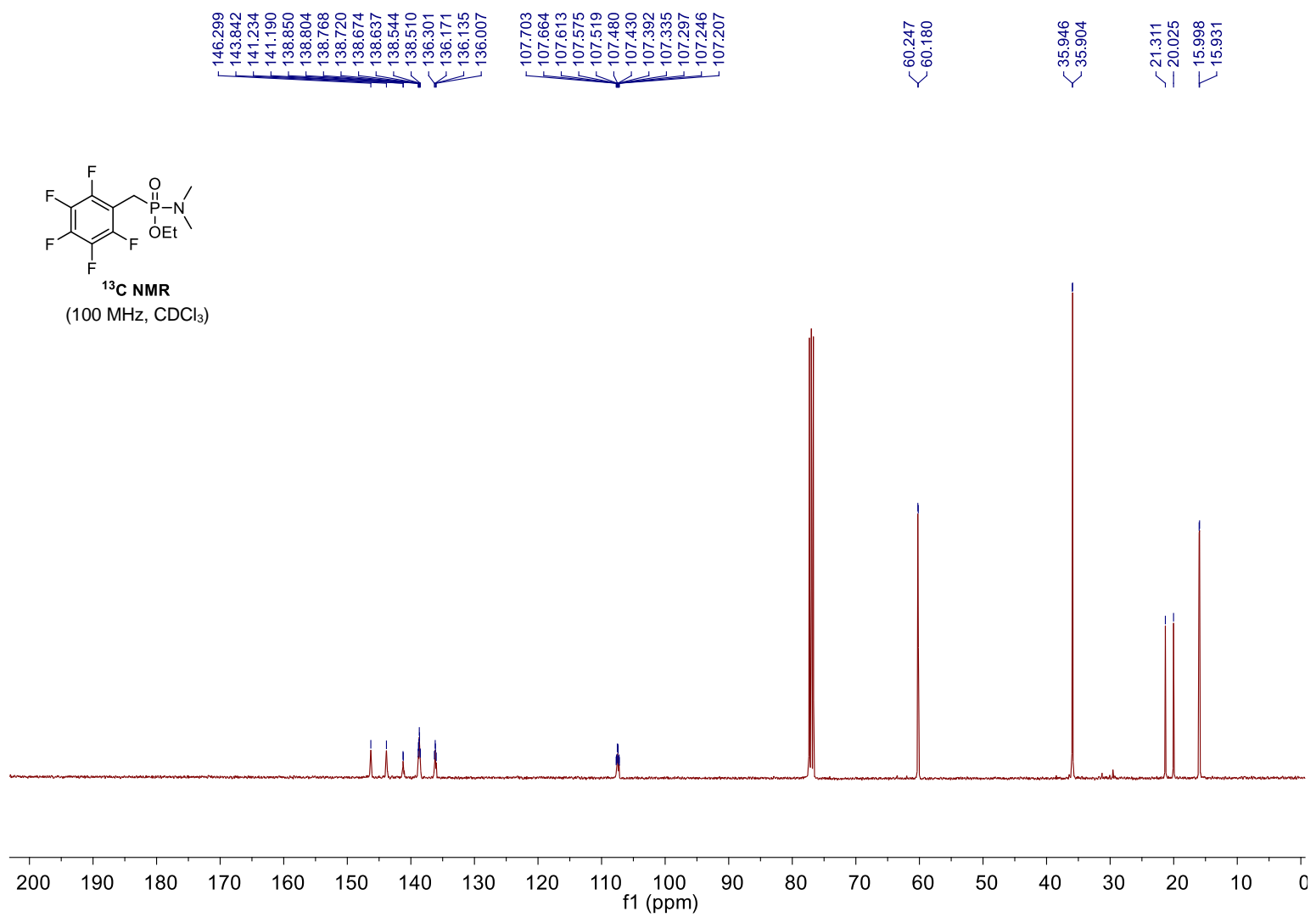
³¹P NMR
(162 MHz, CDCl₃)

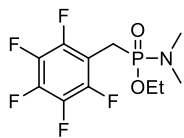




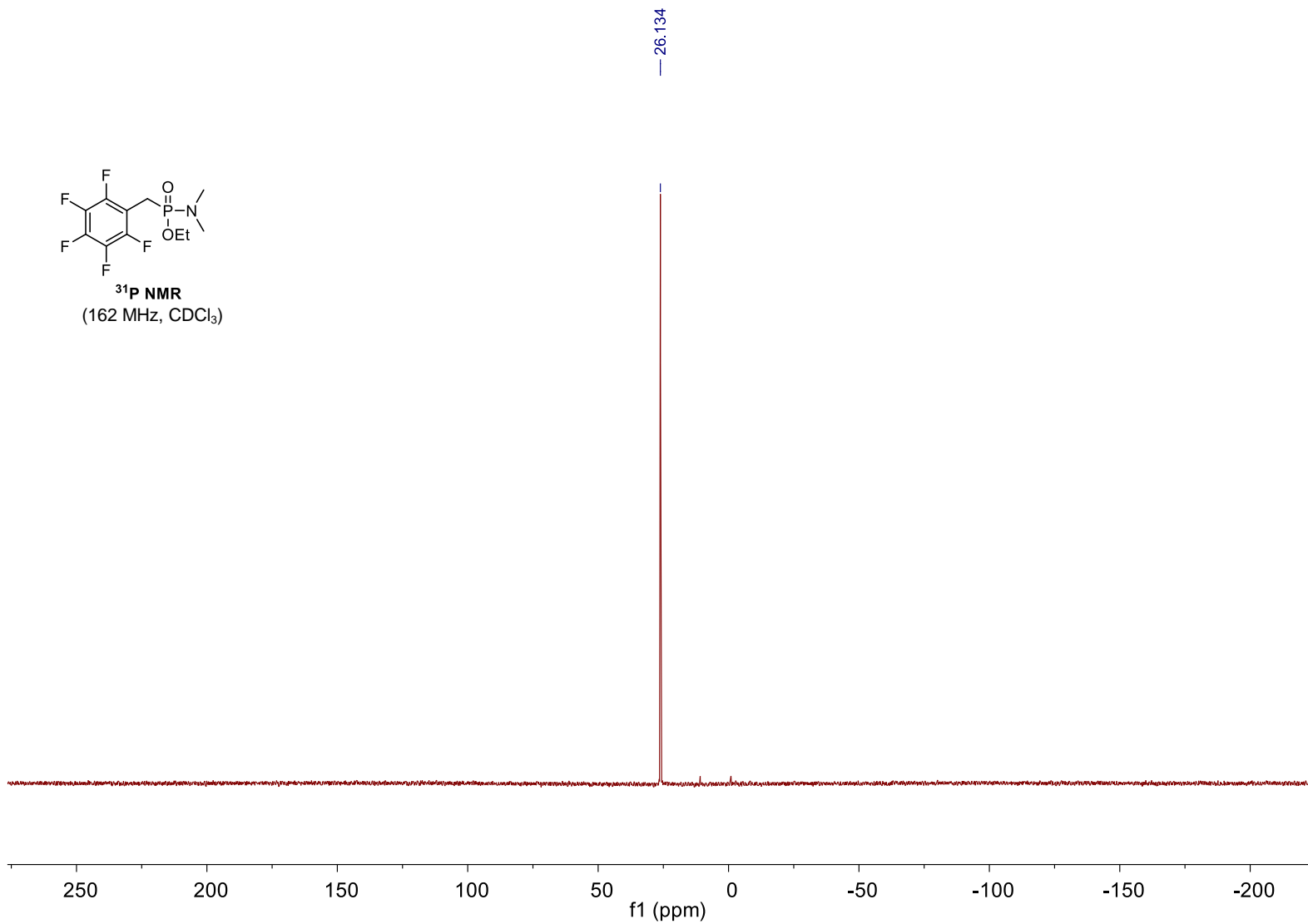
¹H NMR
(400 MHz, CDCl₃)



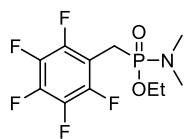




³¹P NMR
(162 MHz, CDCl₃)

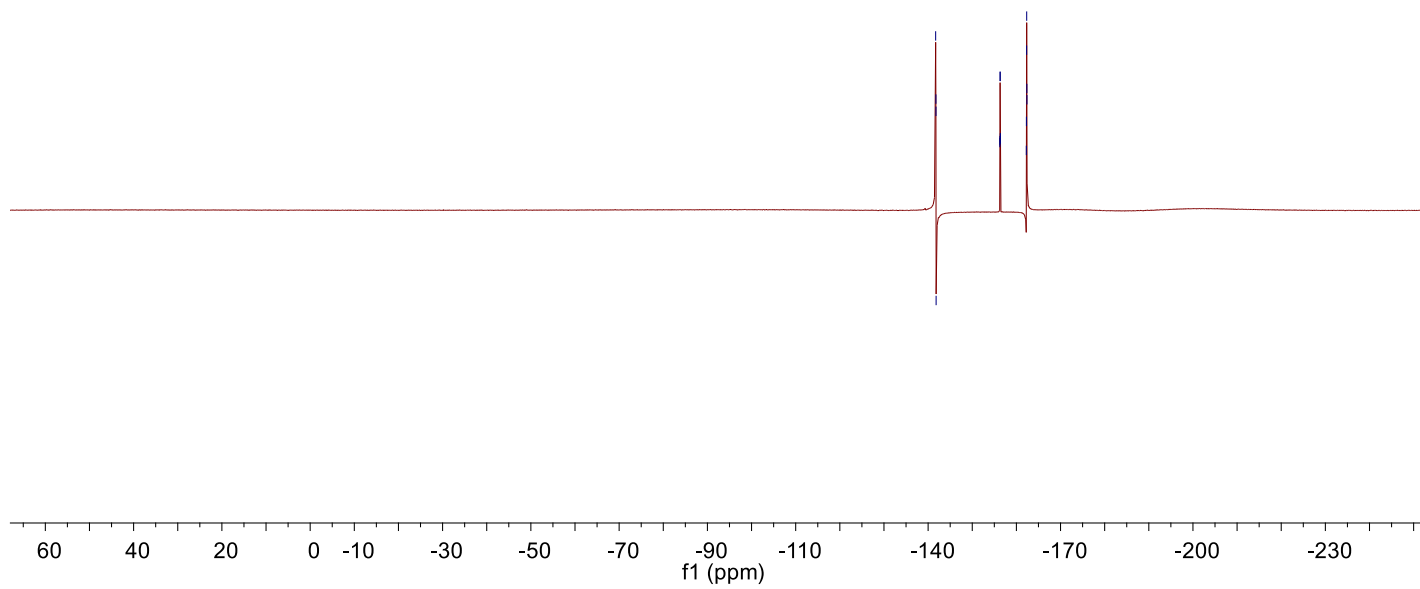


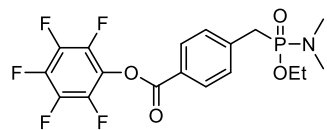
S190



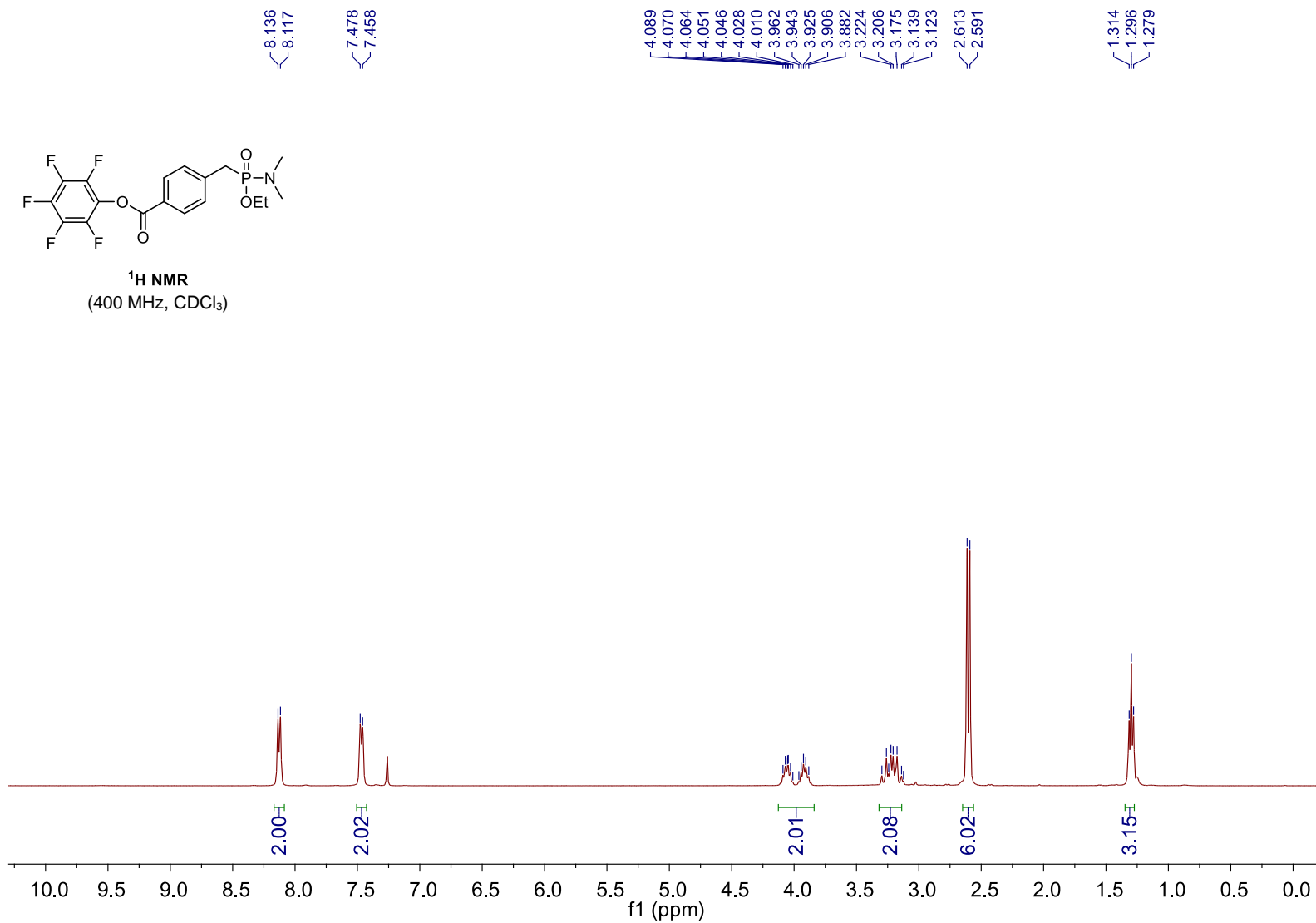
¹⁹F NMR
(376 MHz, CDCl₃)

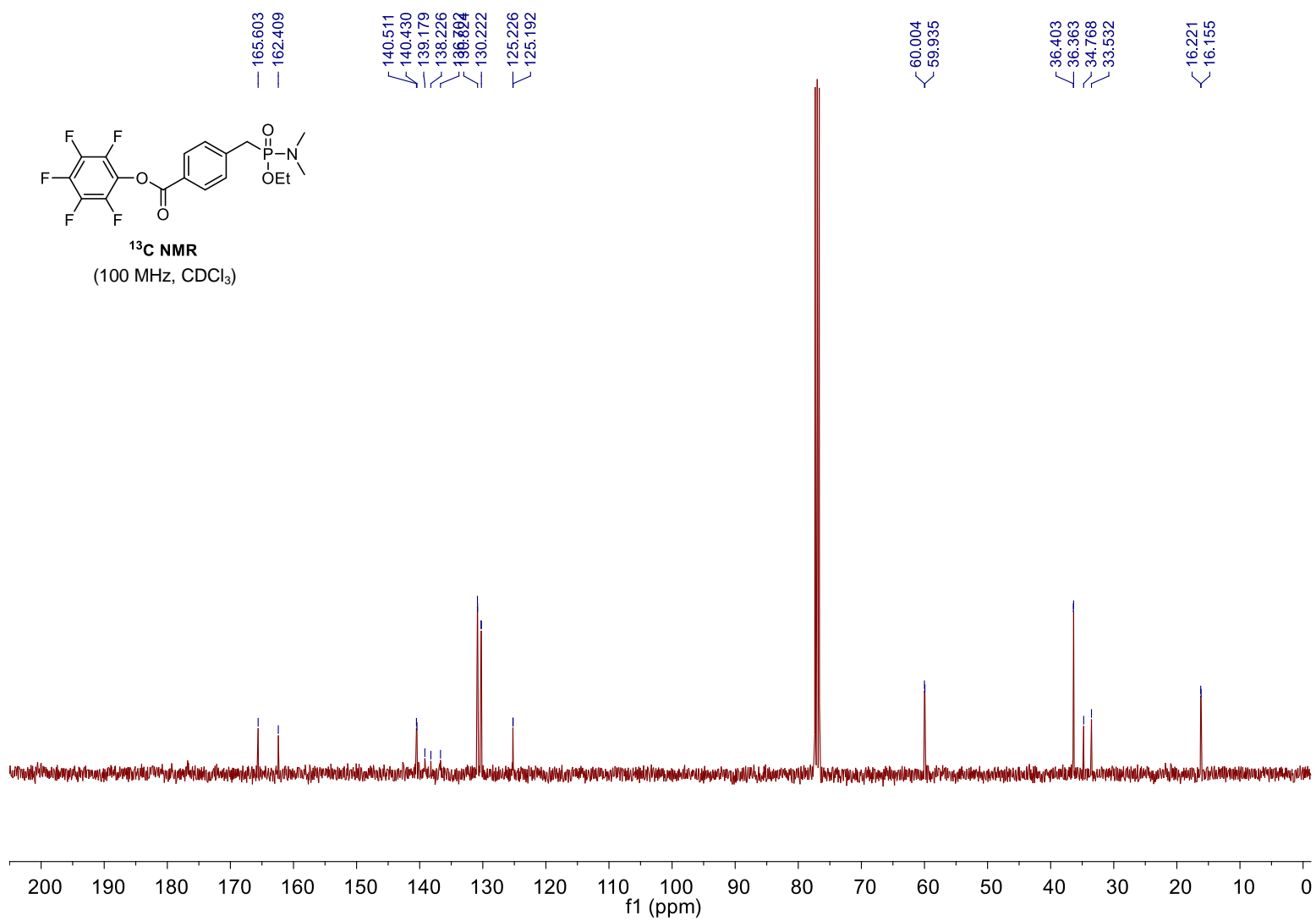
-141.708
-141.766
-141.776
-14.816
-156.312
-156.385
-162.255
-162.277
-162.313
-162.333
-162.370
-162.390

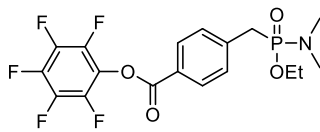




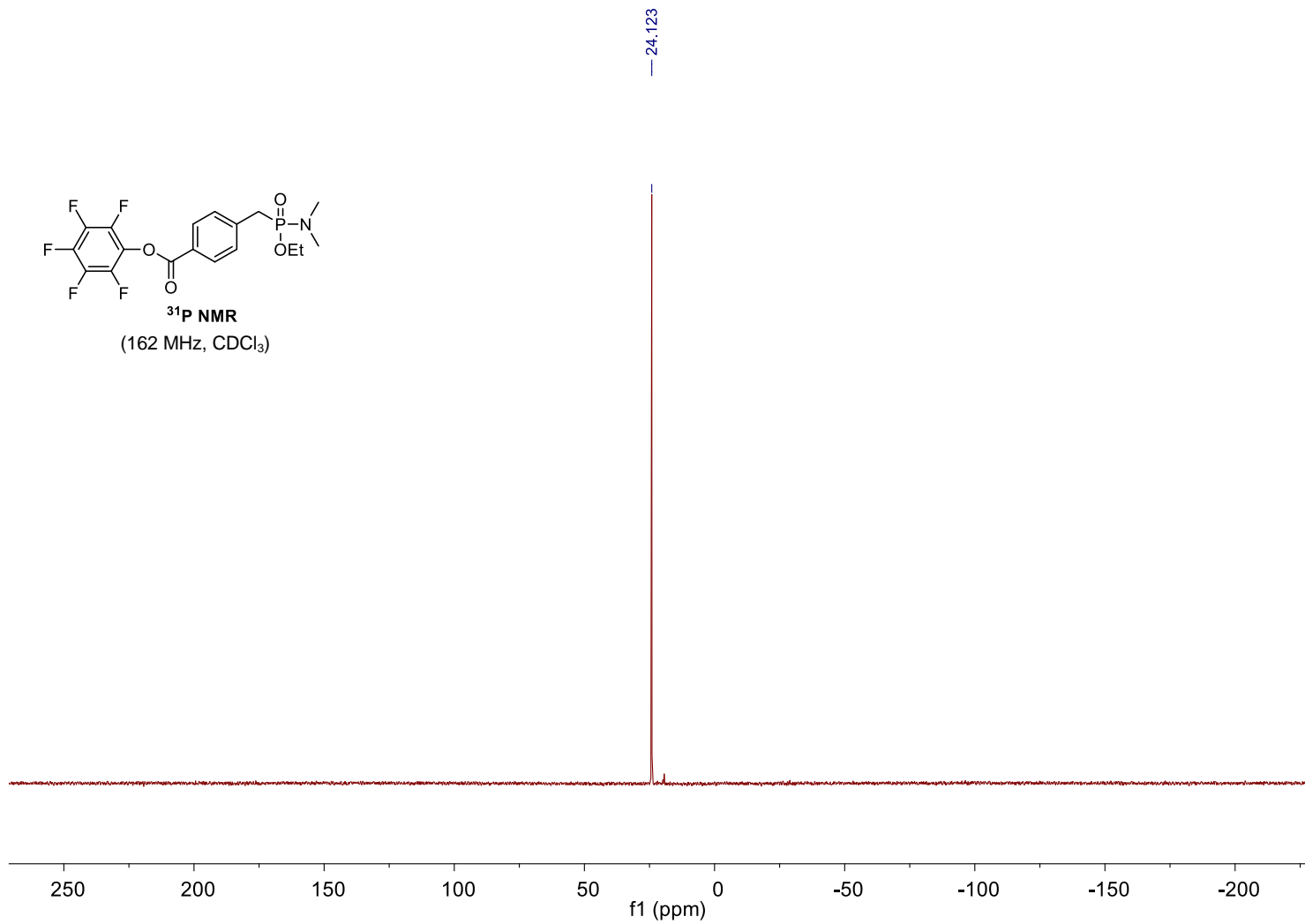
¹H NMR
(400 MHz, CDCl₃)

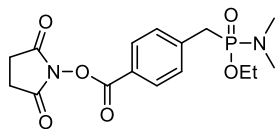




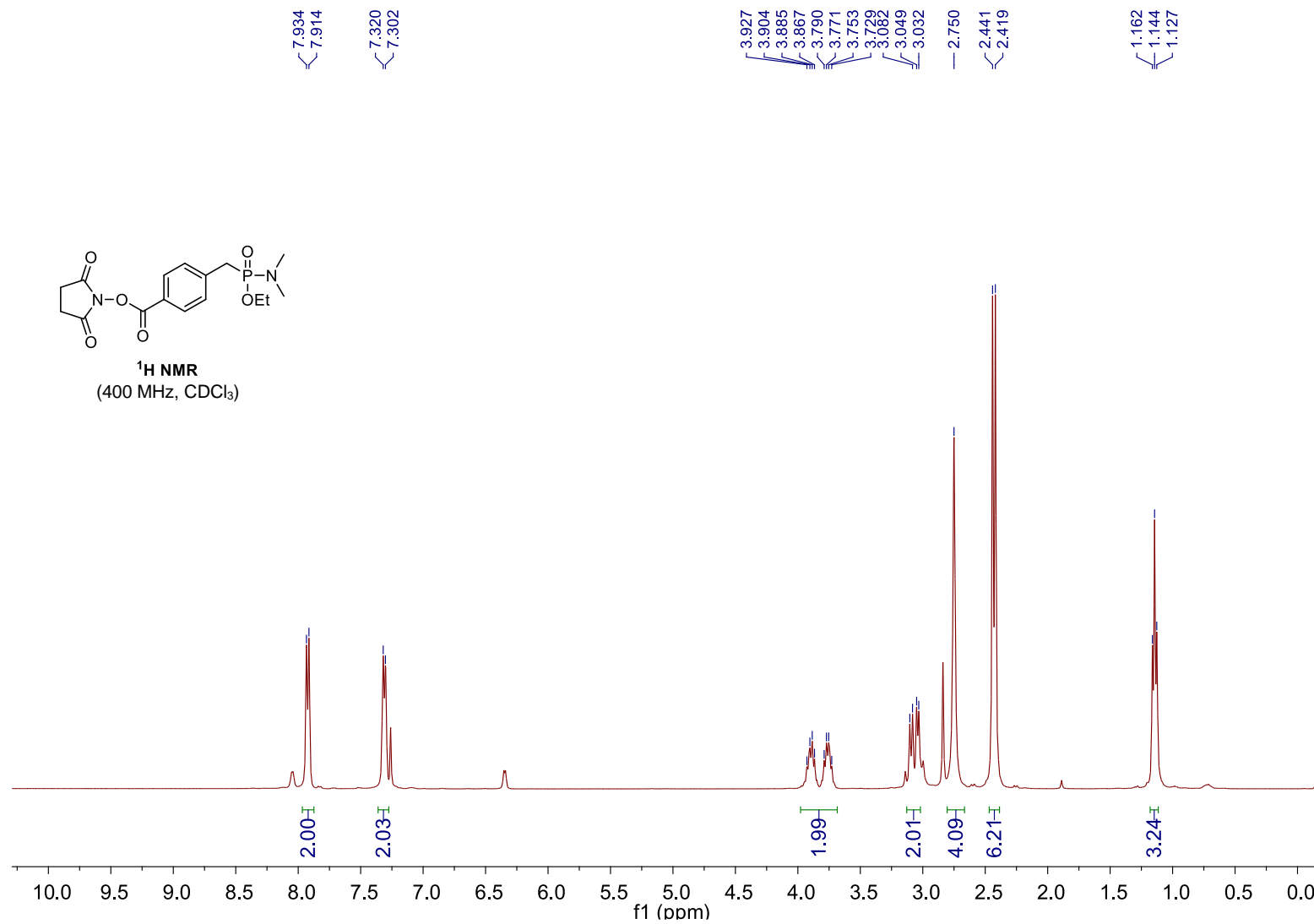


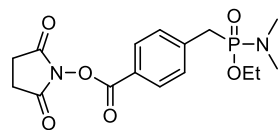
³¹P NMR
(162 MHz, CDCl₃)



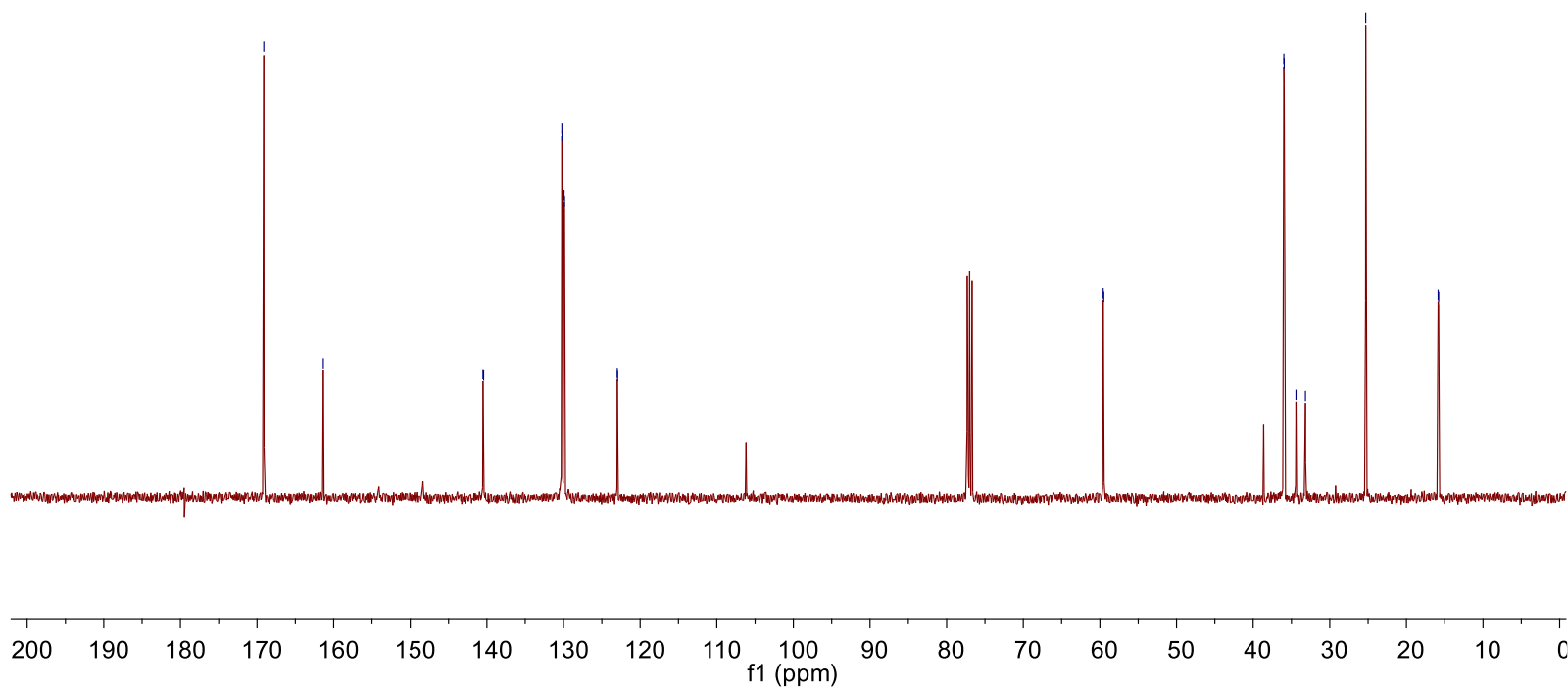


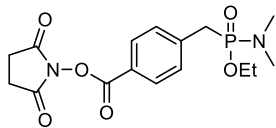
¹H NMR
(400 MHz, CDCl₃)



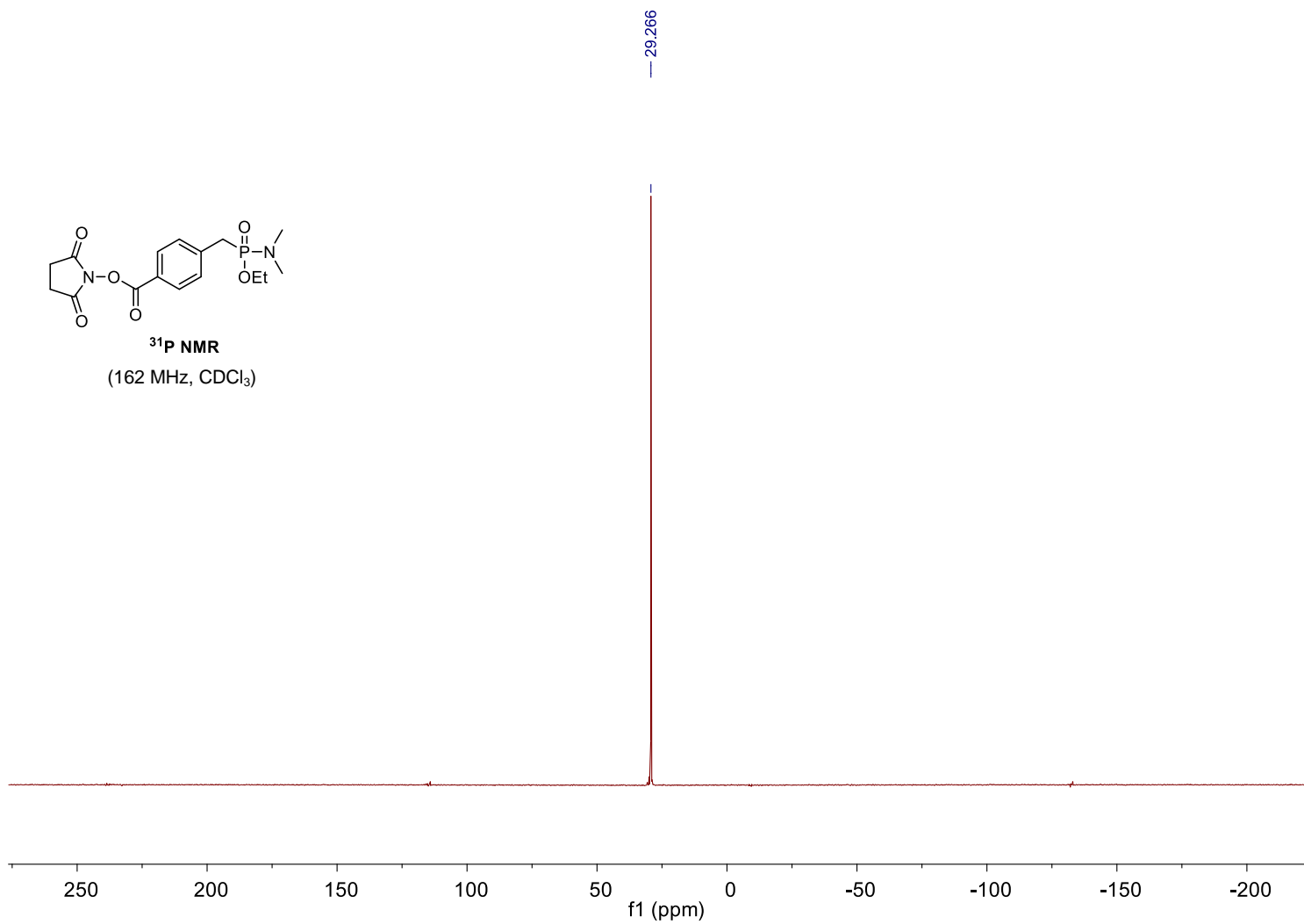


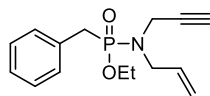
¹³C NMR
(100 MHz, CDCl₃)



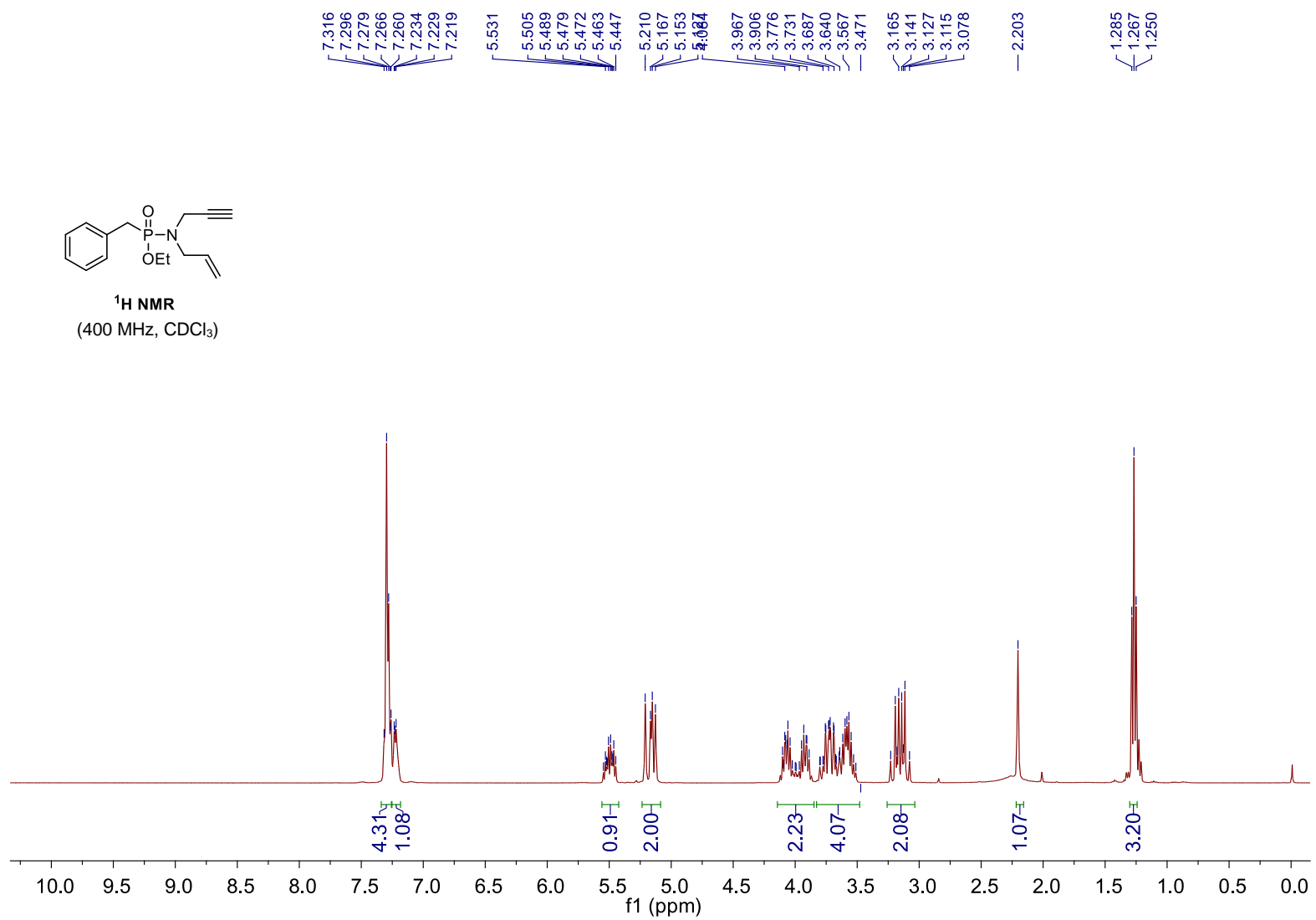


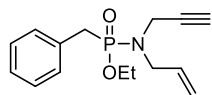
³¹P NMR
(162 MHz, CDCl₃)



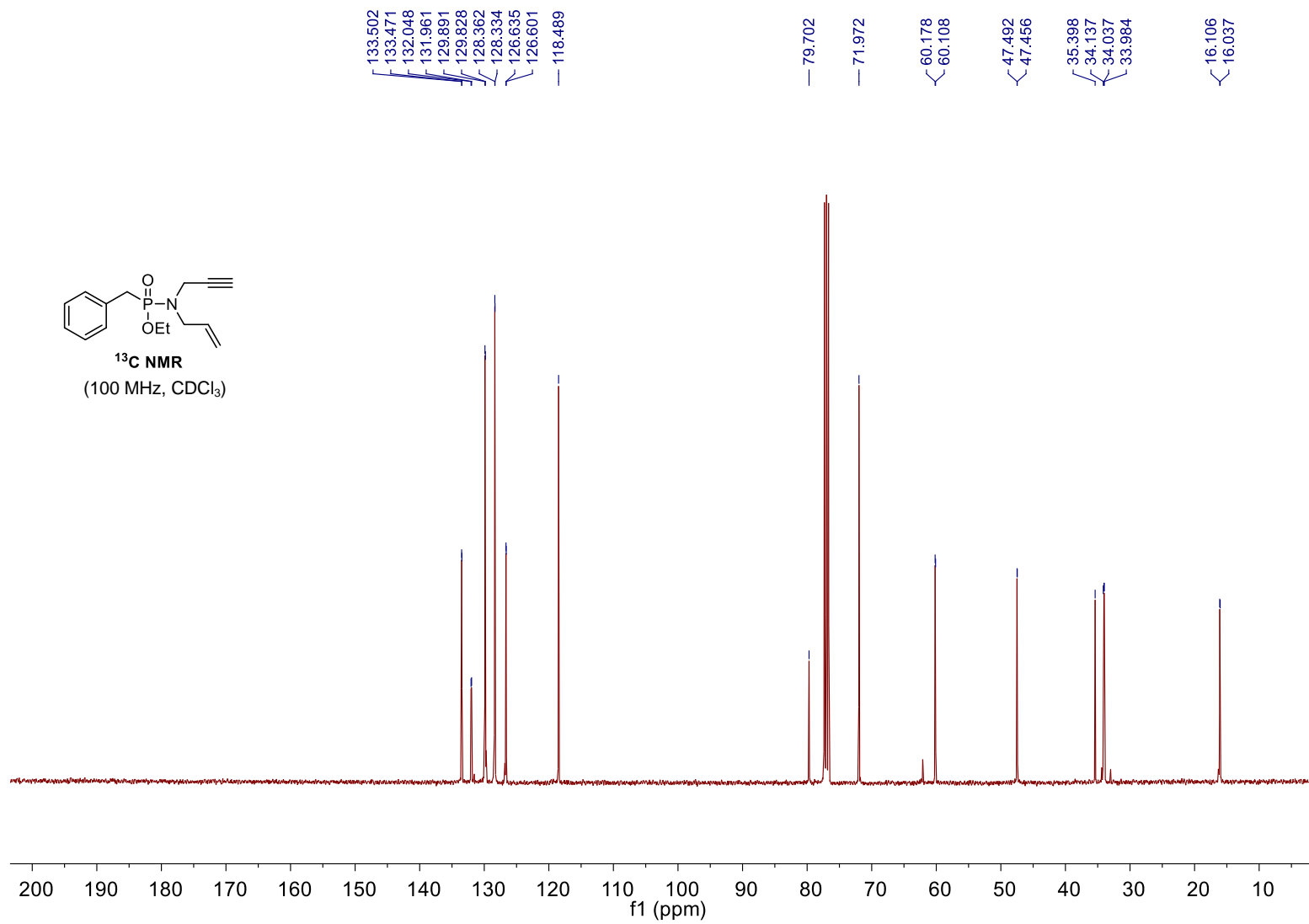


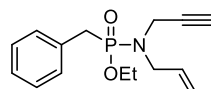
¹H NMR
(400 MHz, CDCl₃)



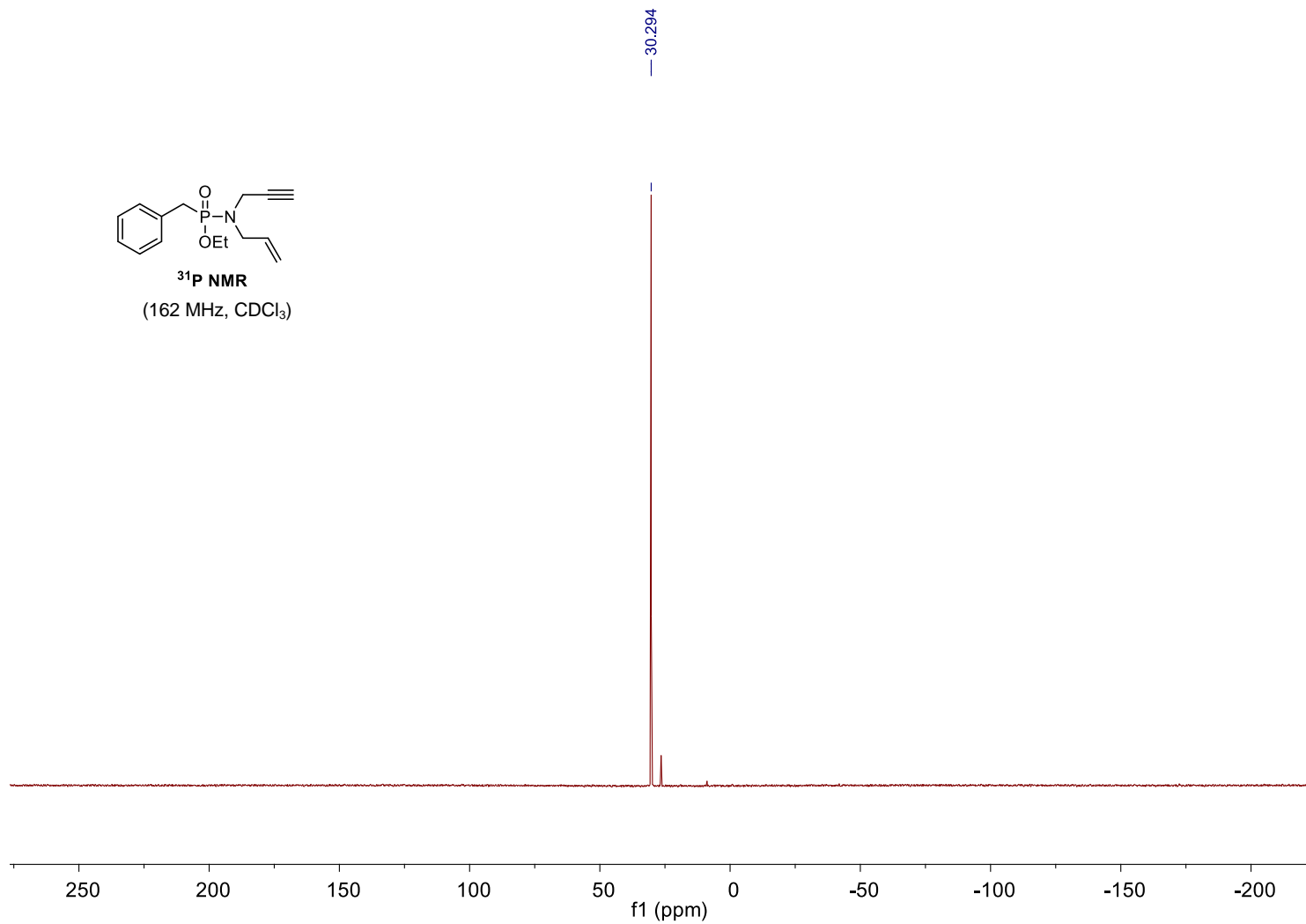


¹³C NMR
(100 MHz, CDCl₃)

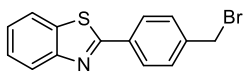




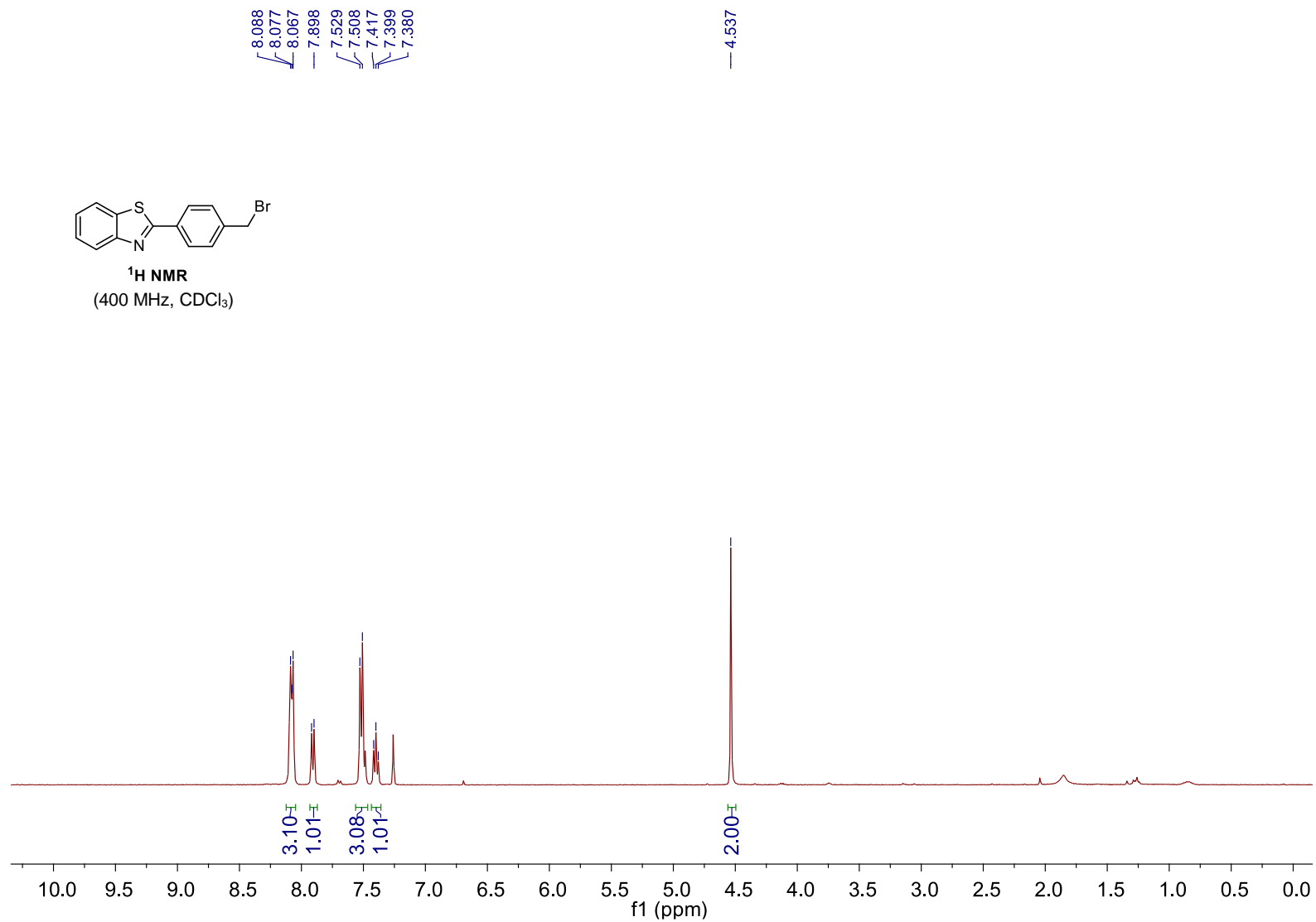
³¹P NMR
(162 MHz, CDCl₃)

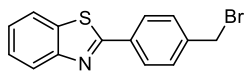


S200

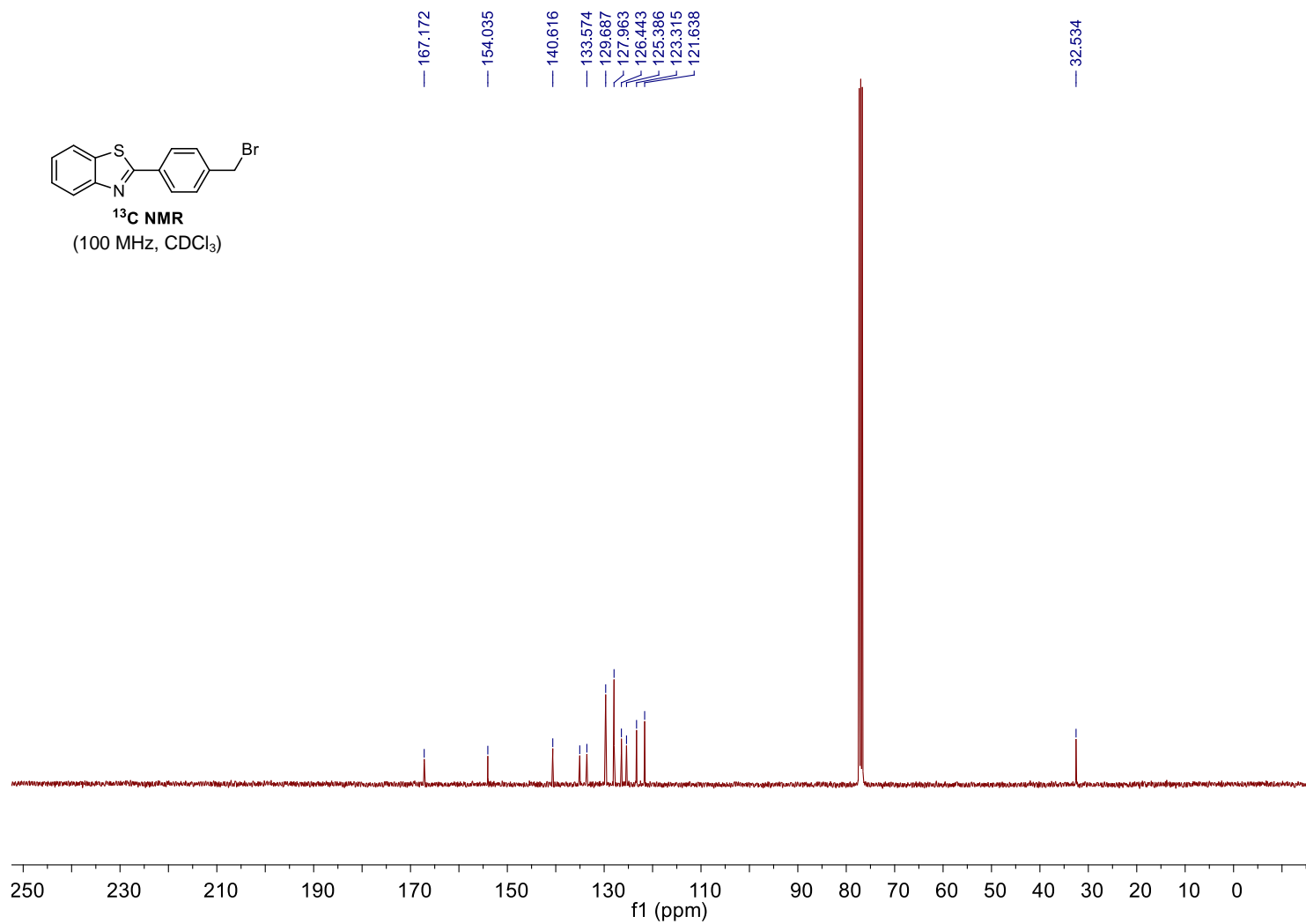


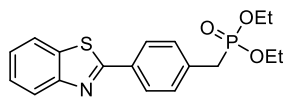
¹H NMR
(400 MHz, CDCl₃)



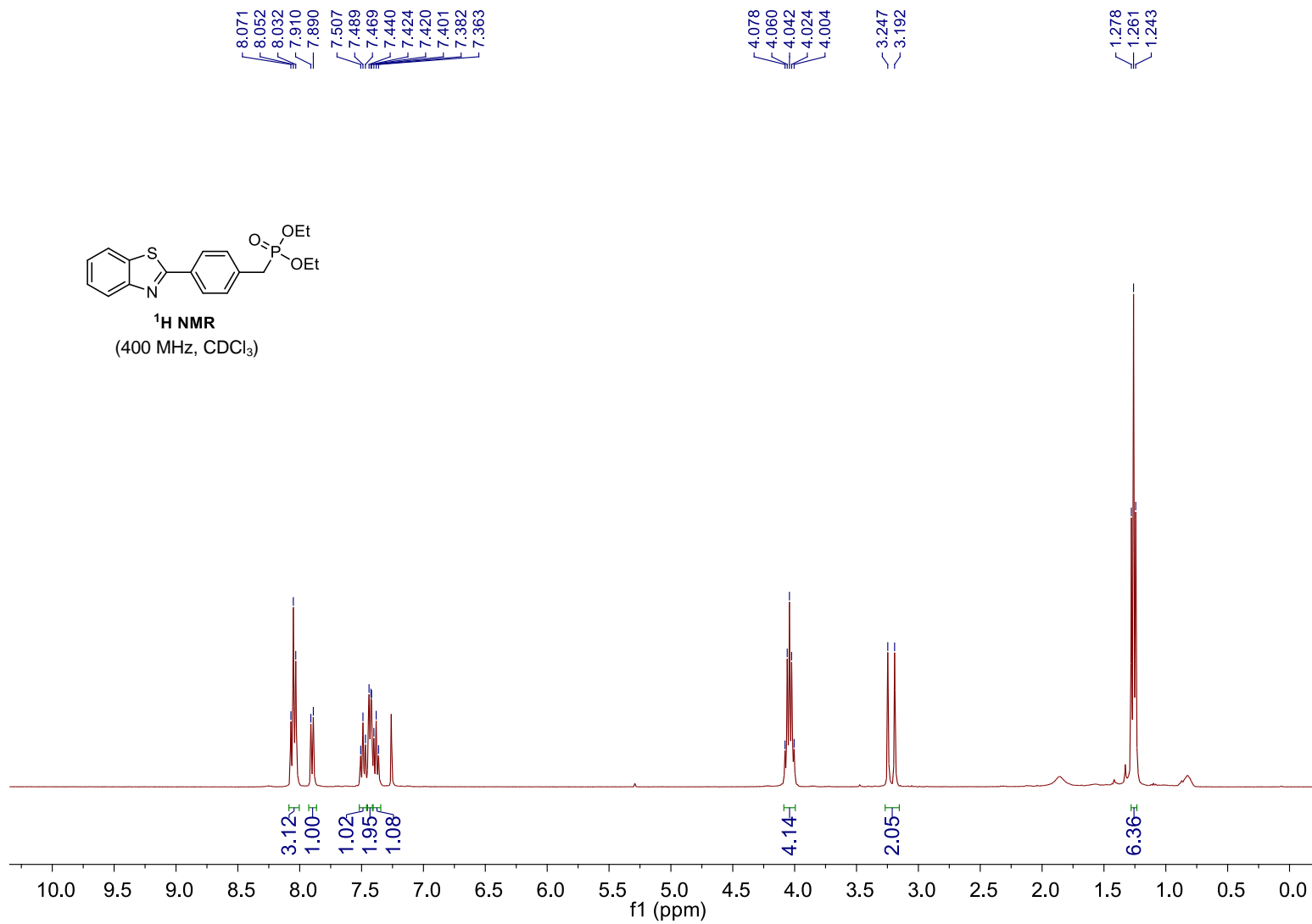


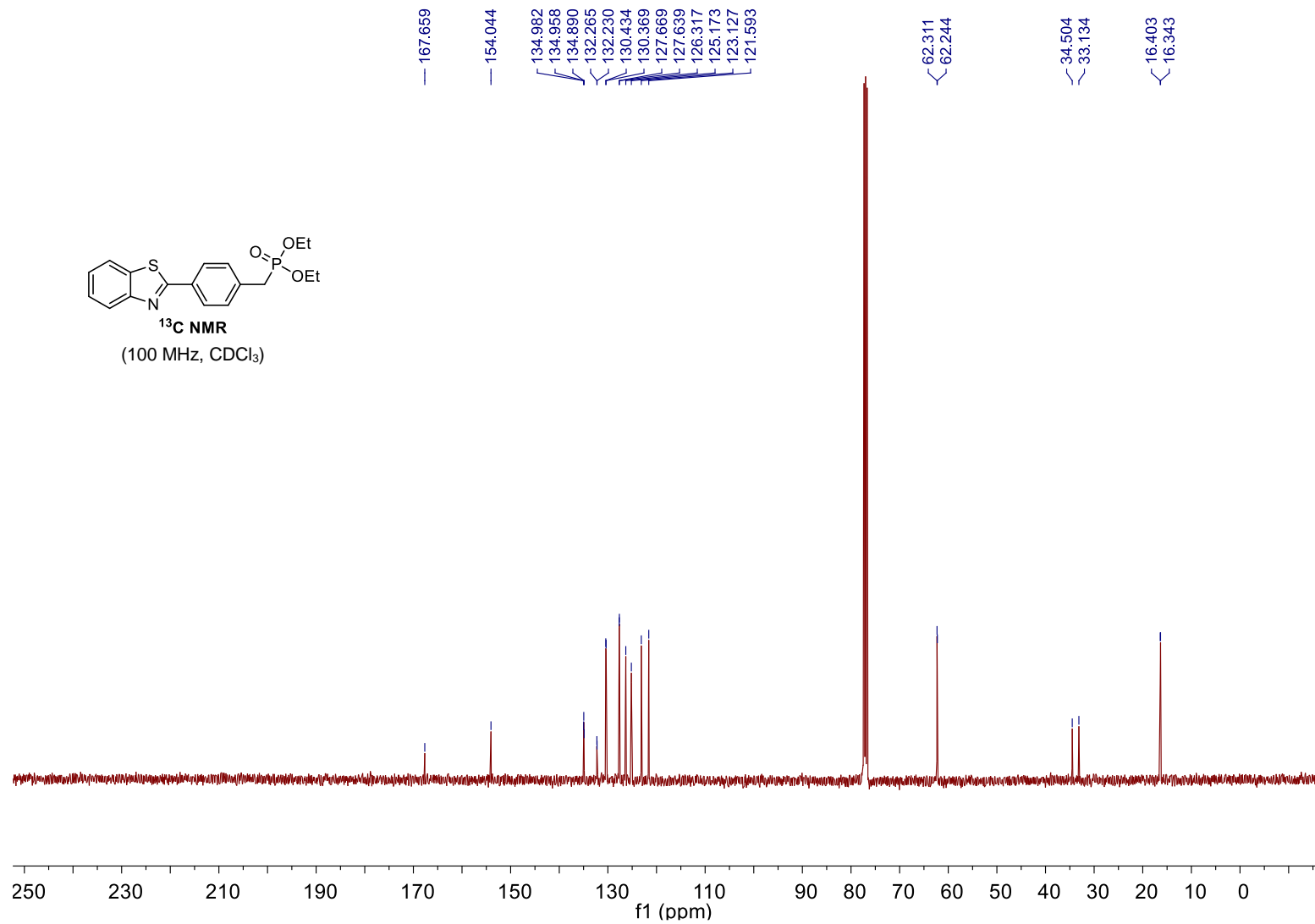
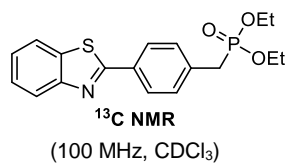
¹³C NMR
(100 MHz, CDCl₃)

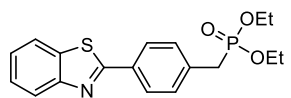




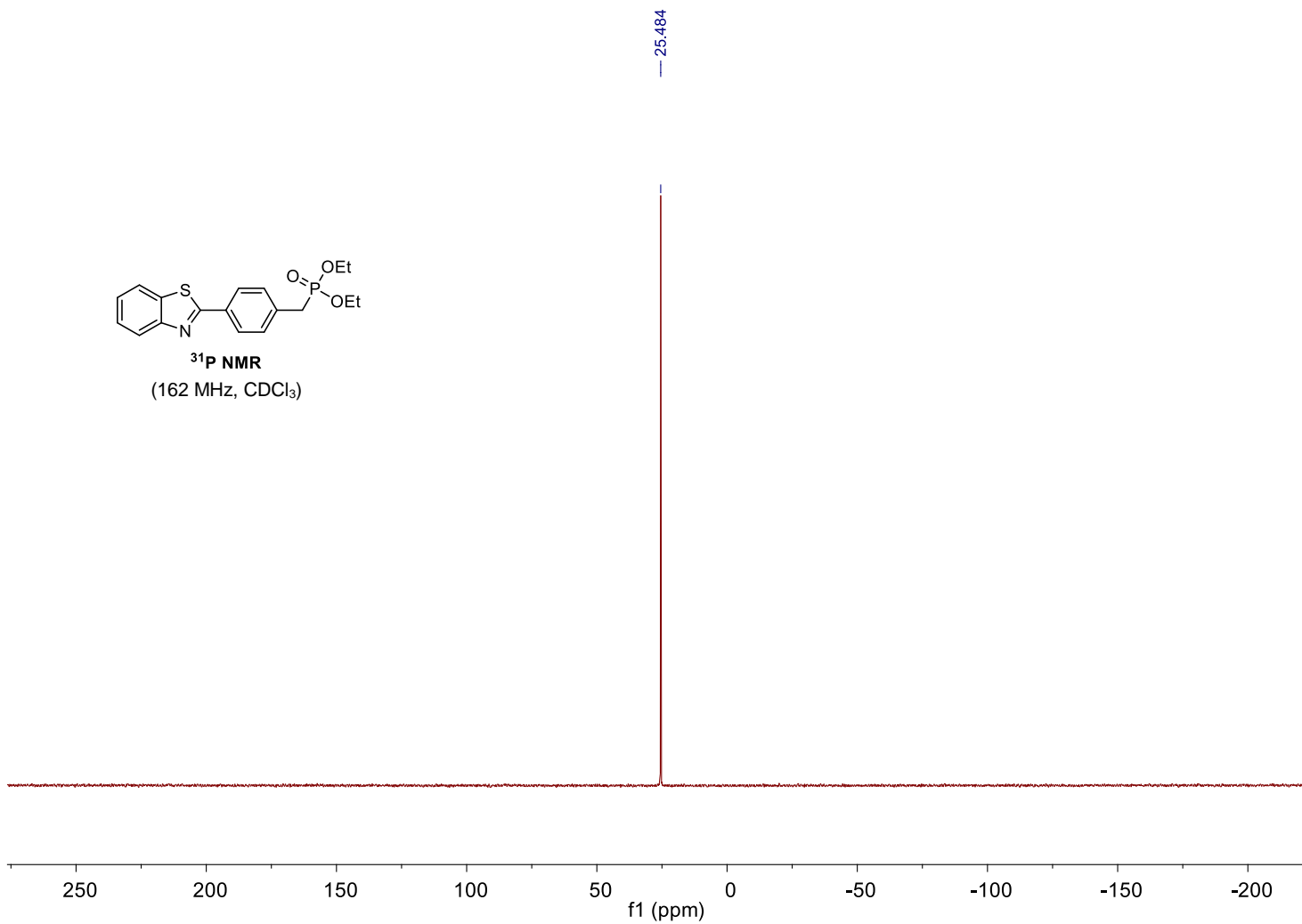
¹H NMR
(400 MHz, CDCl₃)



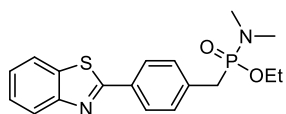




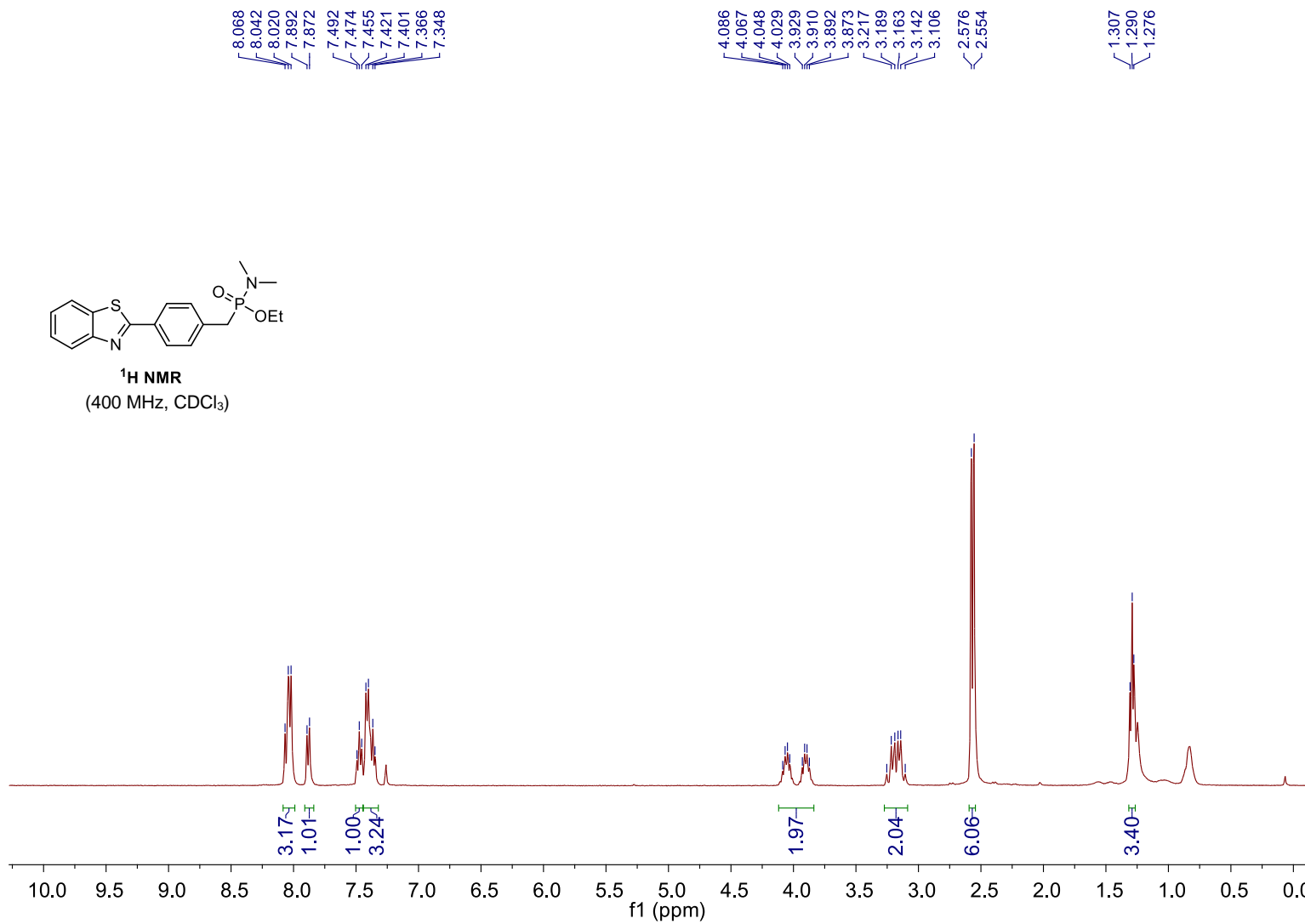
³¹P NMR
(162 MHz, CDCl₃)

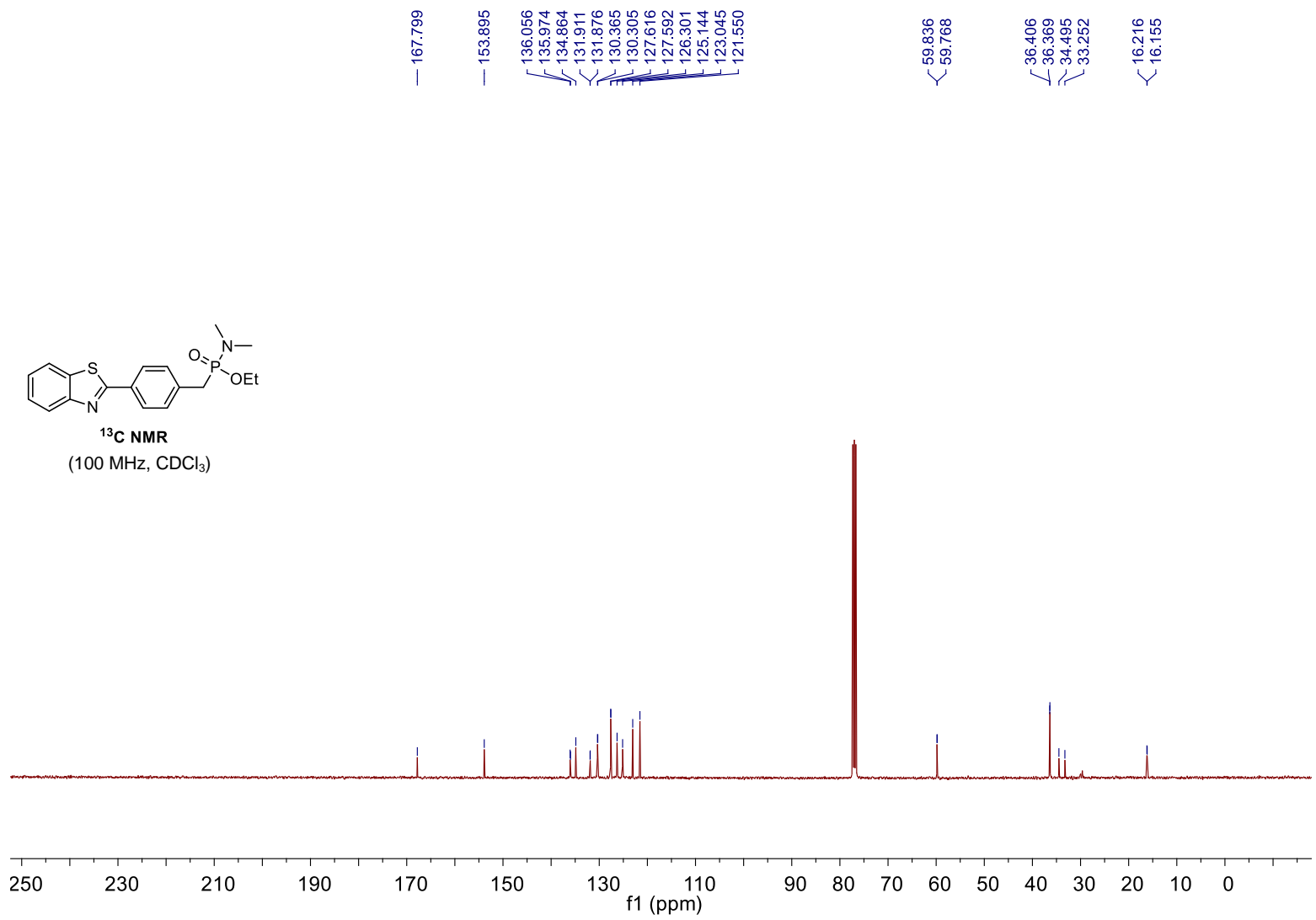


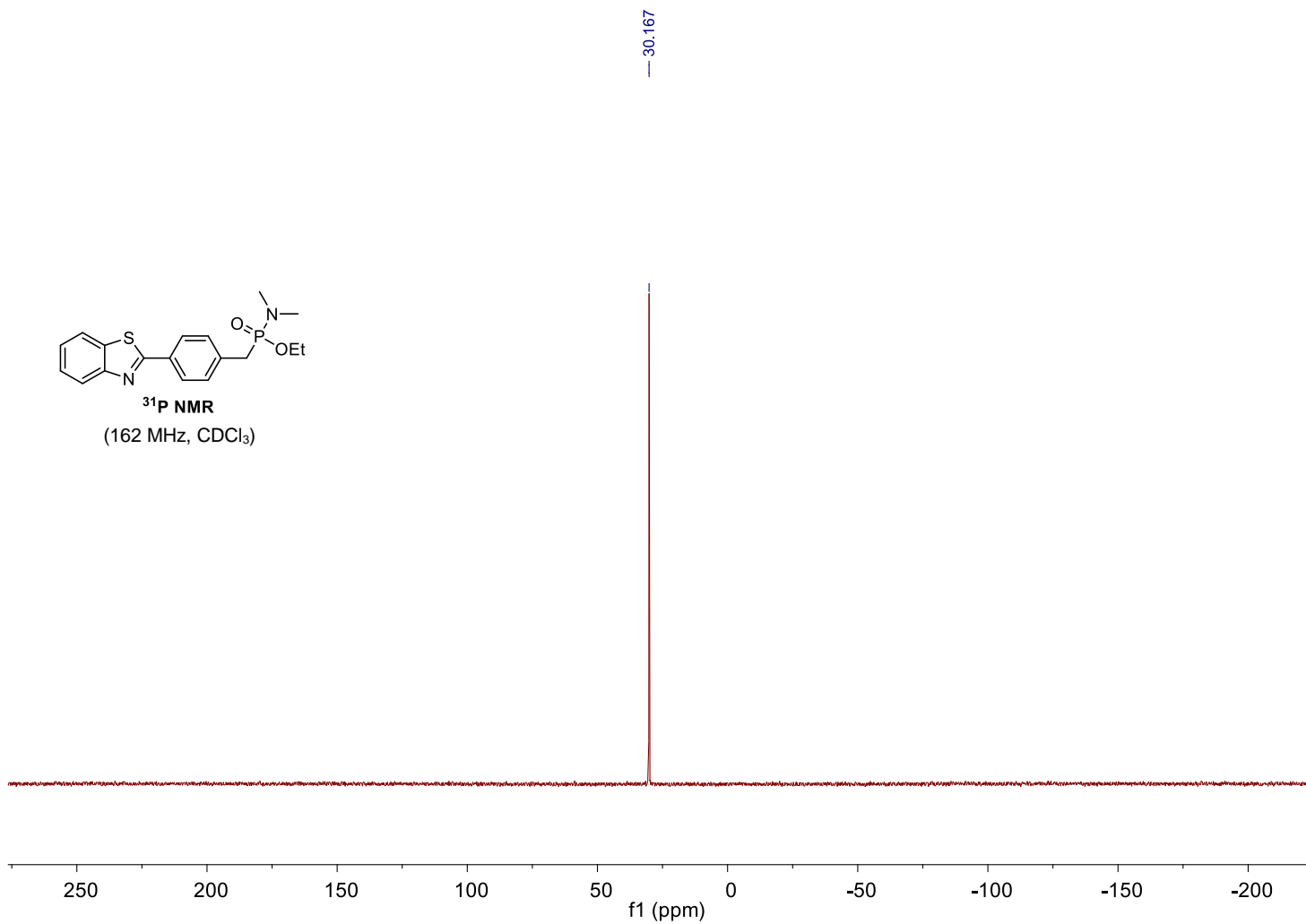
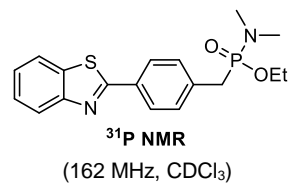
S205

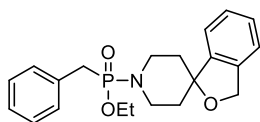


¹H NMR
(400 MHz, CDCl₃)

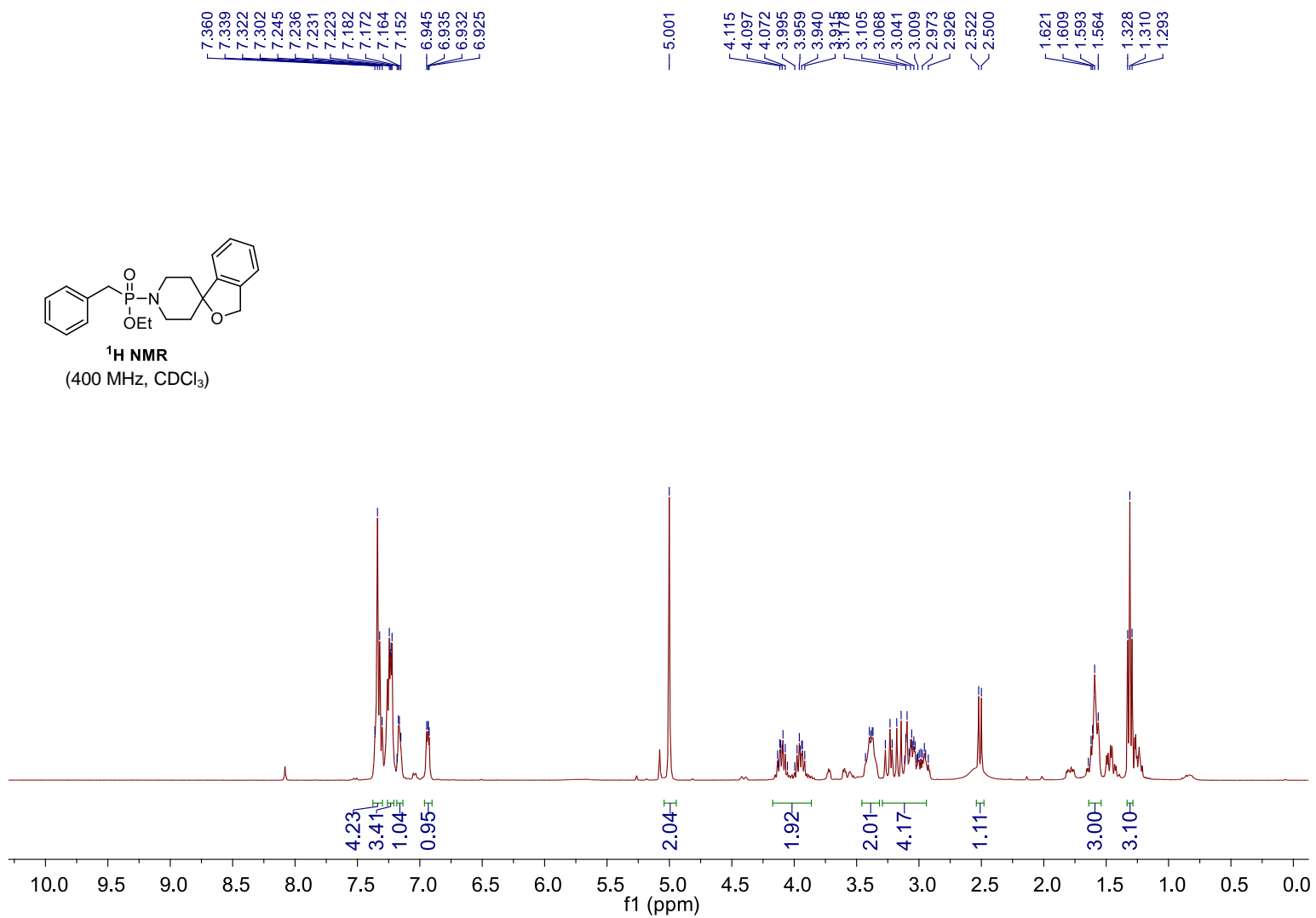


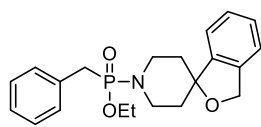




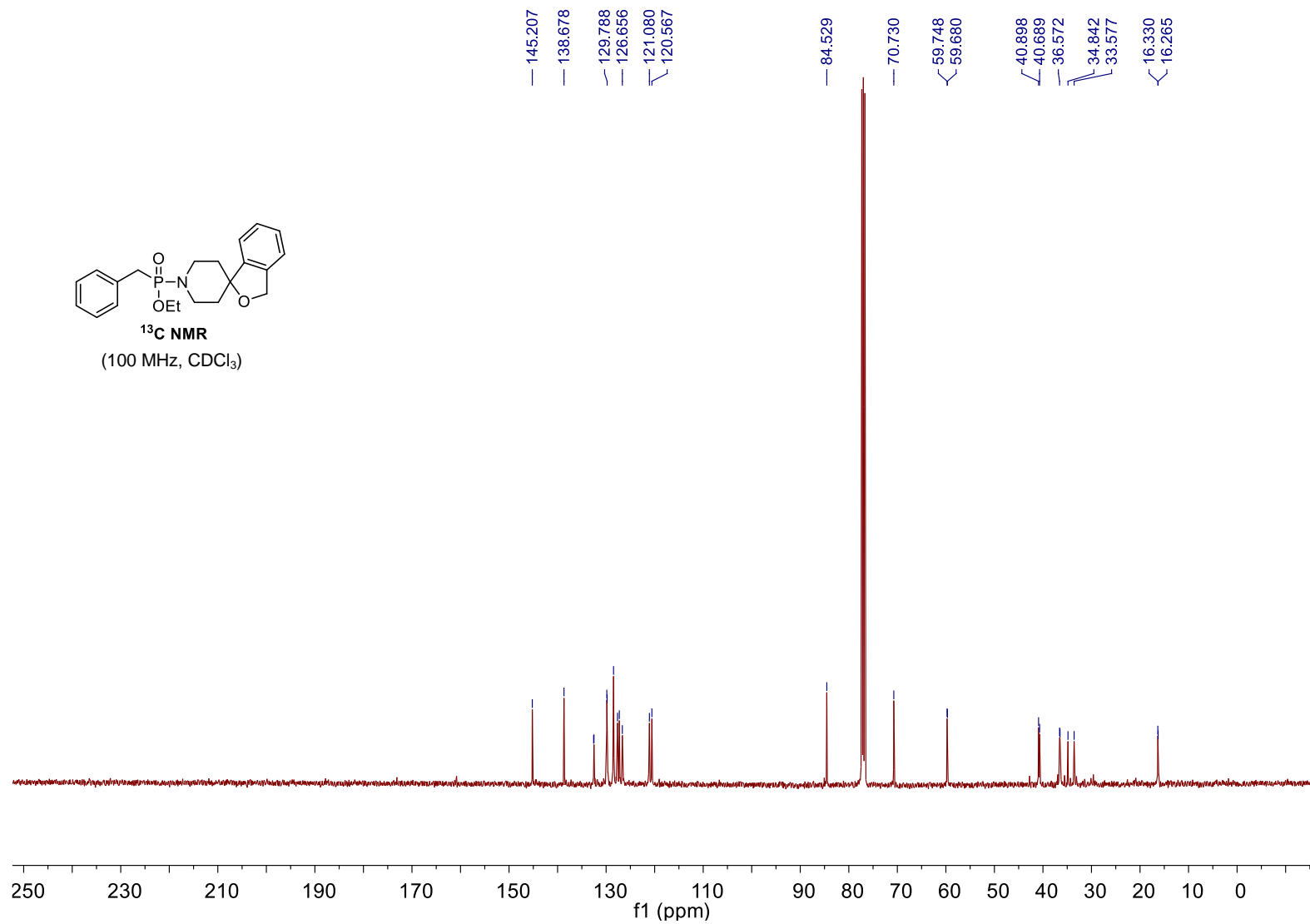


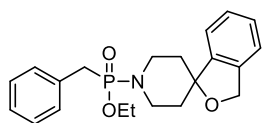
¹H NMR
(400 MHz, CDCl₃)



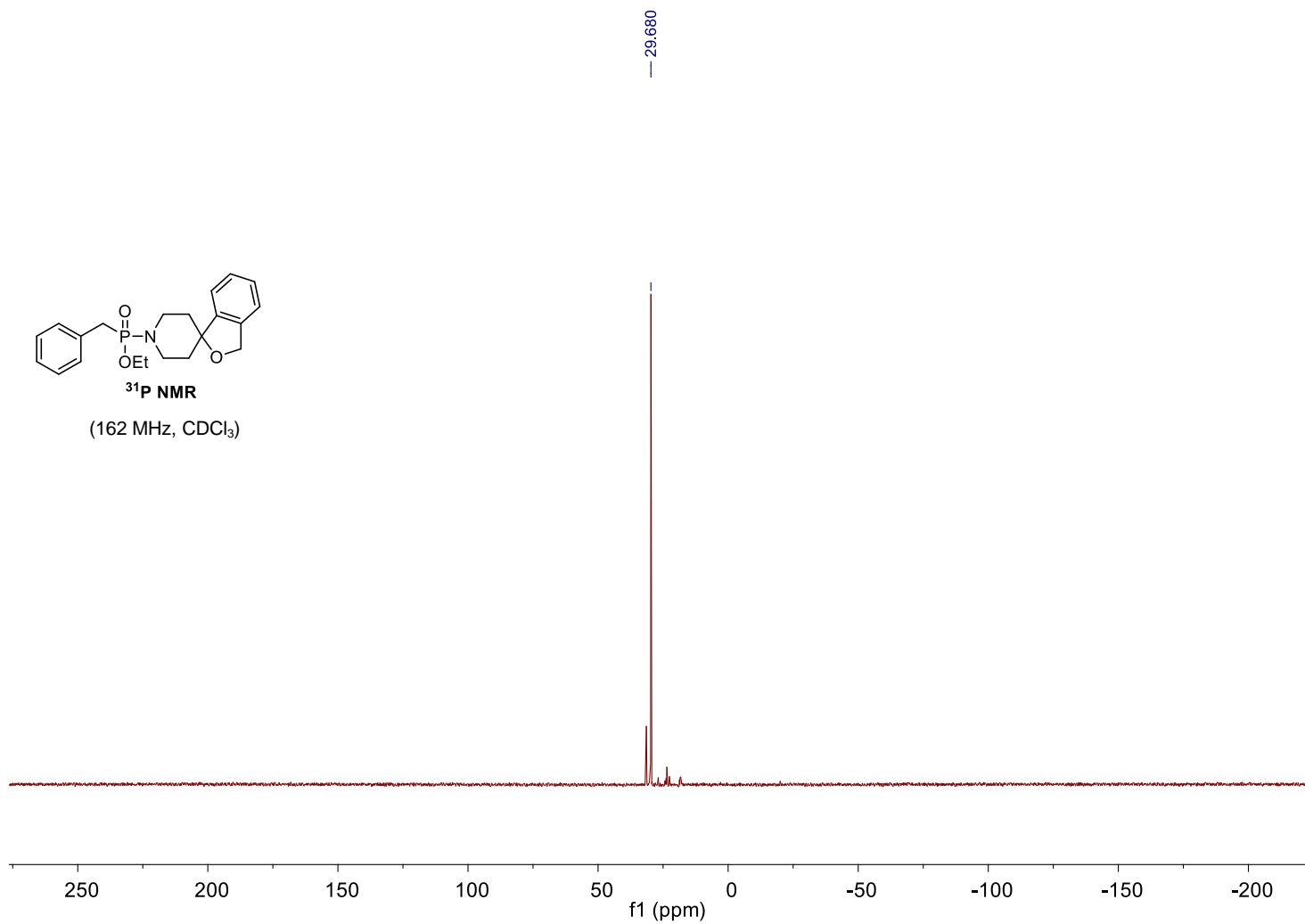


¹³C NMR
(100 MHz, CDCl₃)

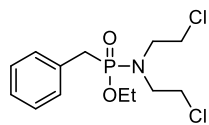




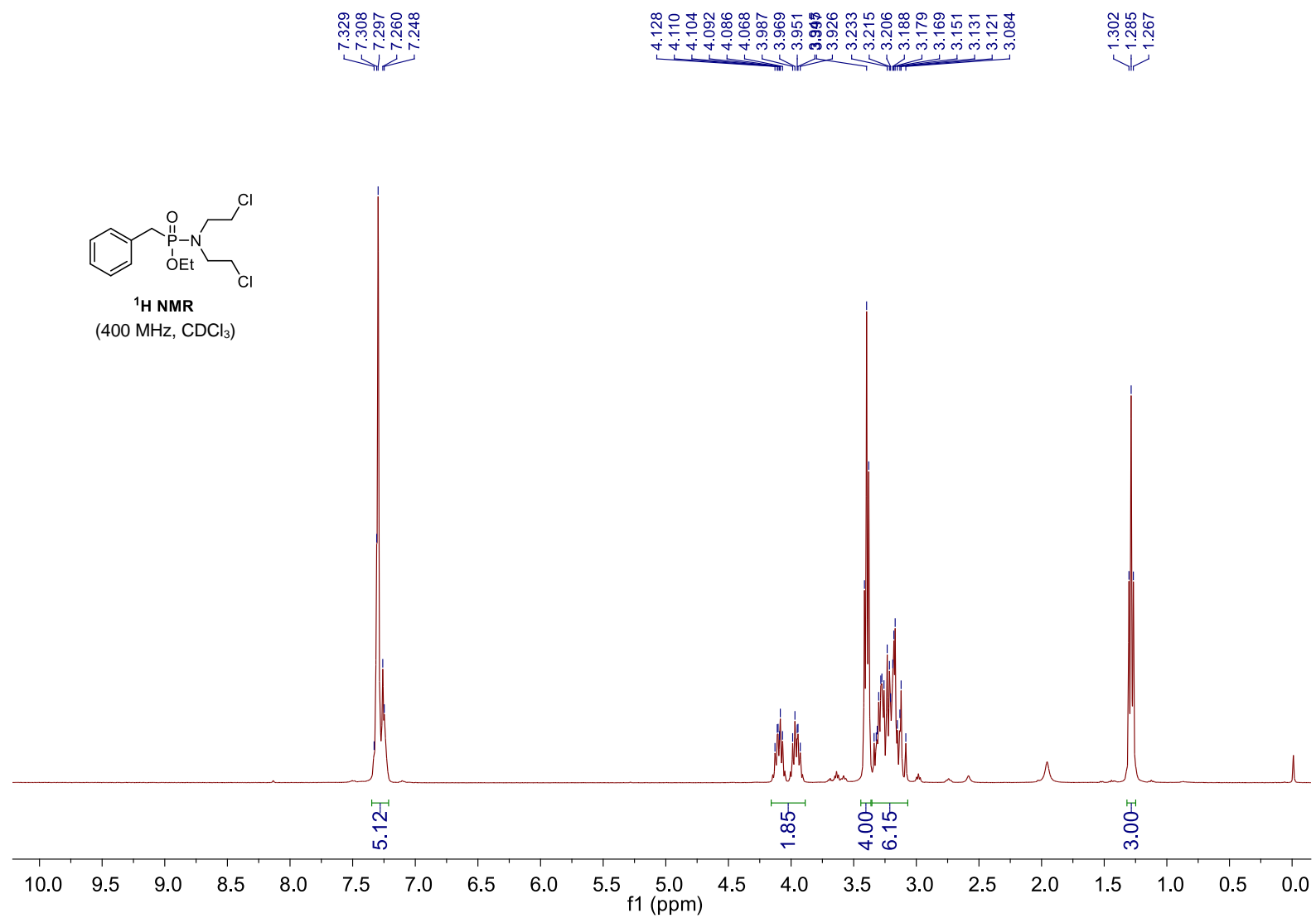
³¹P NMR
(162 MHz, CDCl₃)

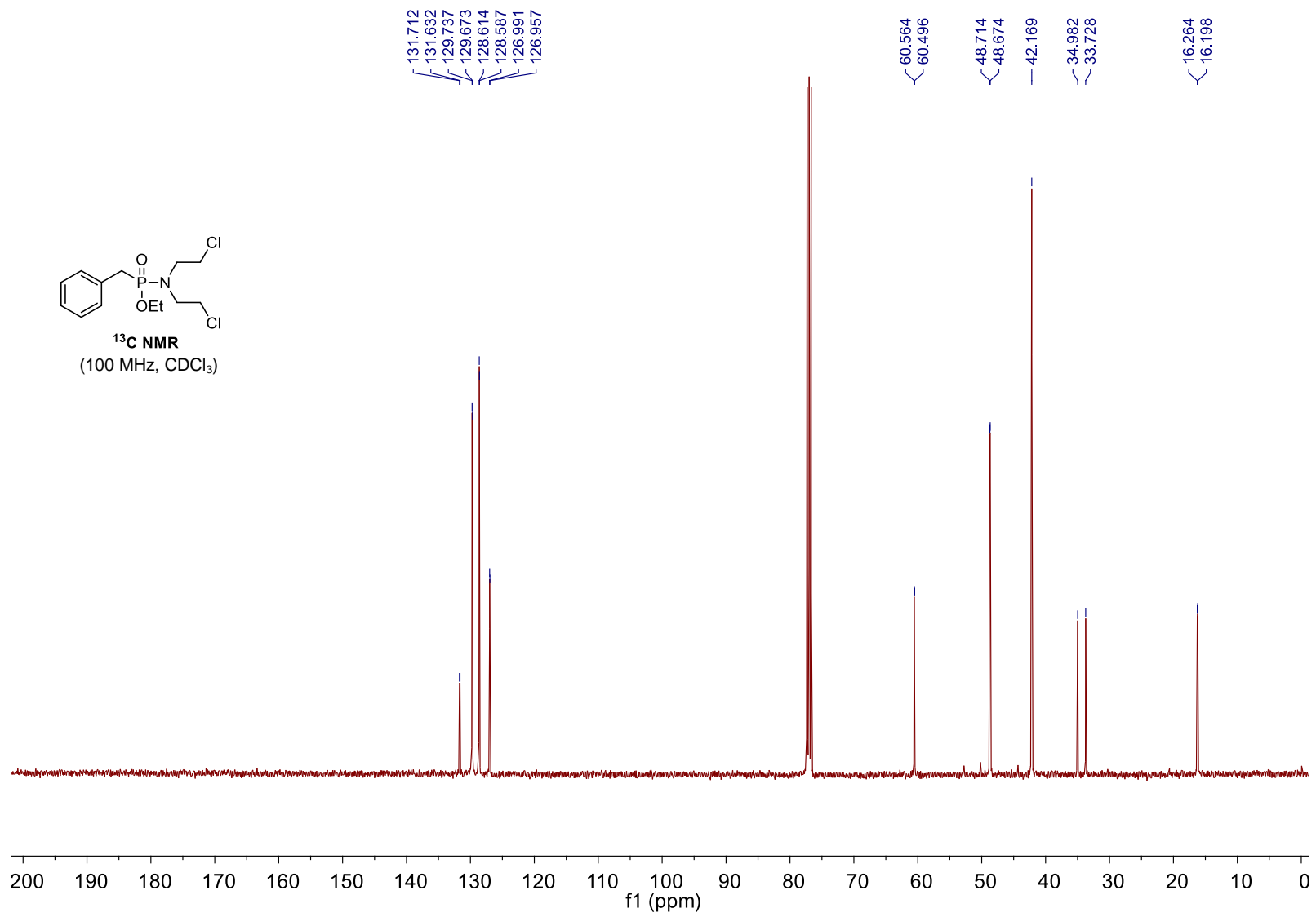
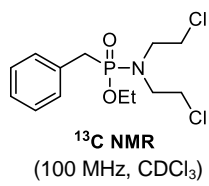


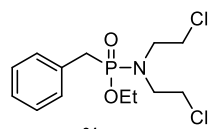
S211



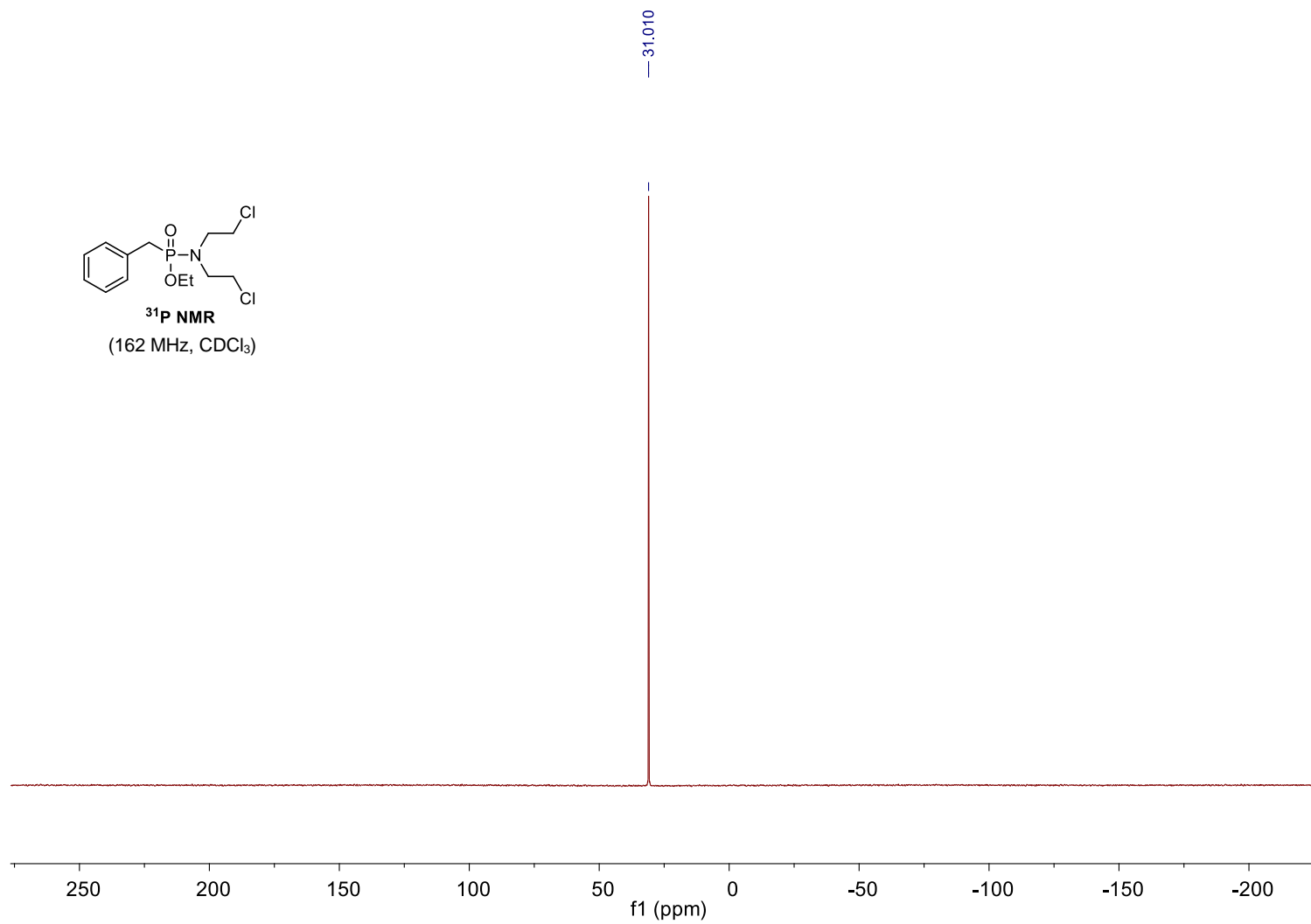
¹H NMR
(400 MHz, CDCl₃)

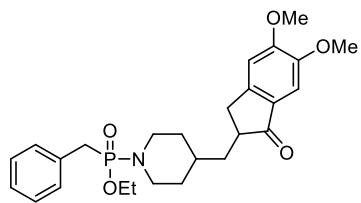




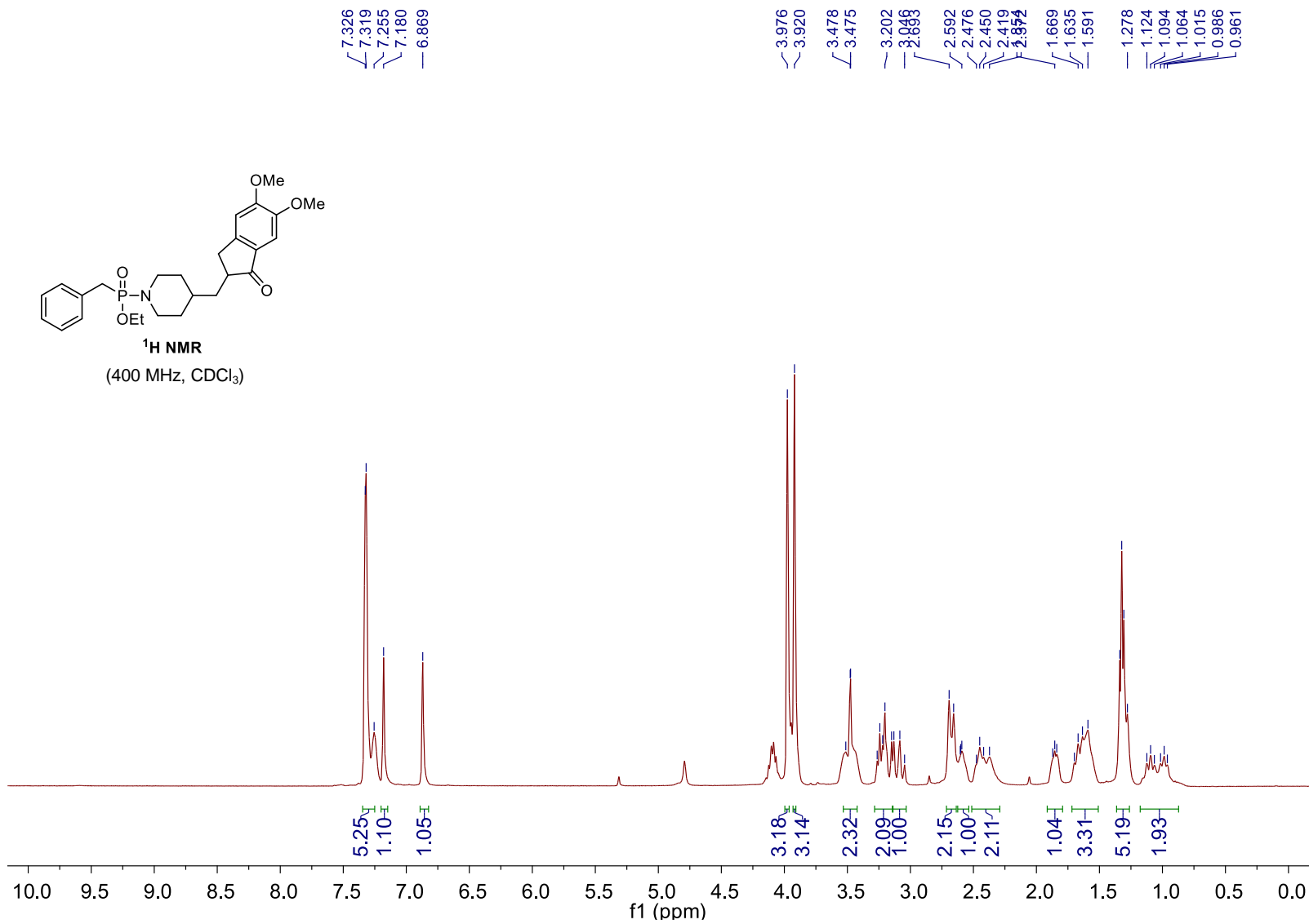


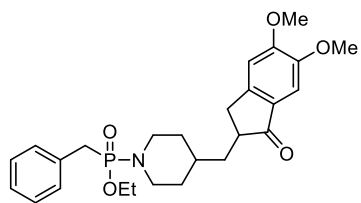
³¹P NMR
(162 MHz, CDCl₃)



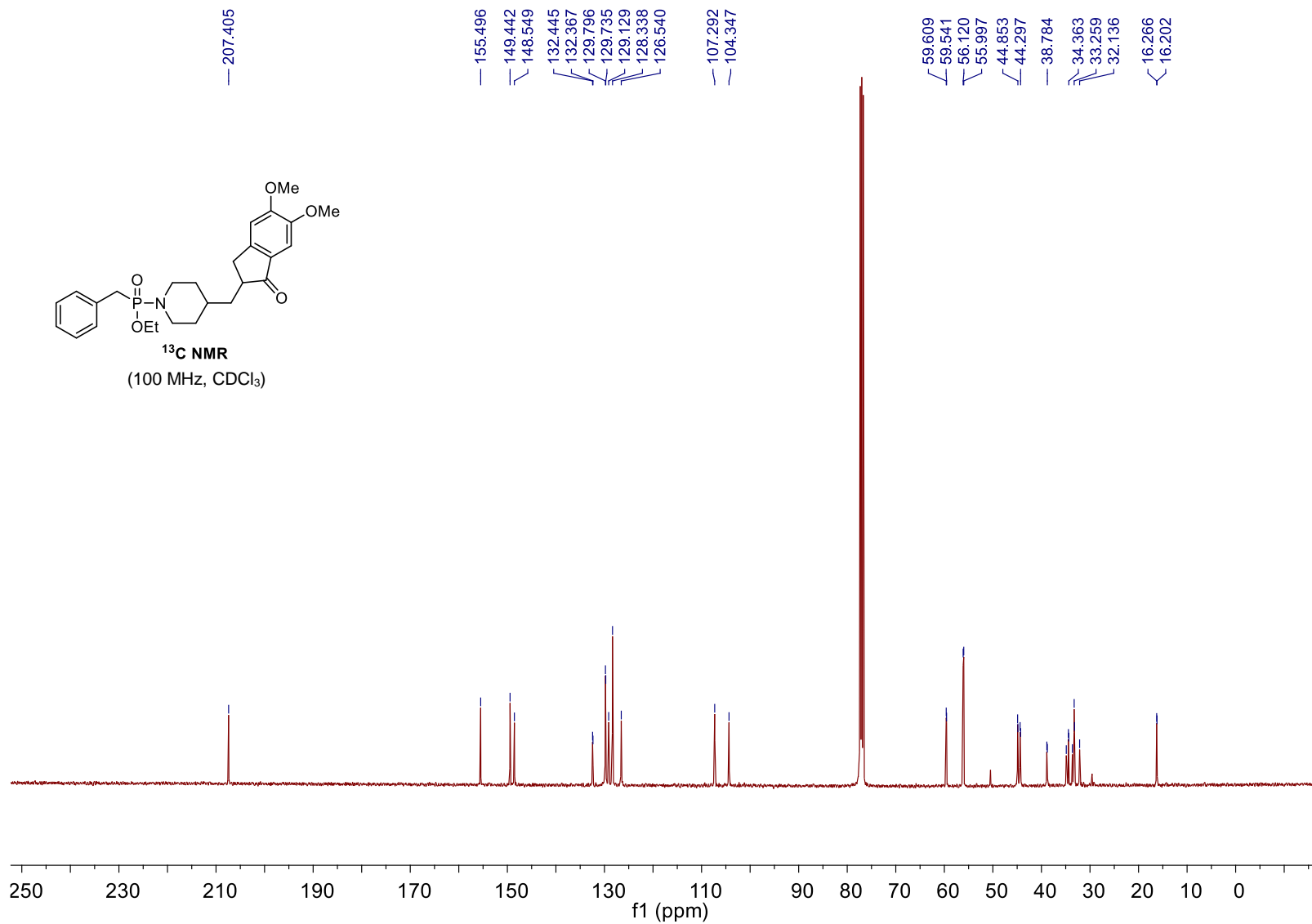


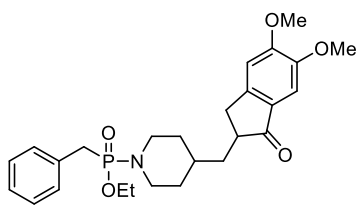
¹H NMR
(400 MHz, CDCl₃)



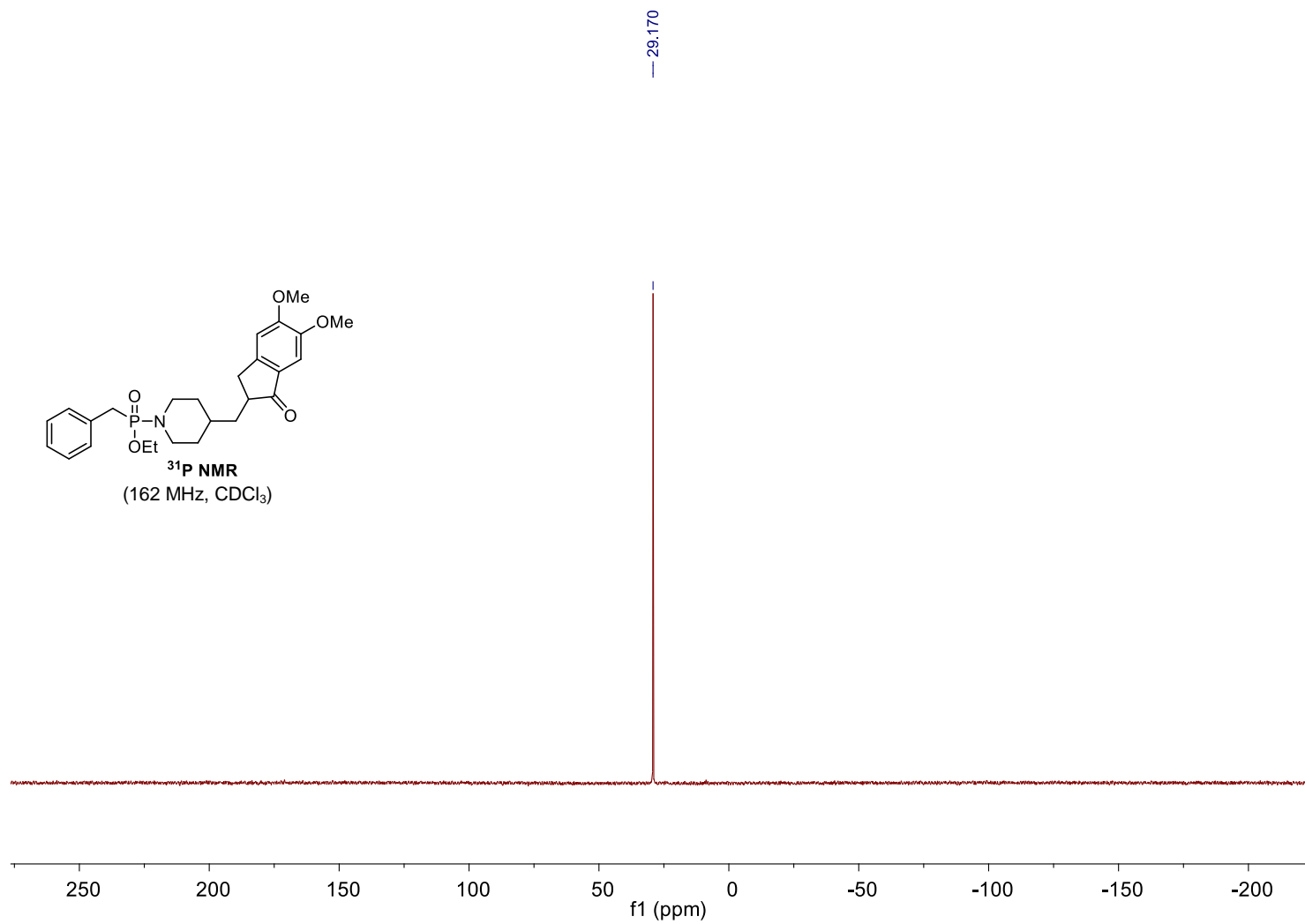


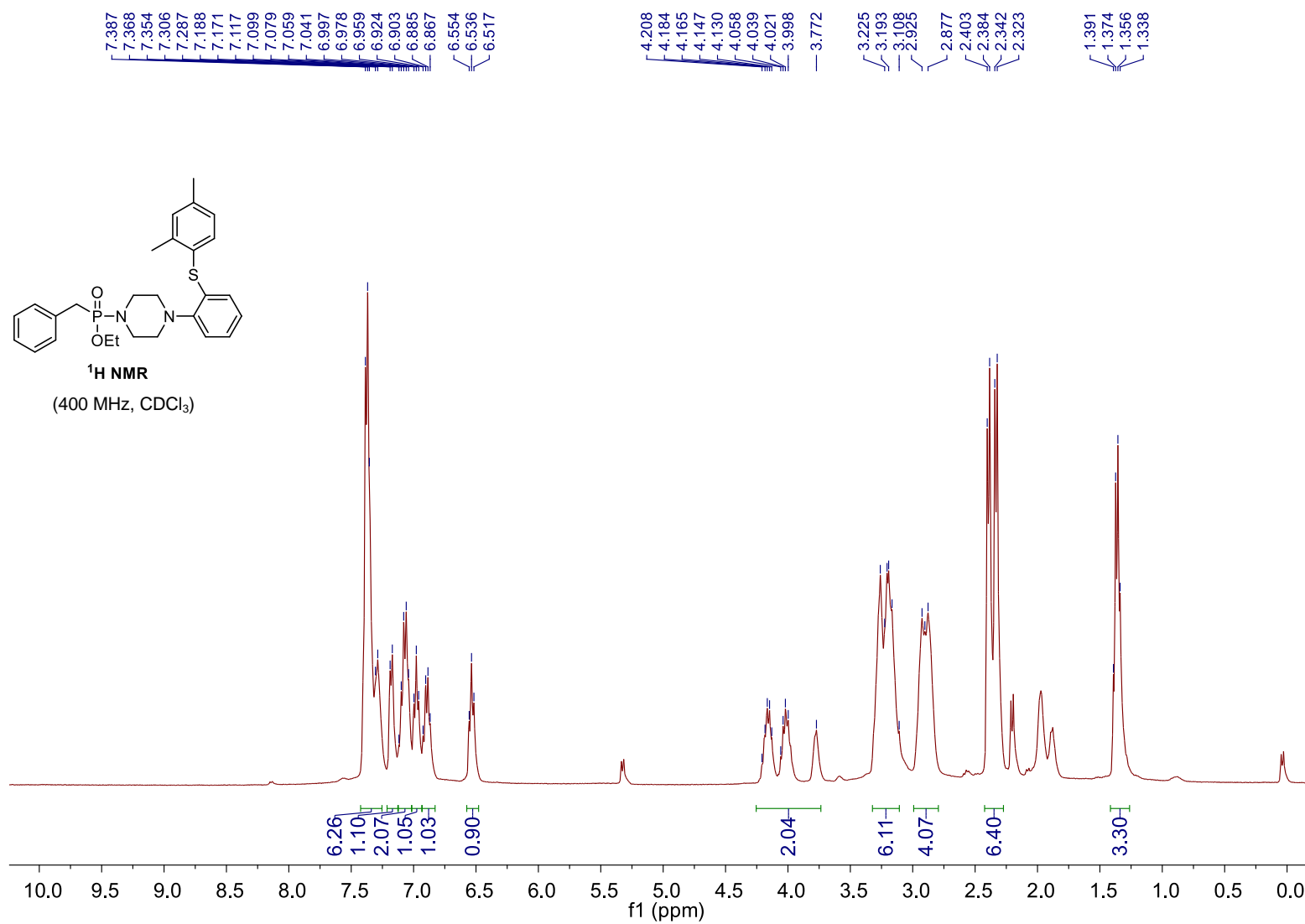
¹³C NMR
(100 MHz, CDCl₃)

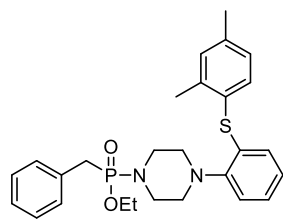




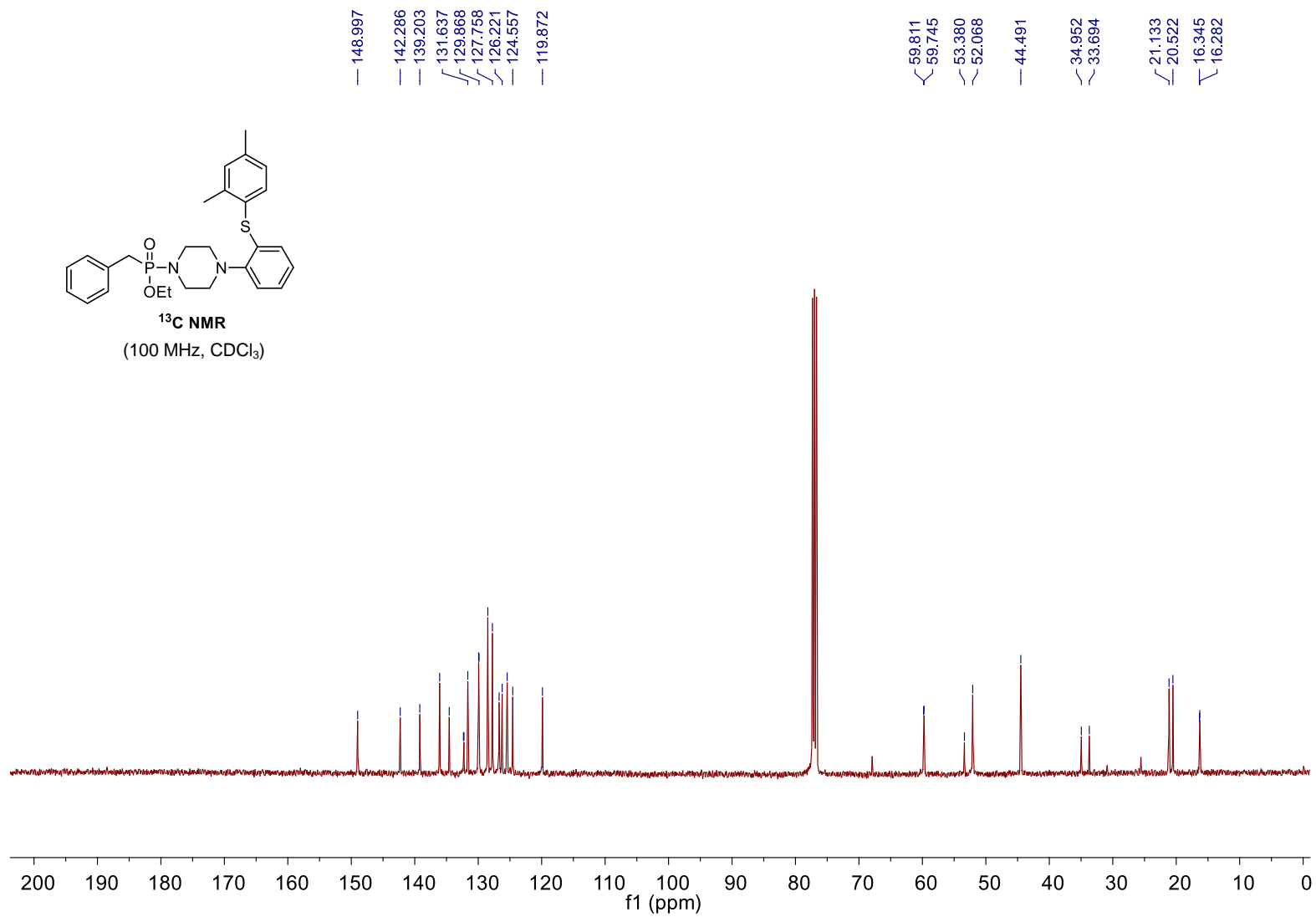
³¹P NMR
(162 MHz, CDCl₃)

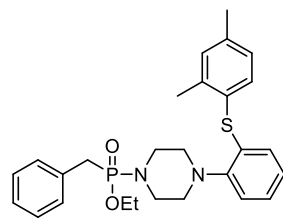




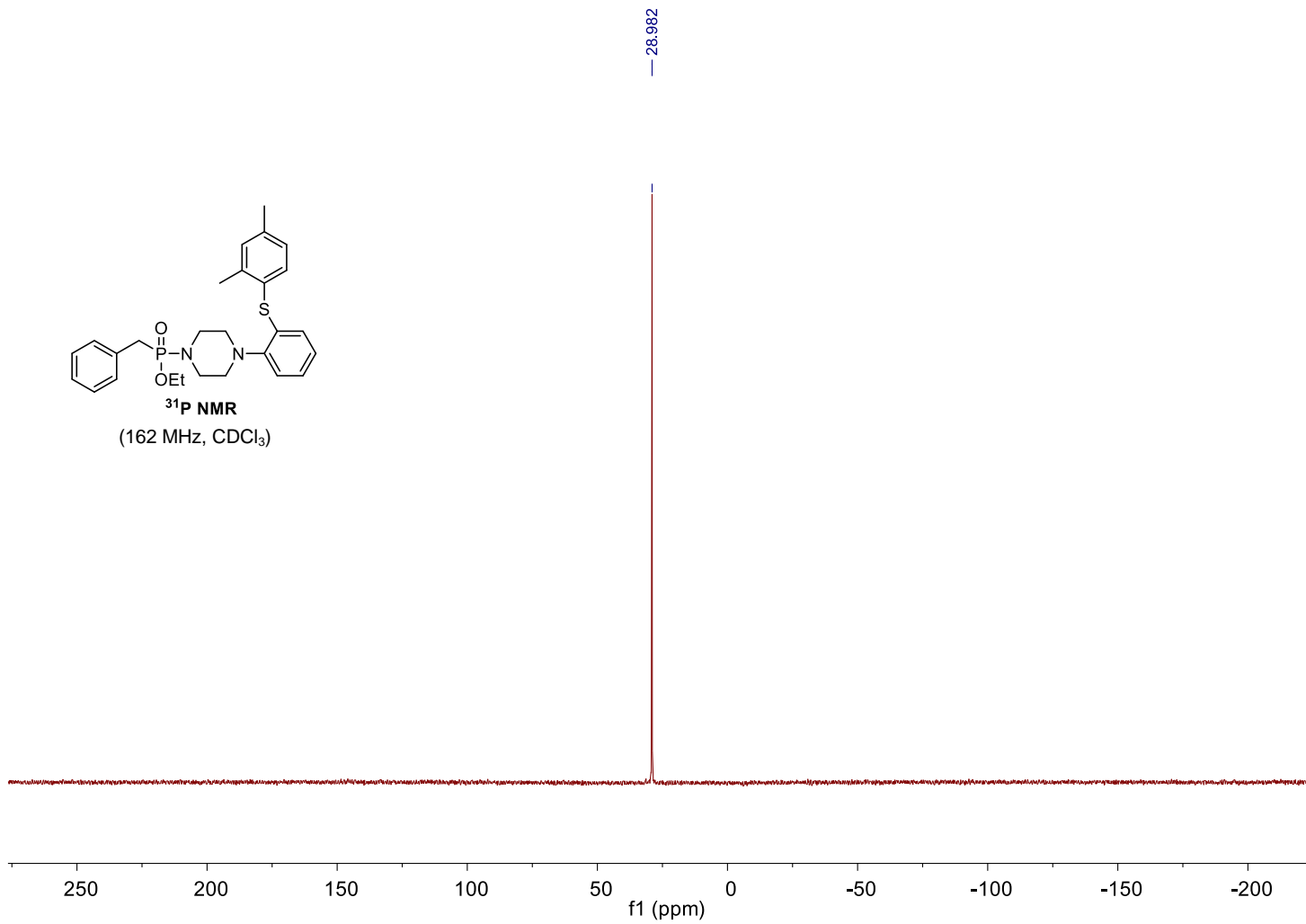


¹³C NMR
(100 MHz, CDCl₃)

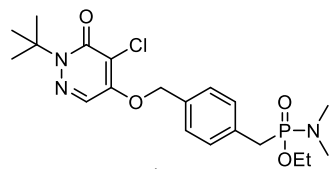




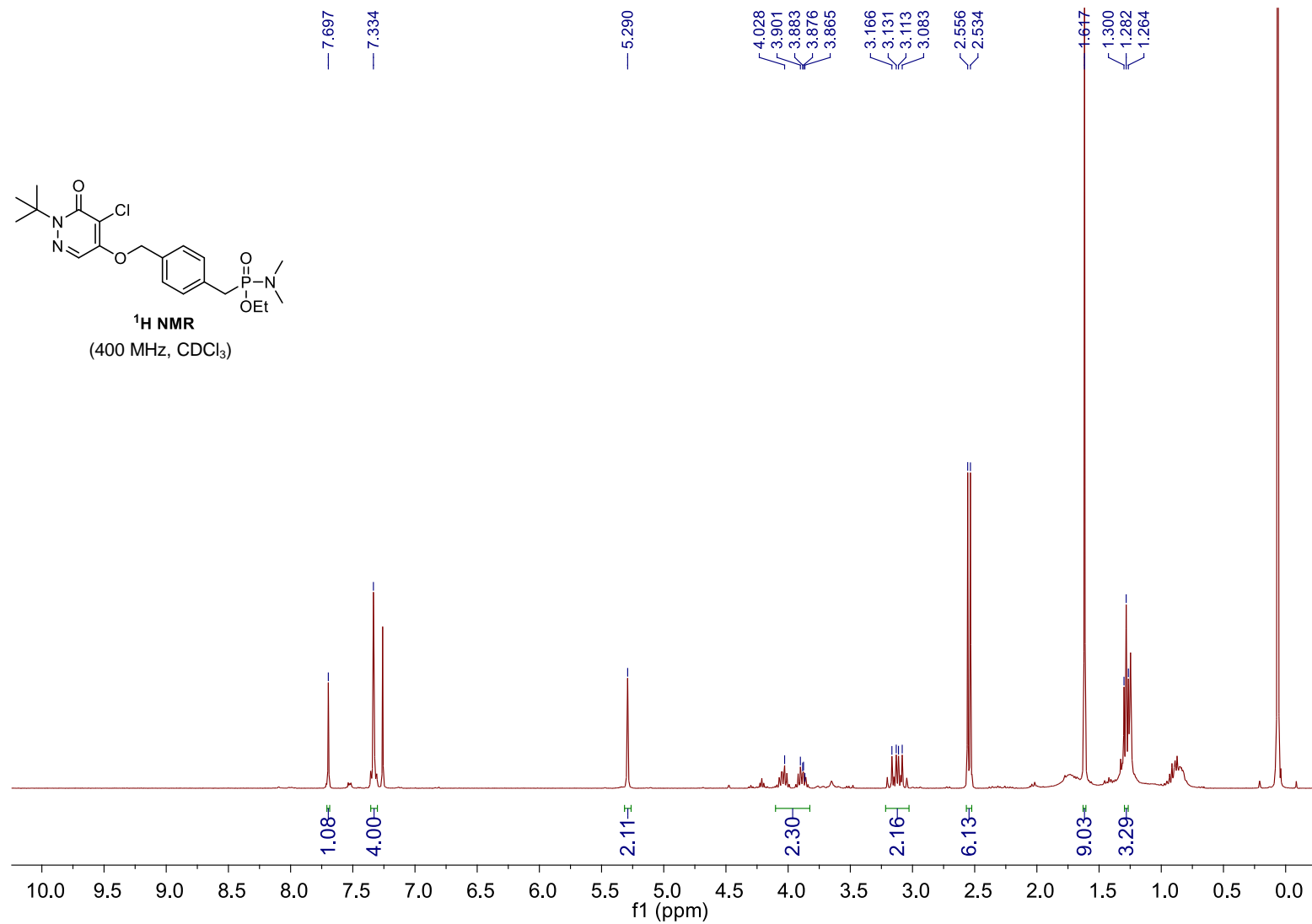
³¹P NMR
(162 MHz, CDCl₃)

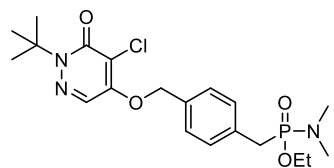


S220

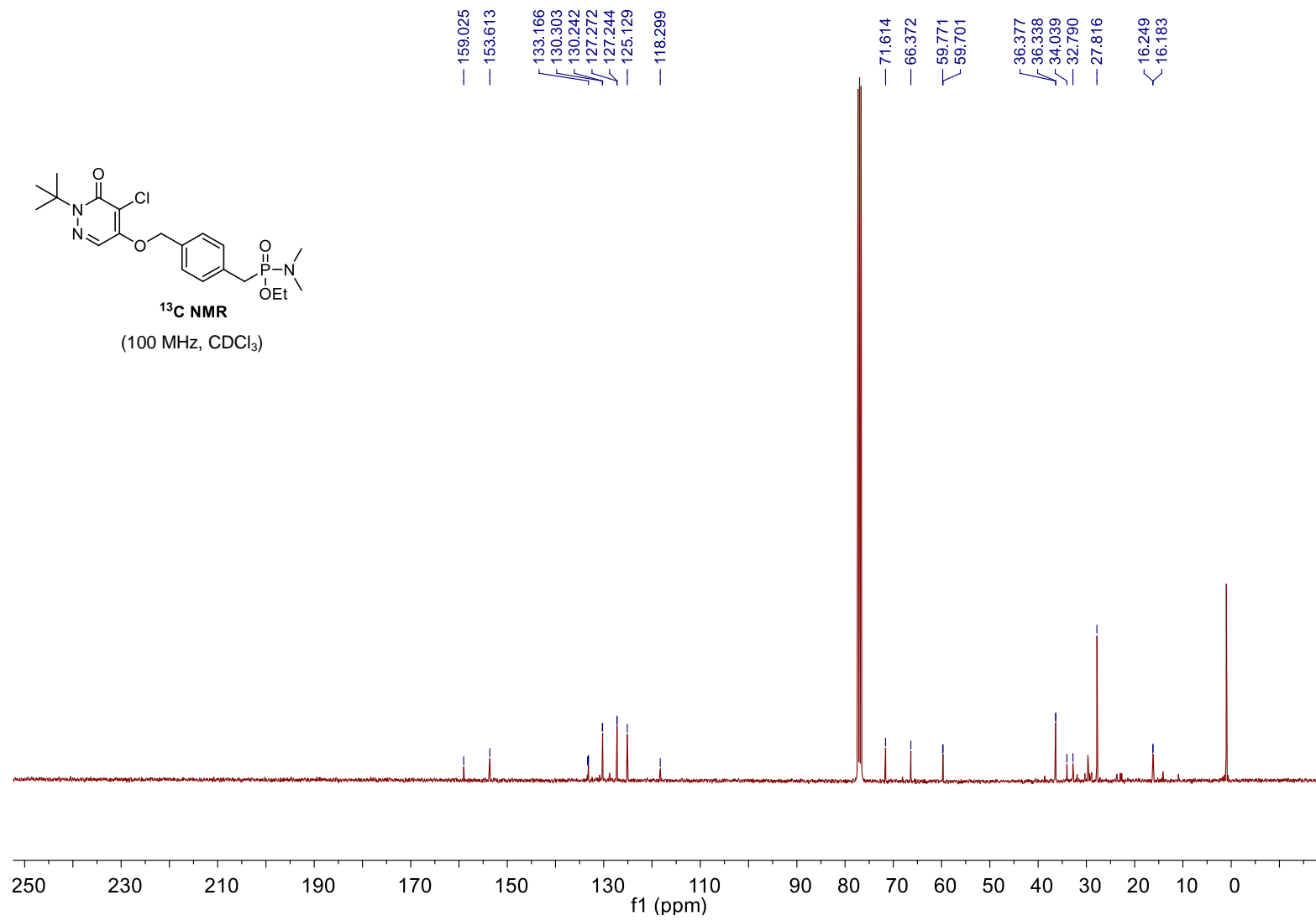


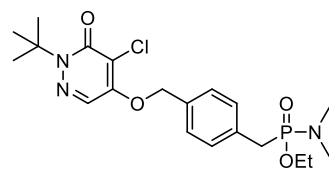
¹H NMR
(400 MHz, CDCl₃)



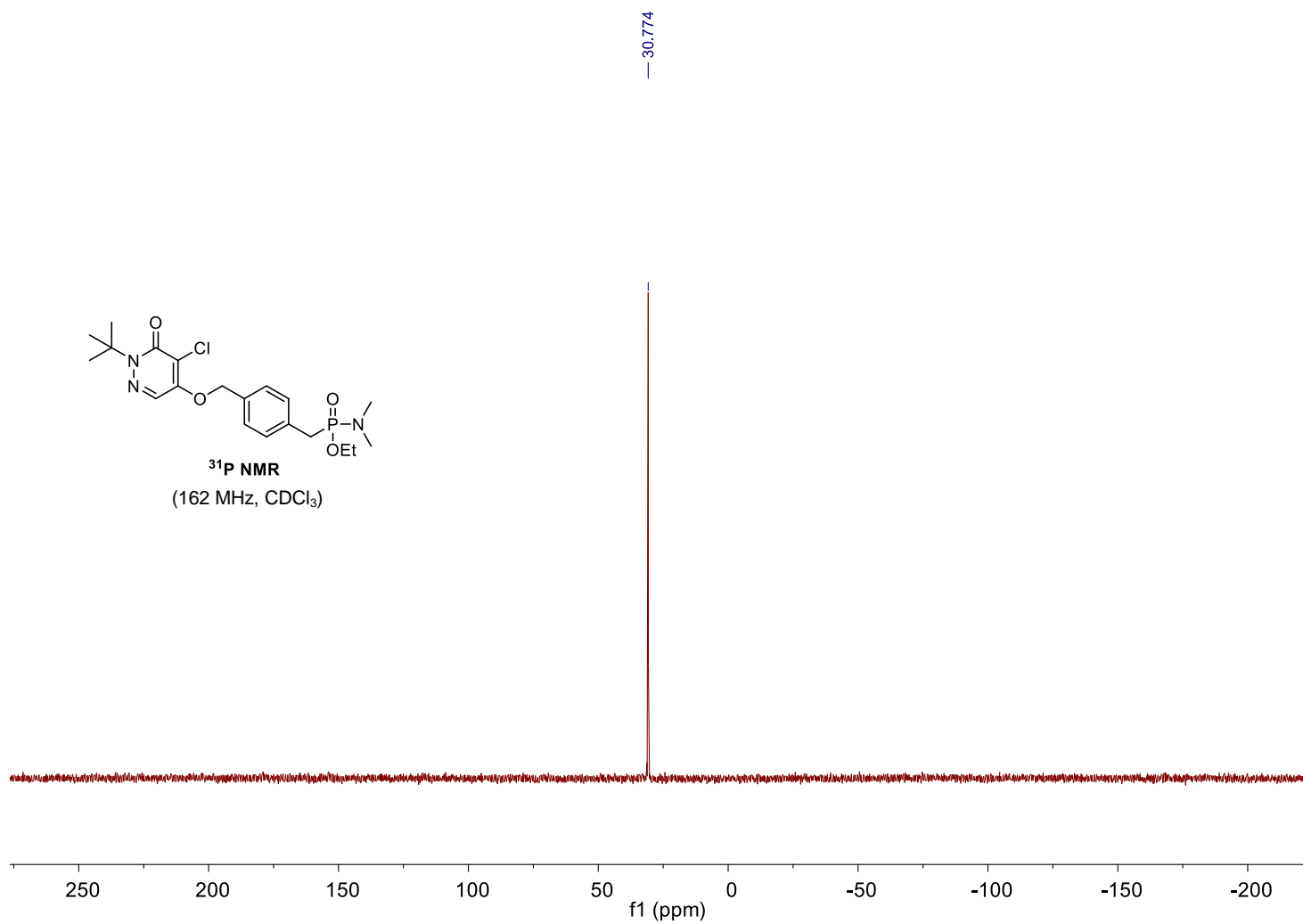


¹³C NMR
(100 MHz, CDCl₃)

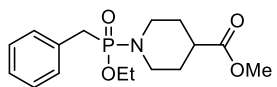




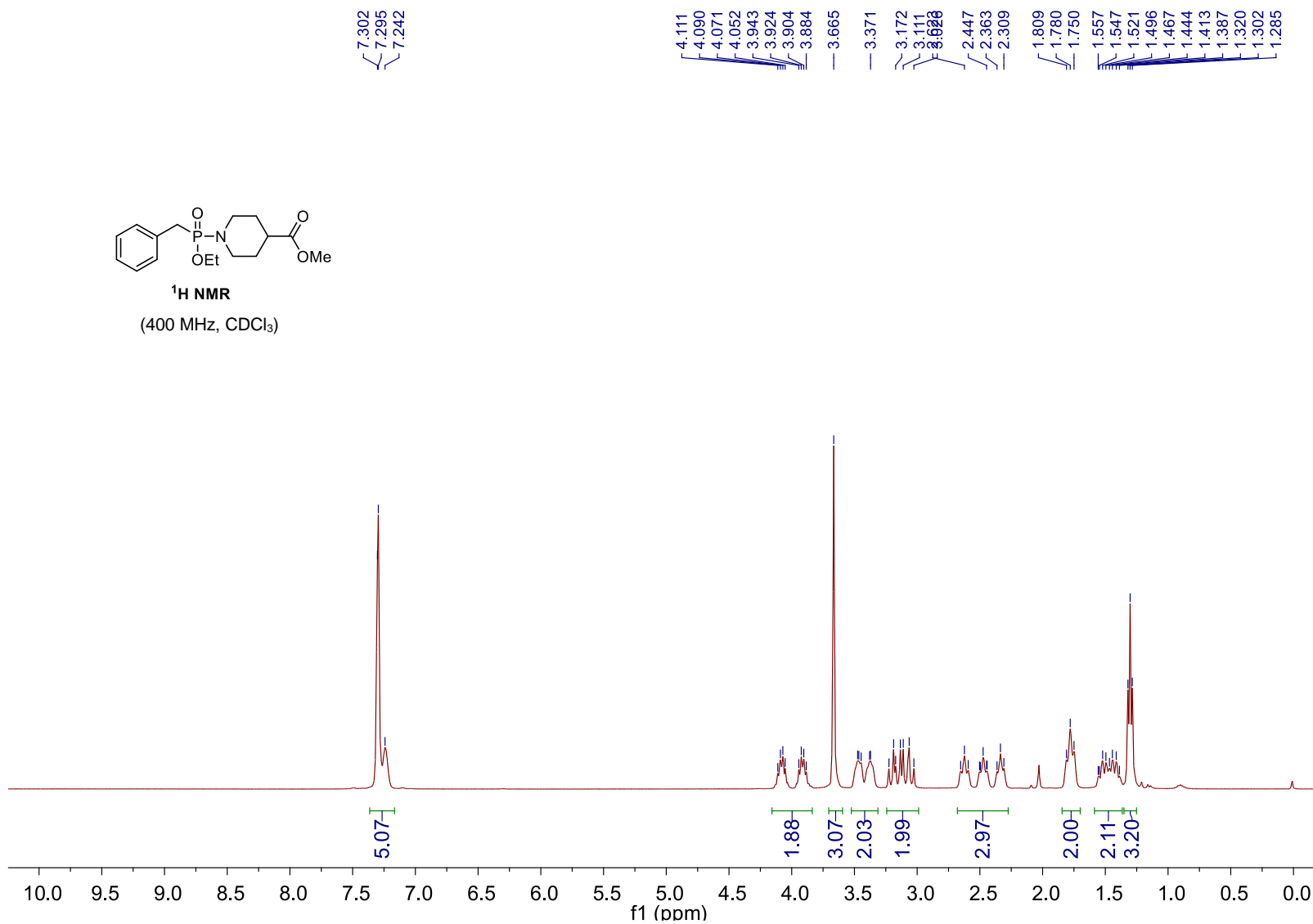
³¹P NMR
(162 MHz, CDCl₃)

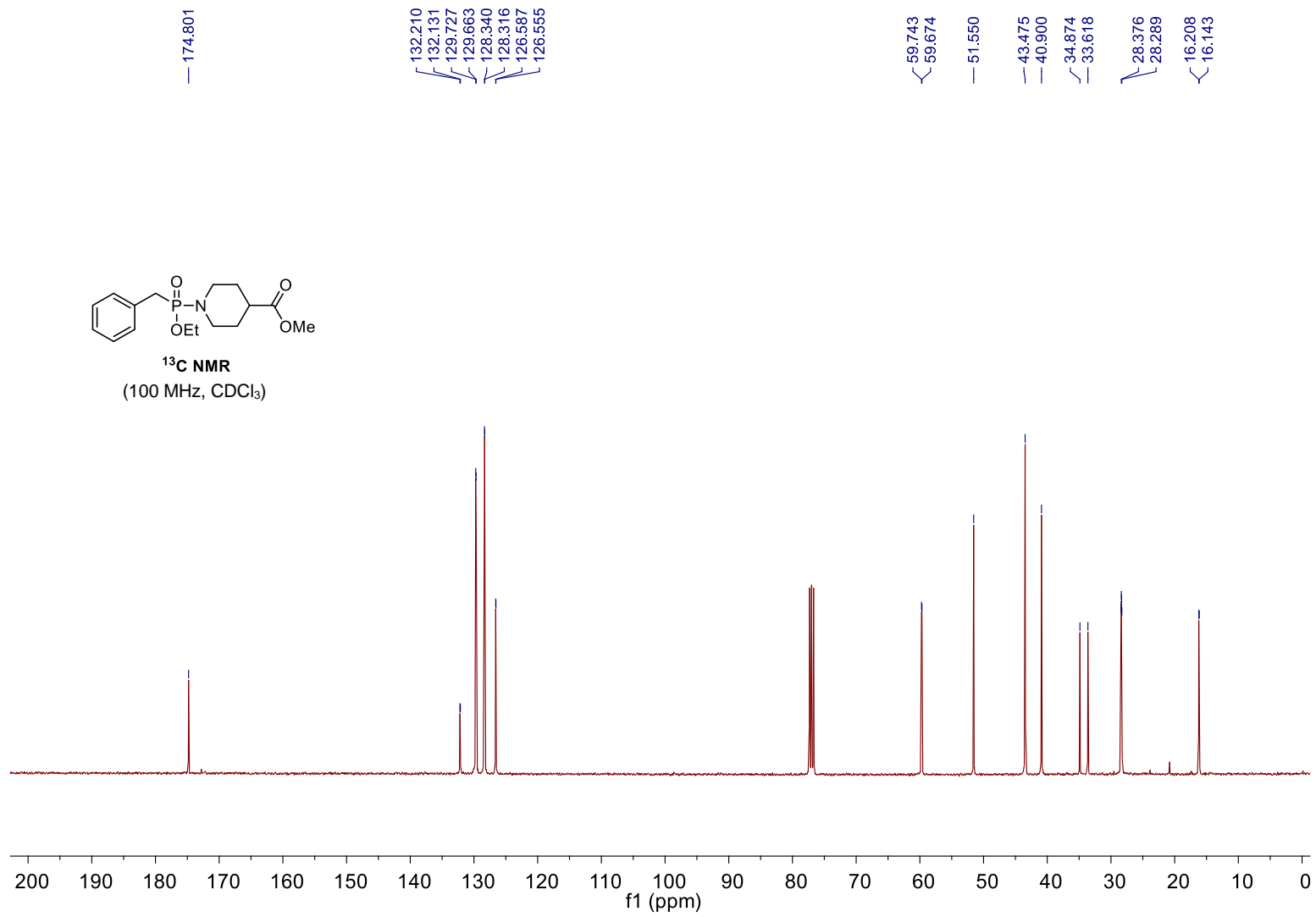


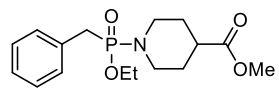
S223



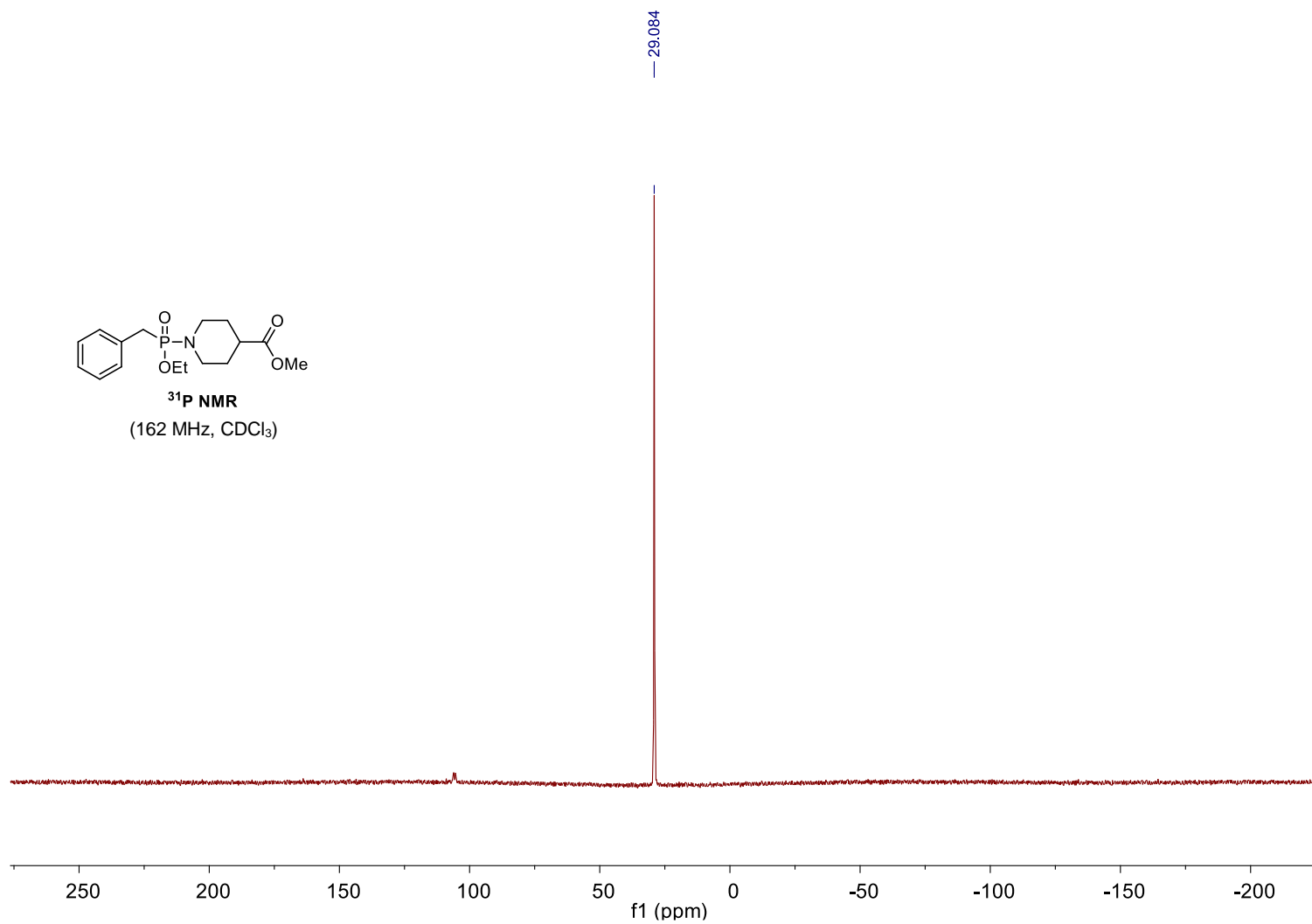
¹H NMR
(400 MHz, CDCl₃)

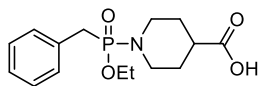






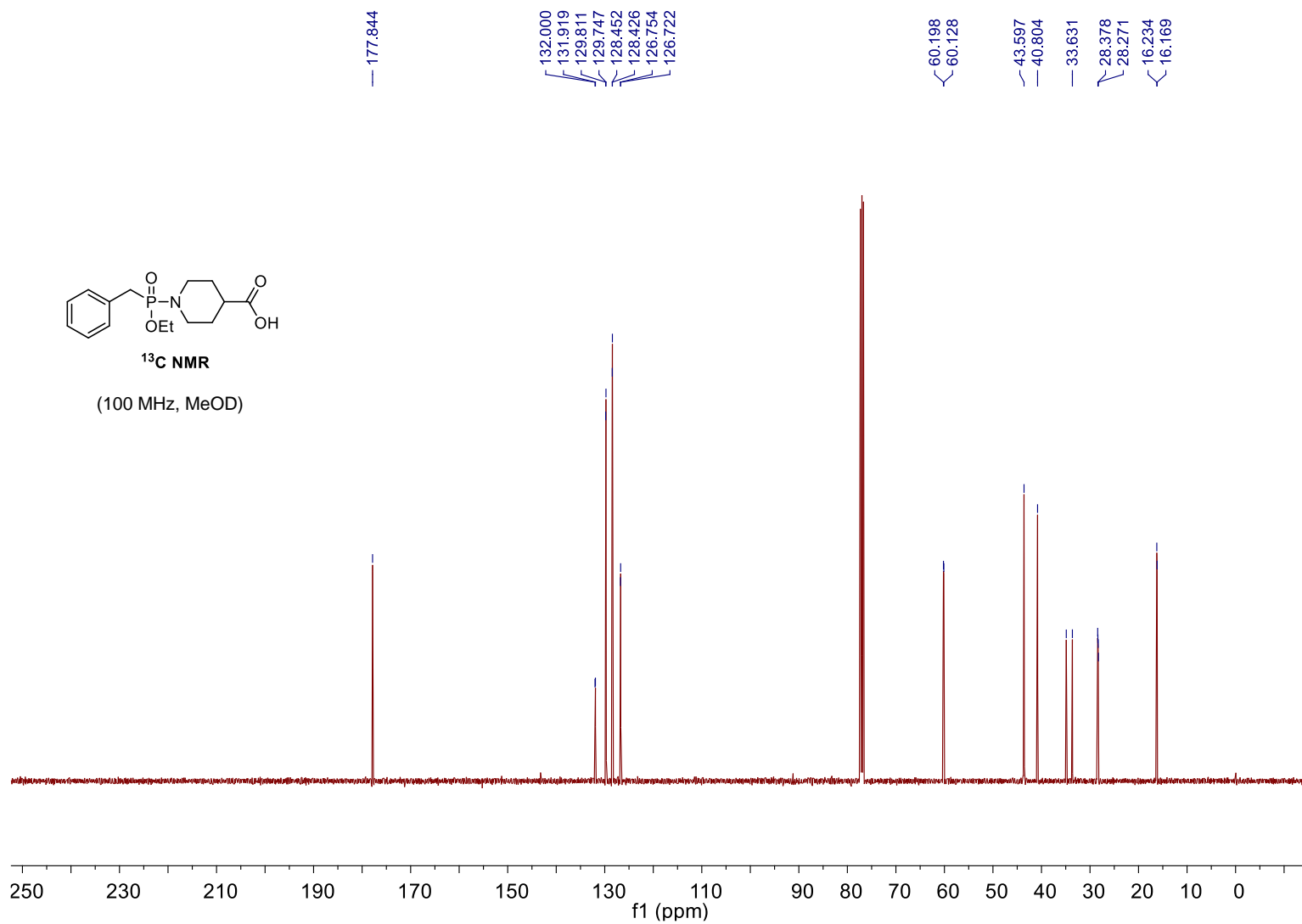
³¹P NMR
(162 MHz, CDCl₃)

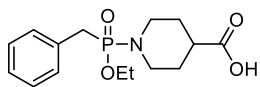




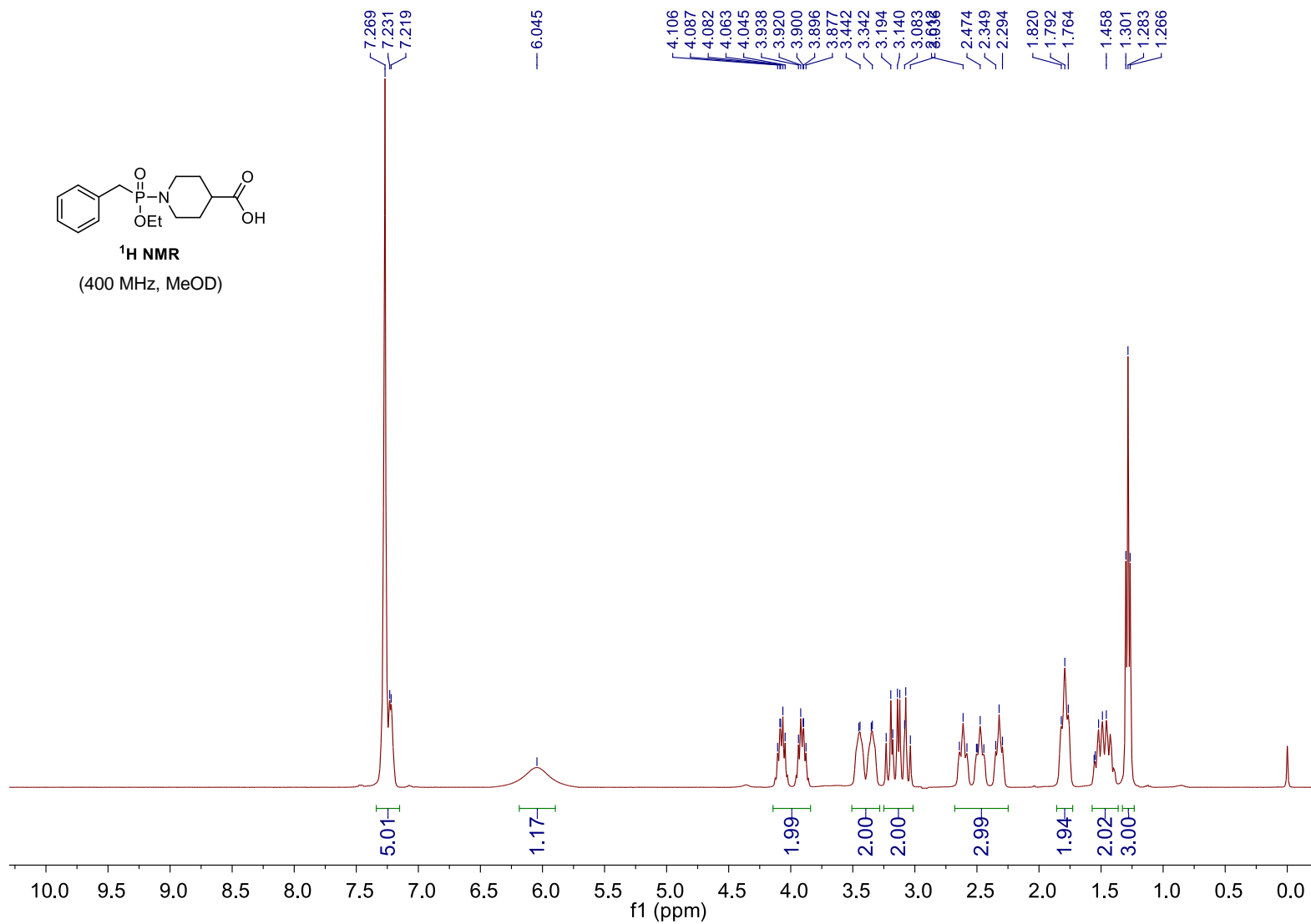
¹³C NMR

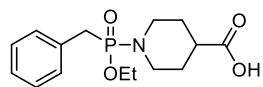
(100 MHz, MeOD)



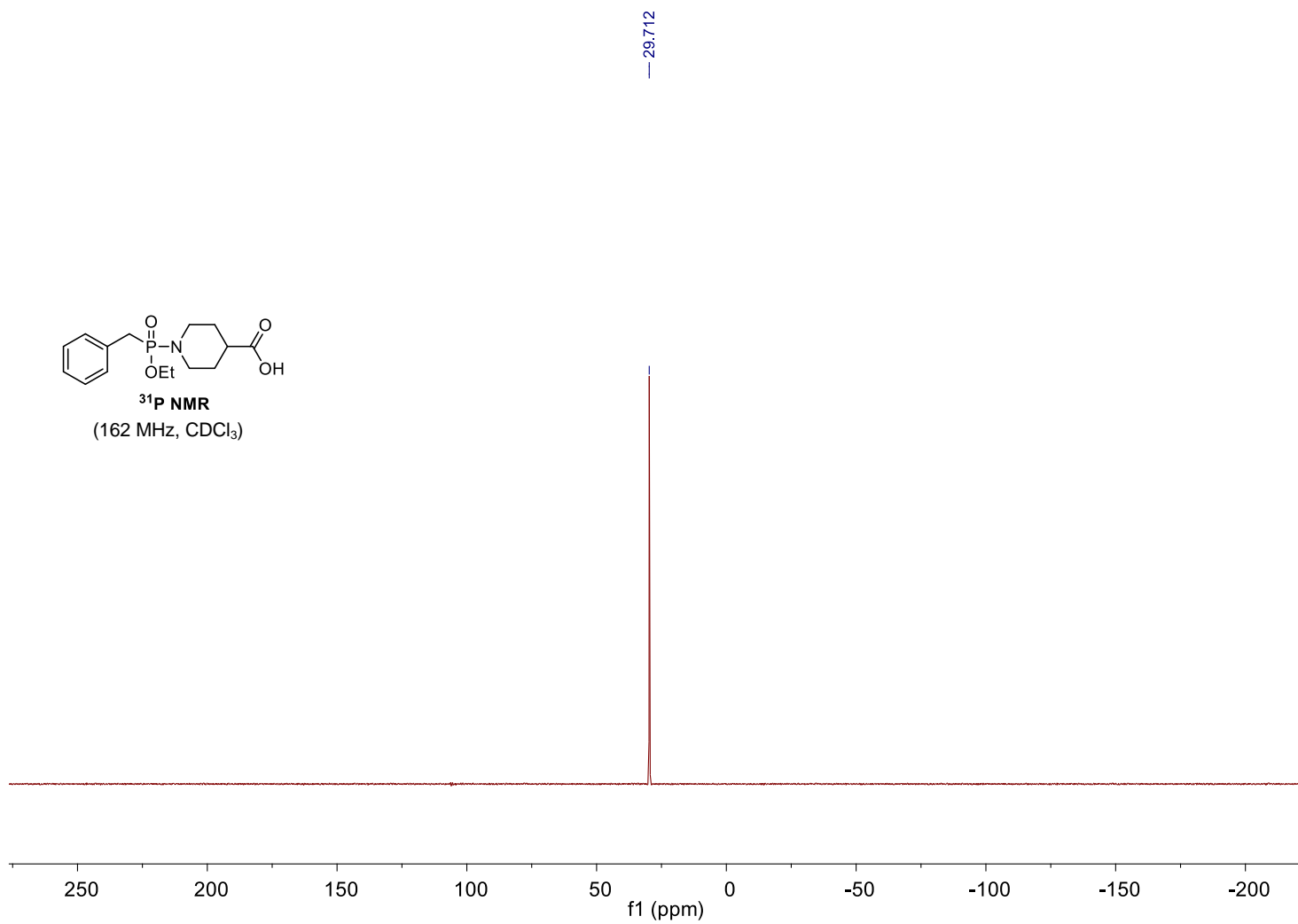


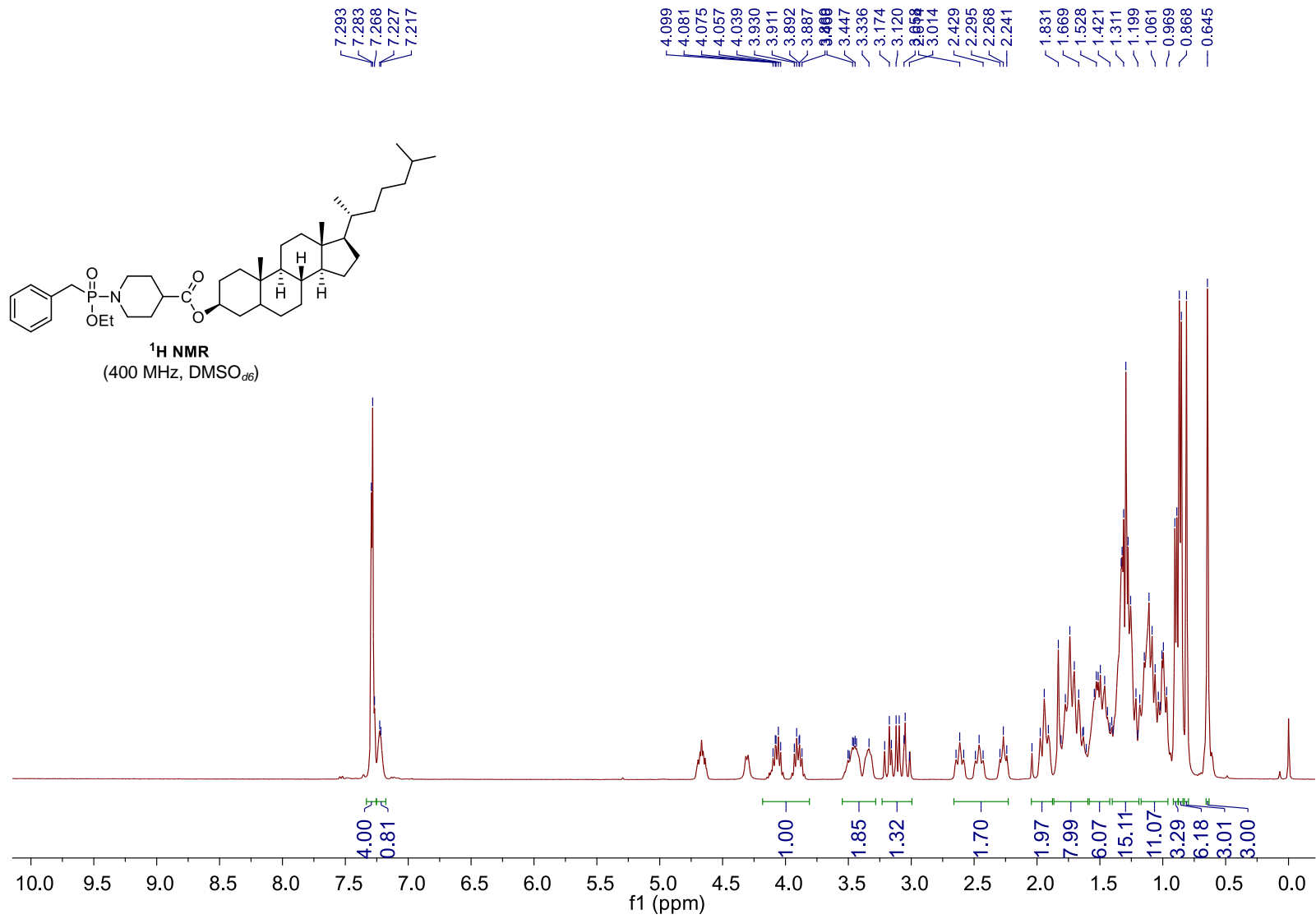
¹H NMR
(400 MHz, MeOD)

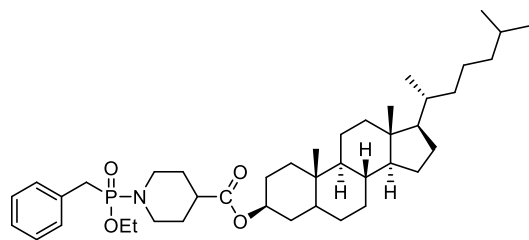




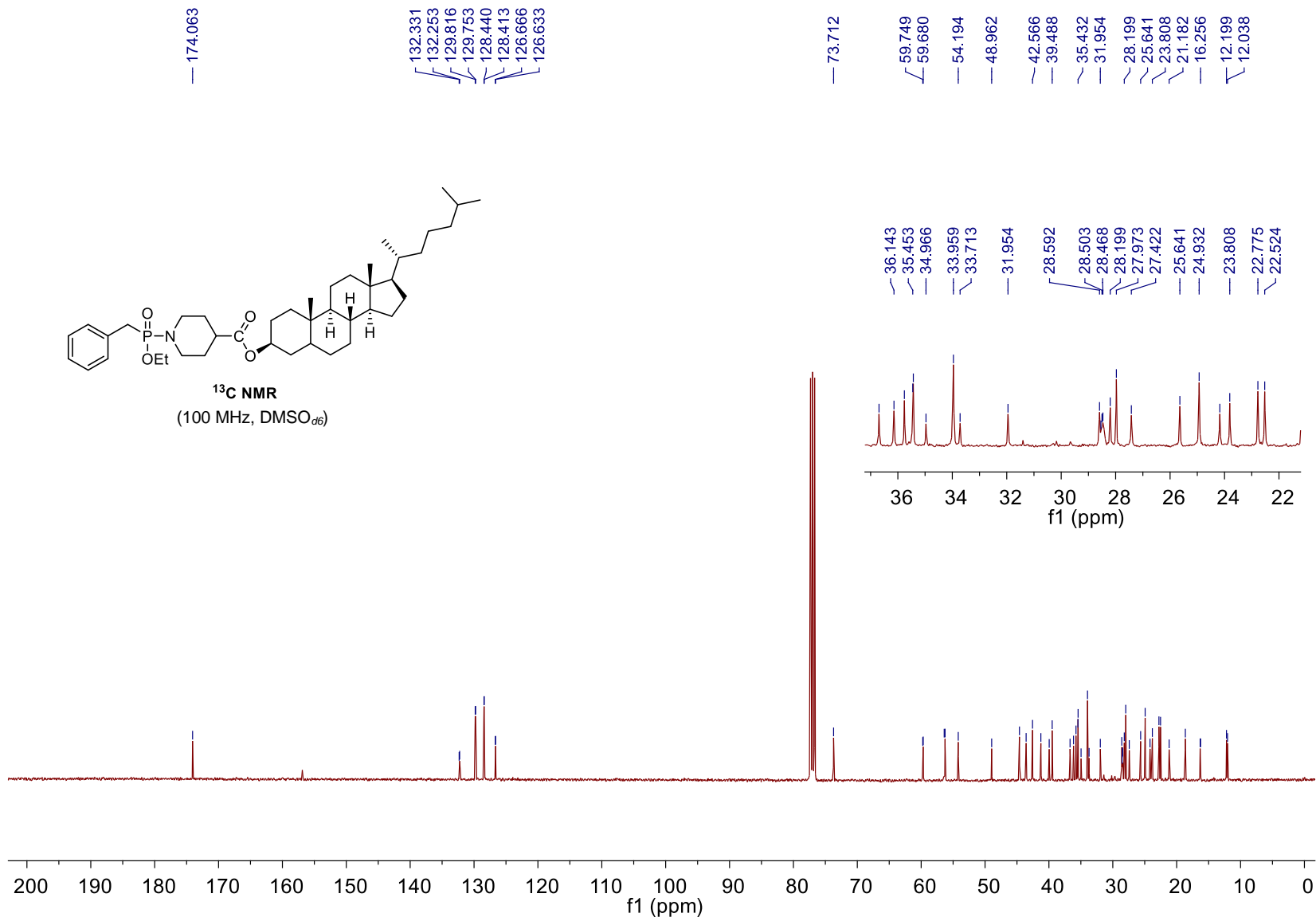
³¹P NMR
(162 MHz, CDCl₃)

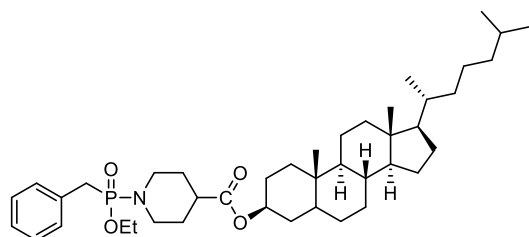




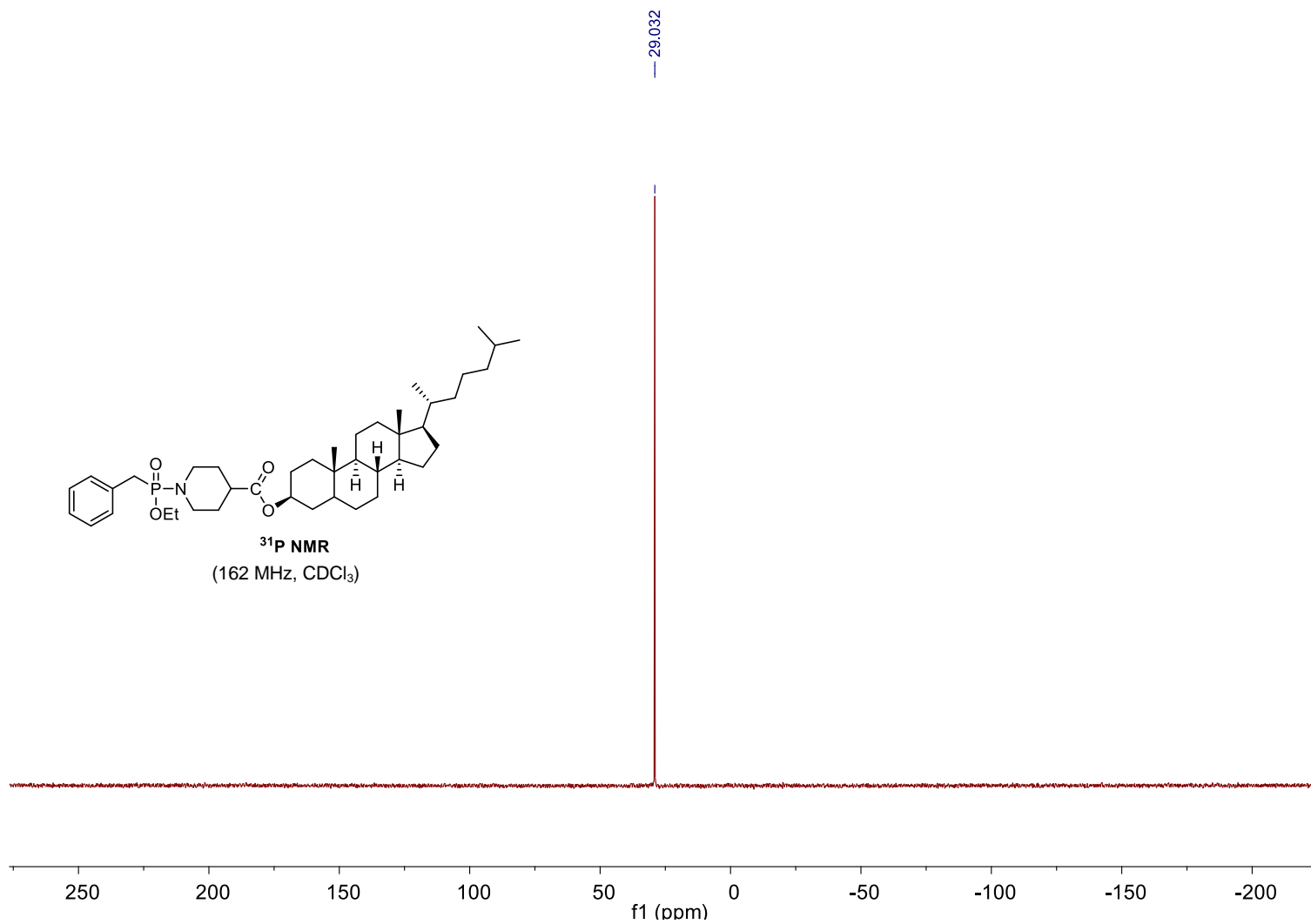


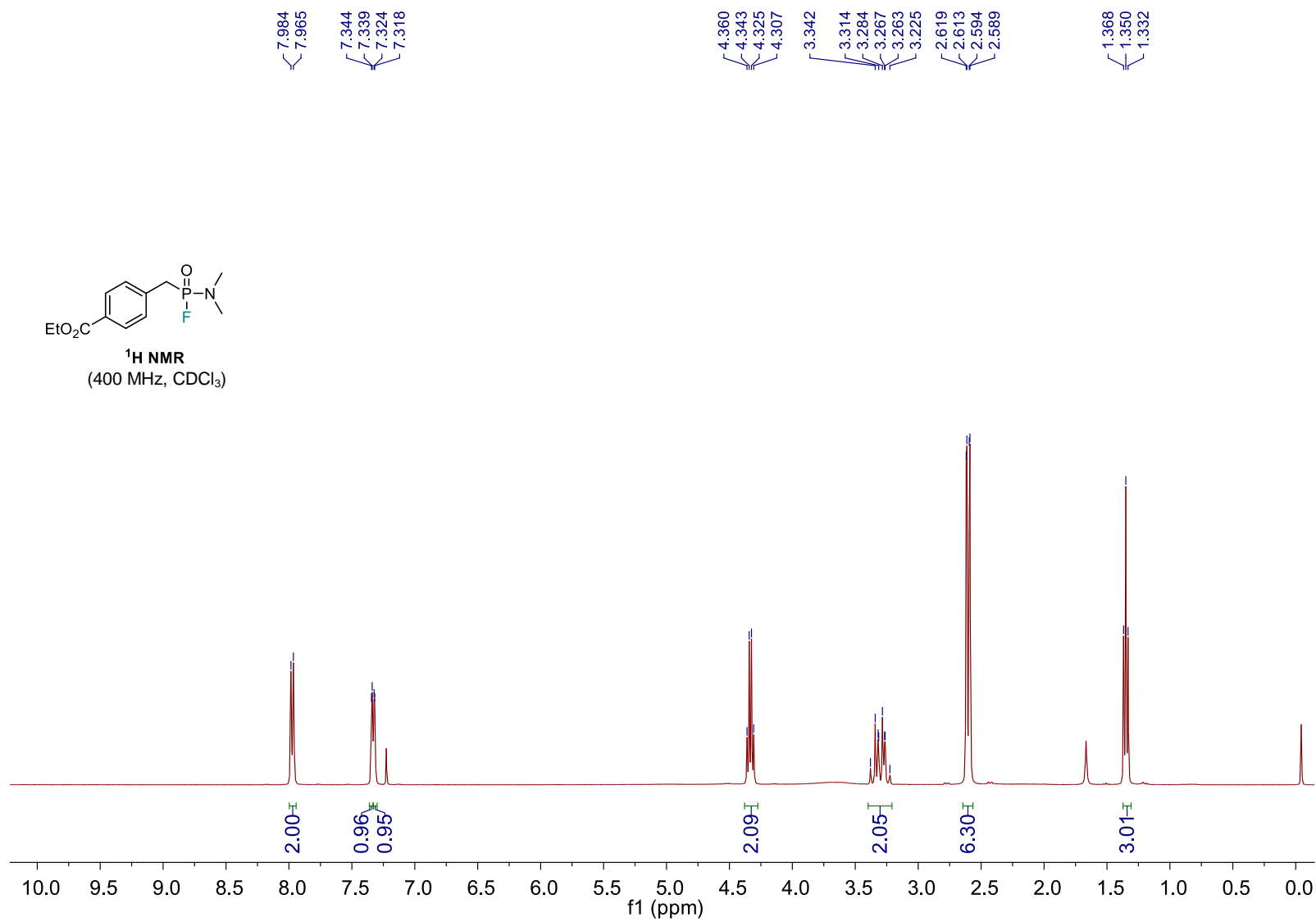
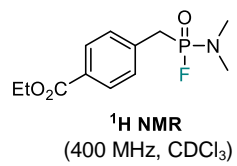
¹³C NMR
(100 MHz, DMSO-*d*₆)

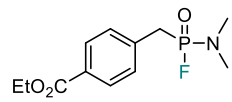




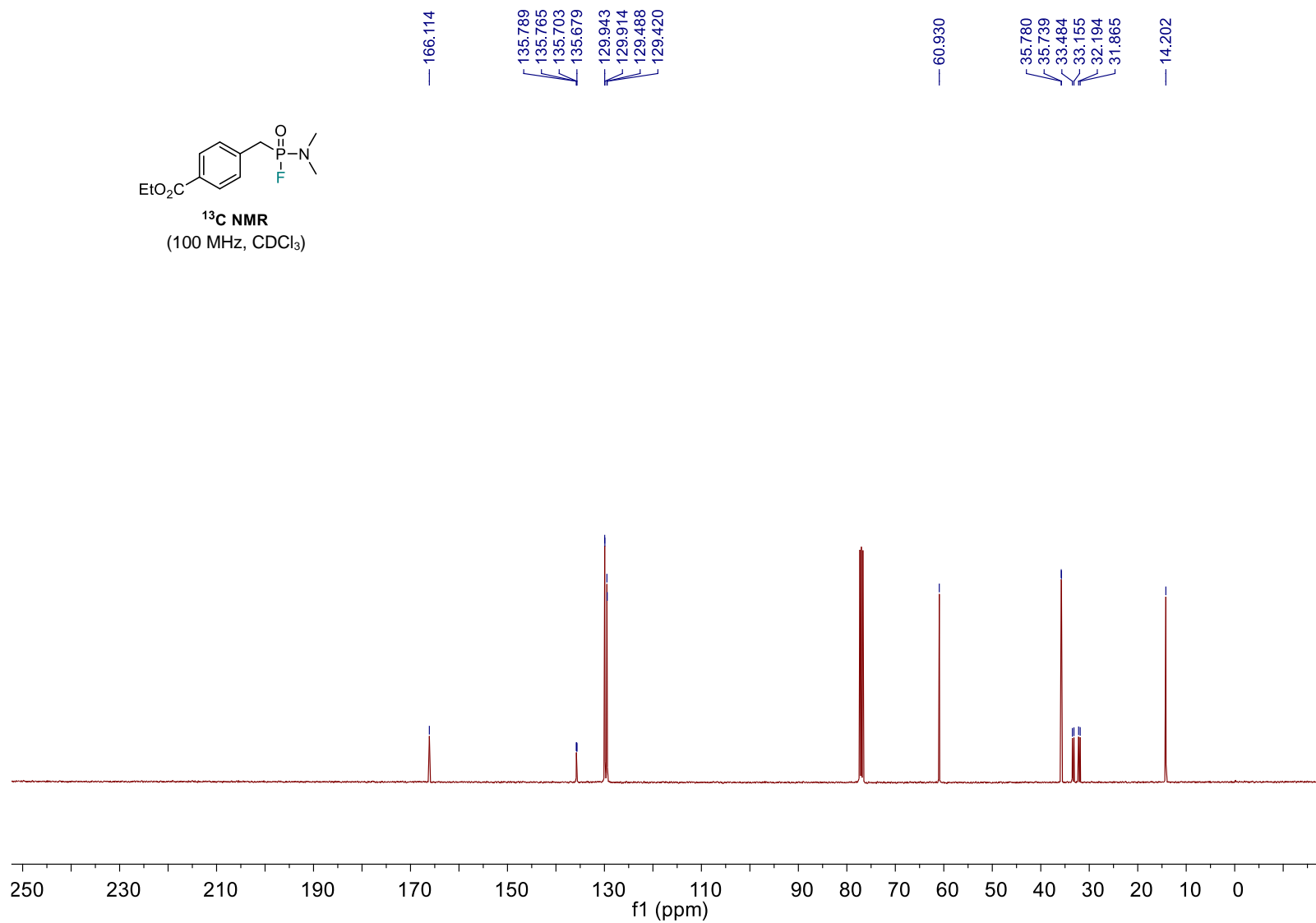
³¹P NMR
(162 MHz, CDCl₃)



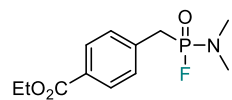




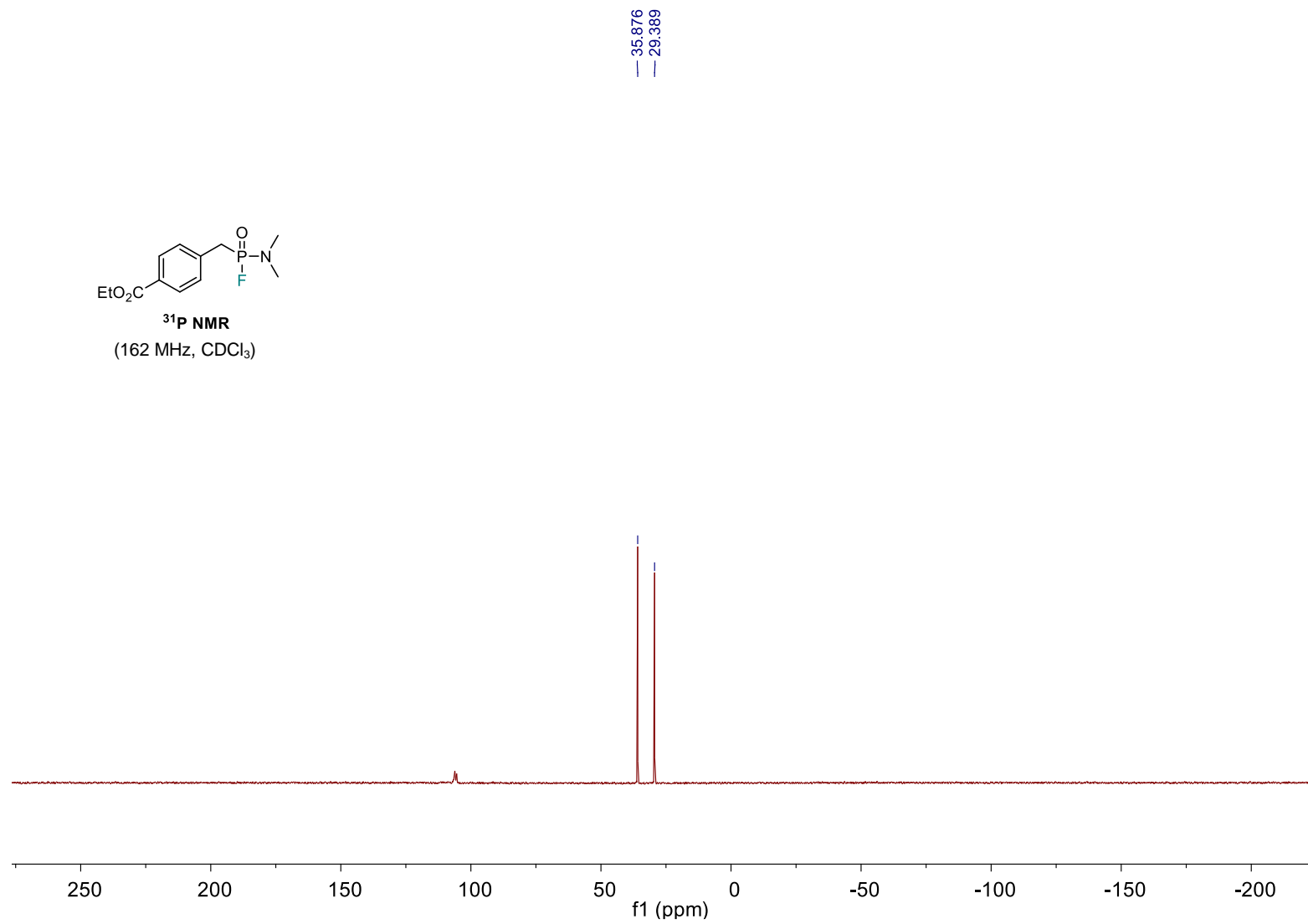
¹³C NMR
(100 MHz, CDCl₃)



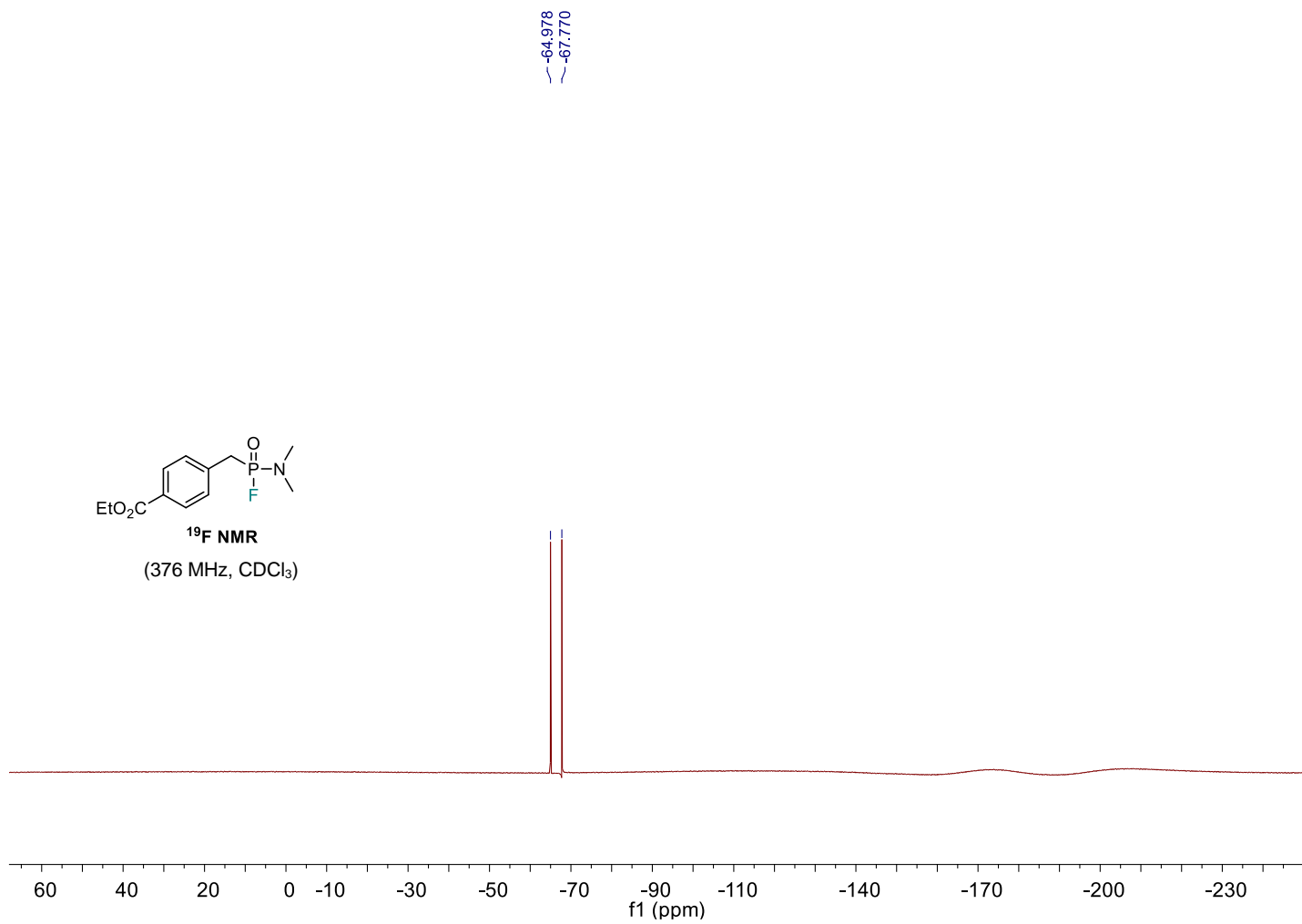
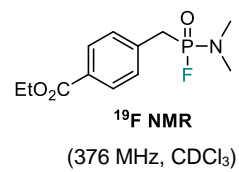
S234

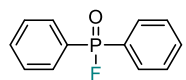


³¹P NMR
(162 MHz, CDCl₃)

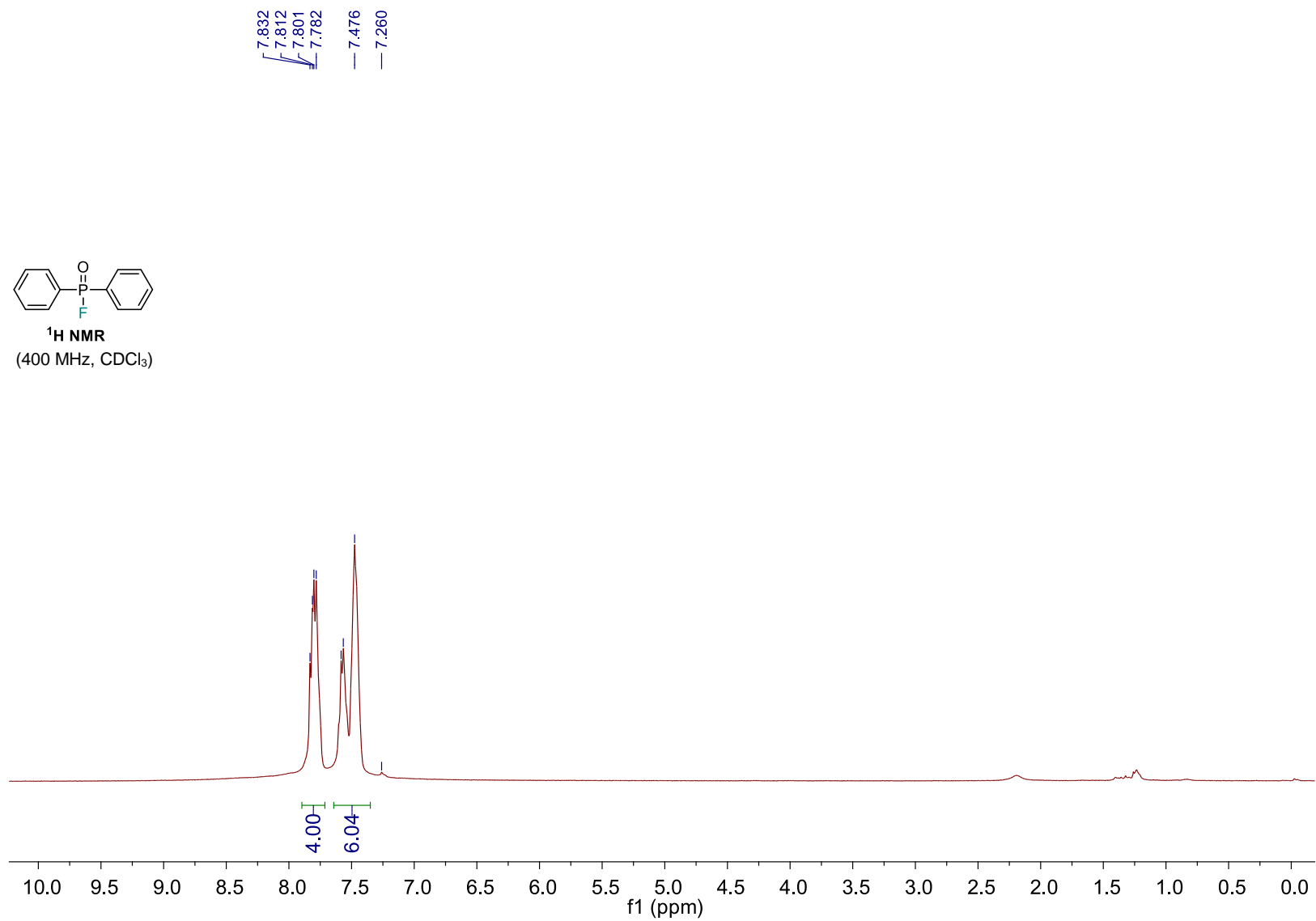


S235

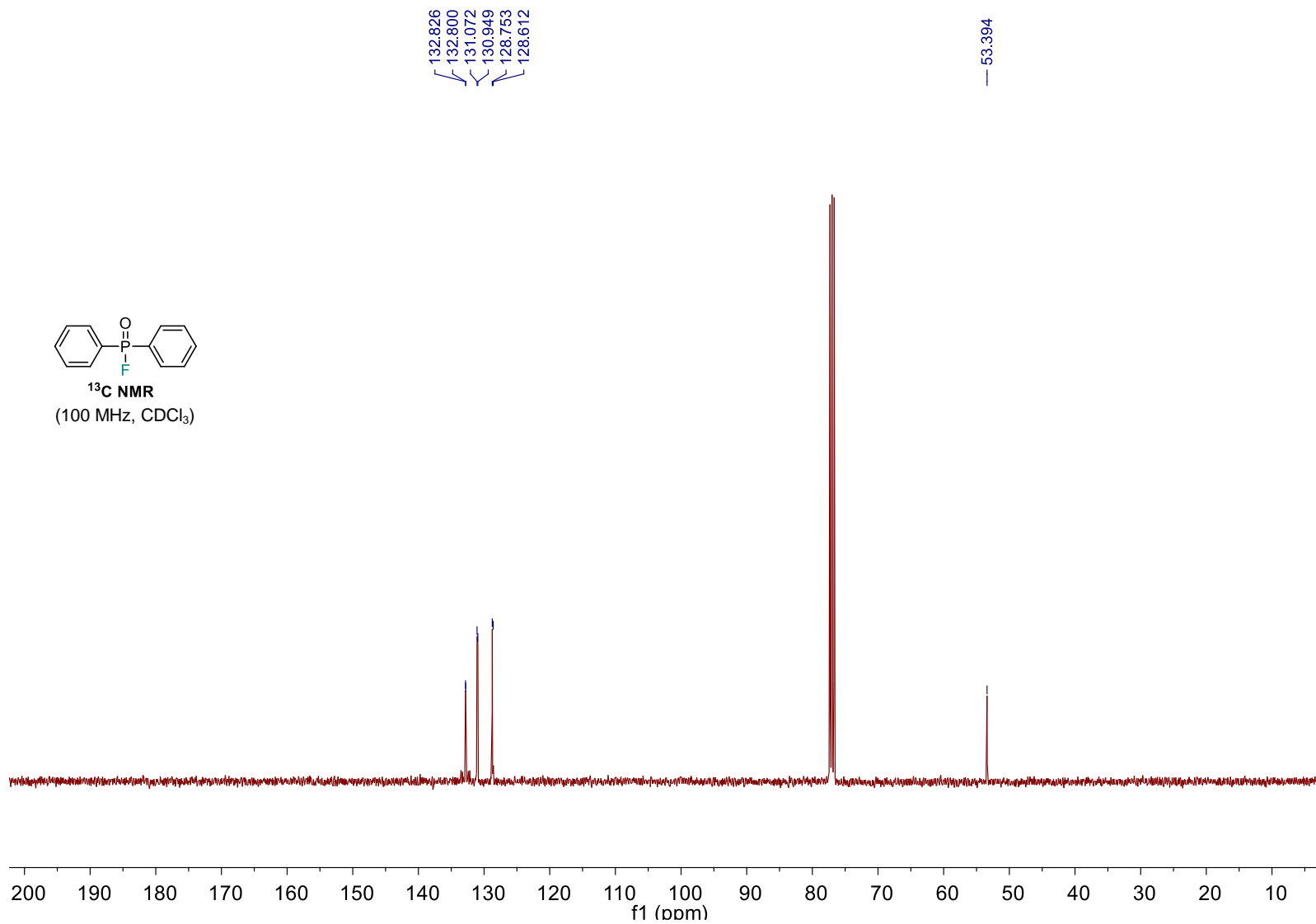
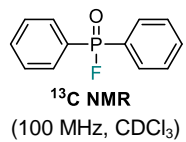


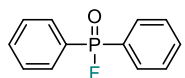


¹H NMR
(400 MHz, CDCl₃)

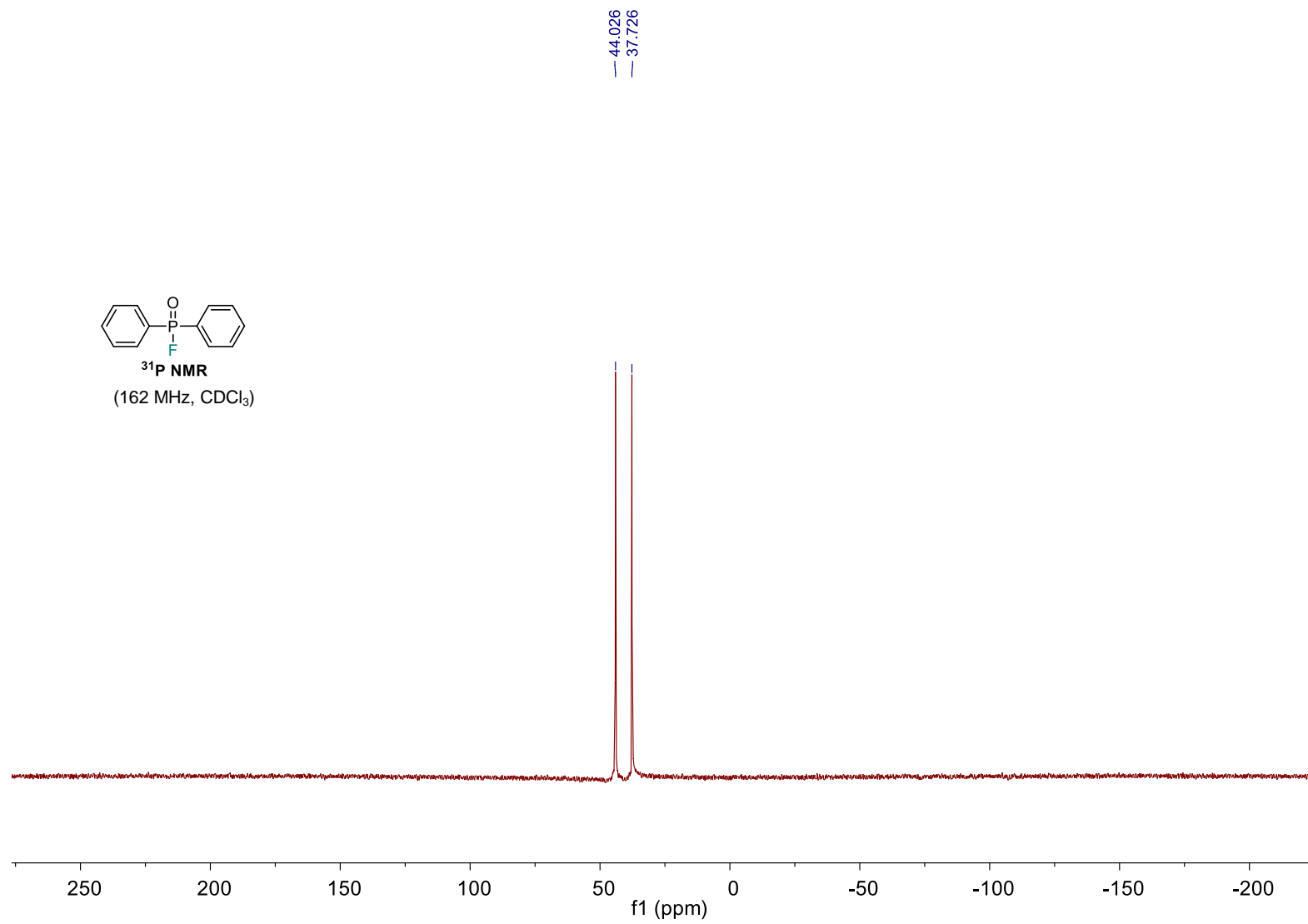


S237

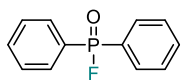




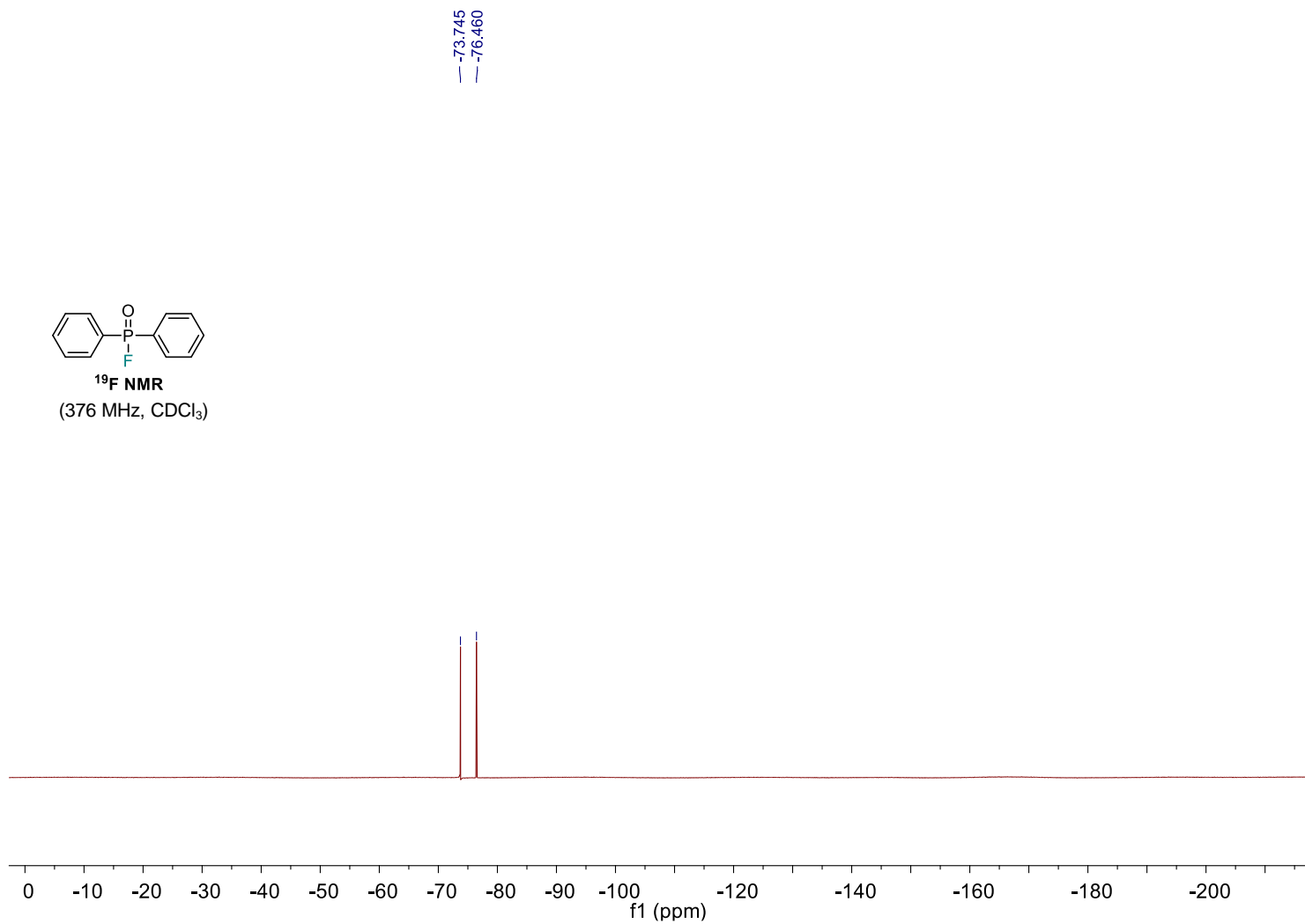
³¹P NMR
(162 MHz, CDCl₃)

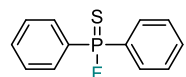


S239

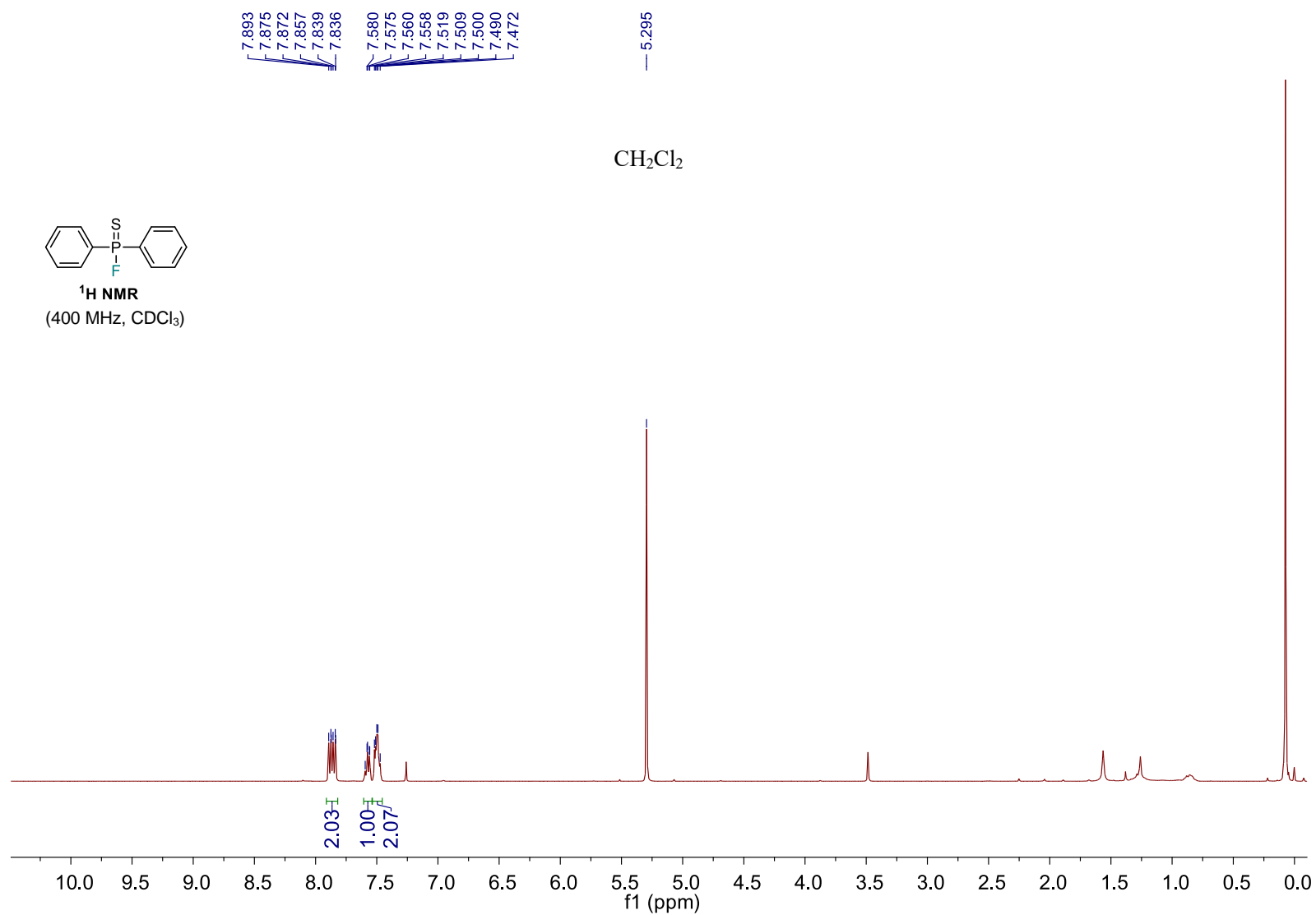


¹⁹F NMR
(376 MHz, CDCl₃)



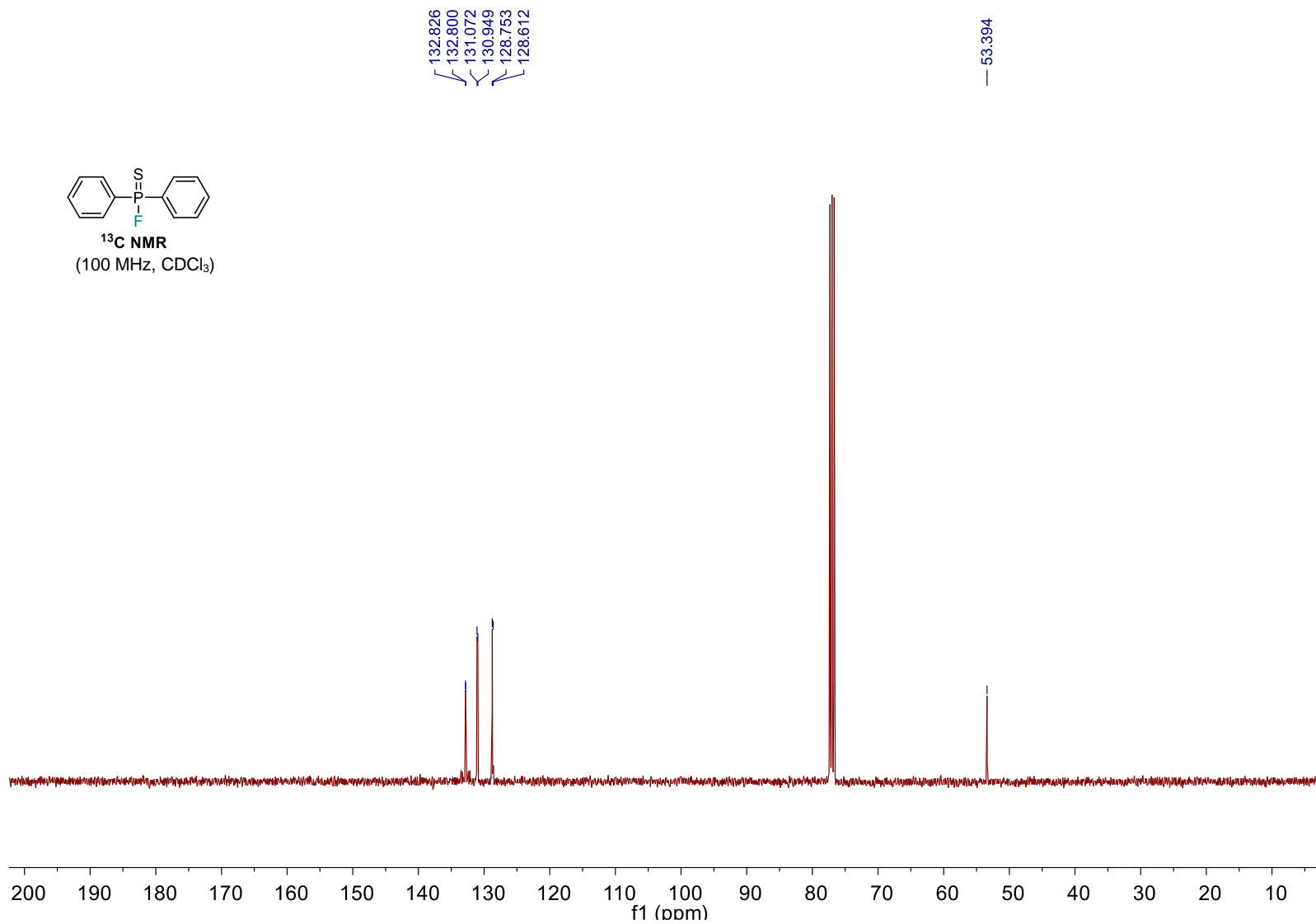


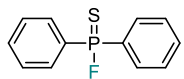
¹H NMR
(400 MHz, CDCl₃)





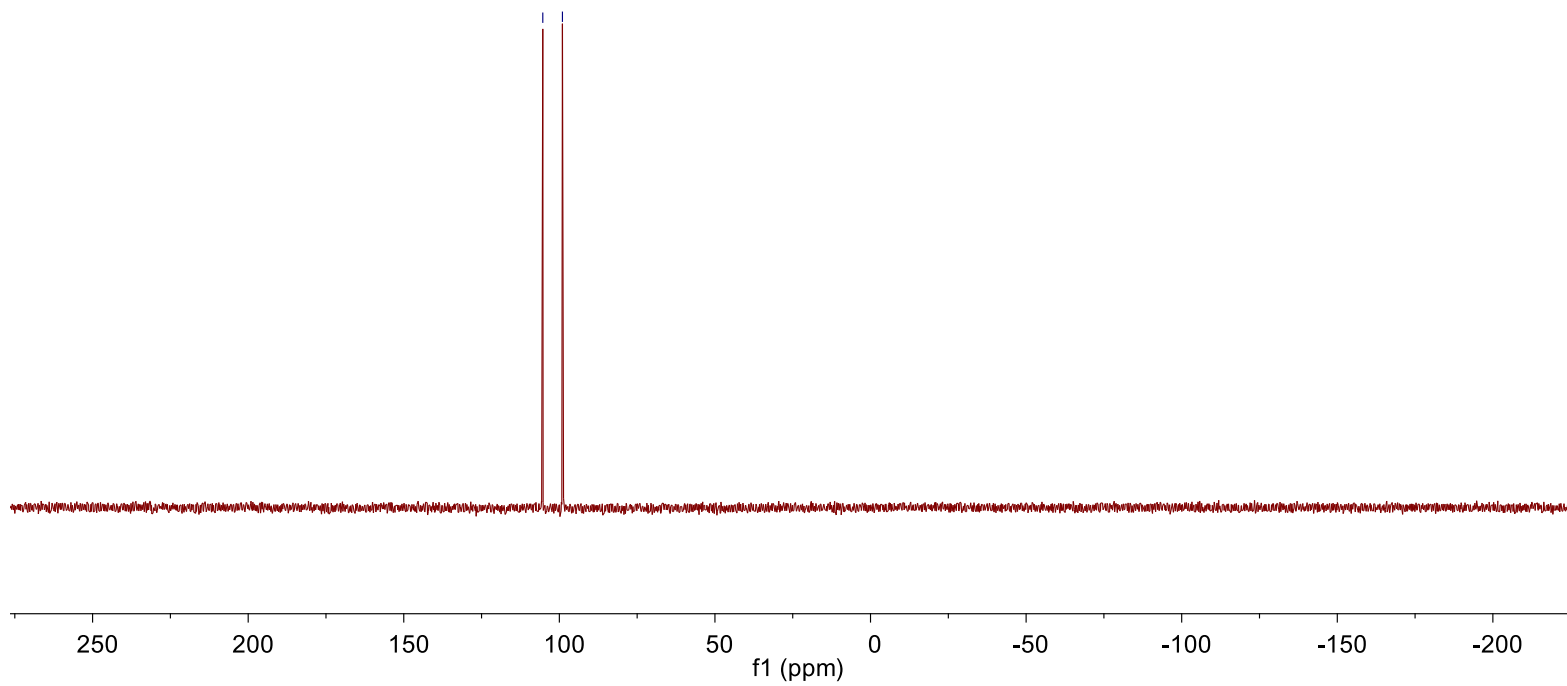
¹³C NMR
(100 MHz, CDCl₃)



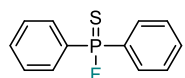


³¹P NMR
(162 MHz, CDCl₃)

105.321
99.010

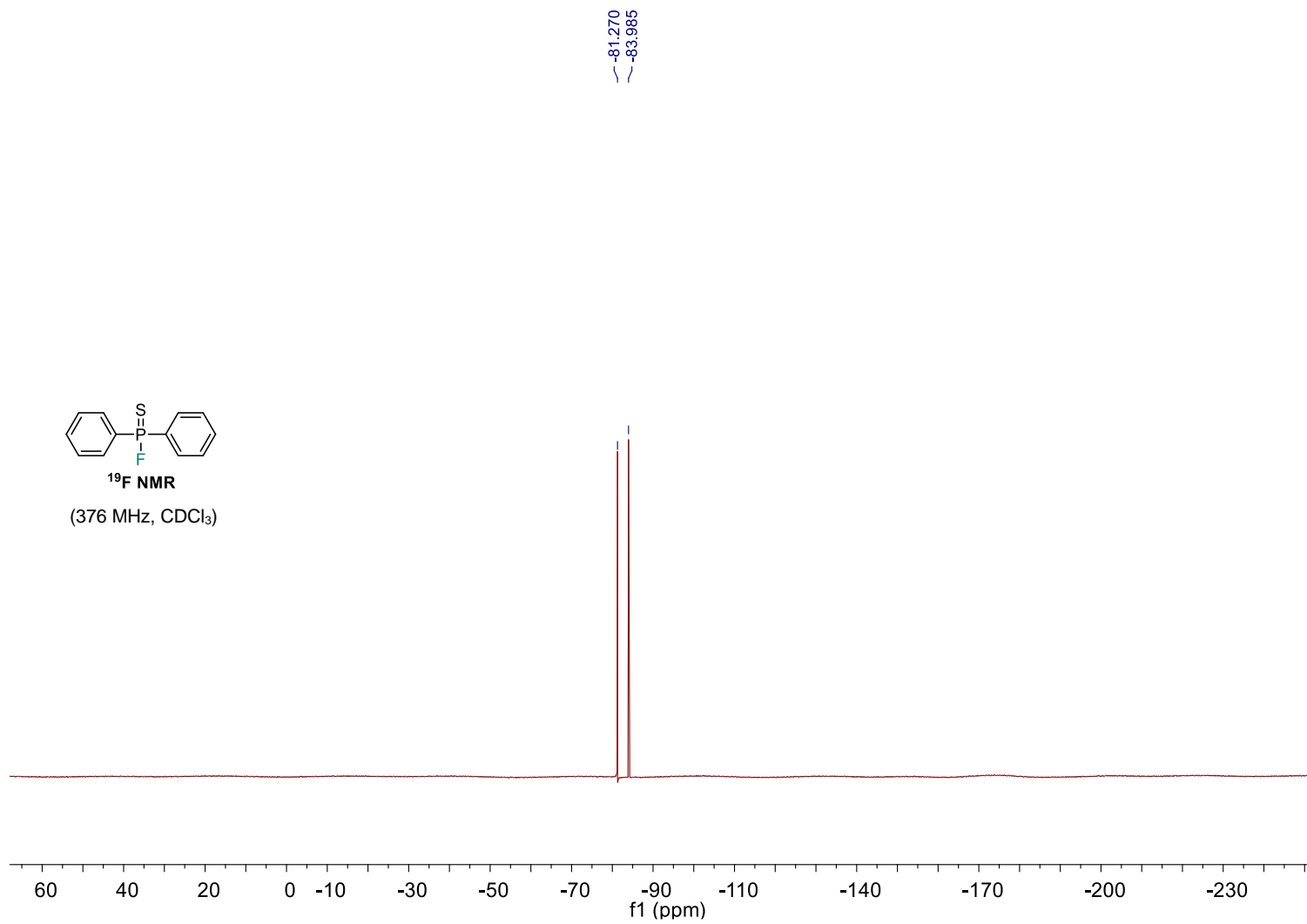


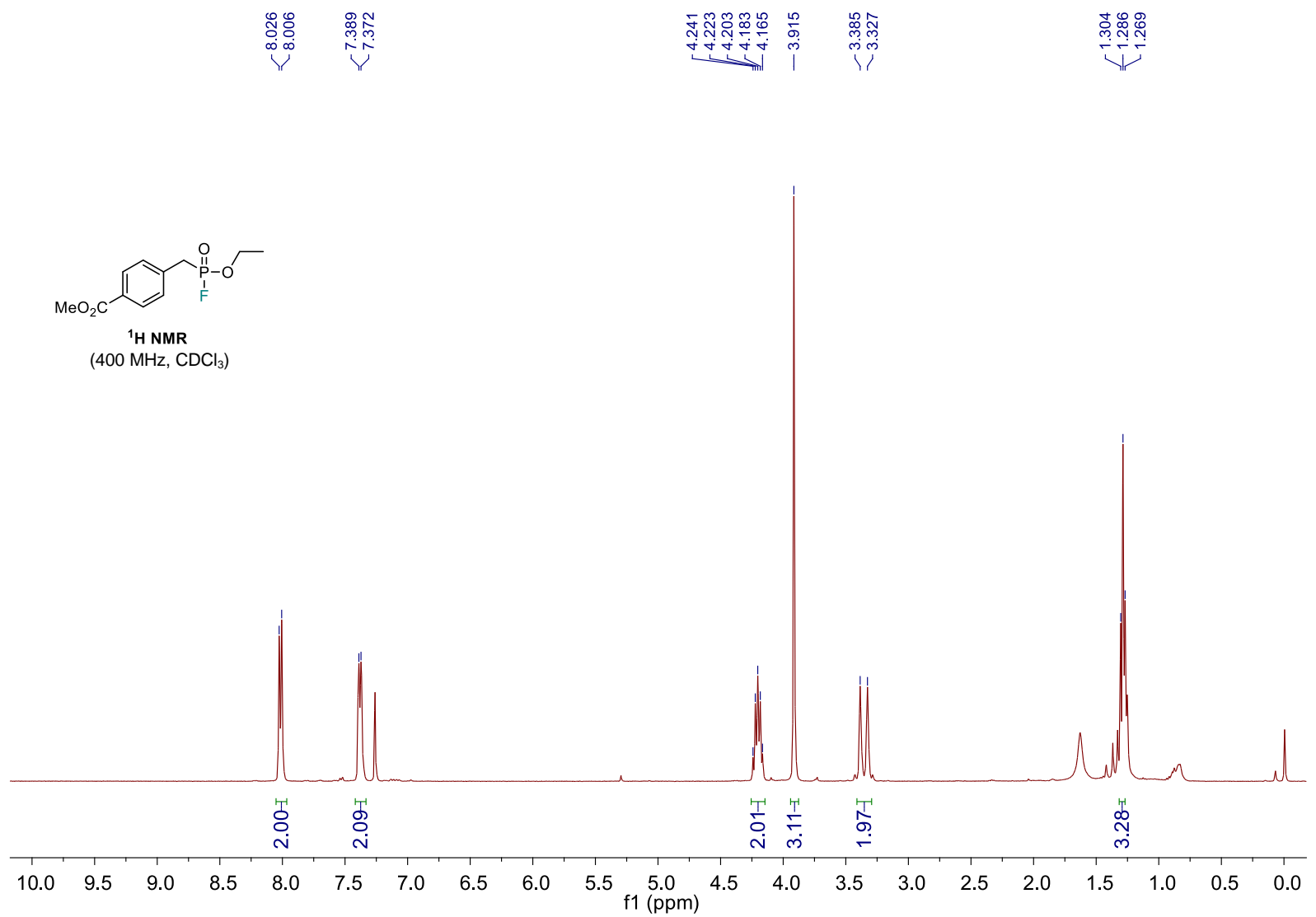
S243

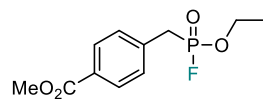


¹⁹F NMR

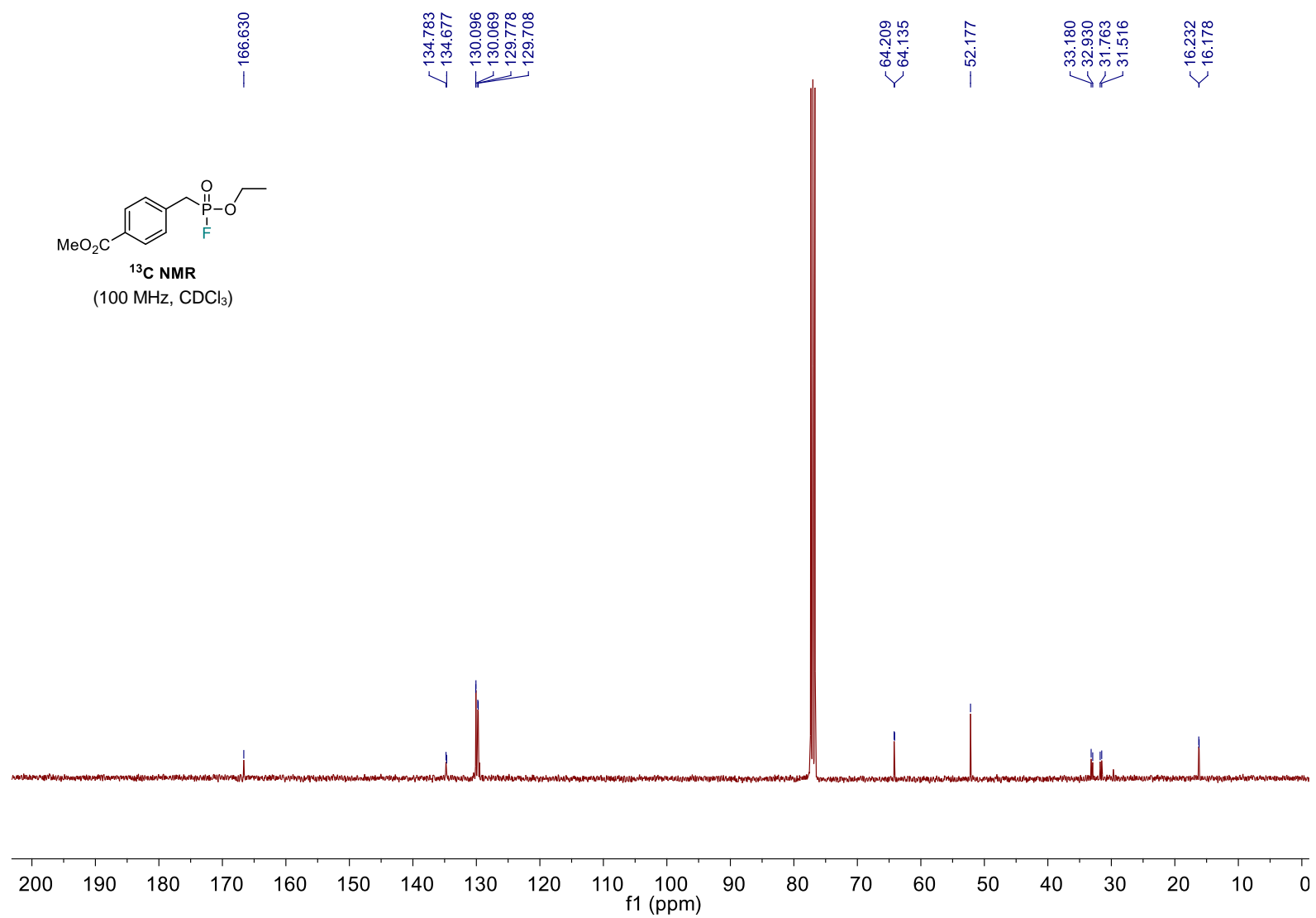
(376 MHz, CDCl₃)

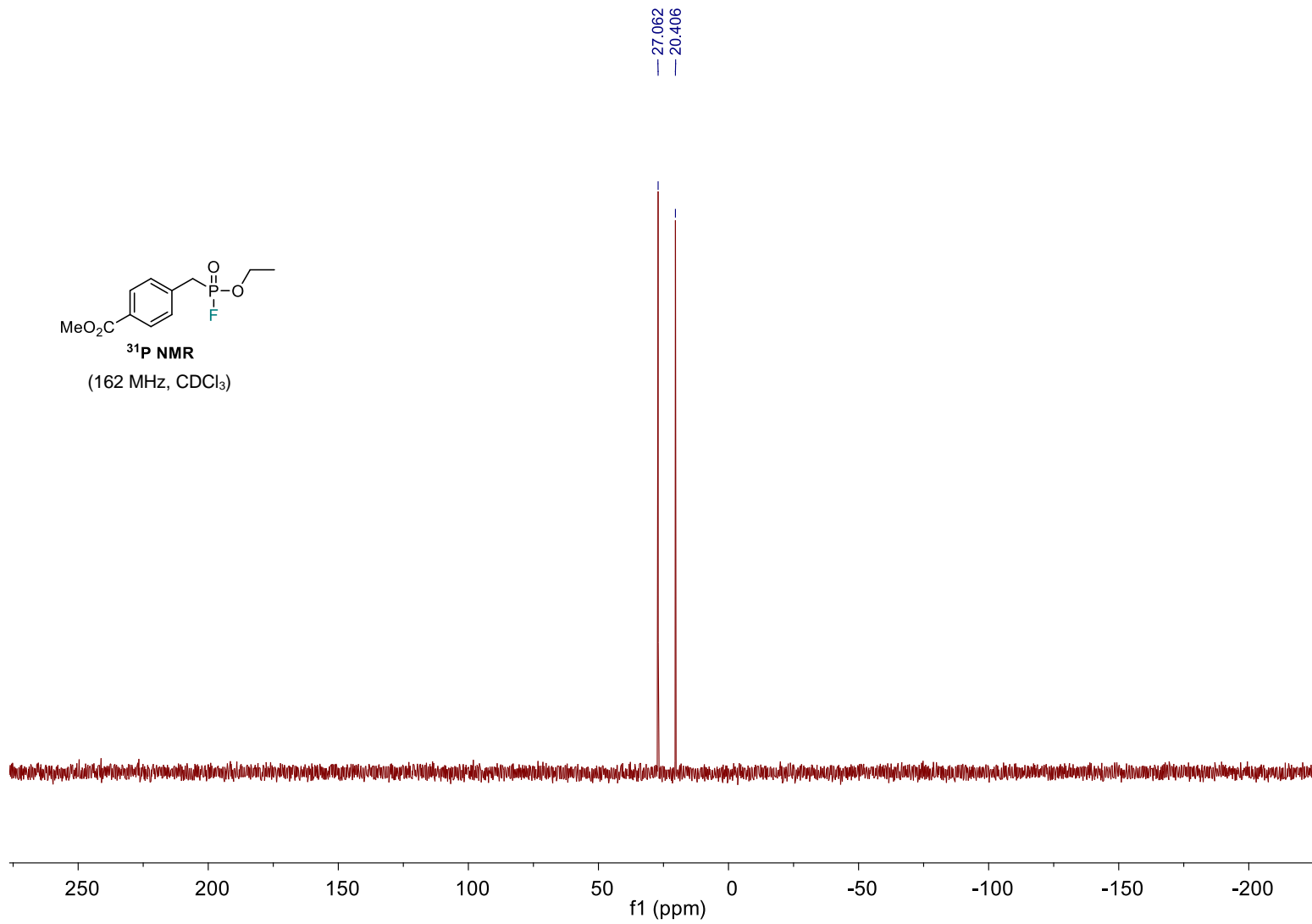
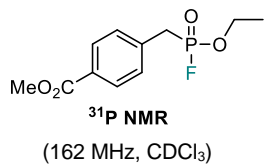


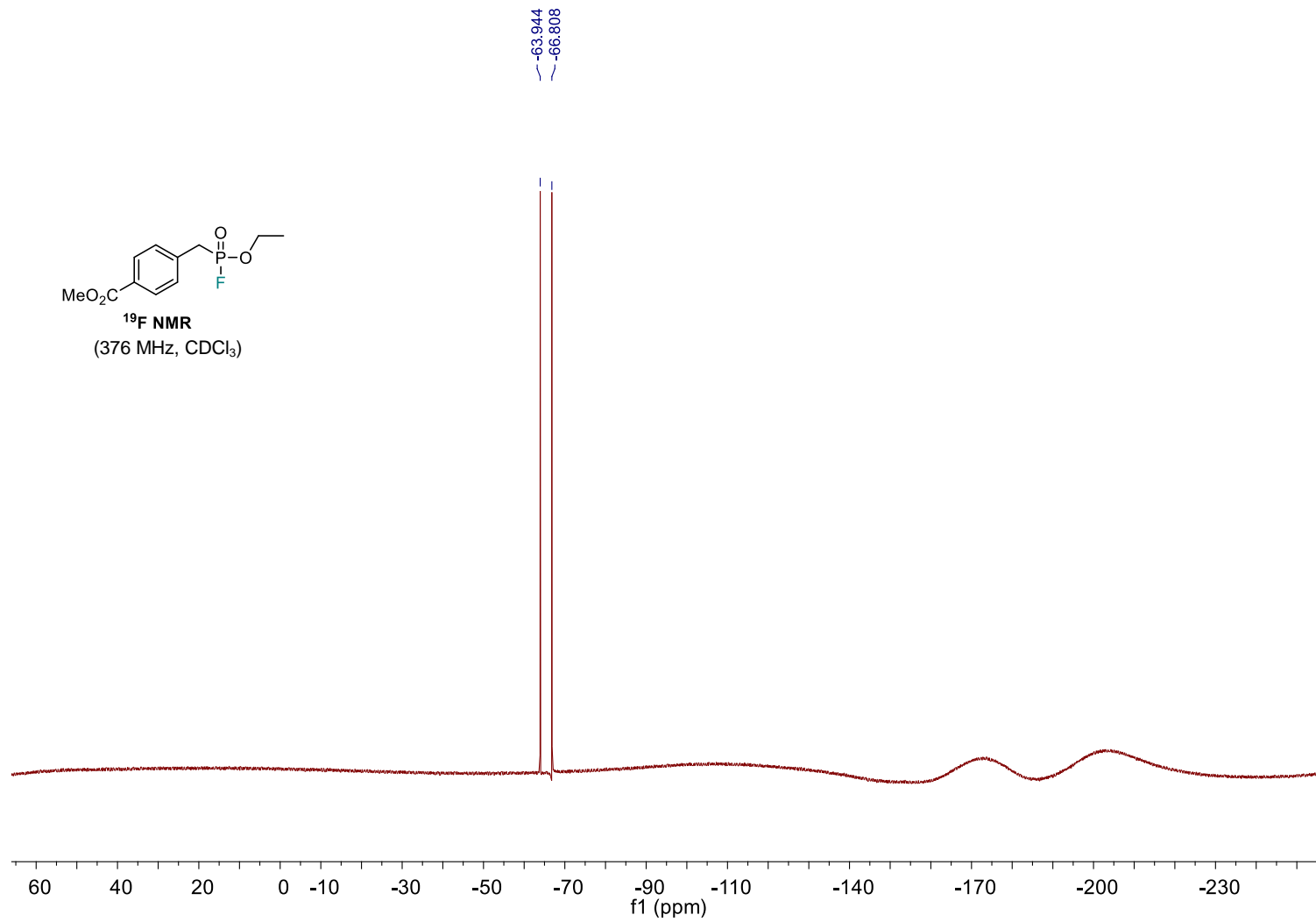


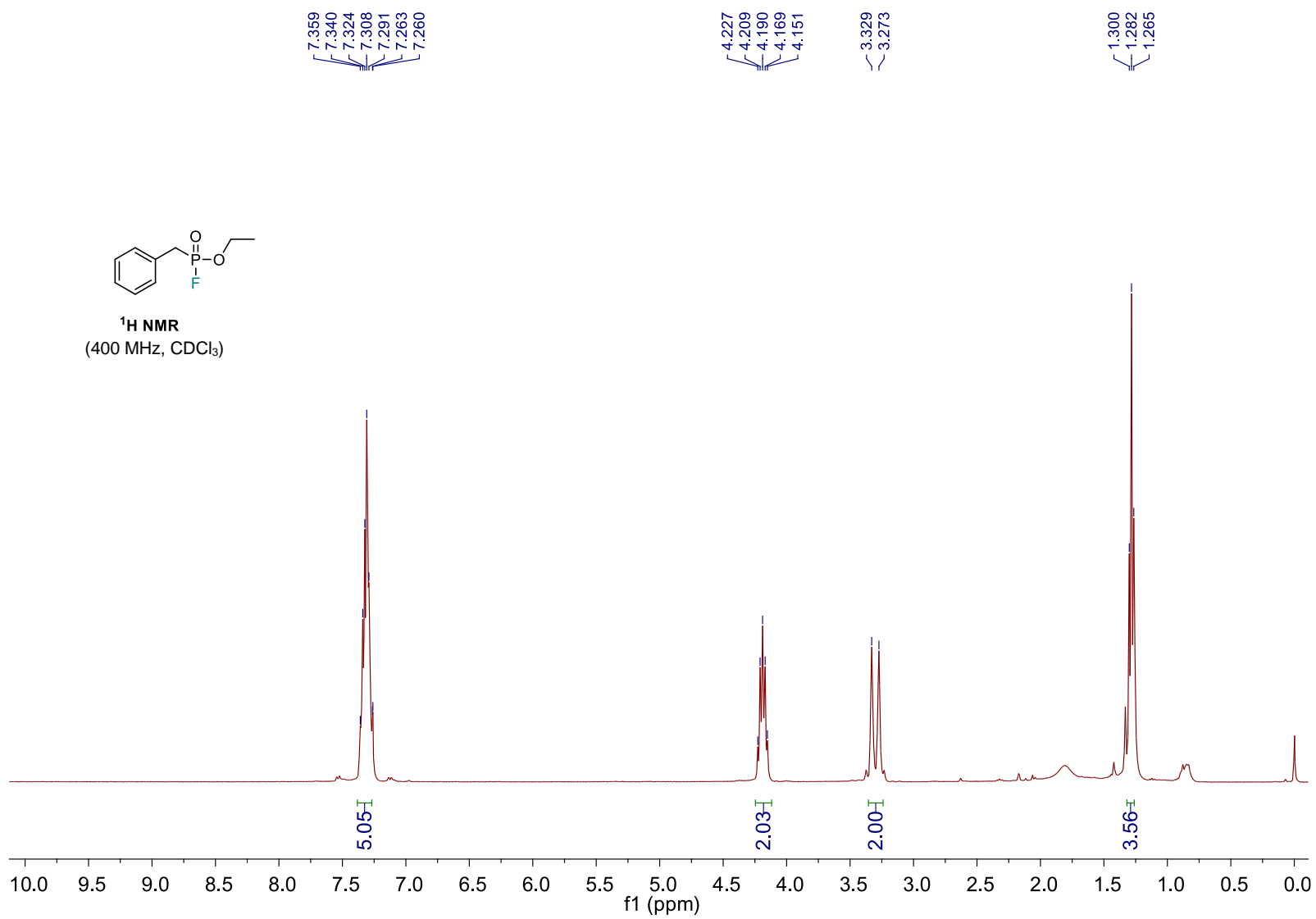
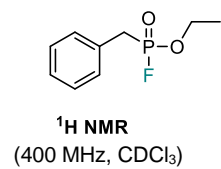


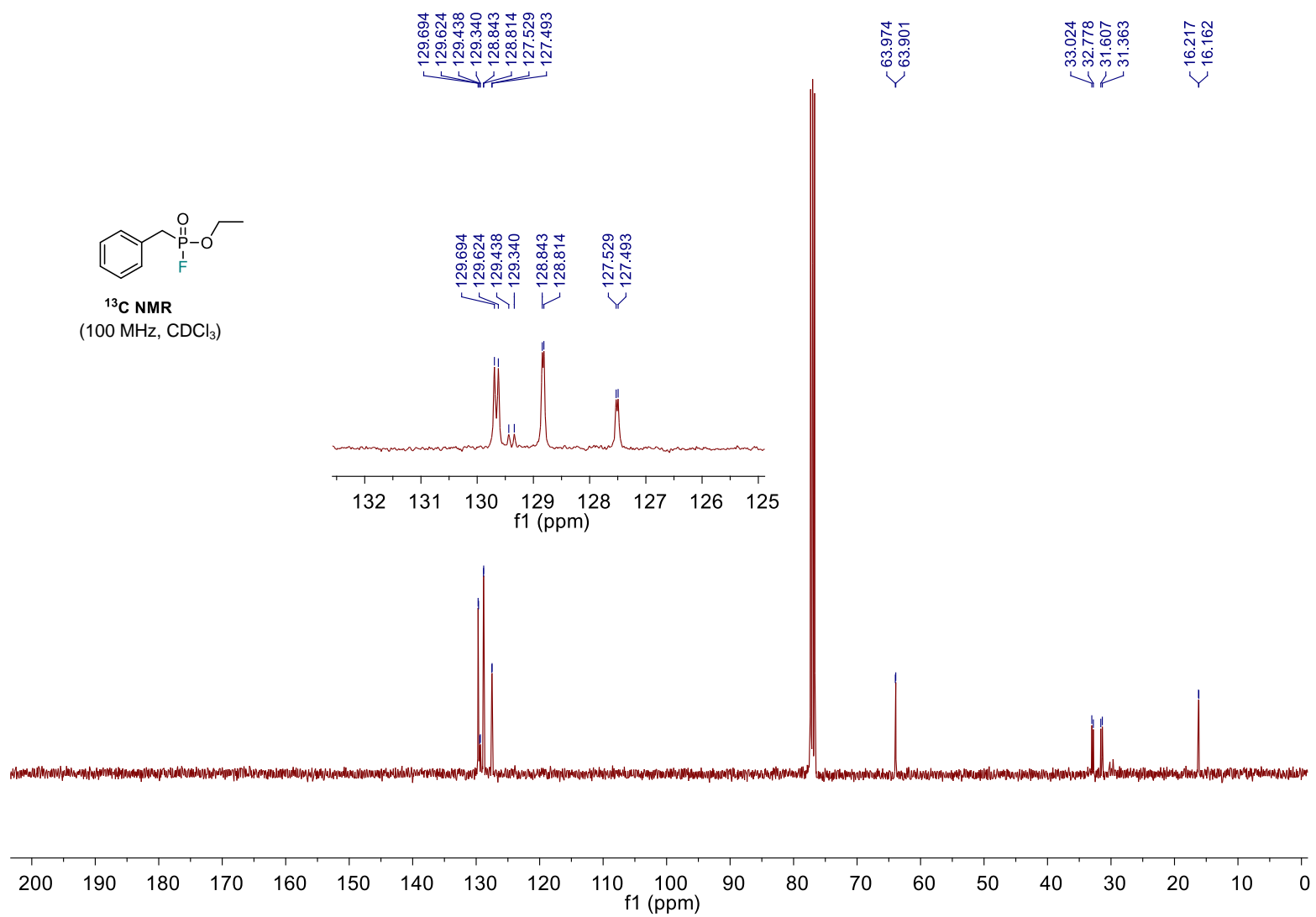
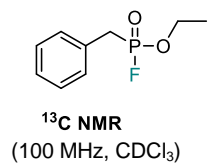
¹³C NMR
(100 MHz, CDCl₃)

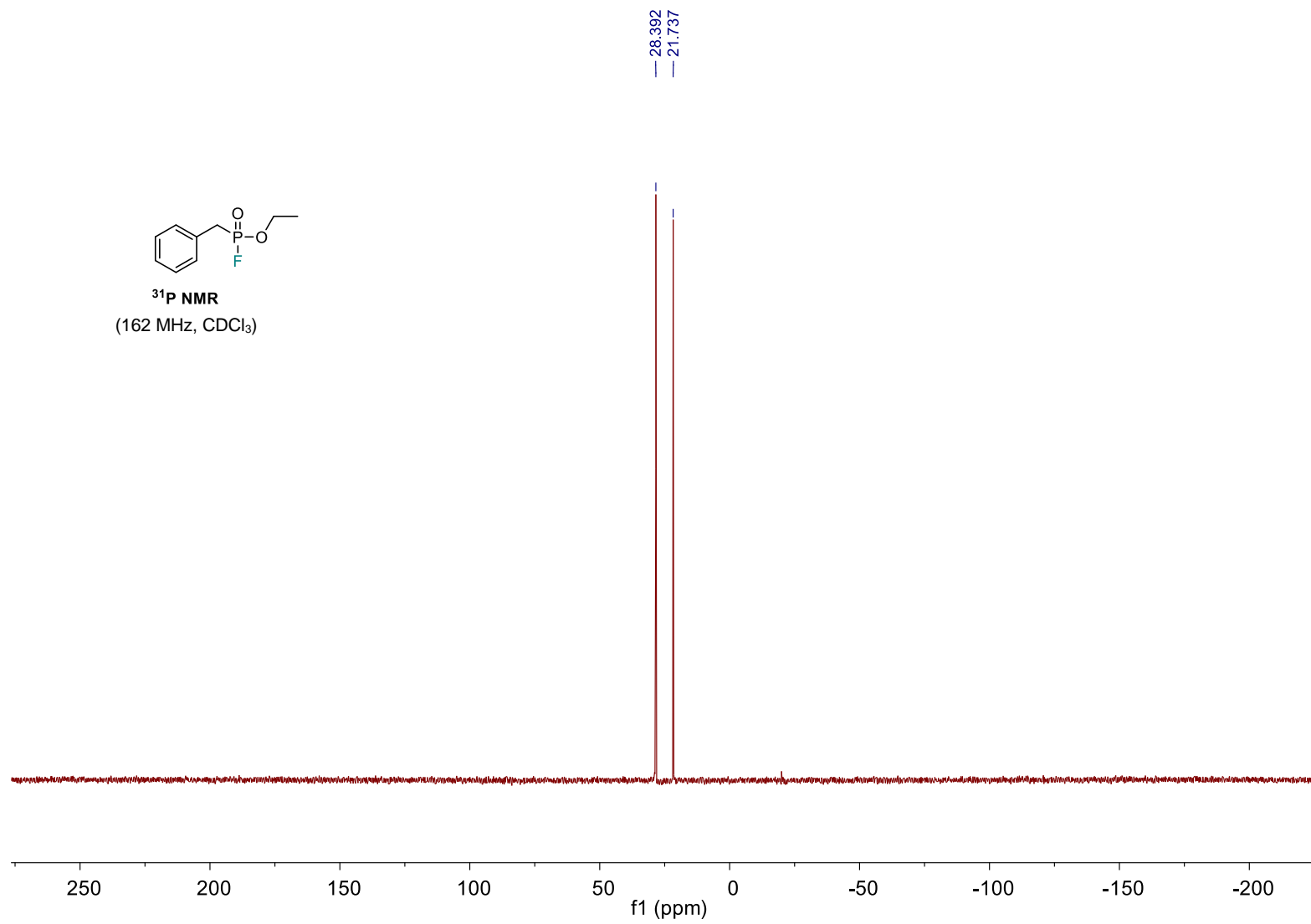
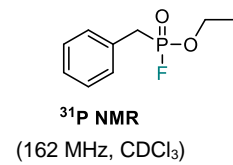




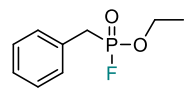




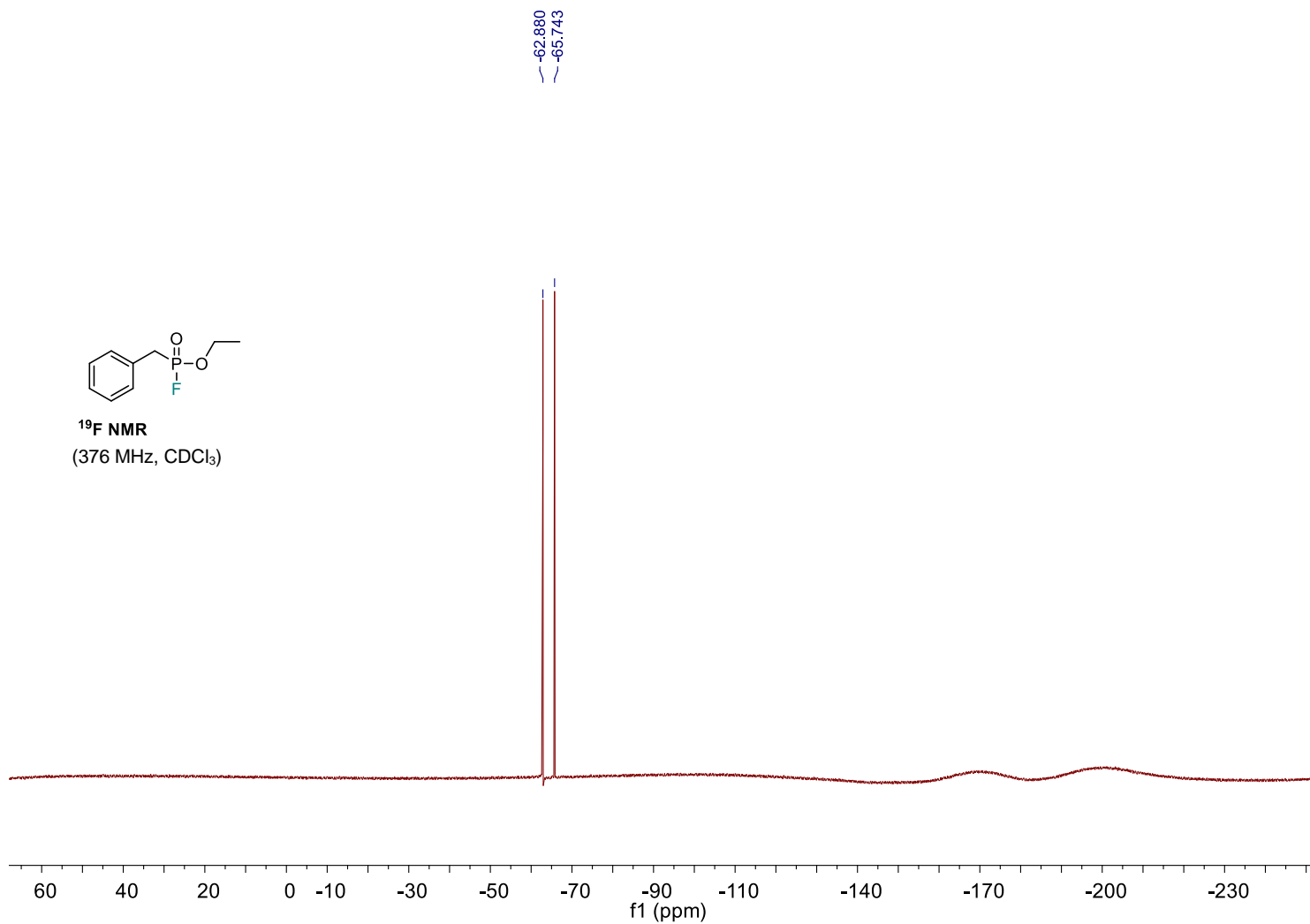


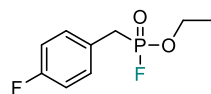


S251

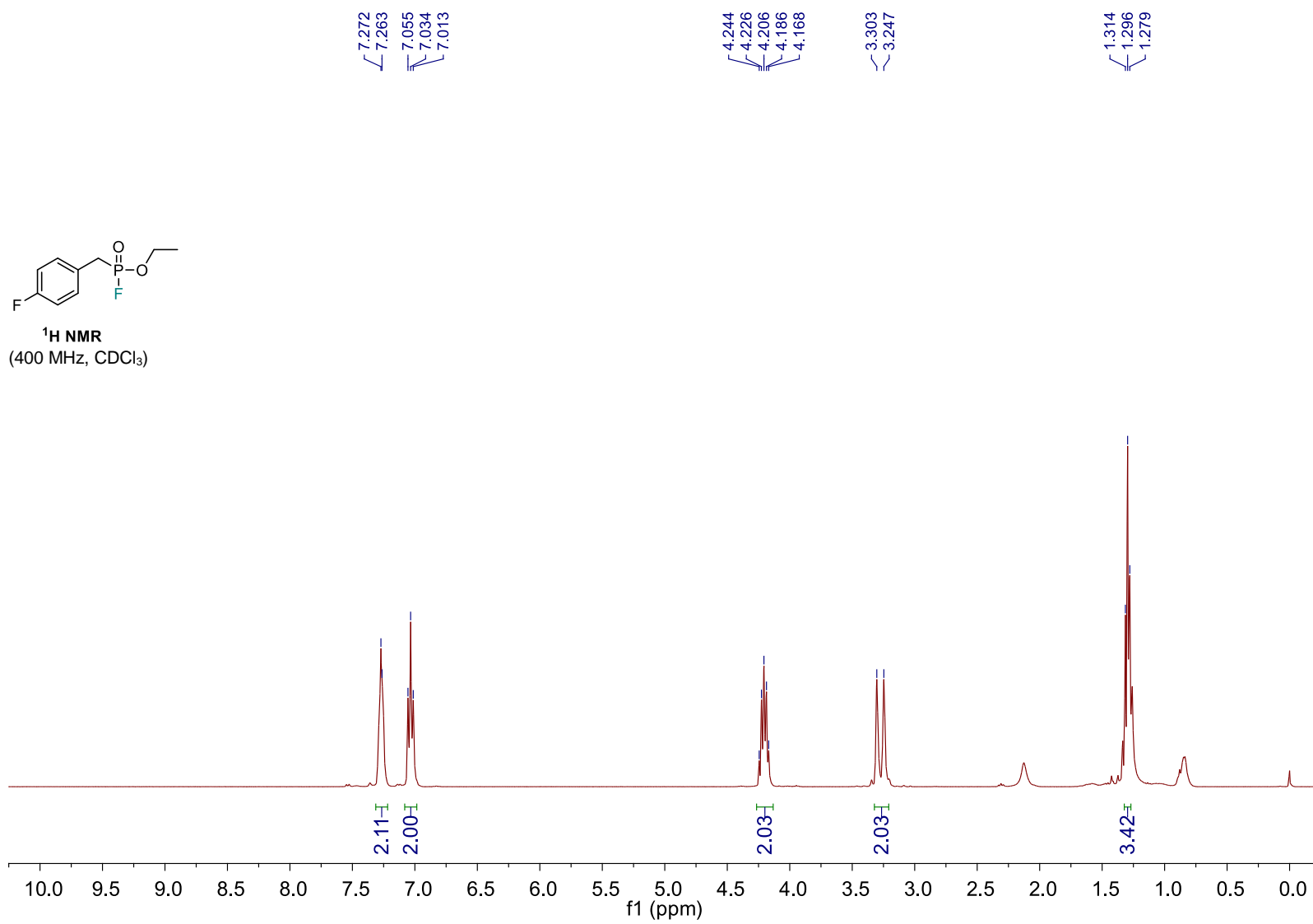


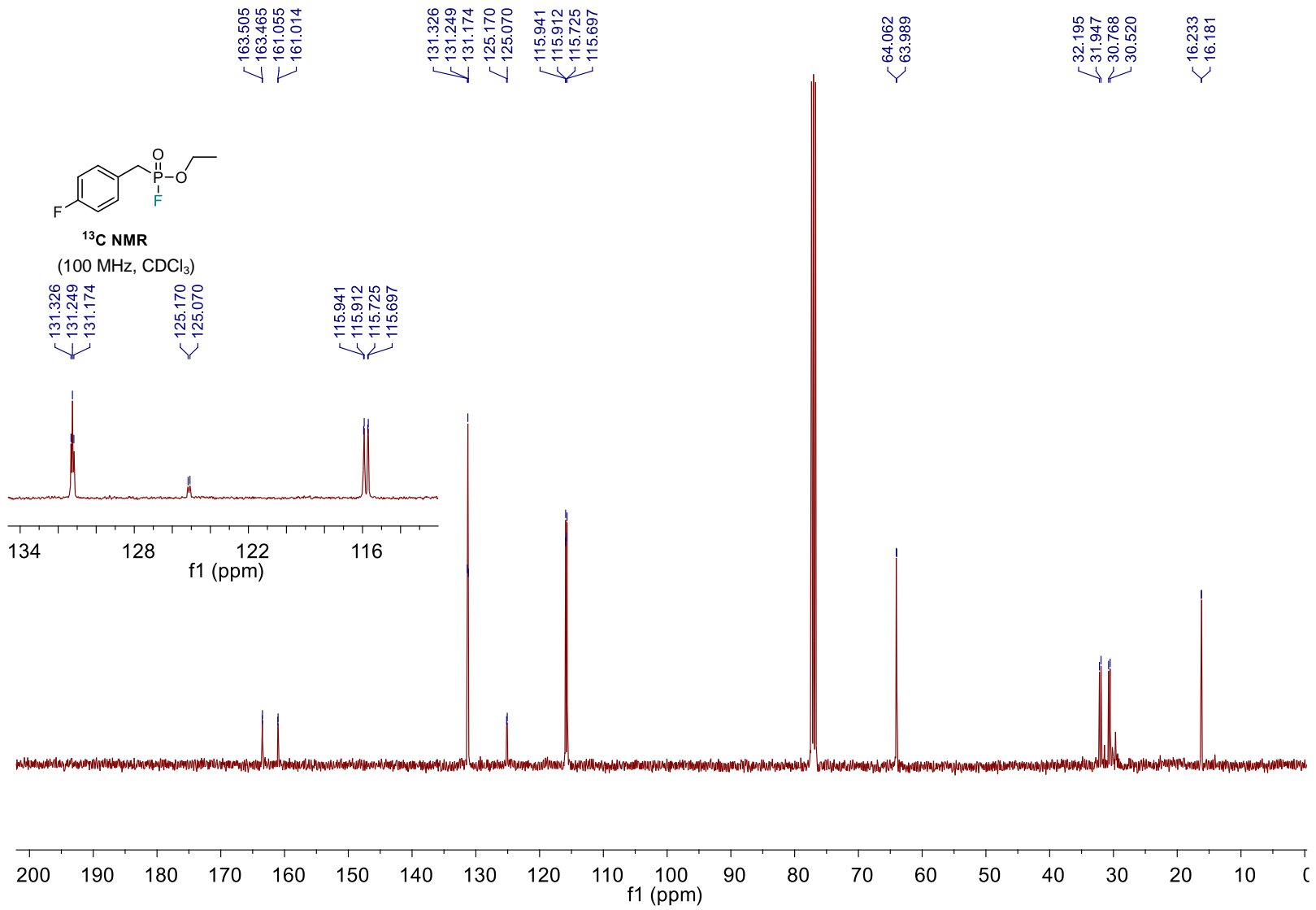
¹⁹F NMR
(376 MHz, CDCl₃)

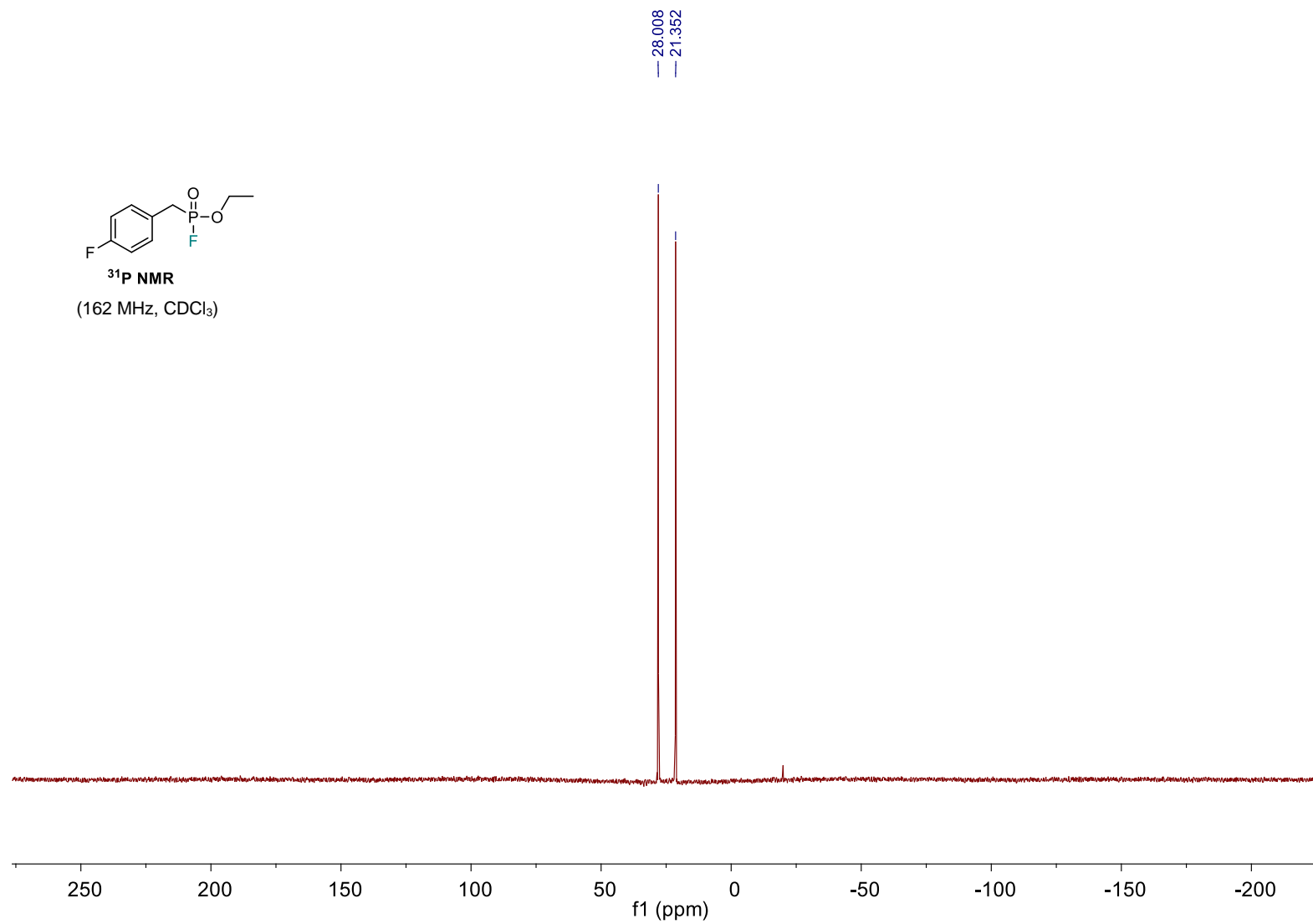
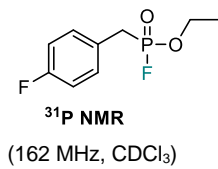




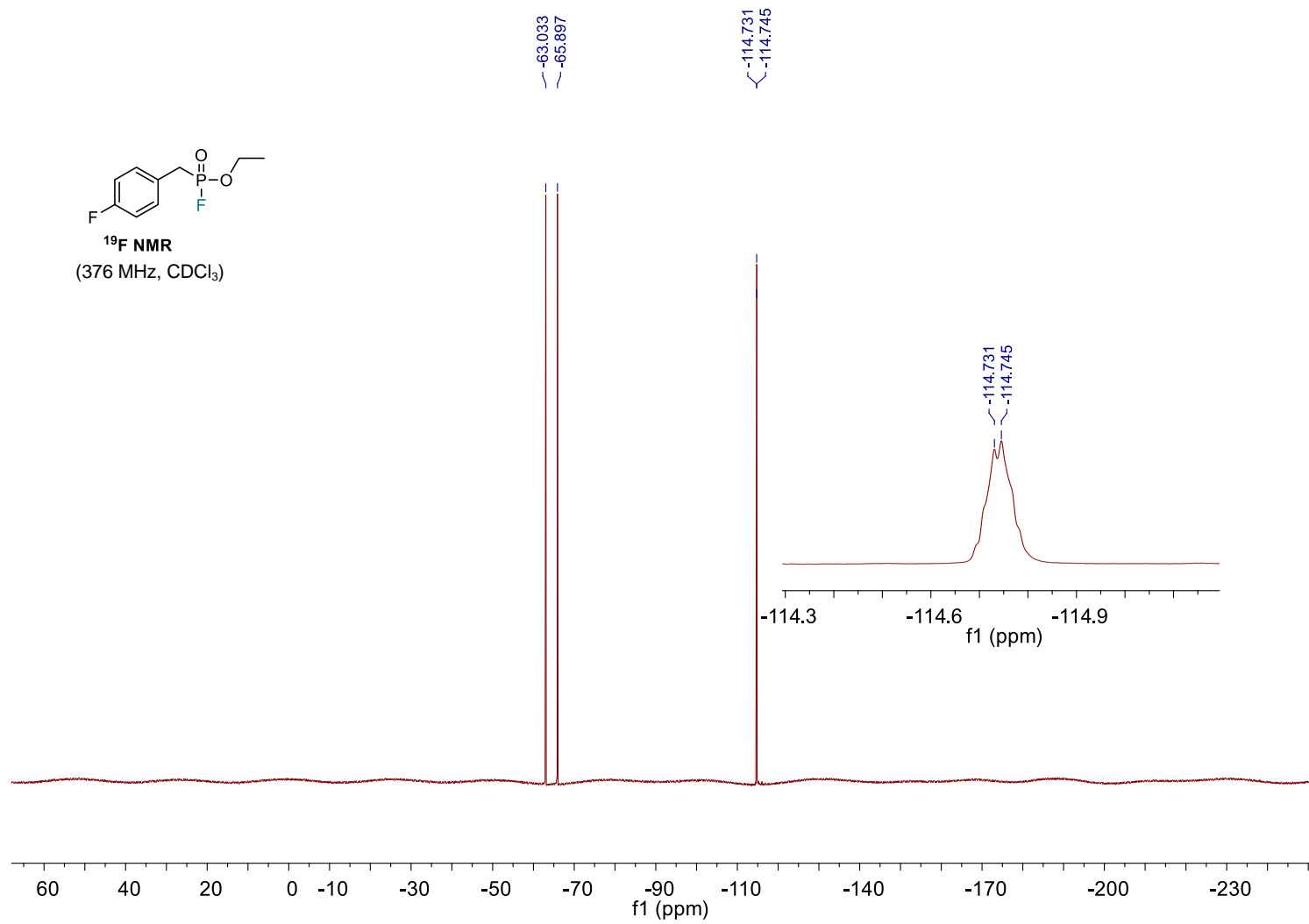
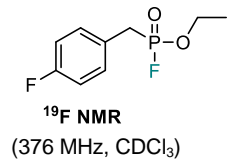
¹H NMR
(400 MHz, CDCl₃)

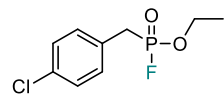




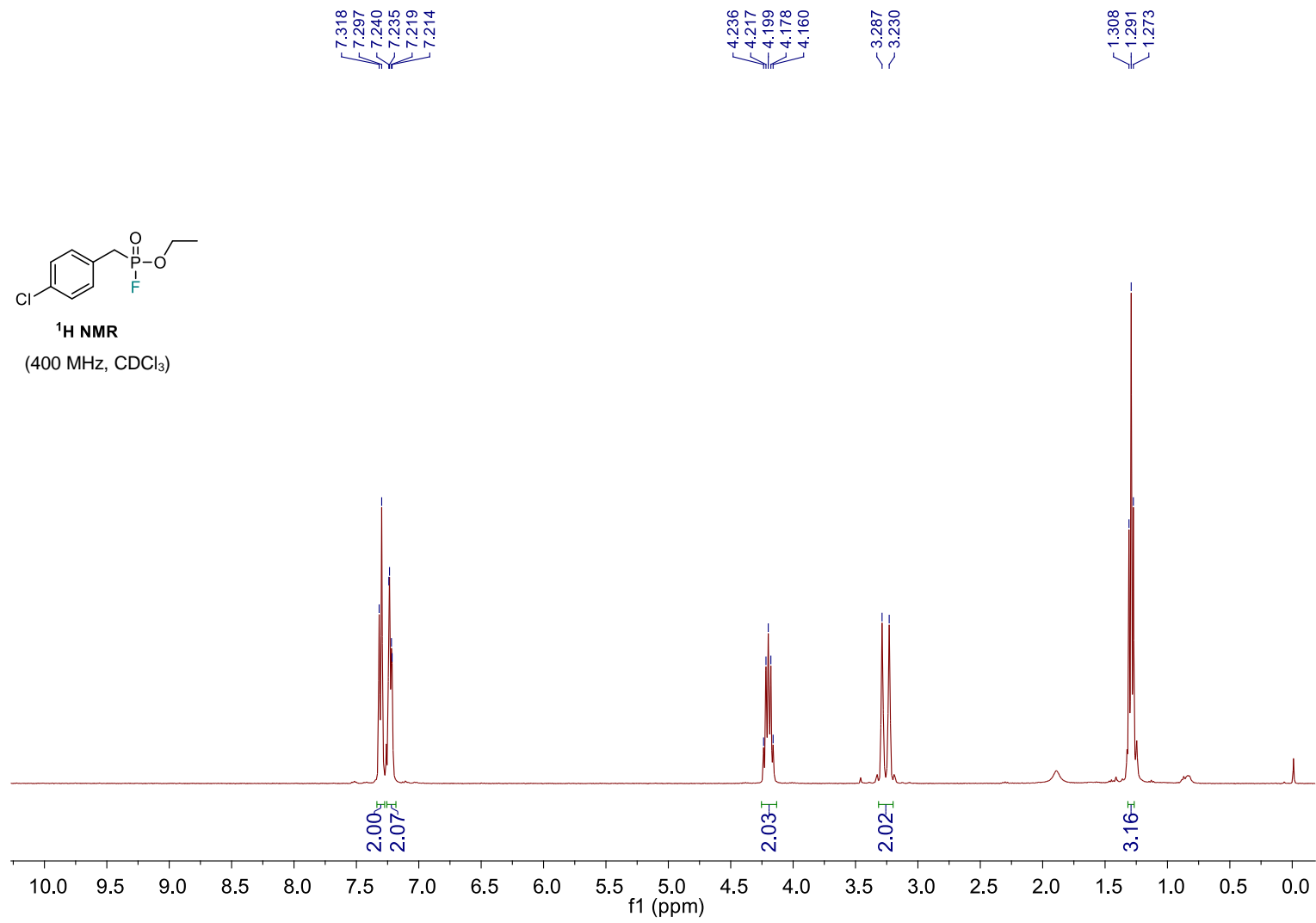


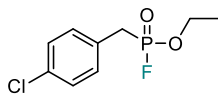
S255



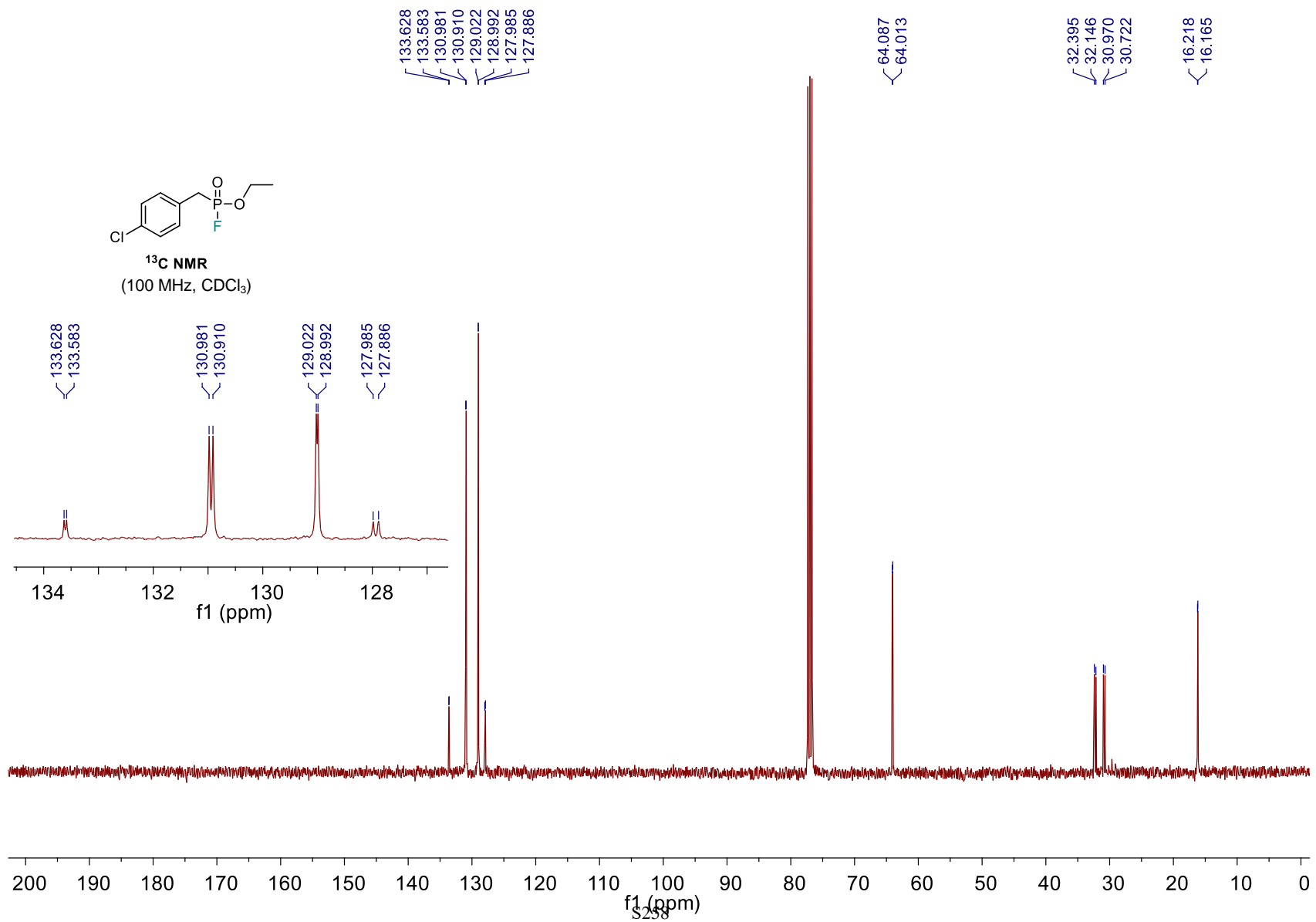


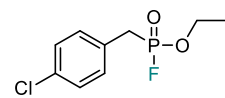
¹H NMR
(400 MHz, CDCl₃)



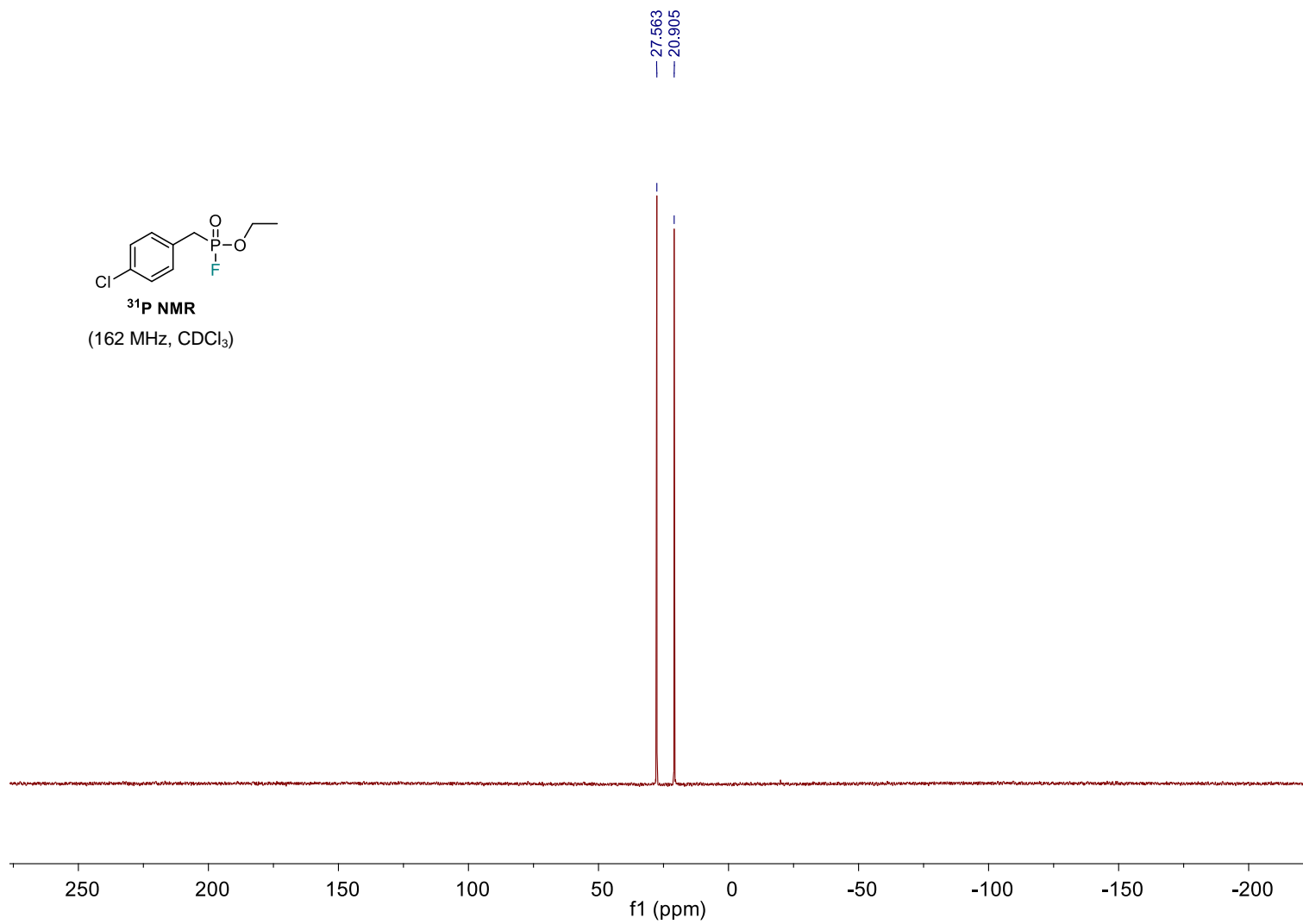


¹³C NMR
(100 MHz, CDCl₃)

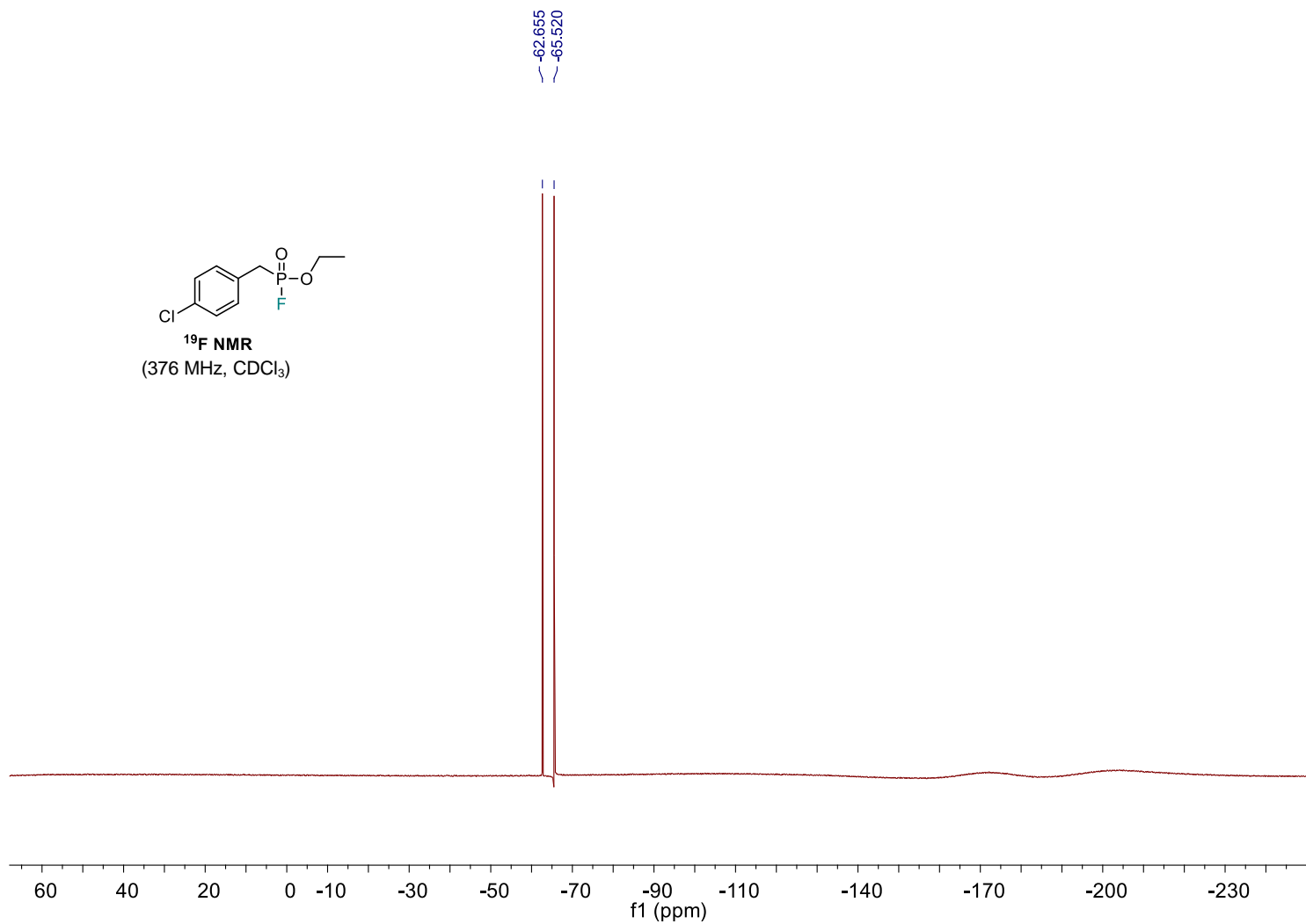
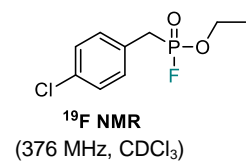


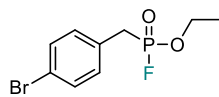


³¹P NMR
(162 MHz, CDCl₃)

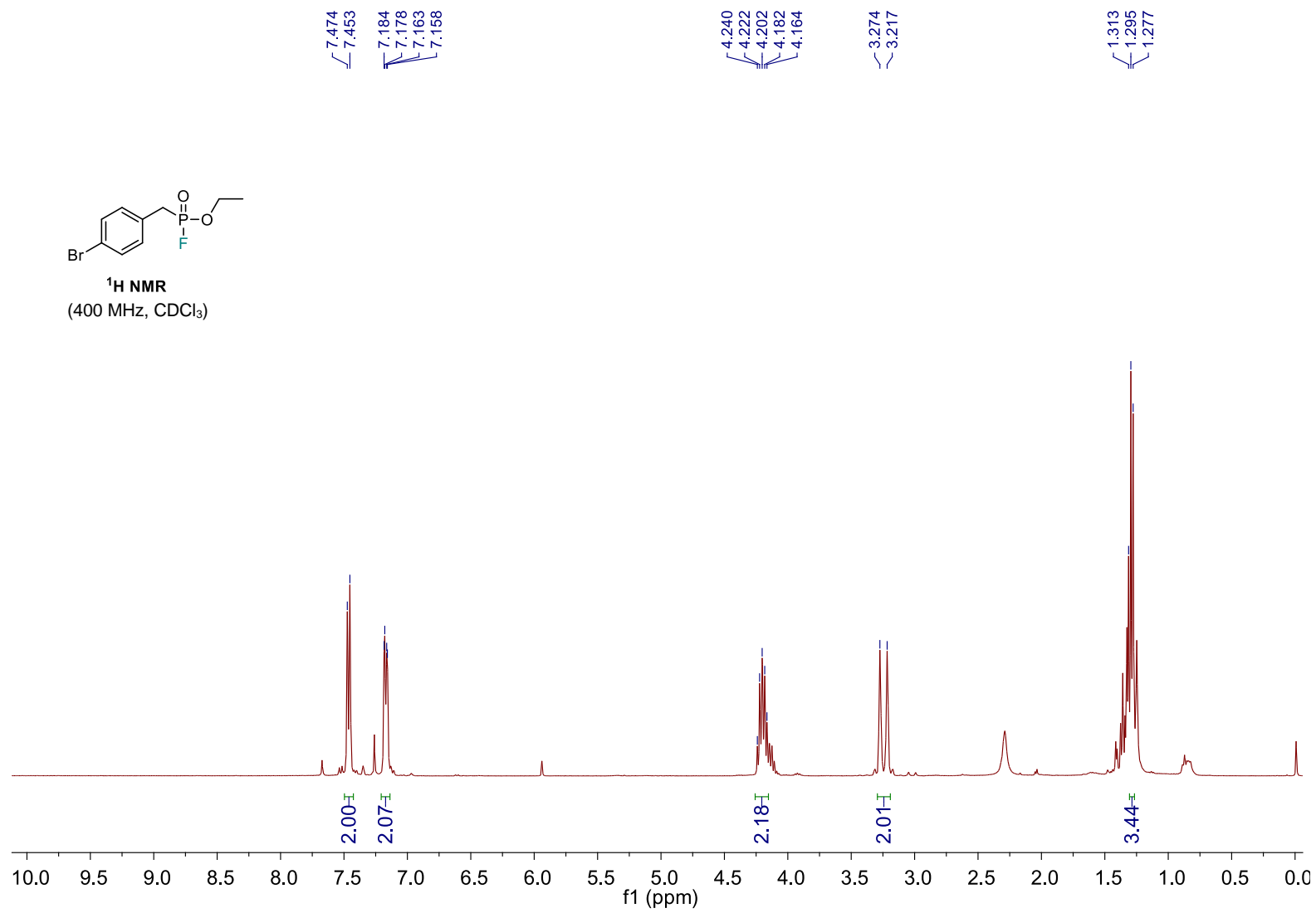


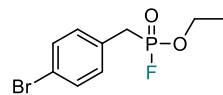
S259





¹H NMR
(400 MHz, CDCl₃)





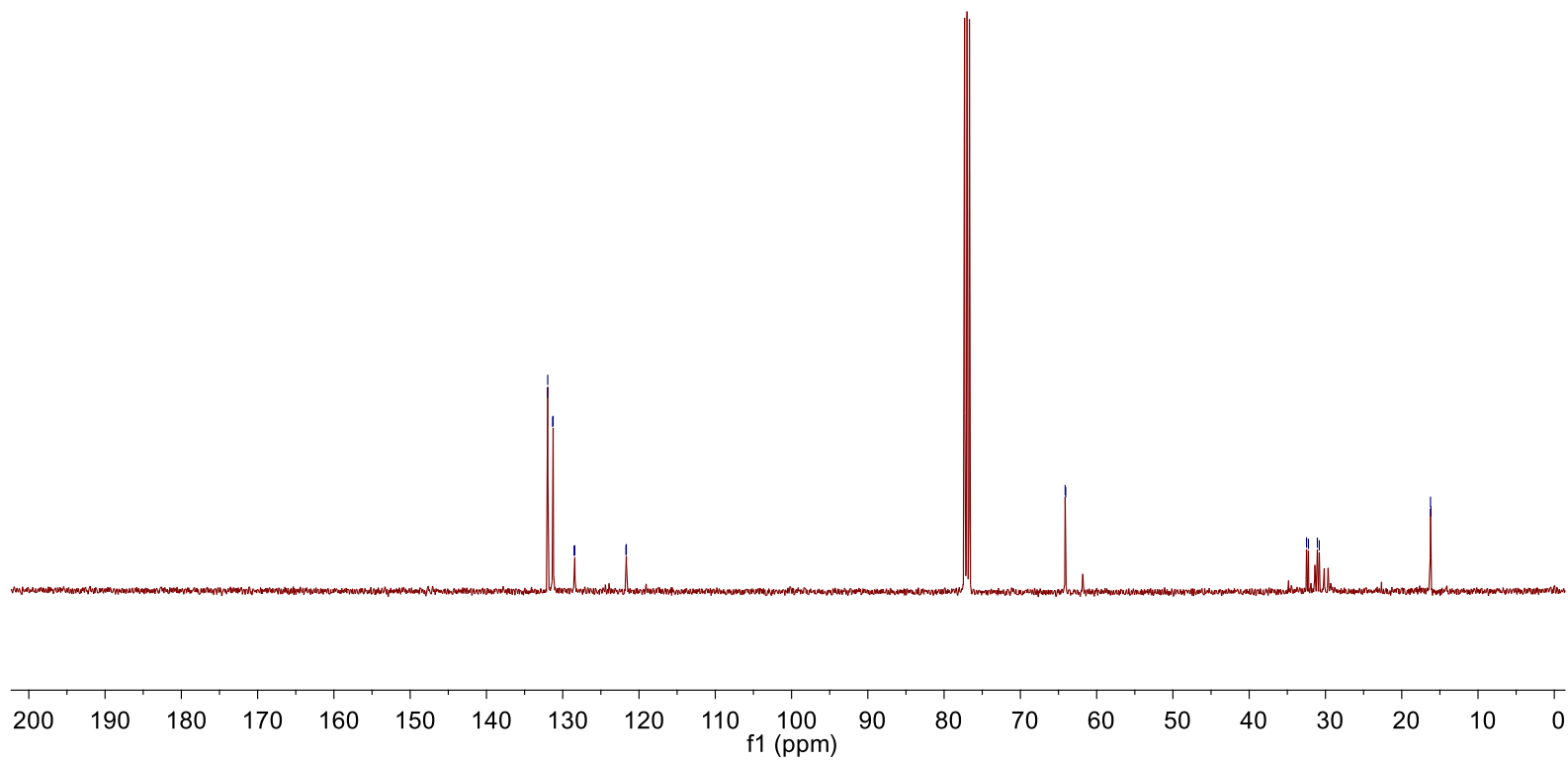
¹³C NMR
(100 MHz, CDCl₃)

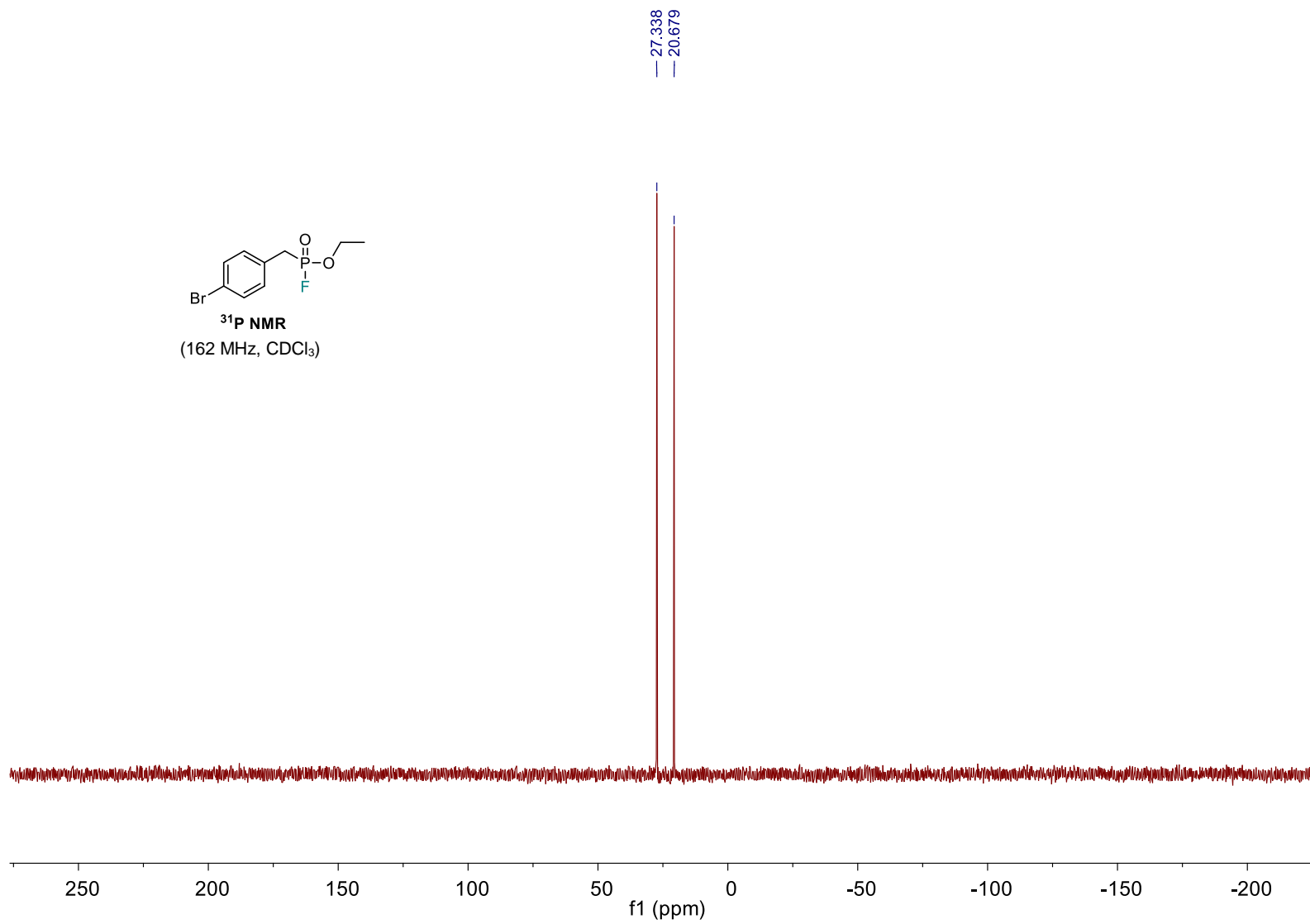
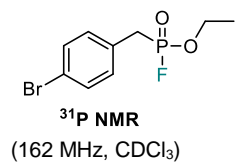
131.995
131.967
131.327
131.256
128.511
128.413
121.709
121.661

64.130
64.057

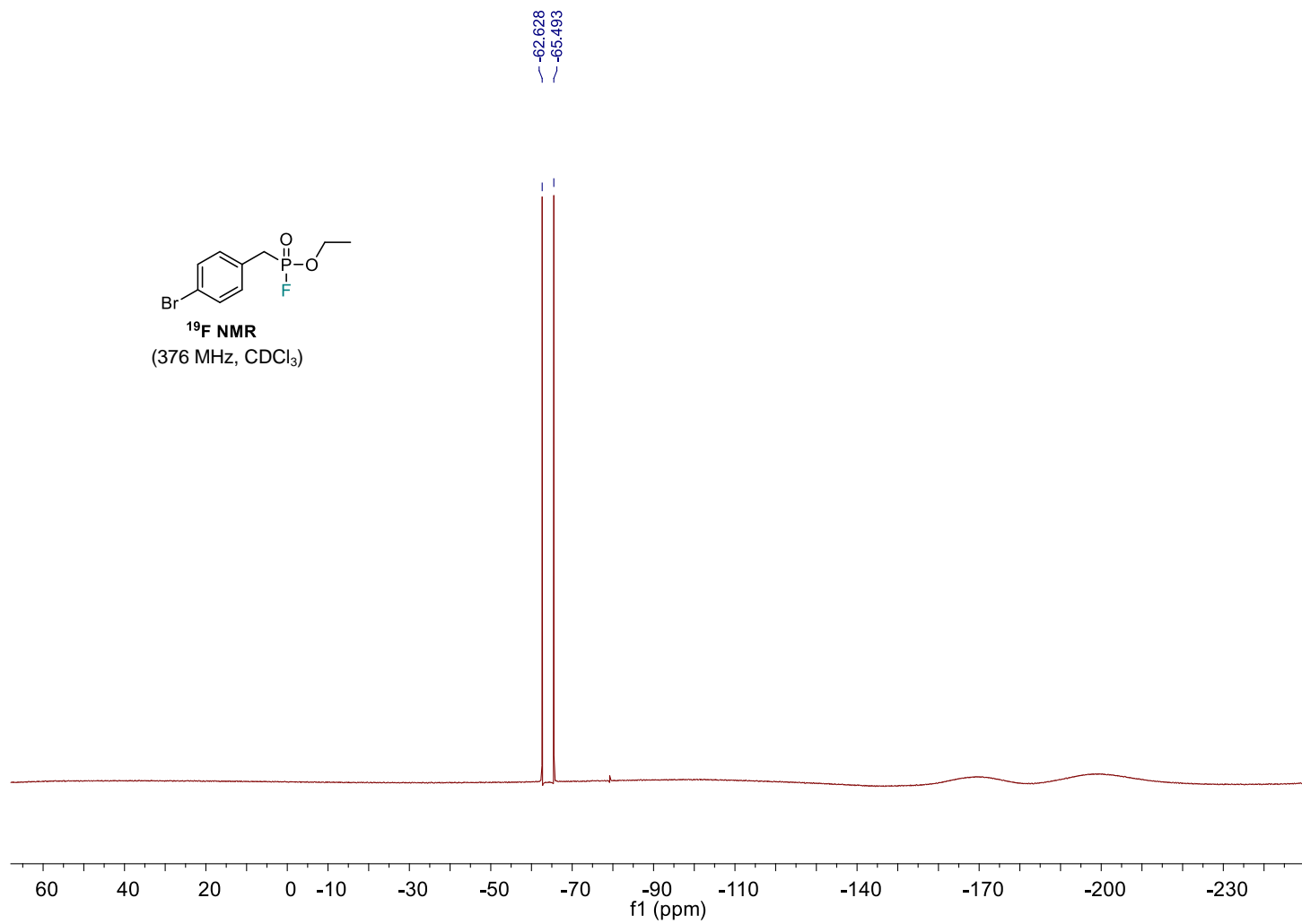
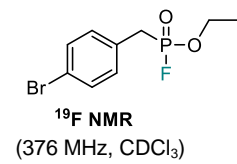
32.484
32.236
31.061
30.813

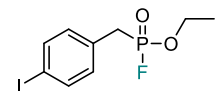
16.232
16.177



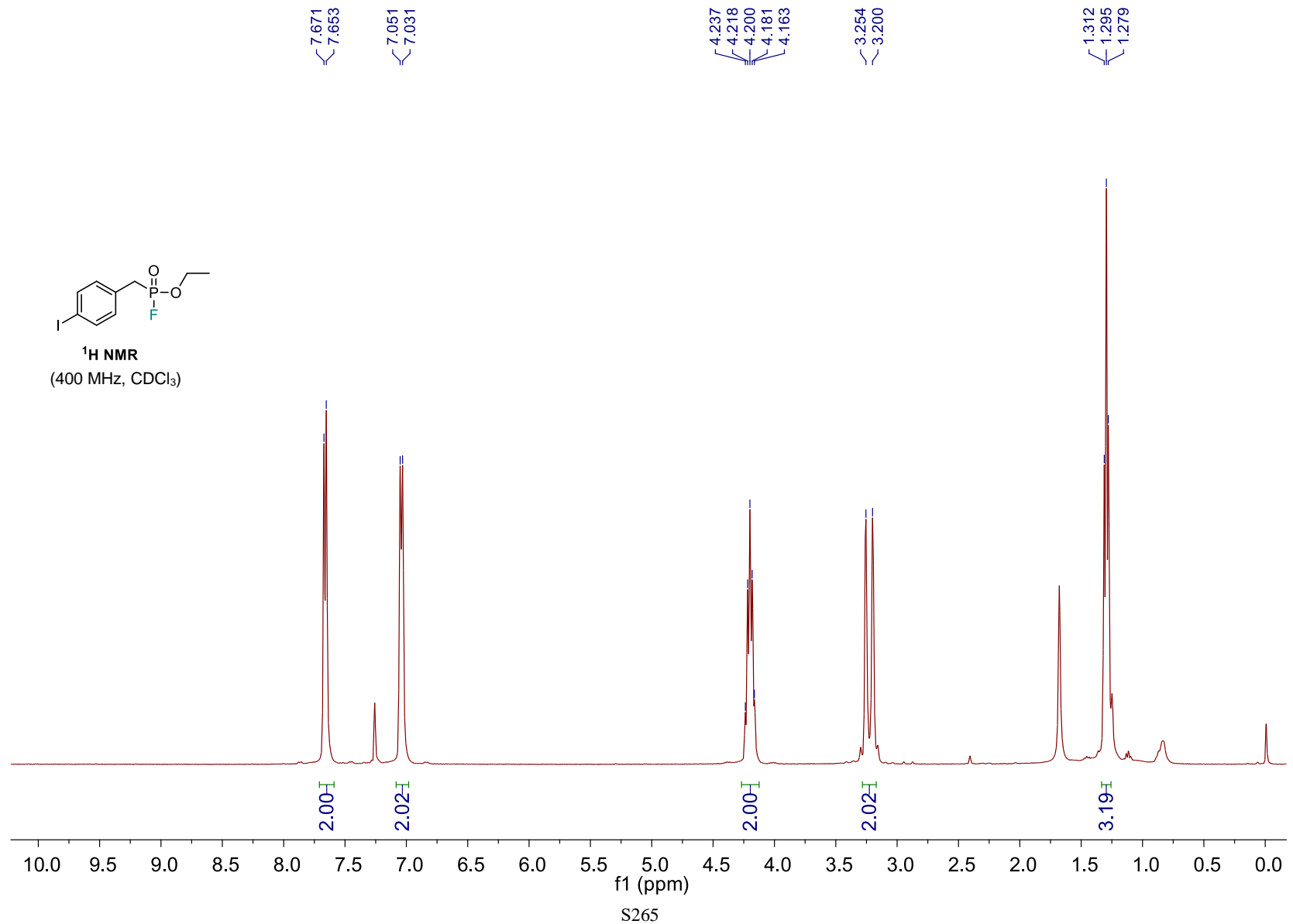


S263

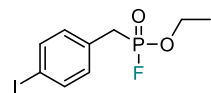




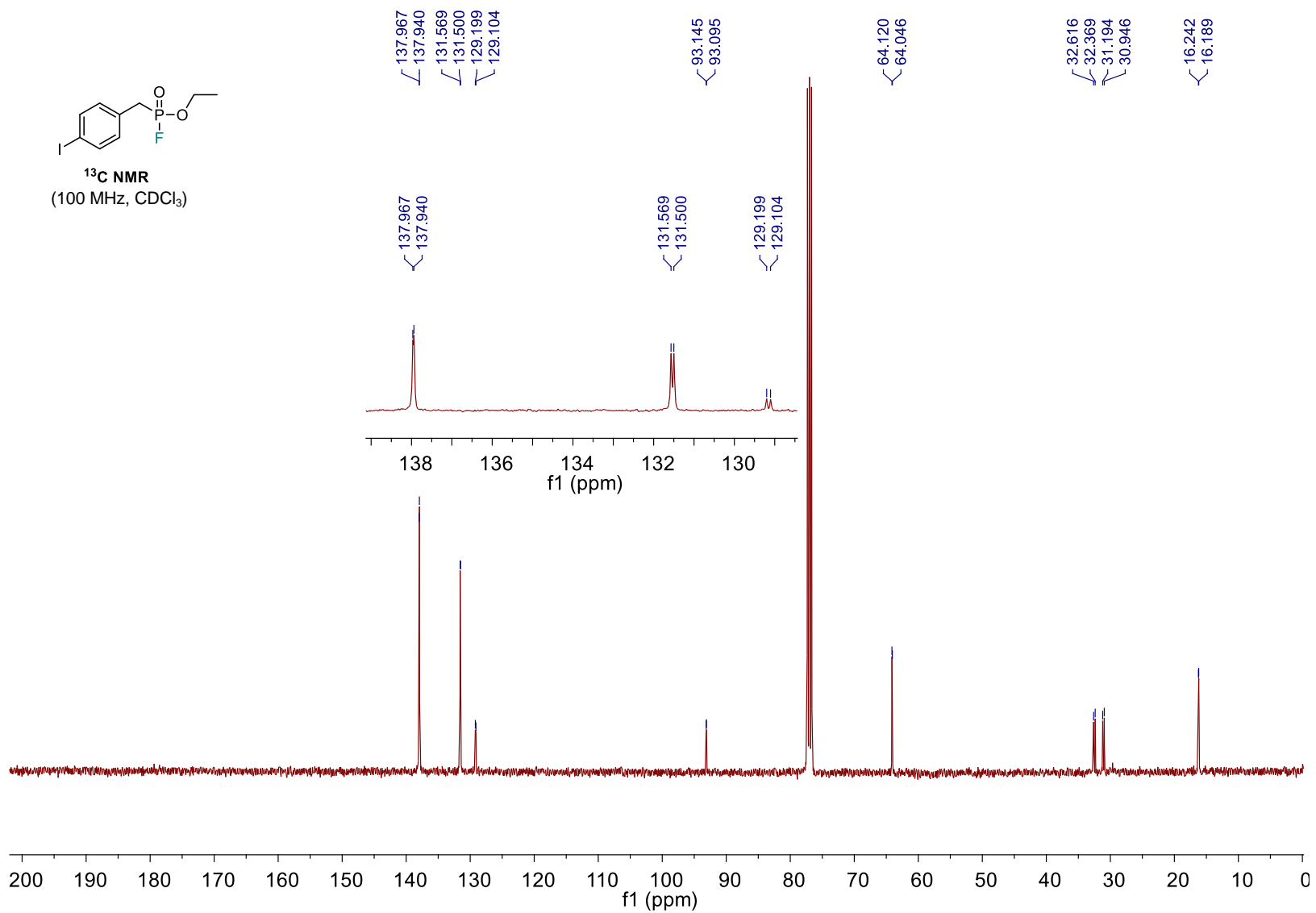
¹H NMR
(400 MHz, CDCl₃)

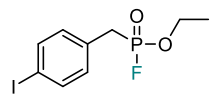


S265

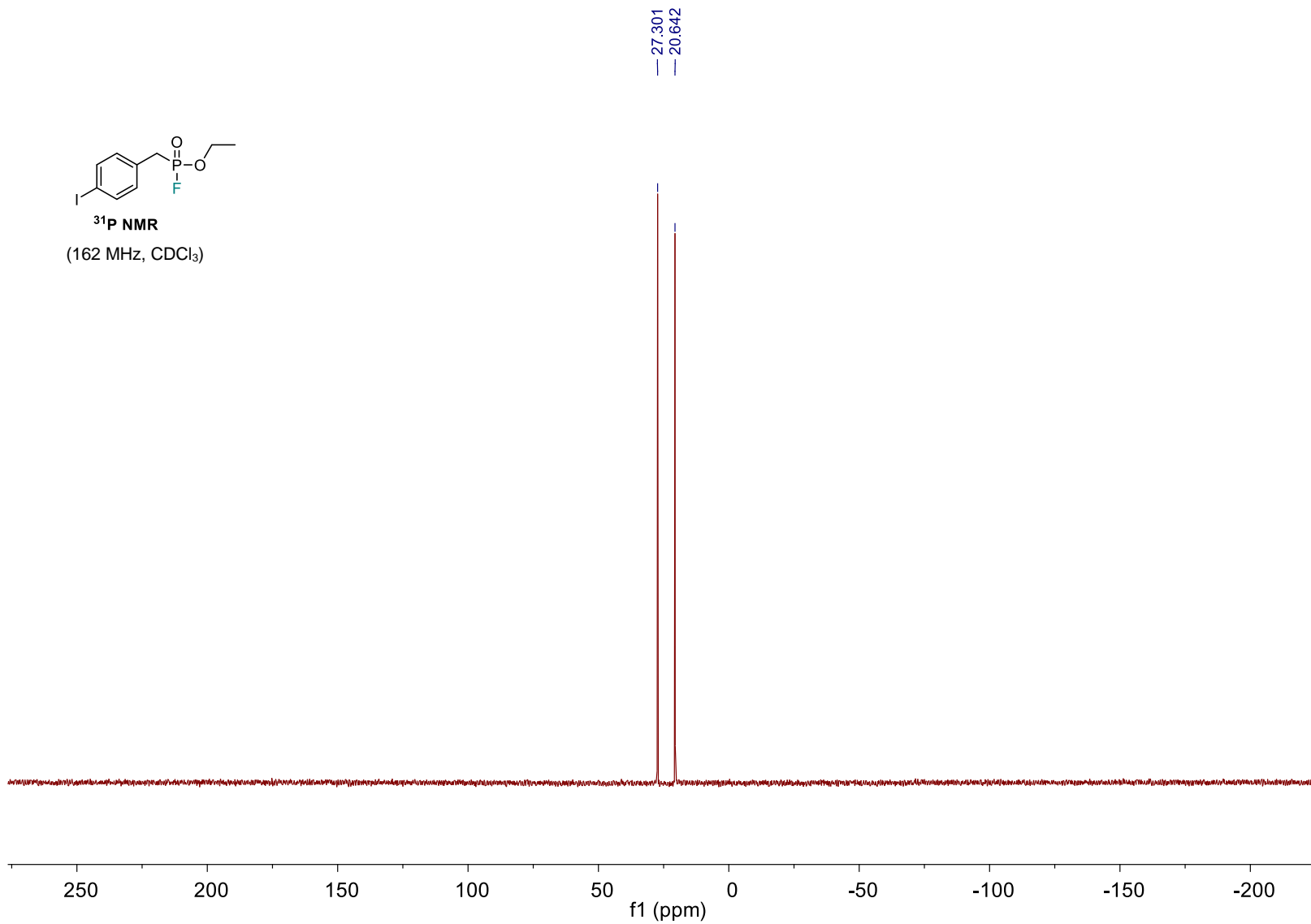


¹³C NMR
(100 MHz, CDCl₃)

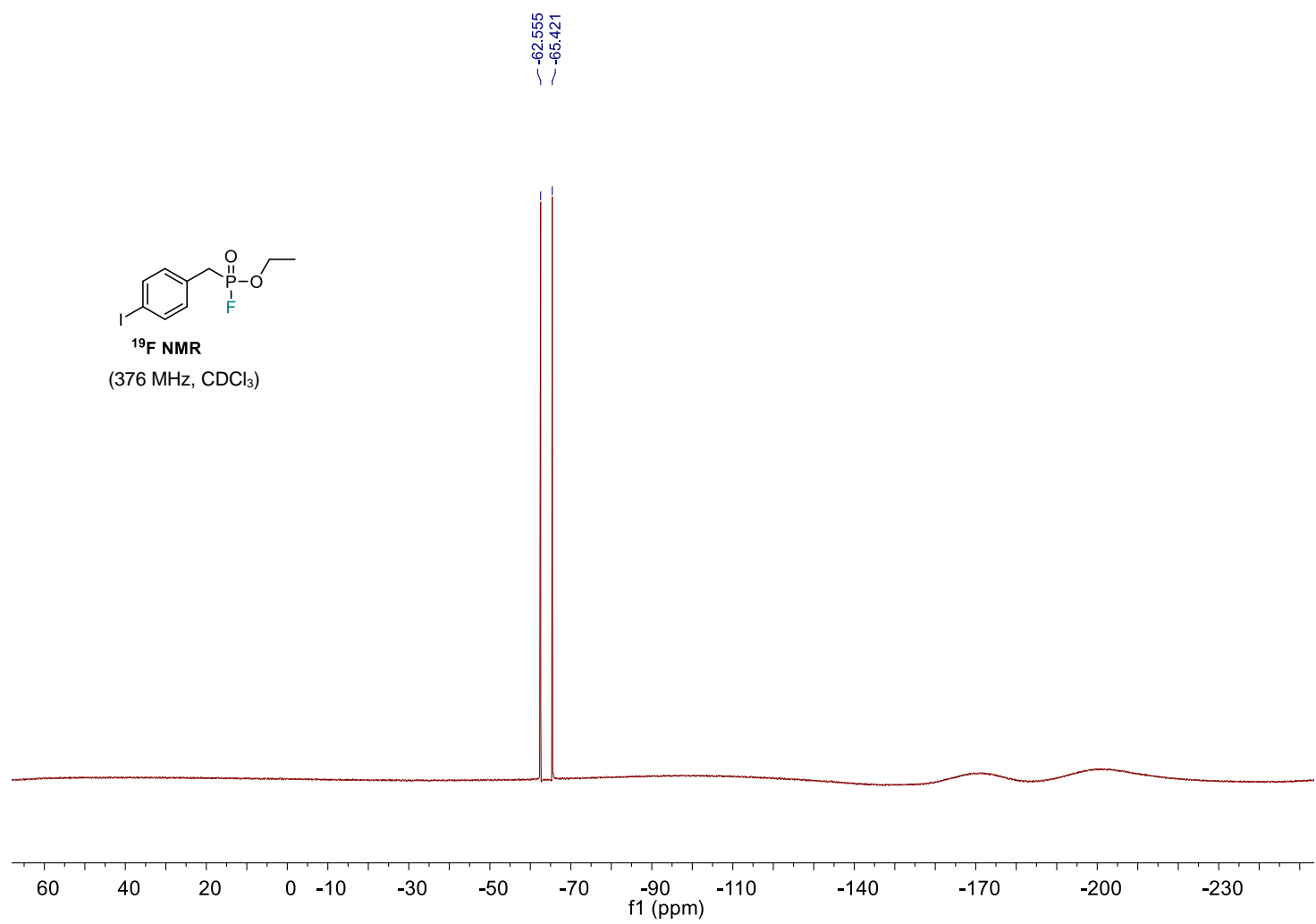
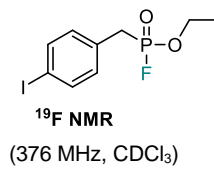


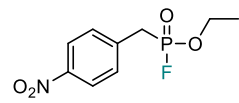


³¹P NMR
(162 MHz, CDCl₃)

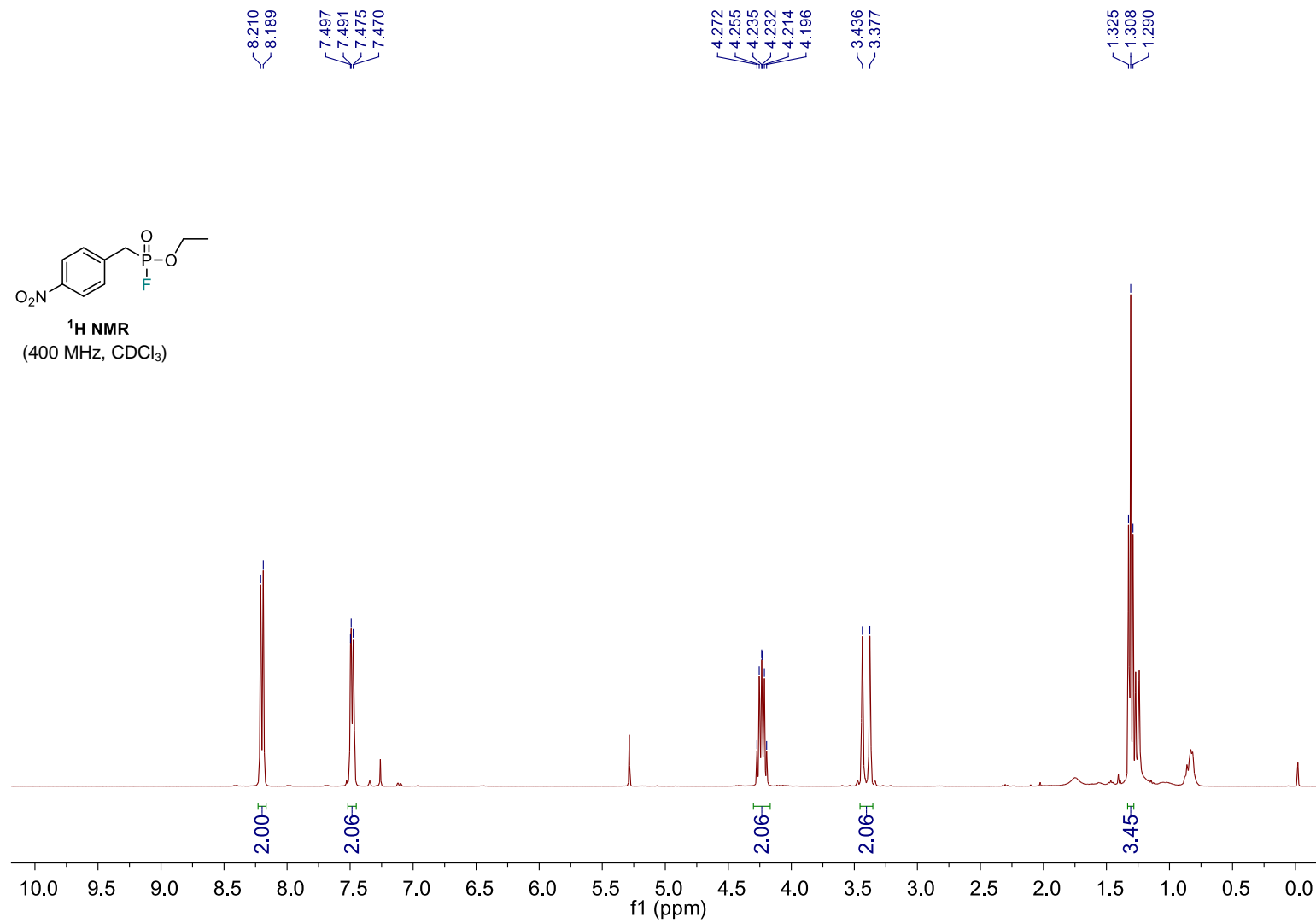


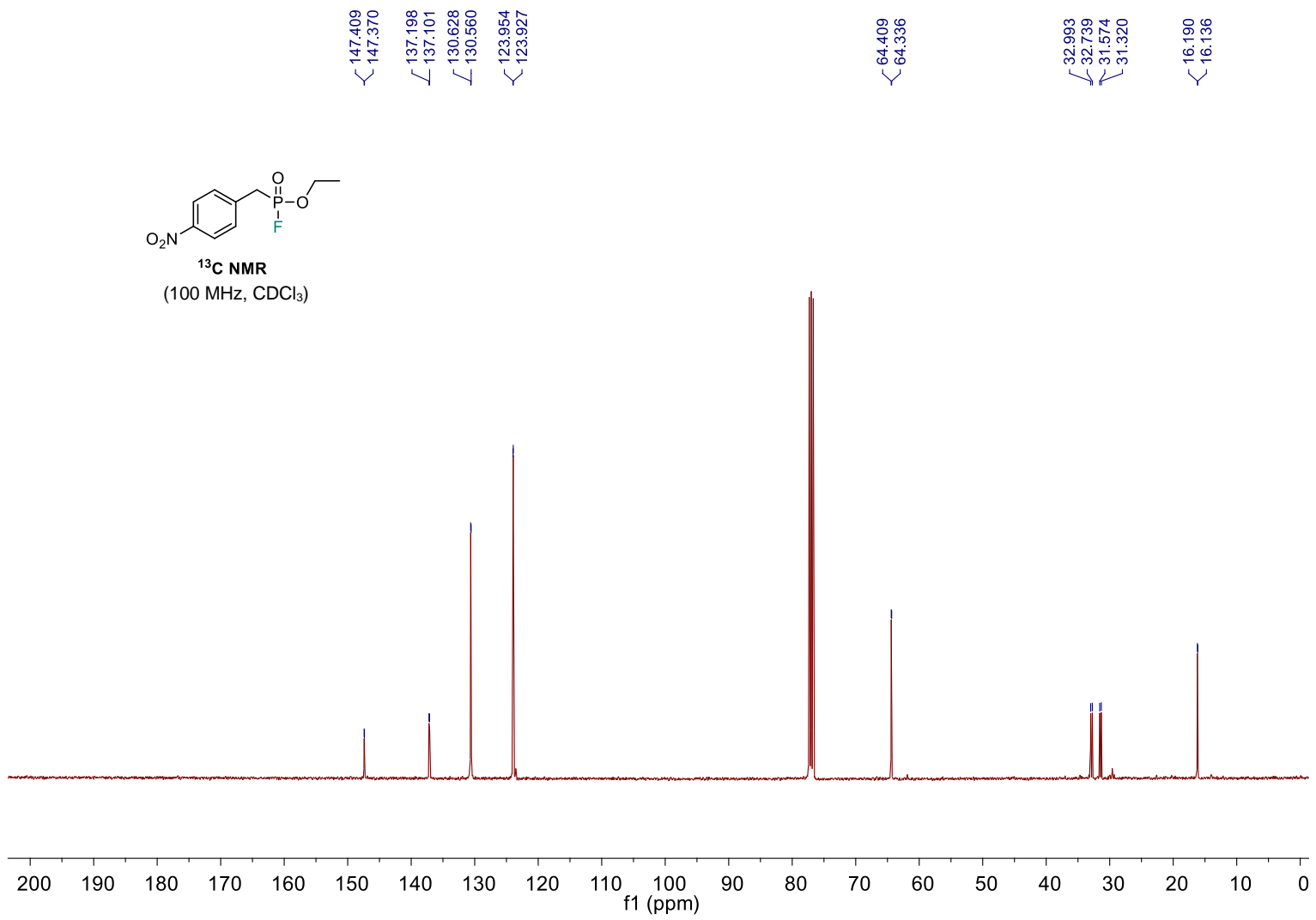
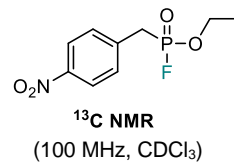
S267

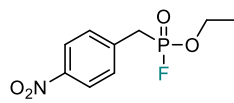




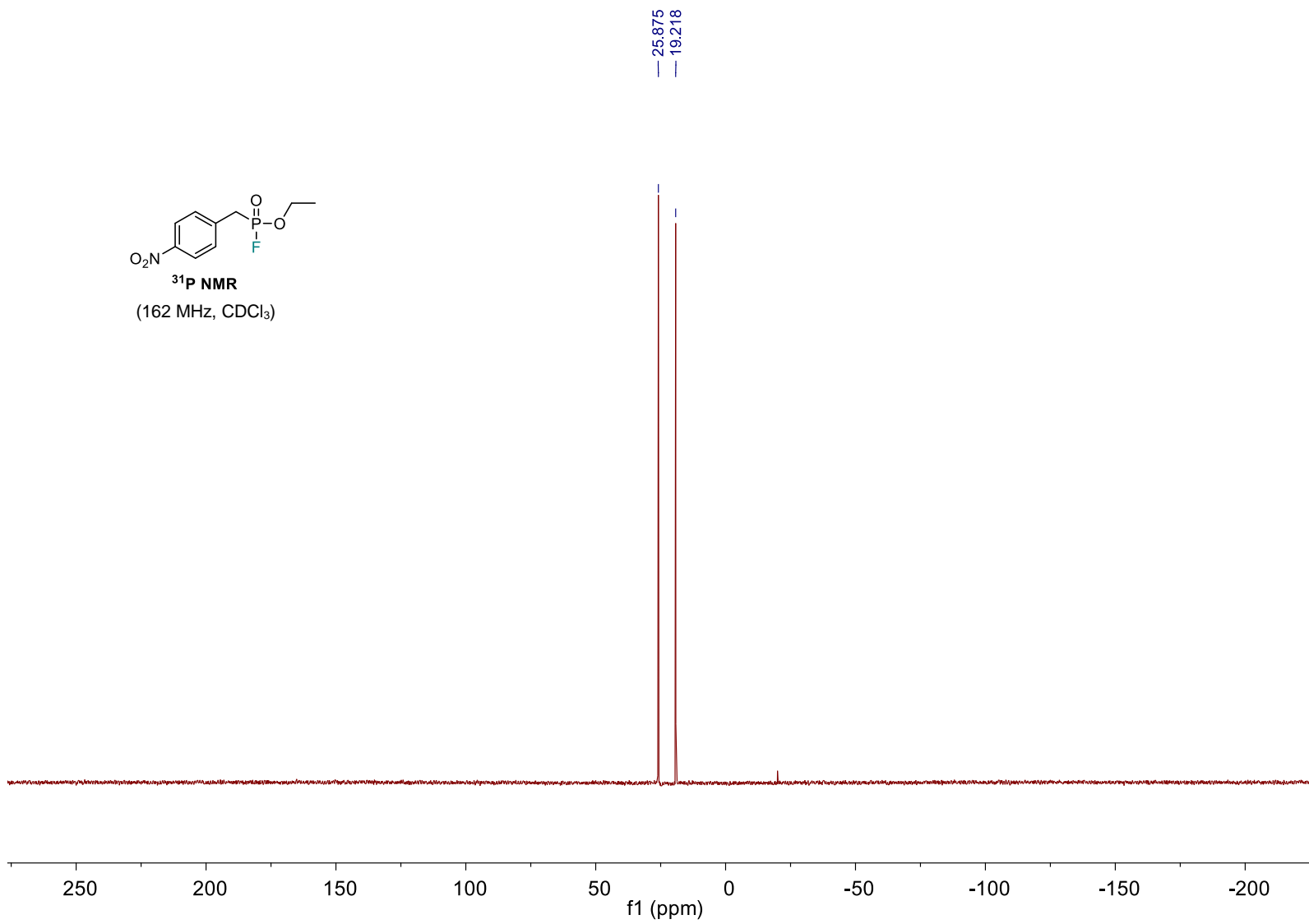
¹H NMR
(400 MHz, CDCl₃)



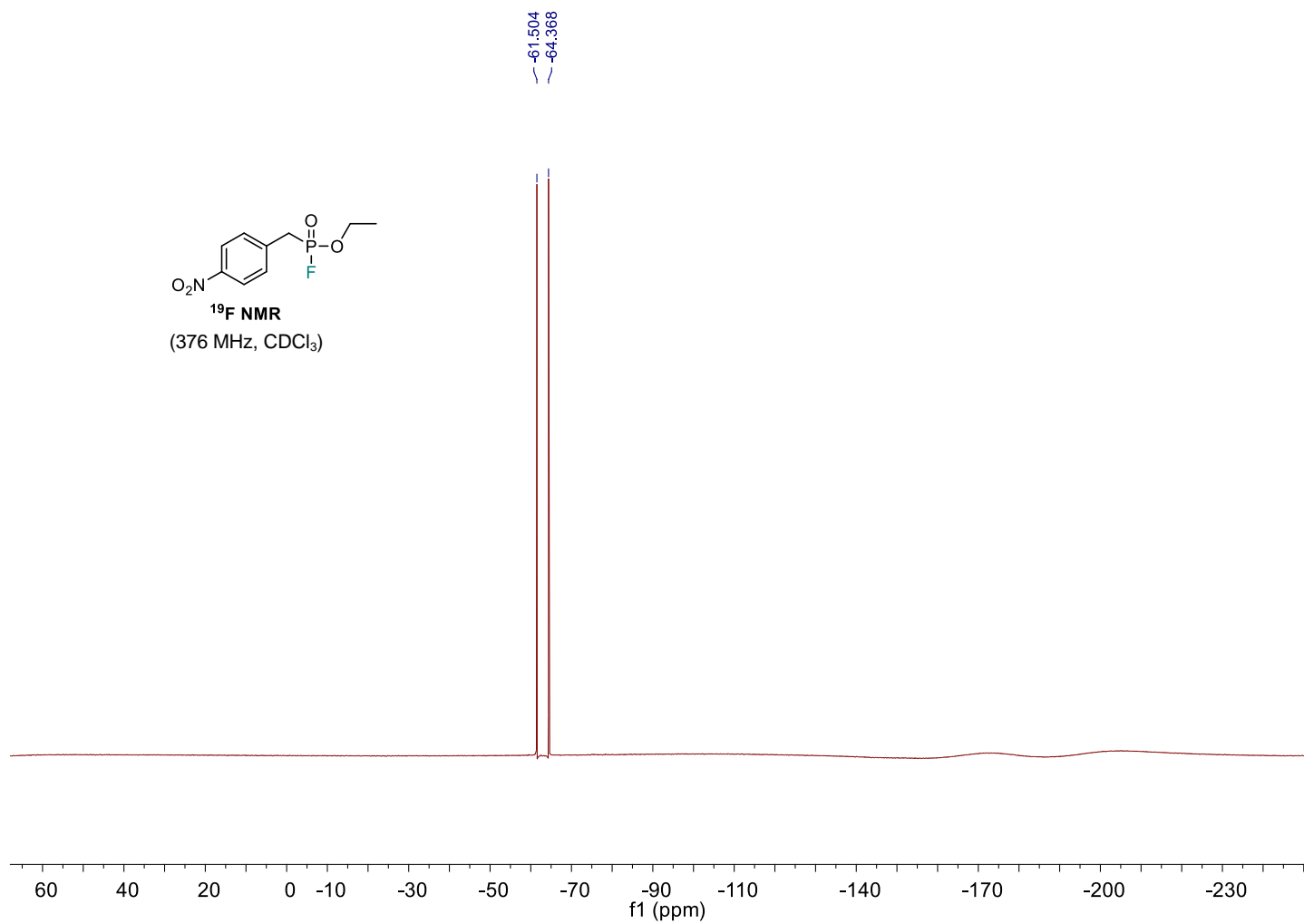
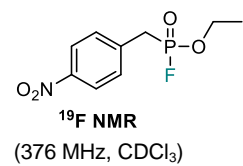


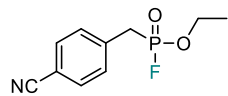


³¹P NMR
(162 MHz, CDCl₃)

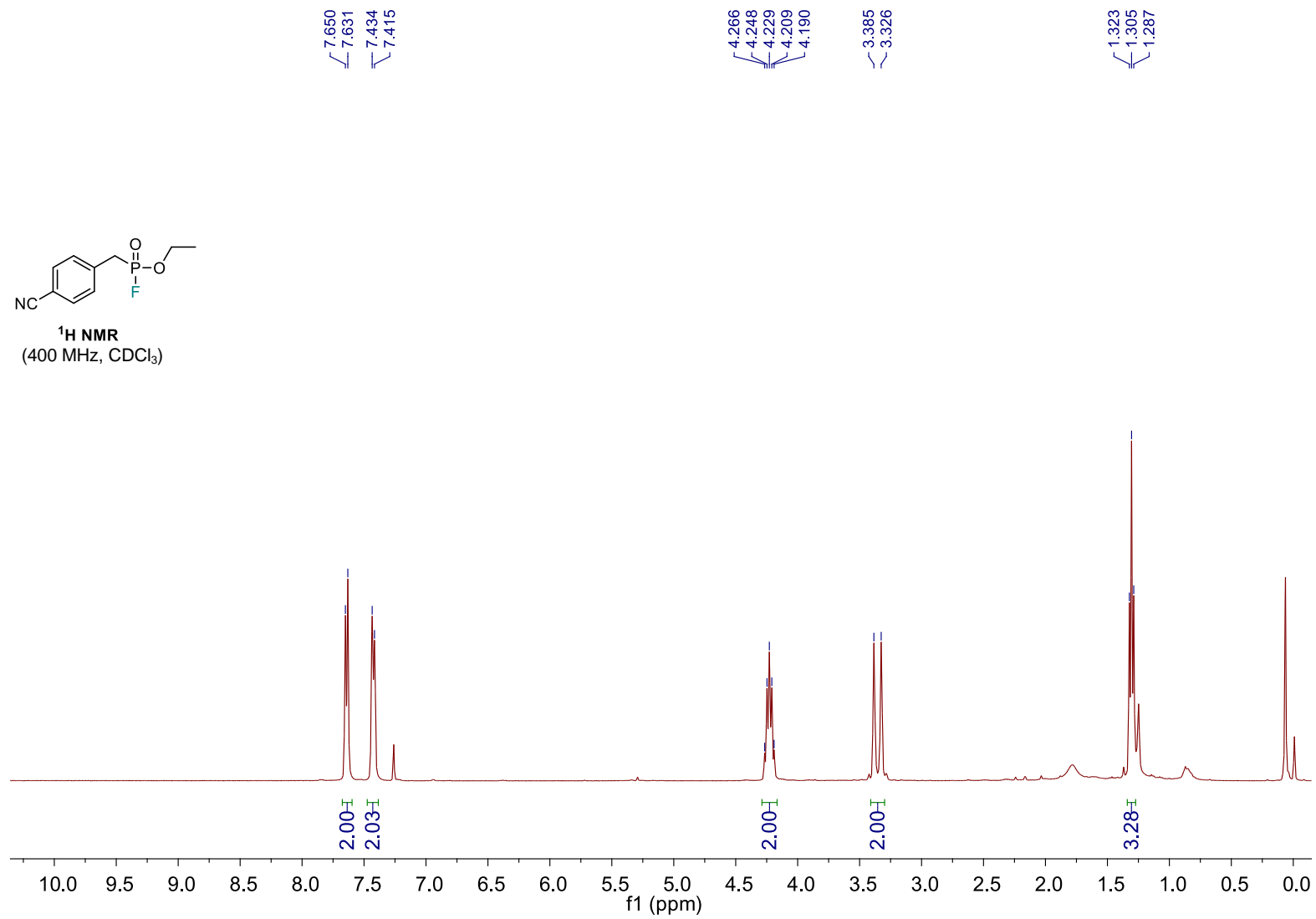


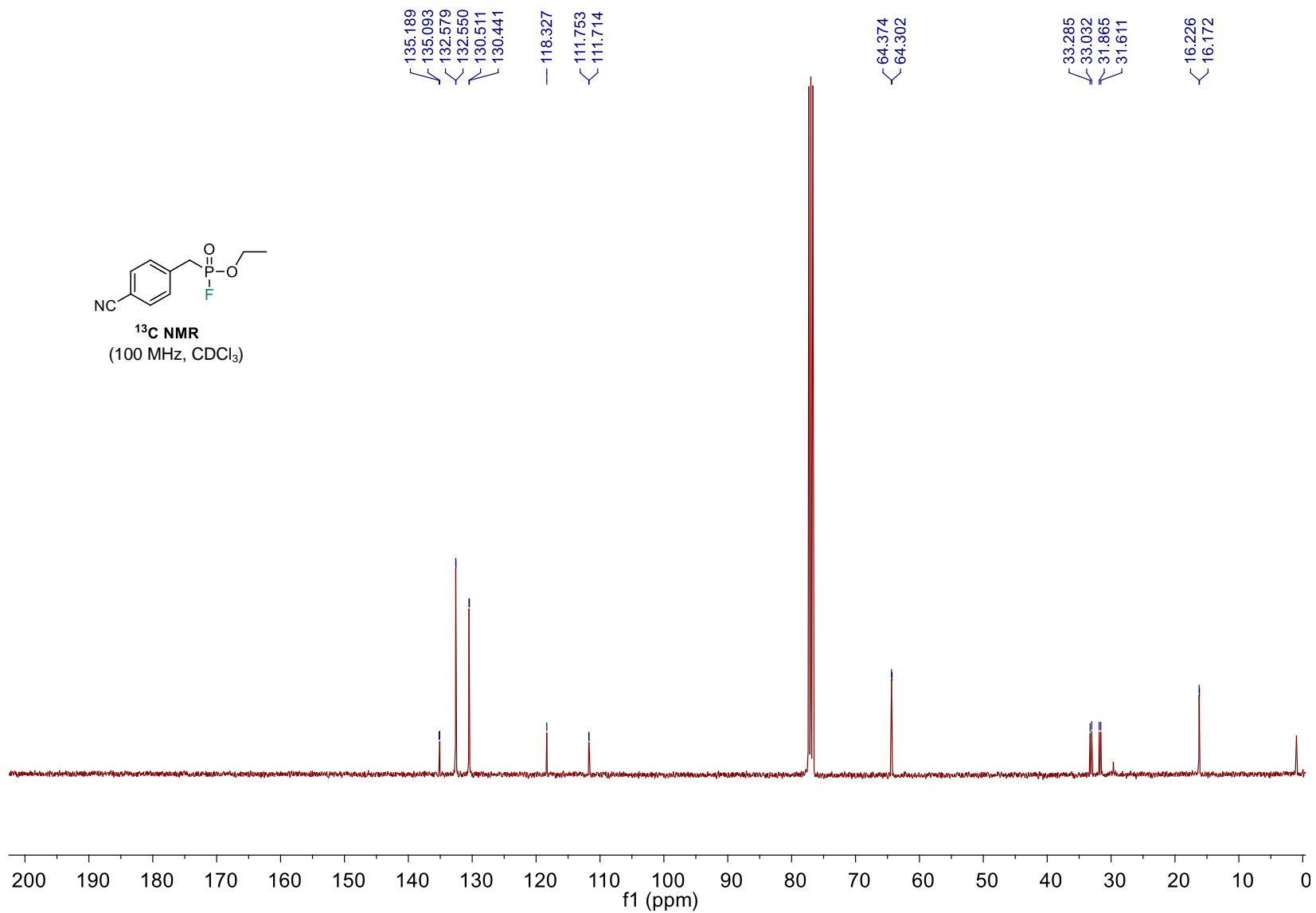
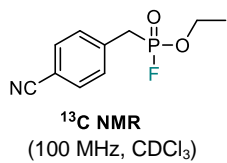
S271

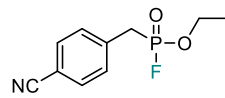




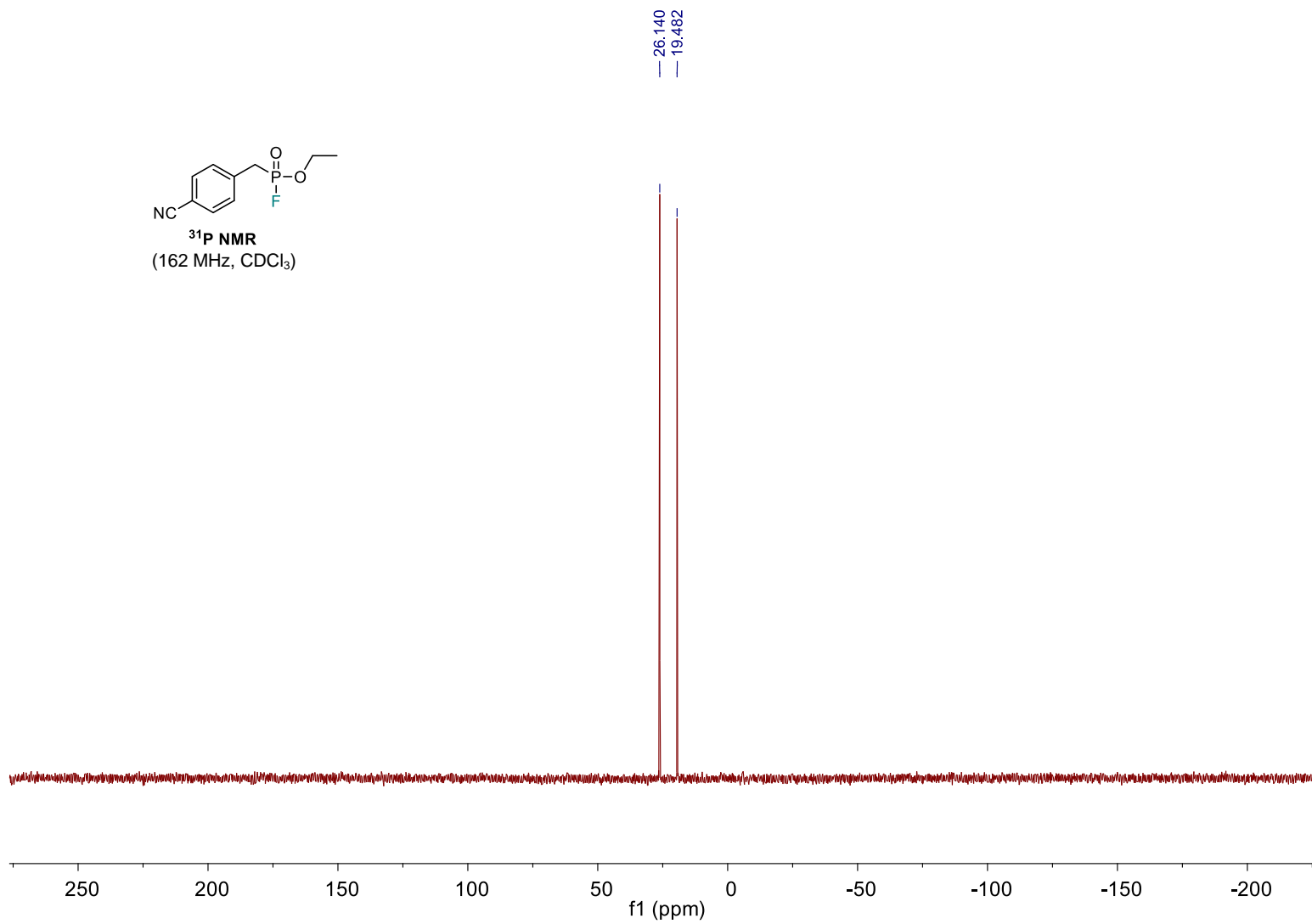
¹H NMR
(400 MHz, CDCl₃)



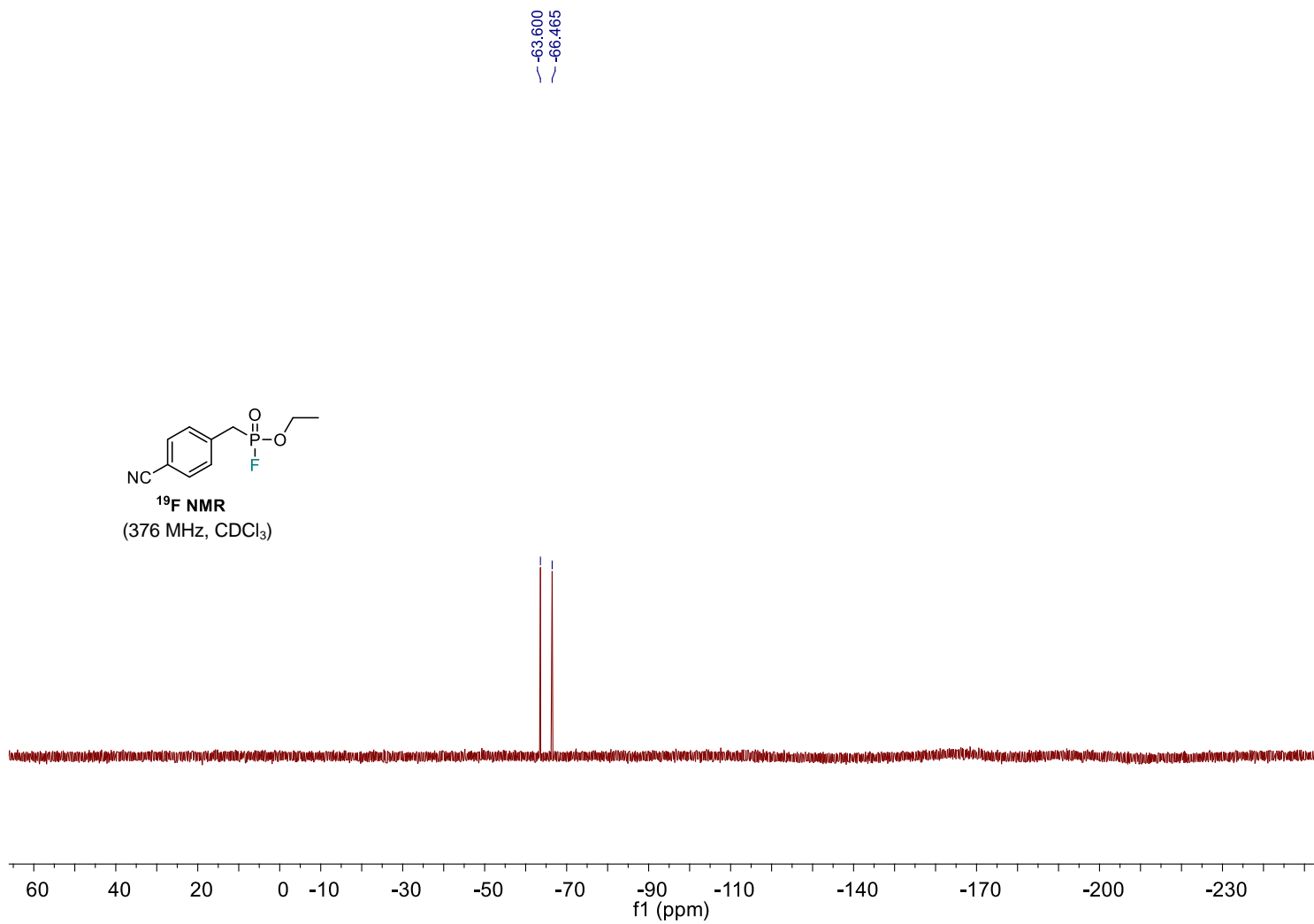
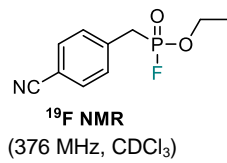


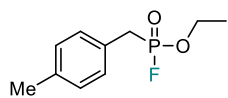


³¹P NMR
(162 MHz, CDCl₃)

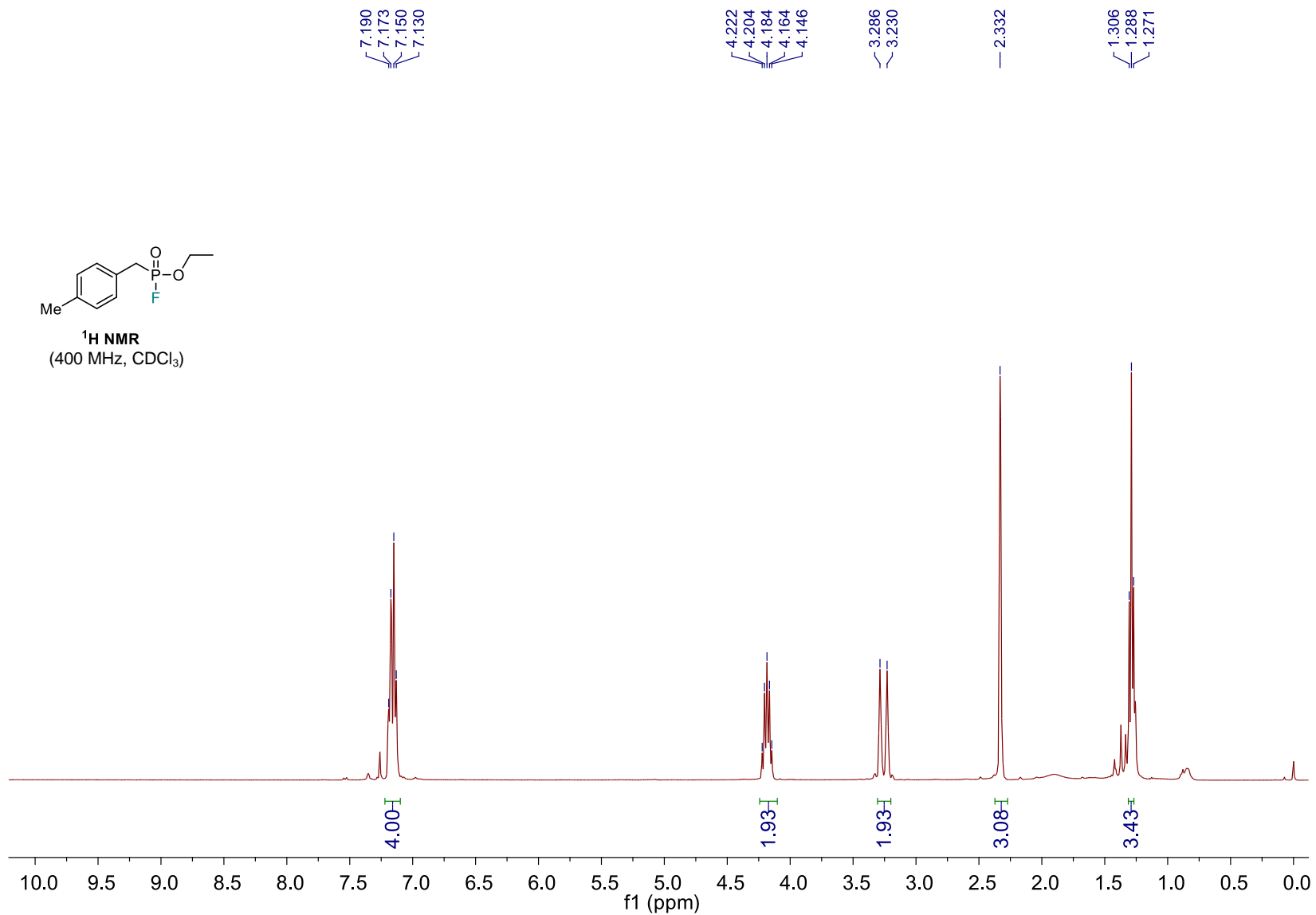


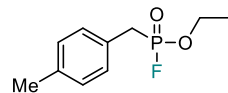
S275





¹H NMR
(400 MHz, CDCl₃)





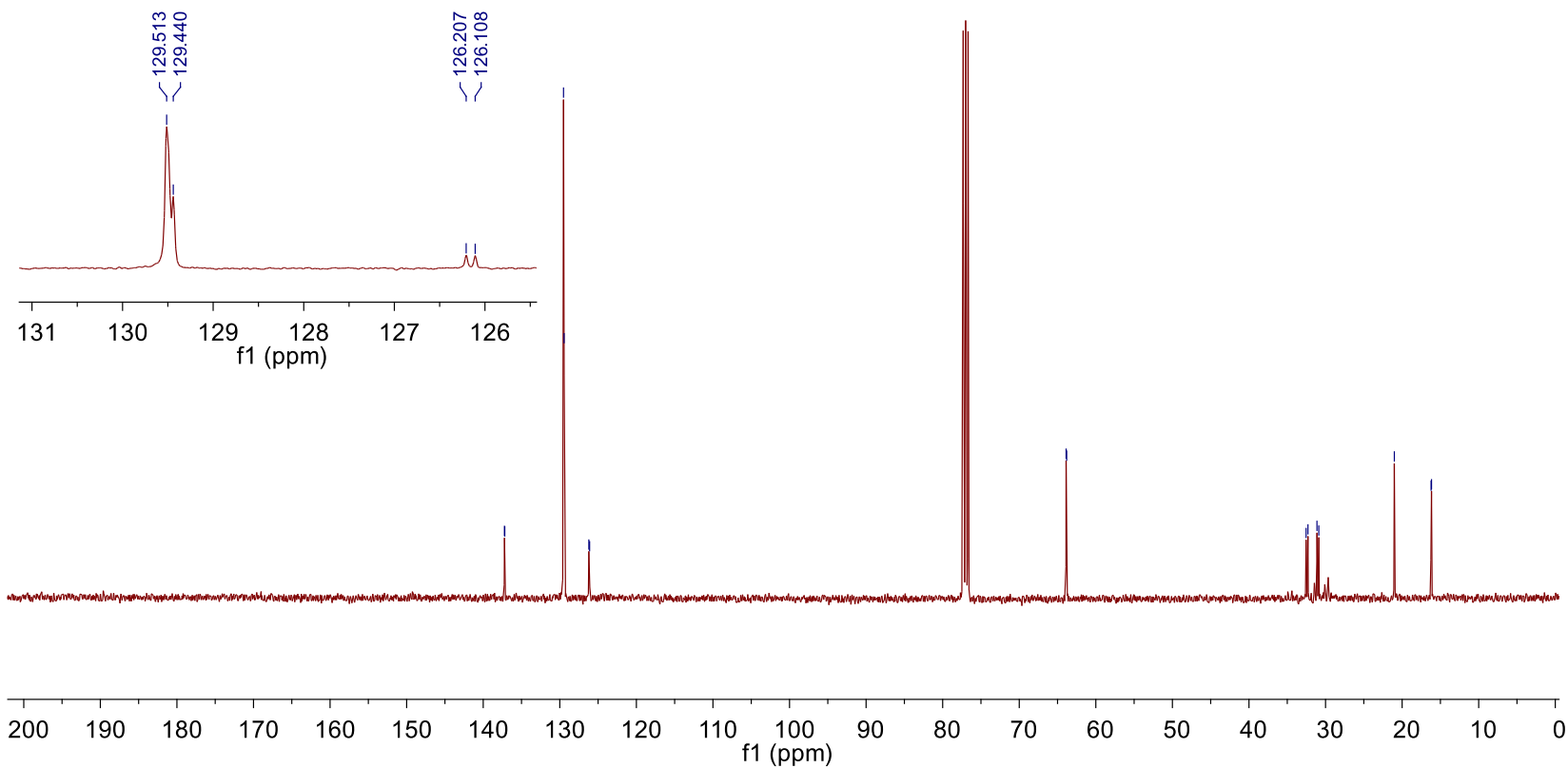
¹³C NMR
(100 MHz, CDCl₃)

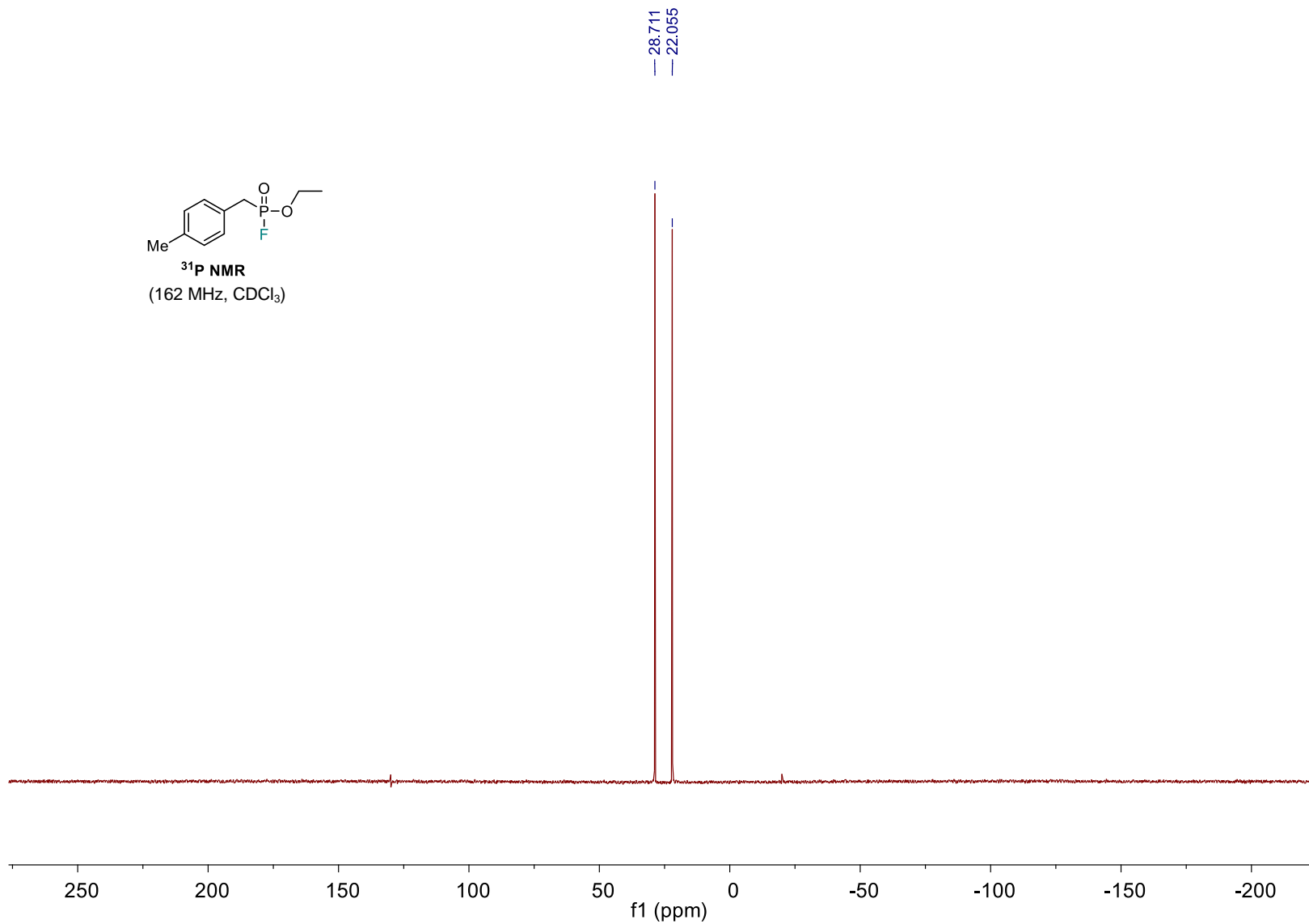
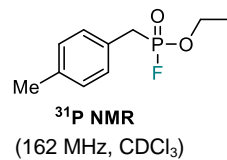
137.226
137.187
129.513
129.440
126.207
126.108

63.880
63.807

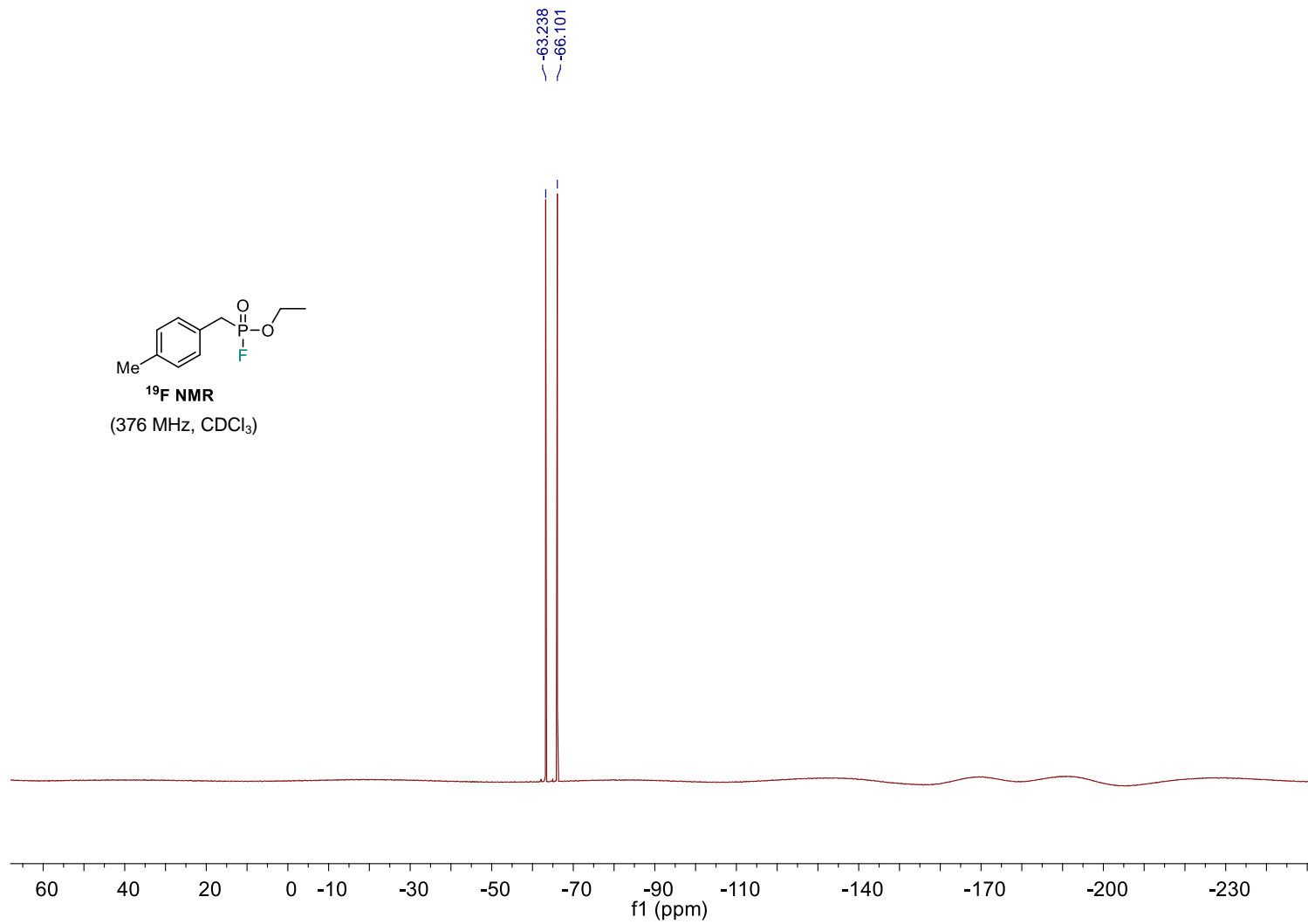
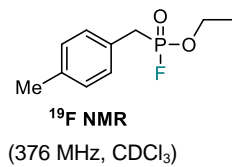
32.540
32.297
31.123
30.880

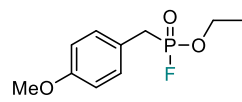
21.018
16.217
16.163



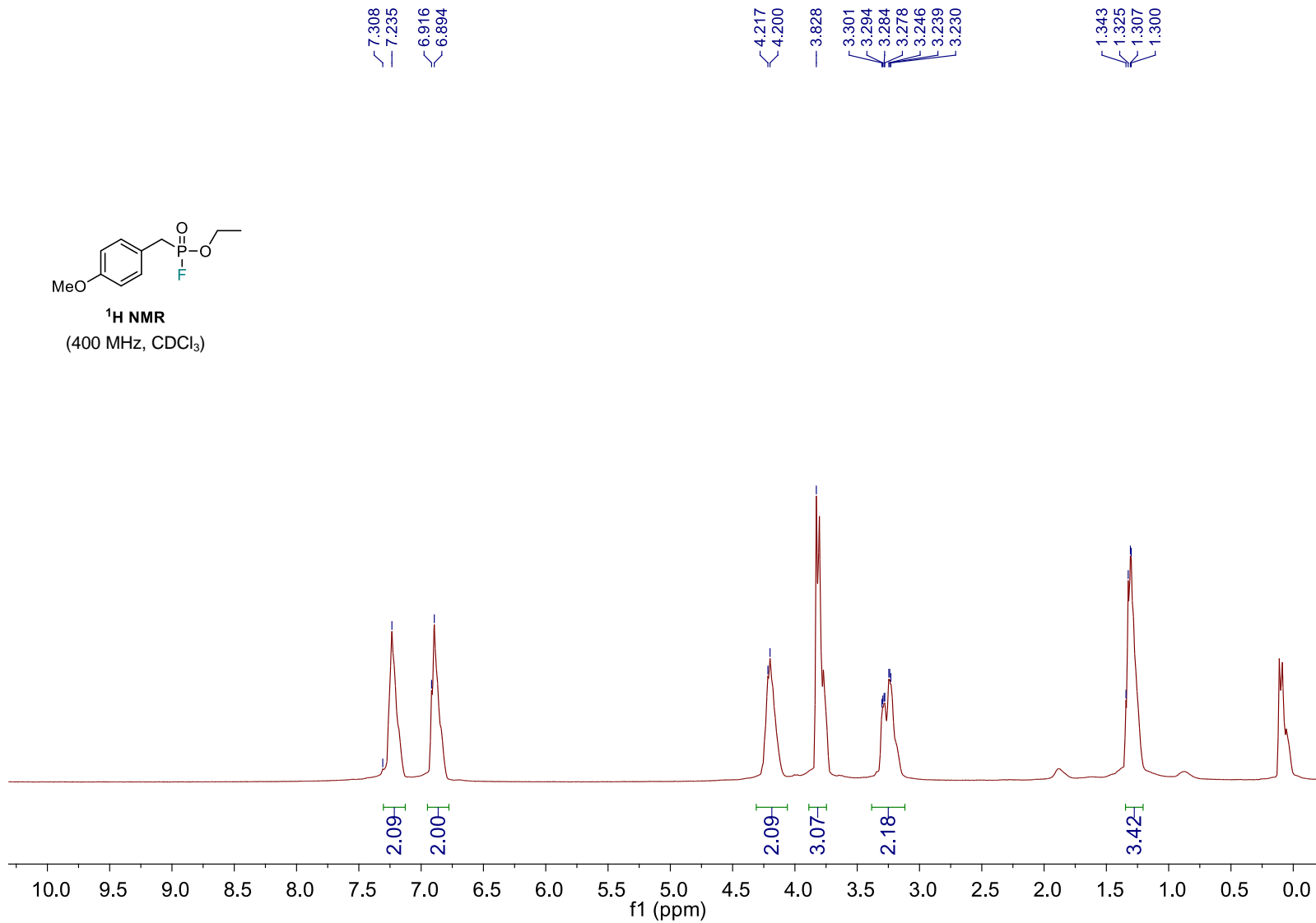


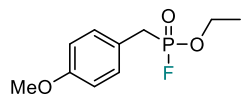
S279



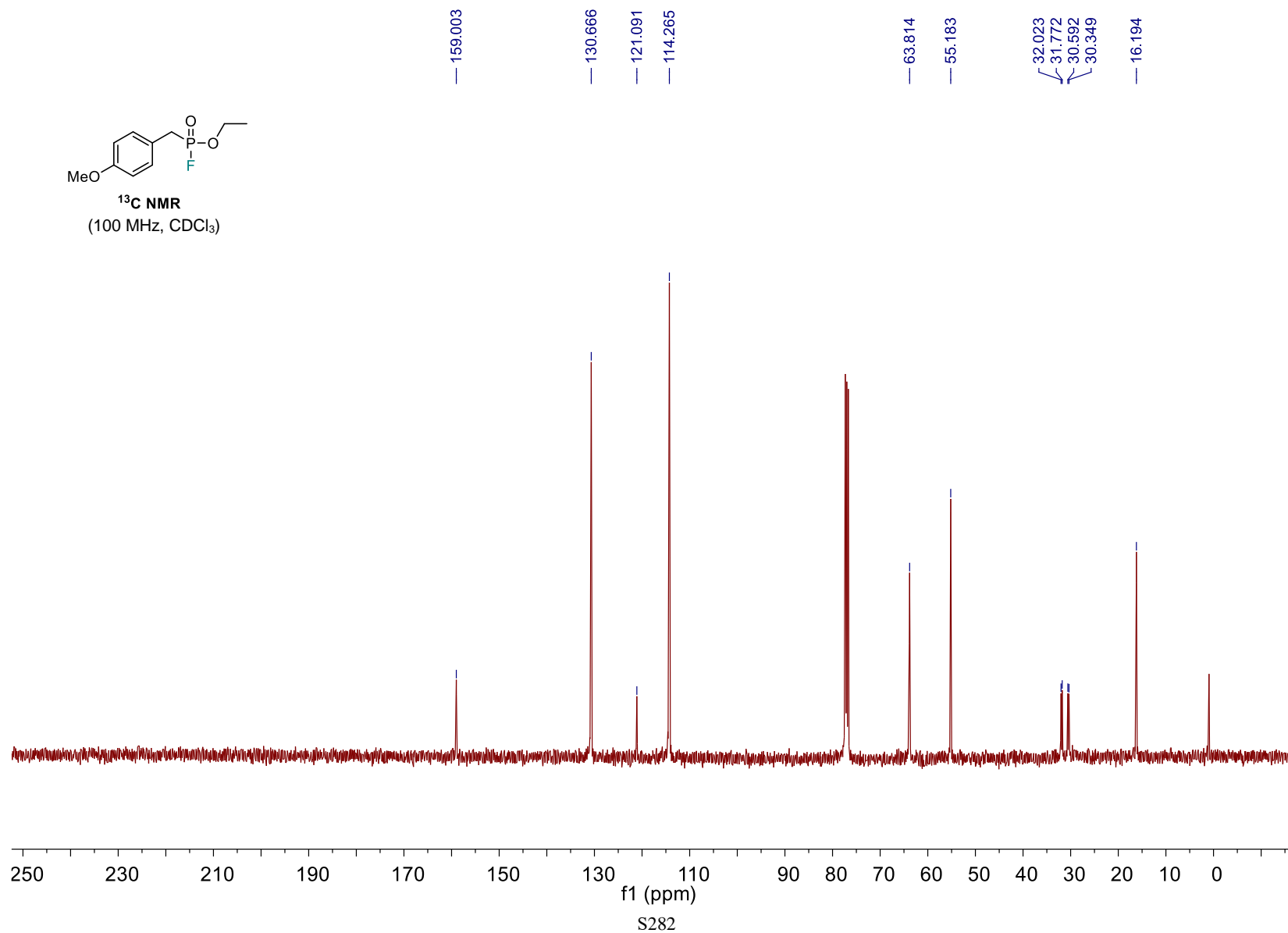


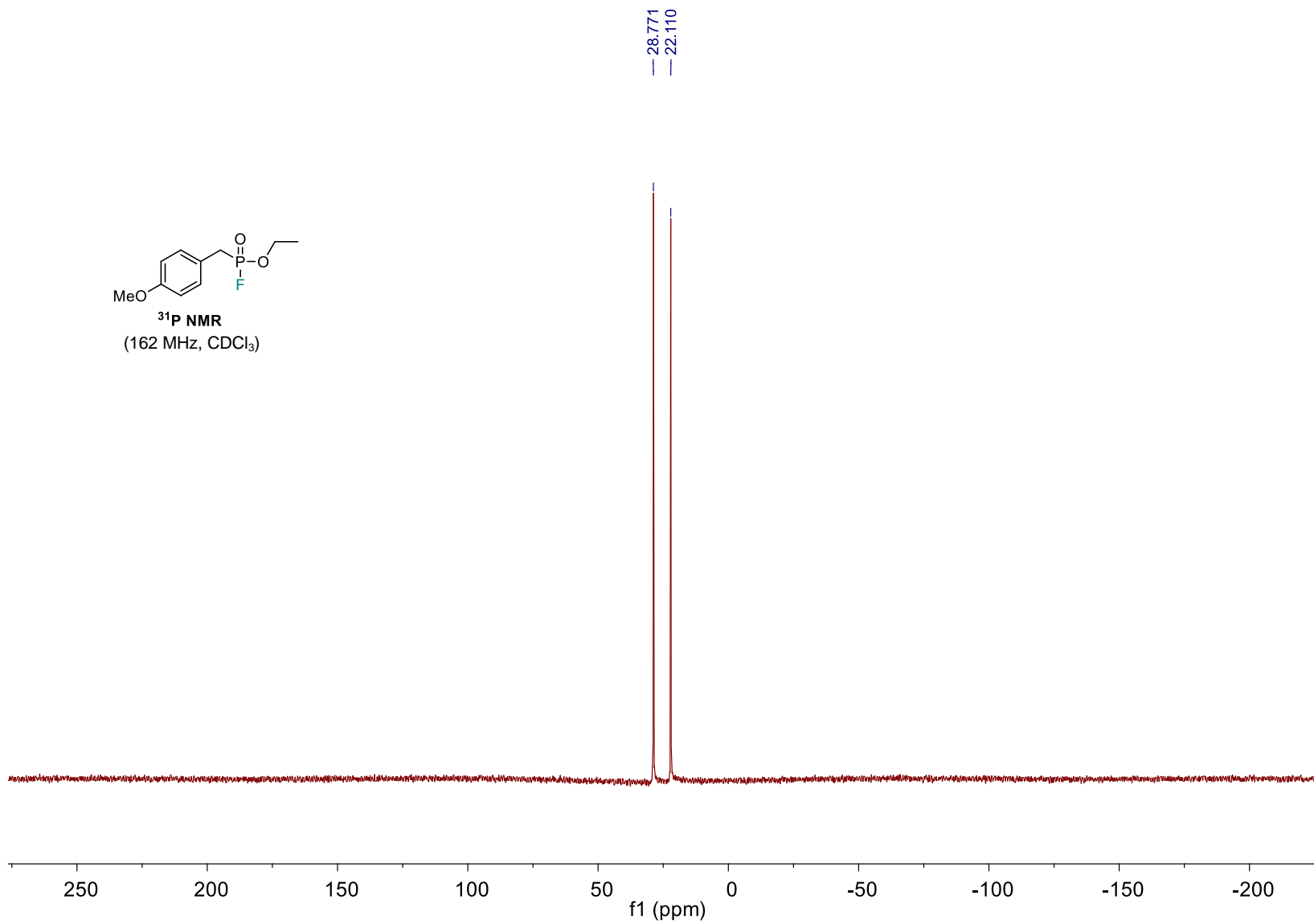
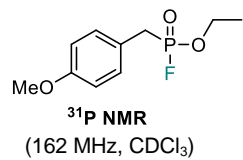
¹H NMR
(400 MHz, CDCl₃)



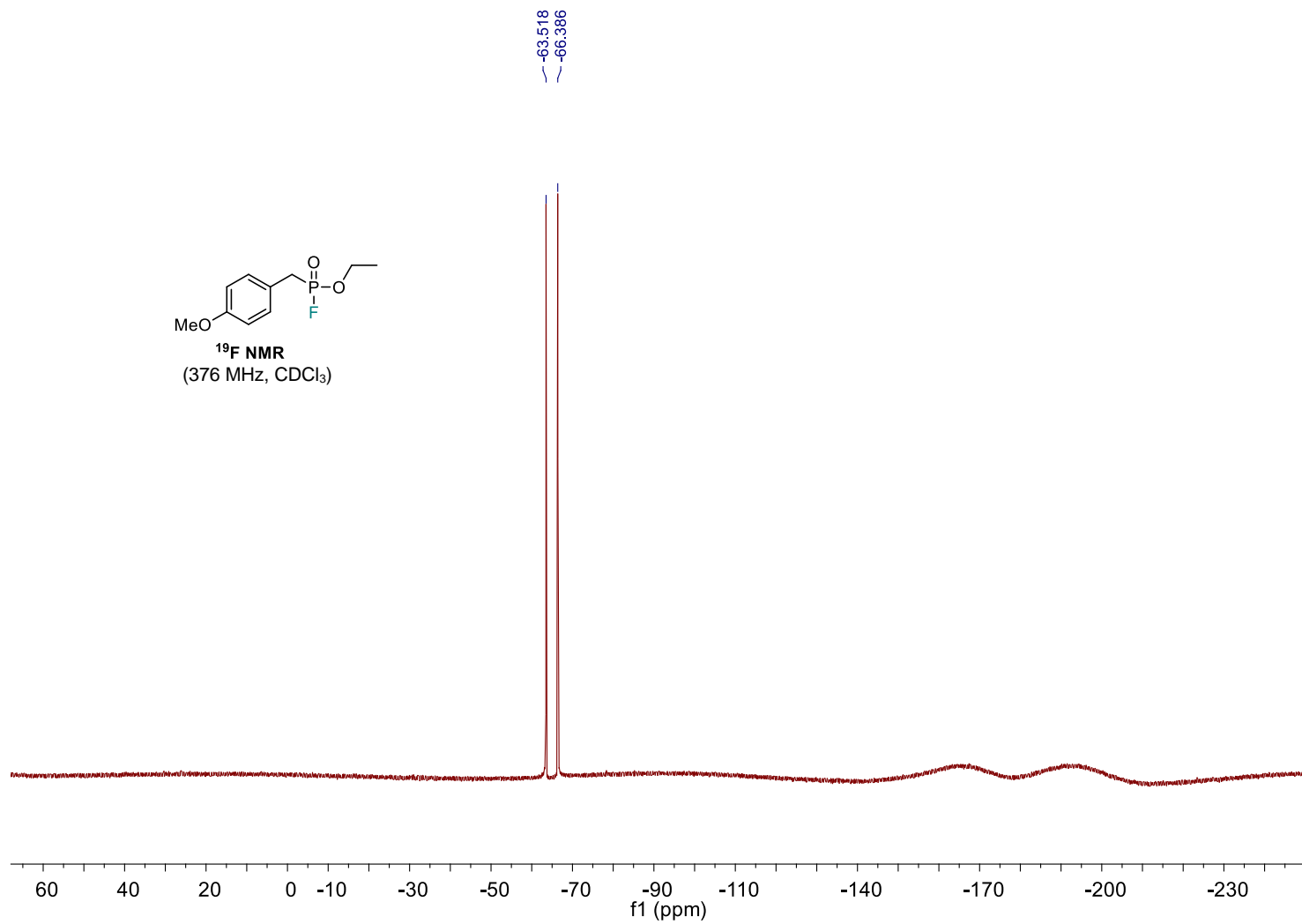
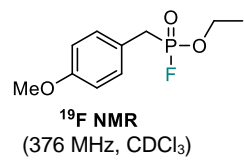


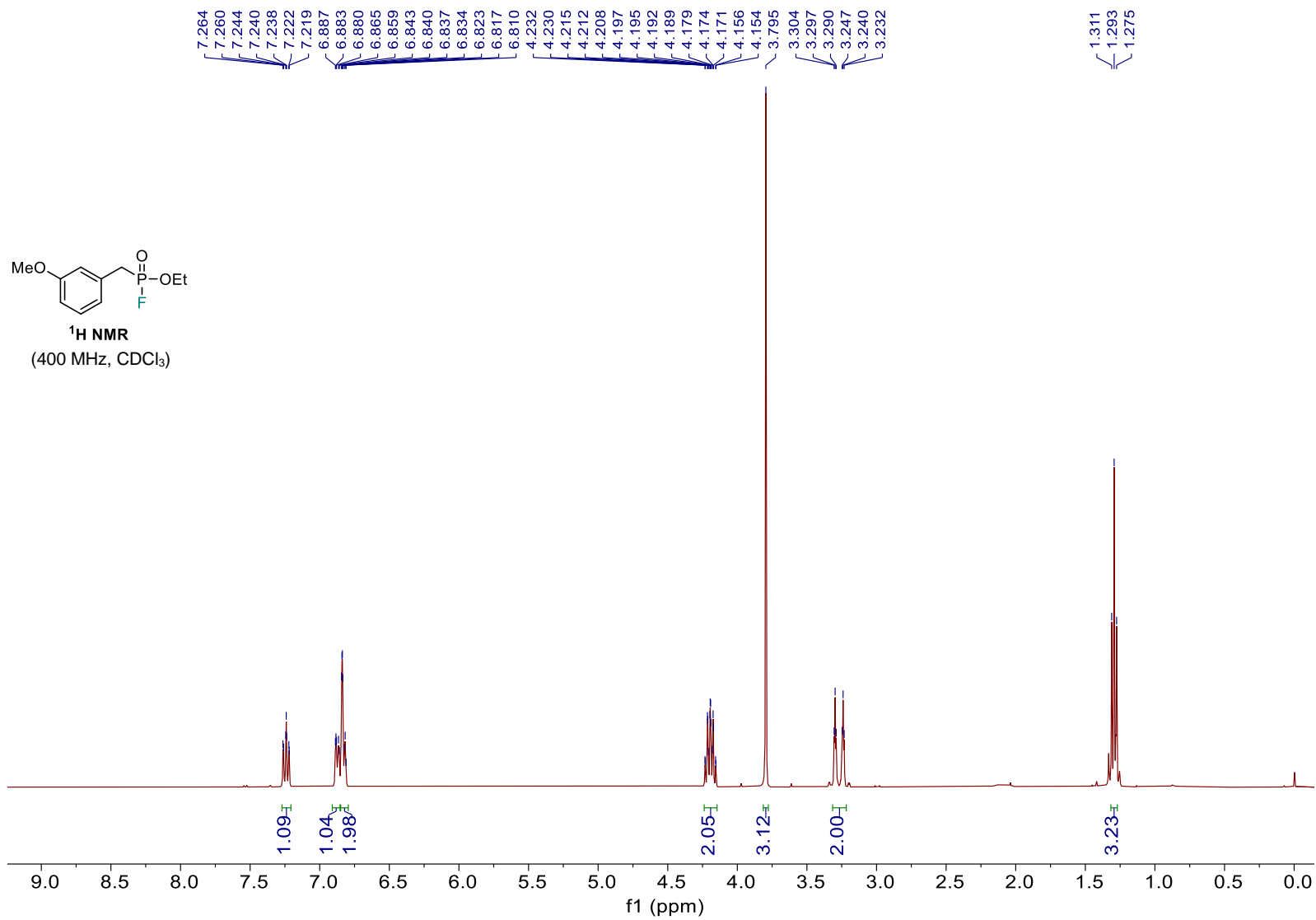
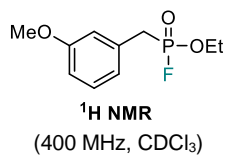
¹³C NMR
(100 MHz, CDCl₃)

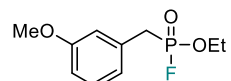




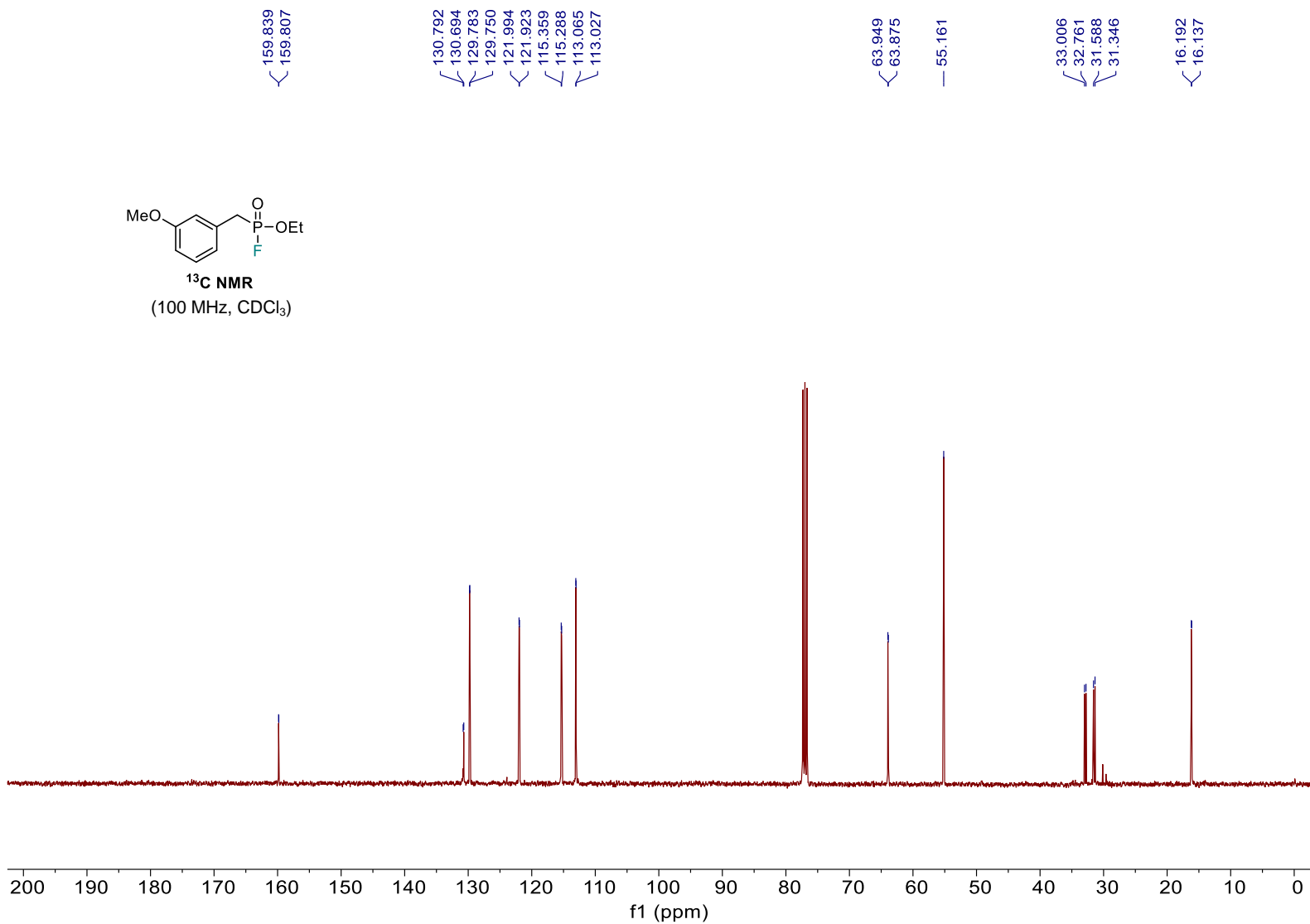
S283

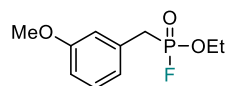




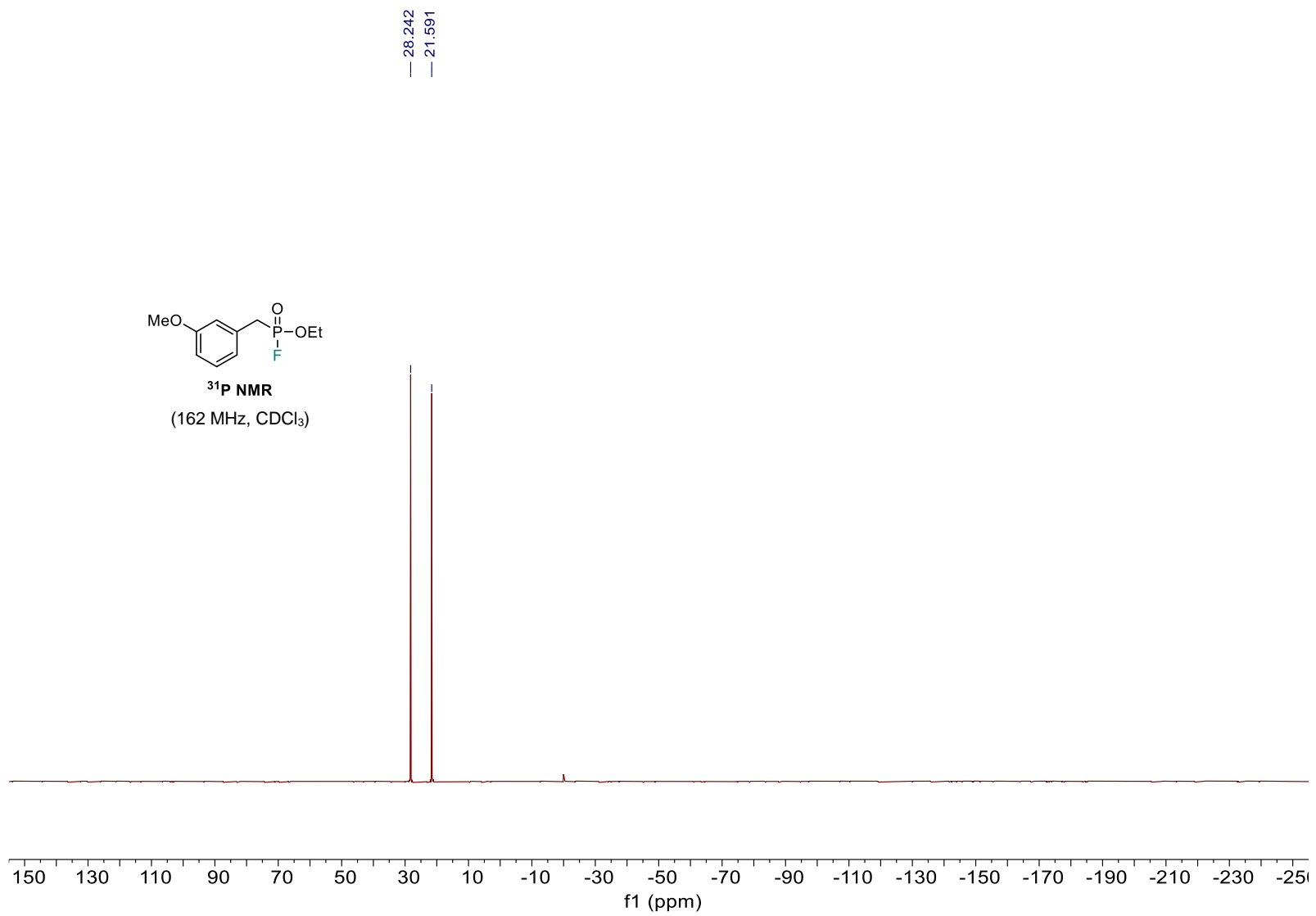


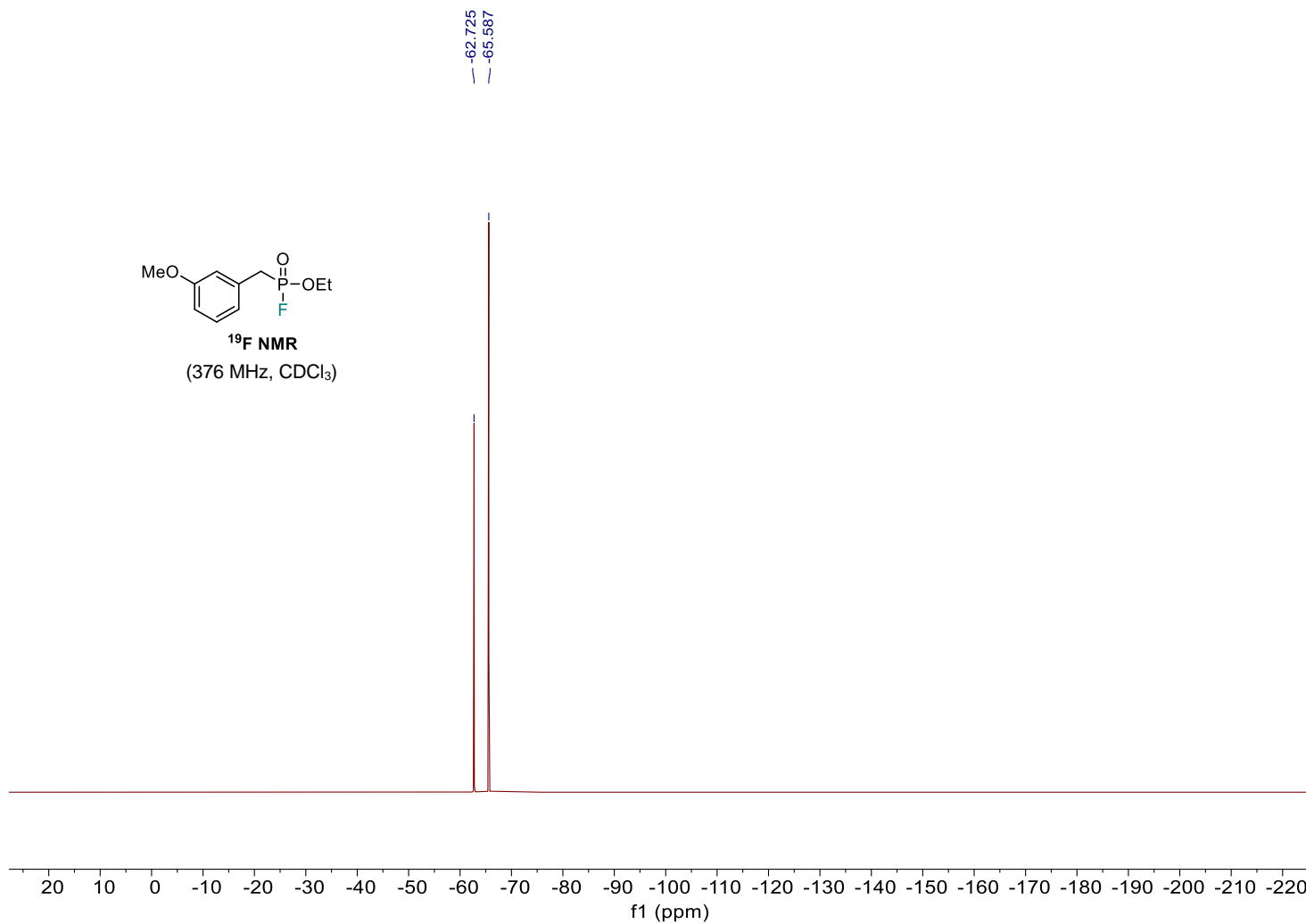
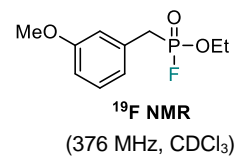
¹³C NMR
(100 MHz, CDCl₃)

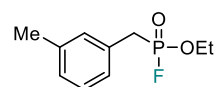




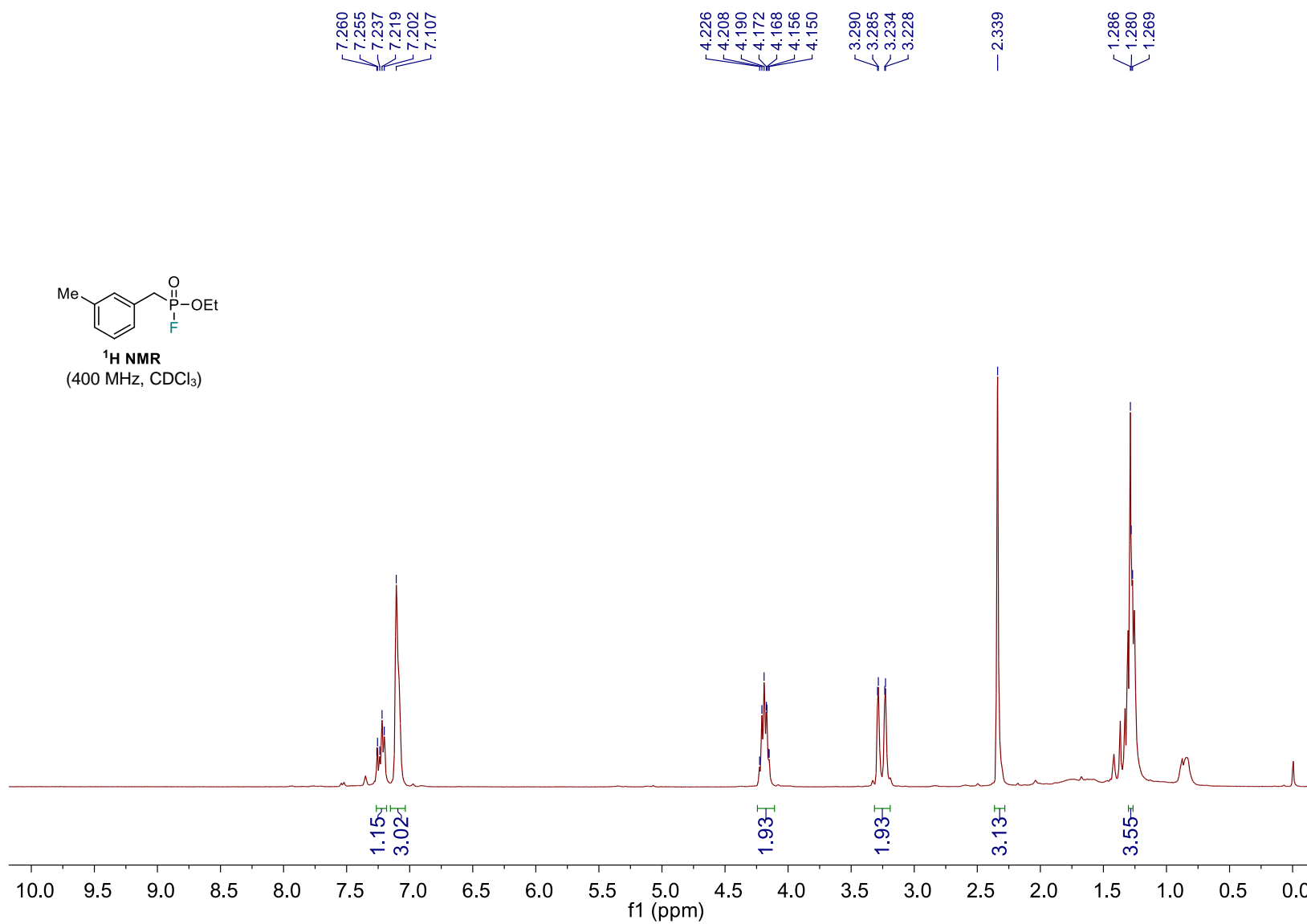
³¹P NMR
(162 MHz, CDCl₃)

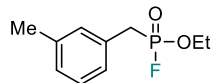




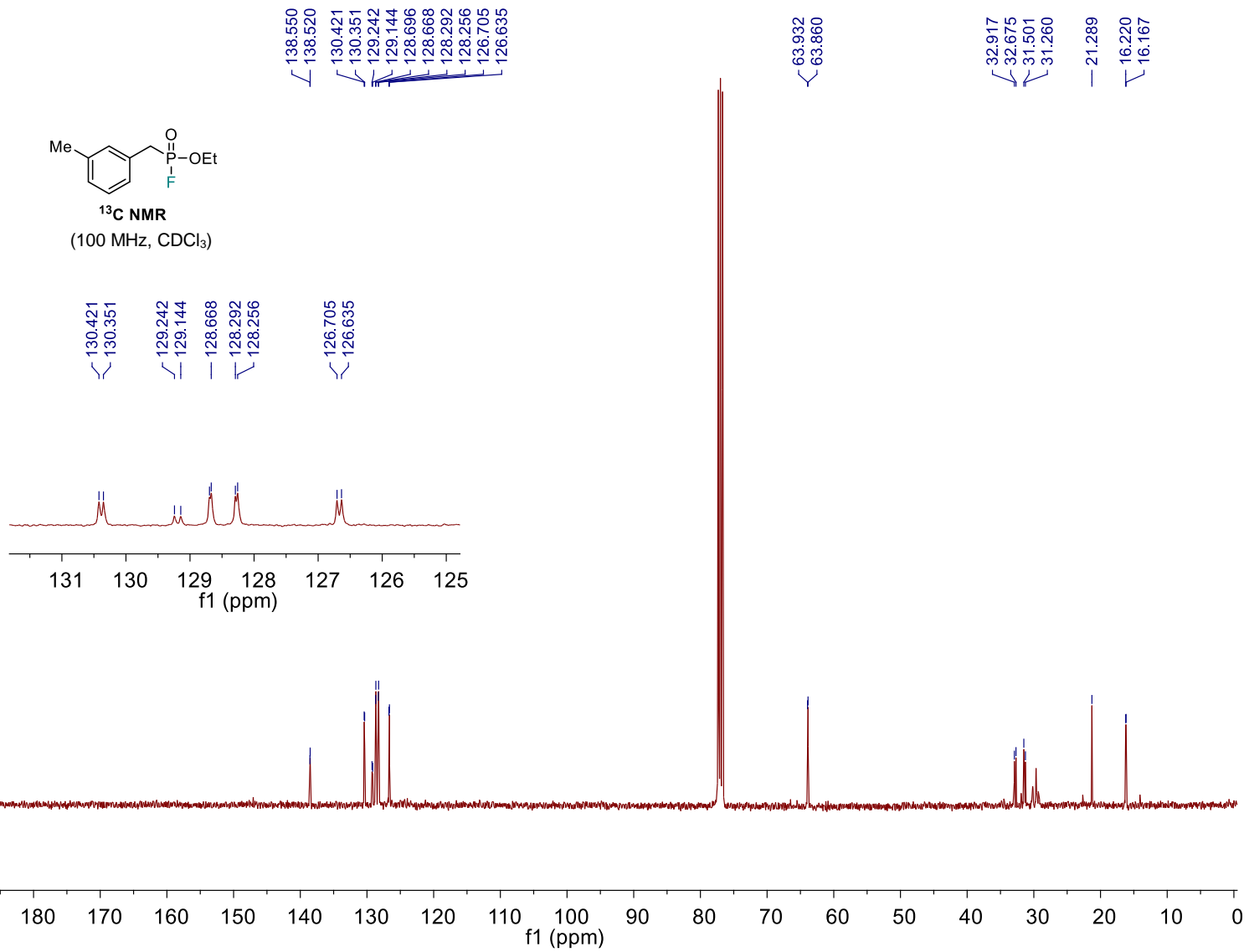


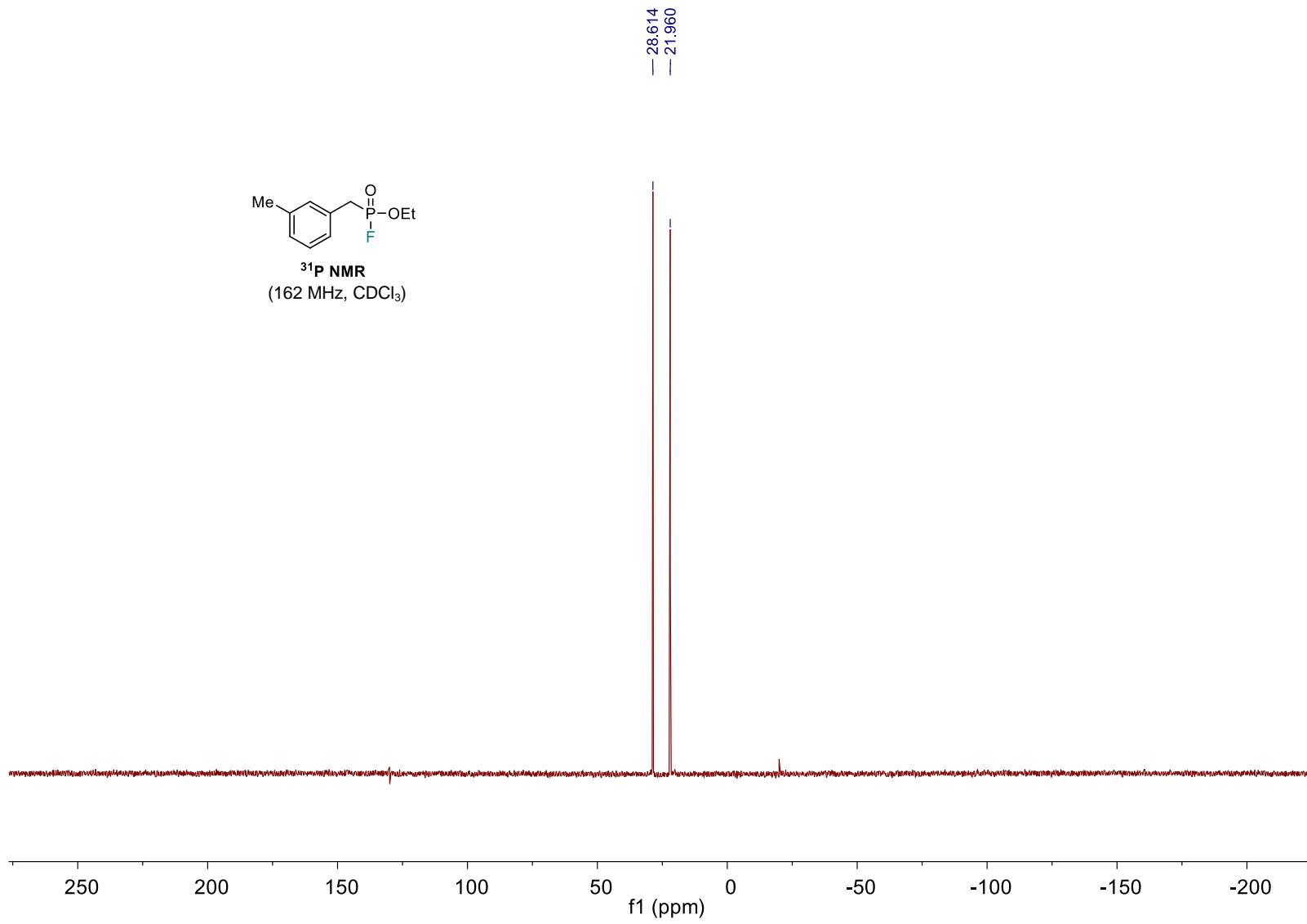
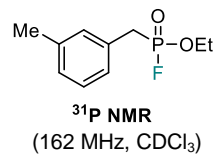
¹H NMR
(400 MHz, CDCl₃)

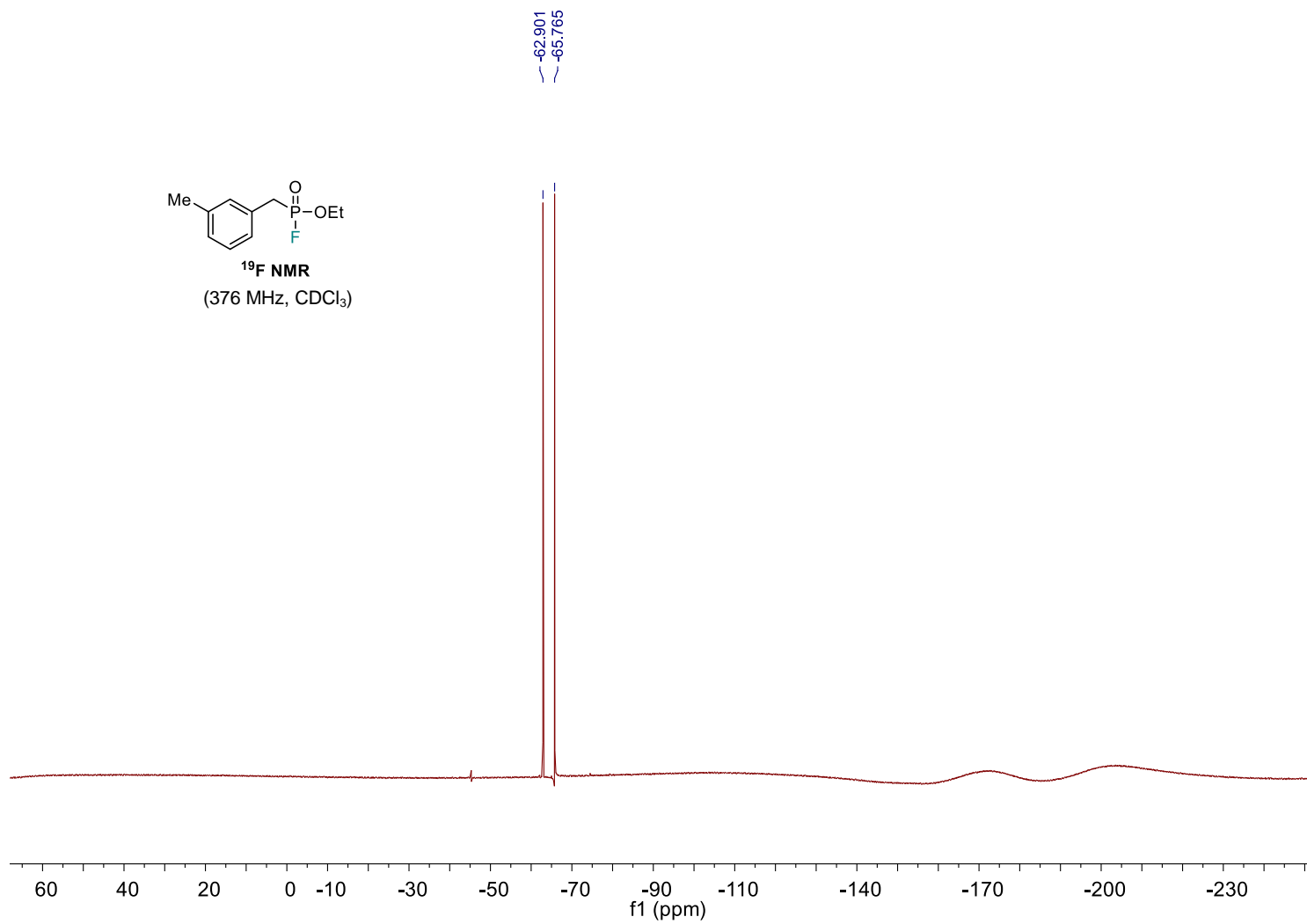
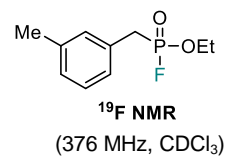




¹³C NMR
(100 MHz, CDCl₃)





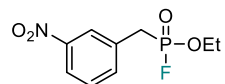


8.177
8.174
8.170
8.161
7.671
7.658
7.554
7.541
7.527

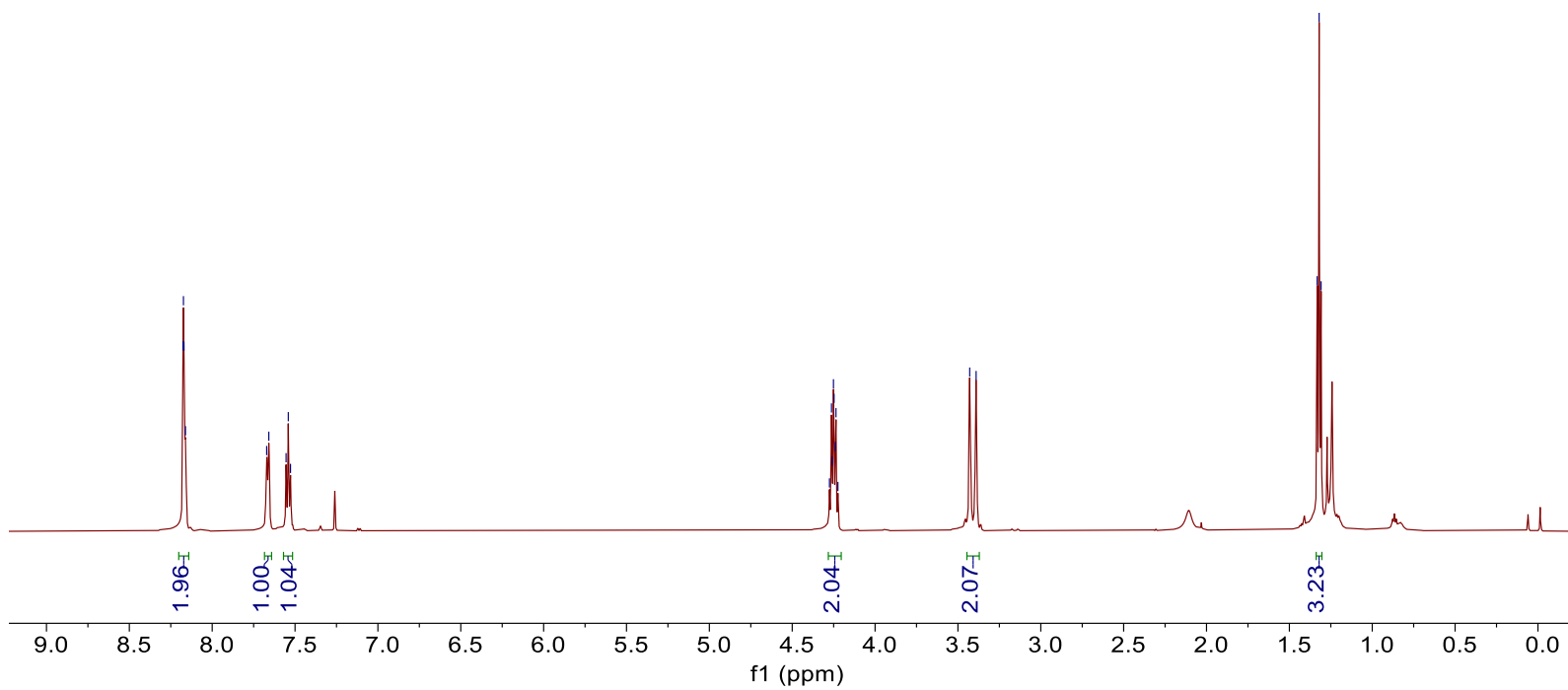
4.275
4.263
4.259
4.251
4.247
4.240
4.236
4.224

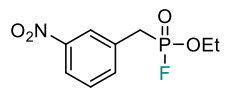
3.429
3.390

1.332
1.321
1.309

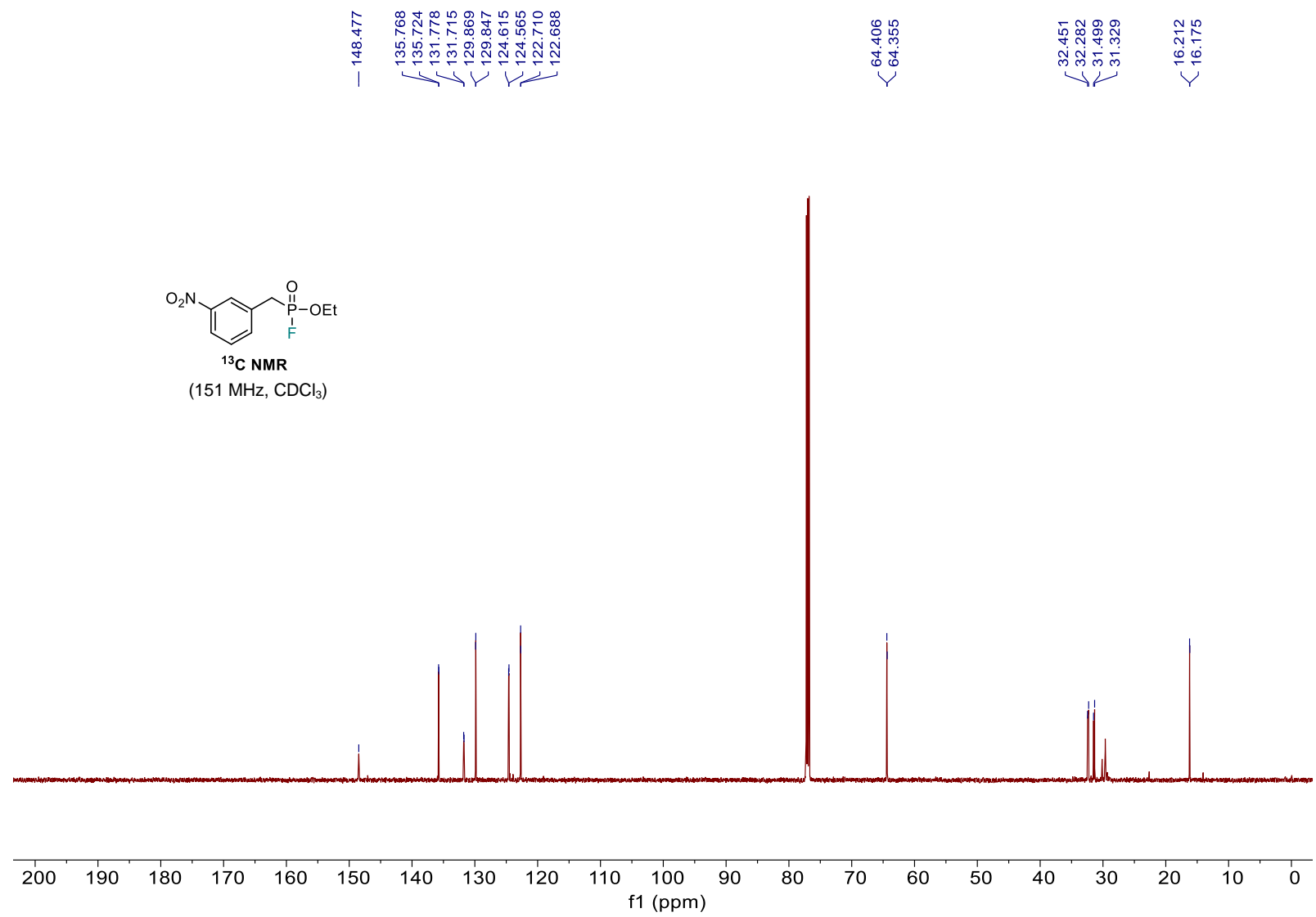


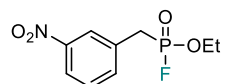
¹H NMR
(600 MHz, CDCl₃)



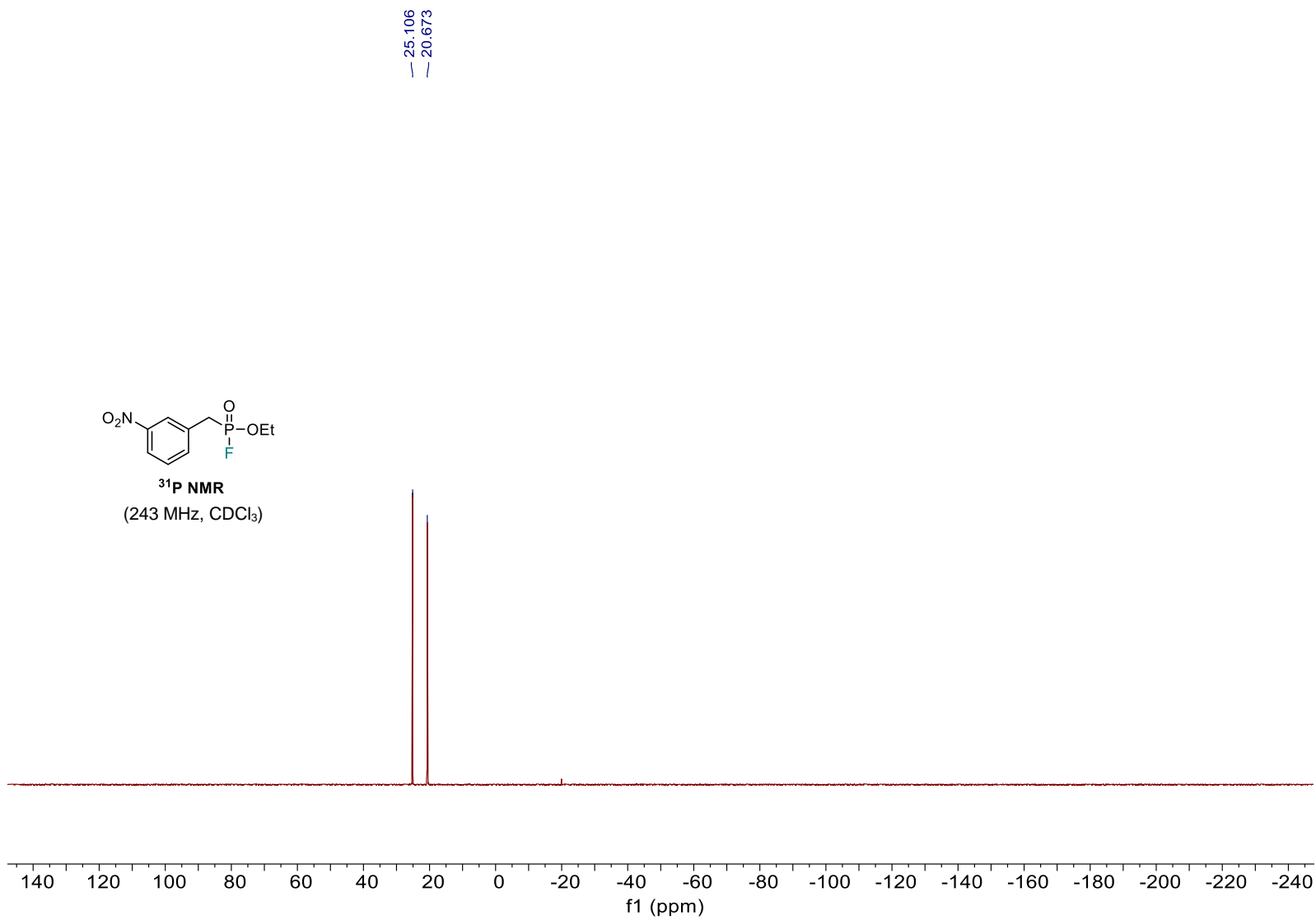


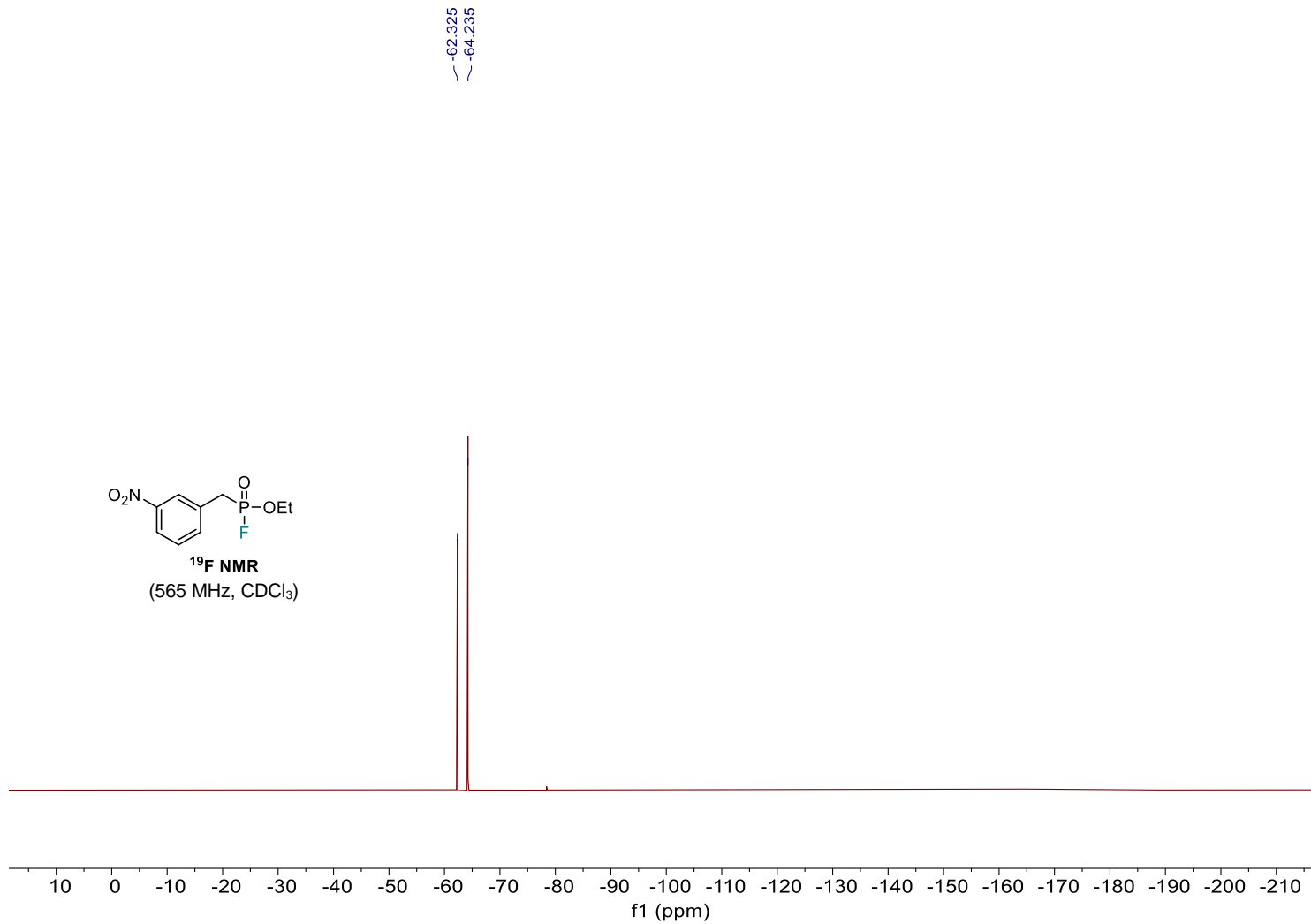
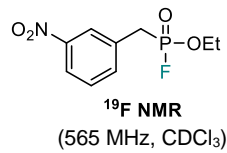
¹³C NMR
(151 MHz, CDCl₃)

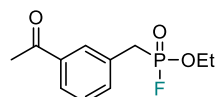




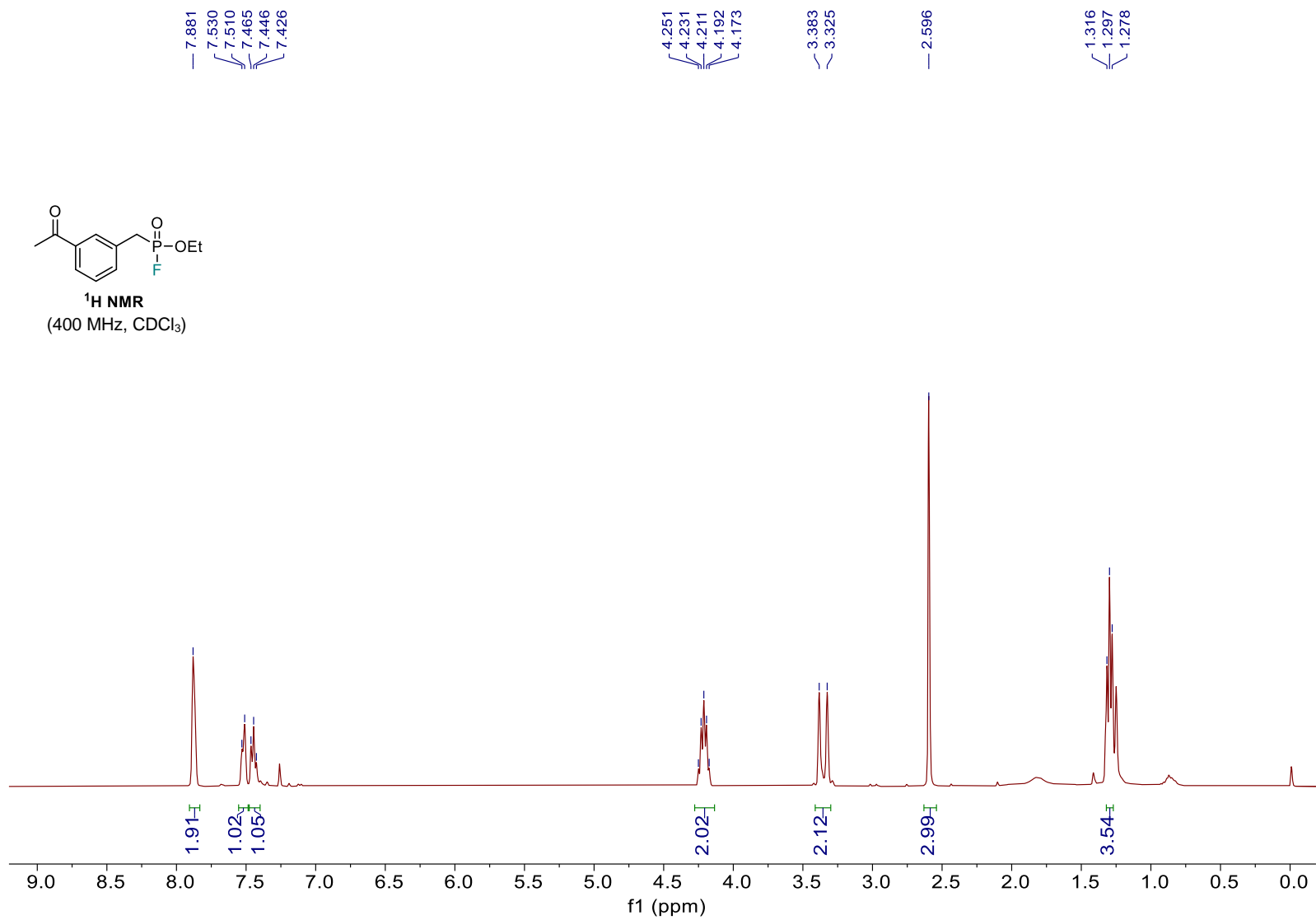
³¹P NMR
(243 MHz, CDCl₃)

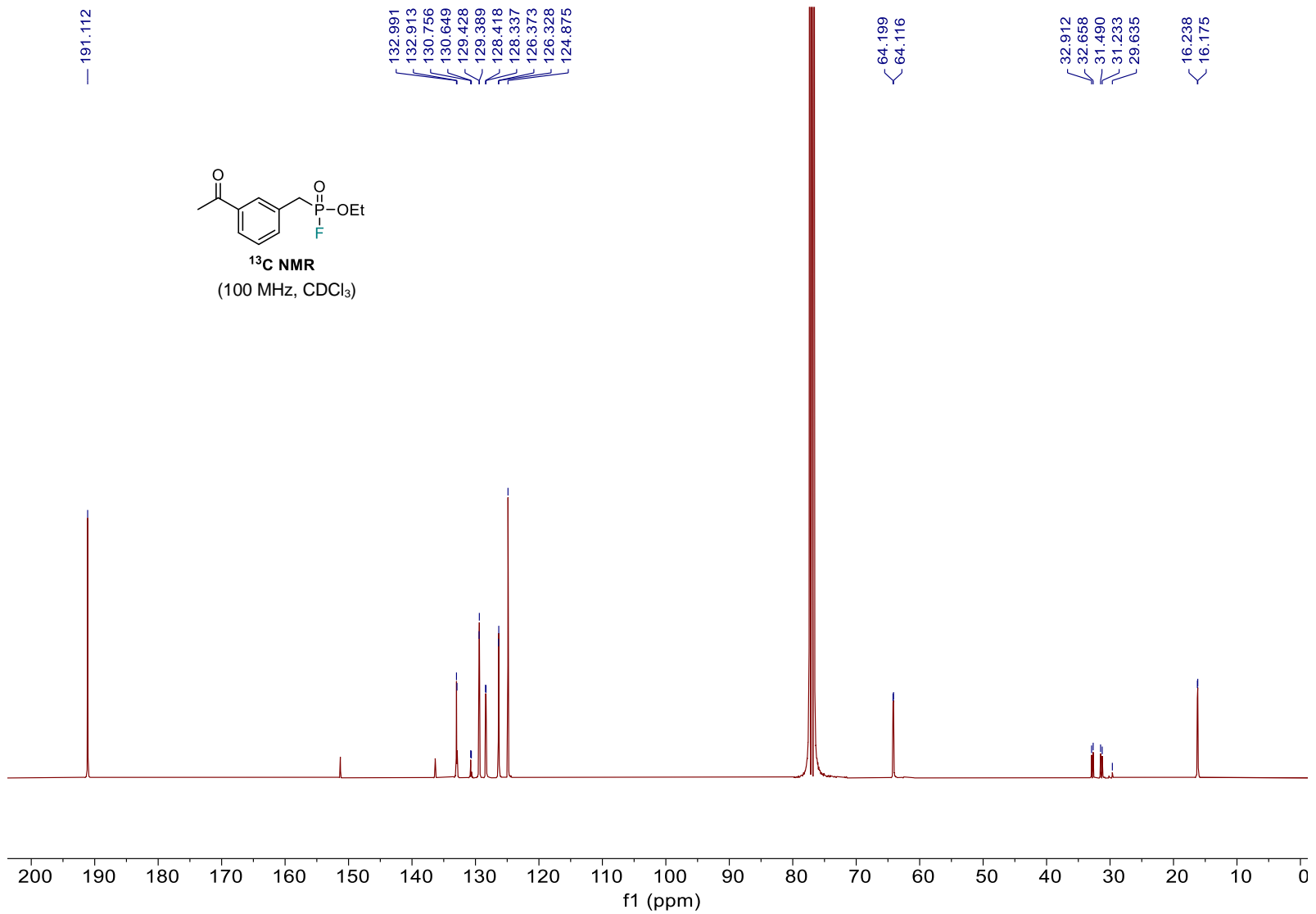


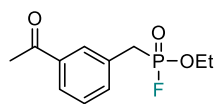




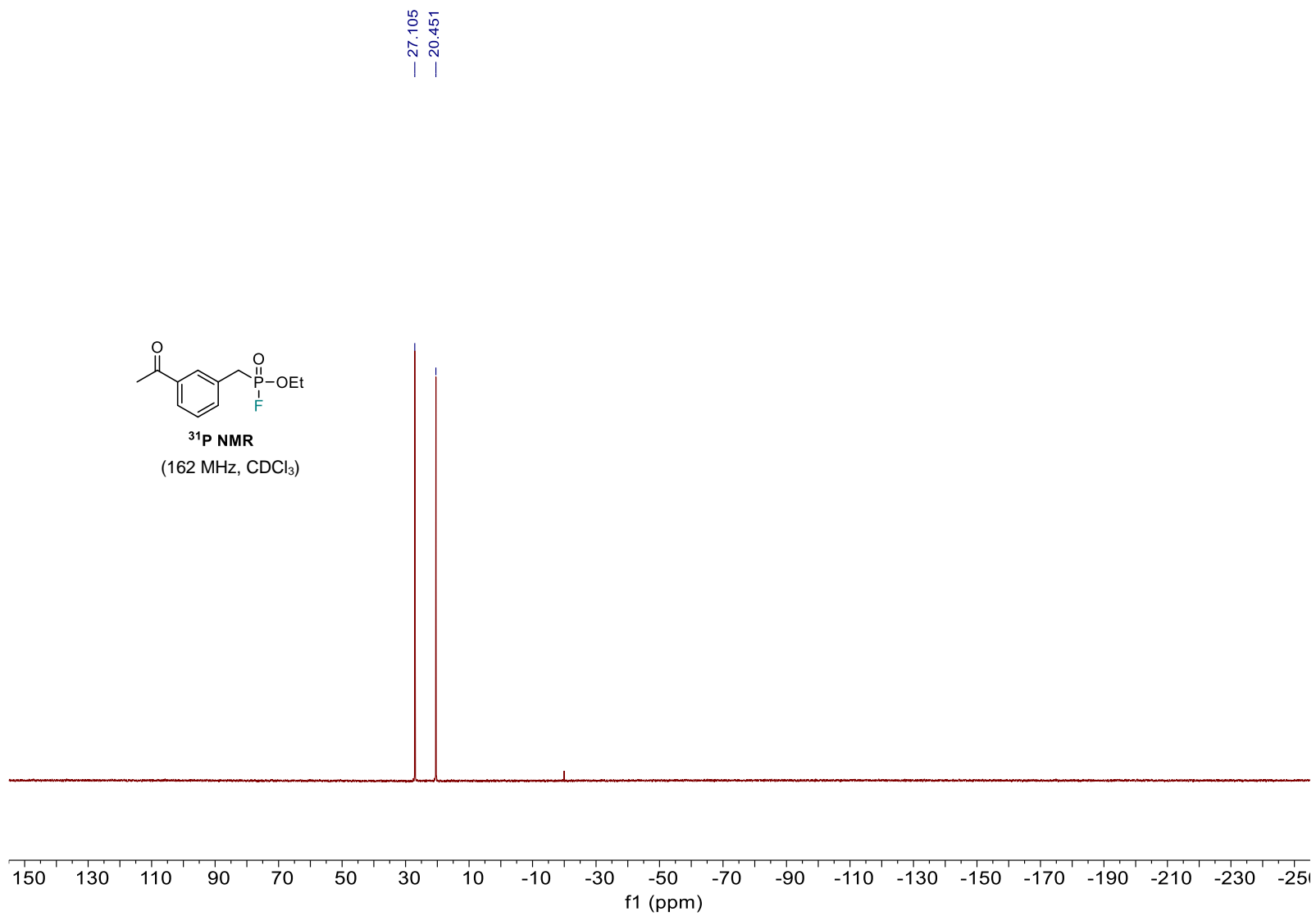
¹H NMR
(400 MHz, CDCl₃)

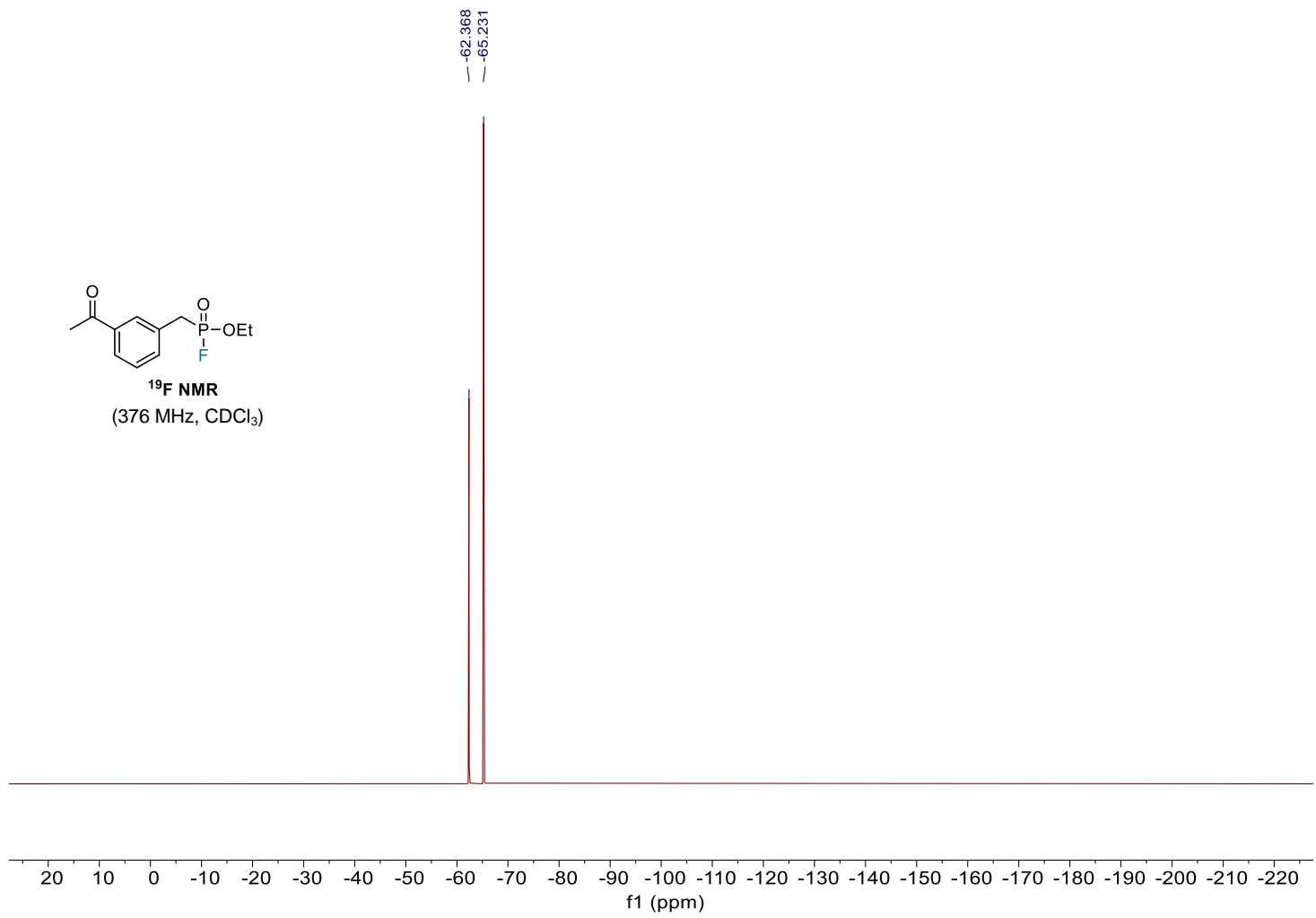
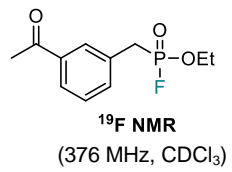


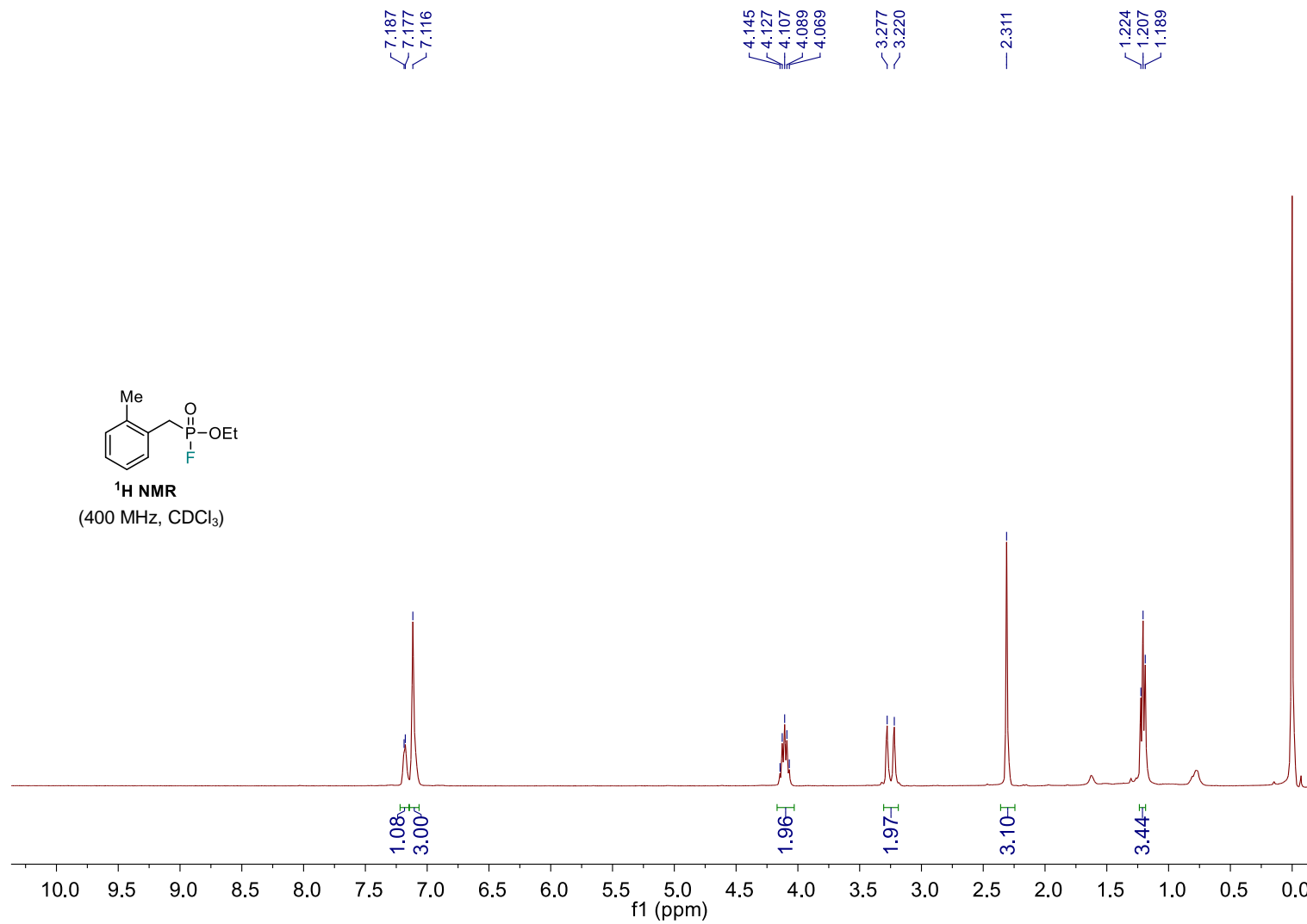
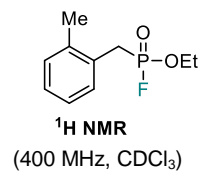


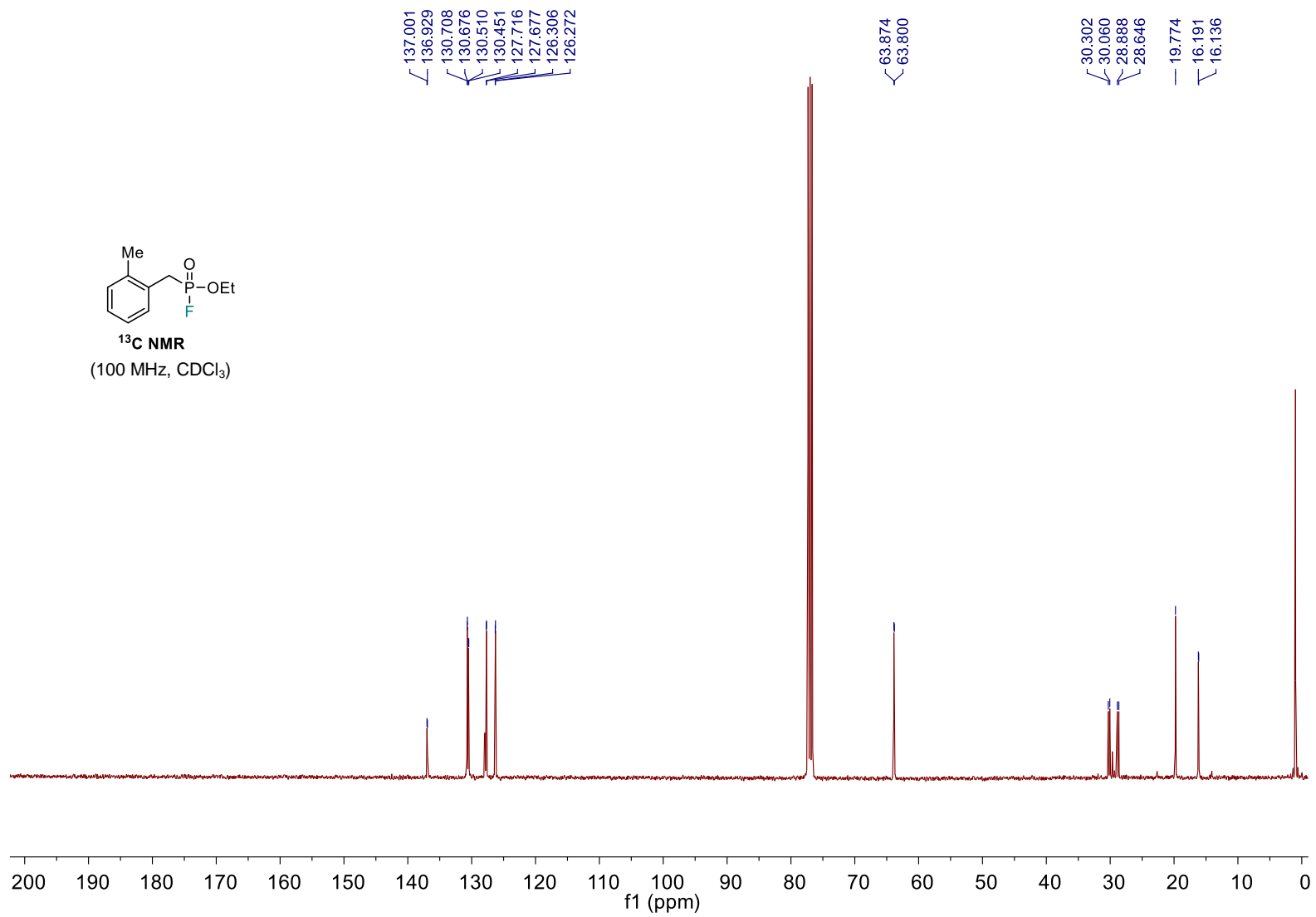
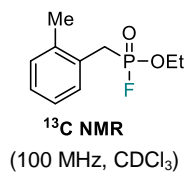


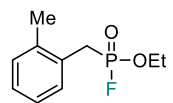
³¹P NMR
(162 MHz, CDCl₃)



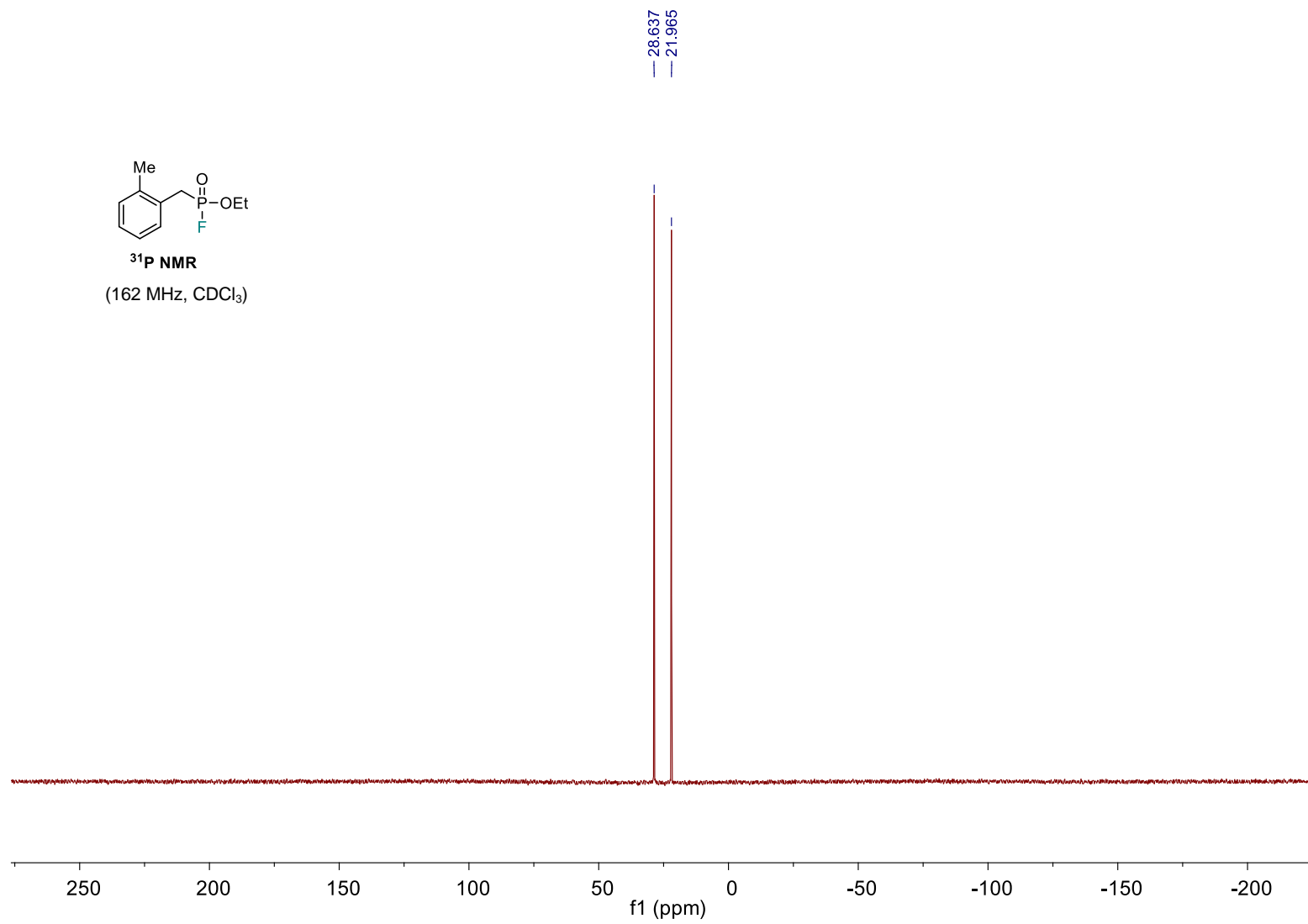




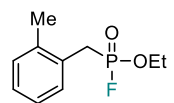




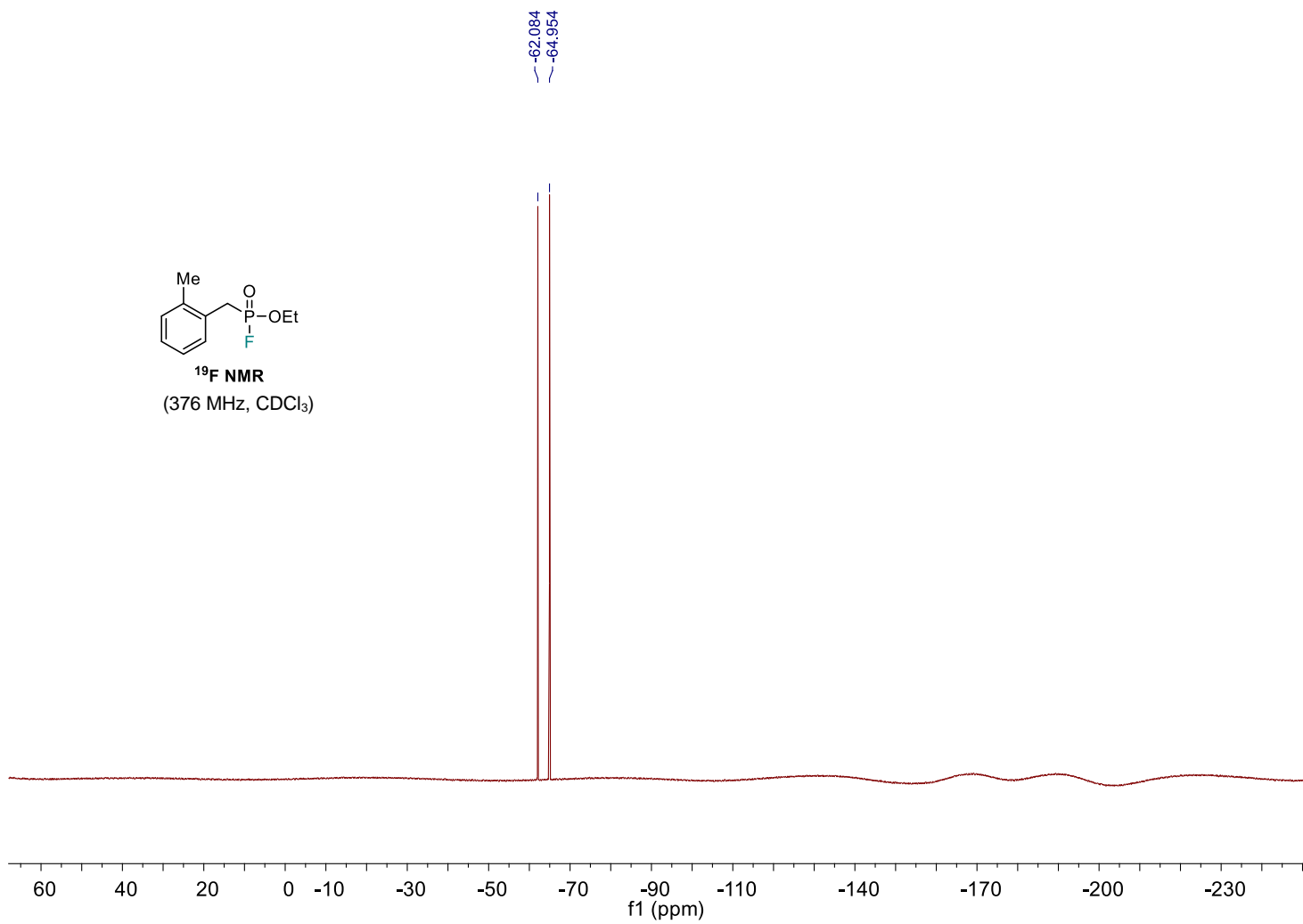
³¹P NMR
(162 MHz, CDCl₃)



S303



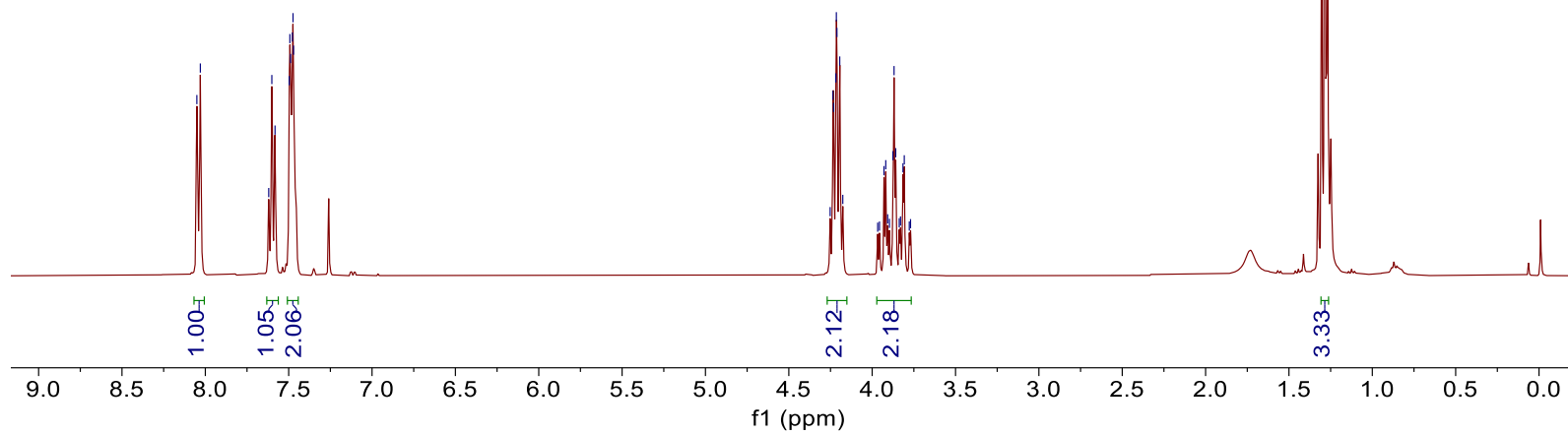
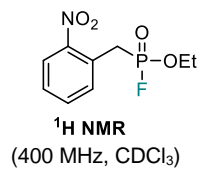
¹⁹F NMR
(376 MHz, CDCl₃)



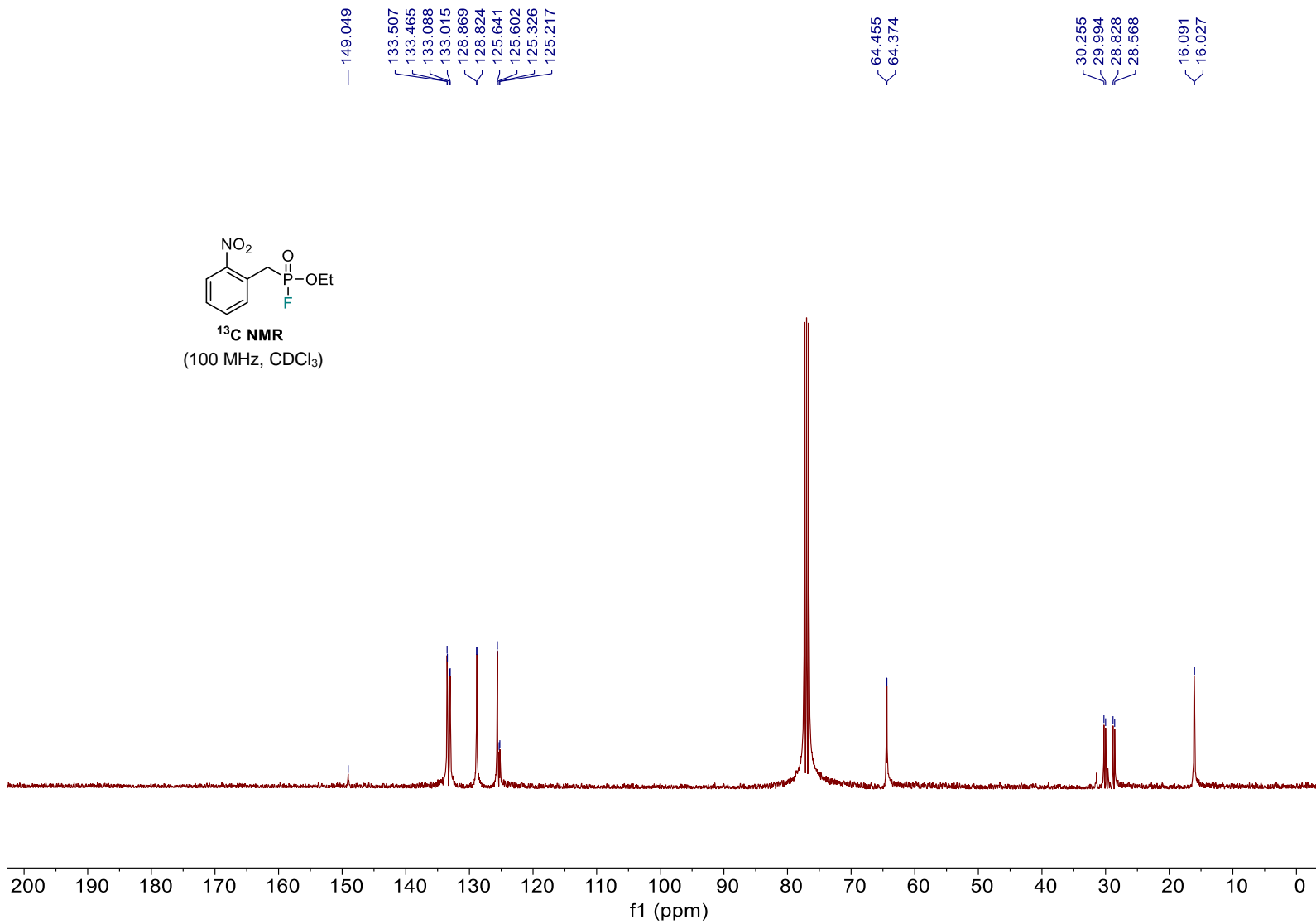
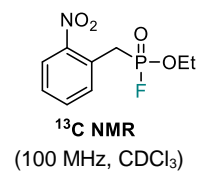
8.051
8.030
7.620
7.601
7.582
7.497
7.493
7.489
7.478
7.474
7.471

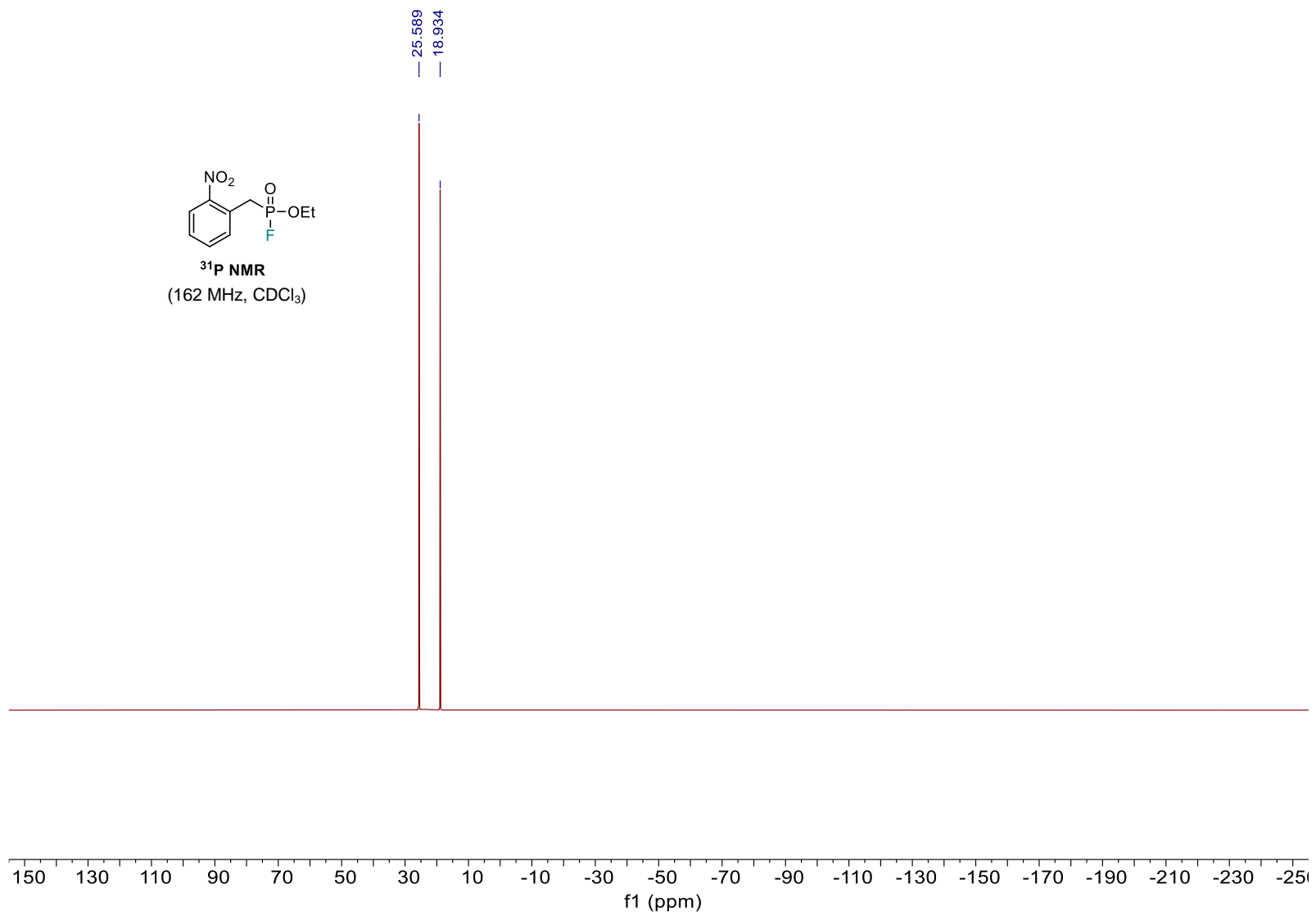
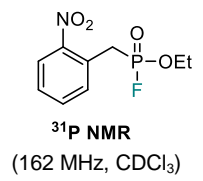
4.252
4.235
4.232
4.218
4.215
4.211
4.194
4.176
3.967
3.956
3.929
3.919
3.907
3.896
3.875
3.869
3.859
3.838
3.830
3.815
3.807
3.777
3.770

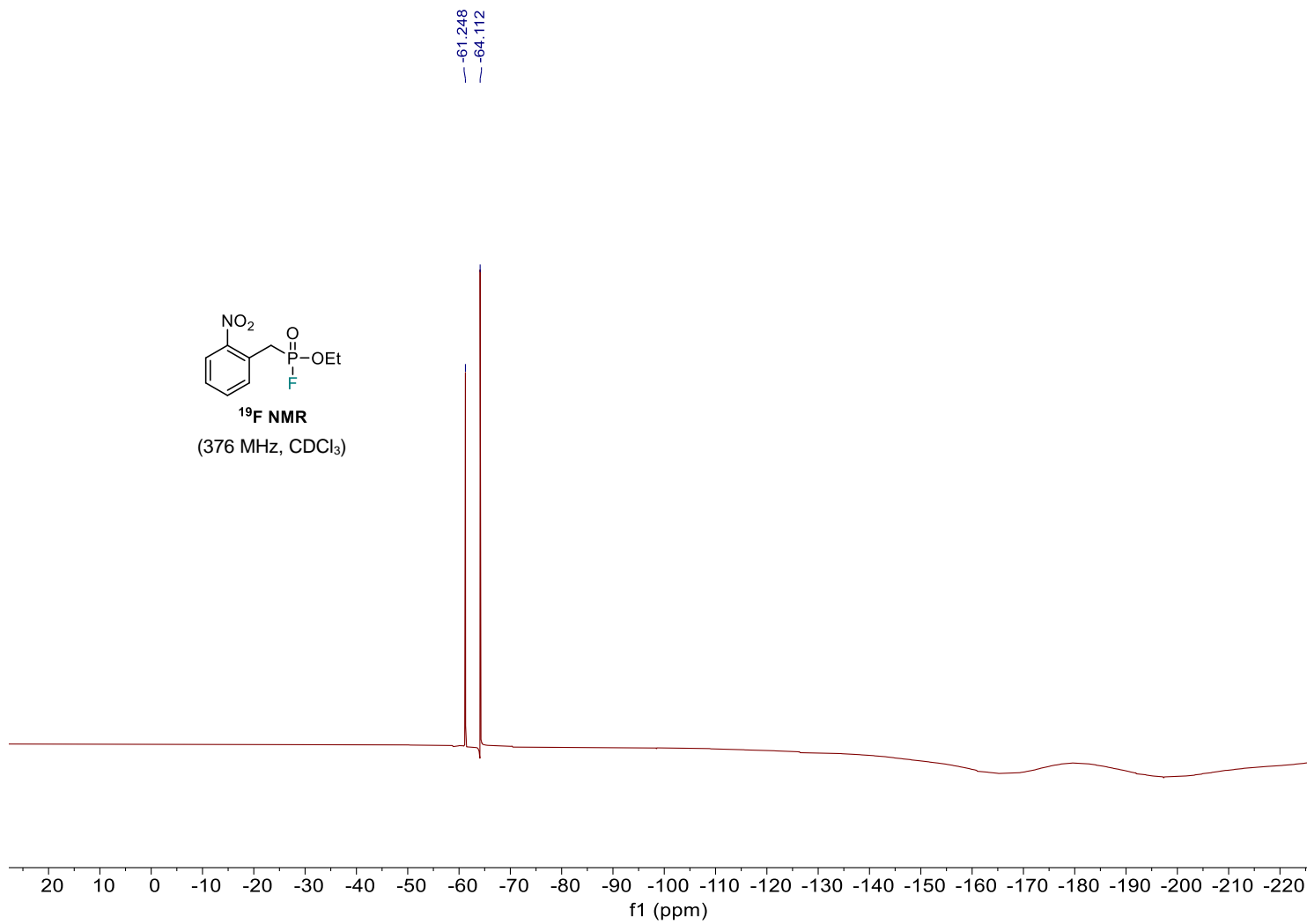
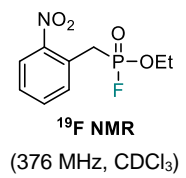
1.304
1.286
1.268

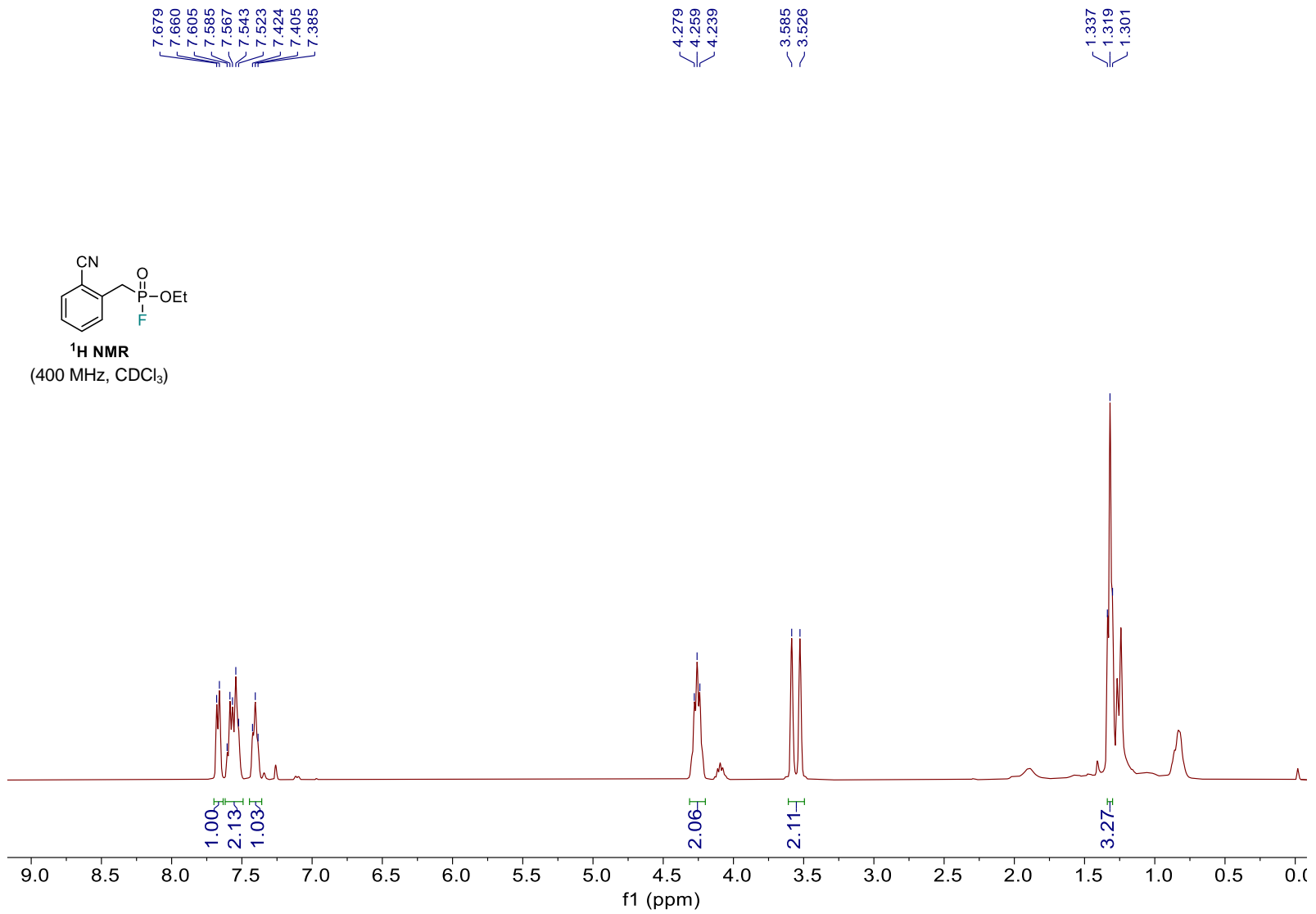
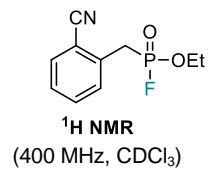


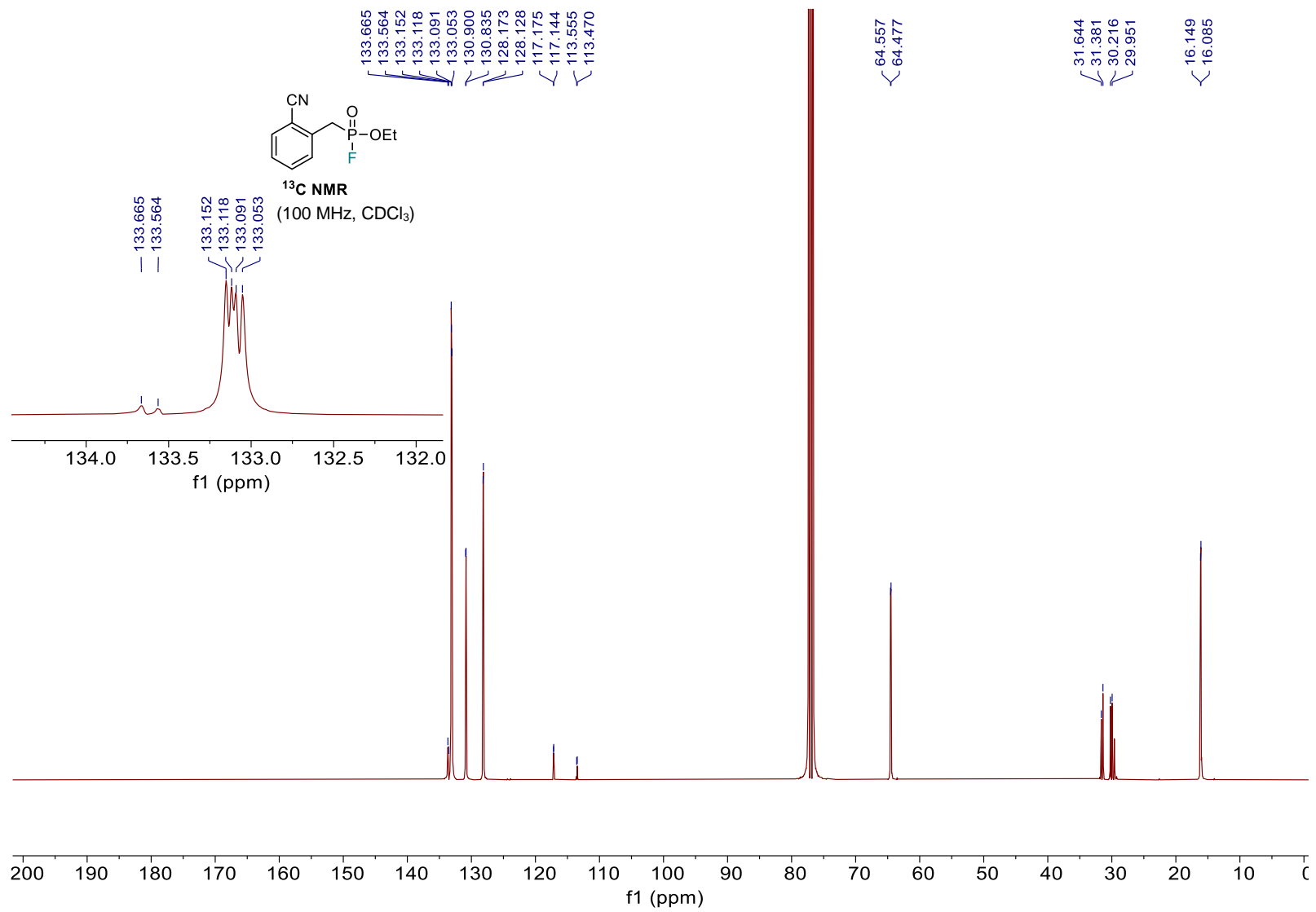
S305

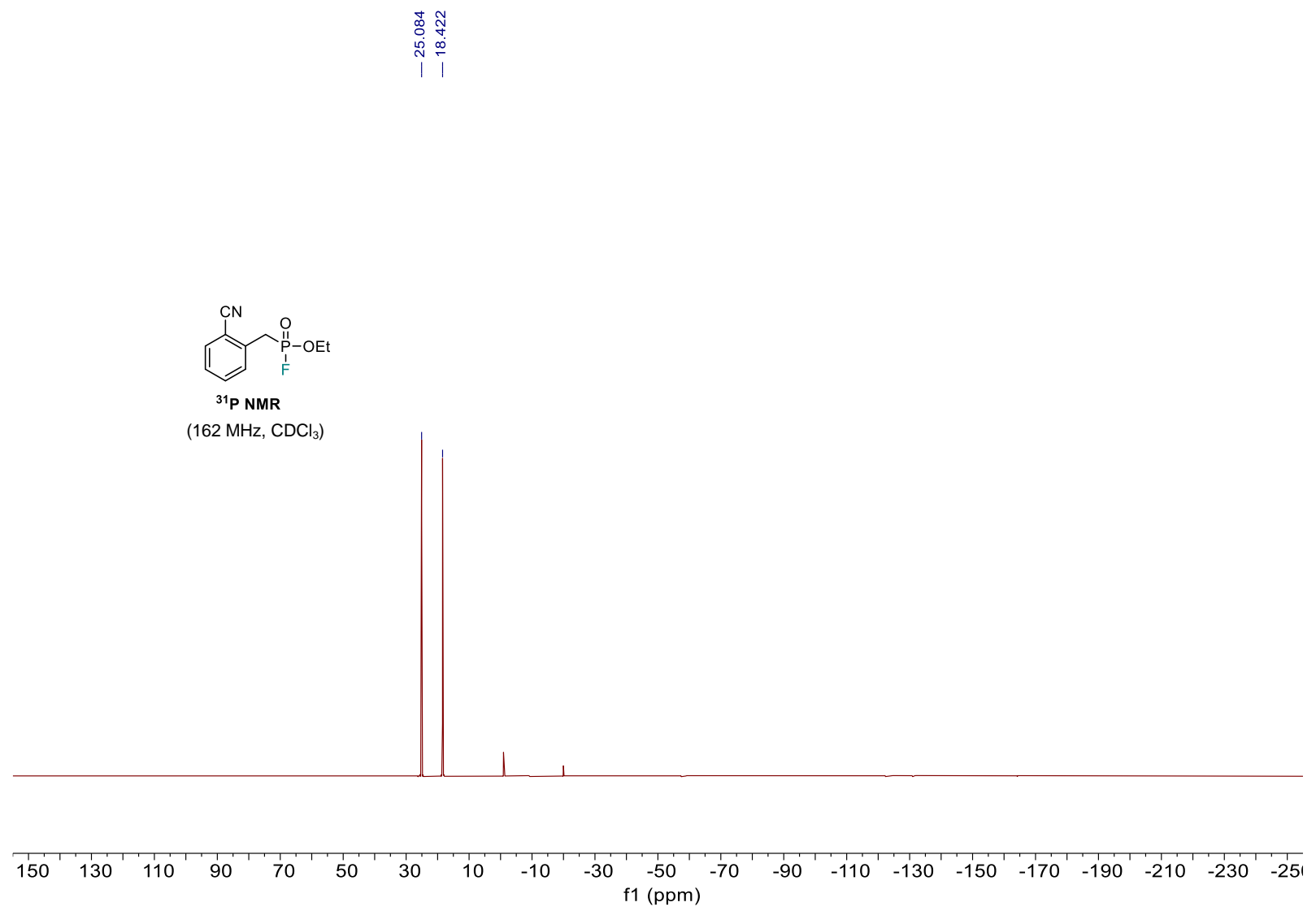
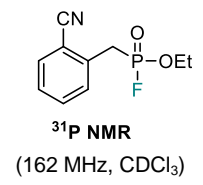


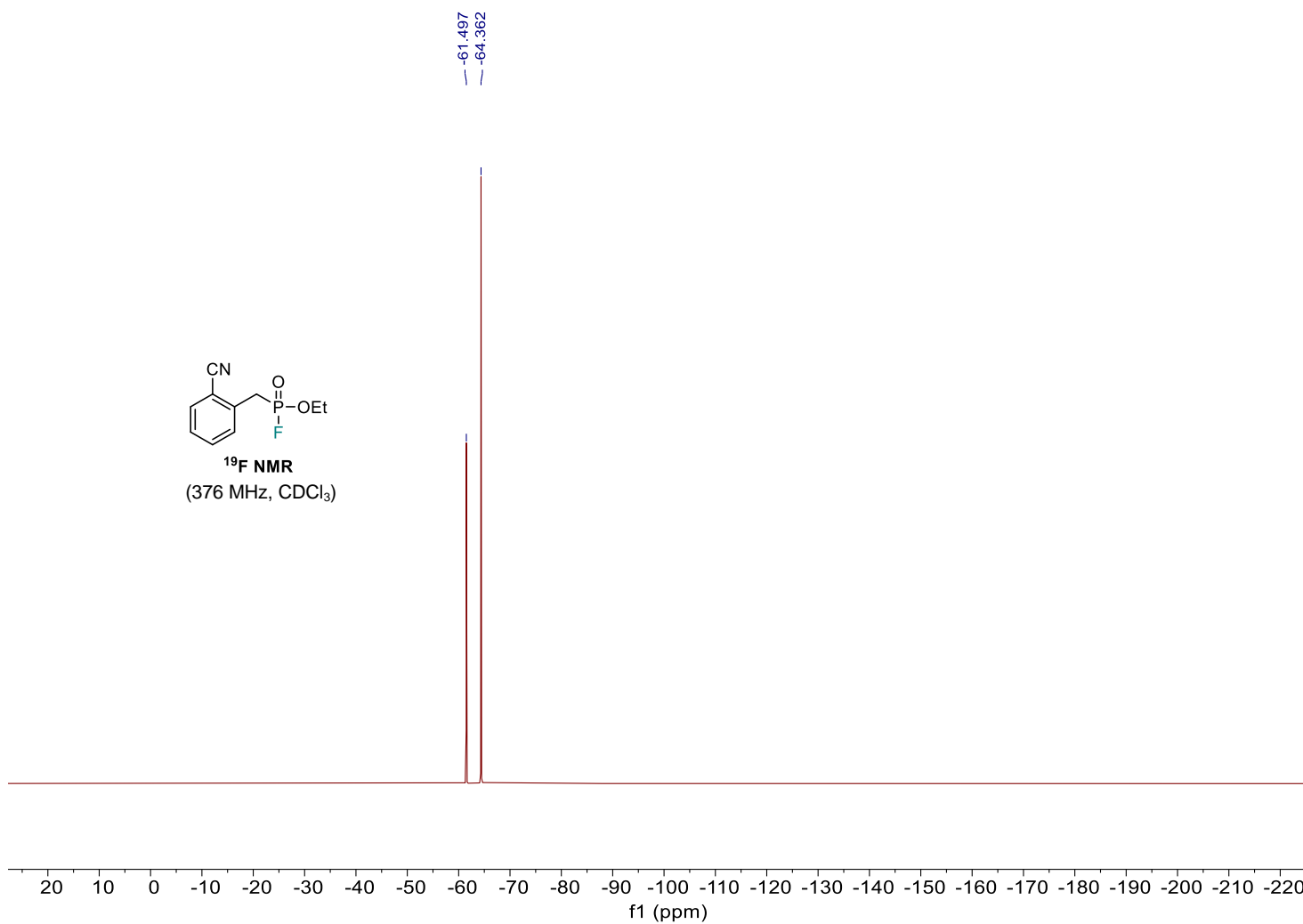
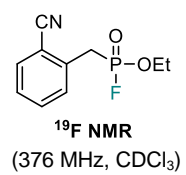


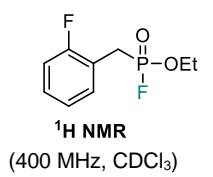
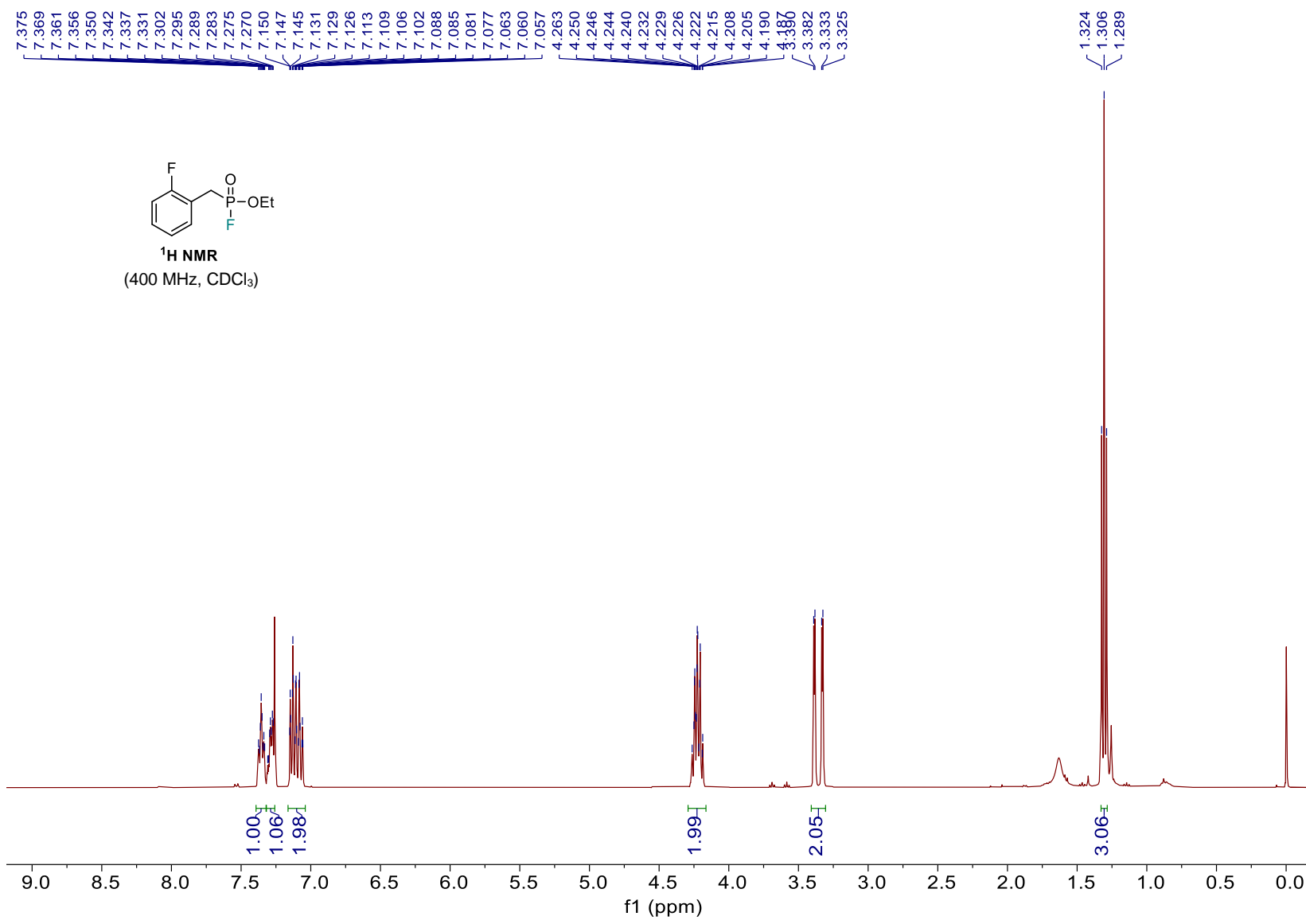


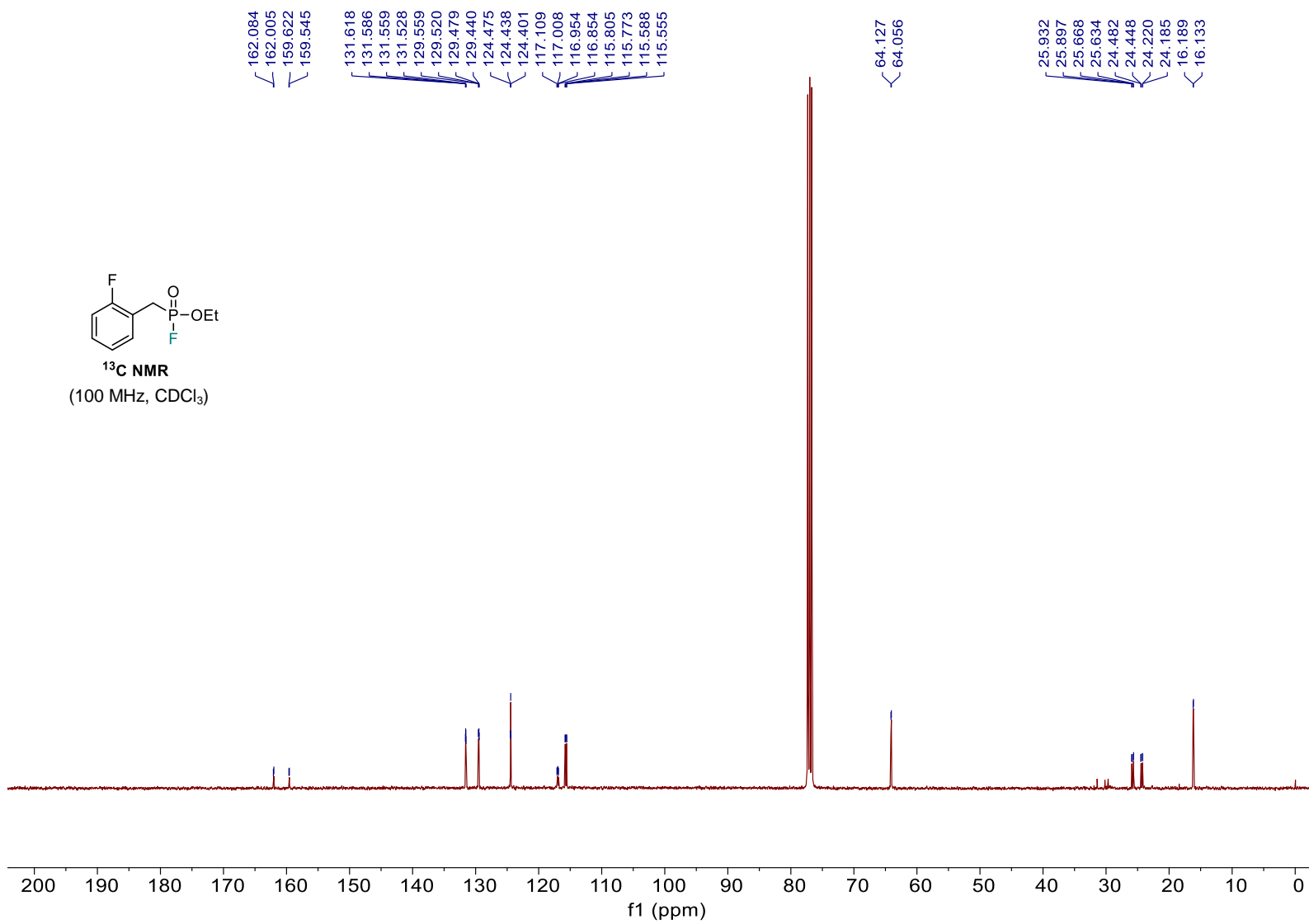
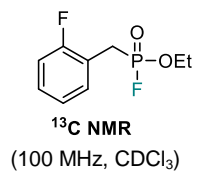


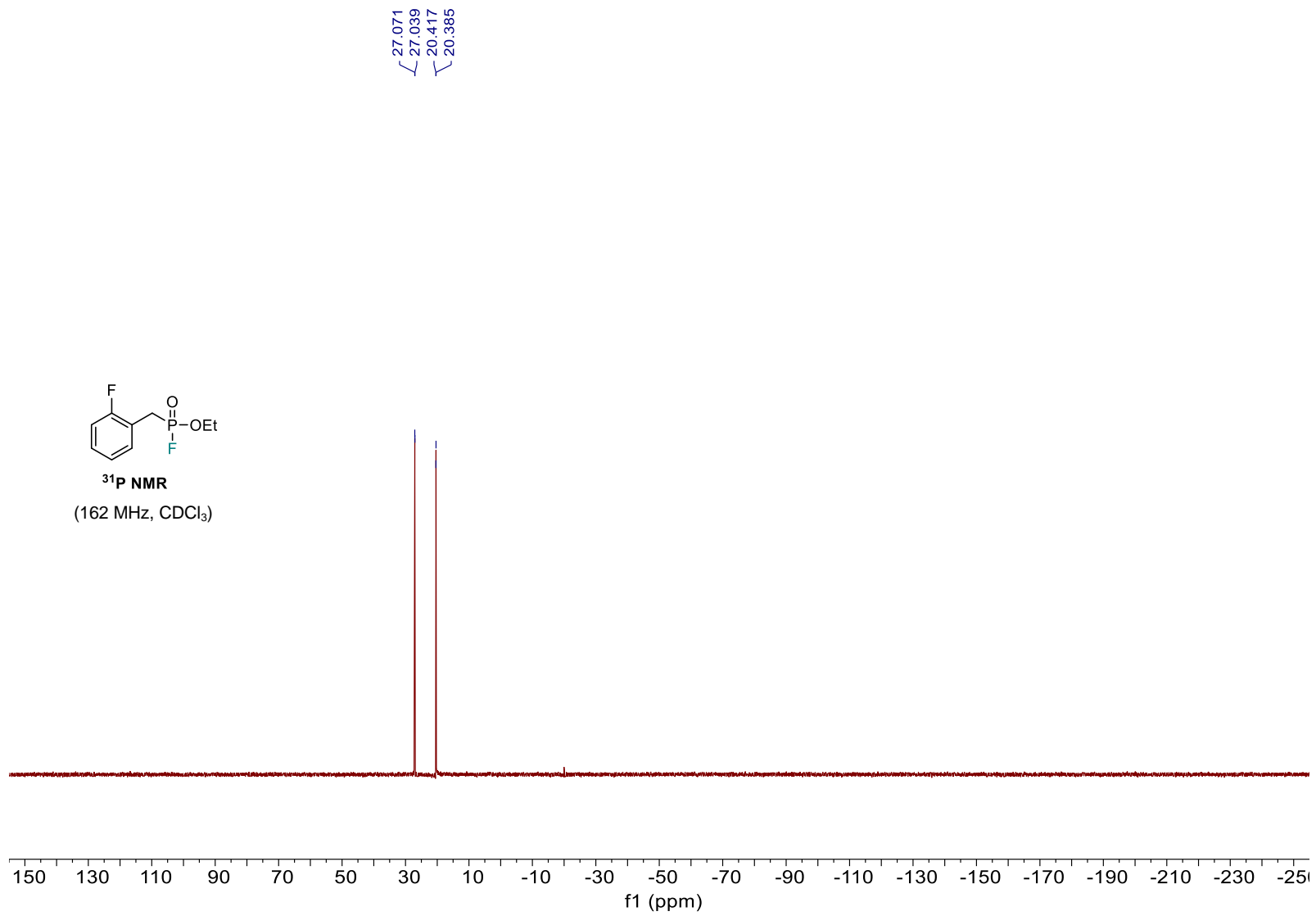
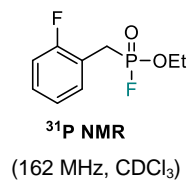


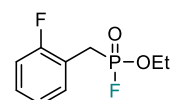




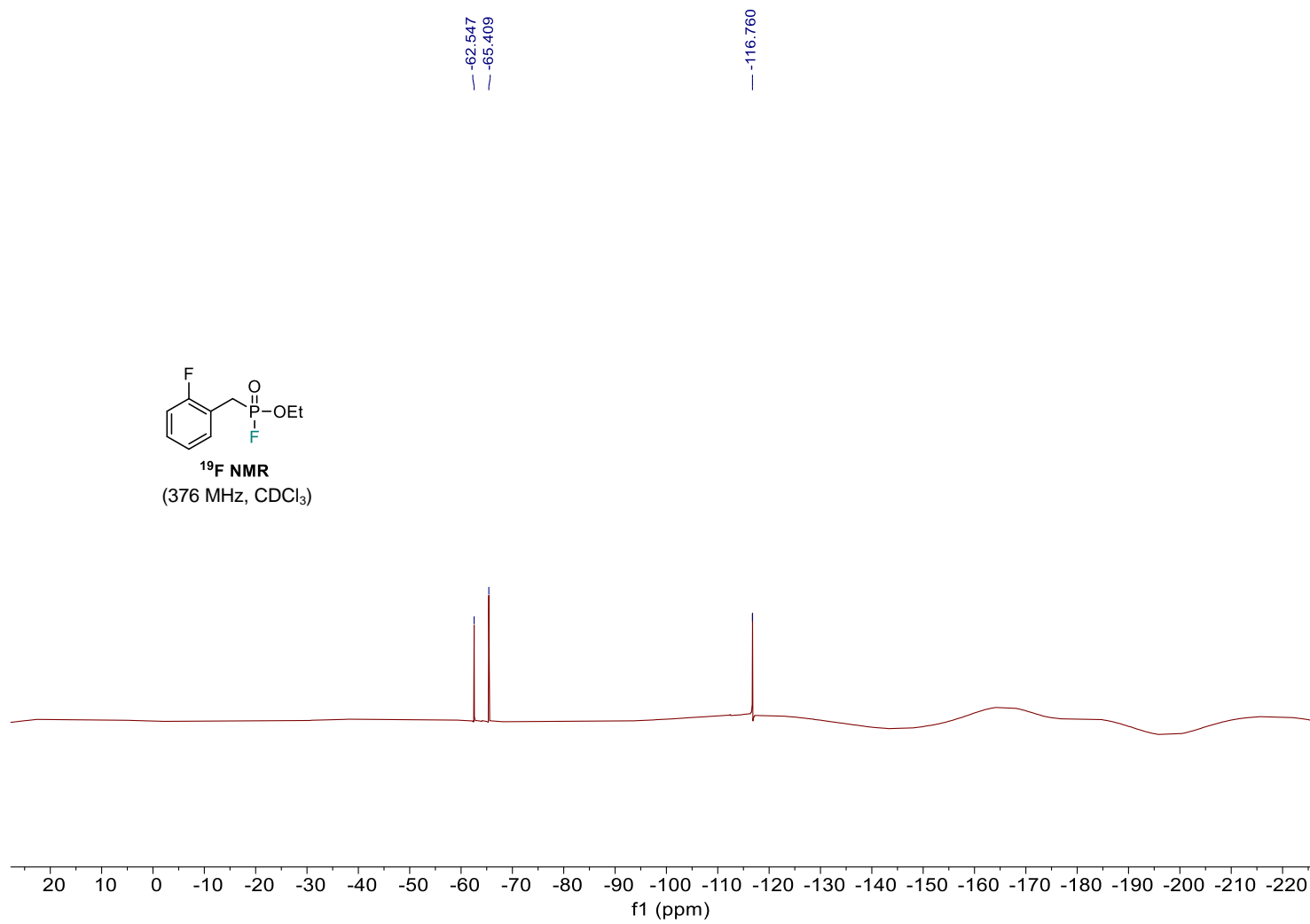


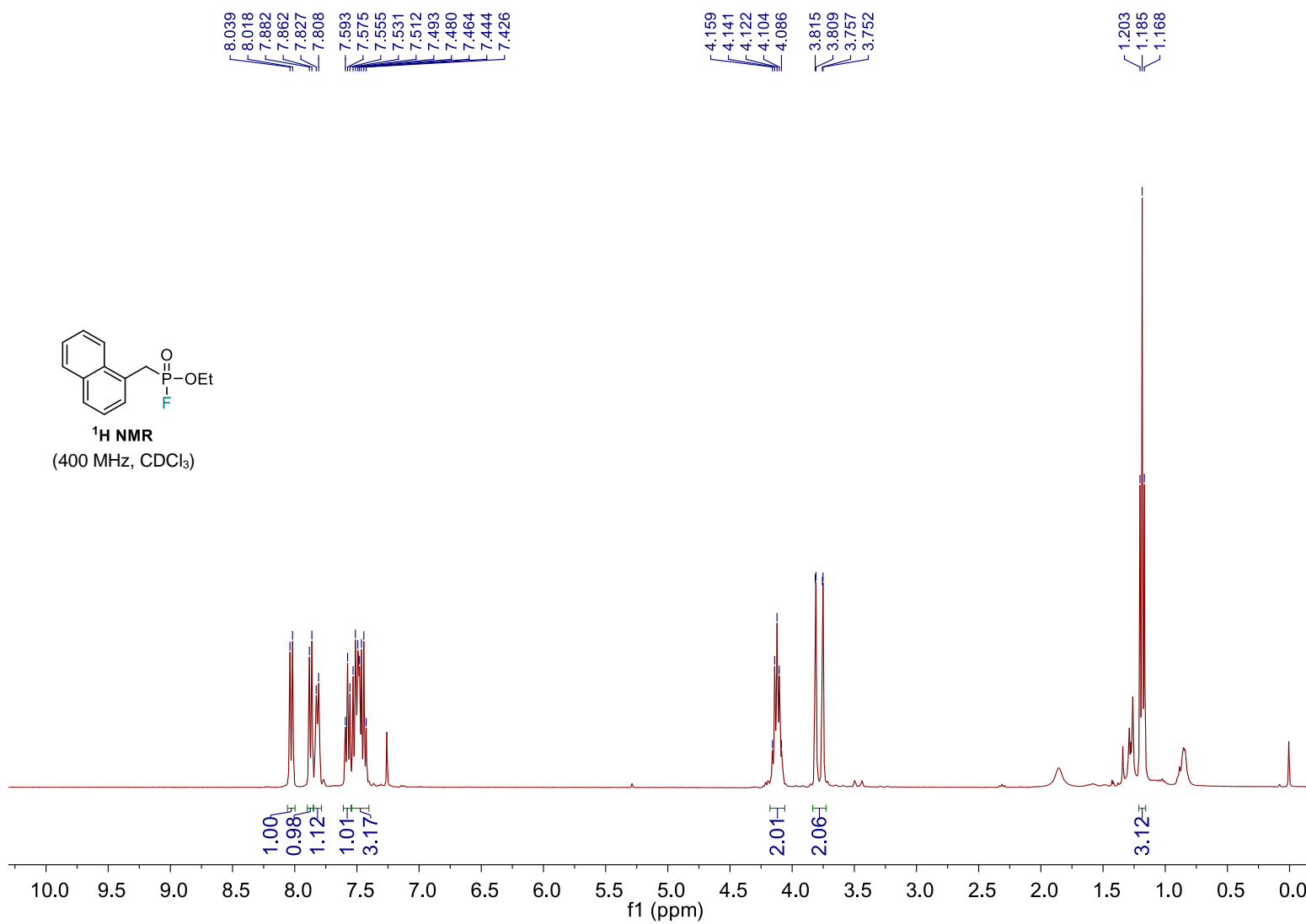


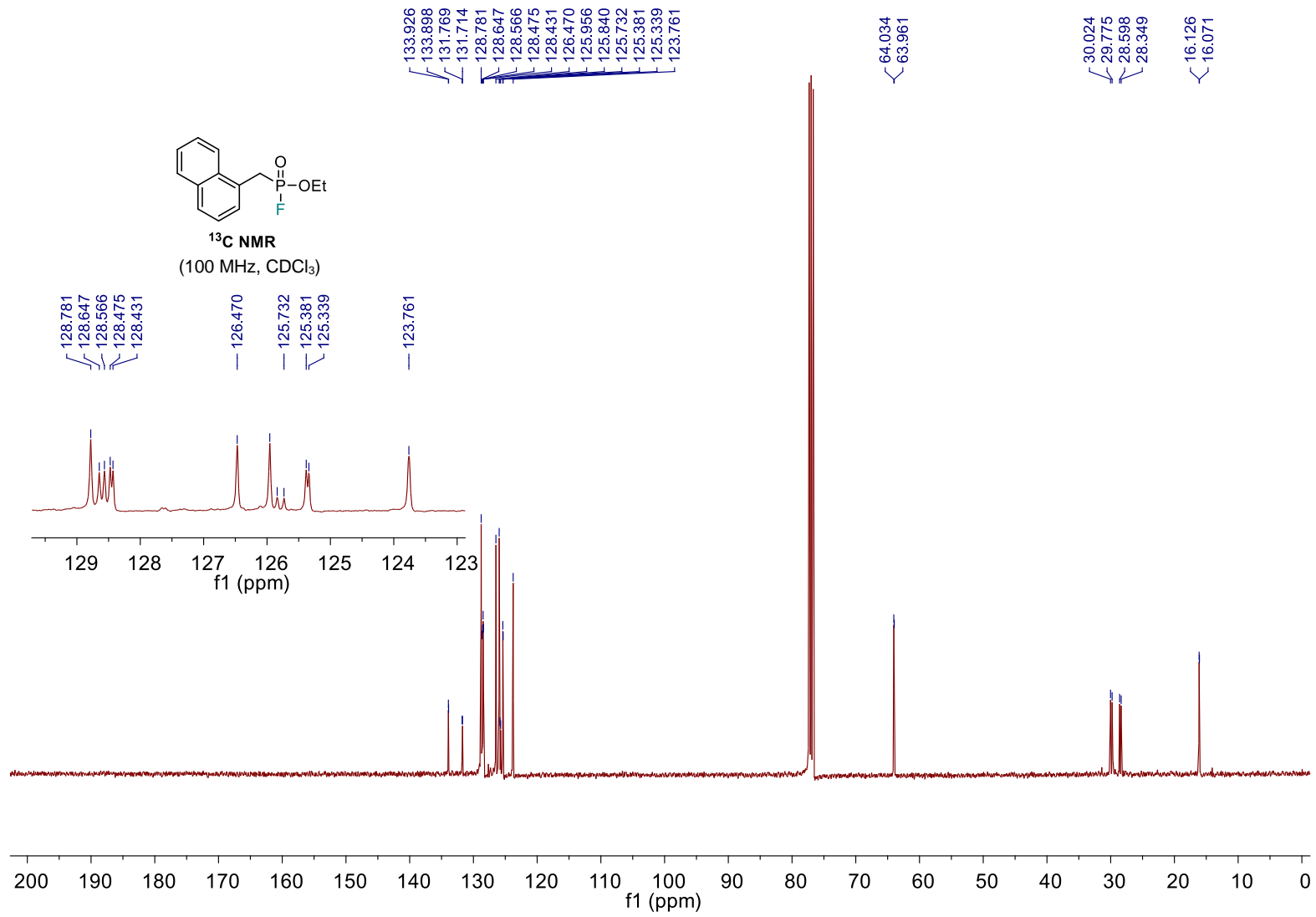


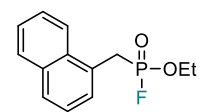


¹⁹F NMR
(376 MHz, CDCl₃)

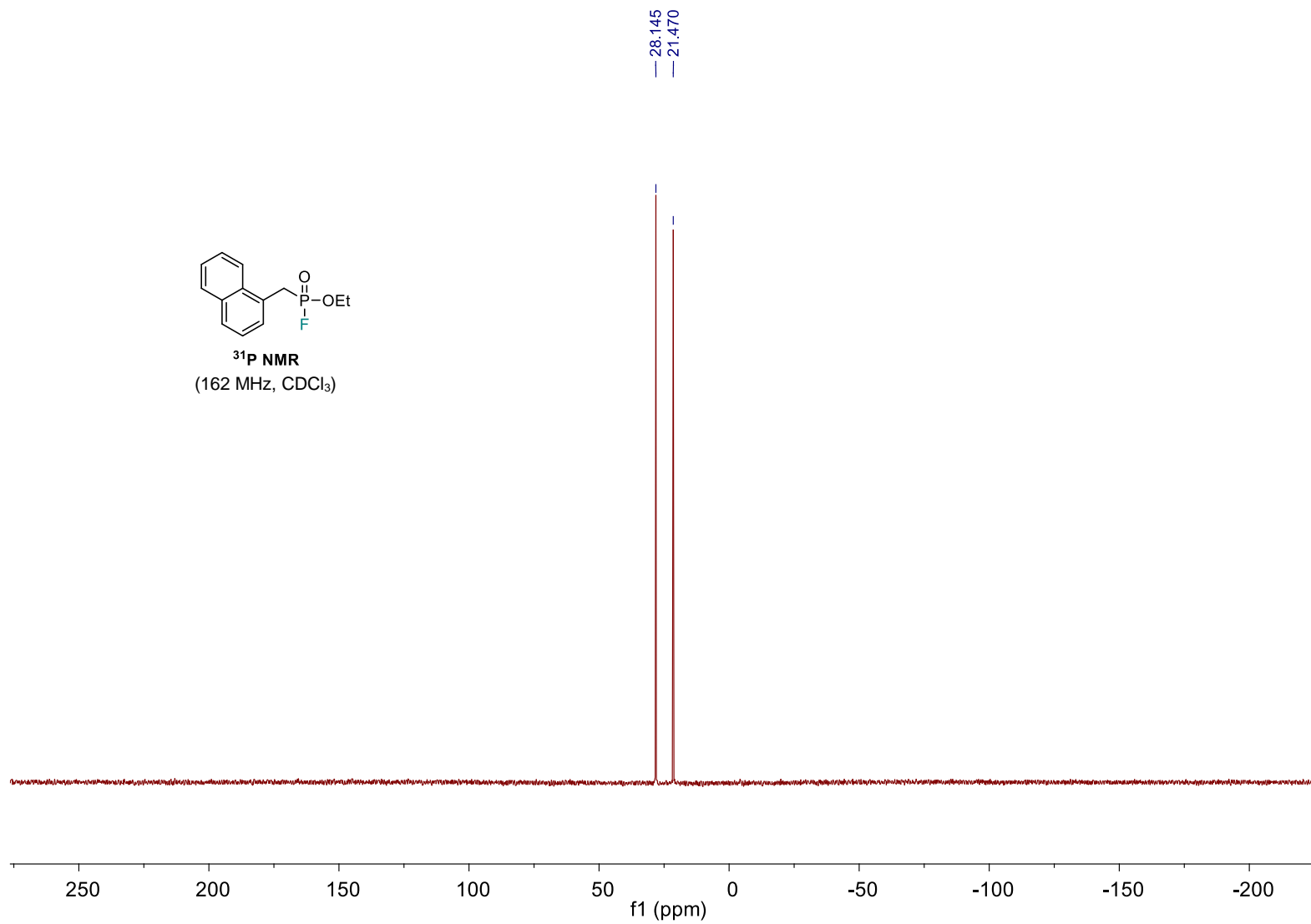




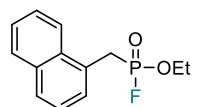




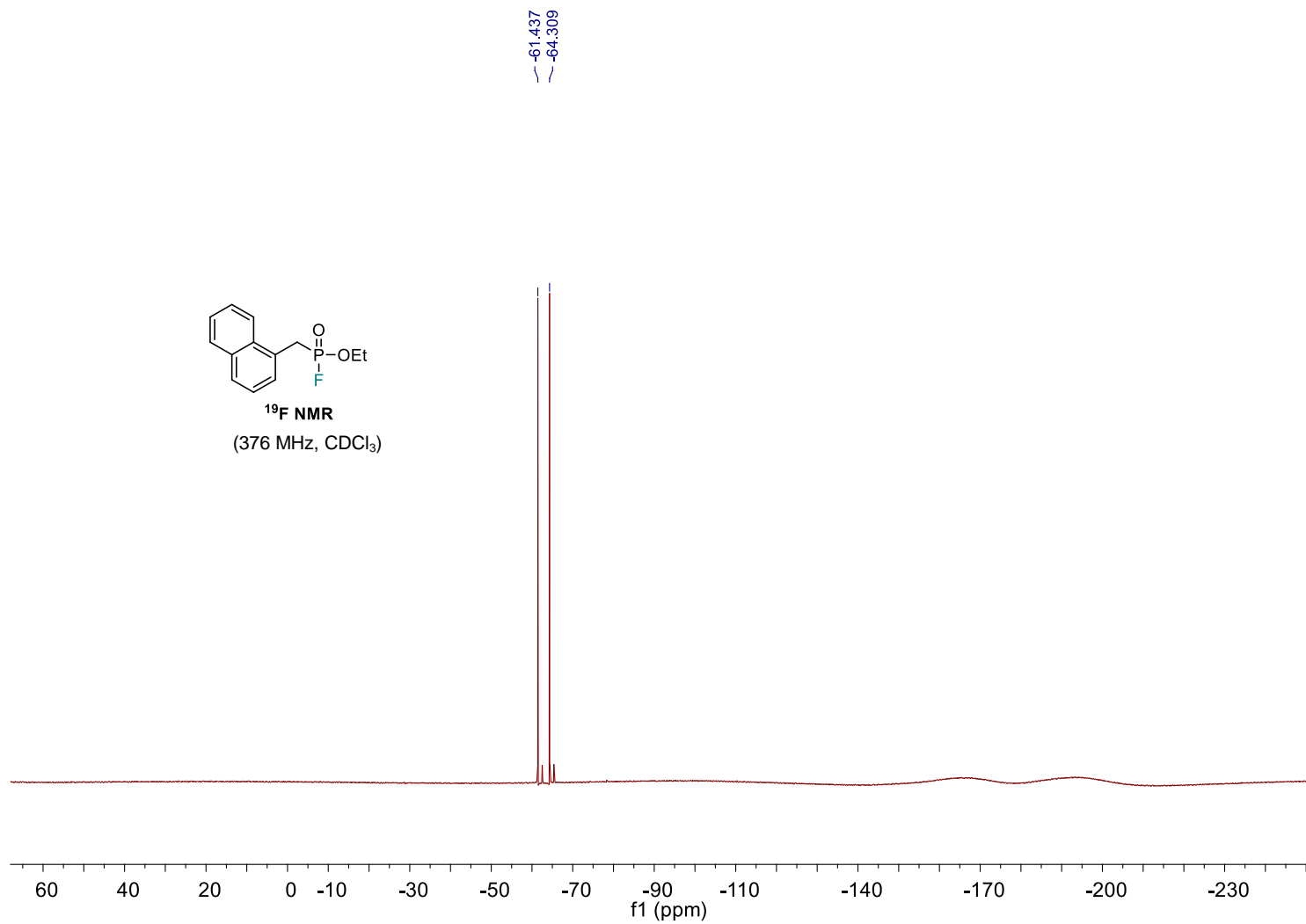
³¹P NMR
(162 MHz, CDCl₃)

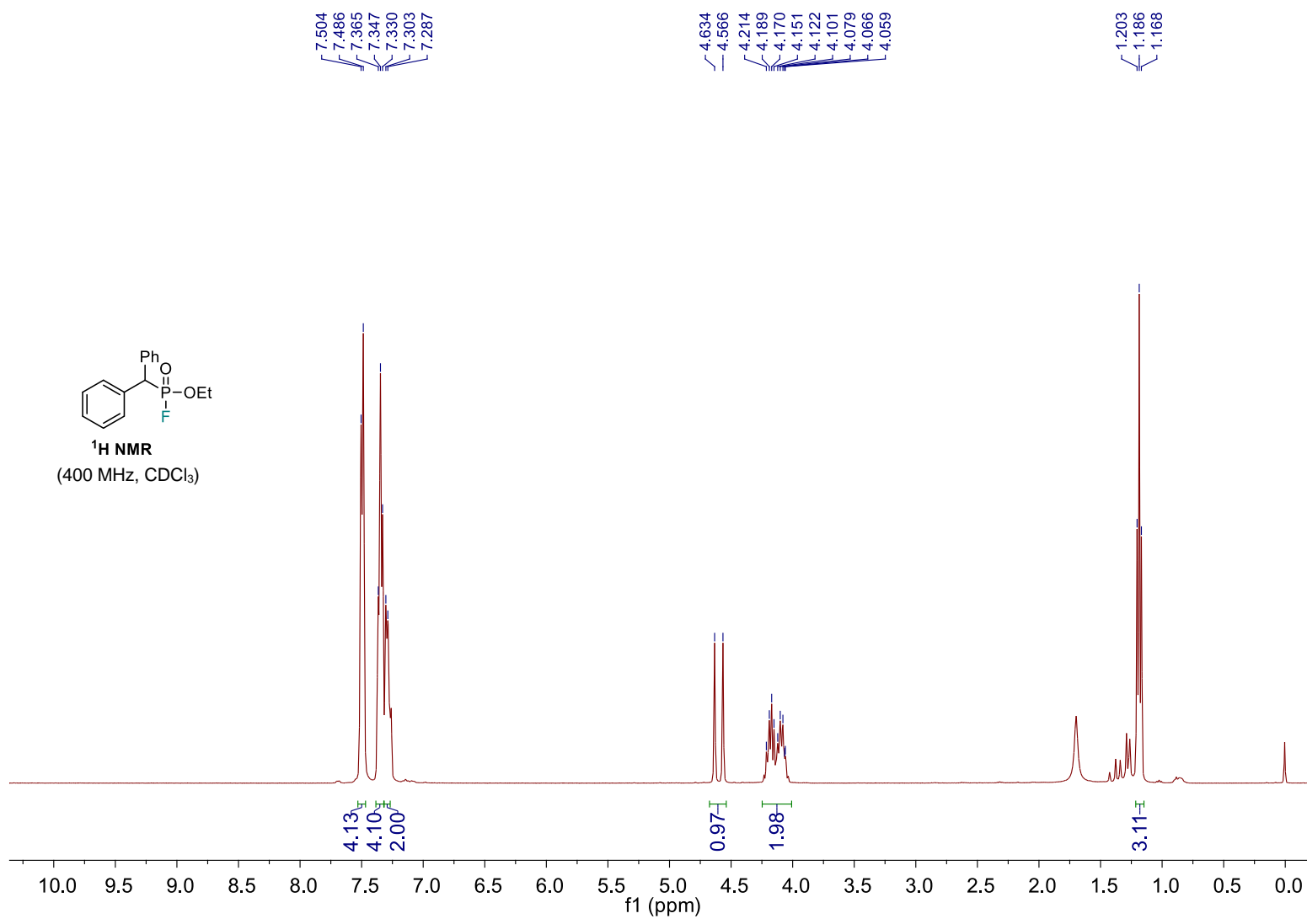


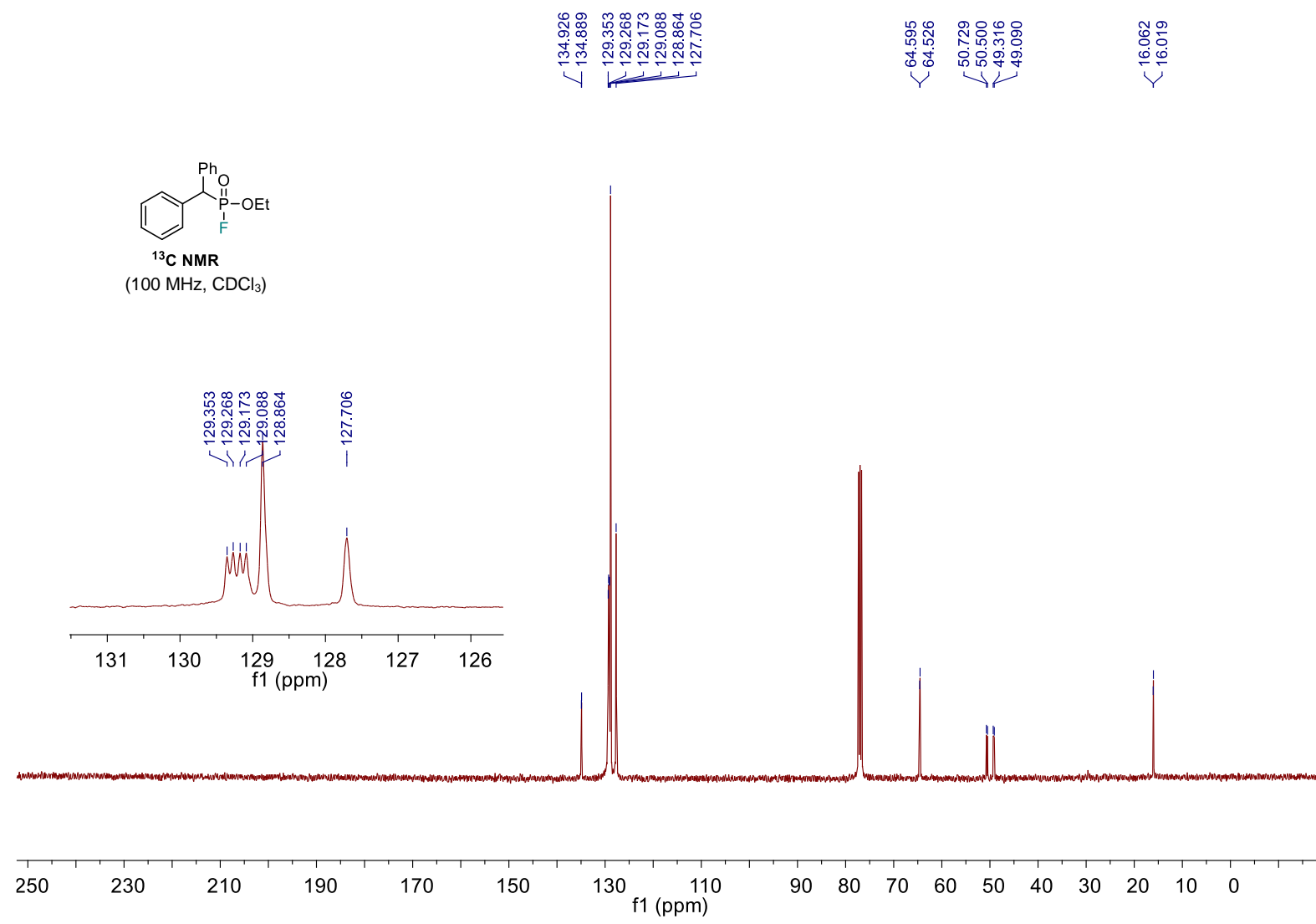
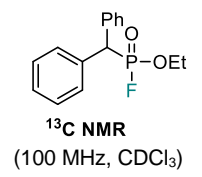
S319

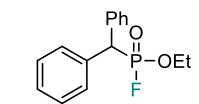


¹⁹F NMR
(376 MHz, CDCl₃)

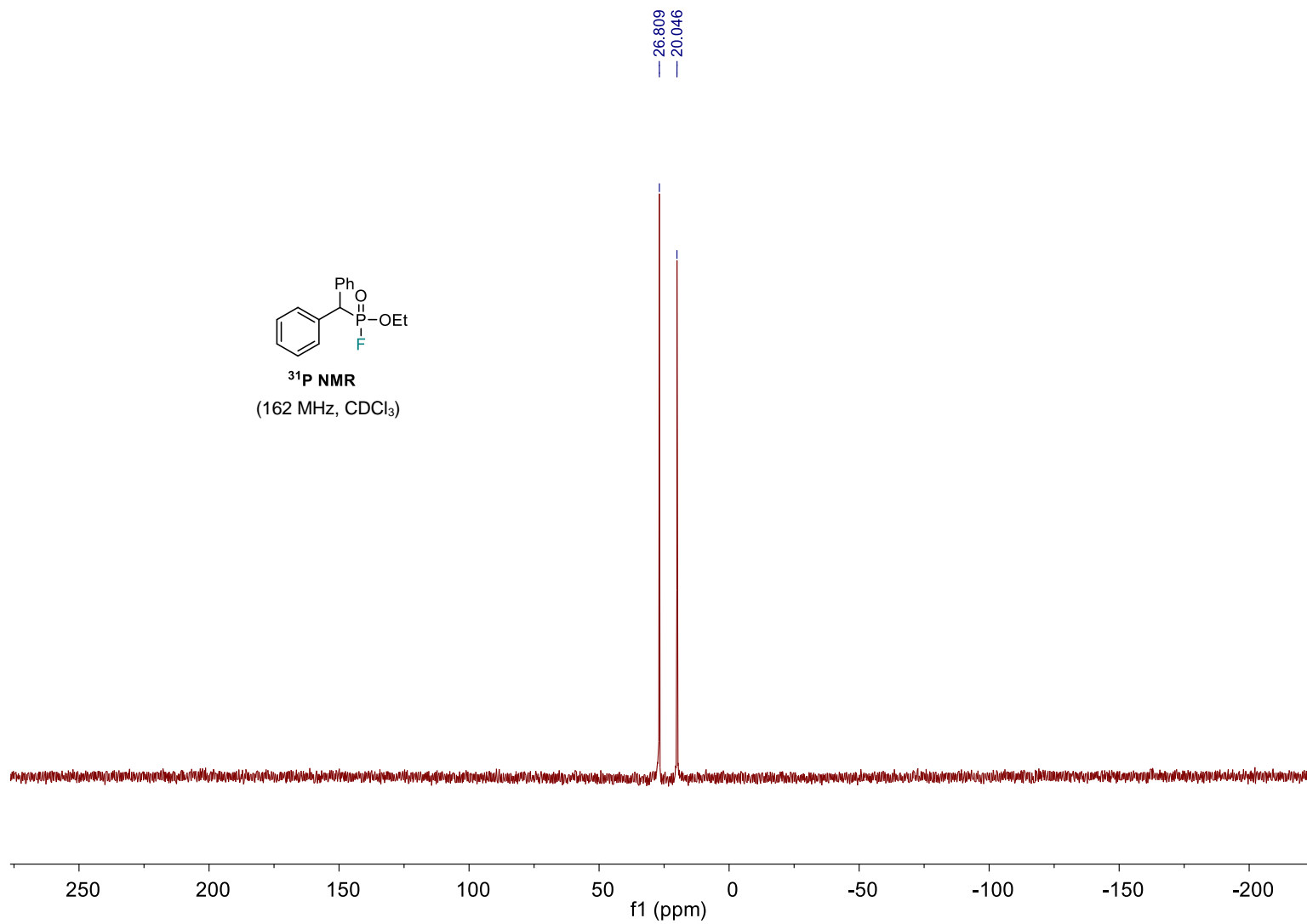




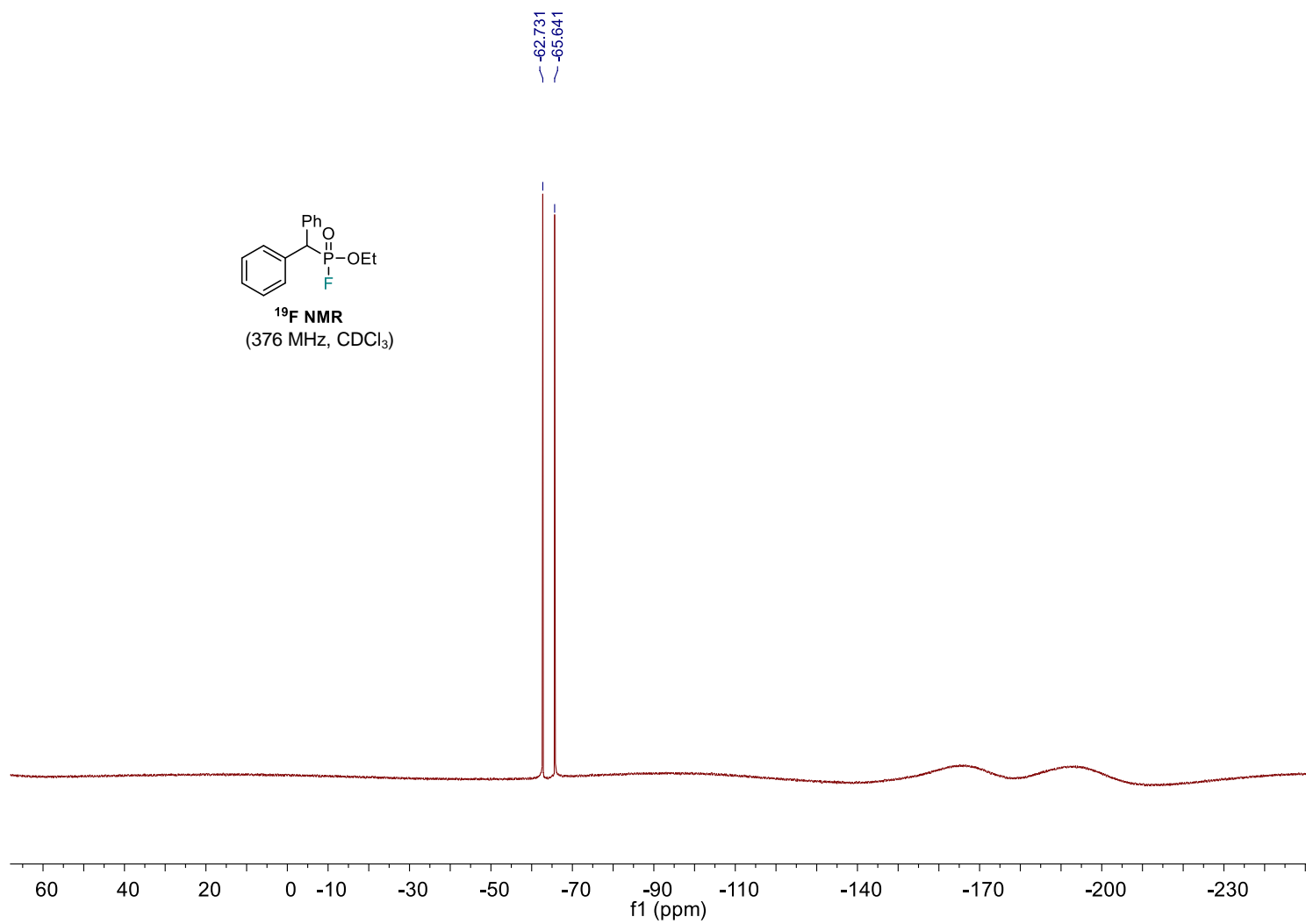
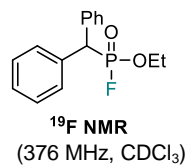


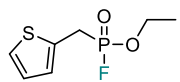


³¹P NMR
(162 MHz, CDCl₃)

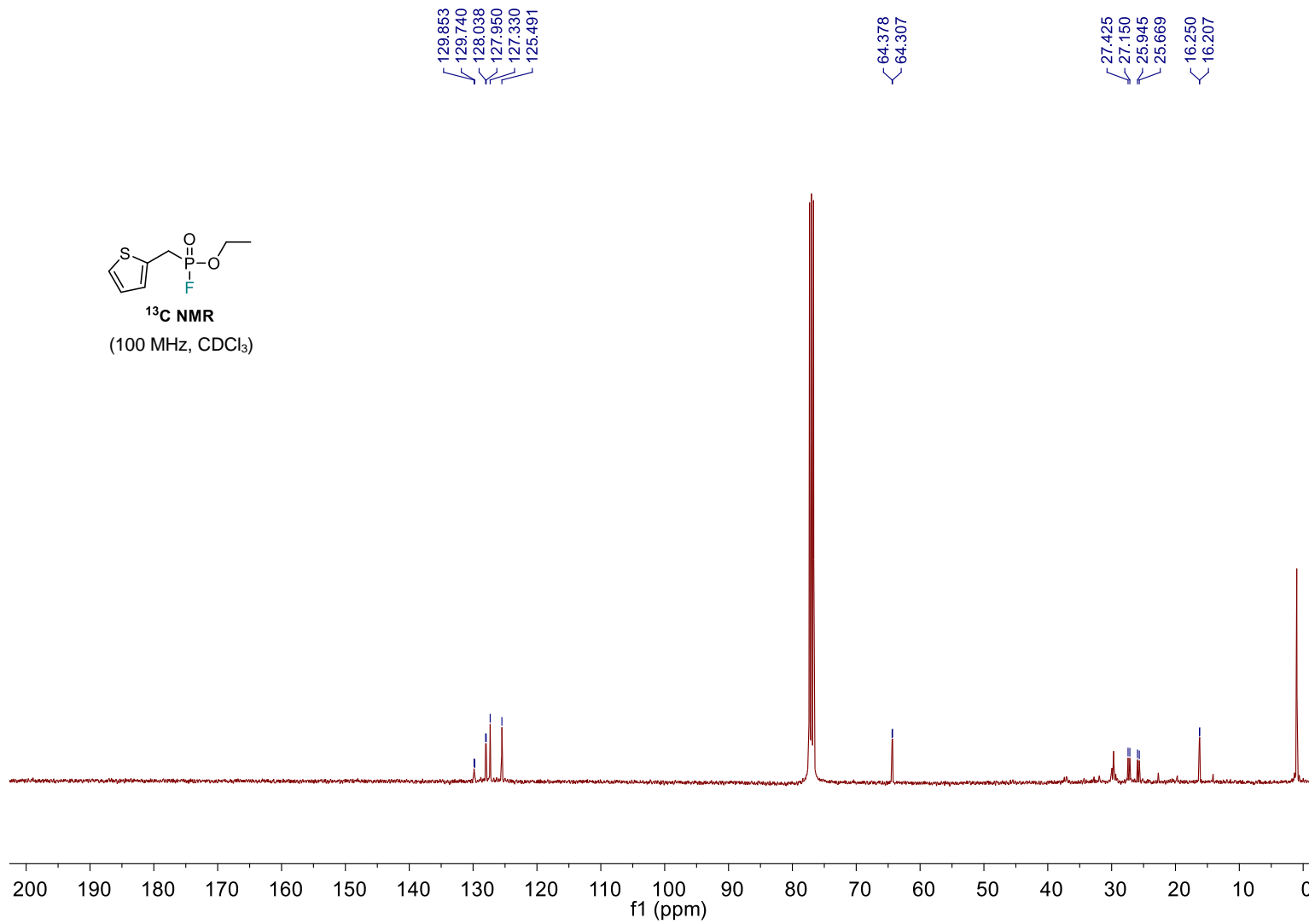


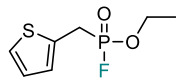
S323



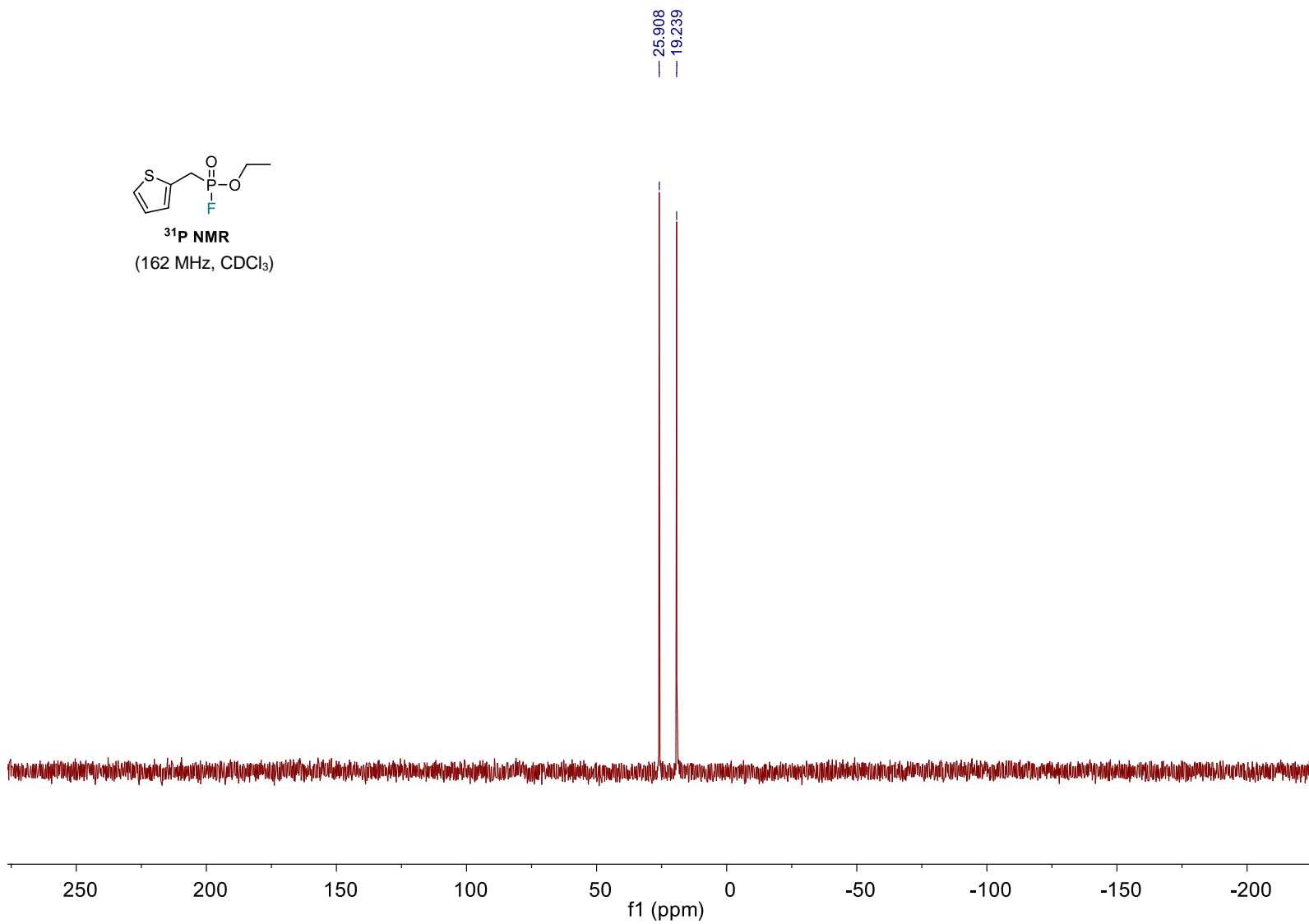


¹³C NMR
(100 MHz, CDCl₃)

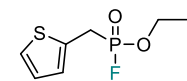




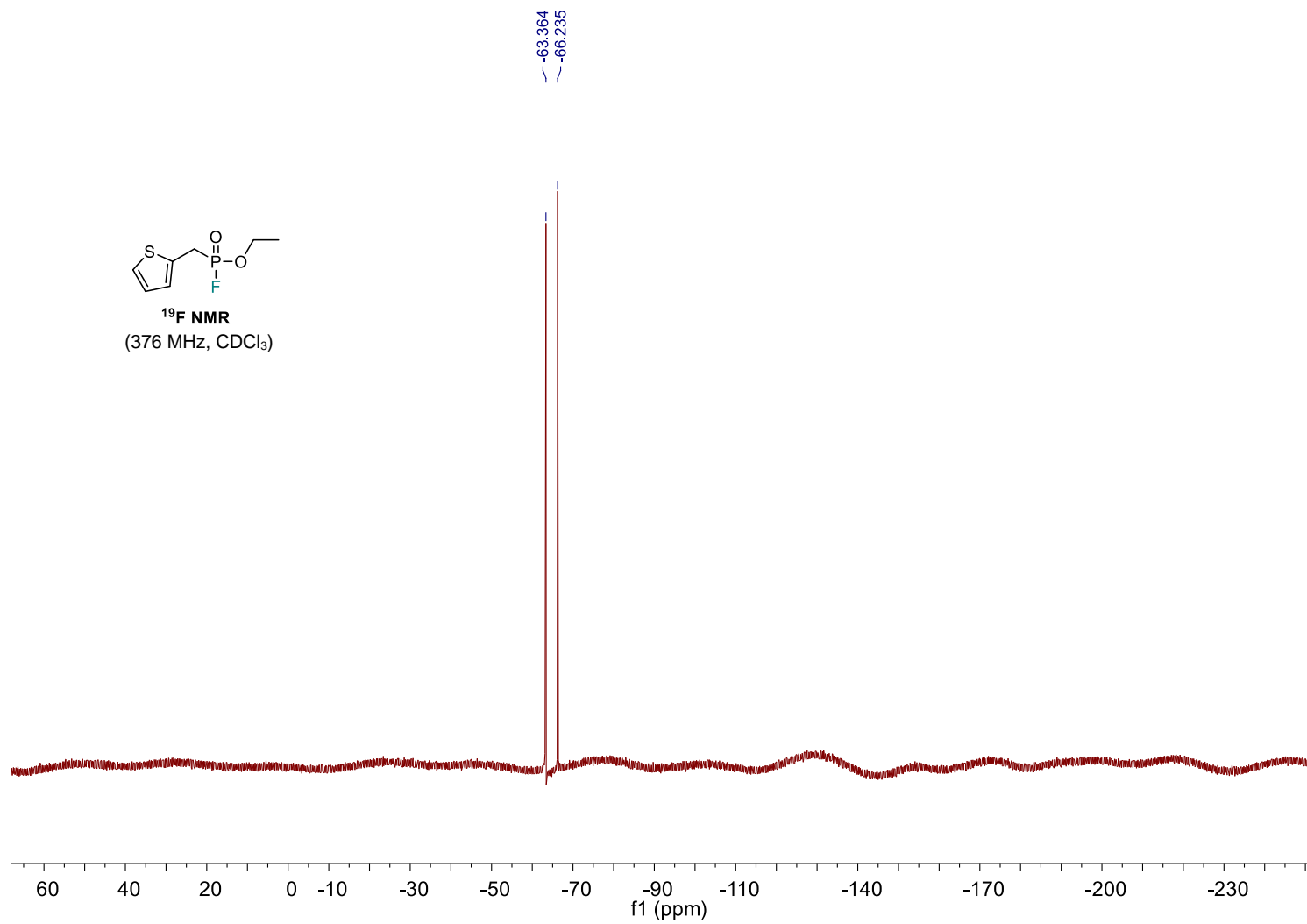
³¹P NMR
(162 MHz, CDCl₃)

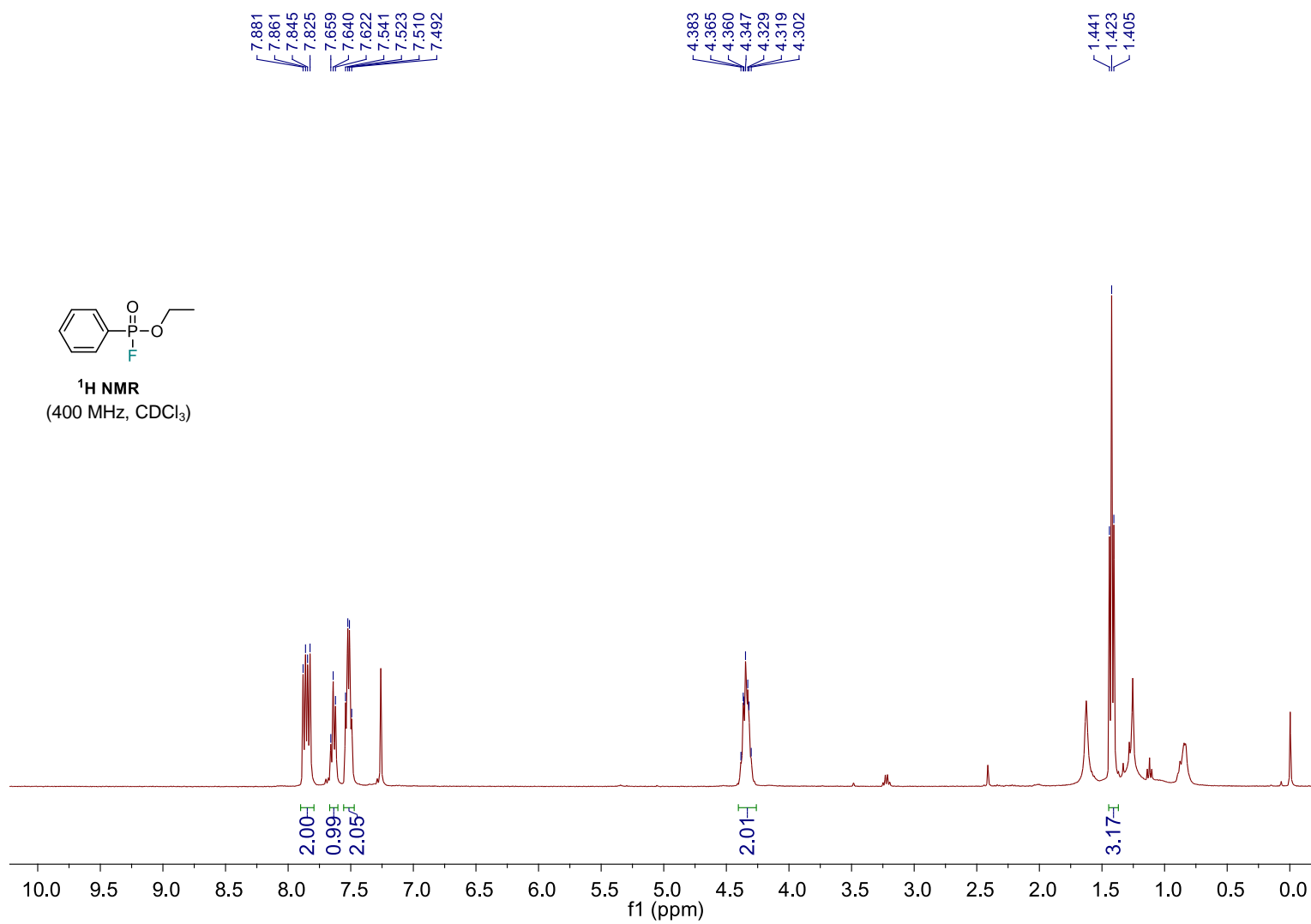
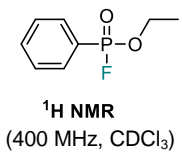


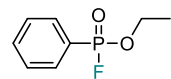
S327



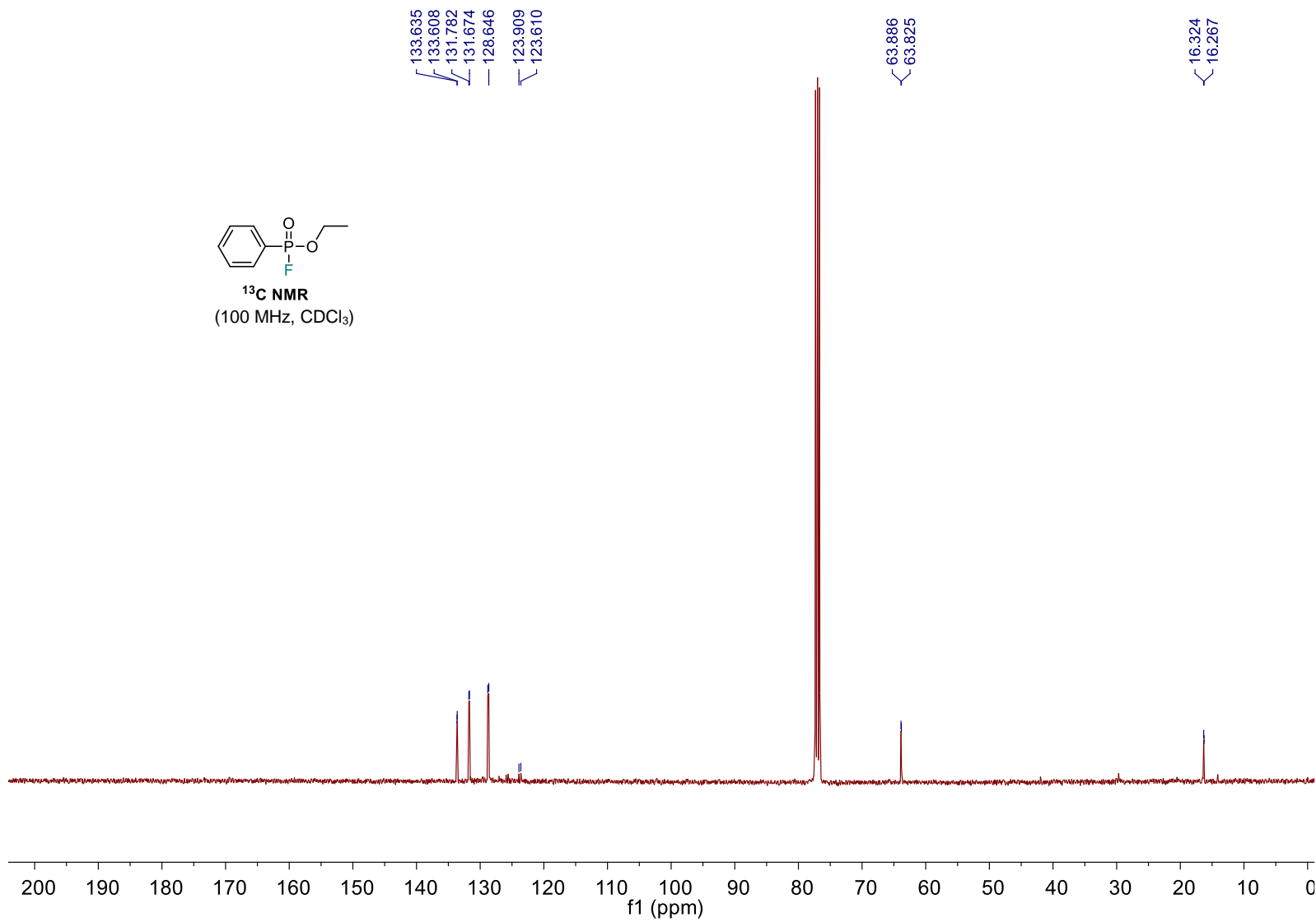
¹⁹F NMR
(376 MHz, CDCl₃)



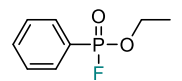




¹³C NMR
(100 MHz, CDCl₃)

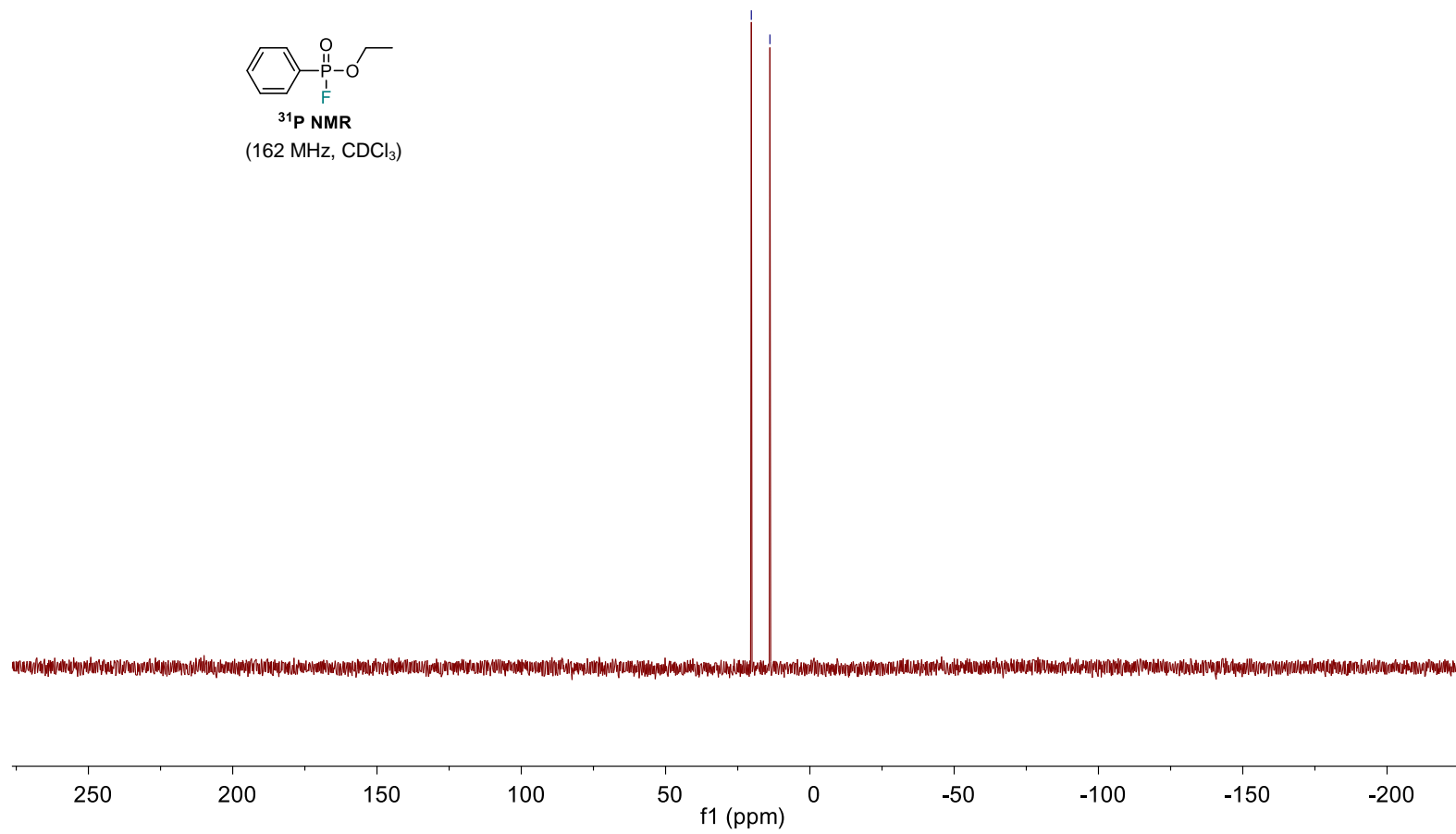


S330

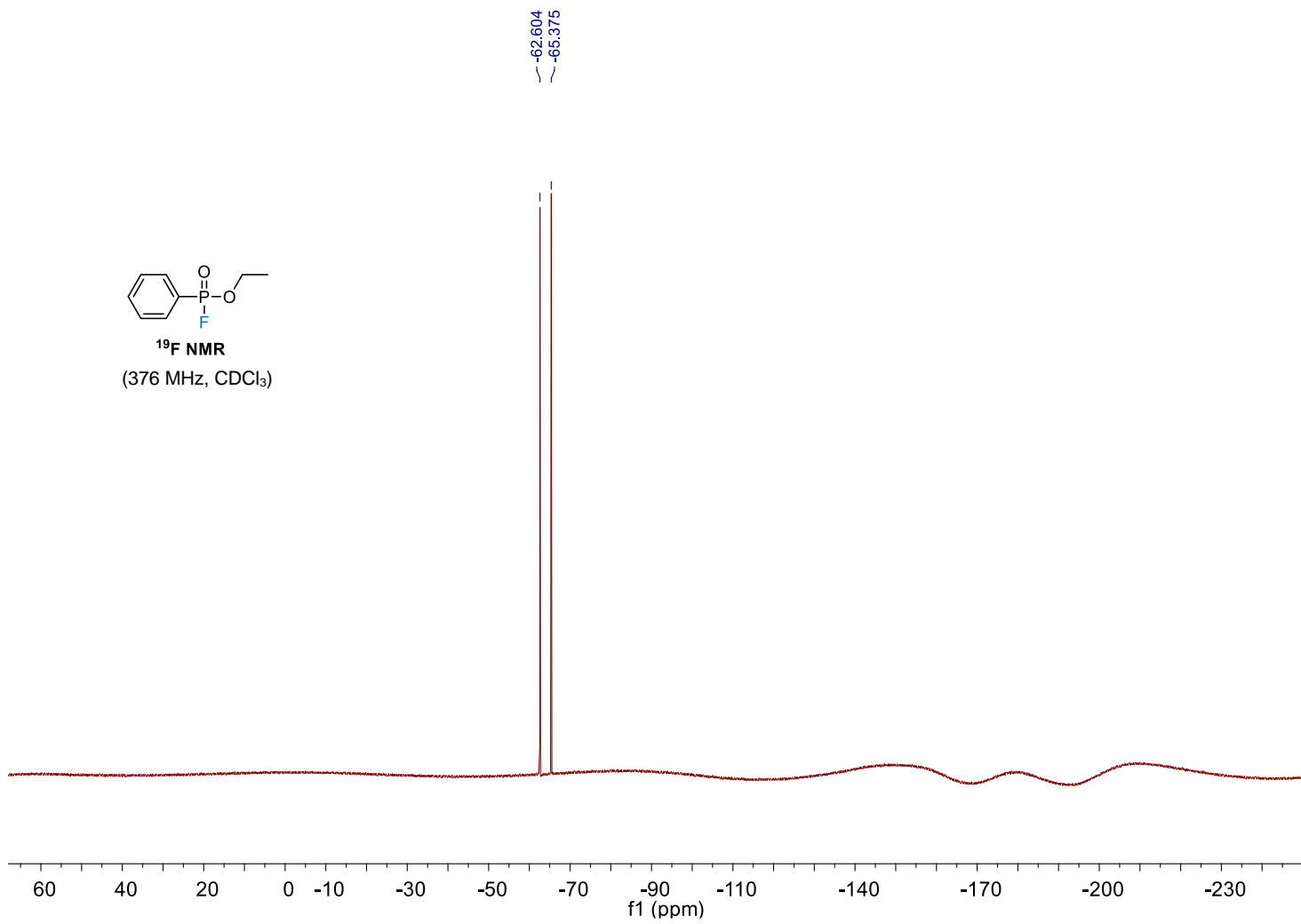
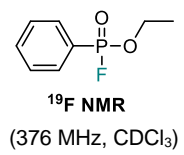


³¹P NMR
(162 MHz, CDCl₃)

— 20.285
— 13.846



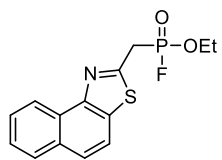
S331



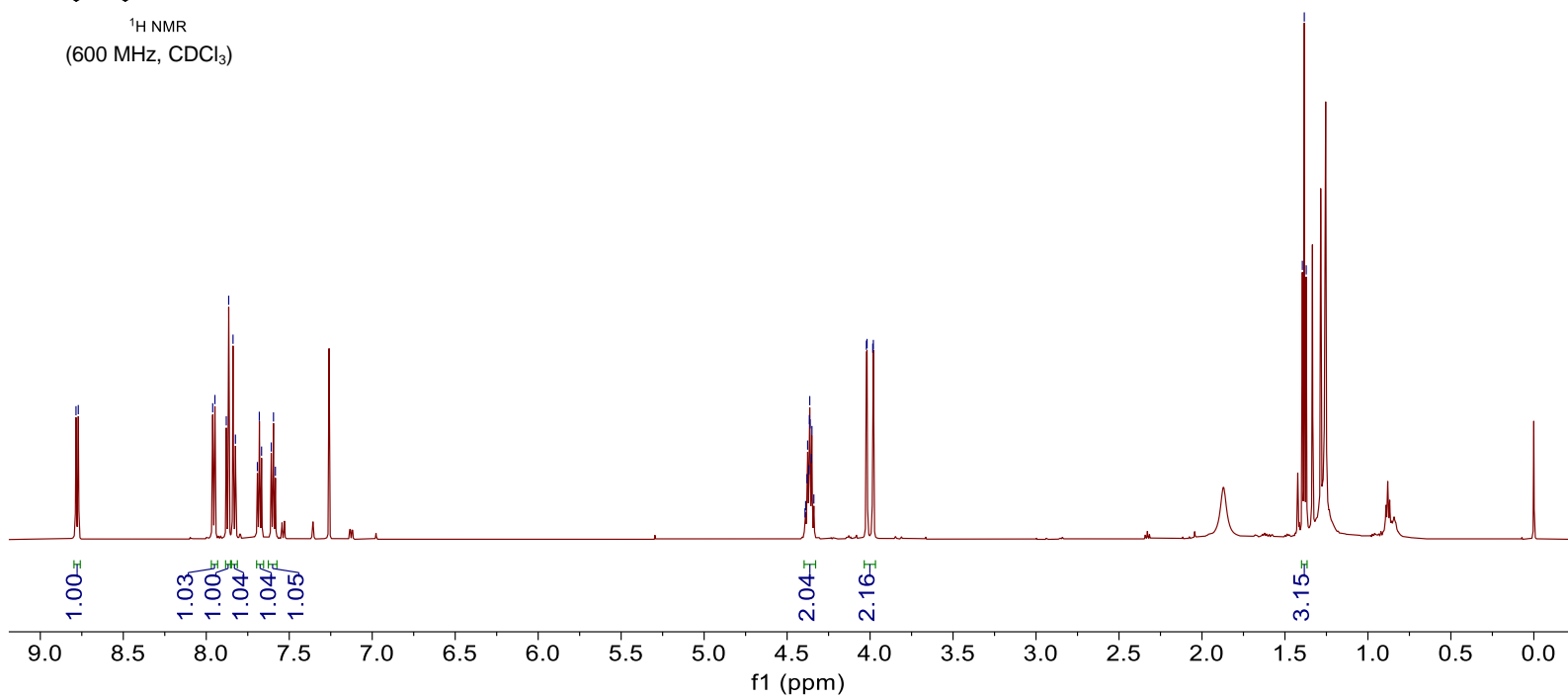
8.785
8.771
7.961
7.948
7.879
7.865
7.839
7.824
7.692
7.680
7.667
7.608
7.595
7.583

4.392
4.388
4.380
4.376
4.368
4.365
4.363
4.362
4.353
4.350
4.339
4.022
4.018
3.984
3.980

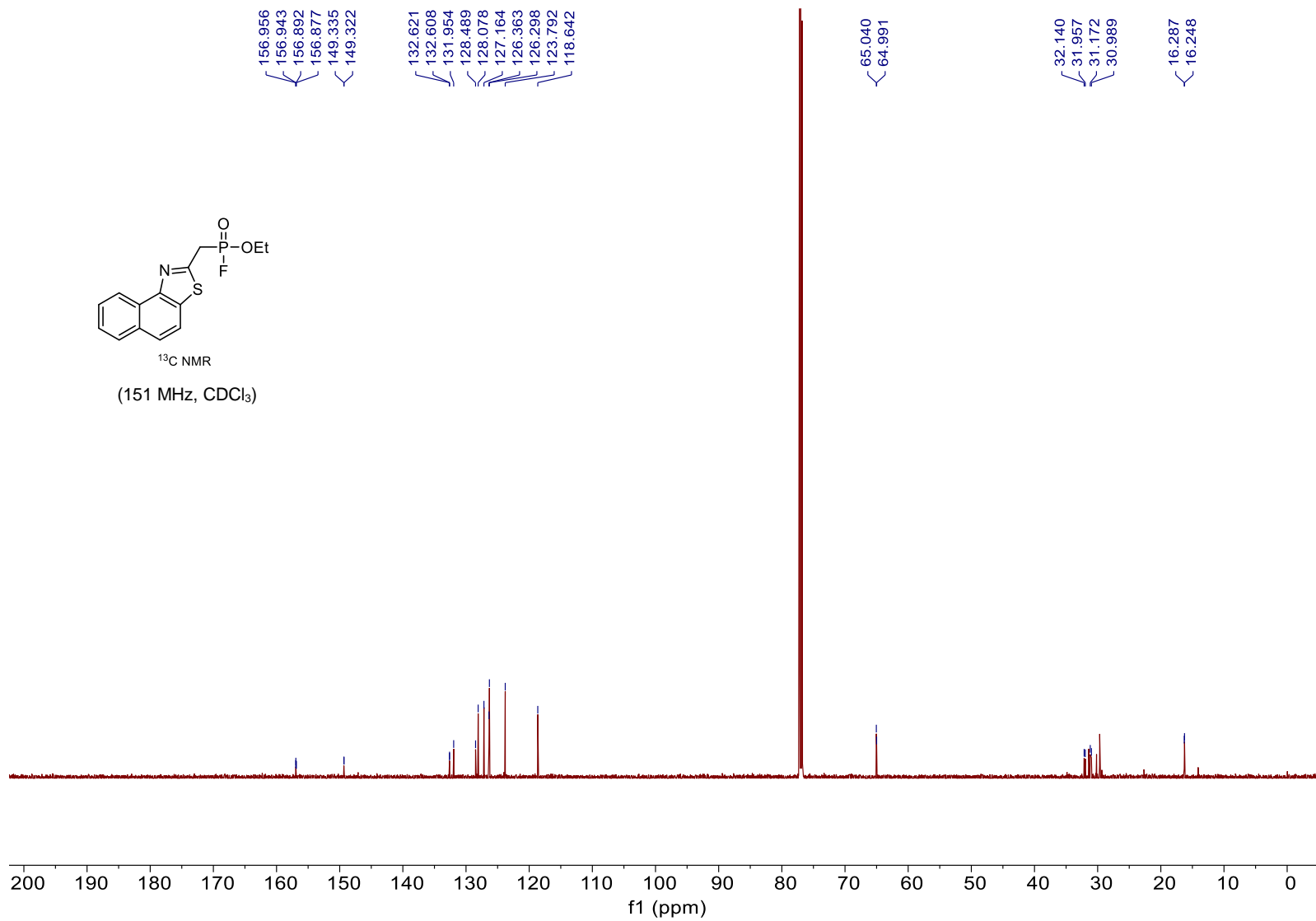
1.395
1.383
1.372

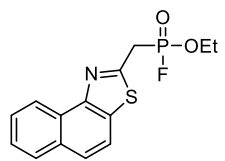


¹H NMR
(600 MHz, CDCl₃)

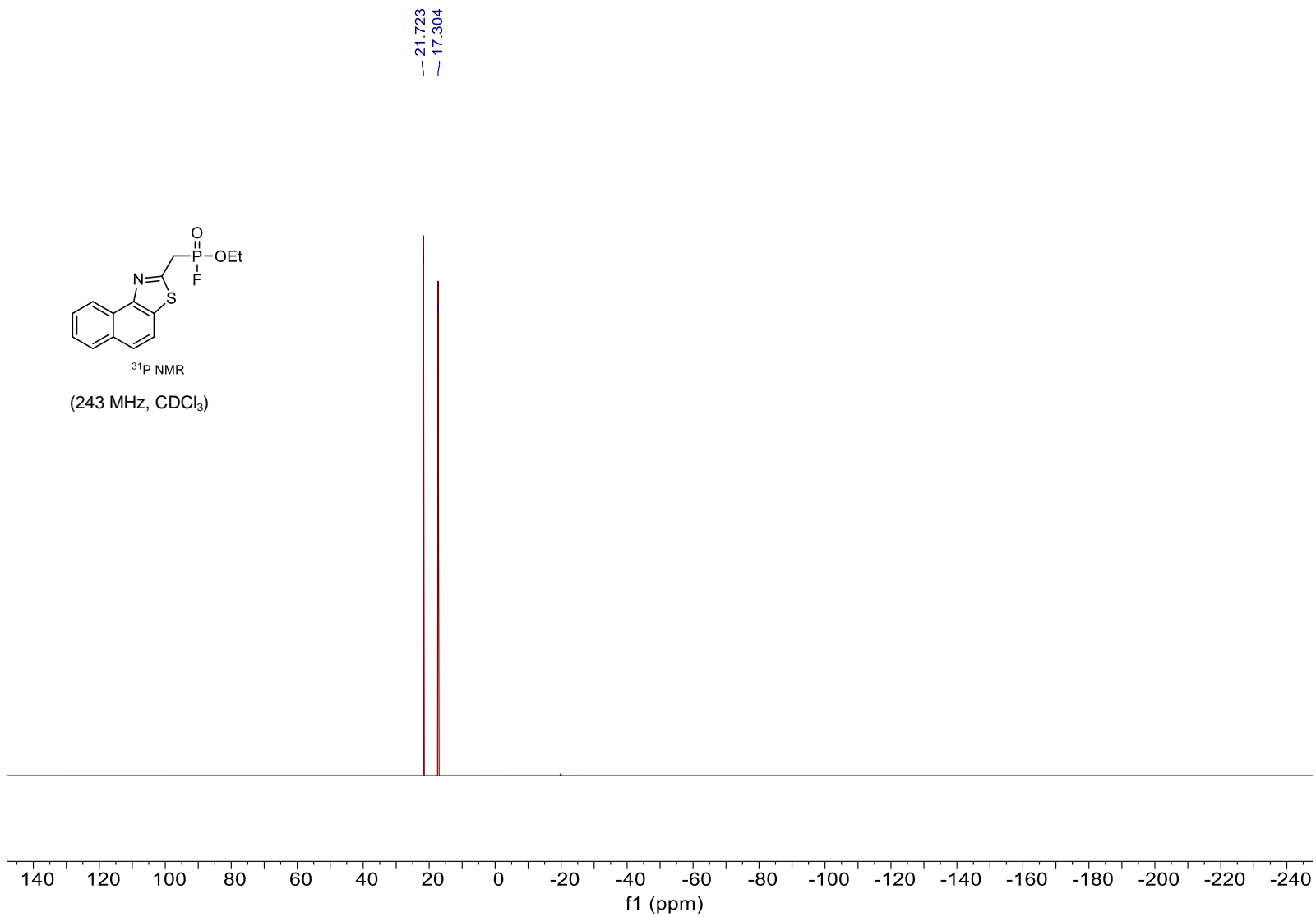


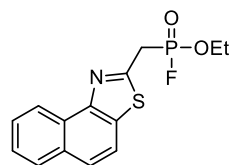
S333



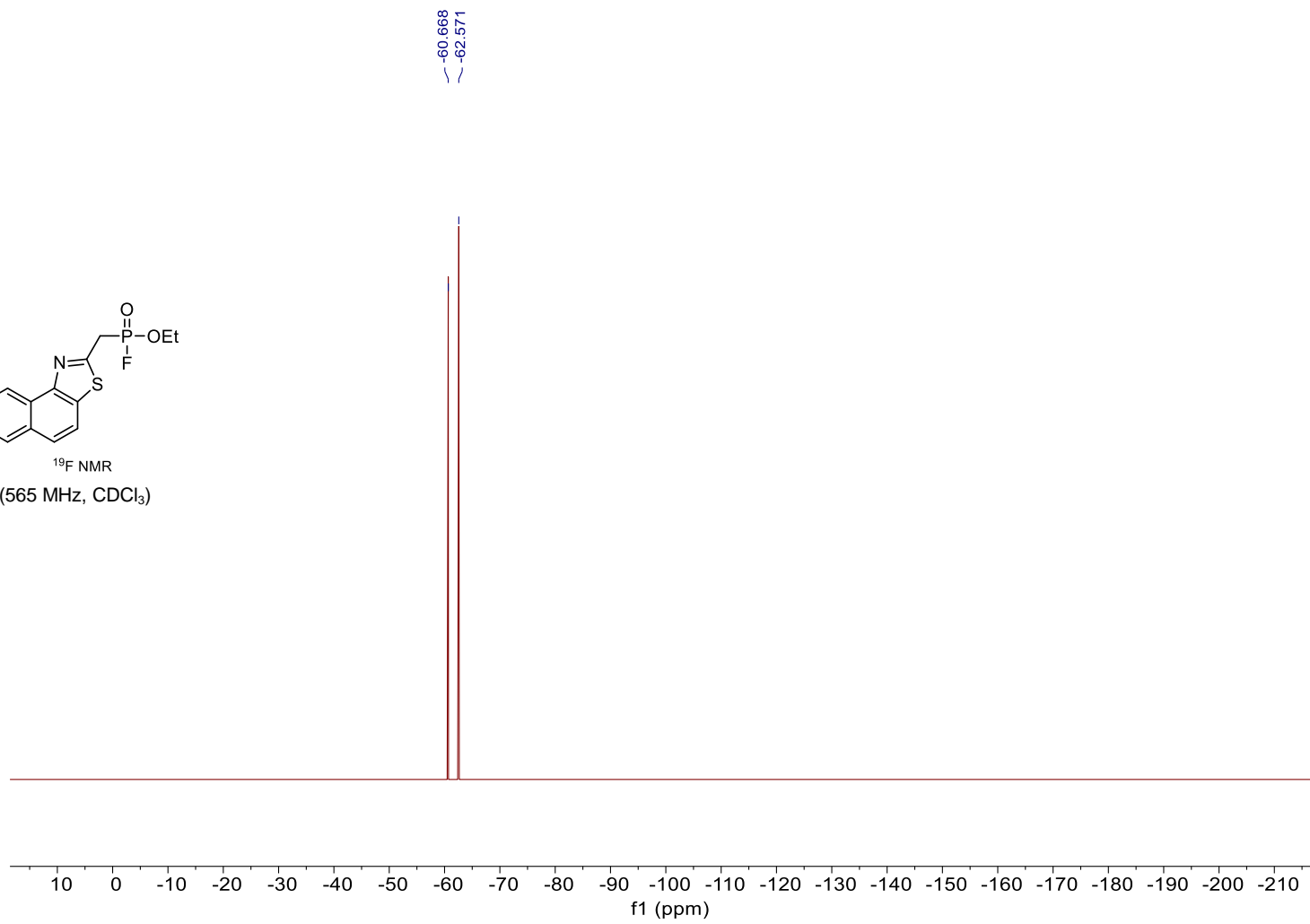


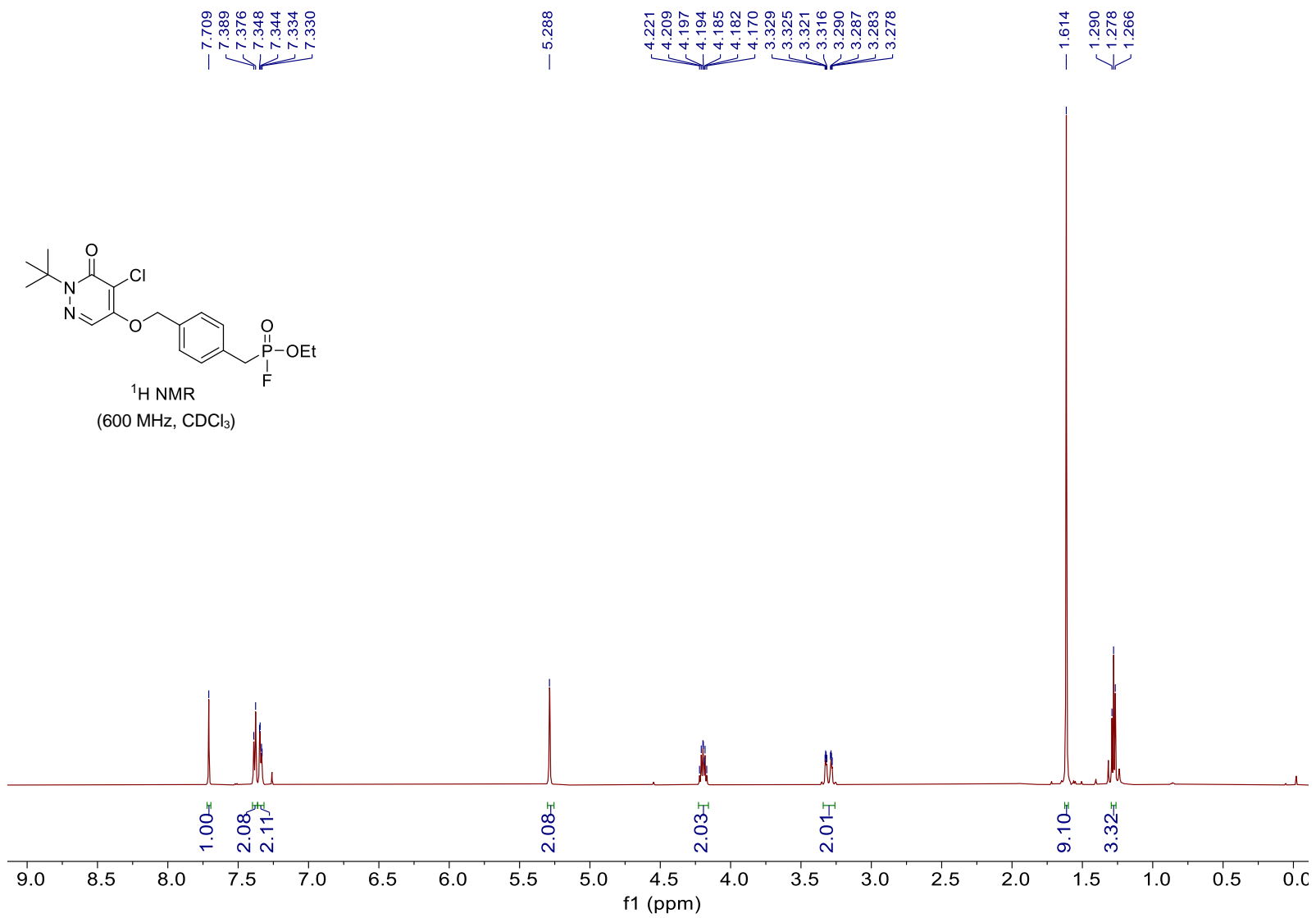
³¹P NMR
(243 MHz, CDCl₃)



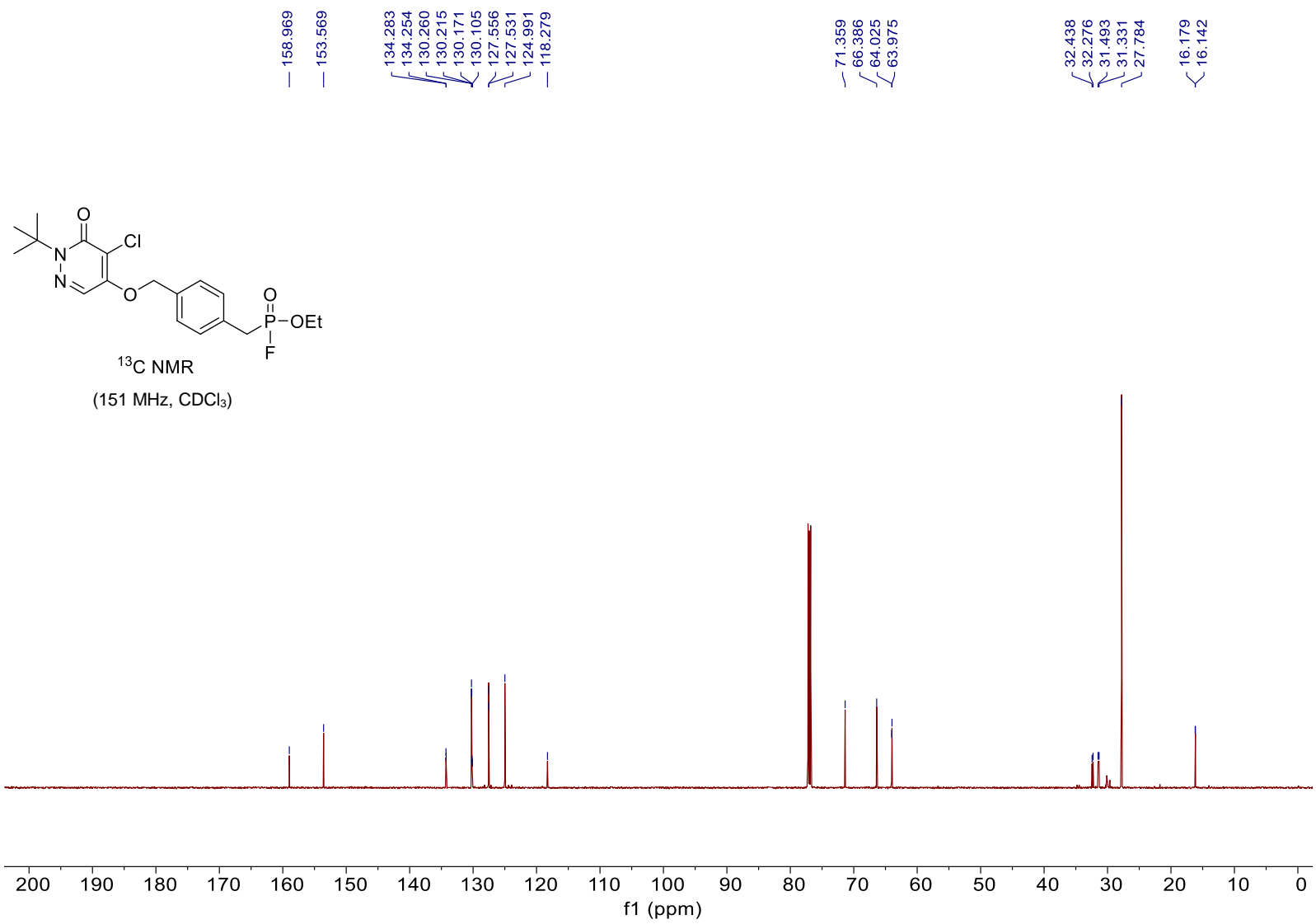


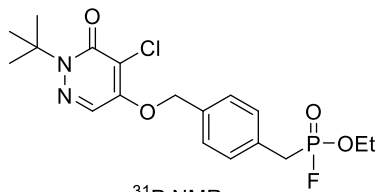
¹⁹F NMR
(565 MHz, CDCl₃)



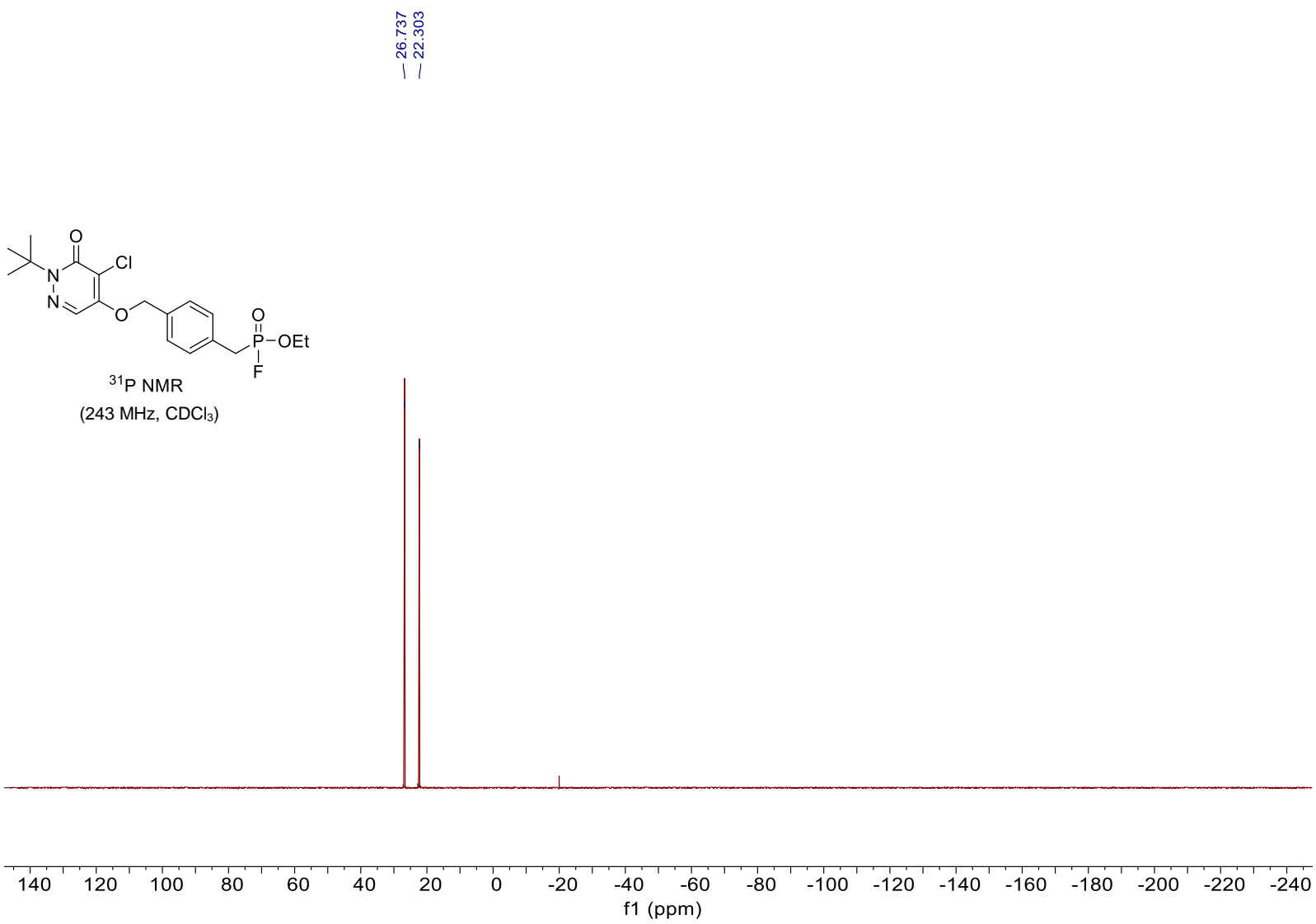


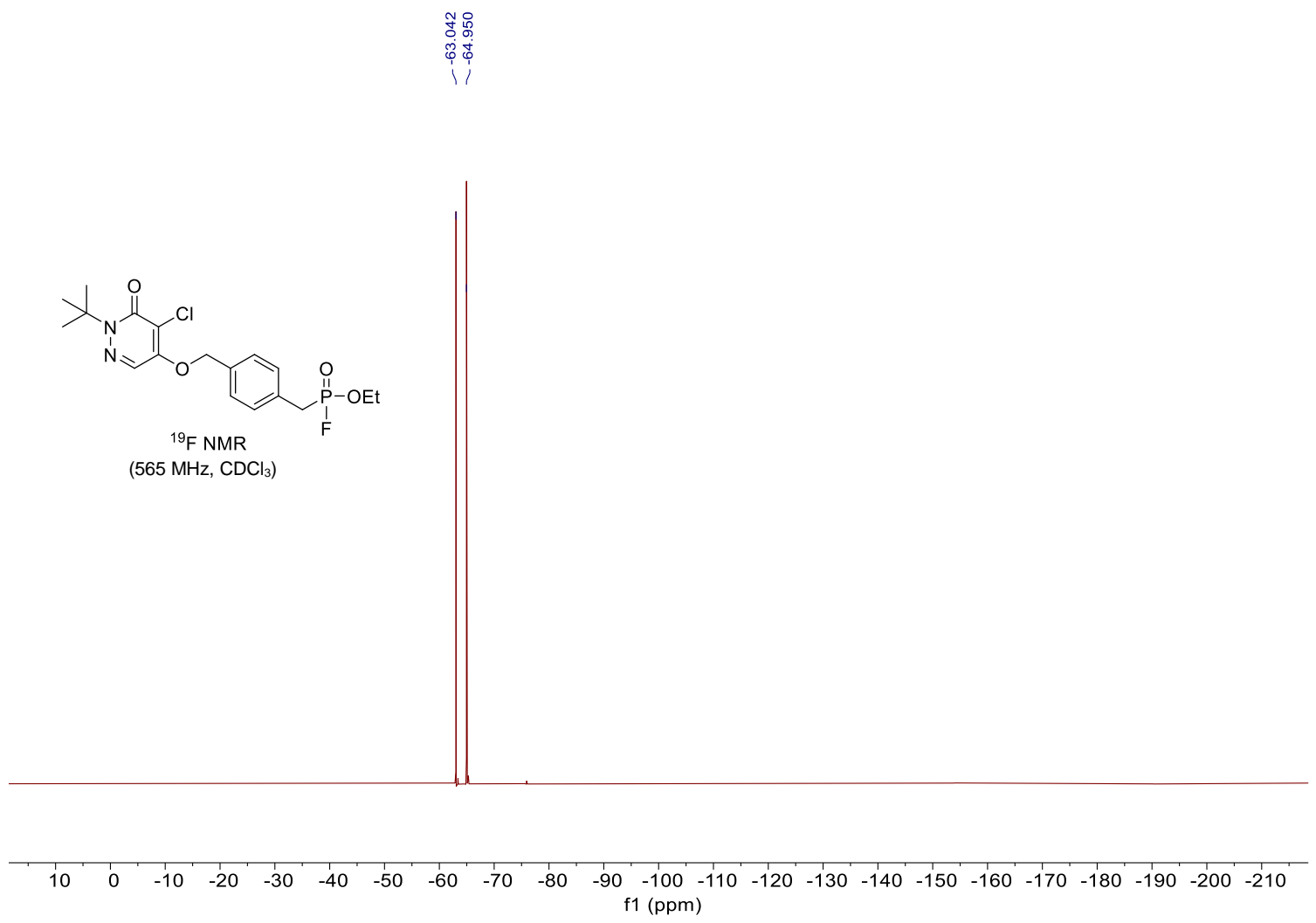
S337

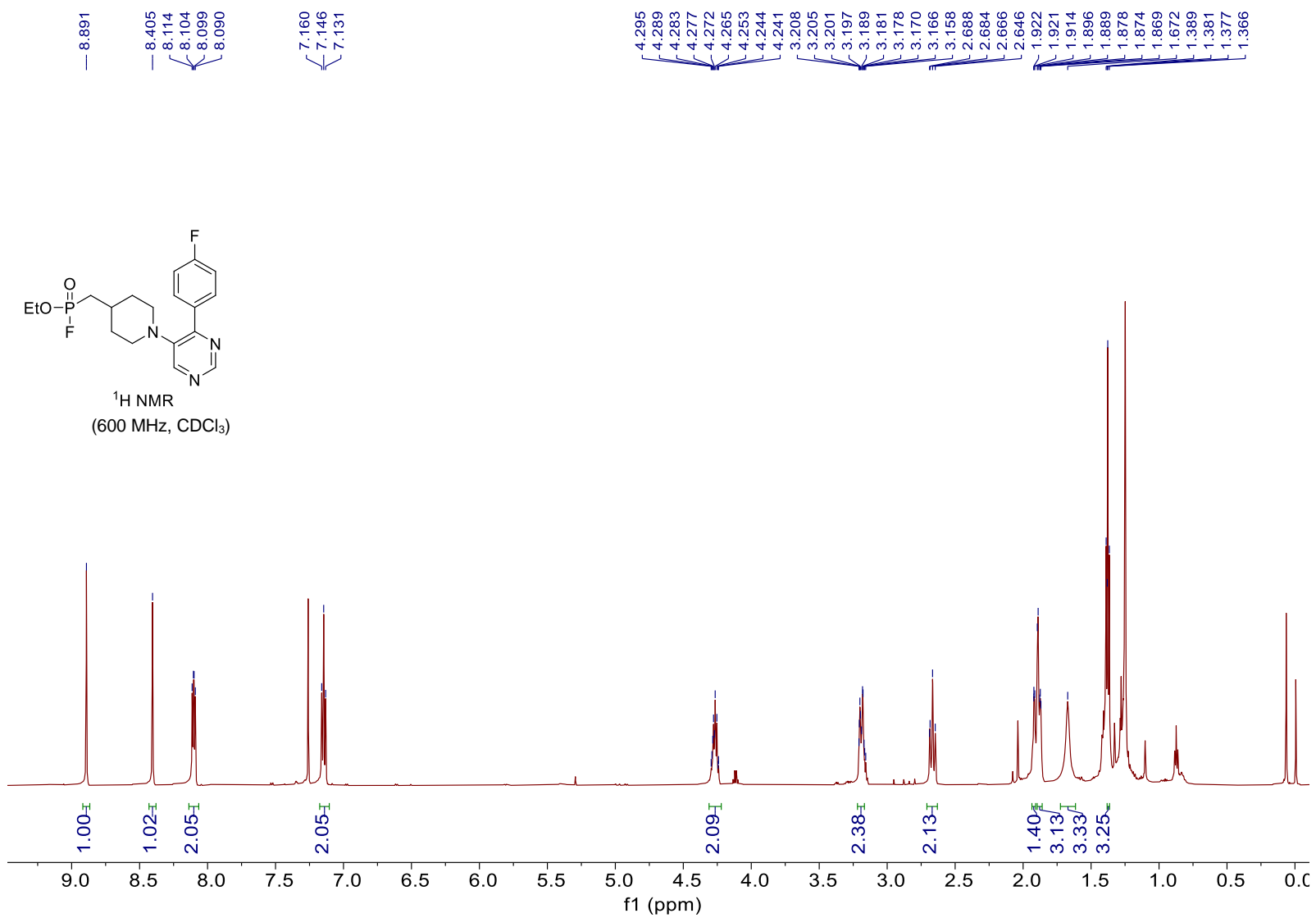


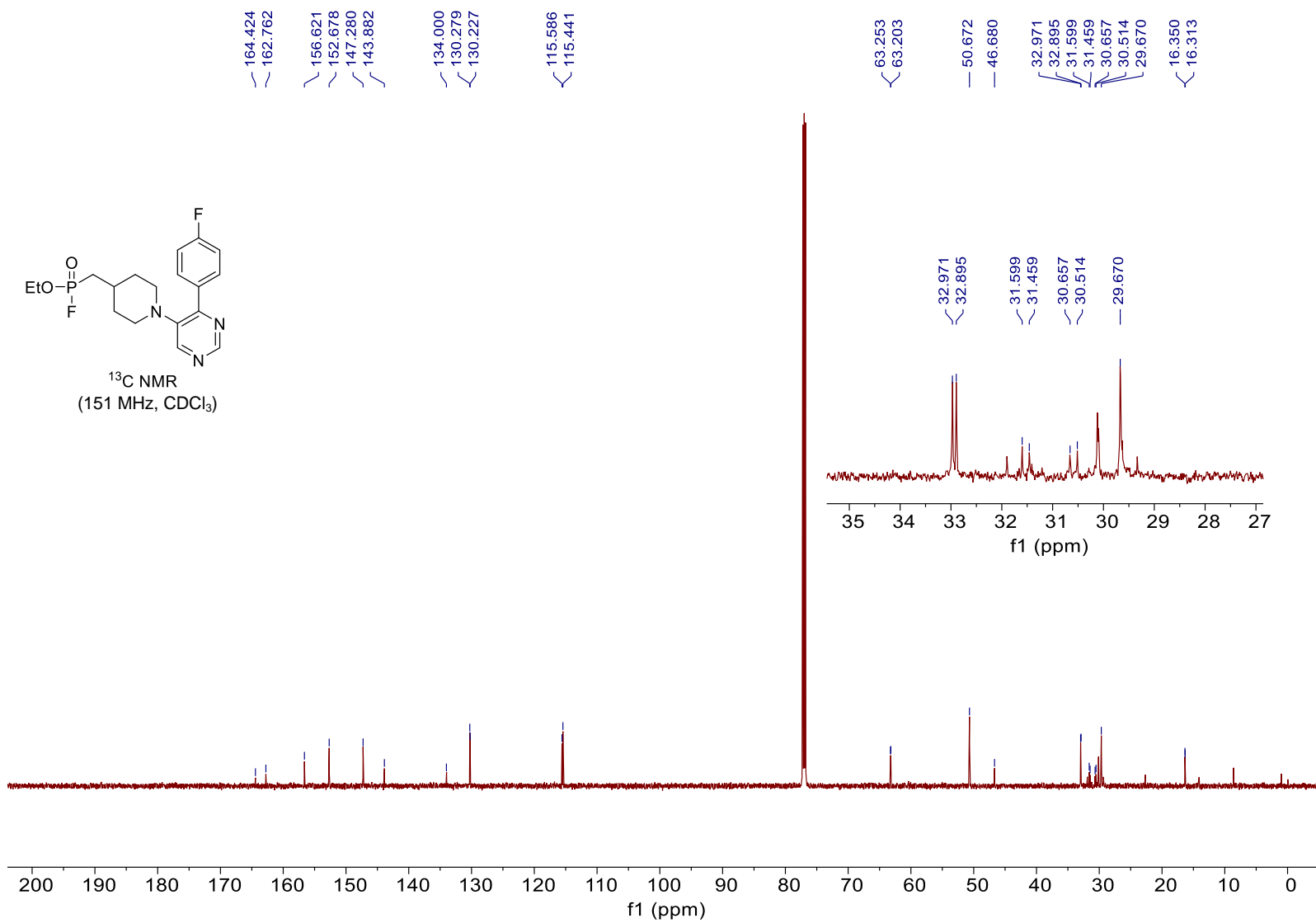


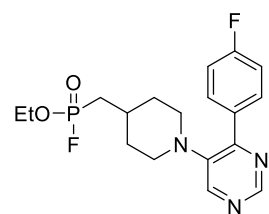
³¹P NMR
(243 MHz, CDCl₃)



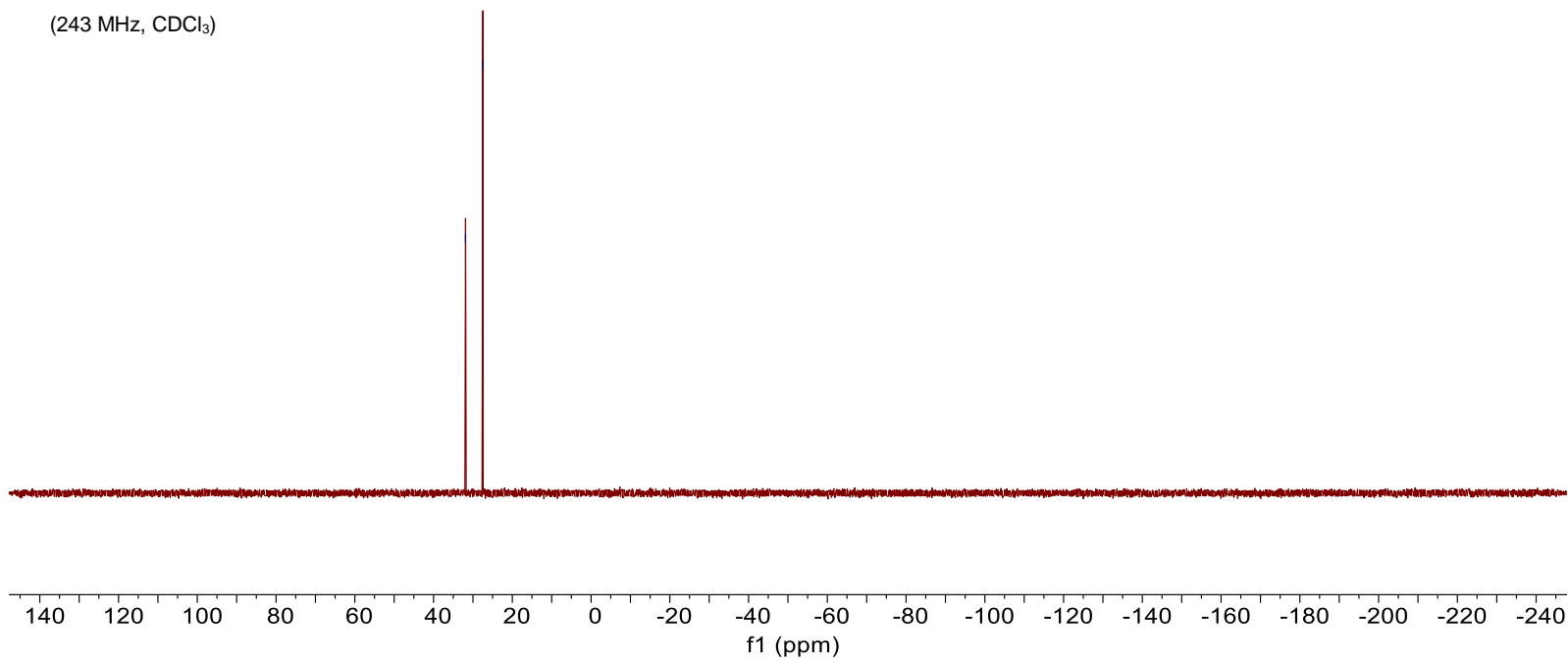


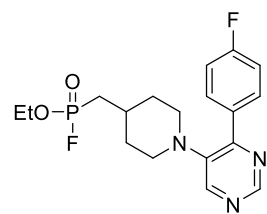




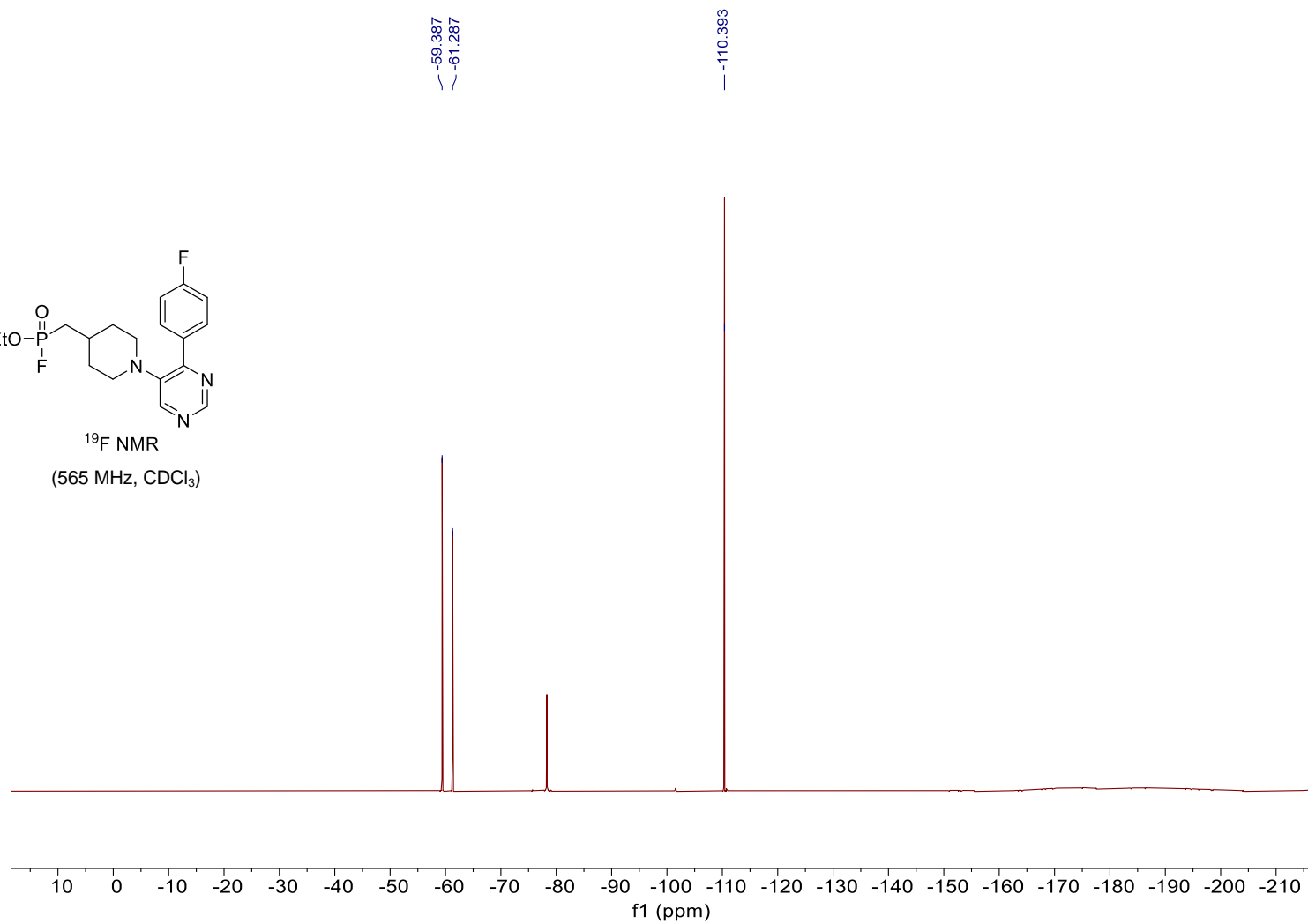


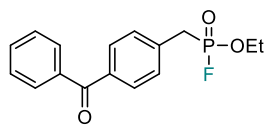
³¹P NMR
(243 MHz, CDCl₃)





¹⁹F NMR
(565 MHz, CDCl₃)





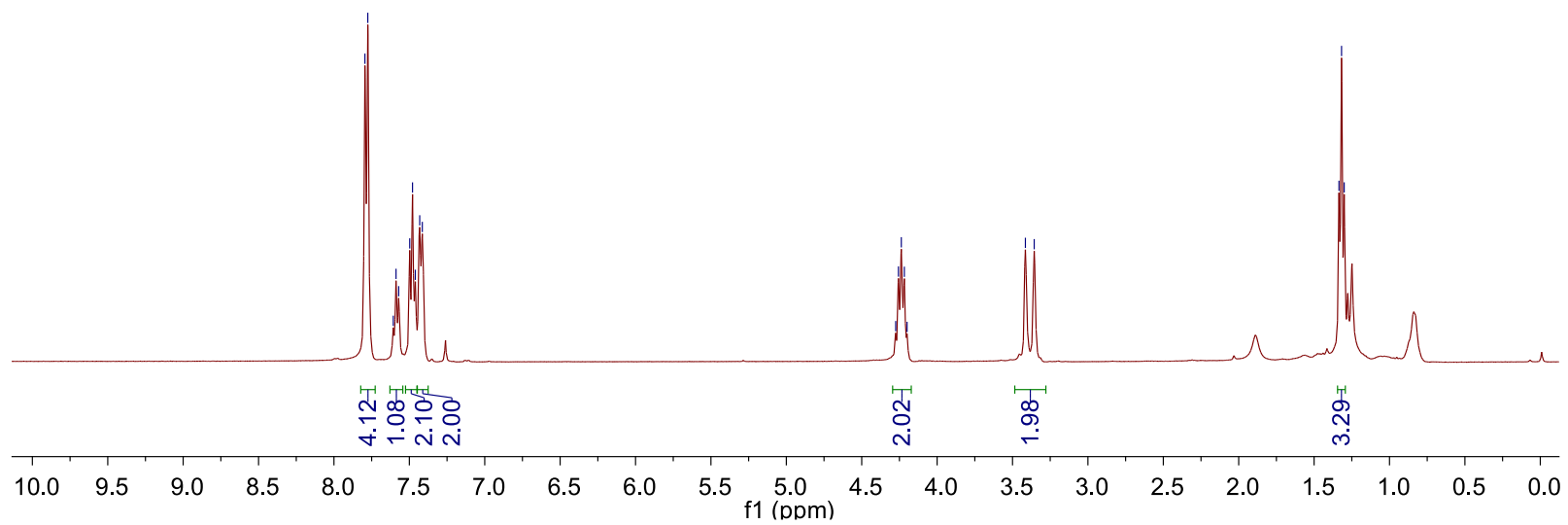
¹H NMR
(400 MHz, CDCl₃)

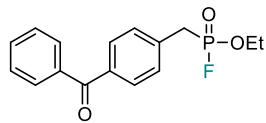
7.795
7.775
7.606
7.588
7.571
7.497
7.479
7.460
7.431
7.414

4.275
4.257
4.237
4.217
4.199

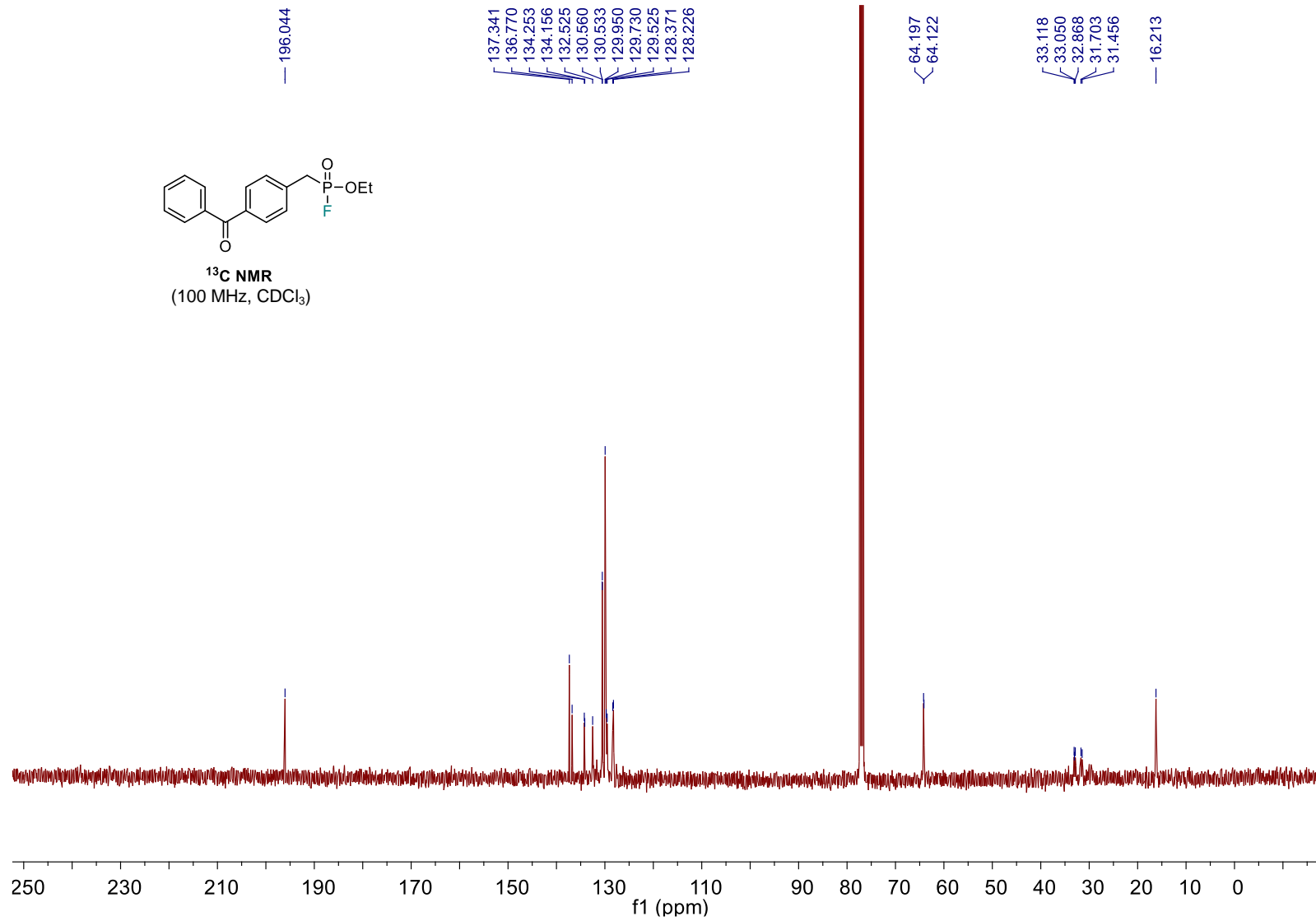
3.415
3.356

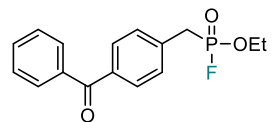
1.335
1.318
1.300





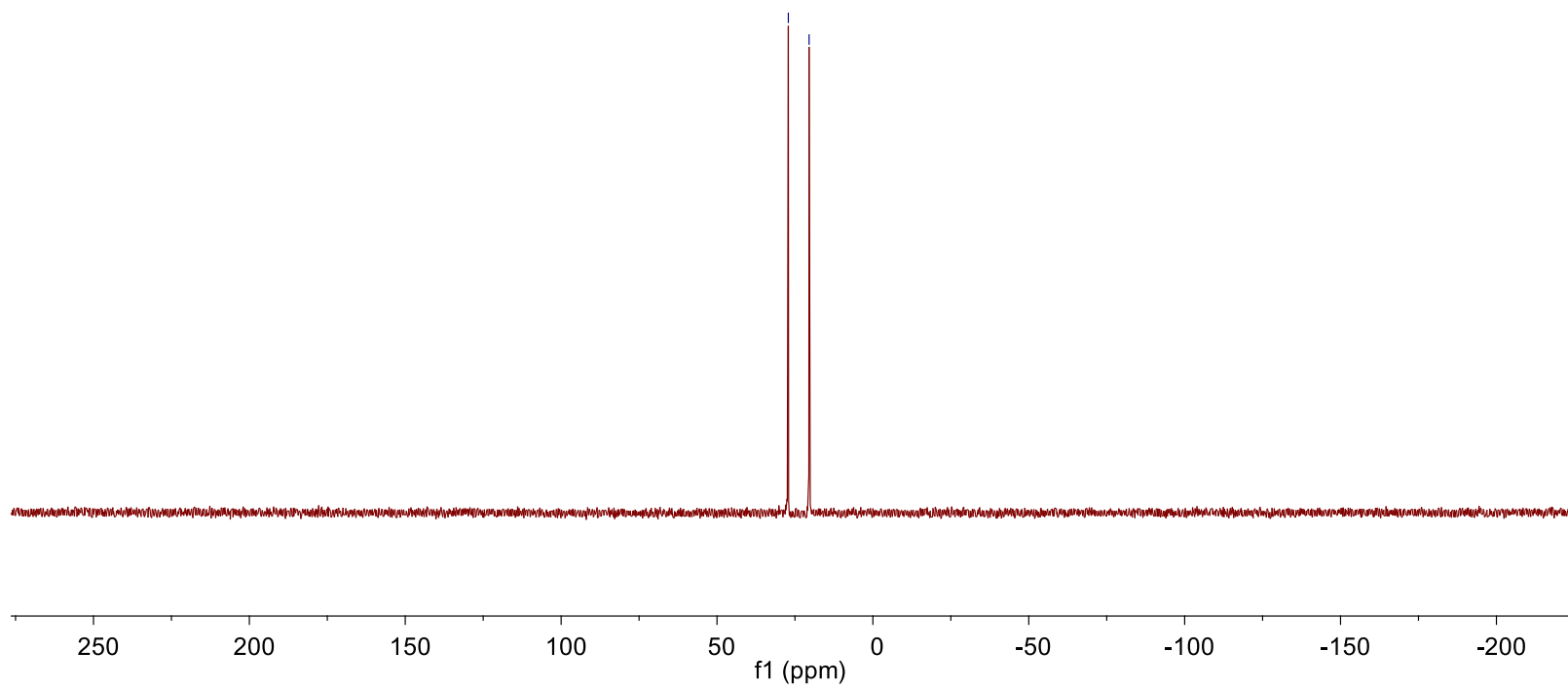
¹³C NMR
(100 MHz, CDCl₃)



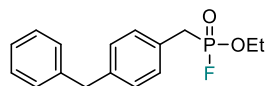


³¹P NMR
(162 MHz, CDCl₃)

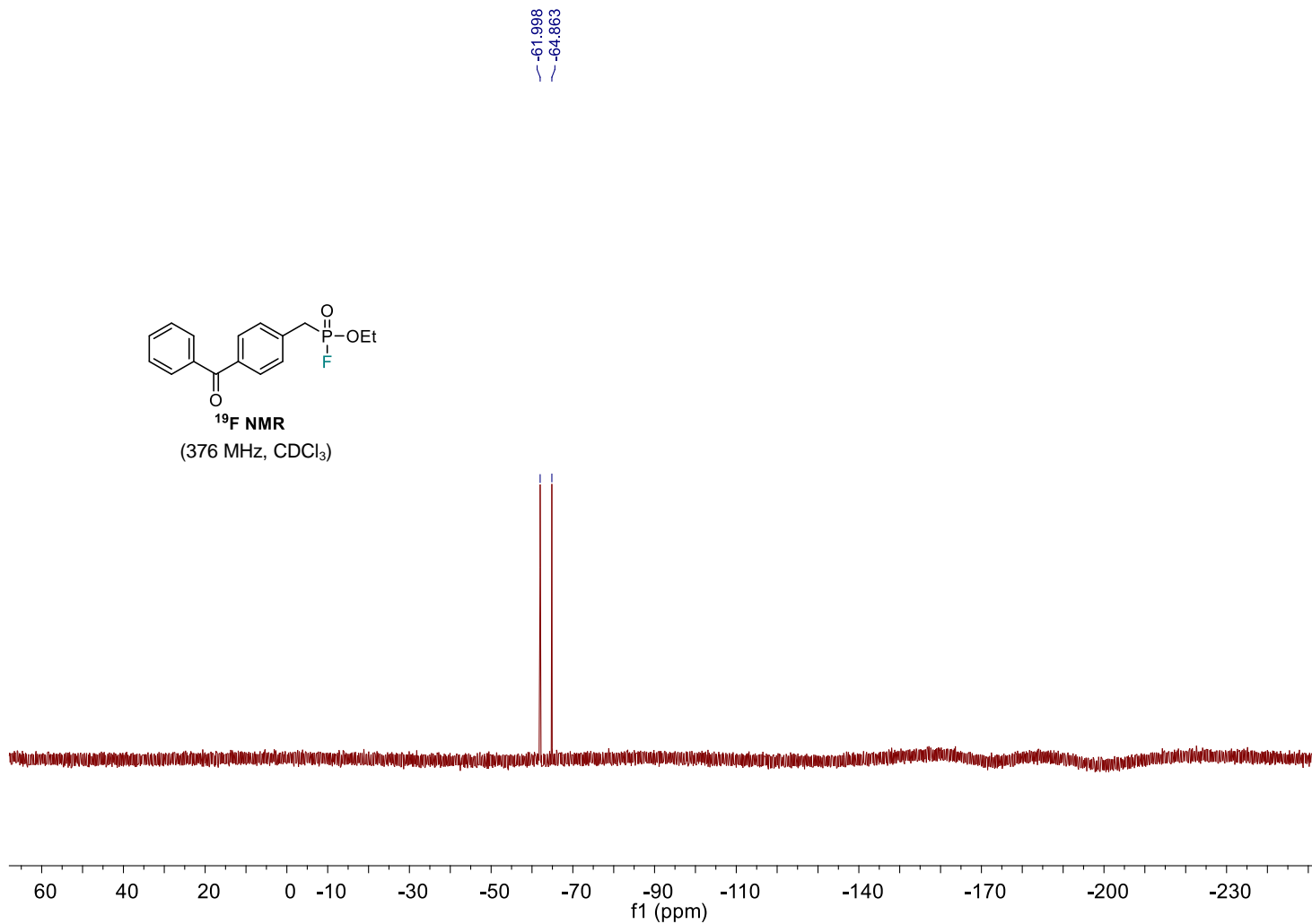
— 27.125
— 20.466

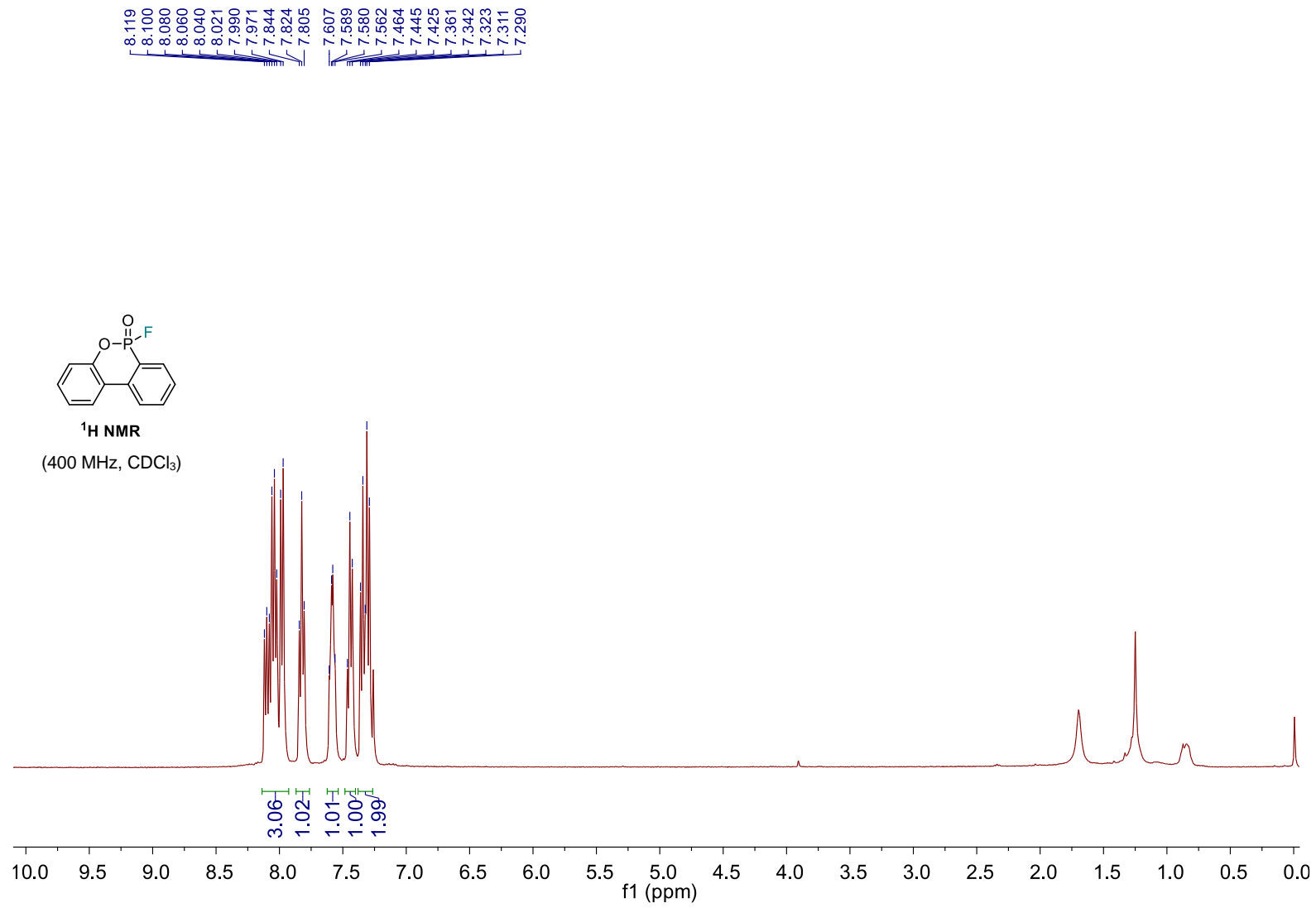


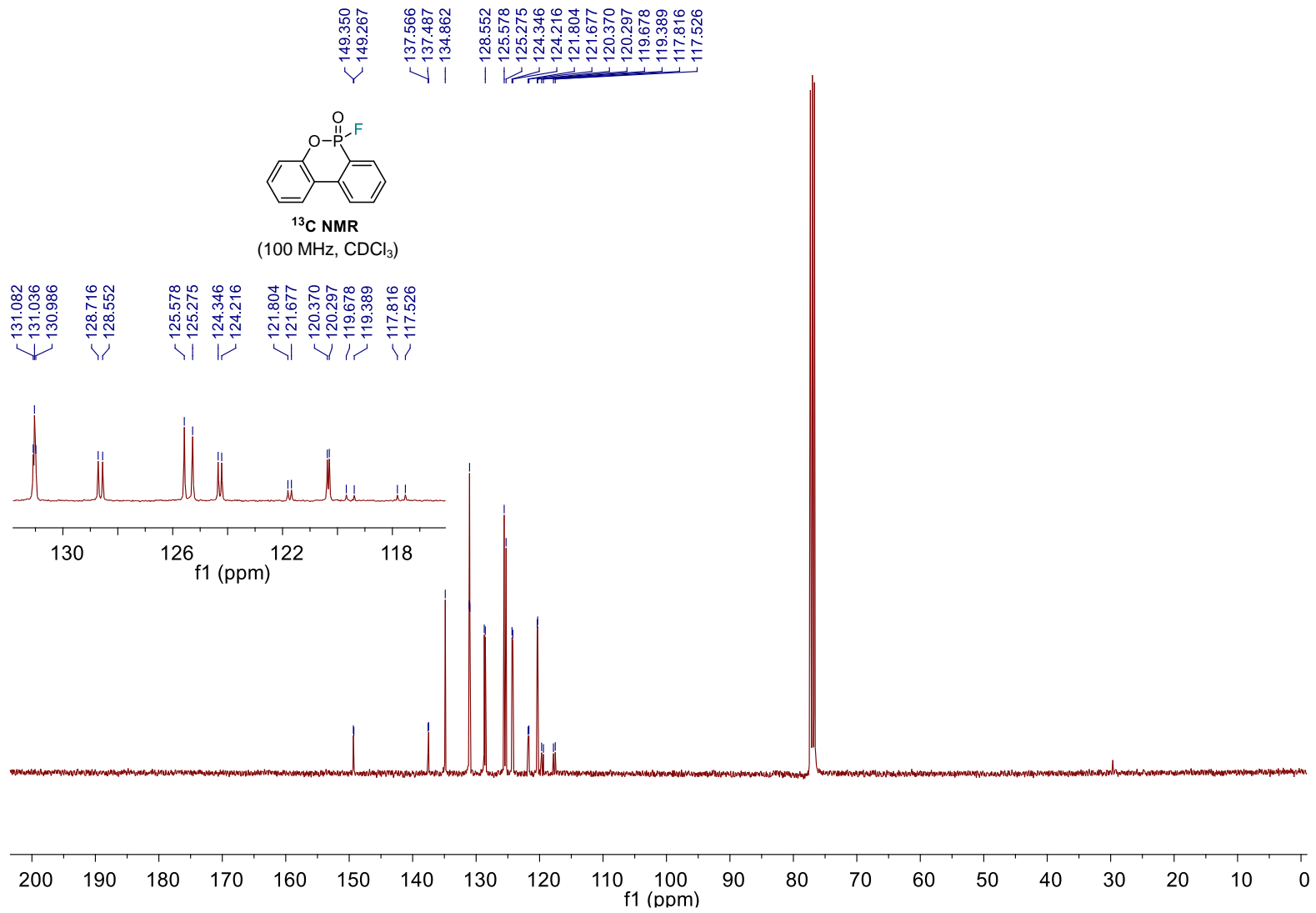
S347

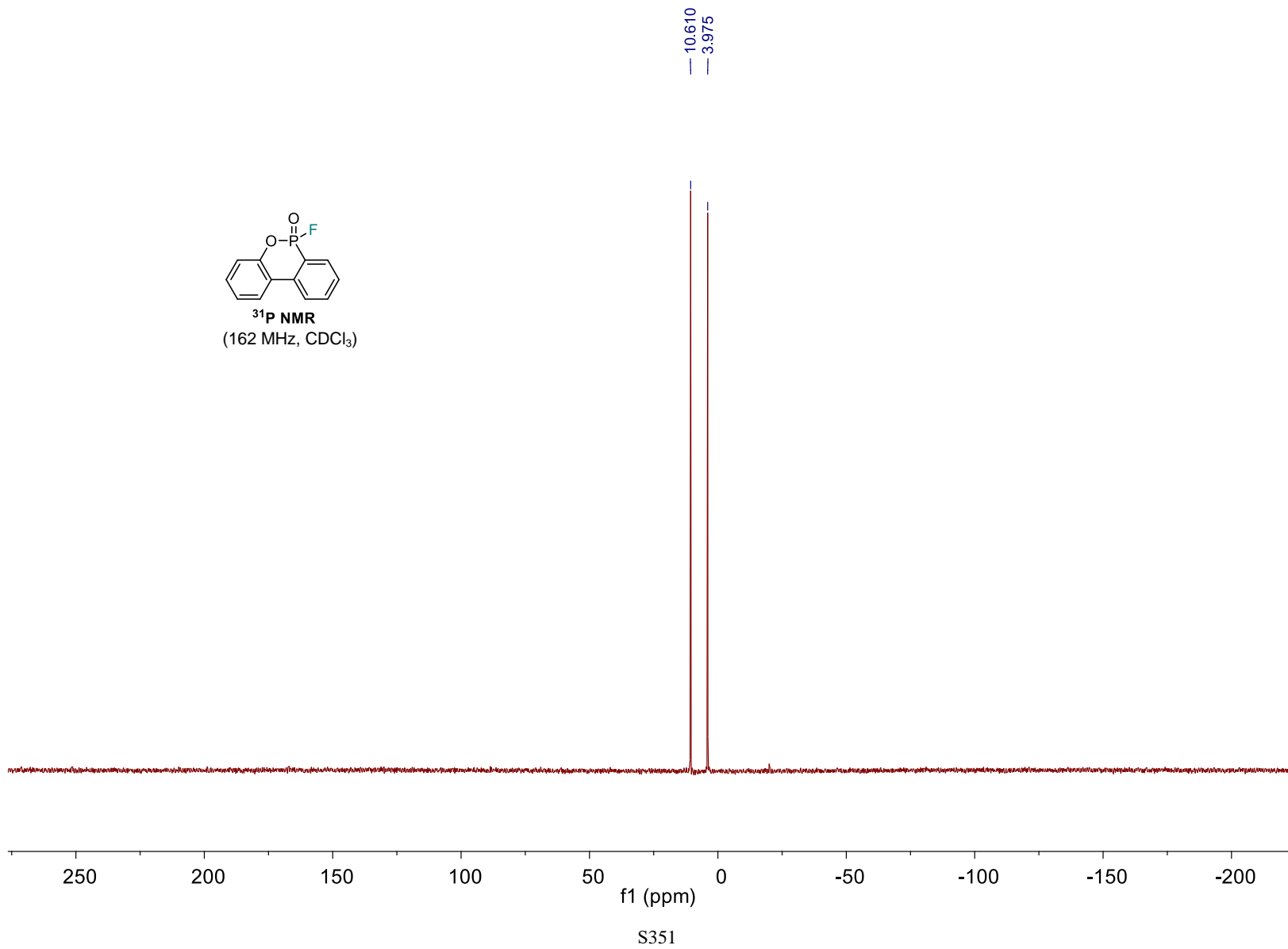
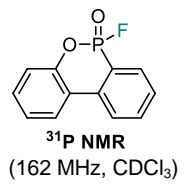


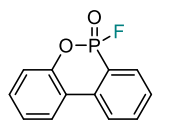
¹⁹F NMR
(376 MHz, CDCl₃)



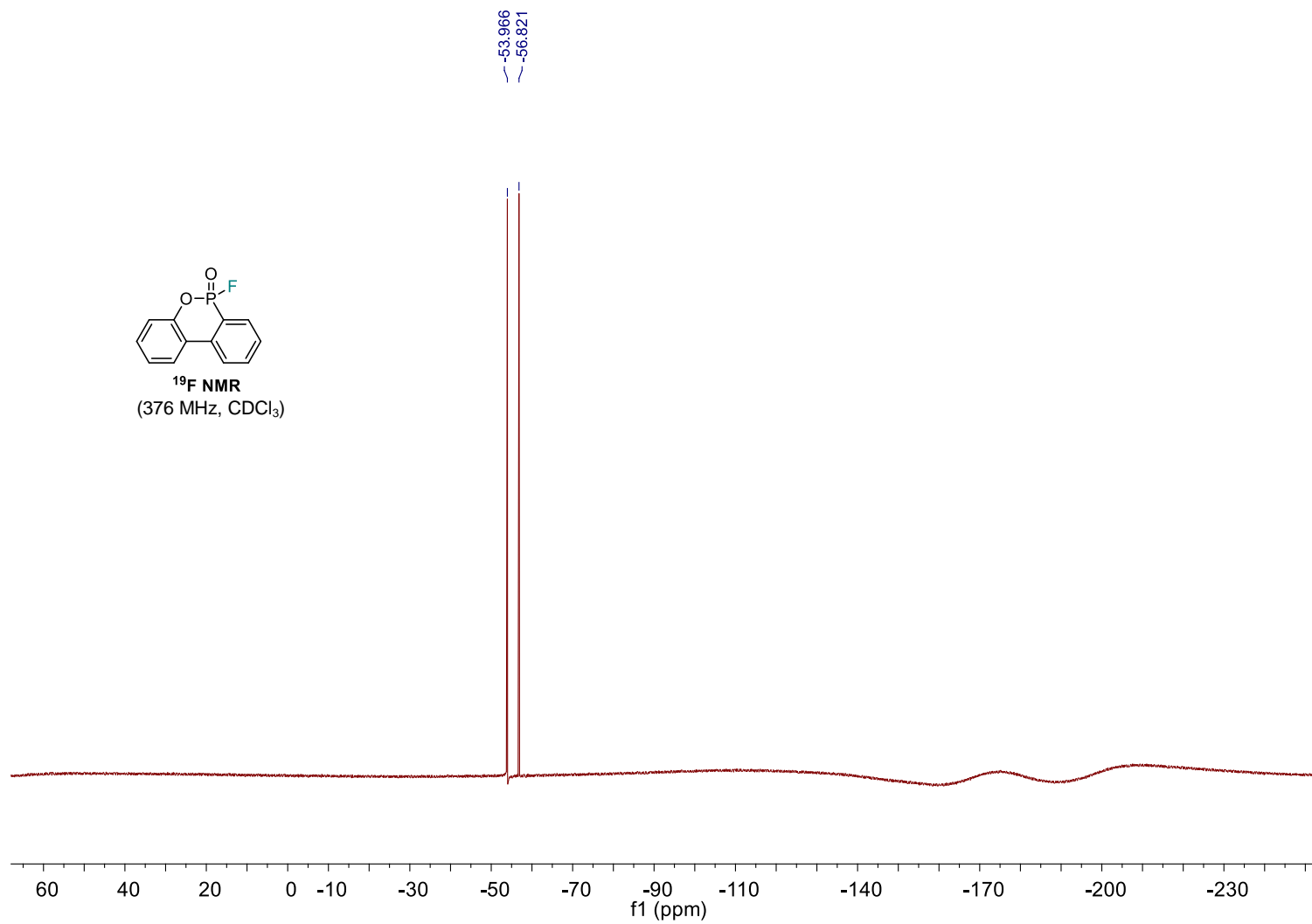


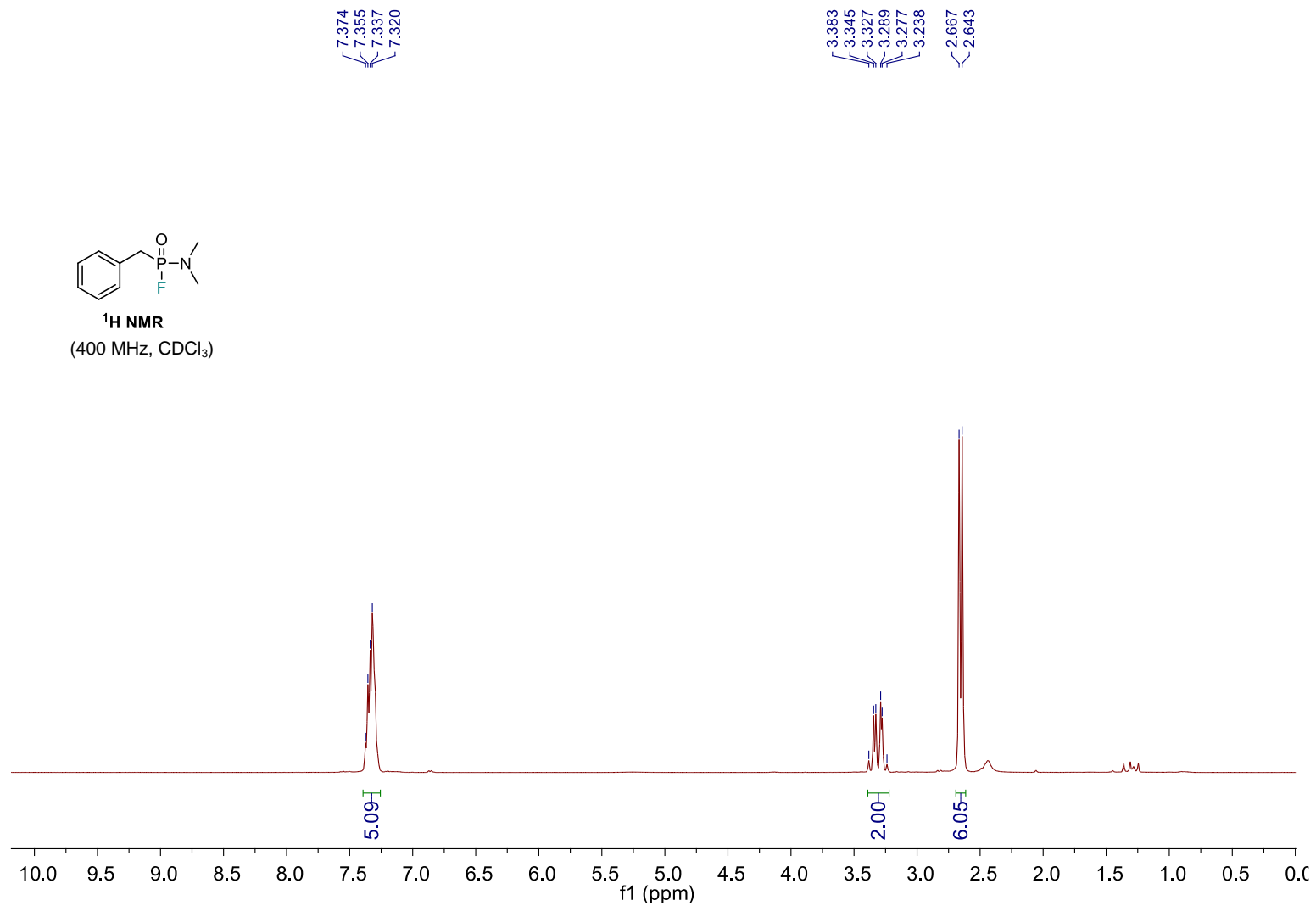
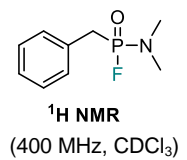


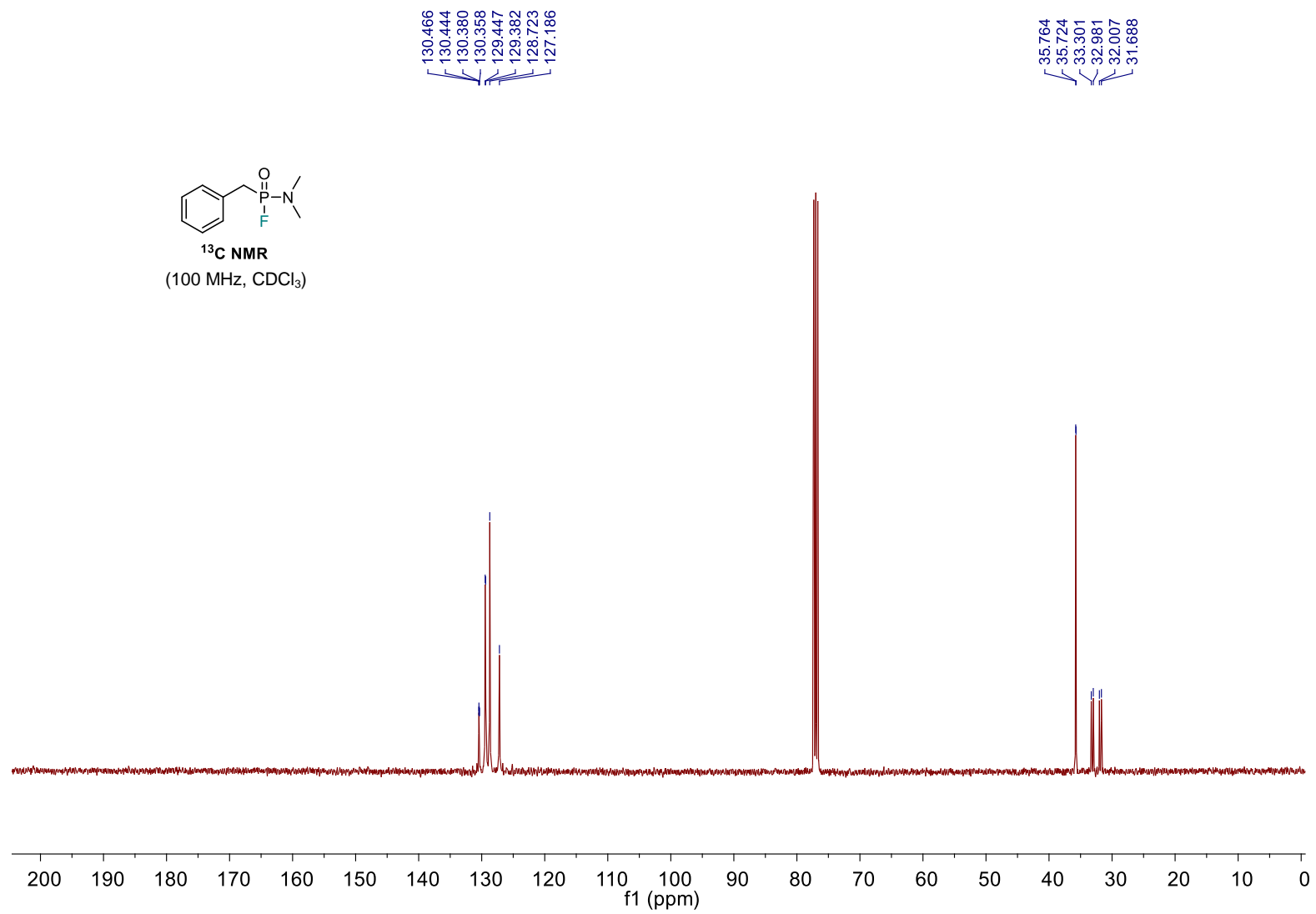
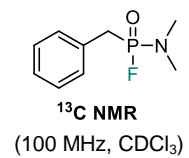


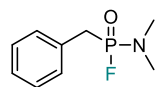


¹⁹F NMR
(376 MHz, CDCl₃)

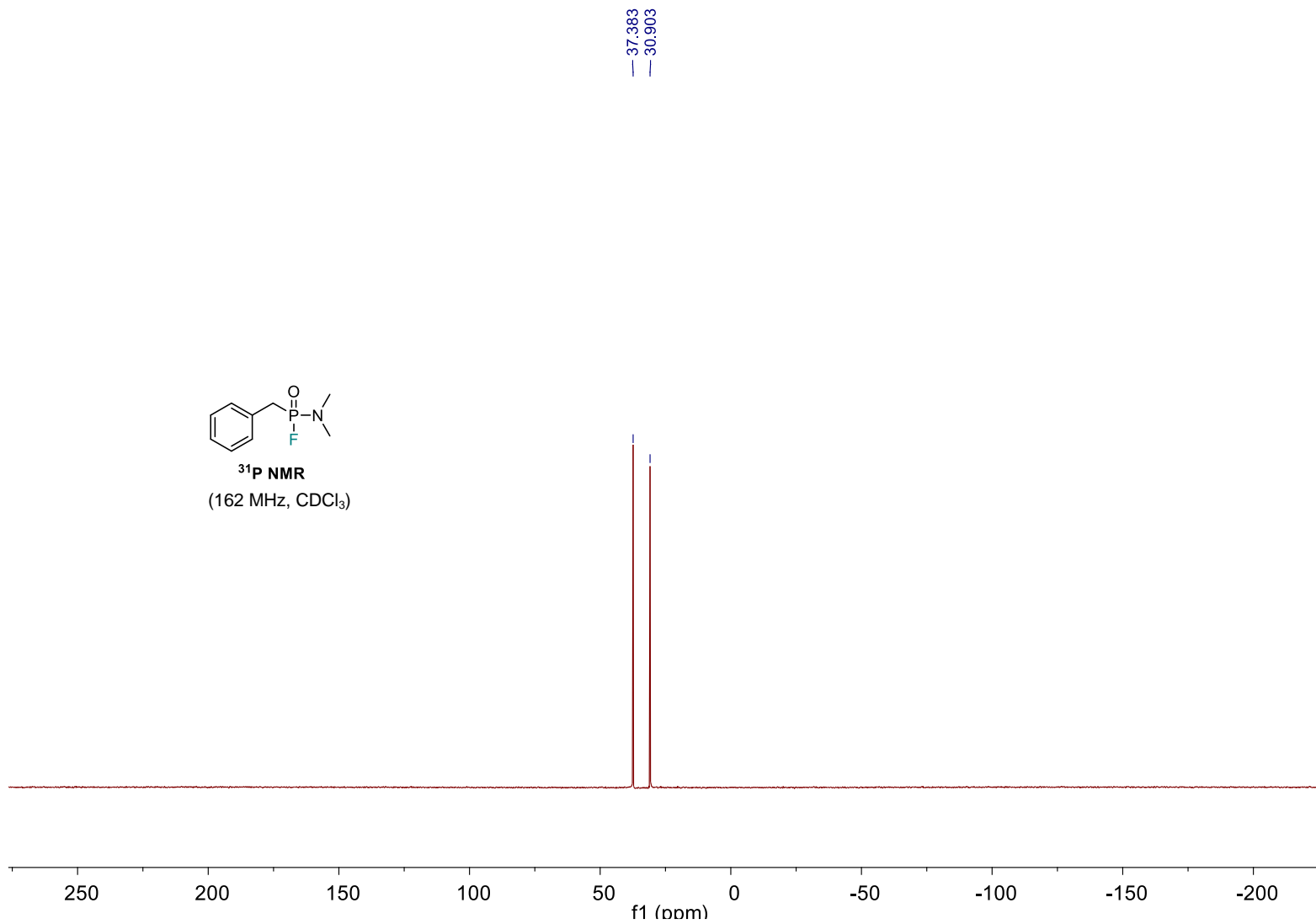




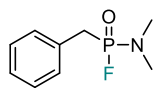




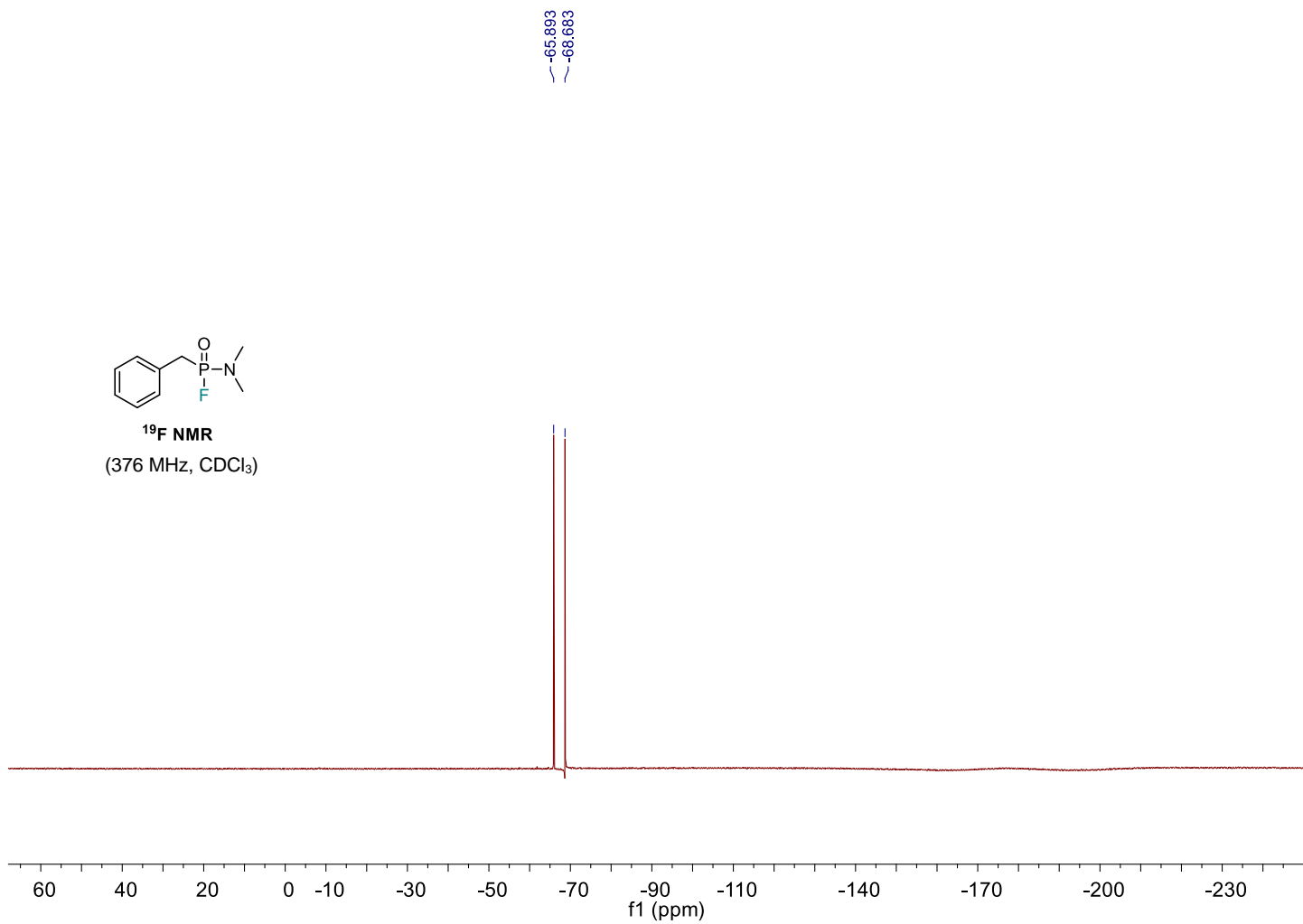
³¹P NMR
(162 MHz, CDCl₃)

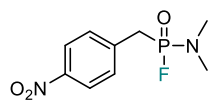


S355

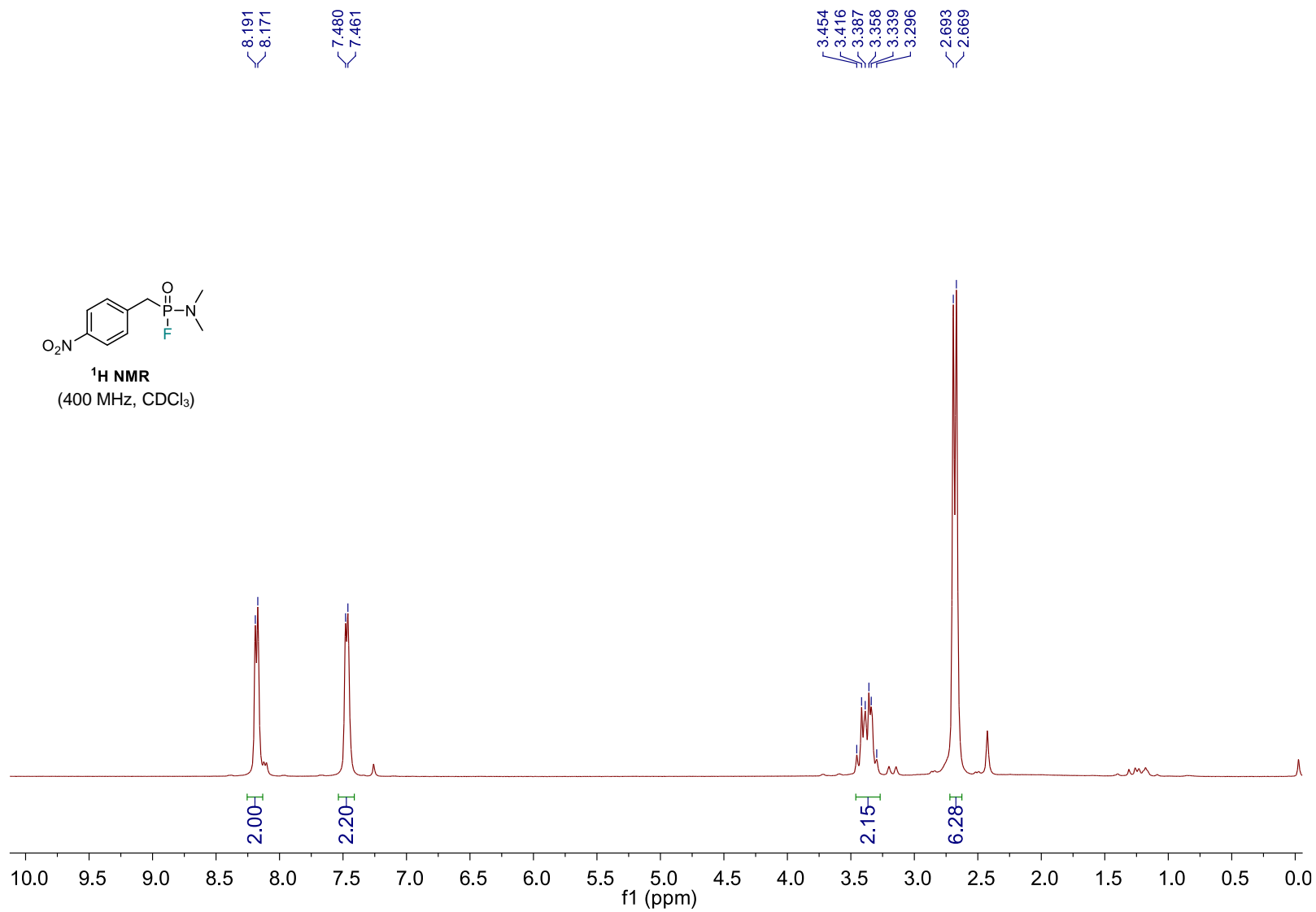


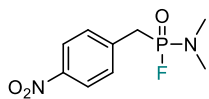
¹⁹F NMR
(376 MHz, CDCl₃)



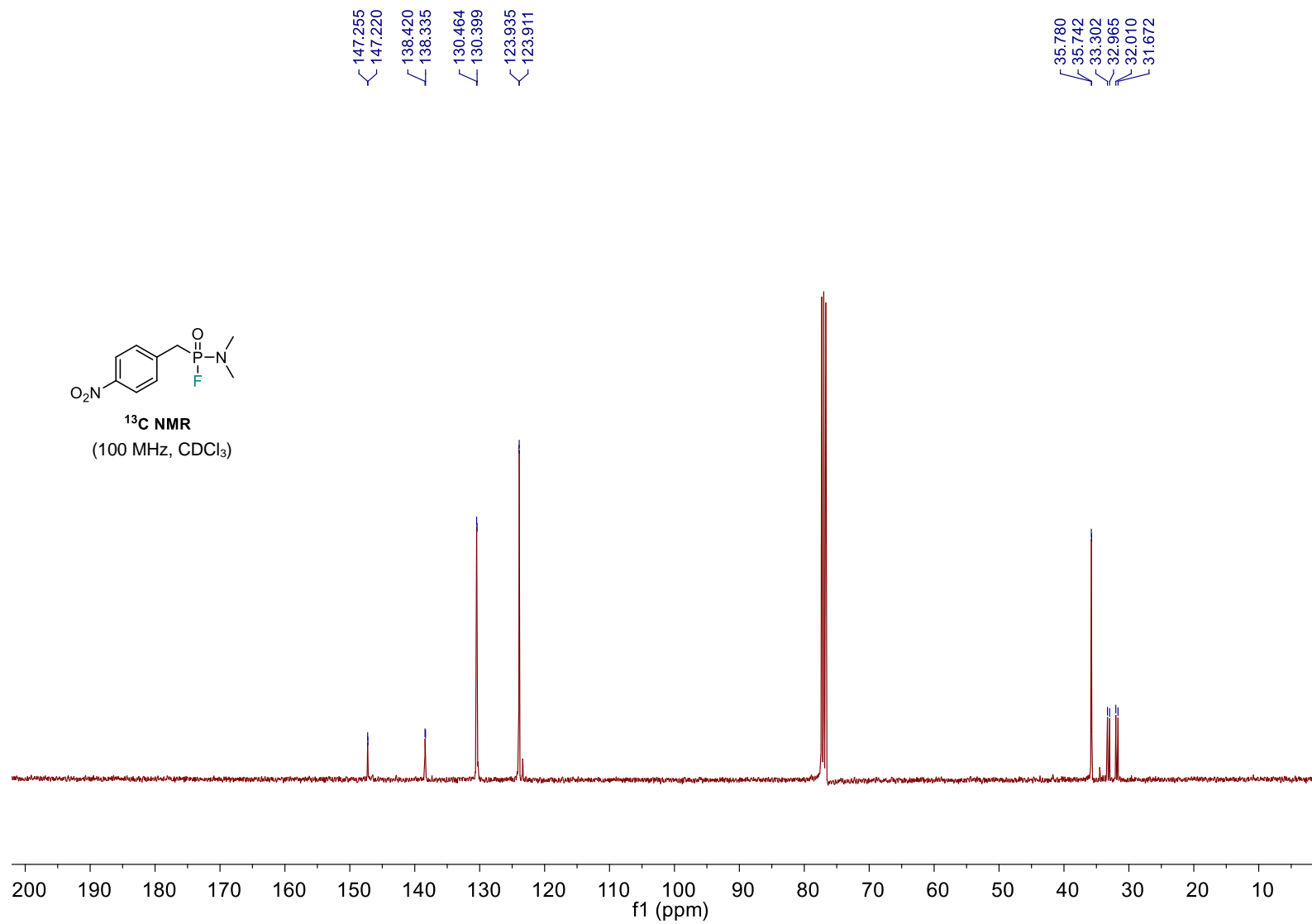


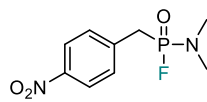
¹H NMR
(400 MHz, CDCl₃)



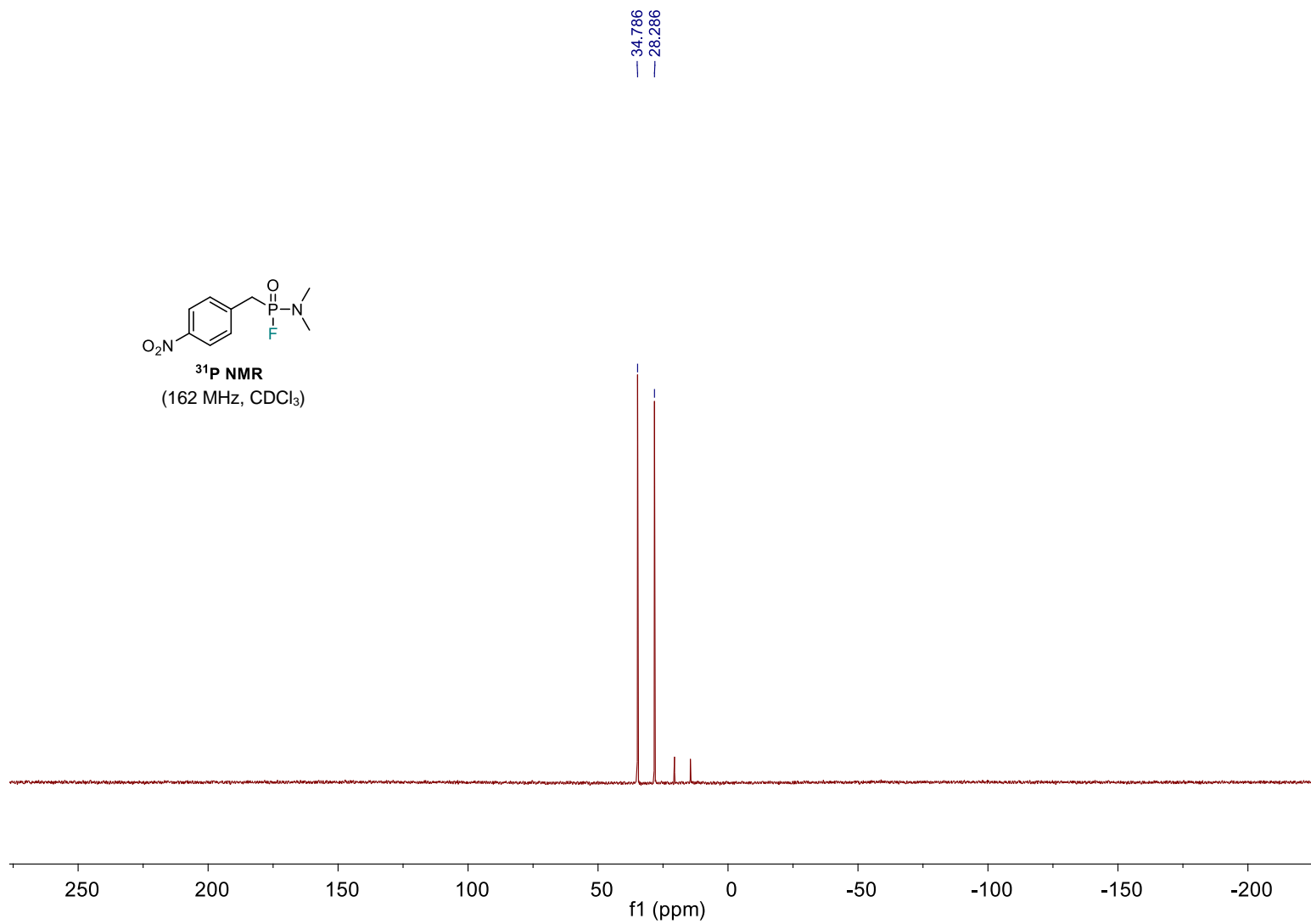


¹³C NMR
(100 MHz, CDCl₃)

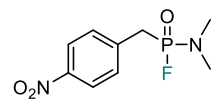




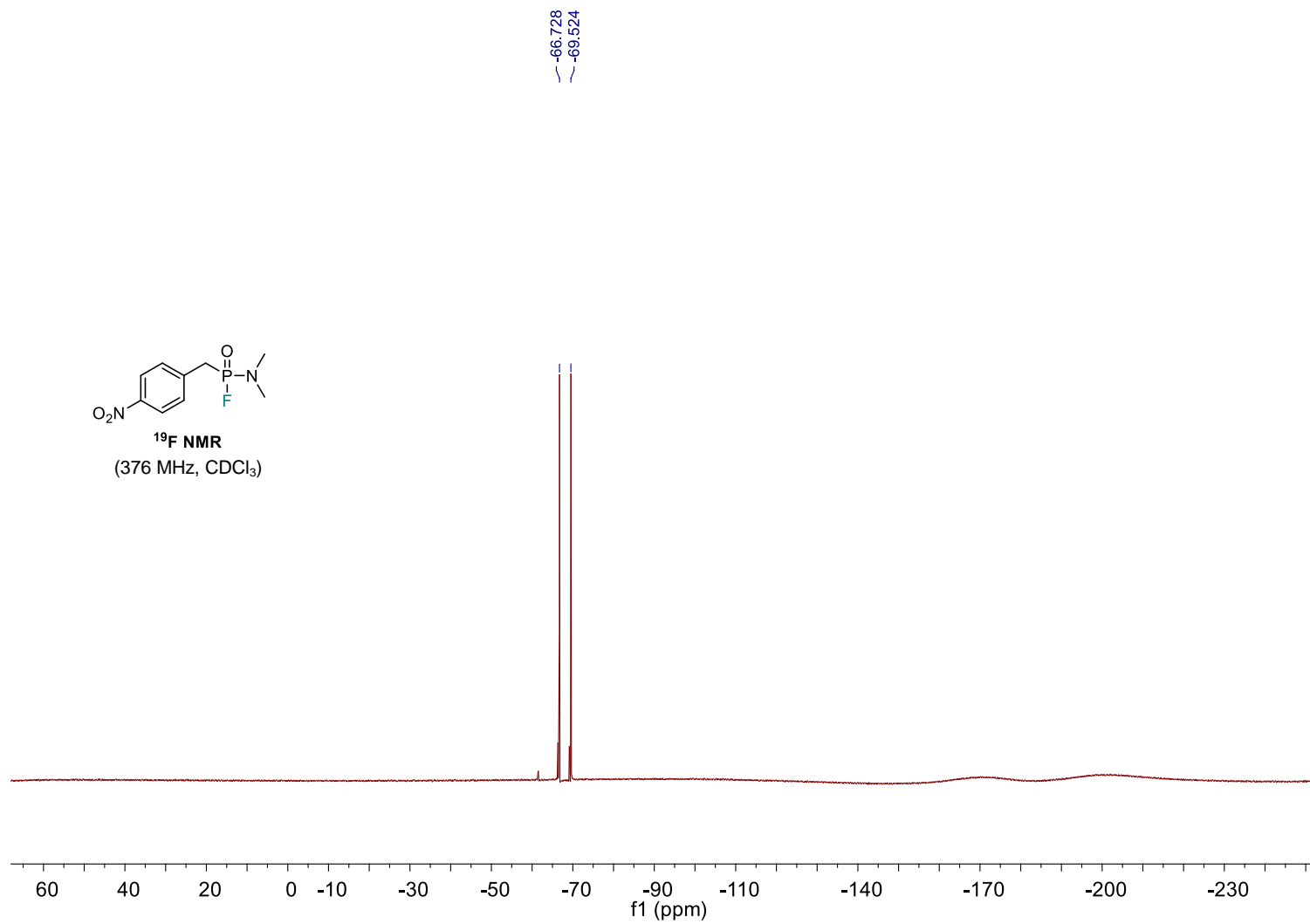
³¹P NMR
(162 MHz, CDCl₃)



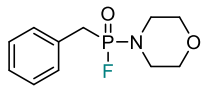
S359



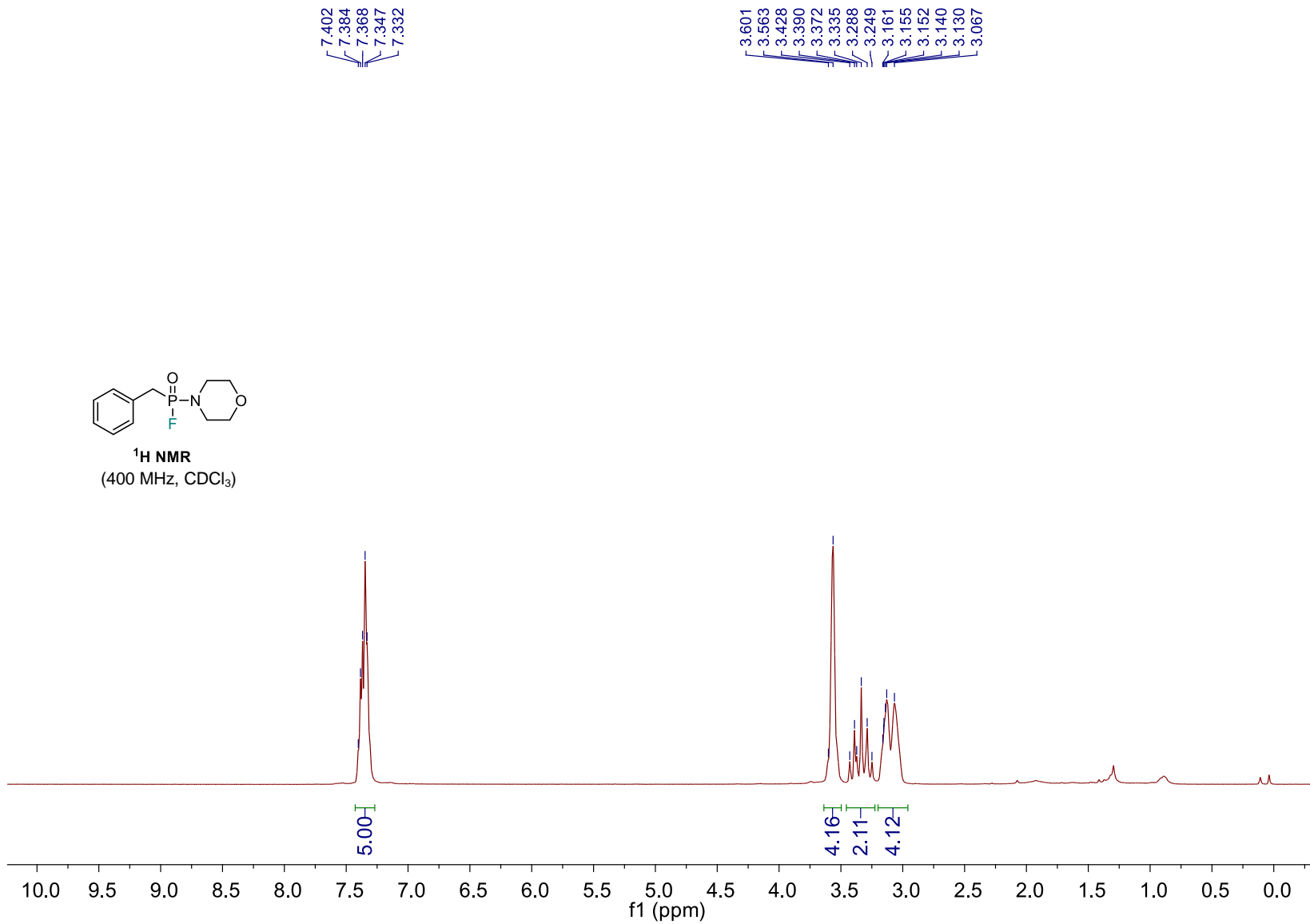
¹⁹F NMR
(376 MHz, CDCl₃)



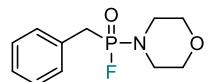
S360



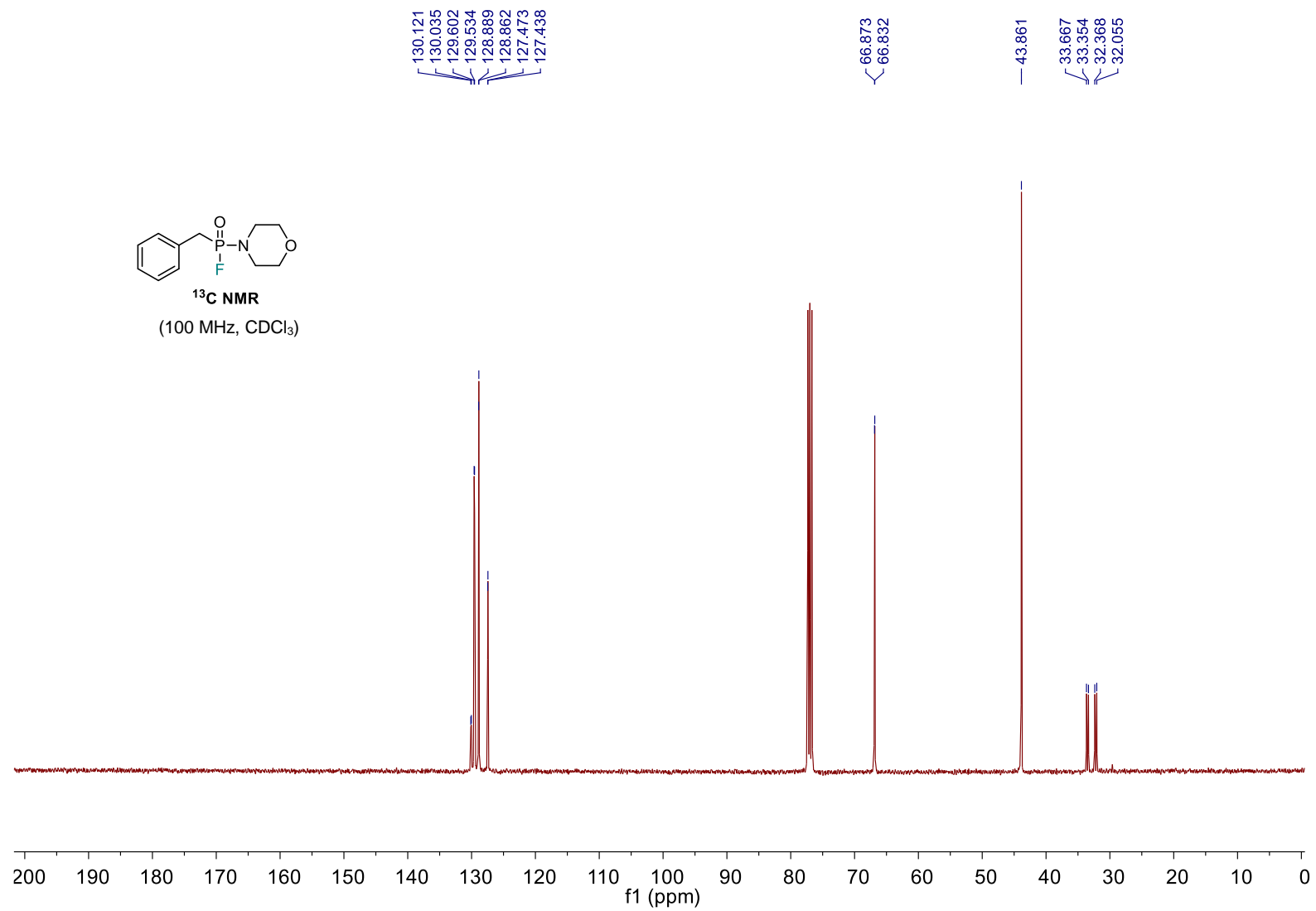
¹H NMR
(400 MHz, CDCl₃)

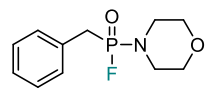


S361

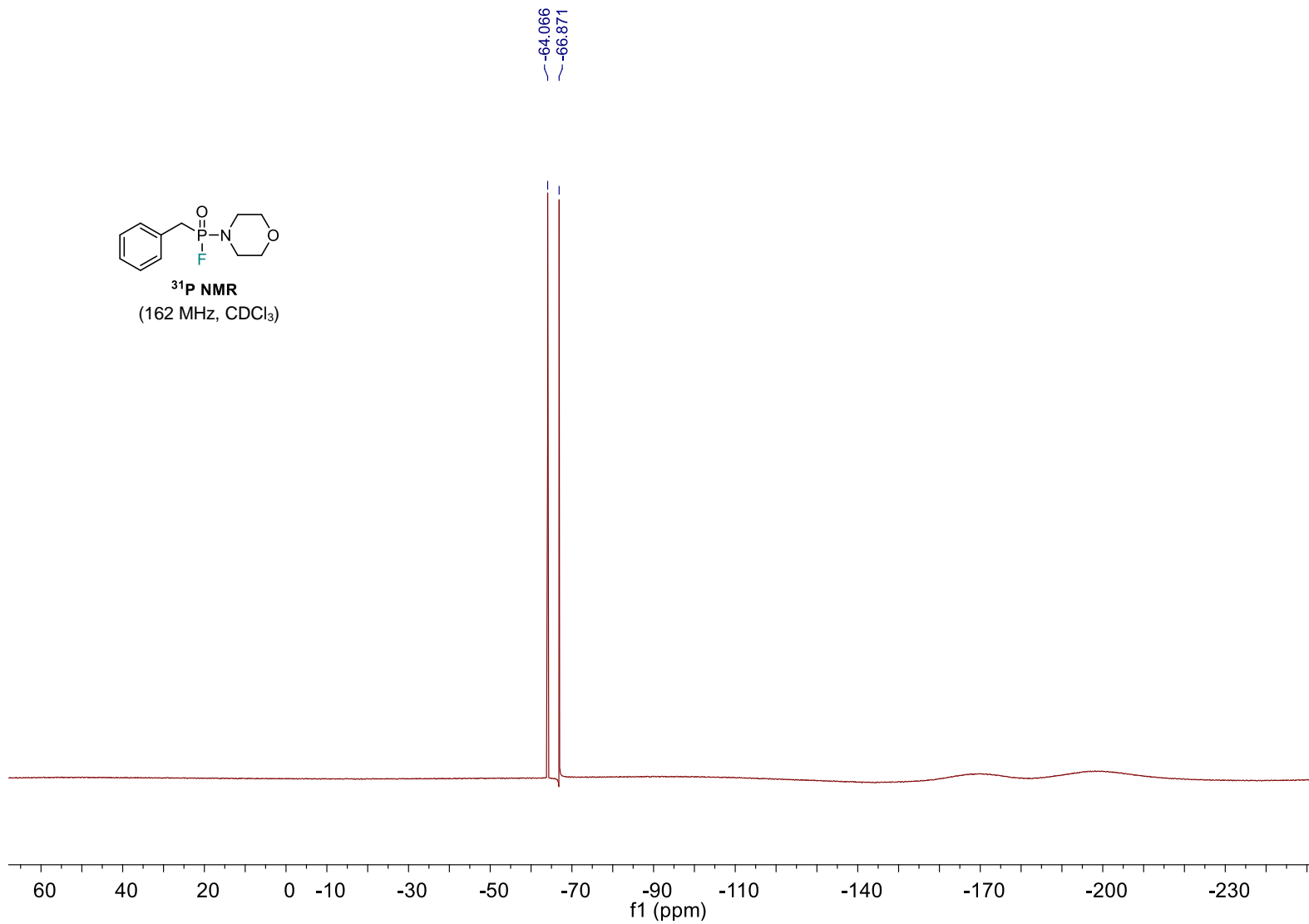


¹³C NMR
(100 MHz, CDCl₃)

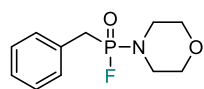




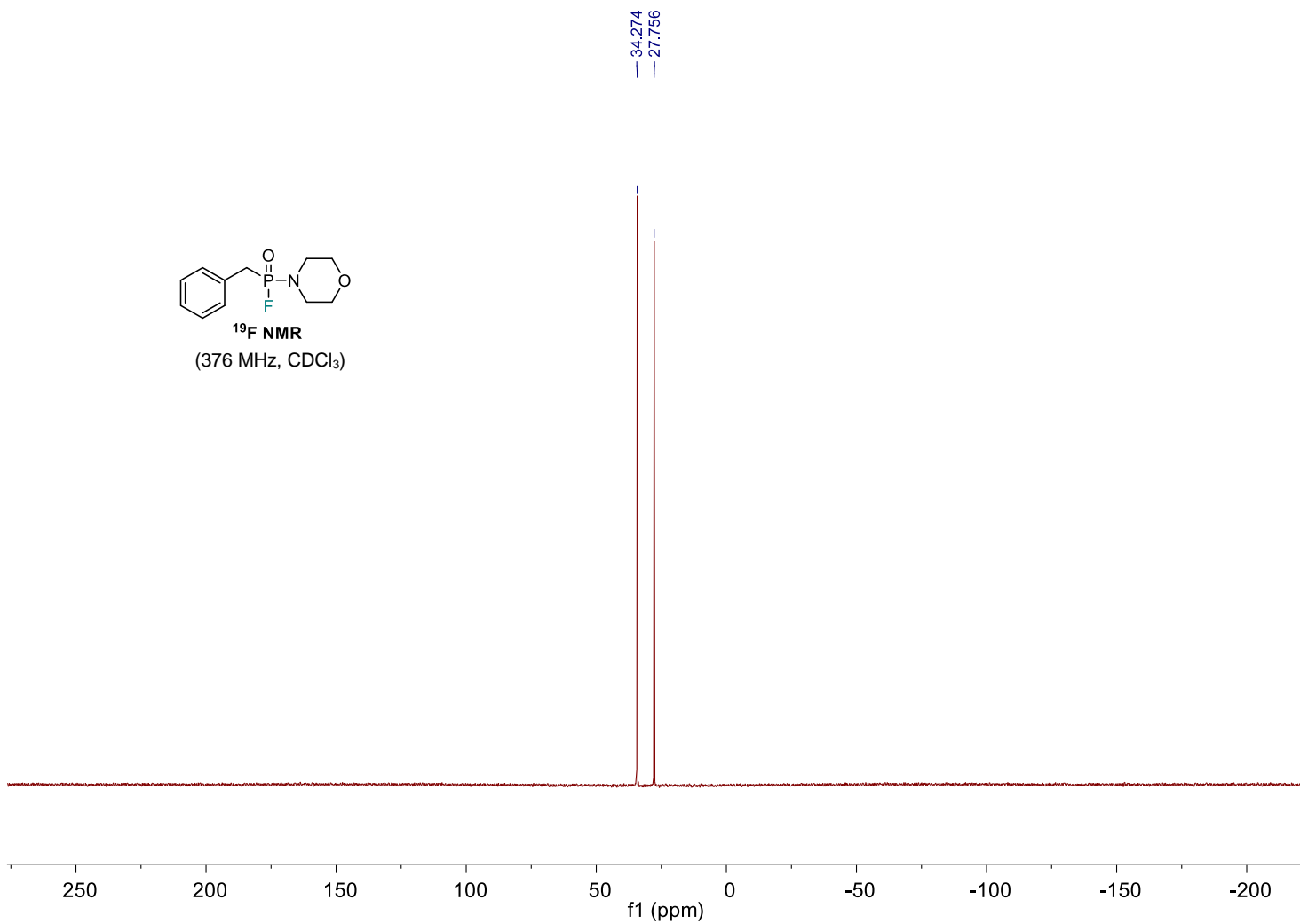
³¹P NMR
(162 MHz, CDCl₃)



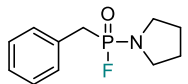
S363



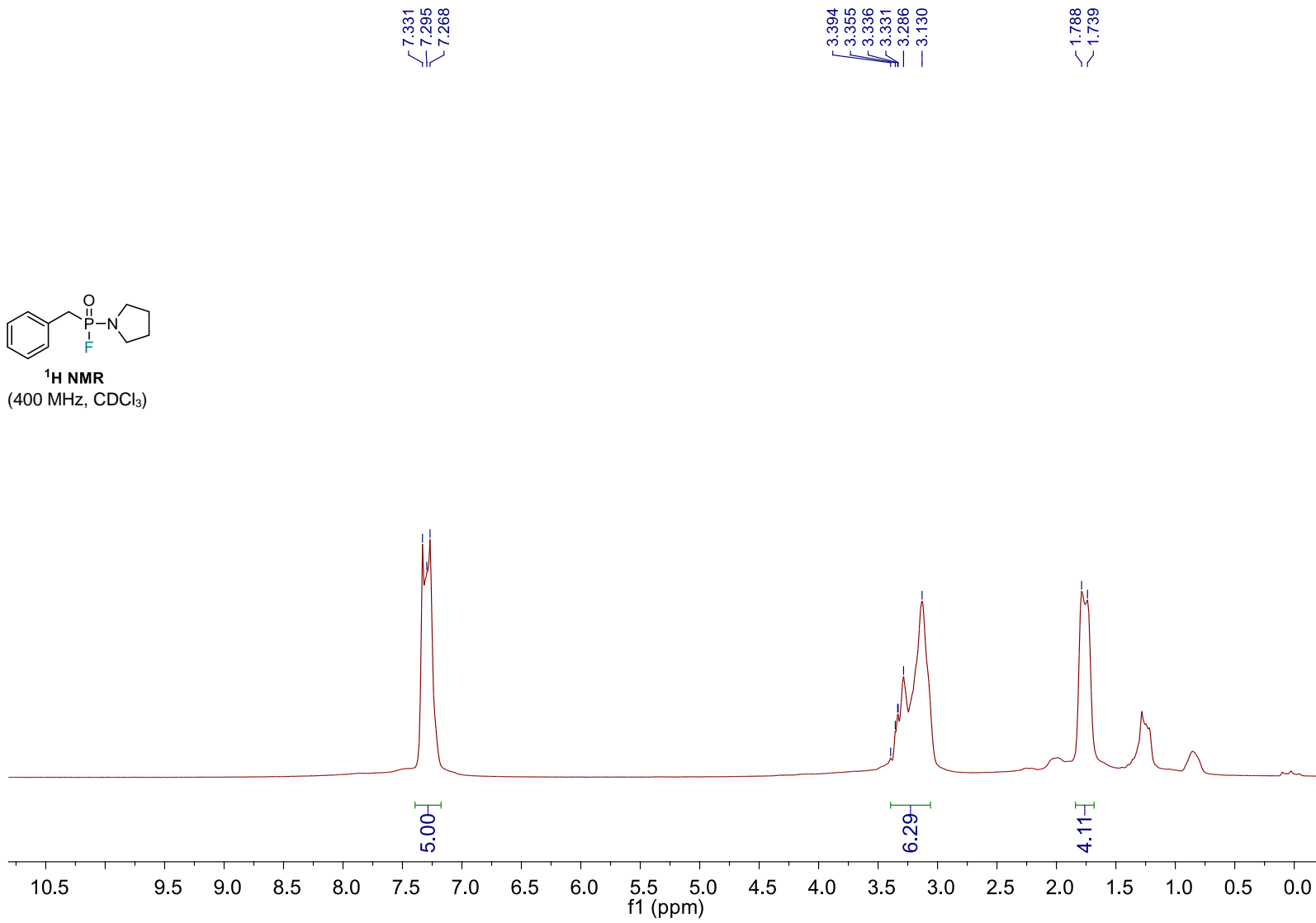
¹⁹F NMR
(376 MHz, CDCl₃)

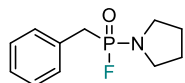


S364

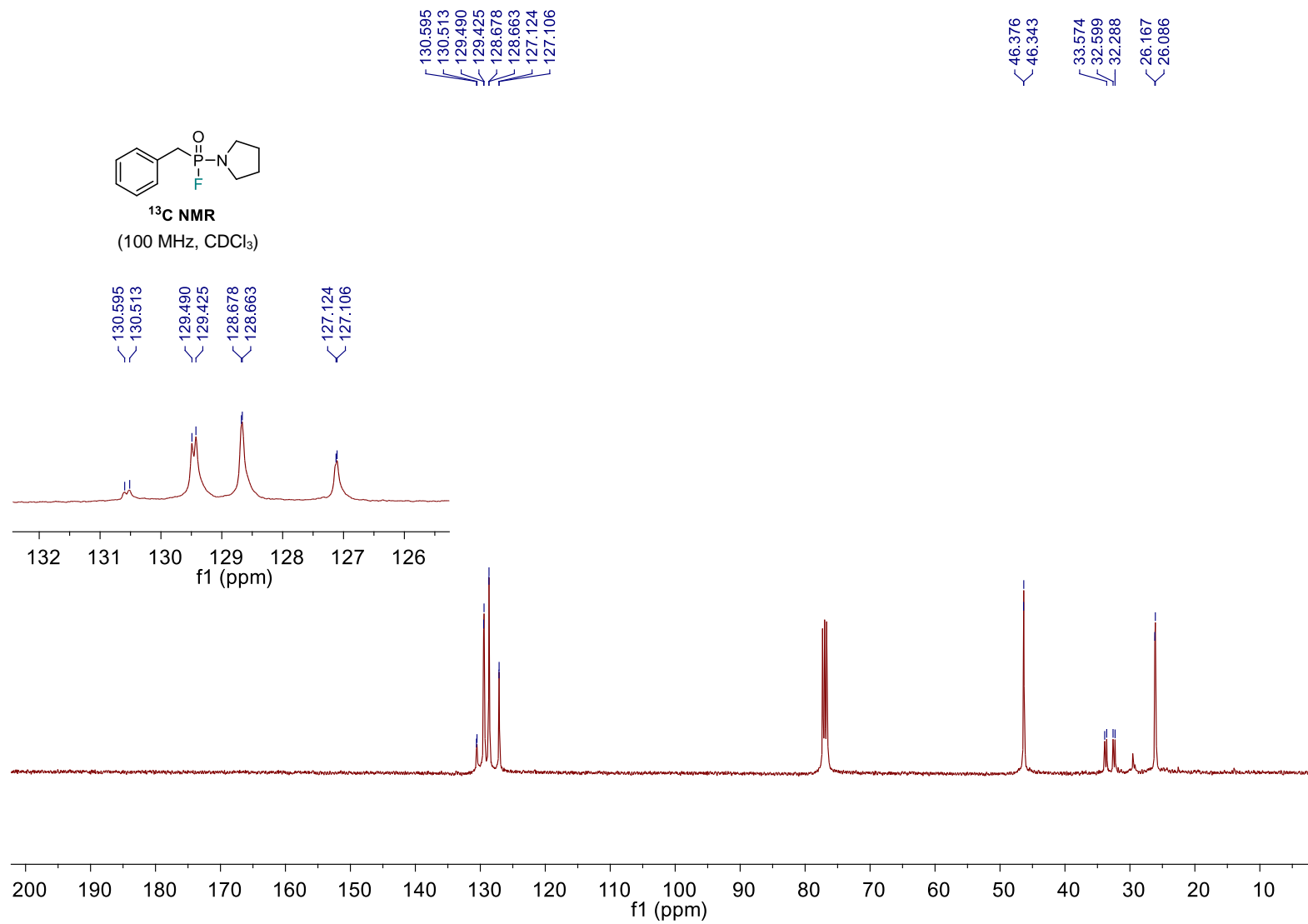


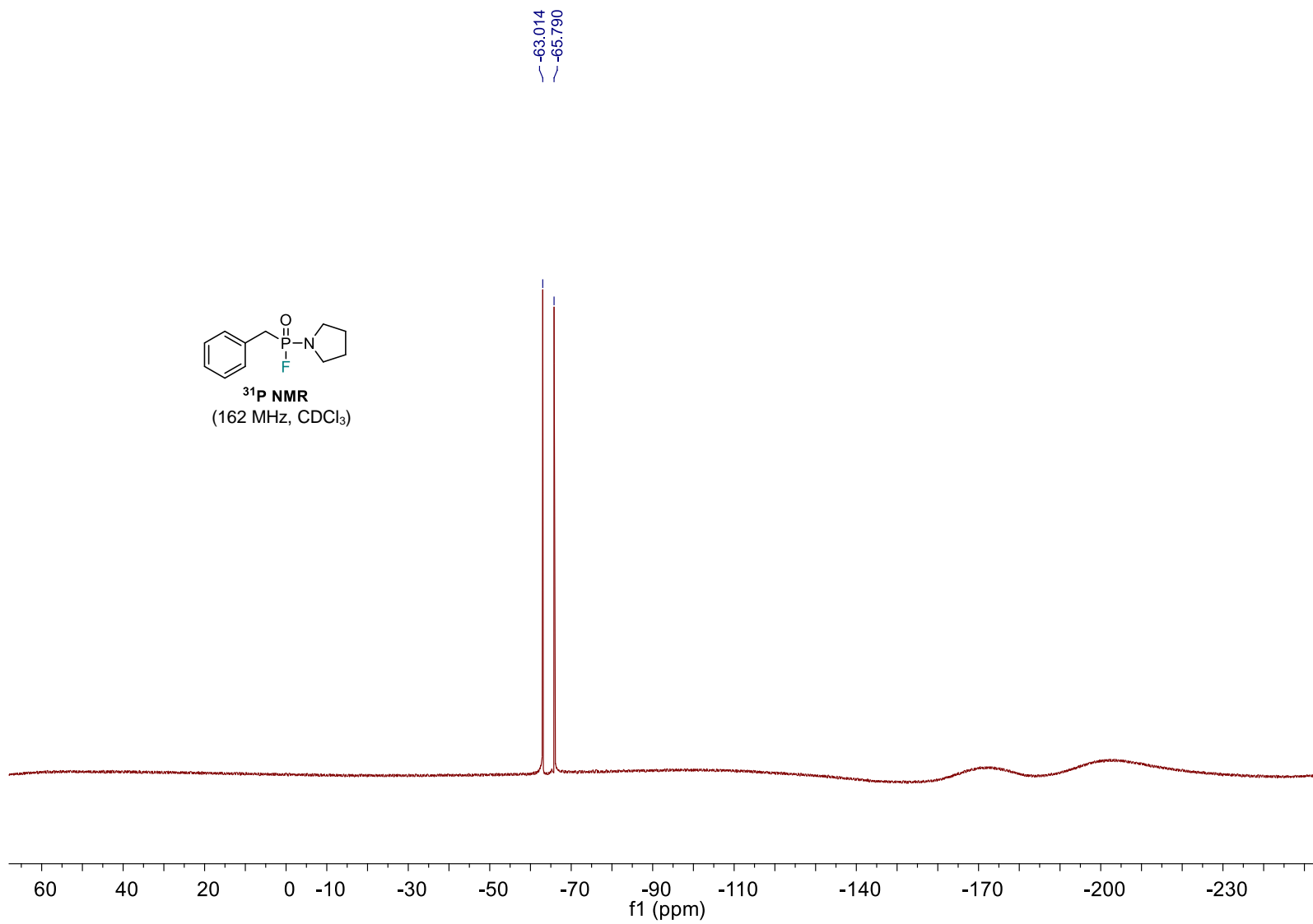
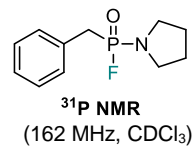
¹H NMR
(400 MHz, CDCl₃)

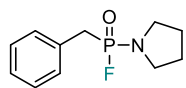




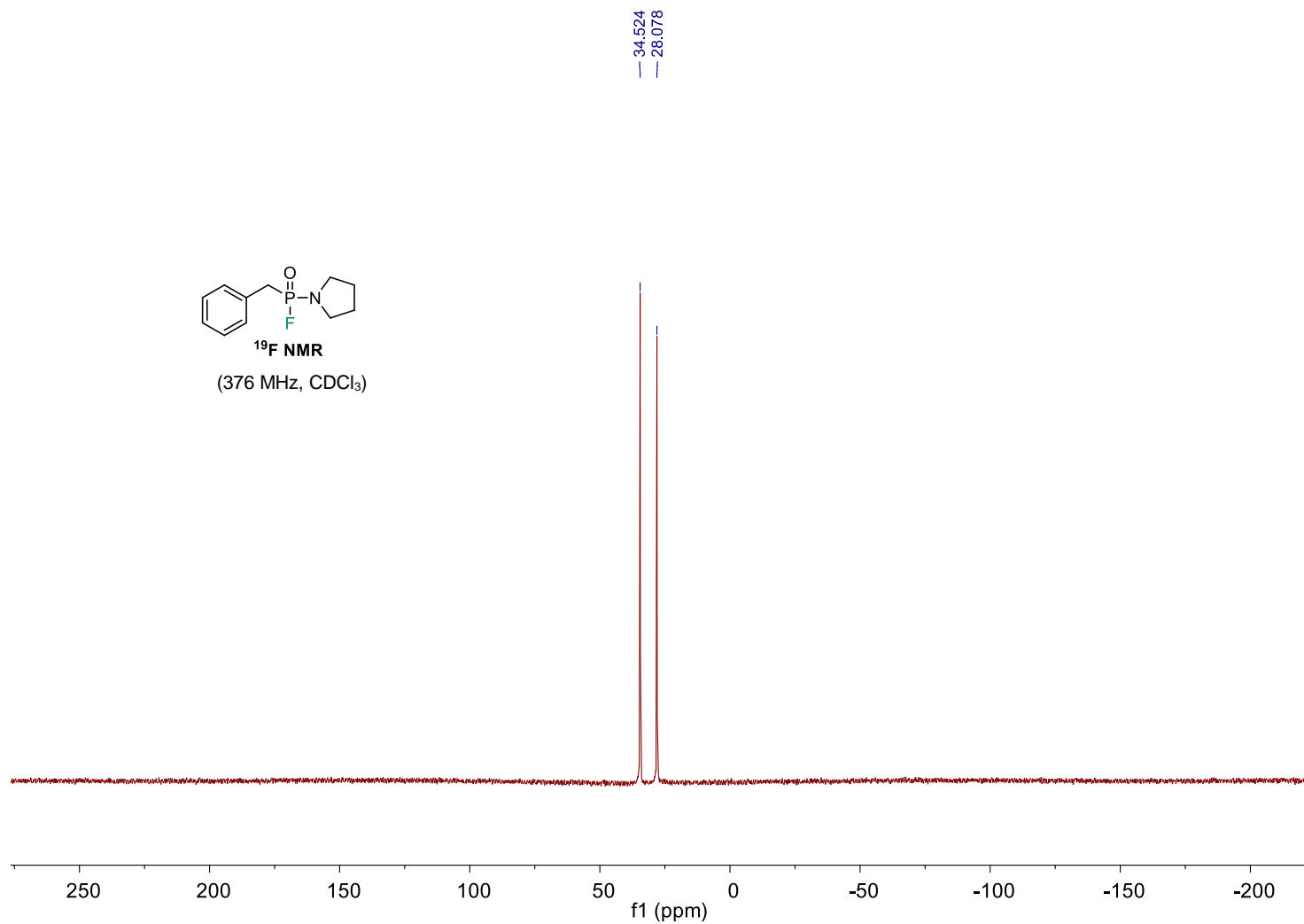
¹³C NMR
(100 MHz, CDCl₃)



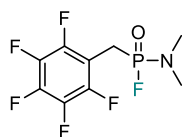




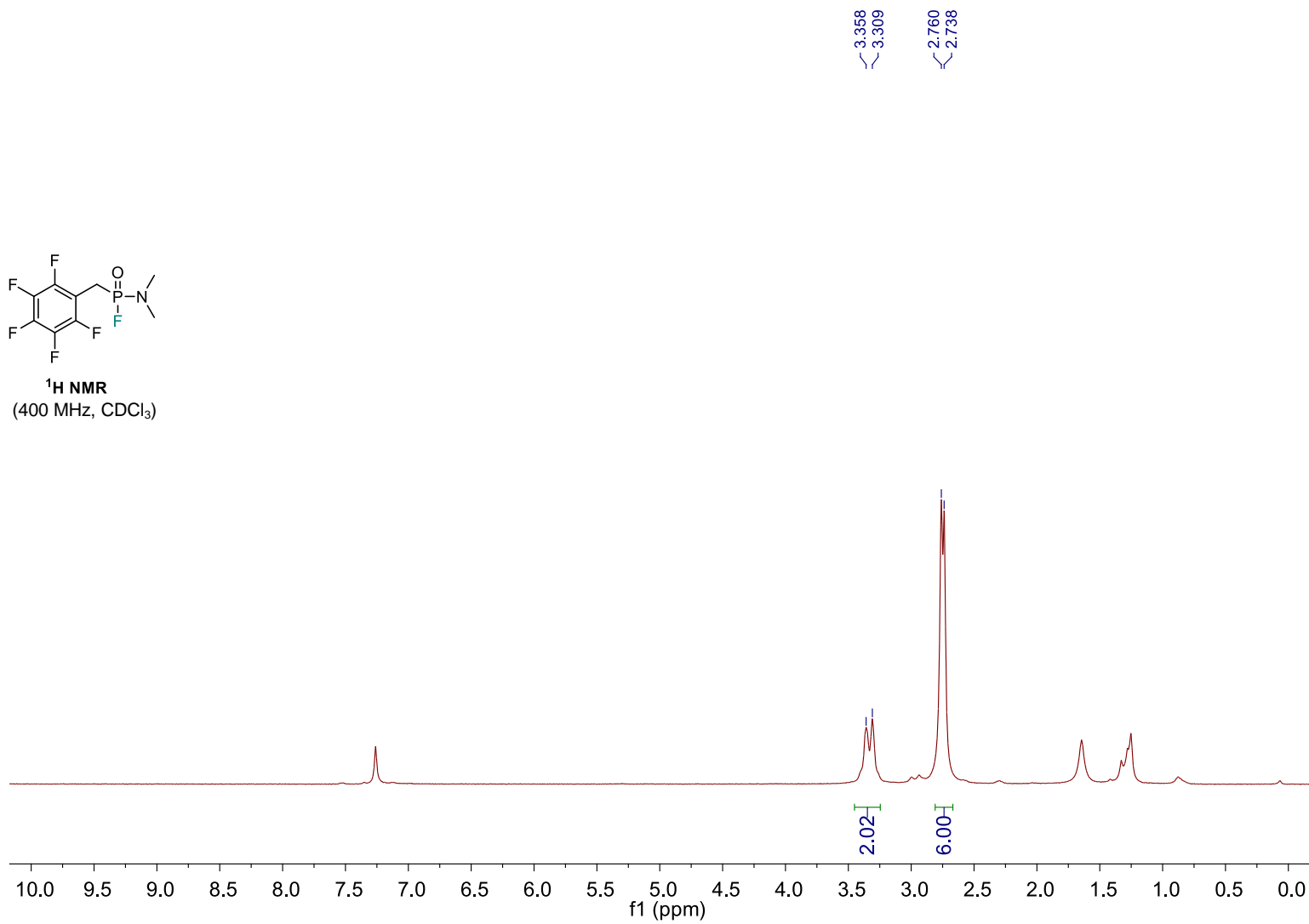
¹⁹F NMR
(376 MHz, CDCl₃)



S368



¹H NMR
(400 MHz, CDCl₃)



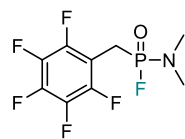
— 179.849

— 146.217
— 143.717
— 141.458
— 138.678

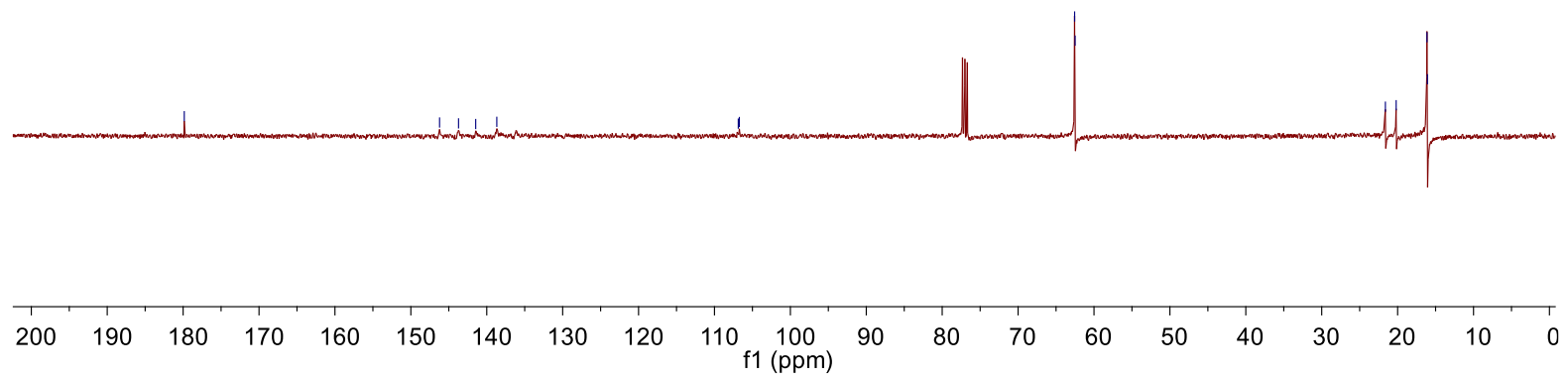
— 106.865
— 106.721

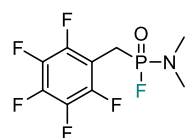
— 62.554
— 62.489

— 21.612
— 20.194
— 16.133
— 16.073

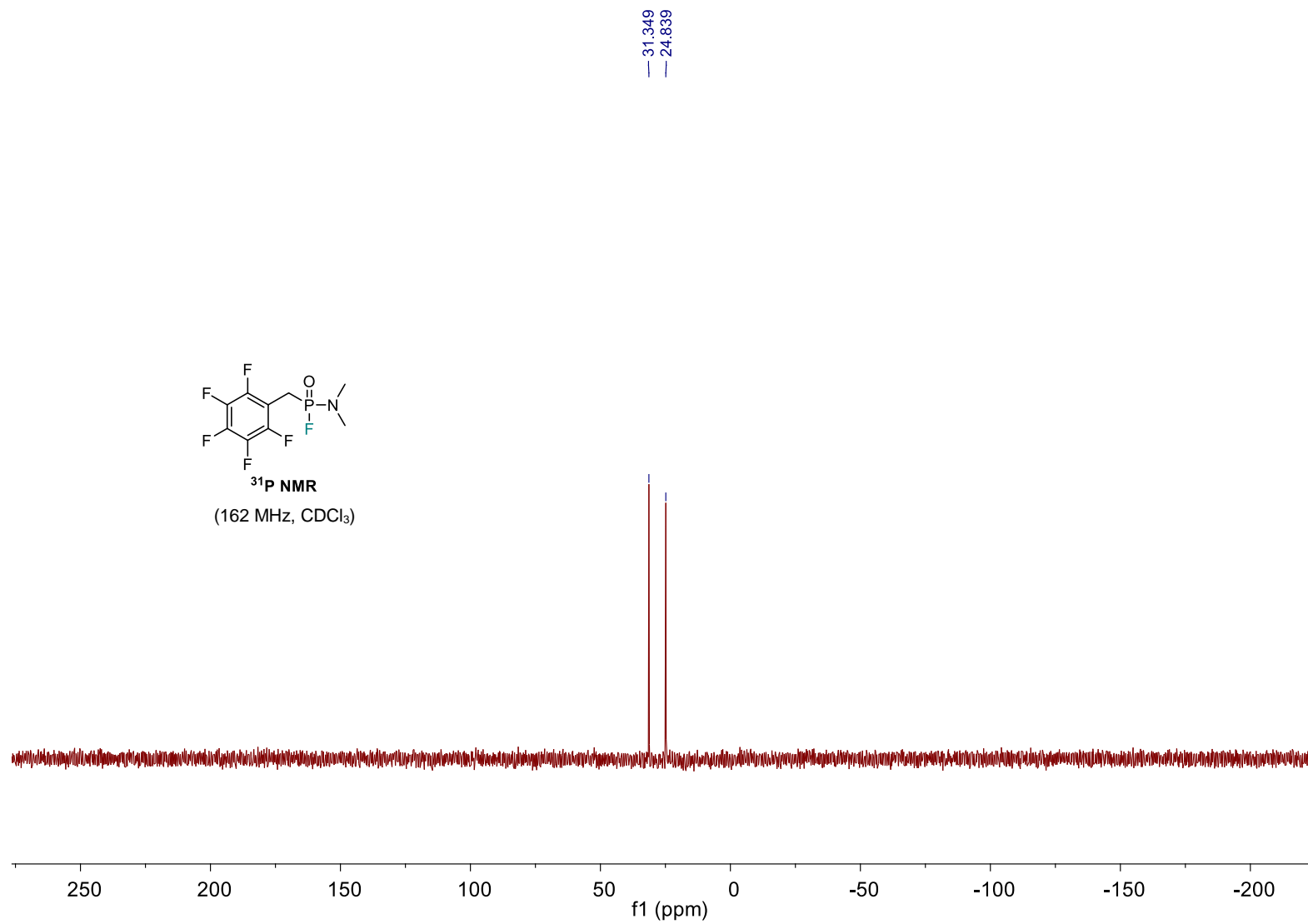


¹³C NMR
(100 MHz, CDCl₃)

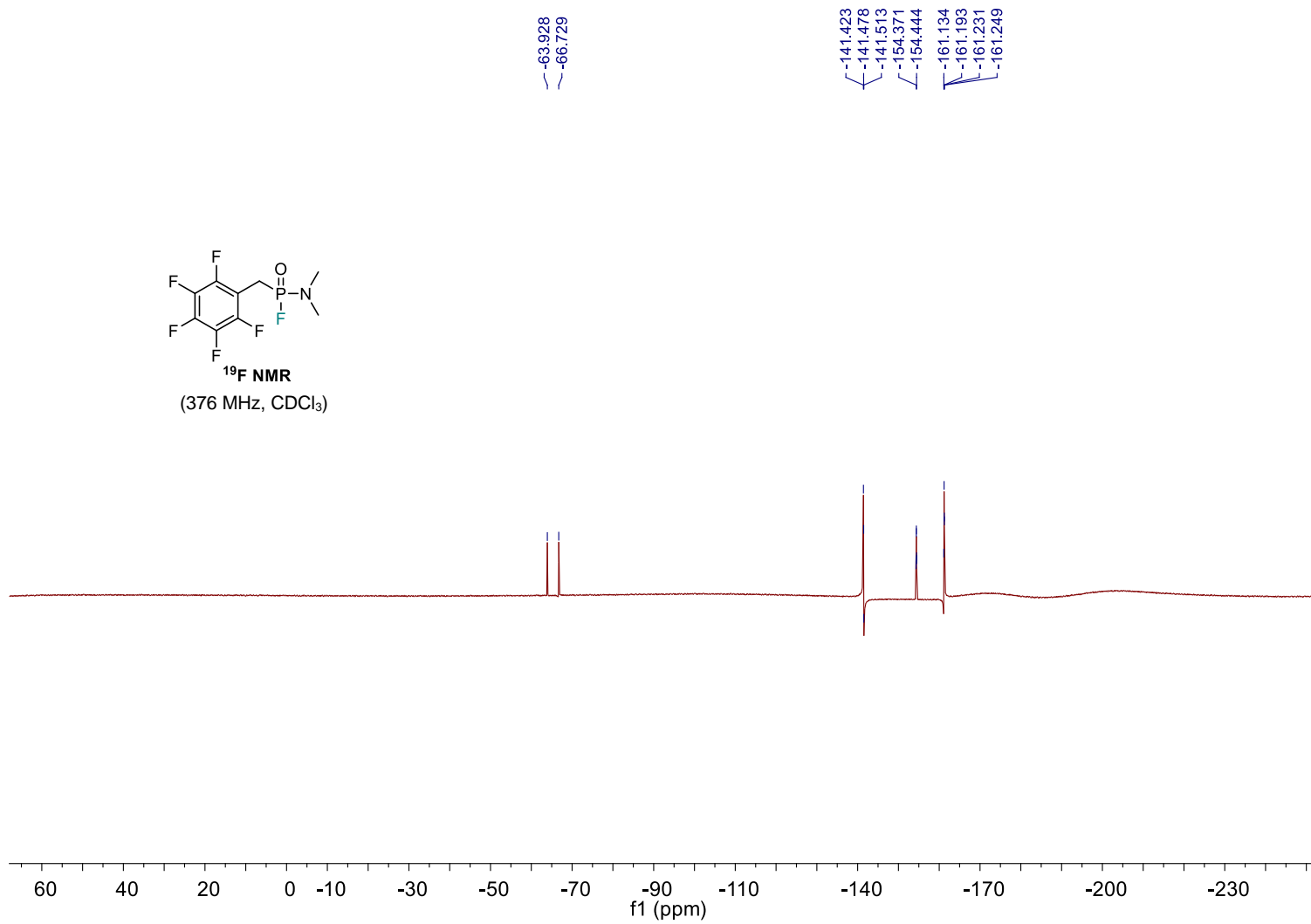
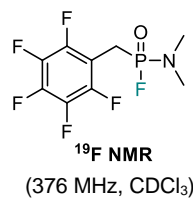


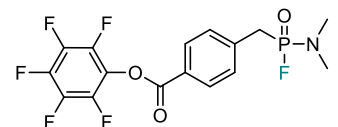


³¹P NMR
(162 MHz, CDCl₃)

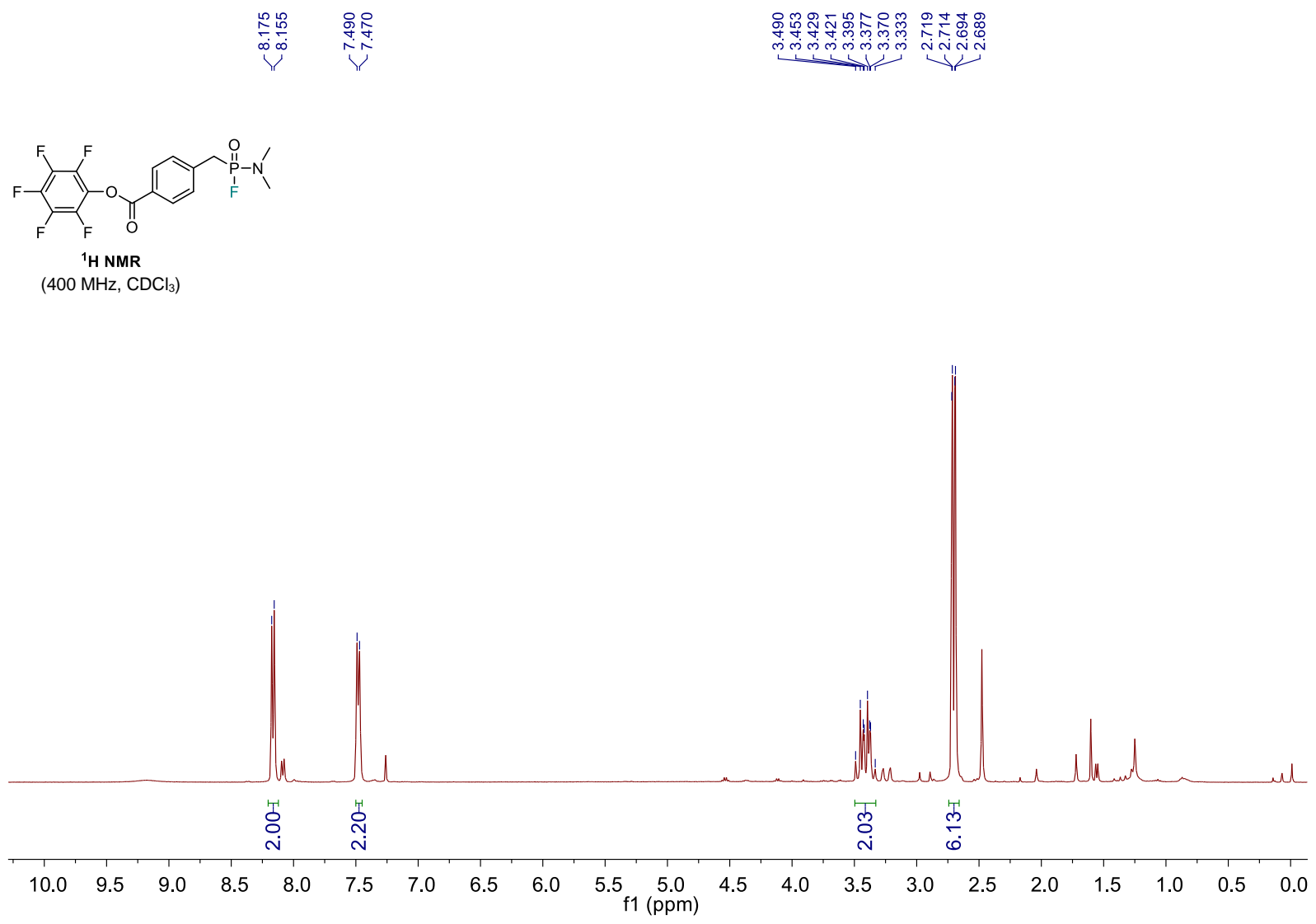


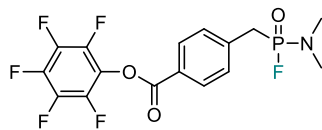
S371



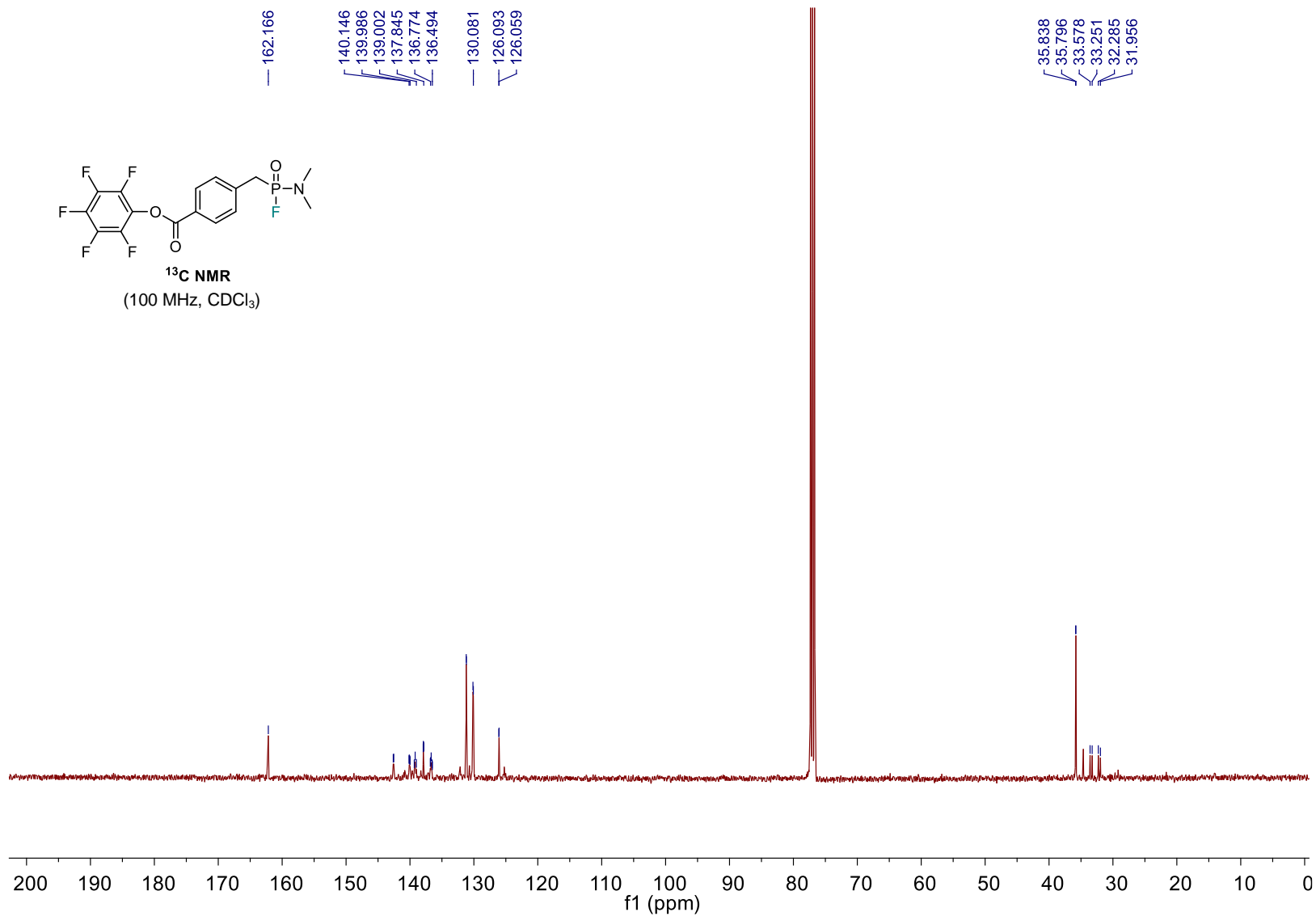


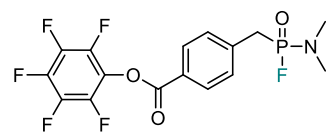
¹H NMR
(400 MHz, CDCl₃)



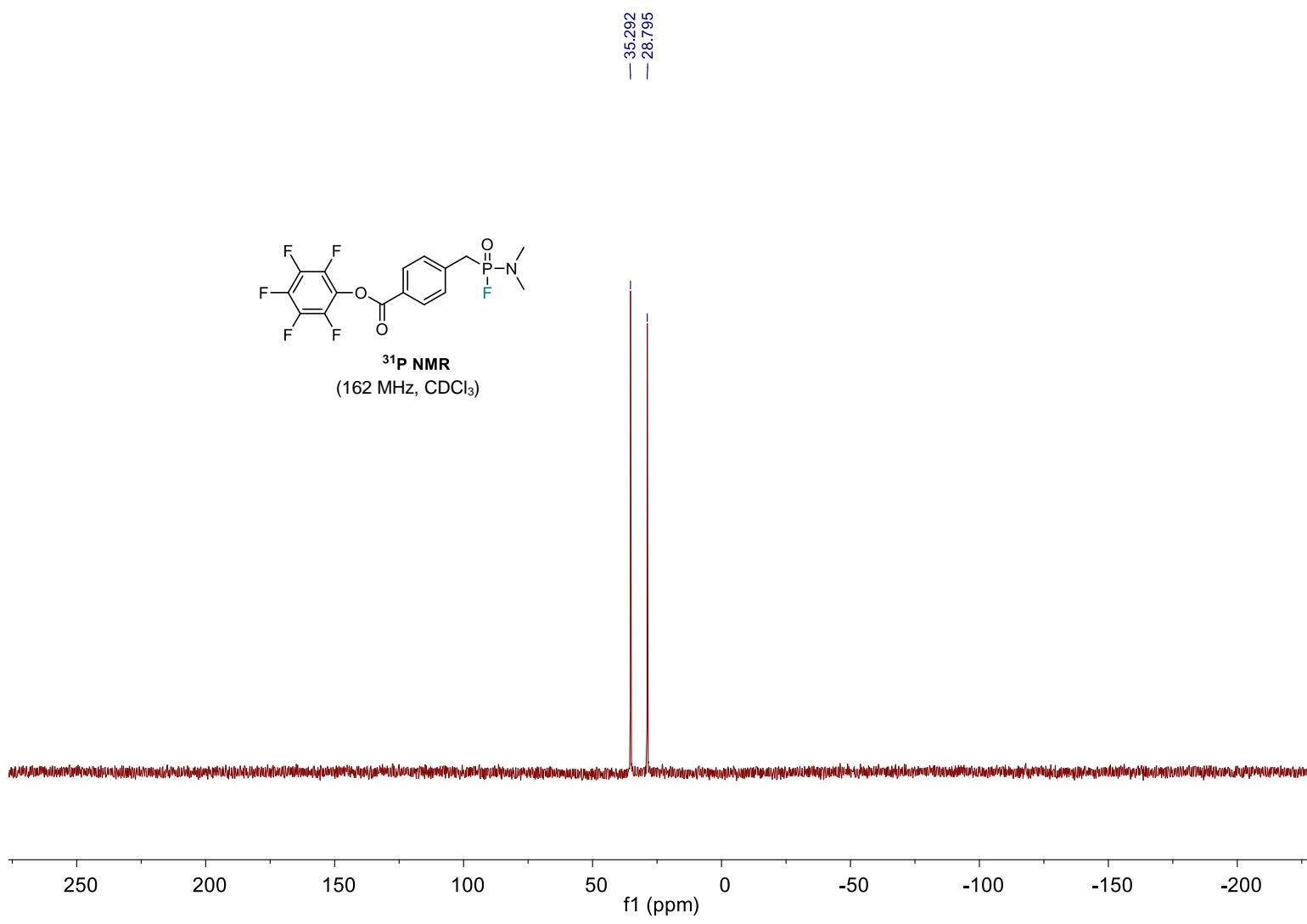


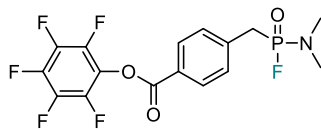
¹³C NMR
(100 MHz, CDCl₃)



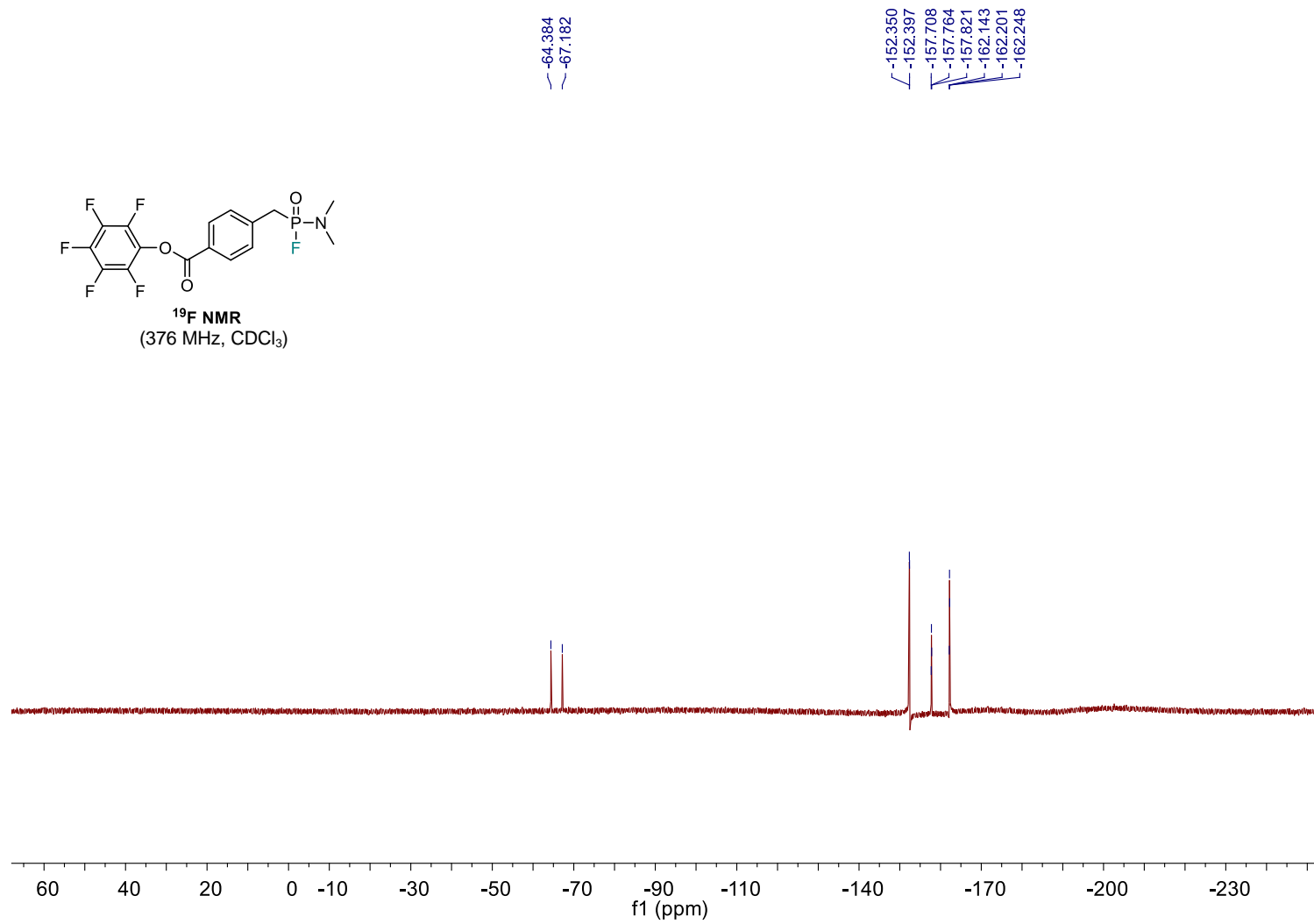


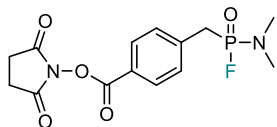
³¹P NMR
(162 MHz, CDCl₃)



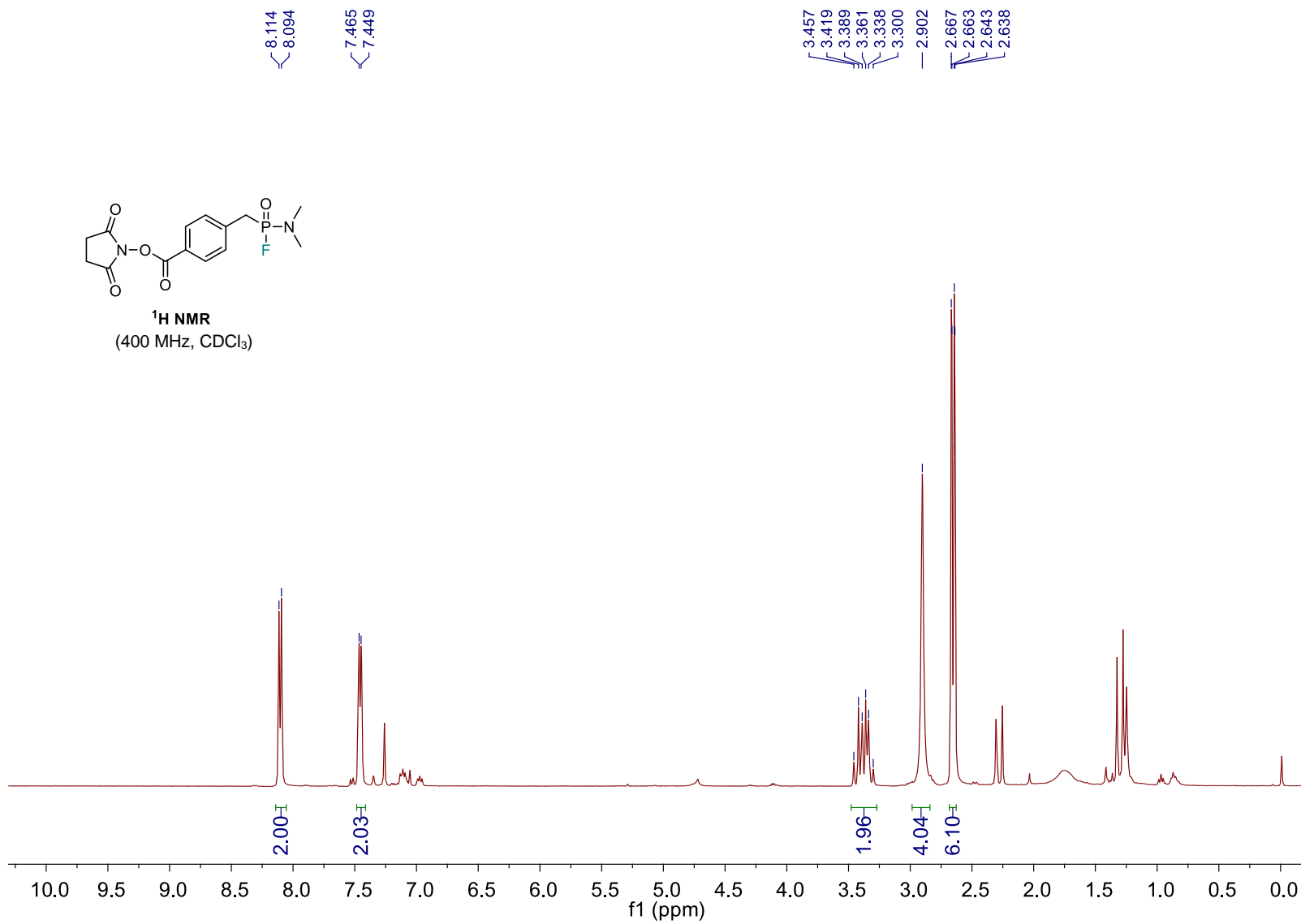


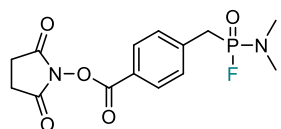
¹⁹F NMR
(376 MHz, CDCl₃)



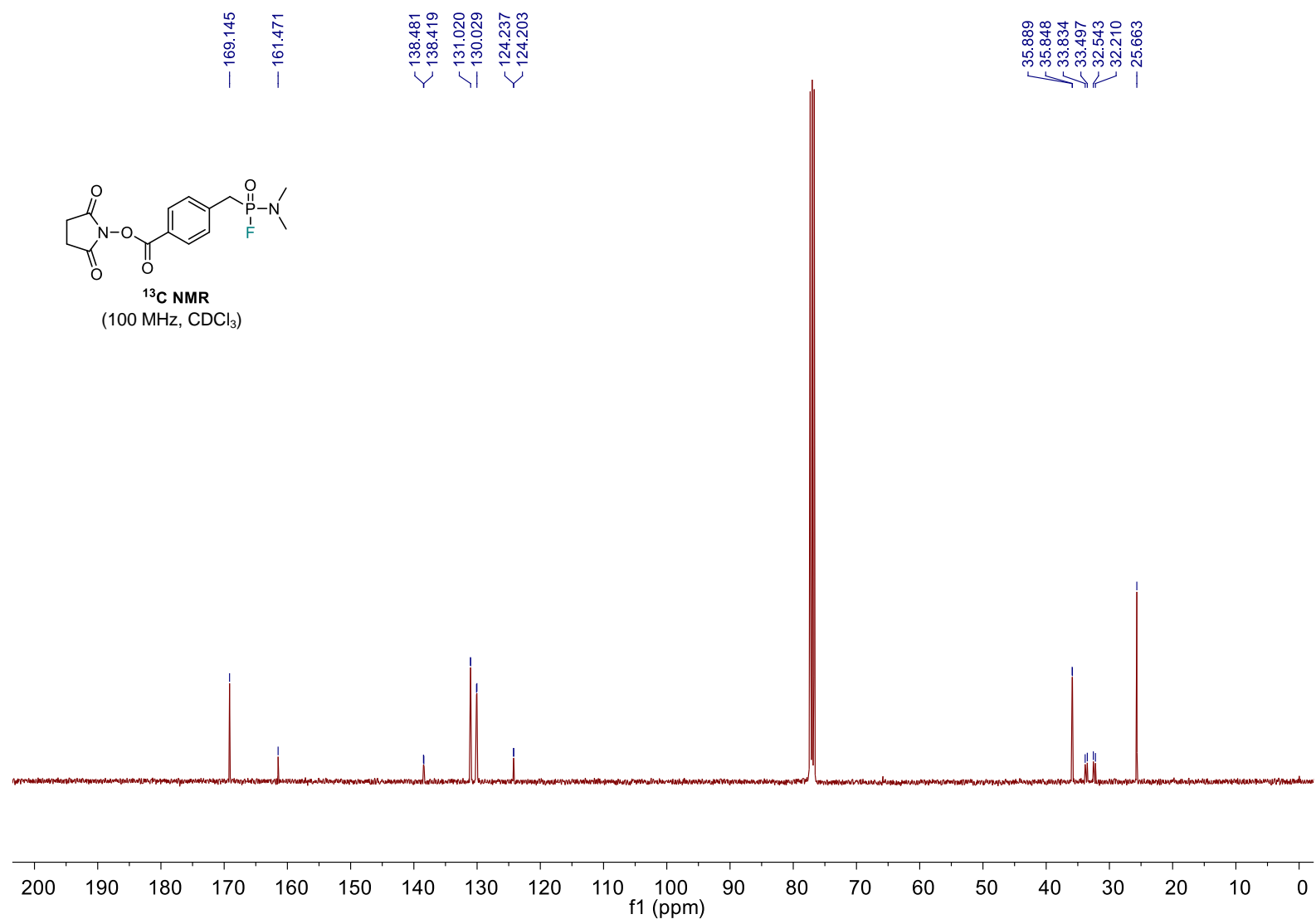


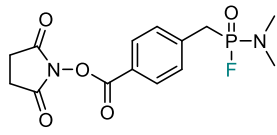
¹H NMR
(400 MHz, CDCl₃)



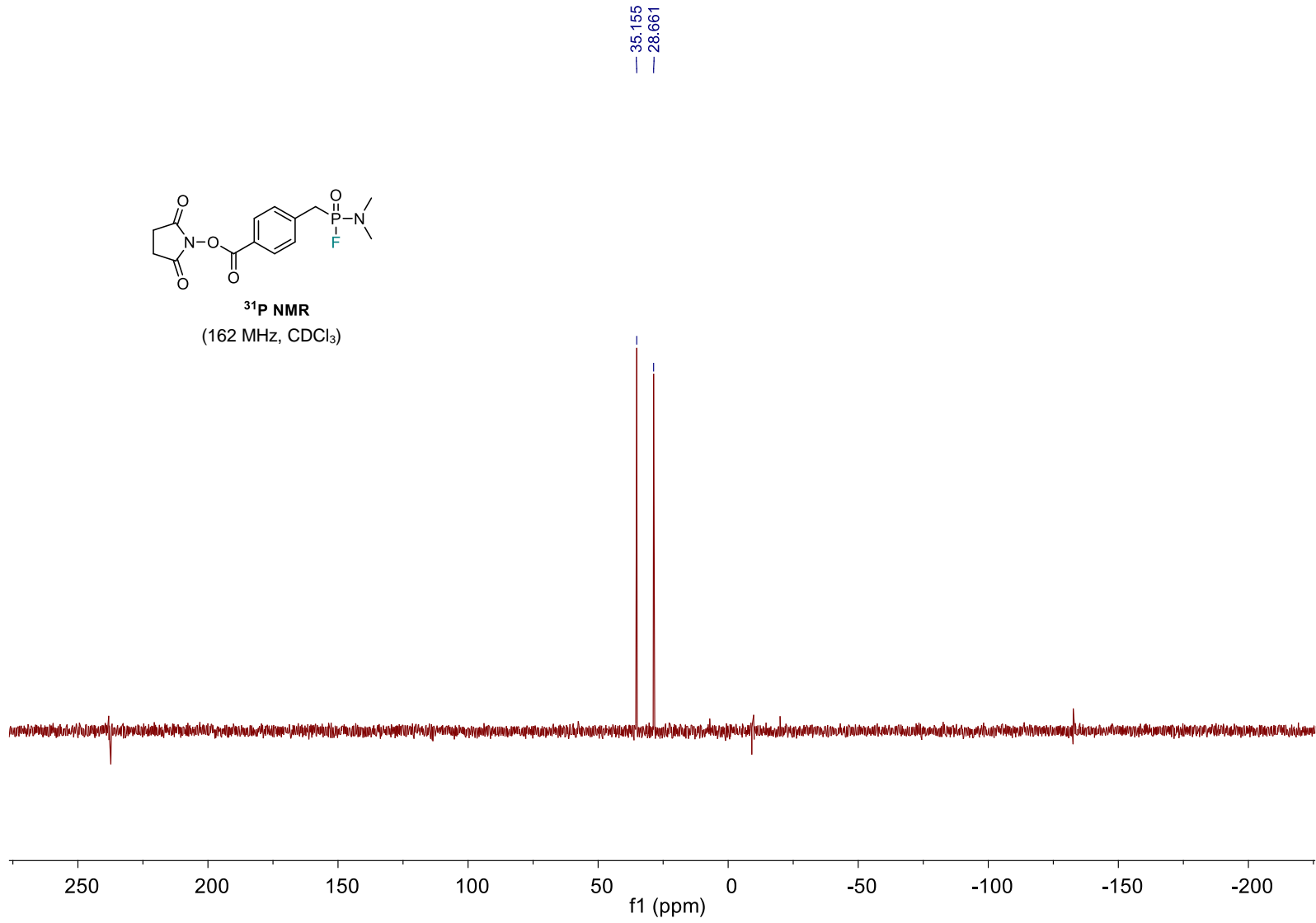


¹³C NMR
(100 MHz, CDCl₃)

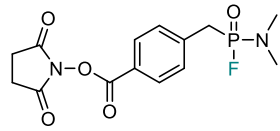




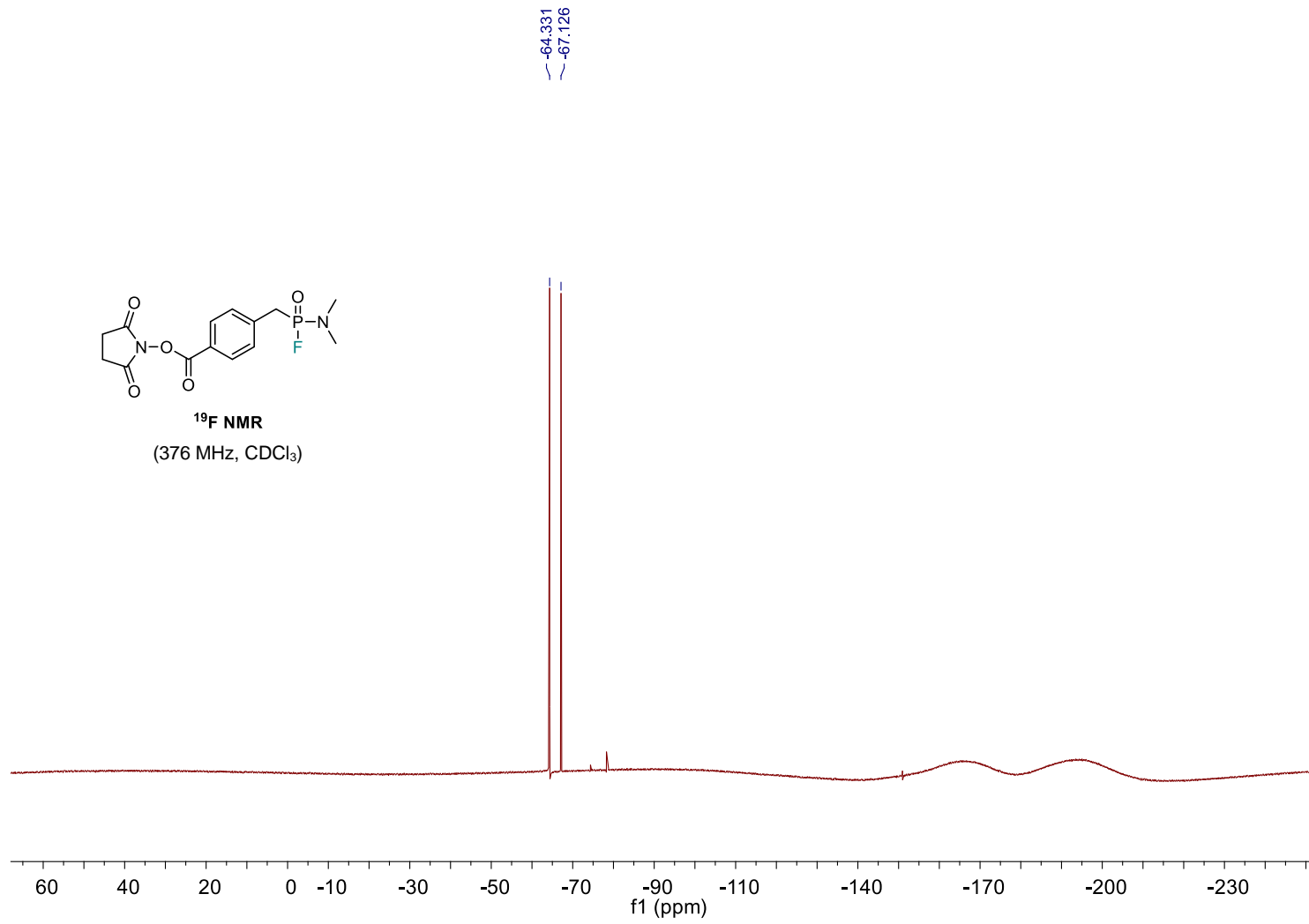
³¹P NMR
(162 MHz, CDCl₃)

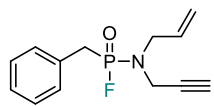


S379

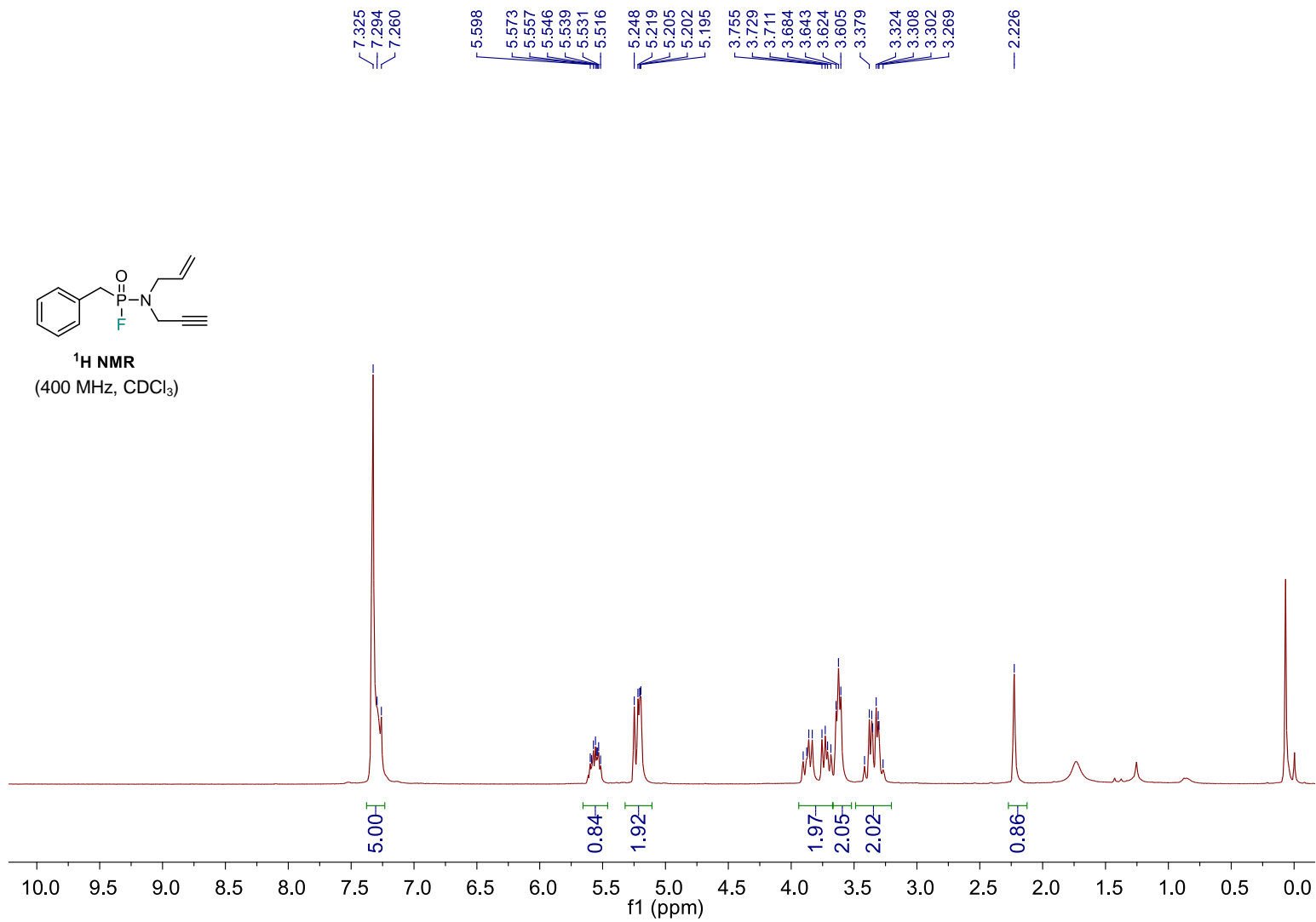


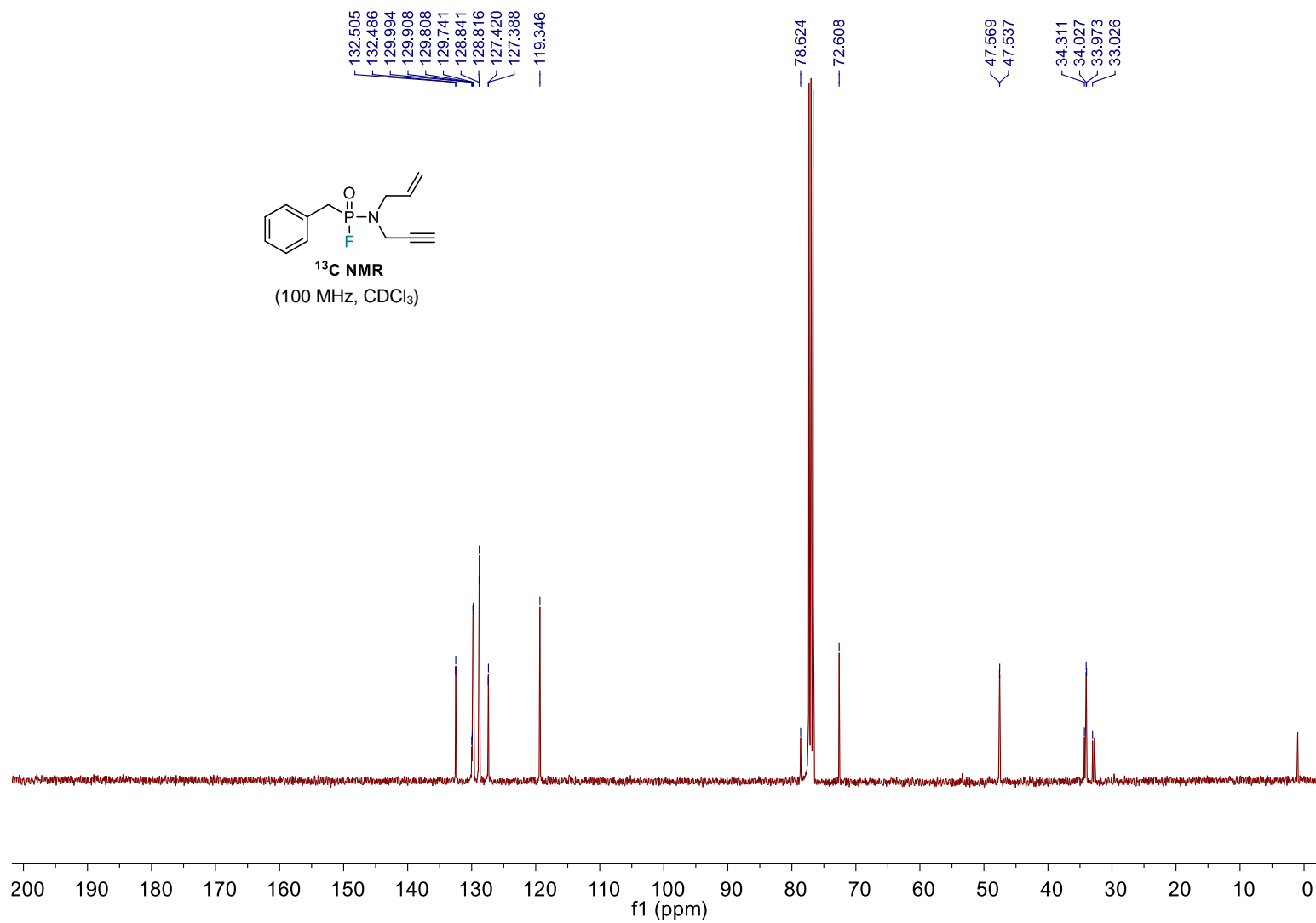
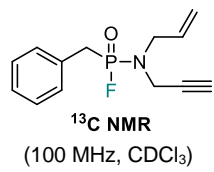
¹⁹F NMR
(376 MHz, CDCl₃)

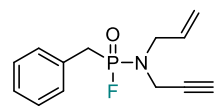




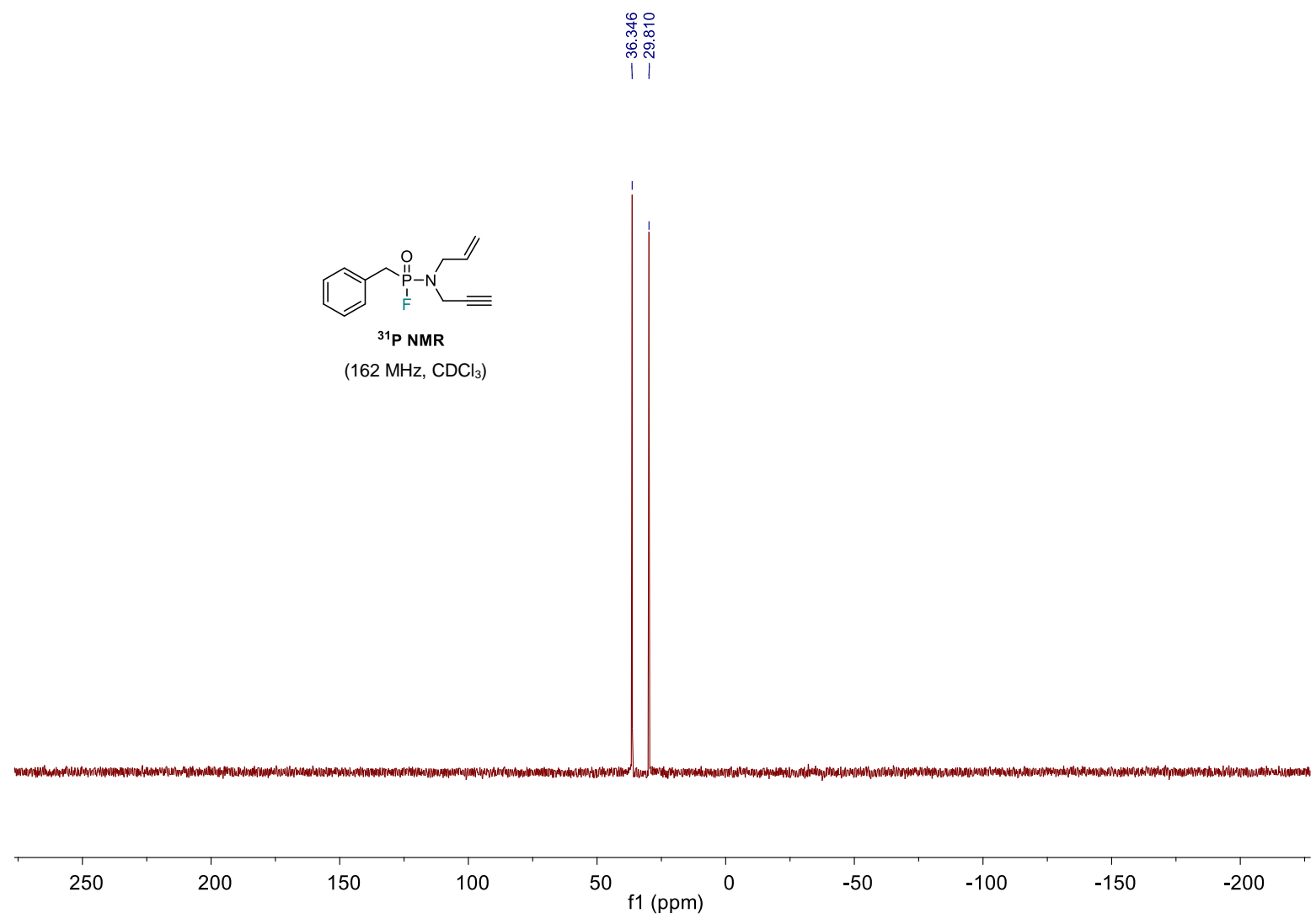
¹H NMR
(400 MHz, CDCl₃)

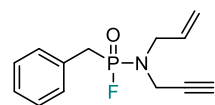




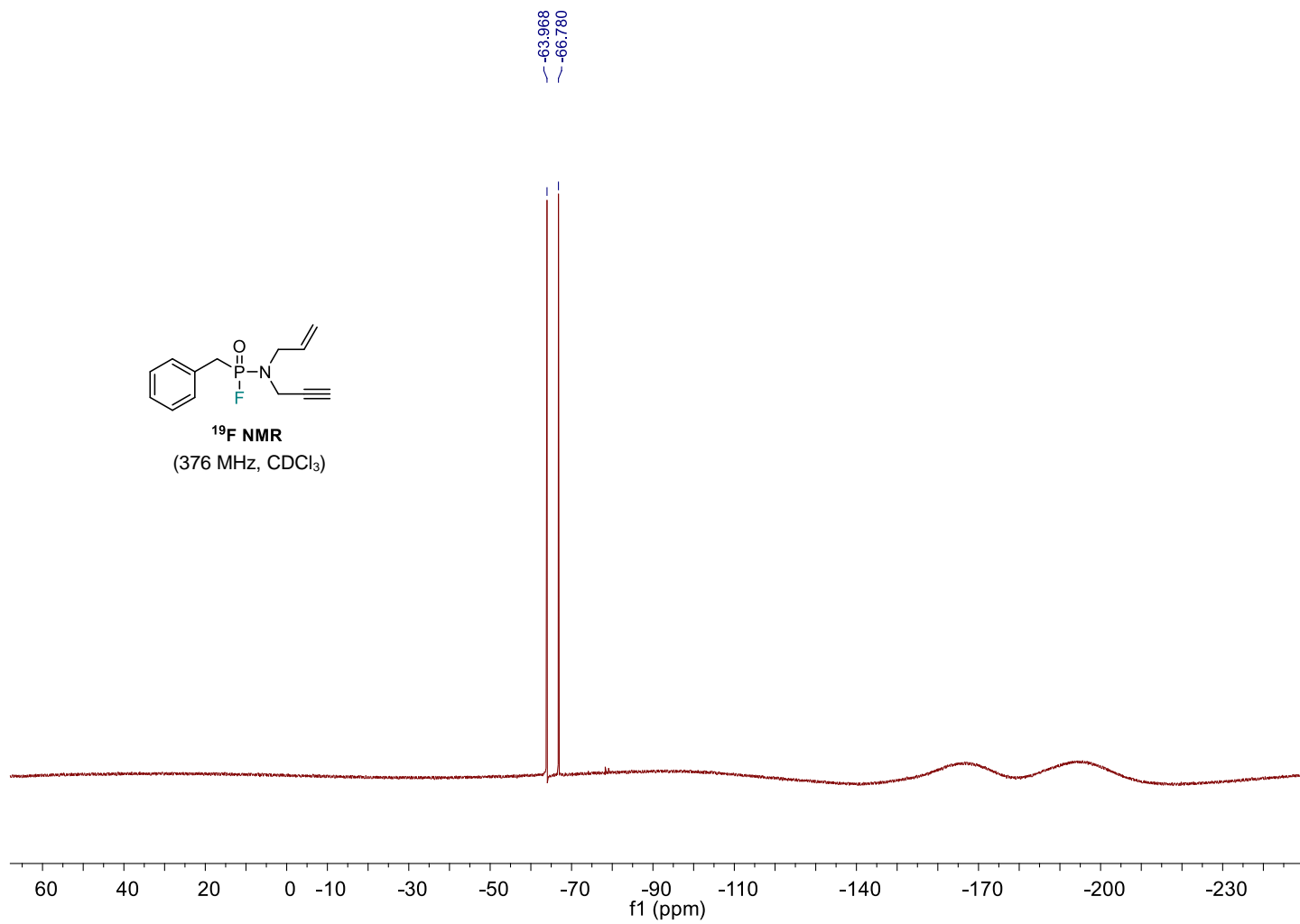


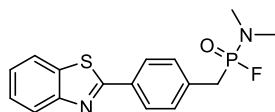
³¹P NMR
(162 MHz, CDCl₃)



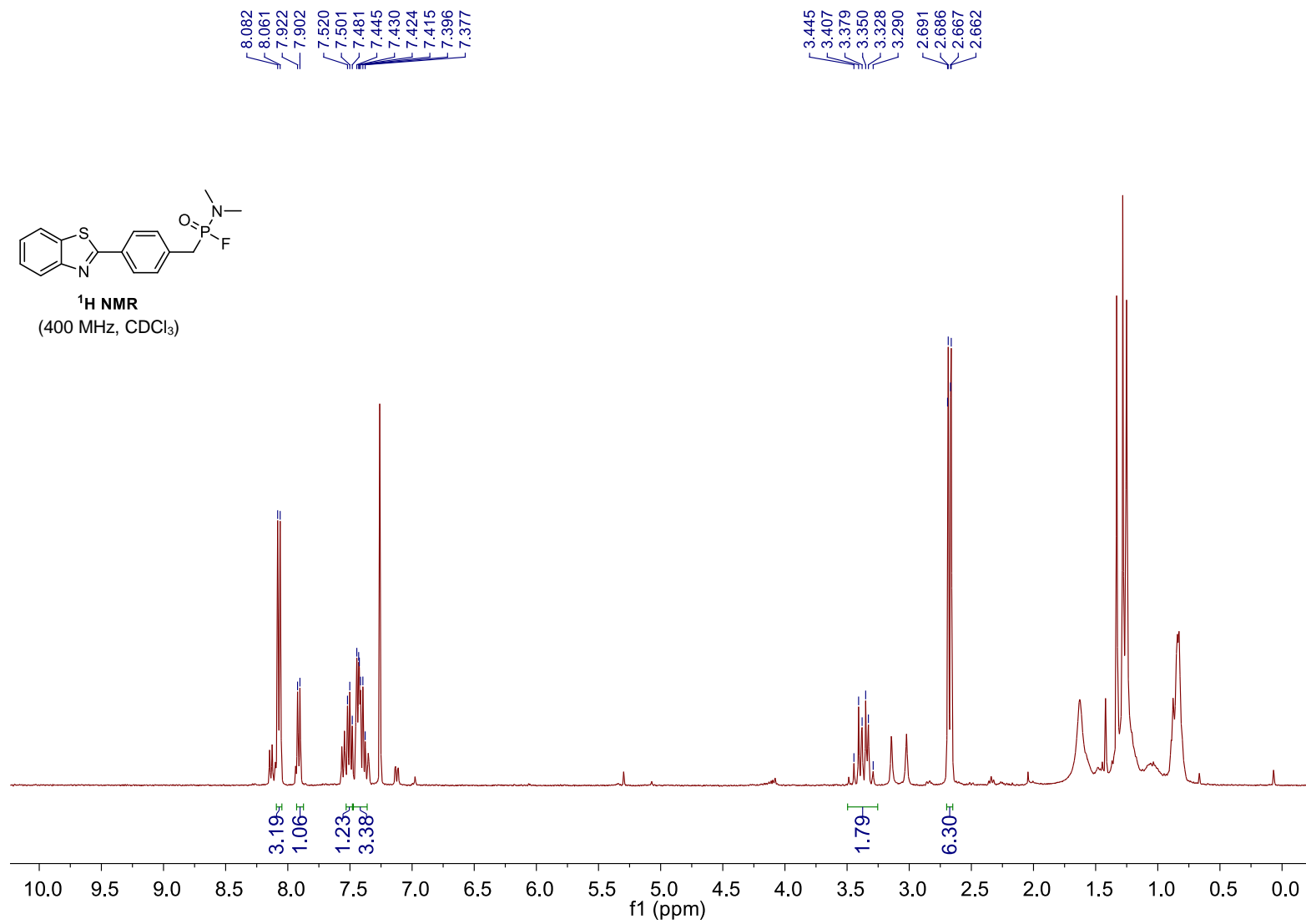


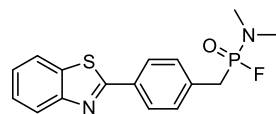
¹⁹F NMR
(376 MHz, CDCl₃)



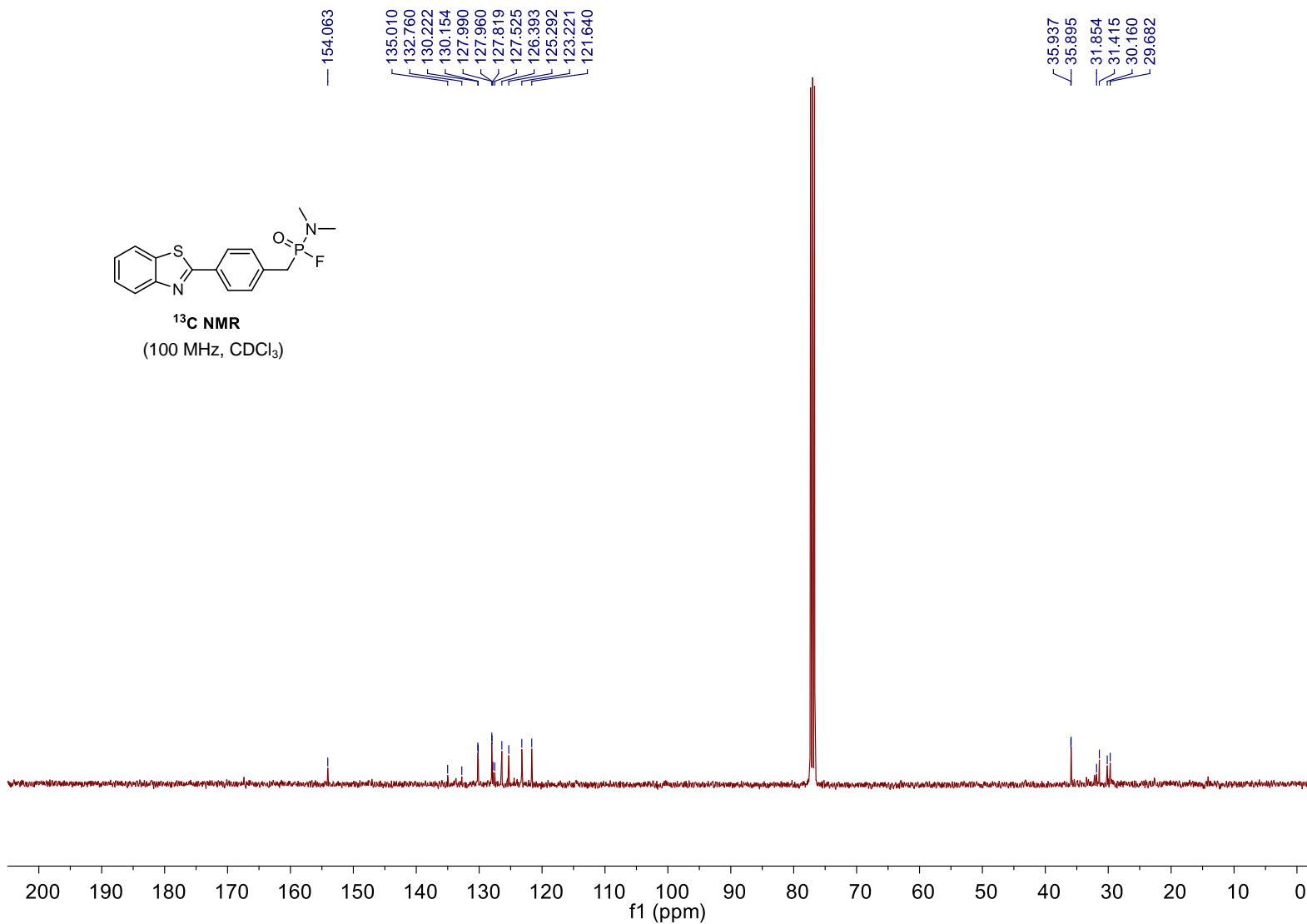


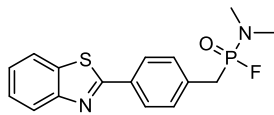
¹H NMR
(400 MHz, CDCl₃)





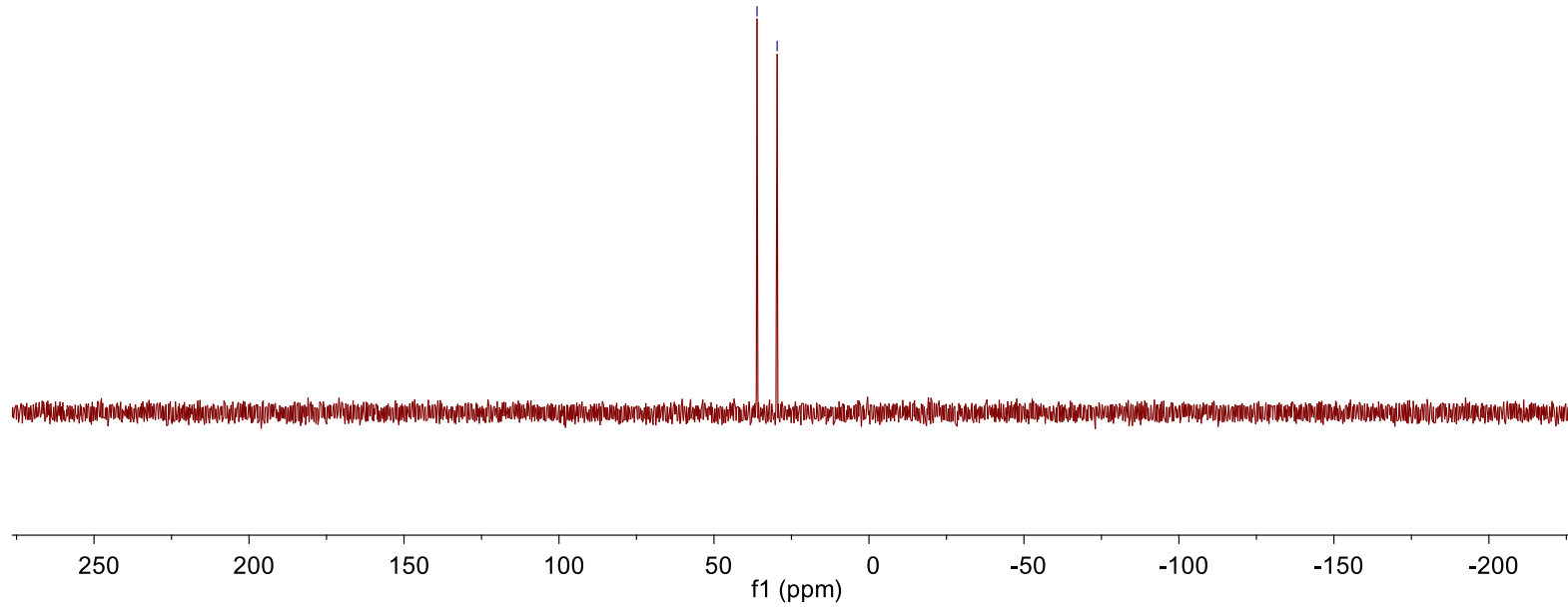
¹³C NMR
(100 MHz, CDCl₃)



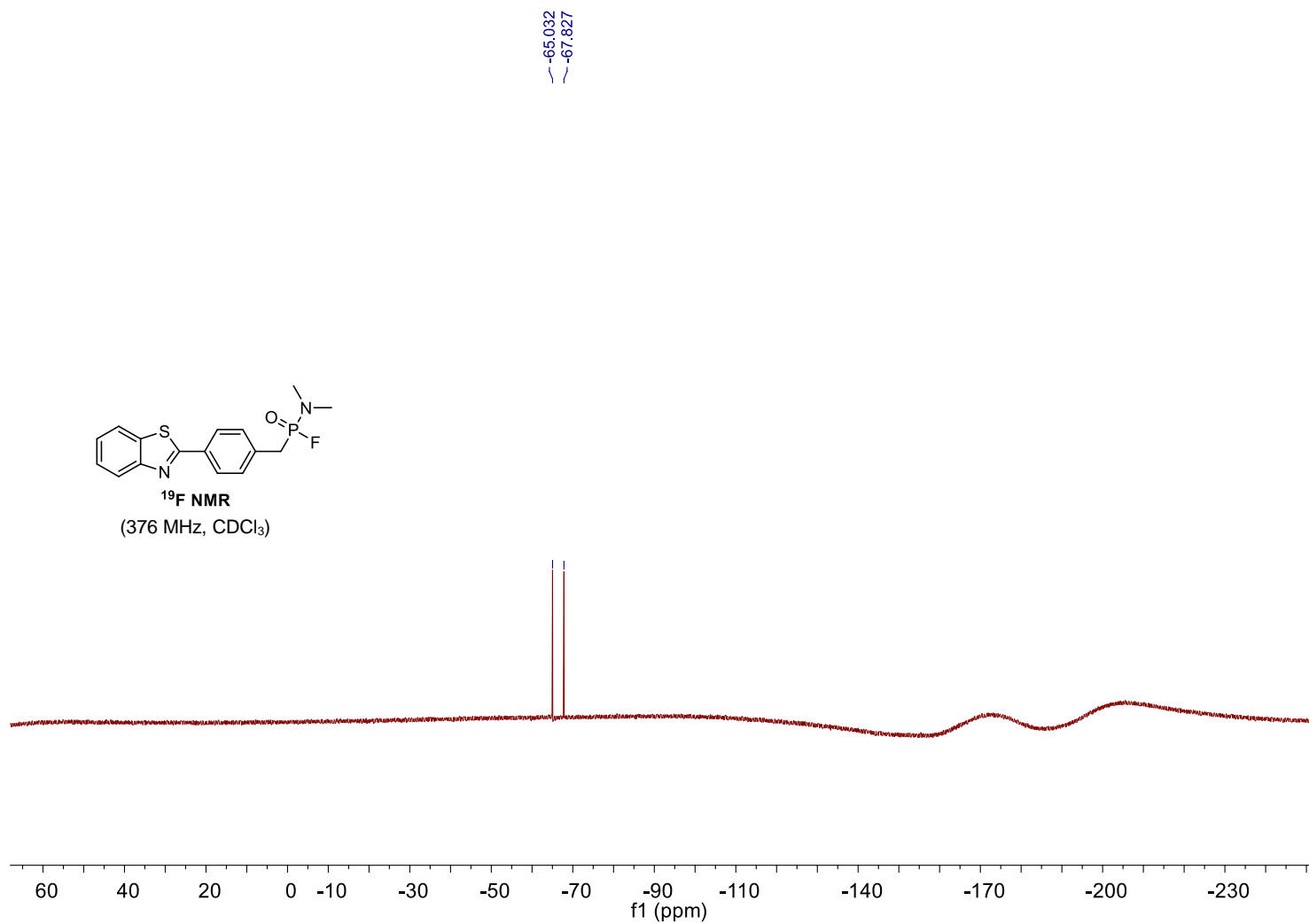
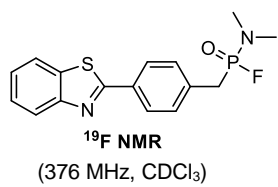


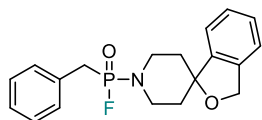
³¹P NMR
(162 MHz, CDCl₃)

— 36.114
— 29.619

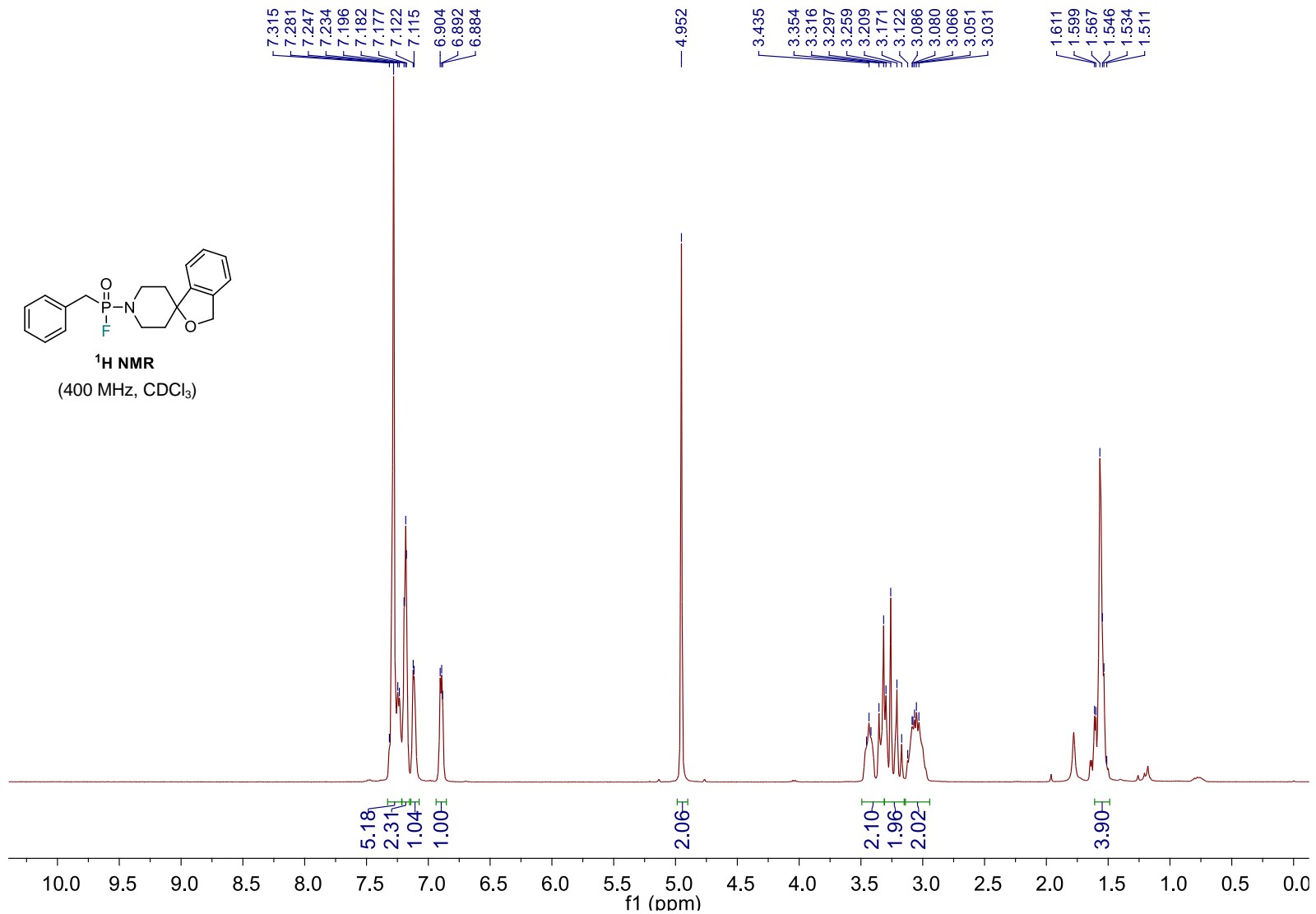


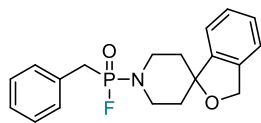
S387



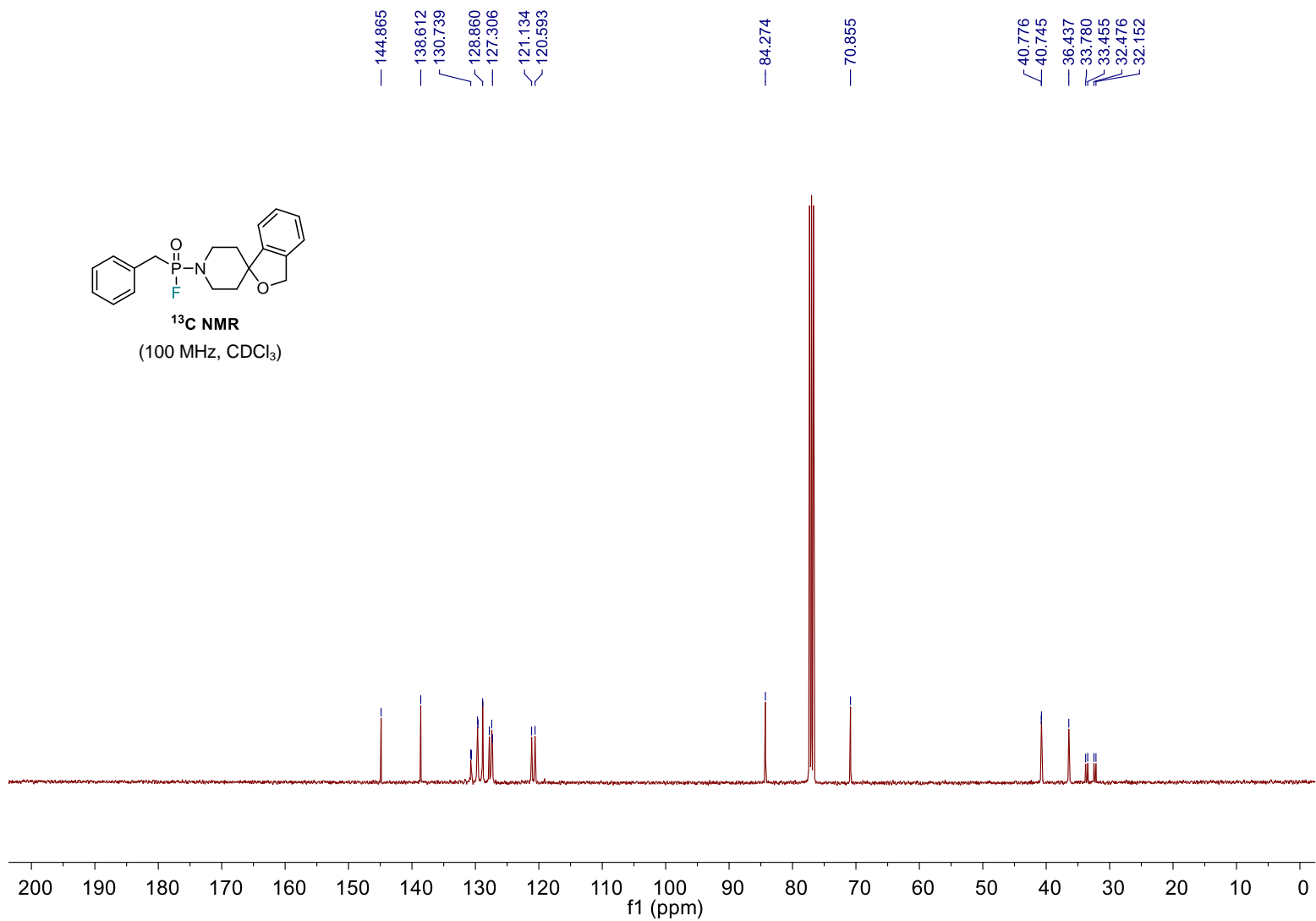


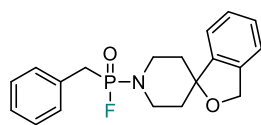
¹H NMR
(400 MHz, CDCl₃)



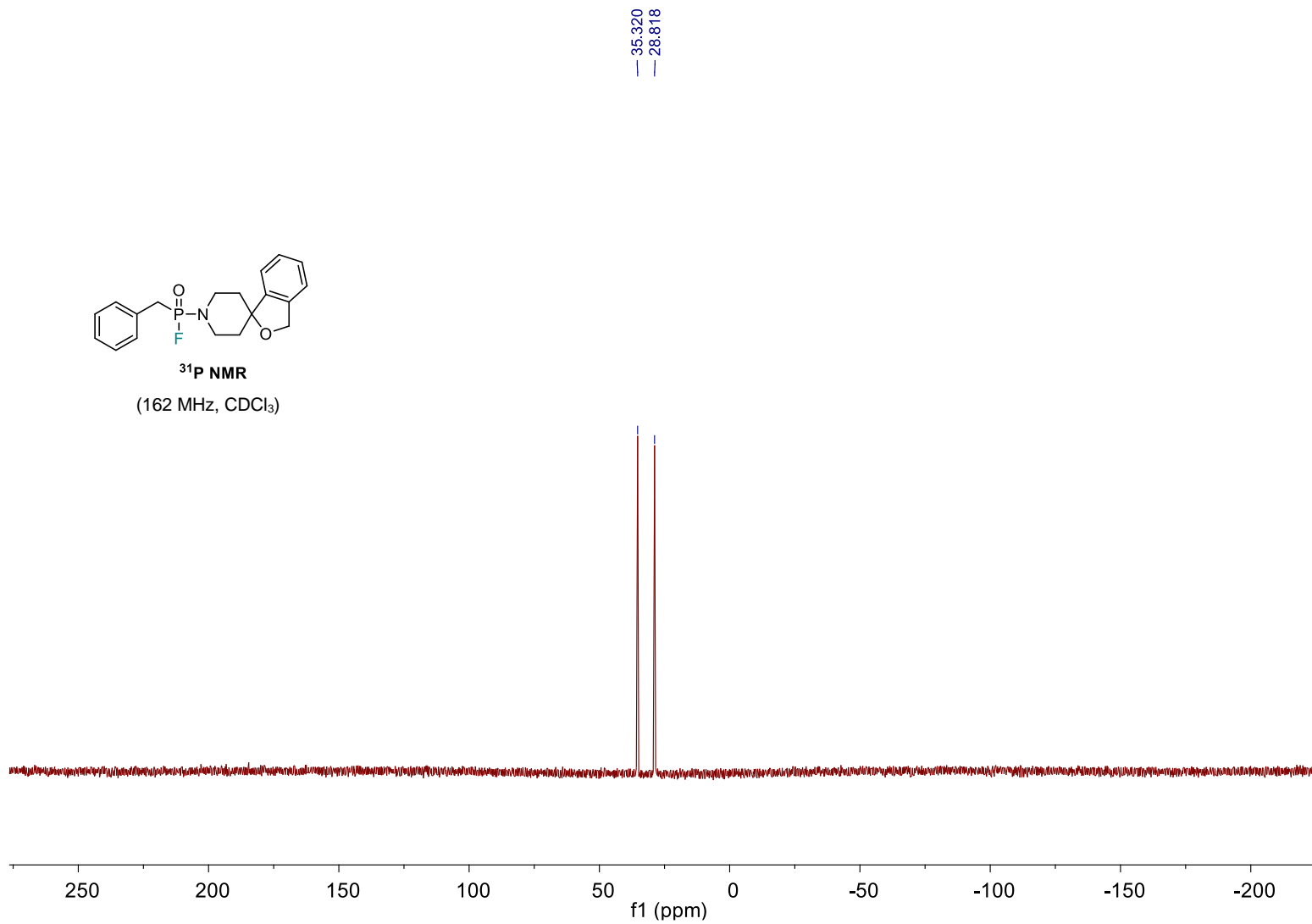


¹³C NMR
(100 MHz, CDCl₃)

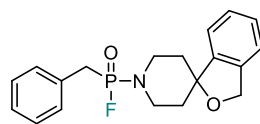




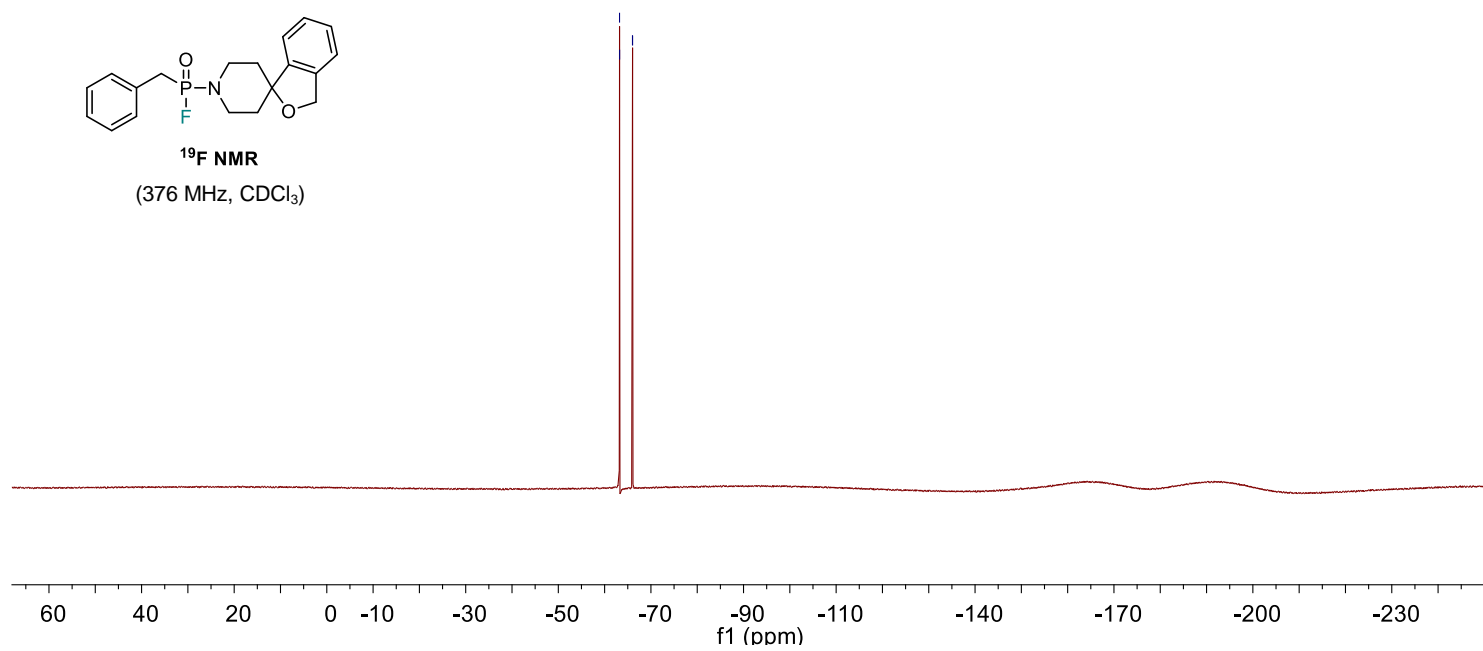
³¹P NMR
(162 MHz, CDCl₃)

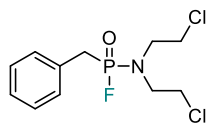


S391



¹⁹F NMR
(376 MHz, CDCl₃)

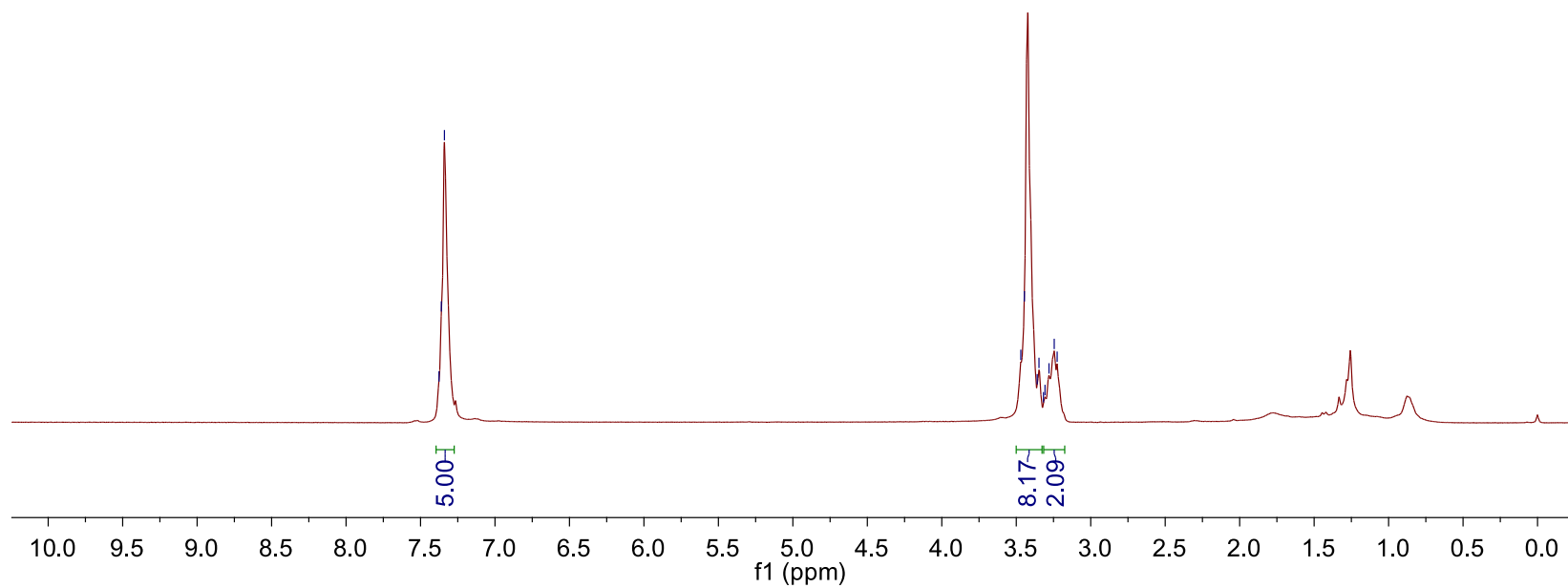




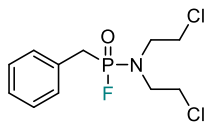
¹H NMR
(400 MHz, CDCl₃)

7.376
7.360
7.340

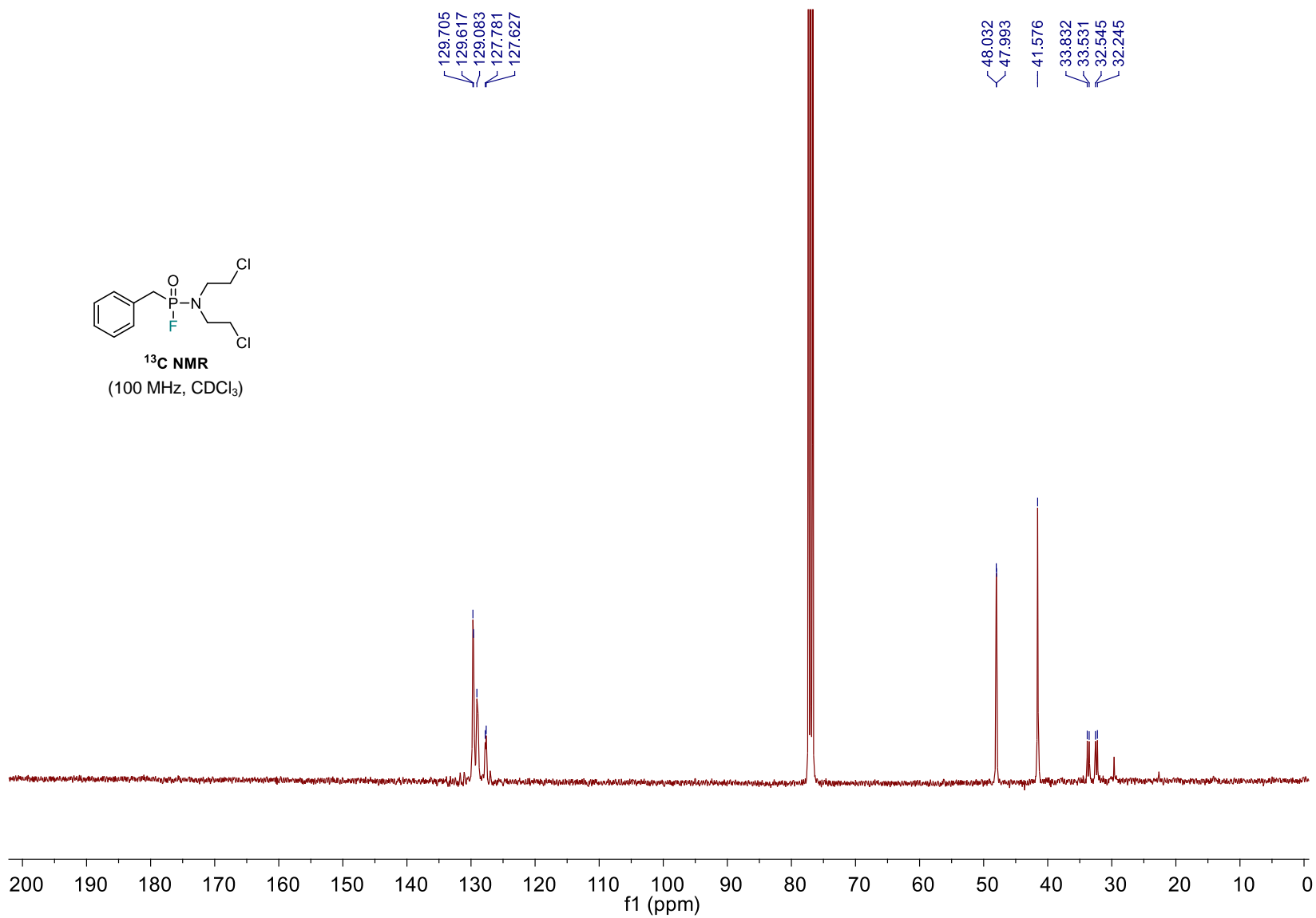
3.469
3.445
3.359
3.347
3.317
3.307
3.280
3.245
3.225



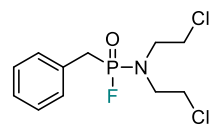
S393



¹³C NMR
(100 MHz, CDCl₃)

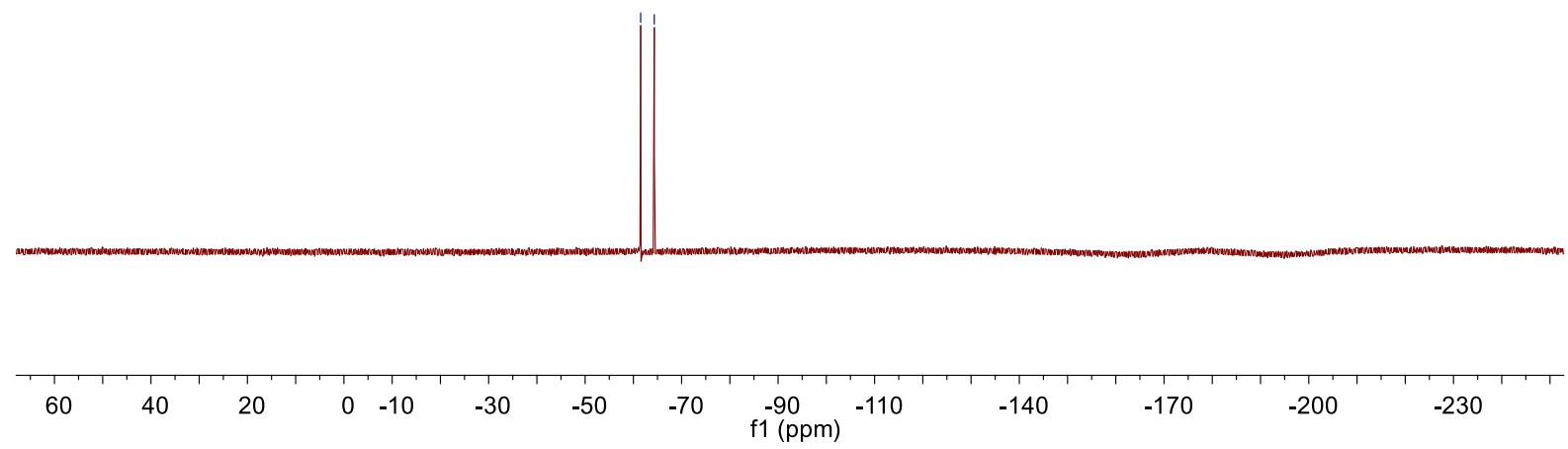


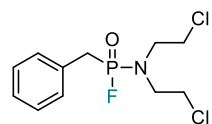
S394



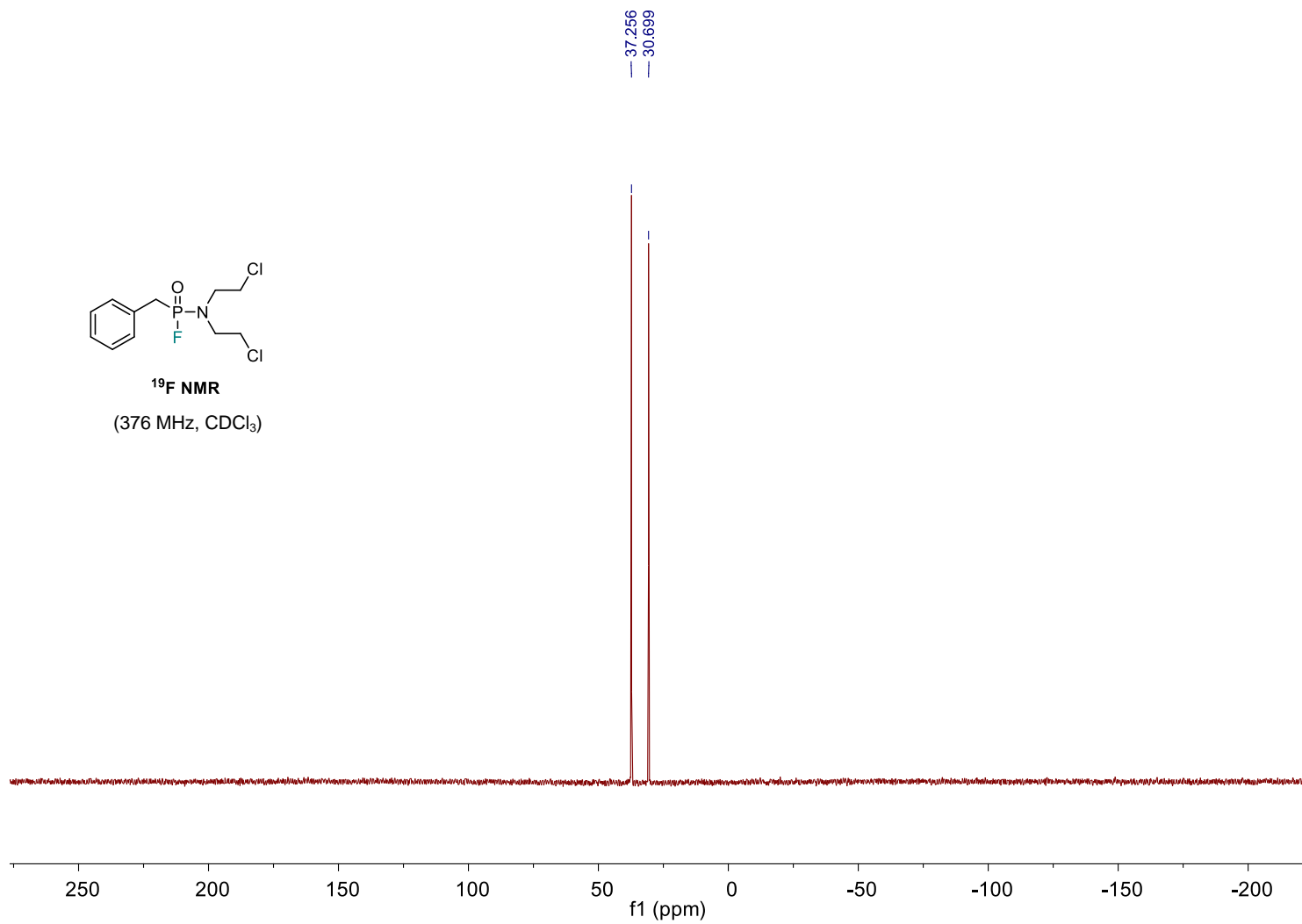
³¹P NMR
(162 MHz, CDCl₃)

-61.511
-64.334

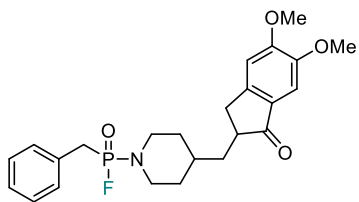




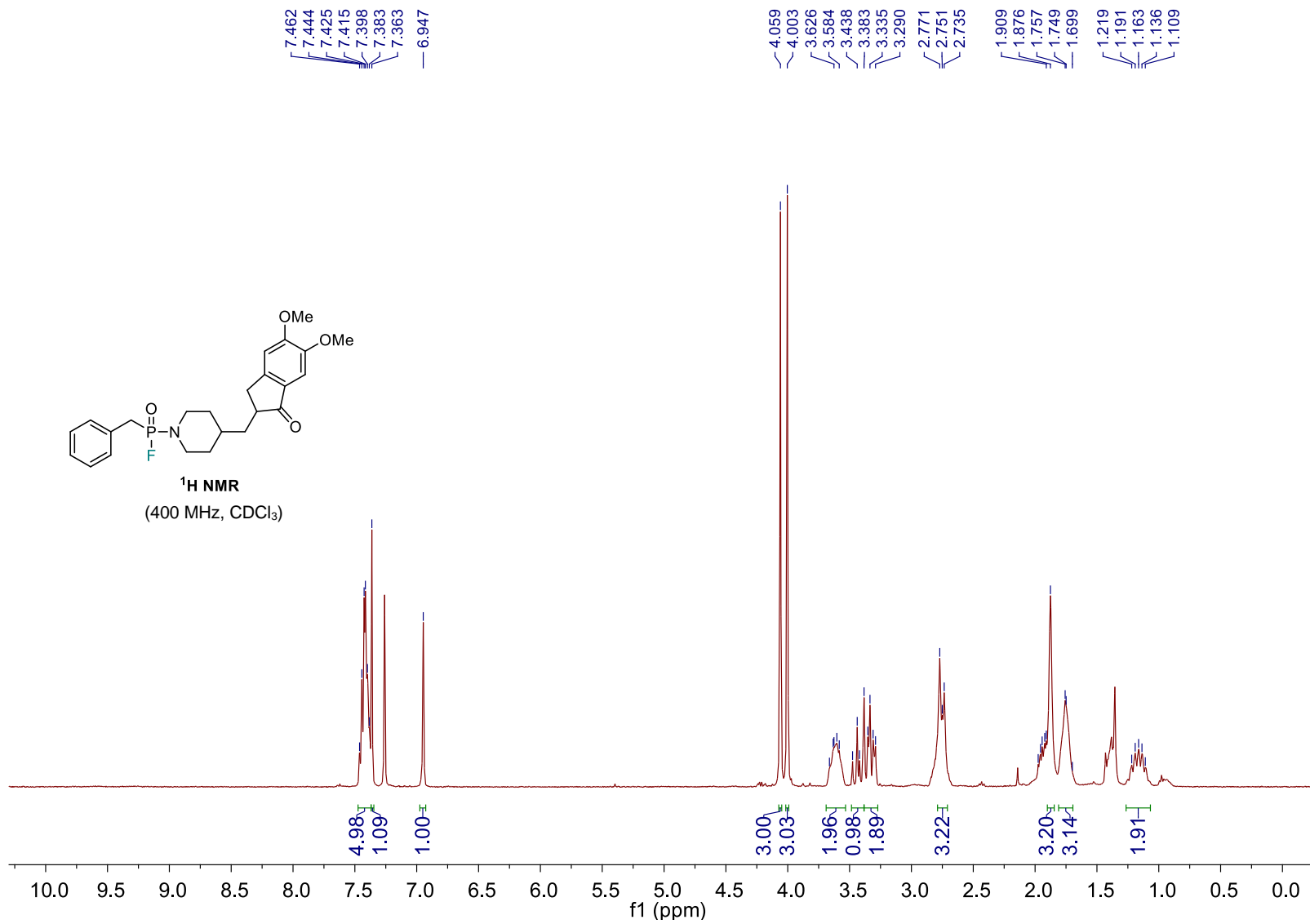
¹⁹F NMR
(376 MHz, CDCl₃)

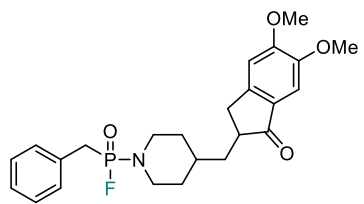


S396

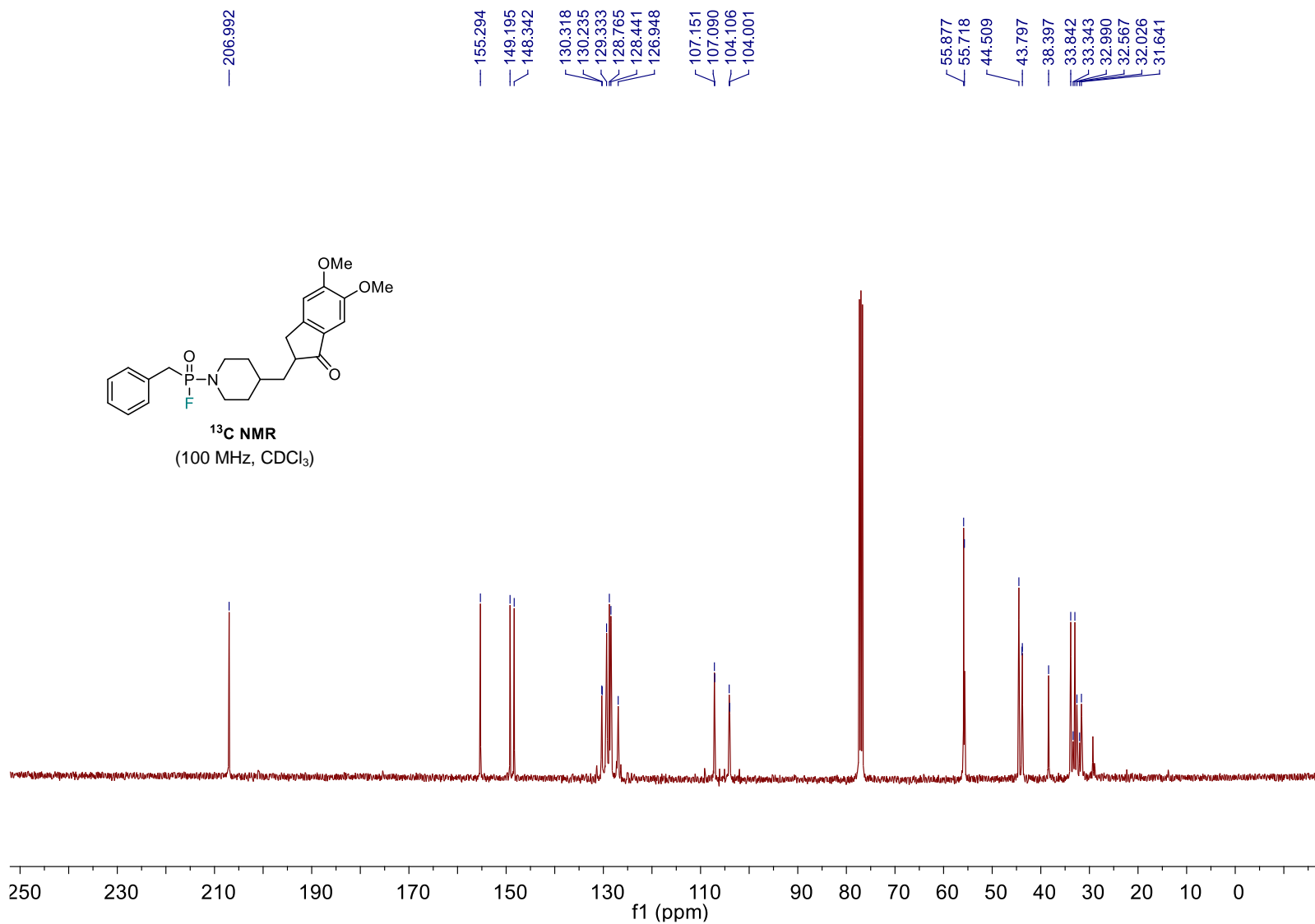


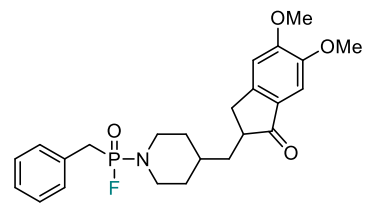
¹H NMR
(400 MHz, CDCl₃)





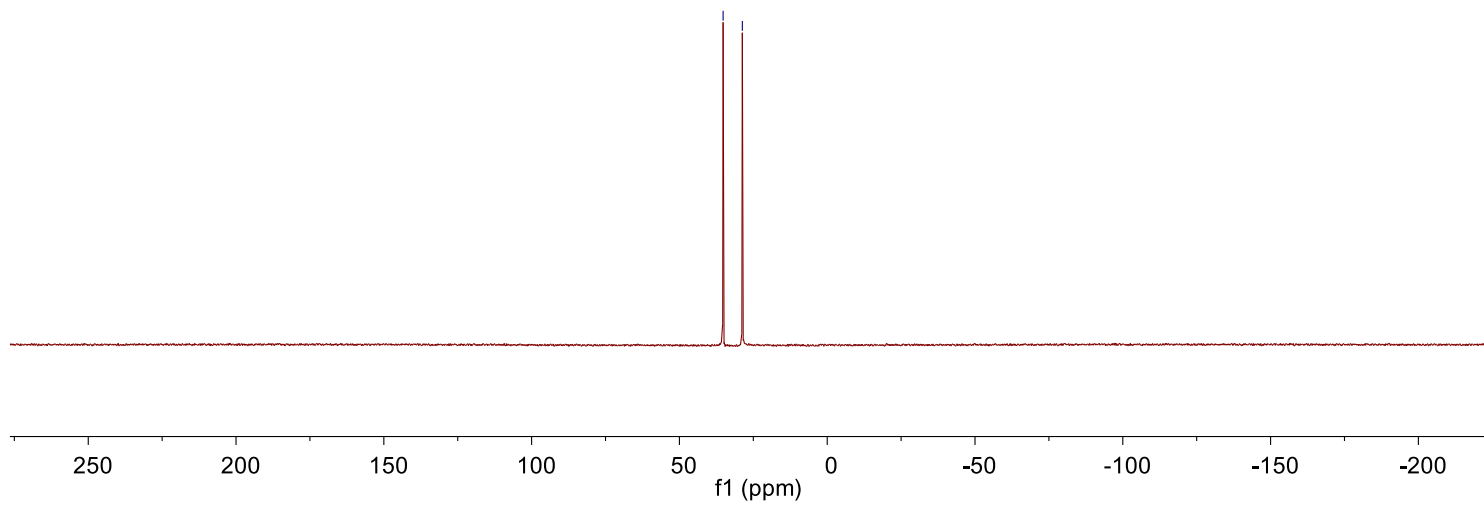
¹³C NMR
(100 MHz, CDCl₃)

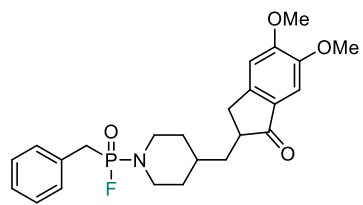




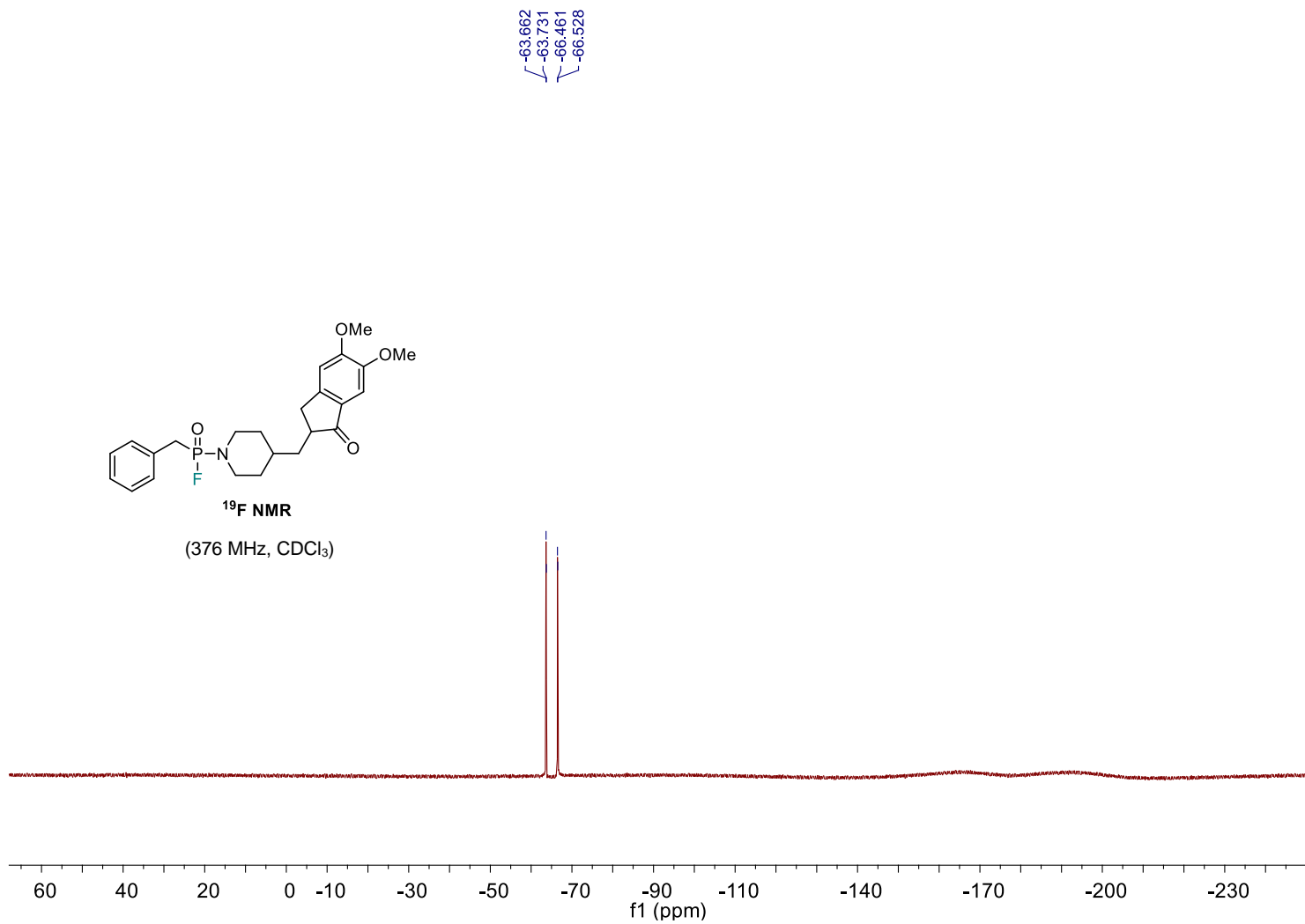
³¹P NMR
(162 MHz, CDCl₃)

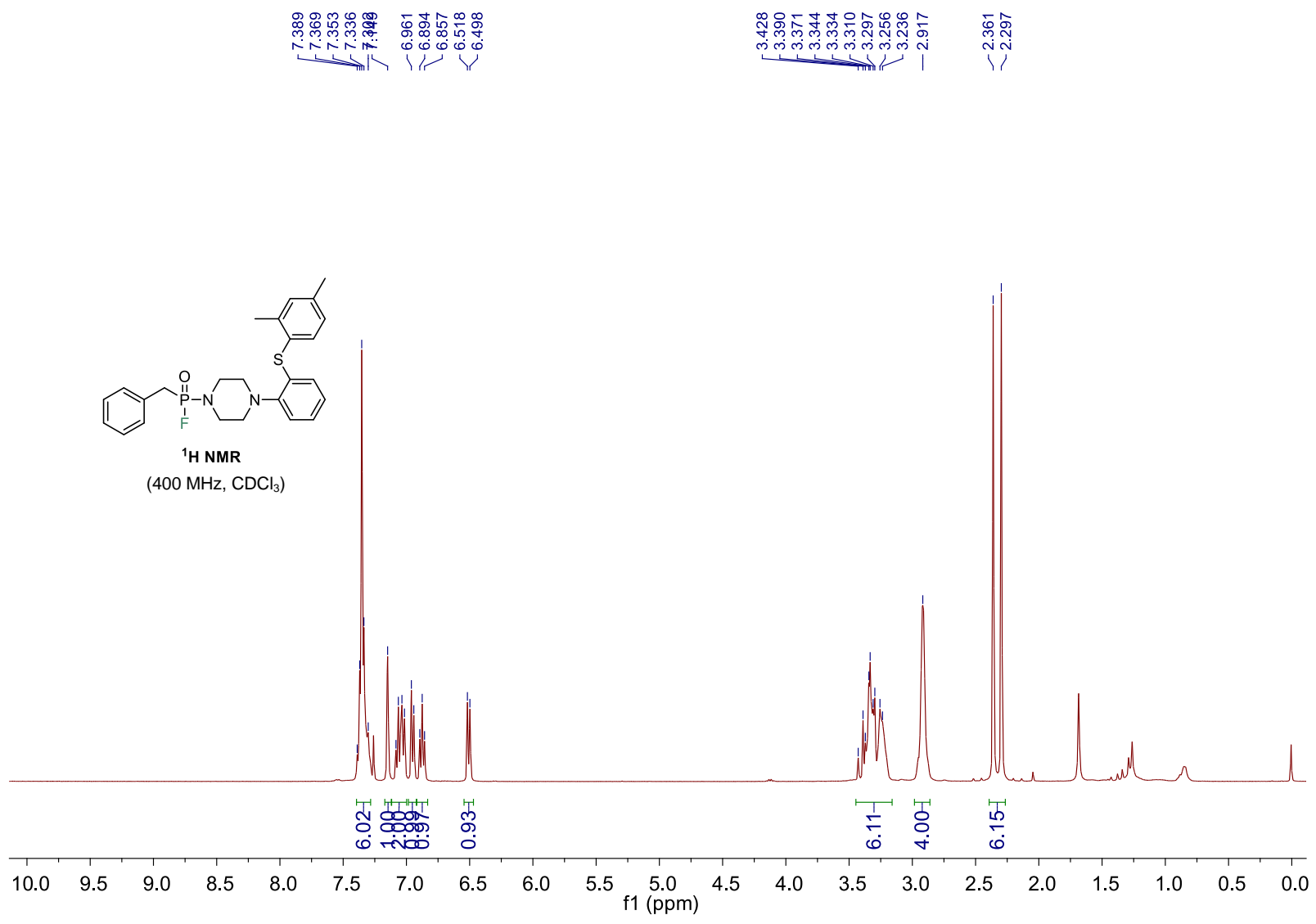
— 35.219
— 28.717

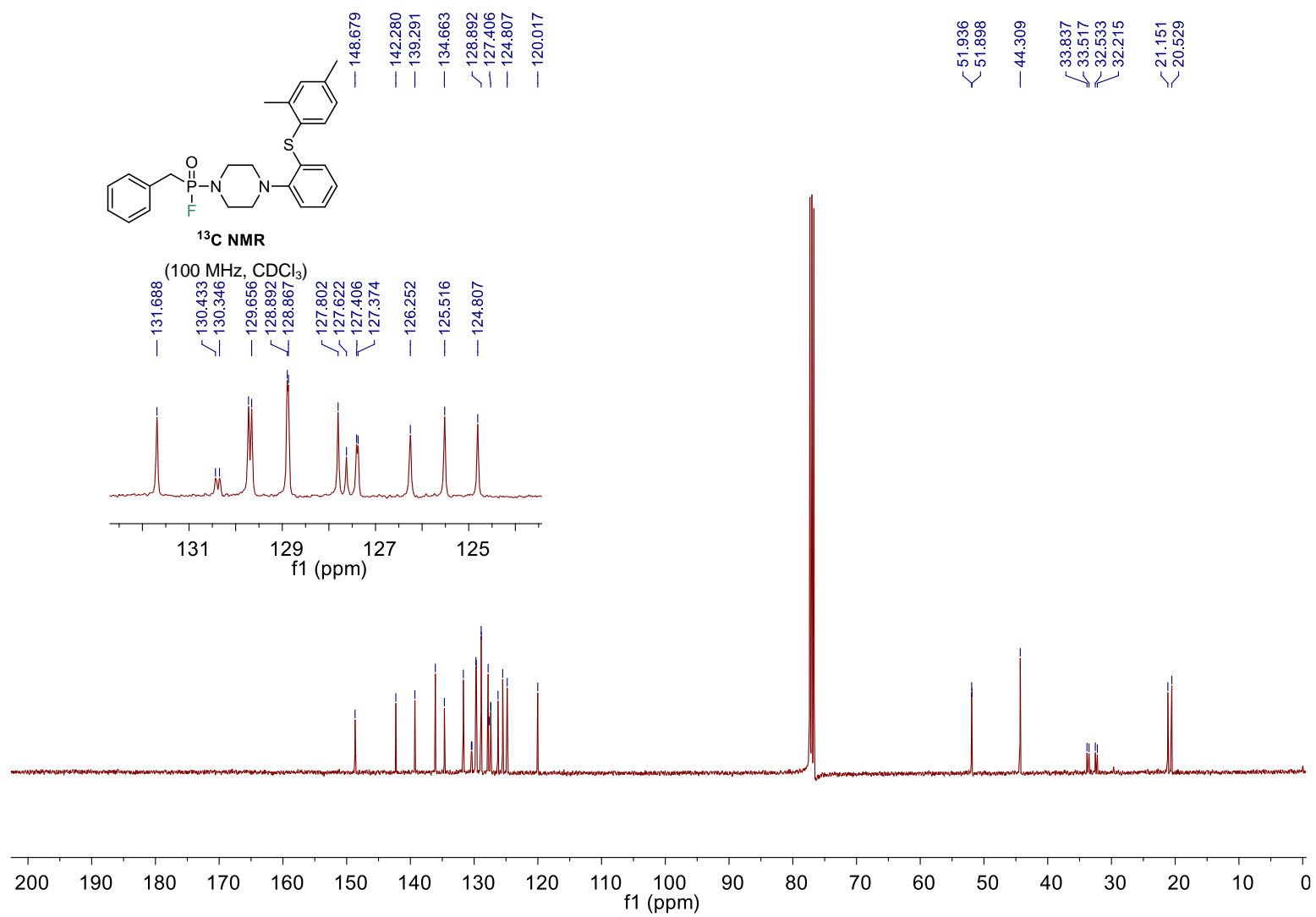


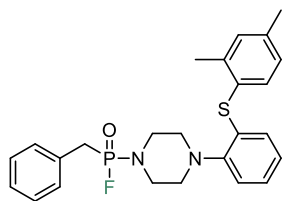


¹⁹F NMR
(376 MHz, CDCl₃)

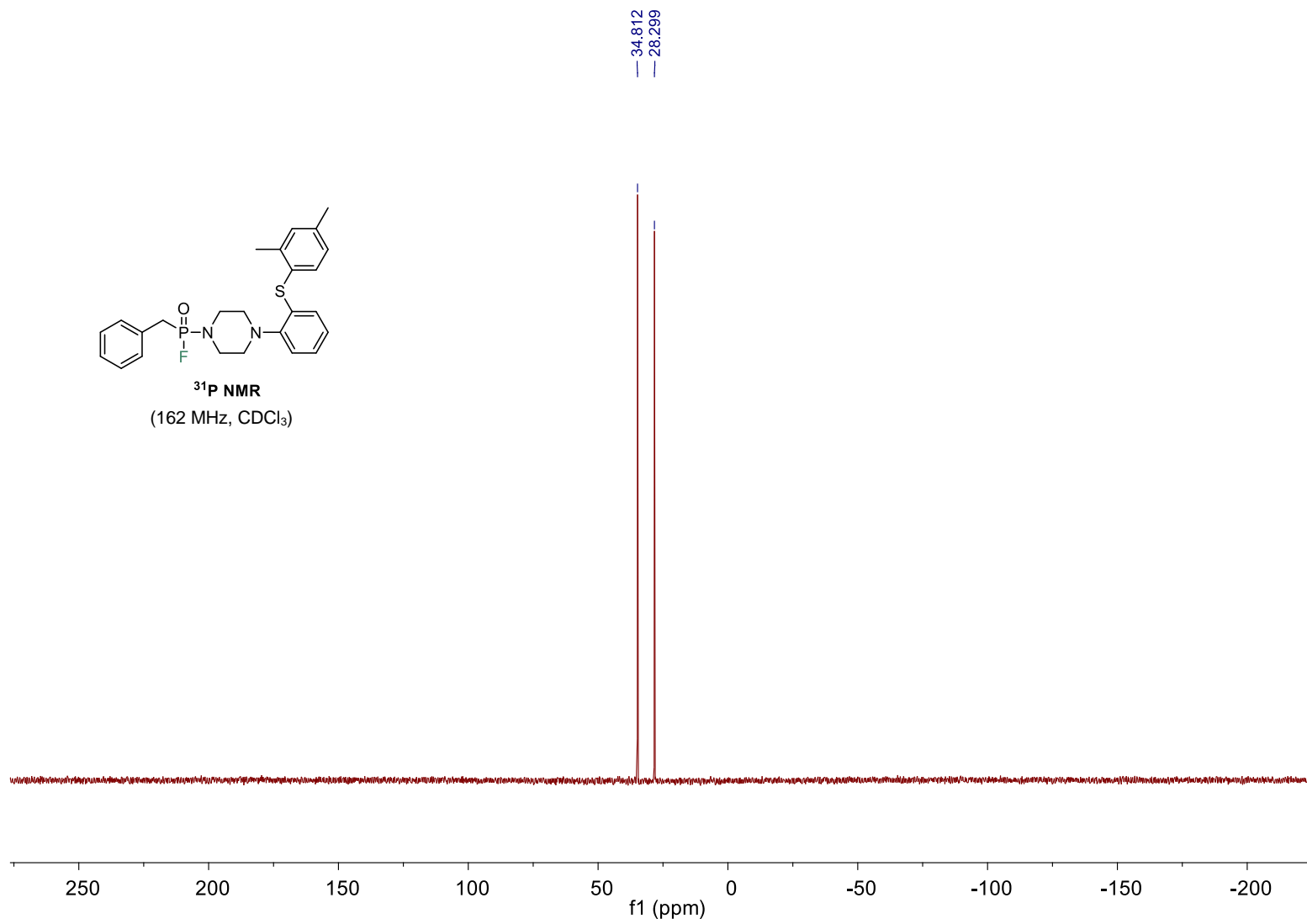




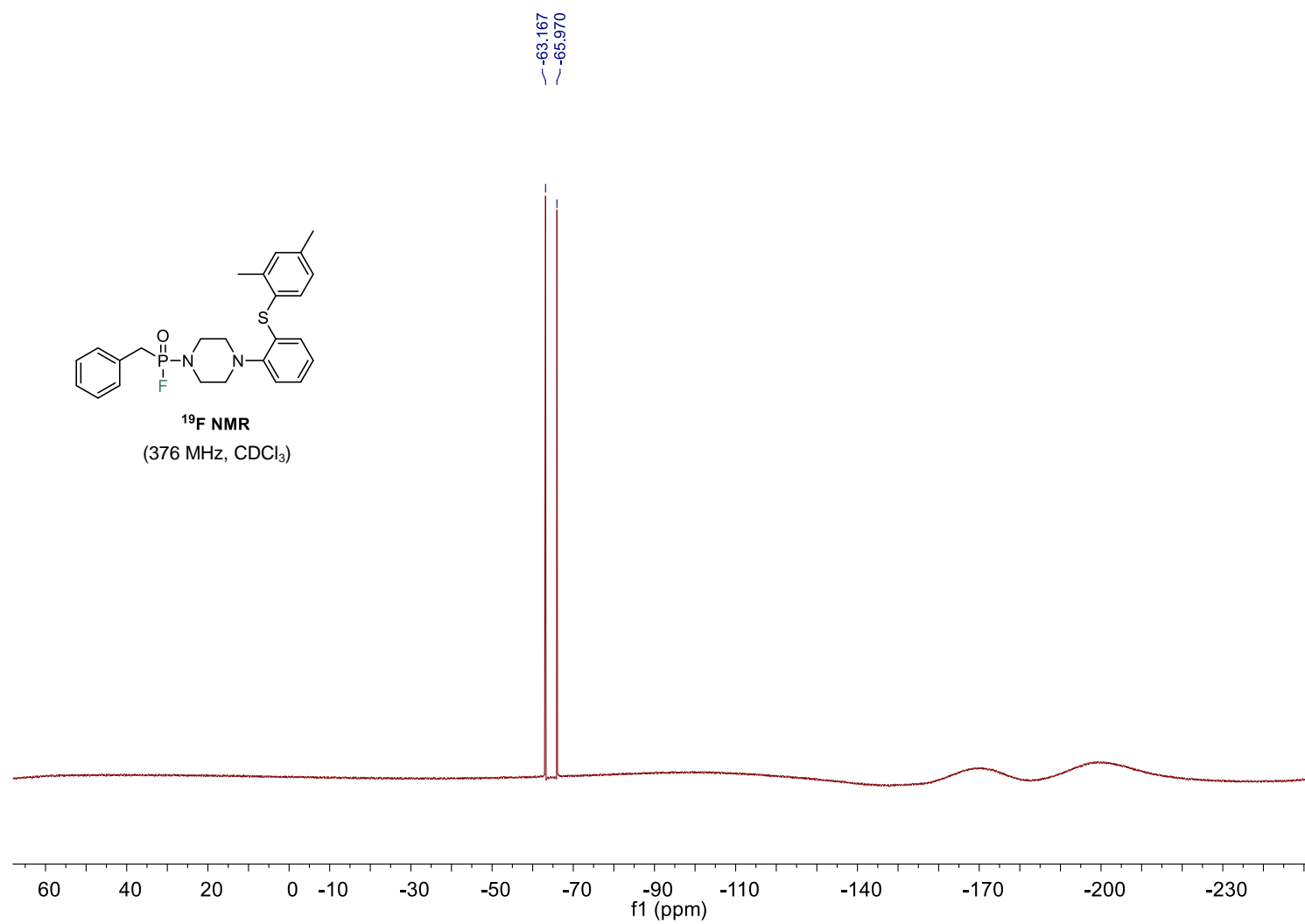


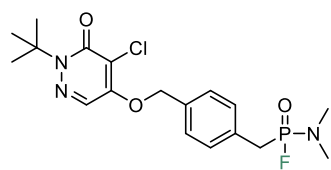


³¹P NMR
(162 MHz, CDCl₃)

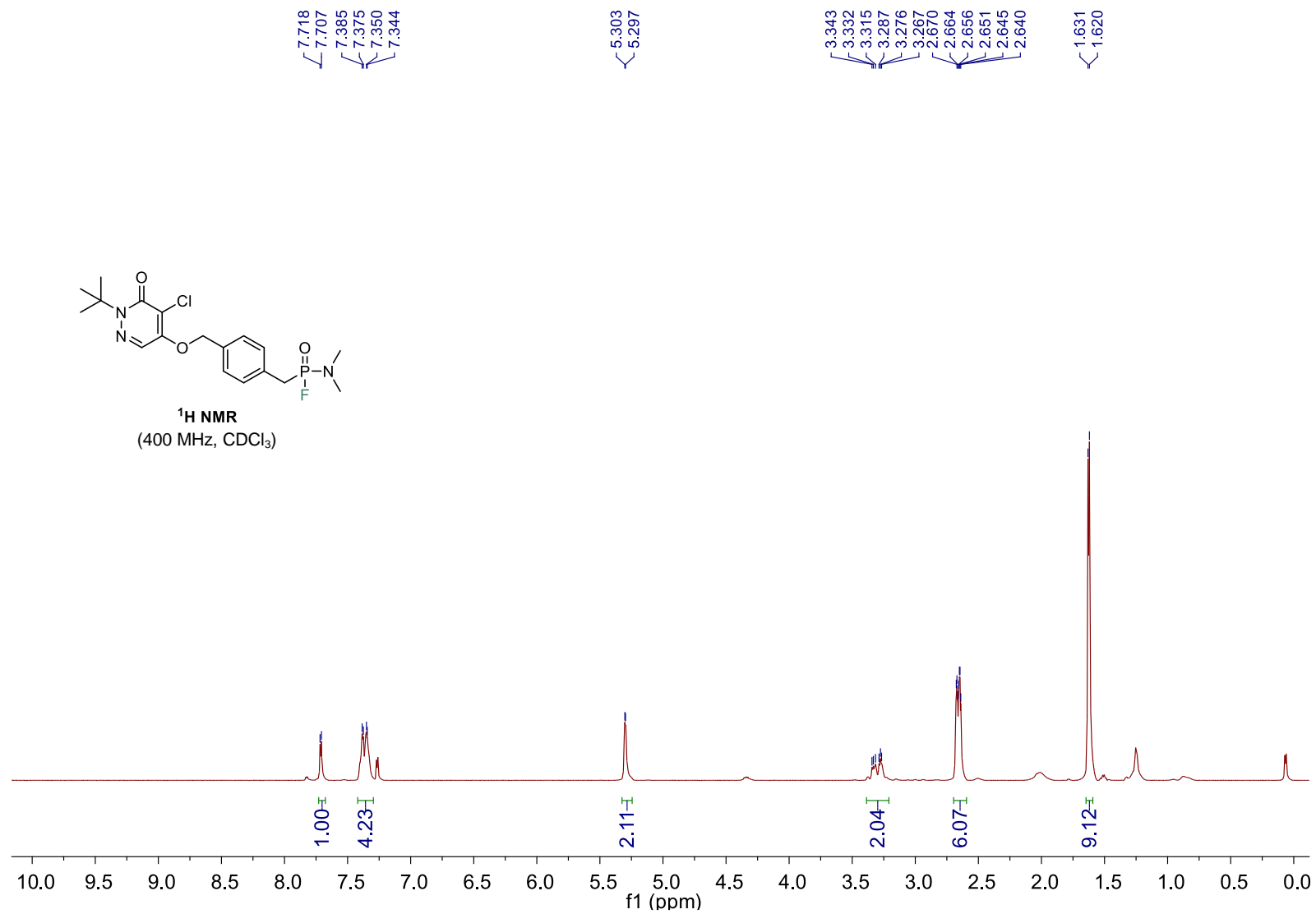


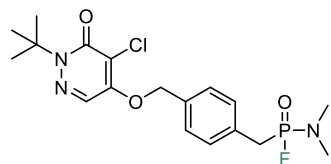
S403



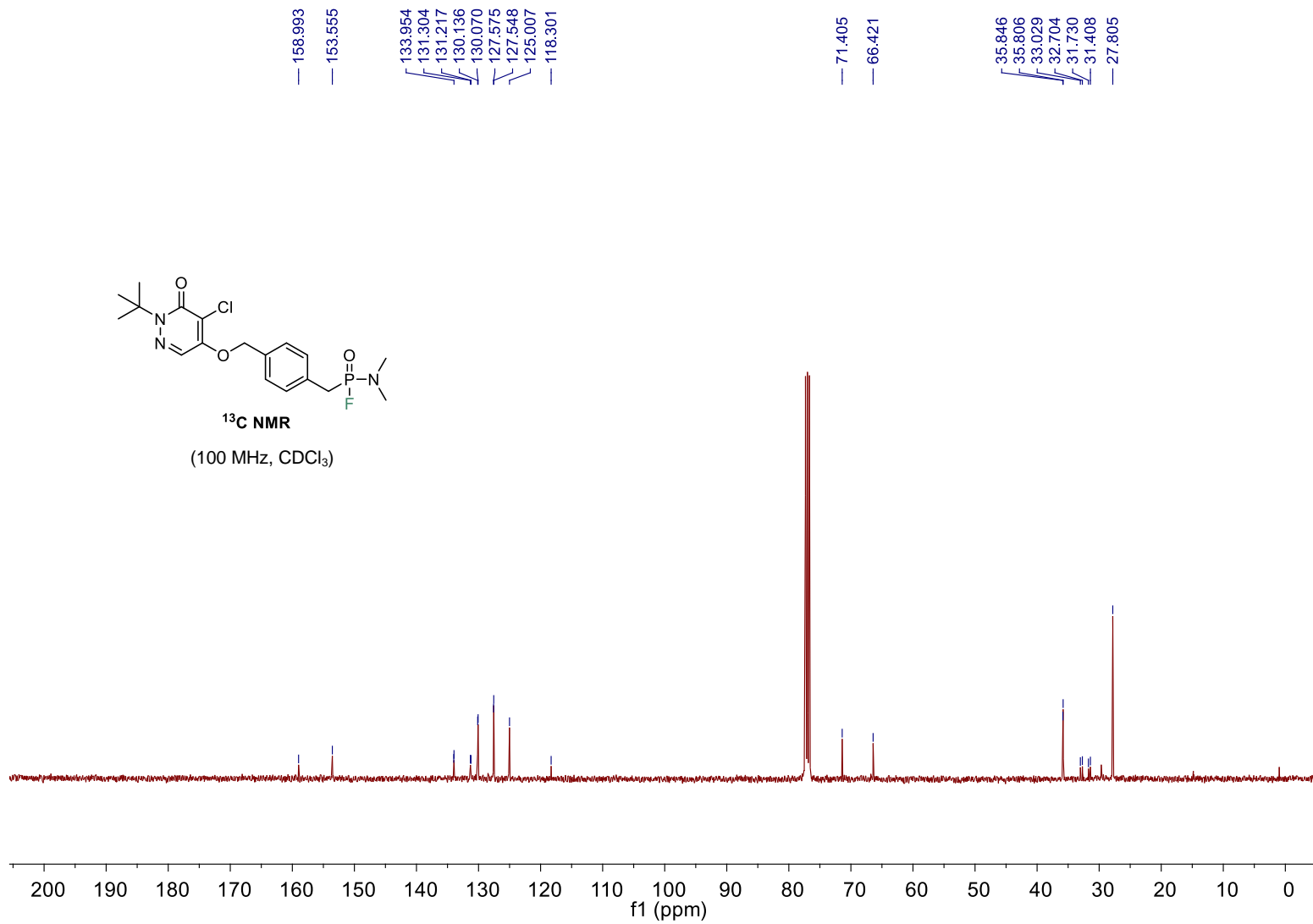


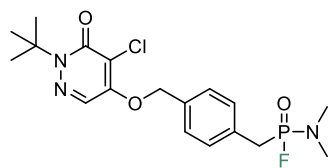
¹H NMR
(400 MHz, CDCl₃)



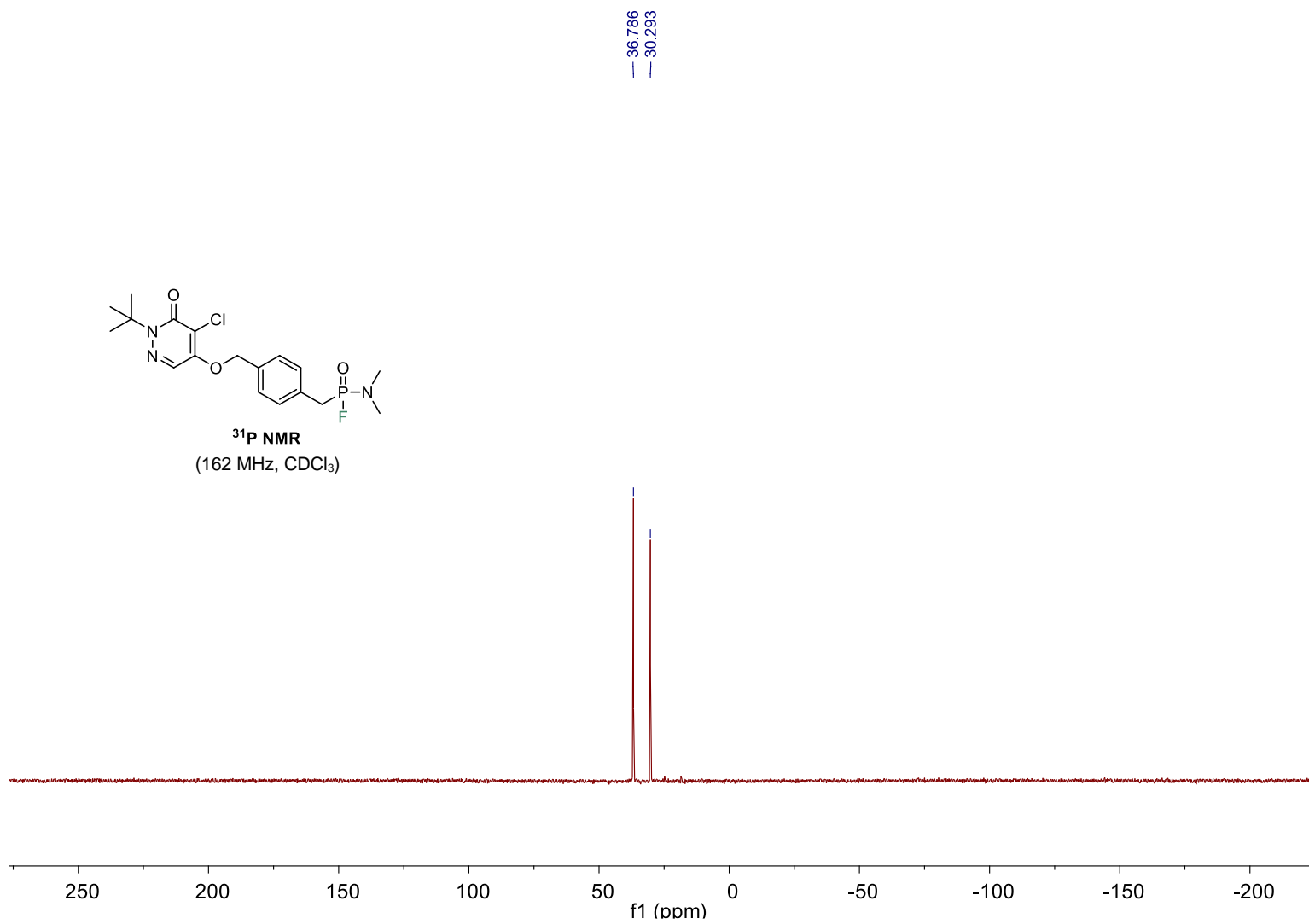


¹³C NMR
(100 MHz, CDCl₃)

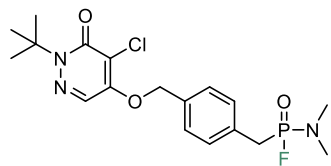




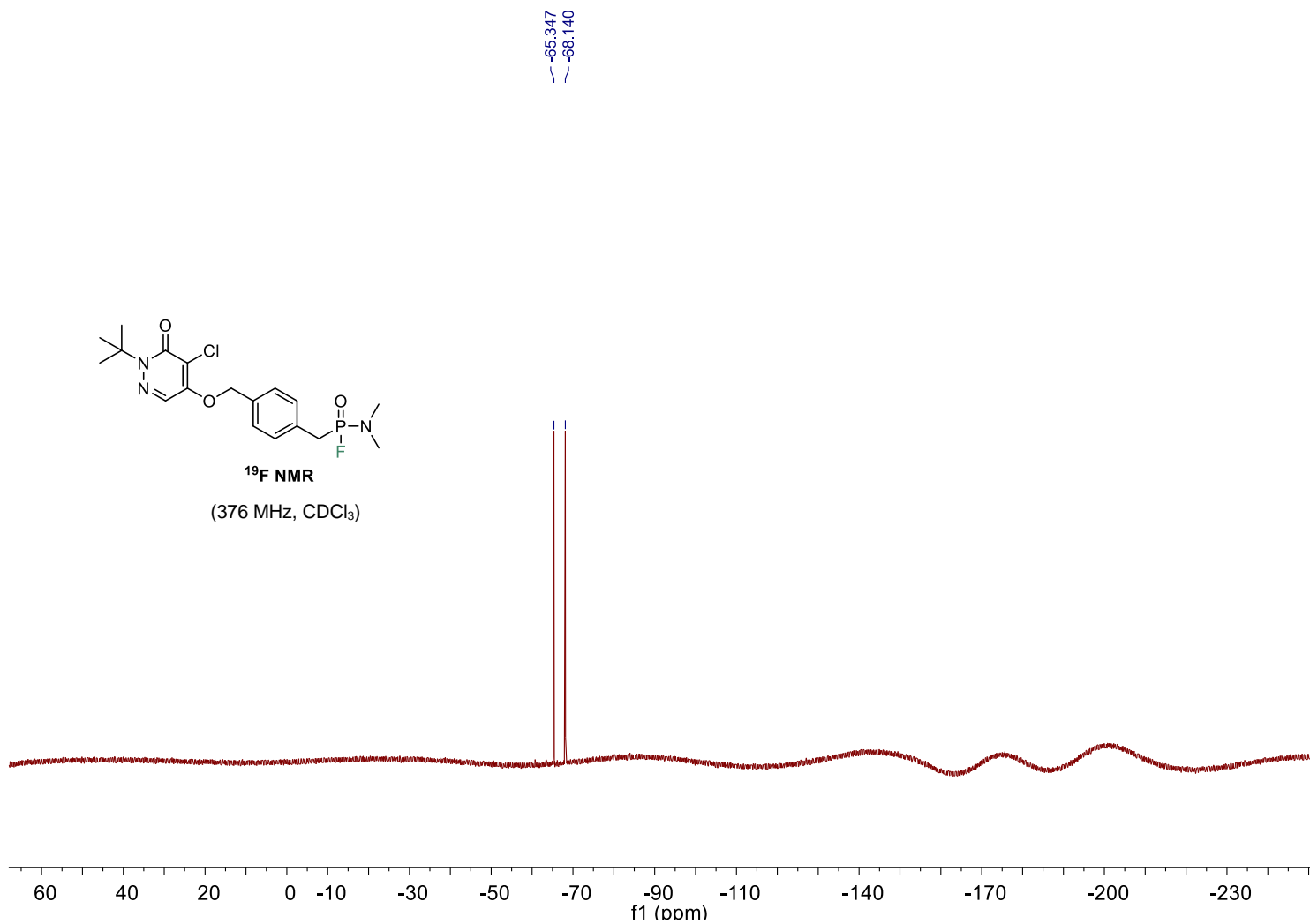
³¹P NMR
(162 MHz, CDCl₃)

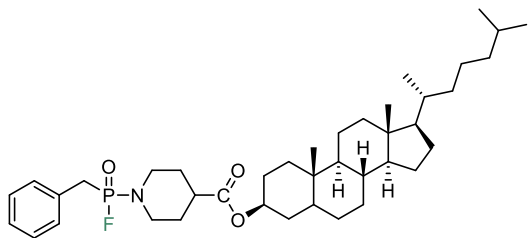


S407

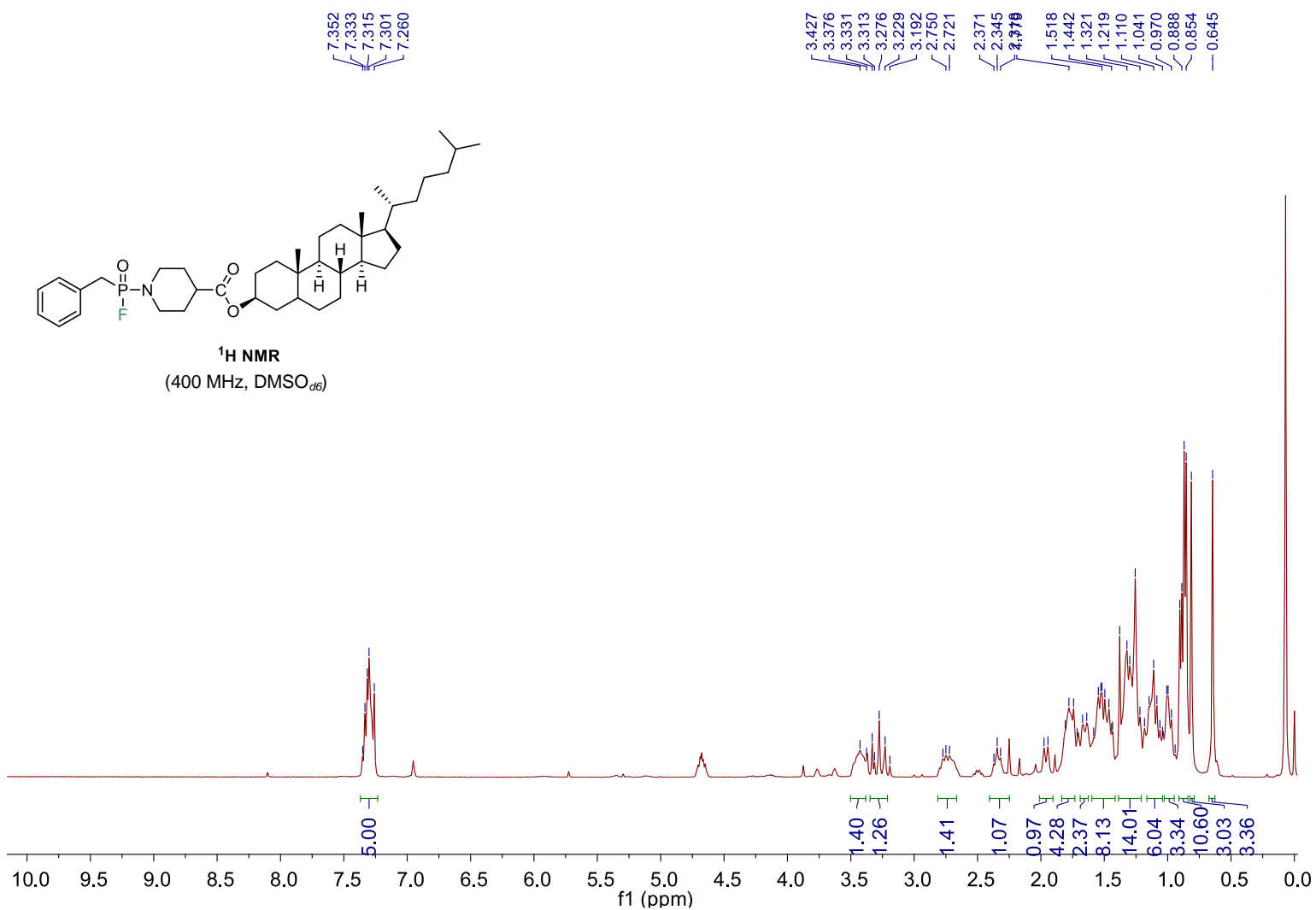


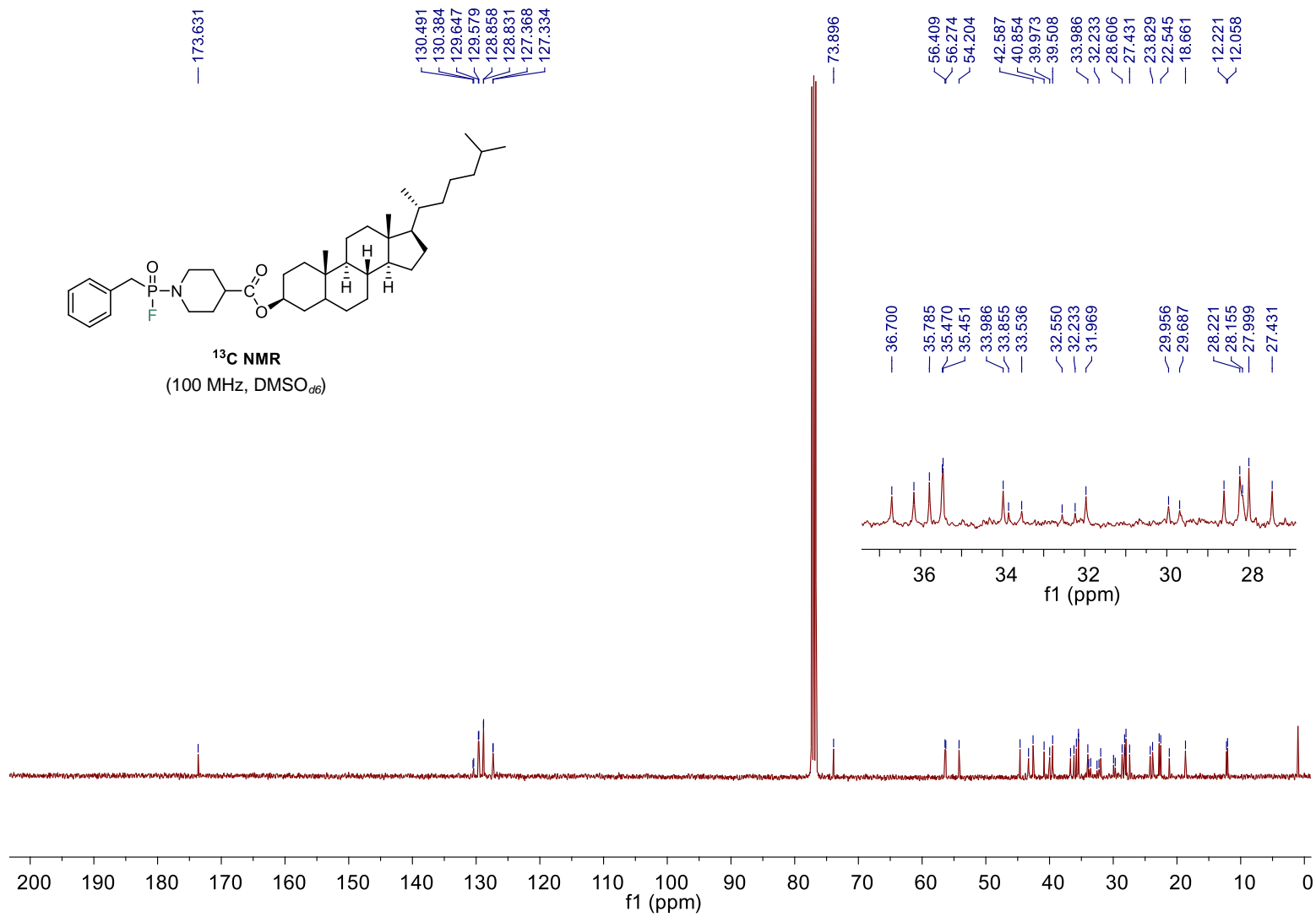
¹⁹F NMR
(376 MHz, CDCl₃)

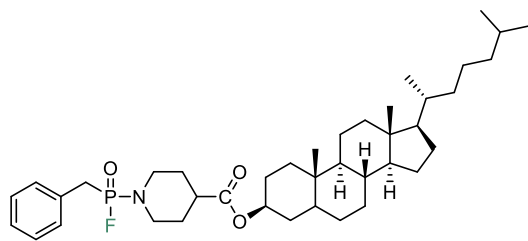




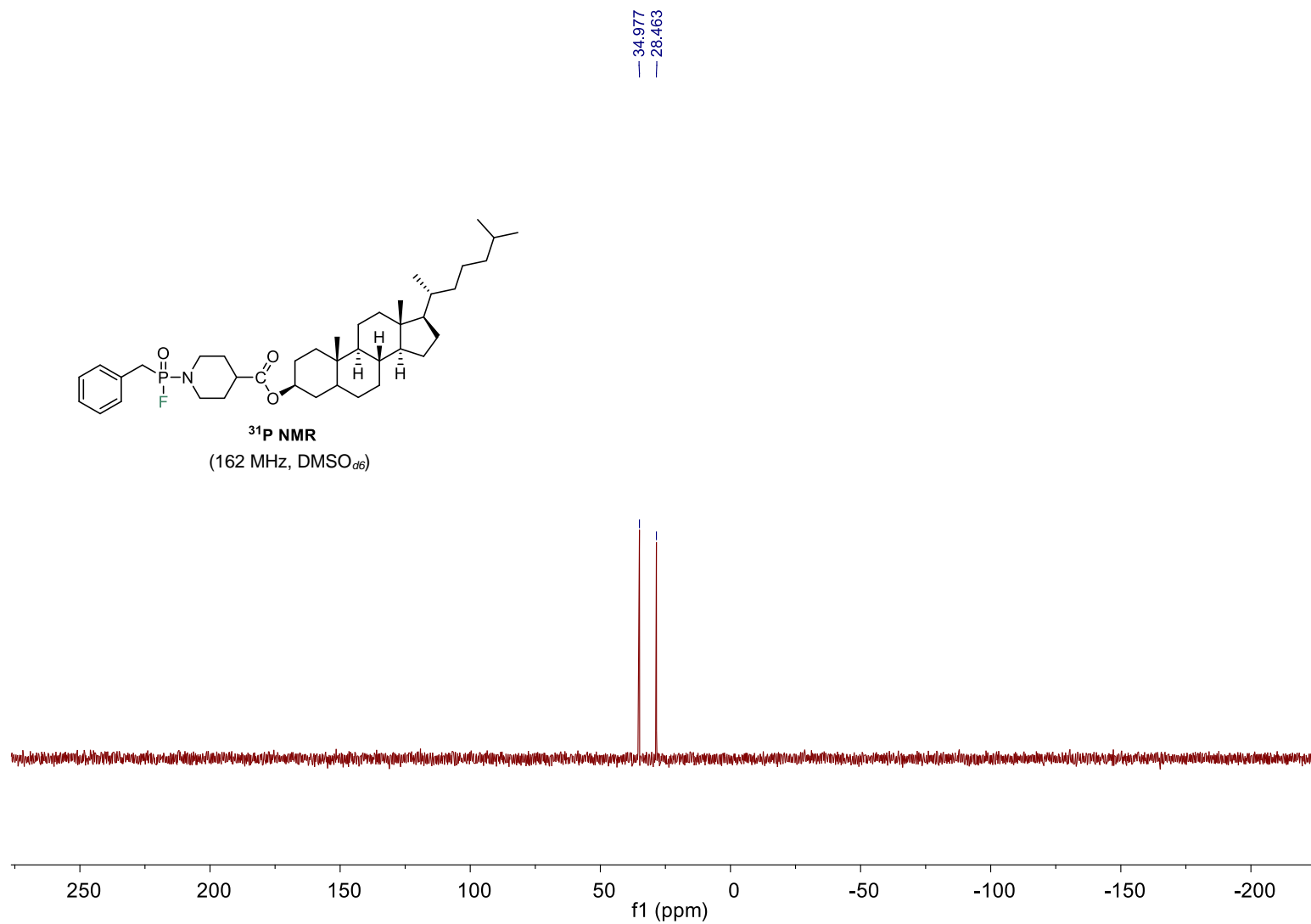
¹H NMR
(400 MHz, DMSO-*d*₆)

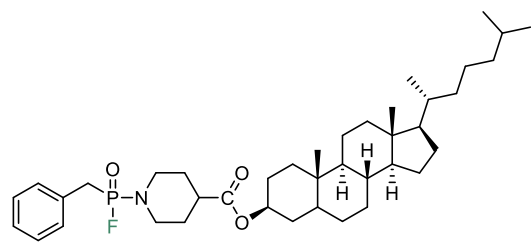




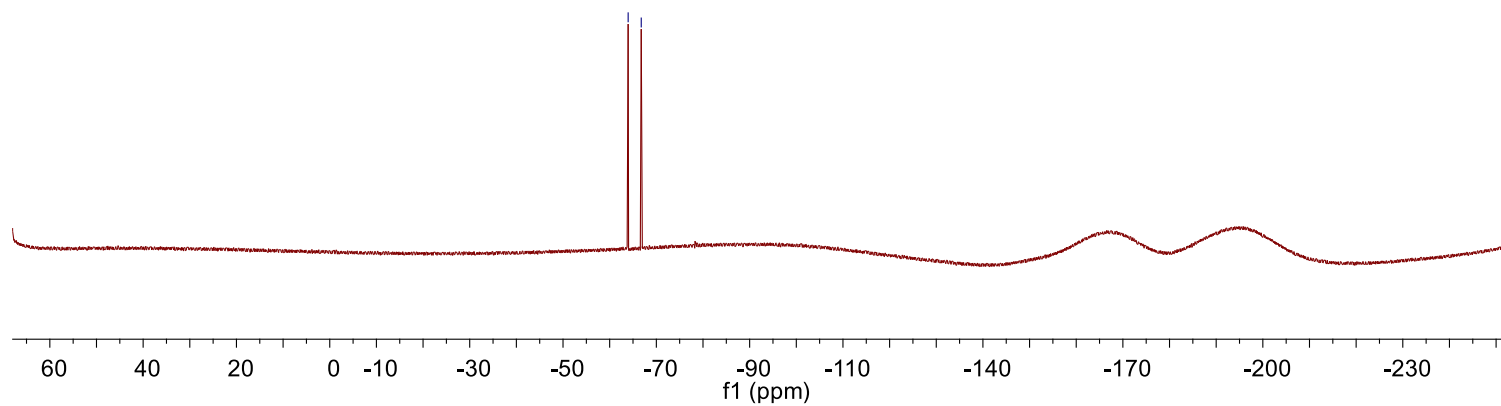


³¹P NMR
(162 MHz, DMSO_{d6})





¹⁹F NMR
(376 MHz, DMSO_{d6})



4 Supplementary references

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