1 Supplementary Figure Legends

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3 Supplementary Figure 1. NSP14 mediates NF-KB activation. (a) HEK293 cells were co-4 transfected with 0.2 μg of the indicated FLAG-tagged SARS-CoV-2 gene along with NF-κB 5 reporter and pRL-SV40. After 48 hours, cells were harvested, and the relative reporter activity 6 was determined by calculating the ratio of firefly luciferase to *Renilla* luciferase. The *p*-value was 7 calculated by one-way ANOVA followed by Dunnett's multiple comparisons test. Lysates were 8 blotted with anti-FLAG and anti- α -tubulin antibodies. (b) HEK293 cells were co-transfected with 9 0.05 μg, 0.1 μg, and 0.15 μg of FLAG-tagged SARS-CoV-2 NSP14 along with NF-κB reporter and 10 pRL-SV40. After 48 hours, cells were harvested, and the relative reporter activity was determined 11 by calculating the ratio of firefly luciferase to Renilla luciferase. Lysates were blotted with anti-12 FLAG and anti- α -tubulin antibodies. (c) 0.1 µg of FLAG-tagged SARS-CoV-2 NSP14 and 0.1 µg 13 of FLAG-tagged SARS-CoV-2 NSP10 were co-transfected with the NF-κB reporter and pRL-SV40 14 into HEK293 cells. Western blotting demonstrates the expression levels of NSP14-FLAG and 15 NSP10-FLAG. The position of NSP14 and NSP10 is denoted. (d) FLAG-tagged SARS-CoV-2 16 NSP14 or the indicated zinc finger mutant was co-transfected with the NF-kB reporter and pRL-SV40 into HEK293 cells. After 48 hours, cells were harvested, and the relative reporter activity 17 18 was determined by calculating the ratio of firefly luciferase to Renilla luciferase. Lysates were 19 blotted with anti-FLAG and anti- α -tubulin antibodies. The *p*-value was calculated by one-way 20 ANOVA followed by Tukey's multiple comparisons test (**b**, **c**, **d**).

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Supplementary Figure 2. NSP14 activates NF-κB via linear ubiquitination. (a) HEK293 cells
were transfected with FLAG-tagged NSP14 genes derived from various coronaviruses. After 48
hours, cell lysates were subjected to immunoprecipitation and subsequent blotting using the
indicated antibodies. (b) Lysates of HEK293 cells and two HOIP knockout cell lines were blotted

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26 as indicated. (c) Blotting analysis of lysates from HEK293 cells and two UBC13 knockout cell 27 lines. (d-e) Wild-type, HOIP knockout (d), and UBC13 (e) knockout HEK293 cells were stimulated 28 with 10 ng/mL TNF α for designated times. Subsequently, cell lysates were analyzed by Western 29 blotting. (f) Wild-type HEK293 cells, along with two UBC13 knockout and two HOIP knockout 30 HEK293 cell lines, were transfected with FLAG-tagged SARS-CoV-2 NSP14 along with NF-kB 31 reporter and pRL-SV40. After 48 hours, cells were collected, and the relative reporter activity was 32 determined by calculating the ratio of firefly luciferase to *Renilla* luciferase. (g) Blotting analysis 33 of lysates from OTULIN wild-type and knockout HEK293 cells. (h) OTULIN wild-type and knockout 34 HEK293 cells were stimulated with 10 ng/mL TNF α for designated times. Cell lysates were blotted 35 using the indicated antibodies. (i) Wild-type and two OTULIN knockout HEK293 cell lines were 36 transfected with FLAG-tagged SARS-CoV-2 NSP14 with the NF-κB reporter and pRL-SV40. The 37 p-value was calculated by two-way ANOVA followed by Sidak's multiple comparisons test (f, i). 38

39 Supplementary Figure 3. NSP14 interacts with HOIP. (a) NSP14-HA was co-transfected with 40 FLAG-tagged HOIP, HOIL-1, or SHARPIN into HEK293 cells. After 48 h, cell lysates were 41 subjected to immunoprecipitation with the anti-FLAG antibody and subsequent blotting with anti-42 FLAG and anti-HA antibodies. (b) NSP14-FLAG or GFP-FLAG was transfected into HEK293 43 cells. After 48 h, cell lysates were immunoprecipitated with the anti-FLAG antibody and blotted 44 using the indicated antibodies. (c) Vector or NSP14-HA was transfected into A549 cells. After 48 45 h, cells were fixed and stained with anti-HOIP (red), anti-HA (green), and DAPI nuclear stain (blue). Scale bar = 10 µm. (d) Schematic of HOIP mutants. PUB: Peptide N-glycanase/UBA or 46 47 UBX-containing proteins; ZnF: Zinc finger; NZF: Npl4 zinc finger; UBA: Ubiguitin-associated; 48 RBR: RING between RING fingers. (e) FLAG-tagged HOIP or the indicated mutant was co-49 transfected with Myc-tagged (NSP14-Myc) into HEK293 cells. After 48 h, cell lysates were 50 immunoprecipitated with the anti-FLAG antibody and blotted as indicated.

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52 Supplementary Figure 4. The IKK complex is required for NSP14-mediated NF-kB 53 activation. (a) Co-transfection of FLAG-tagged NSP14 with IKK α , IKK β , or p65, along with NF-54 κB reporter and pRL-SV40, into HEK293 cells. After 48 h, cells were collected, and the relative 55 reporter activity was determined by calculating the ratio of firefly luciferase to Renilla luciferase. 56 (**b-d**) Wild type, IKKα knockout (**b**), IKKβ knockout (**c**), or NEMO knockout (**d**) HEK293 cells were 57 stimulated with 10 ng/mL TNFα for designated times. Cell lysates were blotted as indicated. (e-58 g) Transfection of vector or FLAG-tagged NSP14 with NF-κB reporter and pRL-SV40 into wild-59 type, IKK α knockout (e), IKK β knockout (f), and NEMO knockout (g) HEK293 cells. After 48 h, 60 cells were collected, and the ratio of firefly luciferase to Renilla luciferase was calculated to 61 determine the relative reporter activity. (h) Vector or FLAG-tagged NSP14 was transfected with 62 the NF-kB reporter and pRL-SV40 into wild-type and TBK1 knockout MEFs. After 48 hours, cells were harvested, and the relative reporter activity was determined by calculating the ratio of firefly 63 luciferase to *Renilla* luciferase. (i) FLAG-tagged NSP14 or NSP14^{K/R} was transfected with Myc-64 tagged IKKα or IKKβ into HEK293 cells, along with NF-κB reporter and pRL-SV40. After 48 hours, 65 66 cells were harvested, and the relative reporter activity was determined by calculating the ratio of 67 firefly luciferase to *Renilla* luciferase. Lysates were blotted with anti-FLAG, anti-Myc, and anti- α -68 tubulin antibodies. The p-value was calculated by two-way ANOVA followed by Sidak's multiple 69 comparisons test (a, e, f, g, h) or one-way ANOVA followed by Tukey's multiple comparisons test 70 (i).

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Supplementary Figure 5. MAVS knockout impairs ISG but not proinflammatory factor expression. (a) MAVS wild-type and knockout A549 cells were infected with HCoV OC43 for the designated times. Cell lysates were blotted as indicated, and band densitometry was calculated using Image J. The ratio of phosphorylated IKK to total IKK in each lane was indicated. (b) HOIP wild-type and knockout H1299 cells were infected with 1 MOI of HCoV OC43 for 16 h. Real-time PCR was conducted to determine the relative mRNA levels of proinflammatory factors and ISGs. (c) MAVS wild-type and knockout A549 cells were infected with 1 MOI of HCoV OC43 for 16 h. Real-time PCR was conducted to determine the relative mRNA levels of proinflammatory factors and ISGs. (d) MTT assays of A549 cells treated with DMSO, 30 μ M HOIPIN-8, or 10 μ M IKK-16 for 5 days. The *p*-value was calculated by two-way ANOVA followed by Sidak's multiple comparisons test.

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Supplementary Figure 6. HOIPIN-8 inhibits HCoV OC43 viral replication. (a) HOIP wild-type 84 85 and knockout A549 cells were infected with 0.01 MOI of HCoV OC43 for the designated days. MTT assays were performed to determine cell viability. (b) HOIP wild-type and knockout H1299 86 87 cells were infected with 0.01 MOI of HCoV OC43 for the designated days. Lysates were blotted 88 as indicated. (c) A549 cells were treated with DMSO, HOIPIN-8, or IKK-16 for 2 h and then 89 infected with 0.01 MOI of HCoV OC43 for the designated days. TCID₅₀ of culture supernatants 90 containing HCoV OC43 were determined on Vero cells. The p-value was calculated by two-way 91 ANOVA followed by Sidak's multiple comparisons test. (d) A549 cells were treated with DMSO or 92 HOIPIN-8 for 2 h and then infected with 0.1 MOI HCoV OC43 for 24 h. Cells were stained with 93 anti-dsRNA (red) and DAPI (blue). The relative ratio of positively stained cells is summarized in 94 the graph. The *p*-value was calculated by *t*-test.

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Figure S1



Figure S2



Figure S3



Figure S4







MX1

Figure S5



Figure S6









Fig. 3f



Fig. 3g

Fig. S3a



Fig. S3b

Fig. S3e











Fig. 5e













Fig. S4c



Fig. S4d



Fig. S4i







Fig. 6e



Fig. S5a



Fig. S6b



Fig. 7b