

## Supporting Information

### PyINETA: Open-source platform for INADEQUATE-JRES integration in NMR metabolomics

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### *In vivo <sup>13</sup>C labeling and sample preparation*

All animal experiments and protocols have been reviewed and approved by the Institutional Animal Care and Use Committee of the Max Planck Institute of Psychiatry. Three 8-week-old male C57BL/6 mice (Charles River Laboratories, Maastricht, The Netherlands) were housed under standard conditions (12-h light/dark cycle, lights on at 0600 h, room temperature  $23 \pm 2$  °C, humidity 60%, tap water and food *ad libitum*) and fed with standard rodent diet (Harlan Laboratories, Inc., Indianapolis, IN, USA) for one week. For adaptation prior to labeling the animals were first fed an unlabeled *Ralstonia eutropha* bacterial protein-based rodent diet (Silantes GmbH, Munich, Germany) for 4 days. The food supply was then switched to <sup>13</sup>C-labeled *Ralstonia eutropha* bacterial diet (Silantes GmbH) for 14 days (Supplementary Tables S2 and S3). Following labeling the animals were sacrificed and organs and blood isolated. The partially <sup>13</sup>C-labeled animals did not show any discernible health effects compared to animals fed with a standard diet and had similar weight gains as animals fed with standard food (data not shown). Tissues were homogenized in 30 volumes of ice cold 80% methanol, homogenates were centrifuged and supernatants were dried. Dried samples were resuspended in 50 µL of deuterated water and methanol (1:4 volume ratio) (Supplementary Table 4).

### *NMR data collection and processing*

Data were collected on a Bruker Avance Neo 900 MHz with a 5-mm TXO cryoprobe (Bruker), using NMR tubes with a diameter of 1.7 mm (Bruker). For the INADEQUATE experiment, a default pulse program with adiabatic 180° pulses (Bruker nomenclature, `inadphppsp`) was used. For JRES, Bruker's default pulse program (`jresdcqf`) was modified to implement an adiabatic 180° pulse after we verified adiabatic 180° pulse is necessary in collecting <sup>13</sup>C-JRES on our 900 MHz magnet<sup>1</sup>. Detailed parameter settings are in Supplementary Table S5. TopSpin 4.0.9 was used to operate the spectrometer.

All the NMR spectra were processed using NMRPipe<sup>2</sup>. Briefly, for both INADEQUATE and JRES, the FIDs were Fourier-transformed after applying a squared sine-bell function and a double zero-filling on both direct and indirect dimensions. JRES spectra were further tilted and symmetrized. Detailed NMRPipe processing parameters are in Supplementary Table S6. Further data processing for JRES spectra was conducted using Metabolomics Toolbox ([https://github.com/edisonomics/metabolomics\\_toolbox](https://github.com/edisonomics/metabolomics_toolbox)) on MATLAB R2022b (MathWorks). Briefly, projection spectra were created from JRES, and they were aligned with the CCOW method (function 'guide\_align1D') and normalized by the probabilistic quotient normalization (PQN) method<sup>3</sup> ('normalize').

All the raw data, NMRPipe processing scripts, processed data, PyINETA output files, and MATLAB scripts are available in Metabolomics Workbench with Study ID ST003304.

### Example of configuration file in PyINETA

```
; This is the template for a config.ini file used with pyINETA
; Lines enclosed in "[" indicate section headers.
; Other lines have the options and parameters required by INETA
; Lines starting with a ";" are comments and describe the options
preceding it.

[PeakPick]
Ft_File =
<path_to_the_raw_data_file>/raw_data/pyINETA_AlaRef_2023/4_inad.ft2
Data_Matrix_File =
; (optional if no Ft file)
13C_Ppm_File =
; (optional if no Ft file)
Double_Quantum_File =
; (optional if no Ft file)

Xrange_min = 0
Xrange_max = 200
; The min and max ppm values for the X-axis (13C axis)
Yrange_min = 0
Yrange_max = 400
; The min and max ppm values for the Y-axis (DQ axis)
OutImage_pick_separate = fig_1plotSeparate.eps
OutImage_pick_complete = fig_1plotAll.eps
; Filenames for output images after peak picking

Shift = No
; Yes or No | Yes to shift the spectrum - This functionality is helpful
for aligning the INADEQUATE spectrum by shifting all the peaks uniformly
across one or both axes.
Direction = Pos
; Pos or Neg | Which direction to shift the spectra in
; For eg, Pos will move a peak at 35 ppm to 40 ppm
;          Neg will move a peak at 40 ppm to 35 ppm
Shift13C = 20
; # of units to shift the spectra
; In a typical INADEQUATE spectrum, 20 units ~ 1 ppm
Full13C = 4096
; Size of the 13C dimension in units; number of columns in the data
matrix.
FullDQ = 2048
; Size of the DQ dimension in units; number of rows in the data matrix.

PPmin = 5.7e10
```

```

; Minimum intensity to select a peak
PPmax = 4e11
; Maximum intensity of a peak
steps = 10
; Number of iterations of peak picking between PPmin and PPmax
; has to be at least 2

[ClusterPoints]
PPCS = 1
; All points within this range (in ppm) along the 13C axis will be
clustered into a single point.
;
PPDQ = 2
; All points within this range (in ppm) along the DQ axis will be
clustered into a single point.

OutImage_cluster_separate = fig_2clusterCenterSeparate.eps
OutImage_cluster_complete = fig_2clusterCenterAll.eps
; Filenames for output images after clustering

[FindNetwork]
Select = all
; all or last | use "all" for most cases
; "all" iterates over all steps to find peaks
; "last" selects peaks found in the last iteration only
LevelPointsDistance = 0.5
; Typical range: 0.5 - 2
; Distance between points (in ppm) across different levels to be
considered a new peak
; Lower values results in more points.
DQT = 0.2
; Typical range: 0.5 - 2
; (threshold for DQ - diffDQ < DQT for networking)
; (Control horizontal connection to be selected for network)
SumXY = 2
; Typical range: 0.5 - 1
; (threshold for sum 2X equals Y) (Control whether Y=X1+X2)
SDT = 0.5
; Typical range: 0.2 - 0.5
; (Symmetrical/Diagonal tolerance) (Equidistant from diagonal) 0.2-0.5
best if 0.3
CST = 0.05
; (chemical shift tolerance) (Controls vertical connections), 0.07

Network_output_file = file_3Networks.txt
OutImage_network_AllNets = fig_3findNetworkAllNets.eps

```

```

[MatchDatabase]
Database_file = <path_to_pyineta>/db/InetaDB.200922.json

Ambiguity = 1
; Higher values will include more database networks, for example, 0 - will
remove all, 1 - will include all
CSMT = 1
; 0.5,2 (chemical shift match tolerance,near) Higher value will increase
the x-axis range for a match between a network and db peak
Match_tolerance = 2
; NCMT, number of peak matches, Higher values will require more number of
matching peaks to consider a database hit
DQMT = 4
; (double quantum match tolerance) Higher value will increase the y-axis
range for a match between a network and db peak
Topology_tolerance = 2
; Higher values will require x and y values to be closer to the db peak to
consider a hit
Hit_Score_threshold = 0.2
; (ratio of matched-number of peaks in db) 0-1. A value of 1 means all db
peaks need to match.A value >1 indicates that multiple candidate peaks
matched the same db peak
Coverage_Score_threshold = 0.5
; (ratio of matched-number of peaks in network) 0-1. A value of 1 requires
all network peaks to match the db peaks

Matches_list_output_file = file_4Matches.txt
Summary_file = file_Summary.txt

[Overlay1D]
1D_File_List = <path_to_1D_file>/raw_data /file1.ft,/file2.ft,/file3.ft
; Comma separated list of 1D ft filenames
Peak_Width_1D = 0.5
; Typical range: 0.2 - 2
; When matching INADEQUATE networks with 1D spectra, this value (in ppm)
determines the peak width used to calculate peak area.
; Range of peak width calculation will be (peakPostion-Peak_Width_1D/2 to
peakPostion+Peak_Width_1D/2)
Intensity_threshold_1D = 10000
; Typical range:
; Intensity threshold for area under the curve around matched peaks to
flag as present or absent.
Match1d_output_file = file_5Match1ds.txt
OutImage_Match1d = fig_5highlight1dMatches.png
; Filename for output file with 1D matching results.
OutImageFormat1D = "png"
; Output image format: can be eps, jpeg, jpg, pdf, pgf, png, ps, raw,

```

```

rgba, svg, svgz, tif, tiff

Shift_1D = No
; Yes or No | Yes to shift all the provided 1D spectra
Direction_1D = Neg
; Pos or Neg; Which direction to shift the spectra in
; For eg, Pos will move a peak at 35 ppm is now at 40 ppm
;           Neg will move a peak at 40 ppm is now at 35 ppm
Shift_1D_val = 930
; 930 - 1.42; 780 - 1.19 ppm
; # of units to shift the spectra (650 units ~ 1 ppm for a spectra fo size
131072 units)
Full_1D = 131072
; Size of the 13C dimension in units

[OverlayJres]
Jres_File_List = /<path_to_Jres_file>/3.tilt.sym.ft2
; 40_cres.tilt.sym.ft
; Comma separated list of Jres ft filenames.

Jres_Projection_Method = max
; Can be sum,max or avg
Peak_Width_Jres = 0.5
; Typical range: 0.2 - 2
; When matching INADEQUATE networks with 1D spectra, this value (in ppm)
determines the peak width used to calculate peak area.
; Range of peak width calculation will be (peakPostion-Peak_Width_1D/2 to
peakPostion+Peak_Width_1D/2)
Intensity_threshold_Jres = 10000
; Typical range:
; Intensity threshold for area under the curve around matched peaks to
flag as present or absent.
MatchJres_output_file = file_5MatchJres.txt
OutImage_MatchJres = fig_5highlightJresMatches.png
; Filename for output file with 1D matching results.
OutImageFormatJres = png
; Output image format: can be eps, jpeg, jpg, pdf, pgf, png, ps, raw,
rgba, svg, svgz, tif, tiff

Shift_Jres = No
; Yes or No | Yes to shift all the provided 1D spectra
Direction_Jres = Neg
; Pos or Neg; Which direction to shift the spectra in
; For eg, Pos will move a peak at 35 ppm is now at 40 ppm
;           Neg will move a peak at 40 ppm is now at 35 ppm
Shift_Jres_val = 930
; 930 - 1.42; 780 - 1.19 ppm

```



```
; # of units to shift the spectra (650 units ~ 1 ppm for a spectra of size  
131072 units)  
Full_Jres = 131072  
; Size of the 13C dimension in units
```

### *Example of Summary file in PyINETA*

```
Summary for PyINETA run on 2024-06-07 15:17:21.483747 :  
# of picked peaks in all steps: 6629  
# of peaks after clustering: 2493  
# of peaks retained after merging steps: 1806  
# of networks found: 67  
# of matches found: 46
```

### Example of Networks file in PyINETA

```
Network1 [27.03 40.91],[13.72 41.01]
Network2 [38.58 56.17],[17.27 55.93]
Network3 [18.7 72.18],[53.2 71.98]
Network4 [31.73 51.09],[19.2 50.97]
Network5 [31.82 52.36],[20.57 52.45]
Network6 [68.52 90.83],[22.24 90.78]
Network7 [22.87 94.29],[22.76 93.85],[70.98 94.12]
Network8 [24.7 51.54],[42.71 69.7 ],[ 42.75 107.44],[26.95
69.7 ],[ 64.76 107.56],[ 64.36 107.25],[26.95 51.62],[23.43 50.35],[26.95
50.46]
Network9 [24.13 53.52],[29.14 53.52]
Network10 [32.59 57.04],[24.3 56.98]
Network11 [ 24.64 201.46],[176.75 201.46]
Network12 [25.69 67.17],[41.51 67.17]
Network13 [183.03 208.87],[ 25.8 209.03]
Network14 [26.42 86.41],[60.42 86.62]
Network15 [48.6 75.22],[26.53 75.15]
Network16 [182.24 243.22],[28.07 89.06],[ 60.97 243.13],[60.99 89.21]
Network17 [28.95 85.85],[56.68 85.82]
Network18 [29.03 70.84],[ 33.61 213.39],[29.04 62.72],[179.7
213.2],[33.57 62.62],[41.66 71. ]
Network19 [176.46 233.69],[29.51 86.95],[175.96 233. ],[57.33
86.94],[ 57.33 233.61]
Network20 [32.53 64.2 ],[31.67 64.35]
Network21 [ 32.4 216.14],[183.72 216.19]
Network22 [32.76 89.8 ],[57.01 89.9 ]
Network23 [176.93 210.88],[ 34.1 210.98]
Network24 [39.26 75.41],[40.64 80.07],[39.28 80.07],[35.96 75.41]
Network25 [57.67 93.92],[36.09 94.03]
Network26 [183.33 219.95],[ 36.39 220. ]
Network27 [ 36.72 221.34],[184.84 221.45]
Network28 [52.6 90.24],[37.61 90.11]
Network29 [ 37.96 214.07],[176.08 214.05],[176.12 238.9 ],[ 62.97 239.03]
Network30 [49.98 88.1 ],[38.15 88.13]
Network31 [41.77 97.25],[55.36 97.25]
Network32 [56.02 98.68],[42.53 98.68]
Network33 [ 43.35 106.54],[ 63.12 106.42]
Network34 [53.69 97.62],[43.76 97.62]
Network35 [ 44. 218.17],[ 43.95 218.75],[174.37 218.7 ],[174.43
218.33],[174.34 233.4 ],[ 59.26 233.4 ]
Network36 [ 60.23 104.39],[ 44.13 104.39]
Network37 [180.62 225.68],[ 44.86 225.79]
Network38 [178.15 223.78],[178.14 224.28],[ 46. 223.94],[ 46.14 224.34]
Network39 [ 46.86 115.12],[49.38 96.25],[46.84 96.37],[ 68.23 115.23]
Network40 [ 52.76 114.37],[ 61.89 114.52]
```

```
Network41 [ 53.12 230.86],[177.71 230.86]
Network42 [ 55.28 229.4 ],[173.87 229.56],[174.04 229.21]
Network43 [177.5 233.48],[ 55.83 233.72]
Network44 [ 58.28 128.55],[ 70.17 128.58]
Network45 [ 60.68 122.31],[ 61.52 122.37]
Network46 [ 68.97 130.51],[ 62.07 130.8 ],[ 61.04 130.4 ]
Network47 [175.75 238.53],[ 63.04 238.26]
Network48 [ 74.1 146.86],[ 72.59 146.83],[ 74.1 137.65],[ 63.49 137.64]
Network49 [ 63.56 142.6 ],[ 78.81 142.35]
Network50 [ 73.62 138.44],[ 64.94 138.43]
Network51 [ 65.74 243.31],[177.61 243.39]
Network52 [ 85.82 153.46],[ 67.59 153.56]
Network53 [ 73.8 142.03],[ 67.95 141.93]
Network54 [171.19 239.93],[ 68.8 239.96]
Network55 [ 71.04 255.69],[184.53 255.64]
Network56 [ 94.87 169.46],[ 74.43 150.18],[ 75.56 150.18],[ 74.41 169.4 ]
Network57 [ 78.72 155.7 ],[ 98.79 175.98],[ 76.99 155.7 ],[ 77.01 176.11]
Network58 [ 77.39 167.15],[ 89.73 167.02]
Network59 [ 79.69 251.91],[172.08 251.96]
Network60 [ 85.51 177.14],[ 91.48 177.14]
Network61 [168.57 273.8 ],[105.15 273.37]
Network62 [126.48 244.29],[117.68 244.4 ]
Network63 [119.39 254.13],[134.66 254.13]
Network64 [121.12 272.98],[151.69 272.98]
Network65 [126.82 281.2 ],[154.51 281.4 ]
Network66 [151.08 278.03],[126.96 278.13]
Network67 [178.26 363.89],[186.07 363.89]
```

Example of Matches file in PyINETA

```

#NetworkNum ID MatchName Solvent AmbiguityScore Hitscore
CoverageScore MatchedConnections UnmatchedConnections
Network1 bmse000041 L_ileucine D20 0.0 0.2 1.0 CX1-CX2-
>C1-C6,
Network1 bmse000319 3_Methyl_2_oxopentanoic_acid D20 0.0 0.2
1.0 CX1-CX2->C5-C8,
Network1 bmse000574 3_Methyl_2_oxopentanoic_acid D20 0.0 0.2
1.0 CX1-CX2->C5-C8,
Network1 bmse000578 ethylmalonic_acid D20 0.0 0.25 1.0 CX1-
CX2->C6-C7,
Network1 bmse000866 L_ileucine D20 0.0 0.2 1.0 CX1-CX2-
>C1-C6,
Network2 bmse000041 L_ileucine D20 0.0 0.2 1.0 CX2-CX1-
>C7-C2,
Network2 bmse000866 L_ileucine D20 0.0 0.2 1.0 CX2-CX1-
>C7-C2,
Network3 bmse000028 L_alanine D20 0.0 0.5 1.0 CX2-CX1->C2-
C5,
Network3 bmse000236 D_alanine D20 0.0 0.5 1.0 CX2-CX1->C2-
C3,
Network3 bmse000282 DL_Alanine D20 0.0 0.5 1.0 CX2-CX1->C4-
C5,
Network3 bmse000403 Ala_Ala D20 0.0 0.25 1.0 CX2-CX1->C6-
C9,
Network4 bmse000052 L_valine D20 0.4 0.25 1.0 CX2-CX1->C7-
C5,
Network4 bmse000466 DL_beta_leucine D20 0.0 0.2 1.0 CX2-
CX1->C8-C4,
Network4 bmse000860 L_valine D20 0.4 0.25 1.0 CX2-CX1->C7-
C5,
Network5 bmse000052 L_valine D20 0.4 0.25 1.0 CX1-CX2->C5-
C7,
Network5 bmse000466 DL_beta_leucine D20 0.0 0.2 1.0 CX1-
CX2->C4-C8,
Network5 bmse000860 L_valine D20 0.4 0.25 1.0 CX1-CX2->C5-
C7,
Network6 bmse000049 L_threonine D20 0.0 0.333 1.0 CX2-CX1-
>C1-C2,
Network6 bmse000794 3_carboxypropyl_trimethyl_ammonium D20 0.571
0.333 1.0 CX2-CX1->C5-C4,
Network6 bmse000859 L_threonine D20 0.0 0.333 1.0 CX2-CX1-
>C1-C2,
Network7 bmse000208 L_lactic_acid D20 0.0 0.5 0.667 CX3-CX1-
>C2-C3,CX3-CX3->C2-C2, CX2-CX3->?-?,
Network7 bmse000269 R_Lactate D20 0.0 0.5 0.667 CX3-CX1->C4-

```

```

C5,CX3-CX3->C4-C4, CX2-CX3->?-?,
Network7  bmse000979 L_lactic_acid D2O 0.0 0.5 0.667 CX3-CX1-
>C2-C3,CX3-CX3->C2-C2, CX2-CX3->?-?,
Network9  bmse000043 L_lysine D2O 0.0 0.2 1.0 CX1-CX2->C6-
C7,
Network9  bmse000072 cadaverine D2O 0.0 0.25 1.0 CX1-CX2->C7-
C6,
Network9  bmse000237 DL_pipecolic_acid D2O 0.0 0.2 1.0 CX1-
CX2->C4-C5,
Network9  bmse000373 isovaleric_acid D2O 0.0 0.25 1.0 CX1-
CX2->C6-C3,
Network9  bmse000411 L_Norleucine D2O 0.0 0.2 1.0 CX1-CX2-
>C7-C5,
Network9  bmse000493 epsilon_caprolactone CDC13 0.0 0.2 1.0
CX1-CX2->C4-C3,
Network9  bmse000571 2_hydroxyhexanoic_acid D2O 0.0 0.2 1.0
CX1-CX2->C6-C4,
Network10 bmse000043 L_lysine D2O 0.0 0.2 1.0 CX2-CX1->C6-
C5,
Network10 bmse000249 methyl_4_aminobutyrate D2O 0.0 0.333 1.0
CX2-CX1->C2-C4,
Network10 bmse000372 epsilon_caprolactam D2O 0.0 0.2 1.0
CX2-CX1->C5-C3,
Network10 bmse000429 DL_2_Aminoadipic_acid D2O 0.0 0.2 1.0
CX2-CX1->C7-C6,
Network10 bmse000442 L_homocitrulline D2O 0.0 0.2 1.0 CX2-
CX1->C7-C9,
Network10 bmse000745 homoarginine D2O 0.0 0.2 1.0 CX2-CX1-
>C7-C8,
Network11 bmse000145 N_acetyl_L_glutamine D2O 0.286 0.2 1.0
CX1-CX2->C1-C2,
Network11 bmse000157 N_acetyl_L_alanine D2O 0.0 0.333 1.0 CX1-
CX2->C9-C7,
Network11 bmse000382 N_Acetyl_L_glutamic_acid D2O 0.0 0.2 1.0
CX1-CX2->C13-C11,
Network11 bmse000423 N_acetyl_L_aspartic_acid D2O 0.0 0.25 1.0
CX1-CX2->C12-C11,
Network11 bmse000468 4_Acetamidobutyric_acid D2O 0.0 0.25 1.0
CX1-CX2->C10-C9,
Network11 bmse000707 N_acetyl_histidine D2O 0.0 0.2 1.0 CX1-
CX2->C14-C12,
Network11 bmse000983 O_acetyl_L_serine D2O 1.0 0.333 1.0 CX1-
CX2->C10-C9,
Network12 bmse000019 D_ornithine D2O 0.0 0.25 1.0 CX1-CX2-
>C4-C5,
Network12 bmse000109 putrescine D2O 0.0 0.333 1.0 CX1-CX2->C5-
C4,

```

```

Network12 bmse000116 spermidine D20 1.0 0.2 1.0 CX1-CX2->C8-
C9,
Network12 bmse000162 L_ornithine D20 0.0 0.25 1.0 CX1-CX2-
>C4-C5,
Network12 bmse000195 N_alpha_acetyl_DL_ornithine D20 0.0 0.333
1.0 CX1-CX2->C8-C11,
Network12 bmse000249 methyl_4_aminobutyrate D20 0.0 0.333 1.0
CX1-CX2->C2-C1,
Network12 bmse000340 gamma_Aminobutyric_acid D20 0.0 0.333 1.0
CX1-CX2->C4-C6,
Network12 bmse000468 4_Acetamidobutyric_acid D20 0.0 0.25 1.0
CX1-CX2->C5-C6,
Network12 bmse000862 putrescine D20 0.0 0.333 1.0 CX1-CX2->C5-
C4,
Network12 bmse000871 gamma_Aminobutyric_acid D20 0.0 0.333 1.0
CX1-CX2->C4-C6,
Network12 bmse000897 D_ornithine D20 0.0 0.25 1.0 CX1-CX2-
>C4-C5,
Network12 bmse000951 spermidine D20 1.0 0.2 1.0 CX1-CX2->C8-
C9,
Network15 bmse000047 L_proline D20 0.0 0.25 1.0 CX2-CX1->C6-
C7,
Network15 bmse000116 spermidine D20 1.0 0.2 1.0 CX2-CX1->C8-
C7,
Network15 bmse000947 L_proline D20 0.0 0.25 1.0 CX2-CX1->C6-
C7,
Network15 bmse000951 spermidine D20 1.0 0.2 1.0 CX2-CX1->C8-
C7,
Network17 bmse000037 L_glutamic_acid D20 0.0 0.25 1.0 CX2-
CX1->C4-C6,
Network17 bmse000038 L_glutamine D20 0.0 0.25 1.0 CX2-CX1-
>C2-C5,
Network17 bmse000194 L_arginine_L_glutamate D20 0.0 0.2 1.0
CX2-CX1->C4-C2,
Network18 bmse000038 L_glutamine D20 0.0 0.5 0.667 CX5-CX3-
>C6-C5,CX5-CX2->C6-C6,CX4-CX2->C7-C6, CX1-CX3->?-?,CX1-CX6->?-?,
Network18 bmse000565 cis_5_dodecenoic_acid CDC13 0.0 0.273 0.667
CX5-CX3->C3-C5,CX5-CX2->C3-C13,CX4-CX2->C14-C13, CX1-CX3->?-?,CX1-CX6-
>?-?,
Network18 bmse000625 4_methylvaleric_acid CDC13 0.0 0.4 0.5
CX5-CX2->C4-C5,CX4-CX2->C8-C5, CX1-CX3->?-?,CX1-CX6->?-?,CX5-CX3->C4-?,
Network19 bmse000019 D_ornithine D20 0.0 0.5 0.8 CX4-CX2-
>C2-C3,CX4-CX5->C2-C2,CX5-CX5->C2-C2,CX5-CX1->C2-C1, CX3-CX5->?-?,
Network19 bmse000029 L_arginine D20 0.0 0.5 0.8 CX4-CX2->C2-
C5,CX4-CX5->C2-C2,CX5-CX5->C2-C2,CX5-CX1->C2-C3, CX3-CX5->?-?,
Network19 bmse000032 L_citrulline D20 0.0 0.5 0.8 CX4-CX2-
>C4-C6,CX4-CX5->C4-C4,CX5-CX5->C4-C4,CX5-CX1->C4-C1, CX3-CX5->?-?,

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Network19  bmse000037  L_glutamic_acid      D20    0.0    0.5    0.8    CX4-
CX2->C4-C6,CX4-CX5->C4-C4,CX5-CX5->C4-C4,CX5-CX1->C4-C1,    CX3-CX5->?-?,
Network19  bmse000038  L_glutamine          D20    0.0    0.5    0.8    CX4-CX2-
>C2-C5,CX4-CX5->C2-C2,CX5-CX5->C2-C2,CX5-CX1->C2-C3,    CX3-CX5->?-?,
Network19  bmse000039  L_histidine          D20    0.333  0.5    0.8    CX4-CX2-
>C5-C3,CX4-CX5->C5-C5,CX5-CX5->C5-C5,CX5-CX1->C5-C6,    CX3-CX5->?-?,
Network19  bmse000162  L_ornithine          D20    0.0    0.5    0.8    CX4-CX2-
>C2-C3,CX4-CX5->C2-C2,CX5-CX5->C2-C2,CX5-CX1->C2-C7,    CX3-CX5->?-?,
Network19  bmse000185  L_glutathione_reduced D20    0.0    0.429  1.0
CX4-CX2->C2-C6,CX4-CX5->C2-C2,CX5-CX5->C2-C2,CX5-CX1->C2-C3,CX3-CX5->C12-
C2,
Network19  bmse000209  D_citrulline         D20    0.0    0.5    0.8    CX4-CX2-
>C6-C11,CX4-CX5->C6-C6,CX5-CX5->C6-C6,CX5-CX1->C6-C2,    CX3-CX5->?-?,
Network19  bmse000289  S_Adenosyl_L_homocysteine D20    0.143  0.222  0.8
CX4-CX2->C13-C15,CX4-CX5->C13-C13,CX5-CX5->C13-C13,CX5-CX1->C13-C21,
CX3-CX5->?-?,
Network19  bmse000449  Npai_Methyl_L_histidine D20    0.0    0.25   0.6
CX4-CX5->C7-C7,CX5-CX5->C7-C7,CX5-CX1->C7-C11,    CX4-CX2->C7-?,CX3-CX5->?-?,
Network19  bmse000711  L_arginine           D20    0.0    0.5    0.8    CX4-CX2->C2-
C5,CX4-CX5->C2-C2,CX5-CX5->C2-C2,CX5-CX1->C2-C3,    CX3-CX5->?-?,
Network19  bmse000858  L_citrulline         D20    0.0    0.5    0.8    CX4-CX2-
>C4-C6,CX4-CX5->C4-C4,CX5-CX5->C4-C4,CX5-CX1->C4-C1,    CX3-CX5->?-?,
Network19  bmse000897  D_ornithine          D20    0.0    0.5    0.8    CX4-CX2-
>C2-C3,CX4-CX5->C2-C2,CX5-CX5->C2-C2,CX5-CX1->C2-C1,    CX3-CX5->?-?,
Network19  bmse000952  L_glutathione_reduced D20    0.0    0.429  1.0
CX4-CX2->C2-C6,CX4-CX5->C2-C2,CX5-CX5->C2-C2,CX5-CX1->C2-C3,CX3-CX5->C12-
C2,
Network21  bmse000483  R_2_Pyrrolidinone_5_carboxylate D20    0.0
0.25  1.0    CX1-CX2->C7-C8,
Network21  bmse000878  R_2_Pyrrolidinone_5_carboxylate D20    0.0
0.25  1.0    CX1-CX2->C7-C8,
Network22  bmse000033  L_cystathionine      D20    0.0    0.333  1.0    CX1-
CX2->C2-C10,
Network22  bmse000043  L_lysine             D20    0.0    0.2    1.0    CX1-CX2->C5-
C2,
Network22  bmse000044  L_methionine         D20    0.0    0.333  1.0    CX1-CX2-
>C5-C2,
Network22  bmse000169  L_selenomethionine  D20    0.0    0.333  1.0    CX1-
CX2->C6-C2,
Network22  bmse000291  Selenomethionine    D20    0.0    0.333  1.0    CX1-
CX2->C7-C5,
Network22  bmse000411  L_Norleucine        D20    0.0    0.2    1.0    CX1-CX2-
>C4-C6,
Network22  bmse000429  DL_2_Aminoadipic_acid D20    0.0    0.2    1.0
CX1-CX2->C6-C8,
Network22  bmse000442  L_homocitrulline     D20    0.0    0.2    1.0    CX1-
CX2->C8-C10,

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Network22  bmse000450  D_Methionine      D2O      0.0      0.333    1.0      CX1-CX2->C5-C6,
Network22  bmse000466  DL_beta_leucine    D2O      0.0      0.2      1.0      CX1-CX2->C4-C5,
Network22  bmse000745  homoarginine      D2O      0.0      0.2      1.0      CX1-CX2->C8-C10,
Network23  bmse000493  epsilon_caprolactone CDC13    0.0      0.2      1.0      CX1-CX2->C8-C6,
Network24  bmse000159  beta_alanine      D2O      0.0      0.5      0.5      CX4-CX1->C2-C6,
           CX1-CX3->C6-?,CX3-CX2->?-?,
Network24  bmse000388  Cysteamine        D2O      0.0      1.0      0.5      CX4-CX1->C4-C3,
           CX1-CX3->C3-?,CX3-CX2->?-?,
Network24  bmse000967  beta_alanine      D2O      0.0      0.5      0.5      CX4-CX1->C2-C6,
           CX1-CX3->C6-?,CX3-CX2->?-?,
Network25  bmse000452  Hypotaurine       D2O      0.0      1.0      1.0      CX1-CX2->C5-C6,
Network26  bmse000037  L_glutamic_acid   D2O      0.0      0.25     1.0      CX1-CX2->C2-C7,
Network26  bmse000794  3_carboxypropyl_trimethyl_ammonium D2O      0.333    1.0      0.571    CX1-CX2->C10-C9,
Network27  bmse000037  L_glutamic_acid   D2O      0.0      0.25     1.0      CX1-CX2->C7-C2,
Network27  bmse000183  succinic_acid     D2O      0.0      0.333    1.0      CX1-CX2->C6-C2,
Network27  bmse000340  gamma_Aminobutyric_acid D2O      0.0      0.333    1.0      CX1-CX2->C5-C7,
Network27  bmse000344  4_Guanidinobutyric_acid D2O      0.0      0.333    1.0      CX1-CX2->C7-C9,
Network27  bmse000376  N_carbamyl_L_glutamate D2O      0.0      0.25     1.0      CX1-CX2->C10-C12,
Network27  bmse000382  N_Acetyl_L_glutamic_acid D2O      0.0      0.2      1.0      CX1-CX2->C9-C12,
Network27  bmse000468  4_Acetamidobutyric_acid D2O      0.0      0.25     1.0      CX1-CX2->C7-C8,
Network27  bmse000871  gamma_Aminobutyric_acid D2O      0.0      0.333    1.0      CX1-CX2->C5-C7,
Network29  bmse000030  L_asparagine      D2O      0.5      0.333    0.5      CX2-CX1->C6-C5,
           CX2-CX3->C6-?,CX3-CX4->?-?,
Network29  bmse000041  L_ileucine        D2O      0.0      0.2      0.5      CX3-CX4->C4-C3,
           CX2-CX1->?-?,CX2-CX3->?-?,
Network29  bmse000049  L_threonine       D2O      0.0      0.333    0.5      CX3-CX4->C4-C3,
           CX2-CX1->?-?,CX2-CX3->?-?,
Network29  bmse000052  L_valine          D2O      0.4      0.25     0.5      CX3-CX4->C3-C2,
           CX2-CX1->?-?,CX2-CX3->?-?,
Network29  bmse000123  trans_4_hydroxy_L_proline D2O      0.0      0.25     0.5      CX3-CX4->C4-C1,
           CX2-CX1->?-?,CX2-CX3->?-?,
Network29  bmse000148  diethyl_oxalacetate D2O      1.0      0.2      0.5

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CX3-CX4->C7-C6,      CX2-CX1->?-?,CX2-CX3->?-?,
Network29  bmse000631 O_phospho_DL_threonine D2O      0.0      0.333  0.5
CX3-CX4->C12-C10,   CX2-CX1->?-?,CX2-CX3->?-?,
Network29  bmse000741 N_methyl_L_aspartic_acid D2O      0.4      1.0      1.0
CX2-CX1->C10-C7,CX2-CX3->C10-C8,CX3-CX4->C8-C6,
Network29  bmse000859 L_threonine D2O      0.0      0.333  0.5      CX3-CX4->
C4-C3,      CX2-CX1->?-?,CX2-CX3->?-?,
Network29  bmse000860 L_valine D2O      0.4      0.25   0.5      CX3-CX4->C3-
C2,      CX2-CX1->?-?,CX2-CX3->?-?,
Network29  bmse000866 L_ileucine D2O      0.0      0.2     0.5      CX3-CX4->
C4-C3,      CX2-CX1->?-?,CX2-CX3->?-?,
Network29  bmse000966 trans_4_hydroxy_L_proline D2O      0.0      0.25   0.5
CX3-CX4->C4-C1,      CX2-CX1->?-?,CX2-CX3->?-?,
Network30  bmse000120 taurine D2O      0.0      1.0     1.0      CX1-CX2->C2-
C7,
Network31  bmse000042 L_leucine D2O      0.333   0.2     1.0      CX1-CX2->C5-
C2,
Network31  bmse000423 N_acetyl_L_aspartic_acid D2O      0.0      0.25   1.0
CX1-CX2->C8-C7,
Network32  bmse000042 L_leucine D2O      0.333   0.2     1.0      CX2-CX1->C5-
C2,
Network32  bmse000423 N_acetyl_L_aspartic_acid D2O      0.0      0.25   1.0
CX2-CX1->C8-C7,
Network32  bmse000453 Ureidosuccinic_acid D2O      0.0      0.333   1.0
CX2-CX1->C9-C8,
Network33  bmse000308 2_Aminoethyl_dihydrogen_phosphate D2O      0.0      1.0
1.0      CX2-CX1->C7-C8,
Network36  bmse000276 Ethanolamine D2O      0.0      1.0     1.0      CX2-CX1->
C4-C3,
Network37  bmse000074 D_carnitine D2O      0.571   0.333   1.0      CX1-CX2->
C9-C8,
Network37  bmse000211 L_carnitine D2O      0.0      0.333   1.0      CX1-CX2->
C1-C2,
Network37  bmse000949 D_carnitine D2O      0.571   0.333   1.0      CX1-CX2->
C9-C8,
Network38  bmse000367 chloroacetic_acid D2O      0.0      1.0     0.5      CX3-
CX3->C4-C4,CX1-CX3->C5-C4,CX1-CX1->C5-C5, CX3-CX2->C4-?,CX4-CX4->?-?,CX2-
CX4->?-?,CX2-CX2->?-?,CX2-CX1->?-?,CX1-CX4->C5-?,
Network38  bmse000658 phenylacetylglycine D2O      0.0      0.333   0.75
CX3-CX3->C10-C10,CX3-CX2->C10-C14,CX2-CX2->C14-C14,CX2-CX1->C14-C7,CX1-
CX3->C7-C10,CX1-CX1->C7-C7, CX4-CX4->?-?,CX2-CX4->C14-?,CX1-CX4->C7-?,
Network41  bmse000028 L_alanine D2O      0.0      0.5     1.0      CX1-CX2->C2-
C3,
Network41  bmse000236 D_alanine D2O      0.0      0.5     1.0      CX1-CX2->C2-
C4,
Network41  bmse000282 DL_Alanine D2O      0.0      0.5     1.0      CX1-CX2->C4-
C6,

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Network43	bmse000019	D_ornithine	D2O	0.0	0.25	1.0	CX1-CX2->C1-C2,
Network43	bmse000038	L_glutamine	D2O	0.0	0.25	1.0	CX1-CX2->C3-C2,
Network43	bmse000040	L_homoserine	D2O	0.0	0.333	1.0	CX1-CX2->C3-C2,
Network43	bmse000042	L_leucine	D2O	0.333	0.2	1.0	CX1-CX2->C3-C2,
Network43	bmse000044	L_methionine	D2O	0.0	0.333	1.0	CX1-CX2->C3-C2,
Network43	bmse000073	L_canavanine	D2O	0.0	0.333	1.0	CX1-CX2->C5-C2,
Network43	bmse000078	creatine	D2O	0.0	1.0	1.0	CX1-CX2->C4-C5,
Network43	bmse000162	L_ornithine	D2O	0.0	0.25	1.0	CX1-CX2->C7-C2,
Network43	bmse000430	DL_homocysteine	D2O	0.0	0.333	1.0	CX1-CX2->C8-C5,
Network43	bmse000450	D_Methionine	D2O	0.0	0.333	1.0	CX1-CX2->C8-C6,
Network43	bmse000897	D_ornithine	D2O	0.0	0.25	1.0	CX1-CX2->C1-C2,
Network43	bmse000950	creatine	D2O	0.0	1.0	1.0	CX1-CX2->C4-C5,
Network43	bmse000983	O_acetyl_L_serine	D2O	1.0	0.333	1.0	CX1-CX2->C8-C6,
Network44	bmse000285	Choline	D2O	0.8	1.0	1.0	CX1-CX2->C3-C4,
Network44	bmse000953	Choline	D2O	0.8	1.0	1.0	CX1-CX2->C3-C4,
Network47	bmse000049	L_threonine	D2O	0.0	0.333	1.0	CX2-CX1->C3-C4,
Network47	bmse000148	diethyl_oxalacetate	D2O	1.0	0.2	1.0	CX2-CX1->C6-C7,
Network47	bmse000631	O_phospho_DL_threonine	D2O	0.0	0.333	1.0	CX2-CX1->C10-C12,
Network47	bmse000859	L_threonine	D2O	0.0	0.333	1.0	CX2-CX1->C3-C4,
Network48	bmse000008	D_allose	D2O	0.667	0.4	0.75	CX4-CX3->C4-C1, CX1-CX3->C9-C1, CX2-CX1->?-?,
Network48	bmse000013	D_galactose	D2O	1.0	0.2	0.75	CX4-CX3->C6-C5, CX1-CX3->C5-C5, CX2-CX1->?-?,
Network48	bmse000017	D_maltose	D2O	0.917	0.2	0.75	CX4-CX3->C6-C2, CX1-CX3->C8-C2, CX2-CX1->?-?,
Network48	bmse000018	D_mannose	D2O	1.0	0.2	0.75	CX4-CX3->C7-C6, CX1-CX3->C6-C6, CX2-CX1->?-?,
Network48	bmse000022	D_sorbose	D2O	0.5	0.4	0.75	CX4-CX3->C8-

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C10,CX1-CX3->C6-C10, CX2-CX1->?=?,
Network48 bmse000023 D_tagatose D20 0.833 0.4 0.75 CX4-CX3->C8-
C10,CX1-CX3->C6-C10, CX2-CX1->?=?,
Network48 bmse000026 D_xylose D20 1.0 0.25 0.75 CX4-CX3->C5-
C4,CX1-CX3->C4-C4, CX2-CX1->?=?,
Network48 bmse000086 alpha_D_glucose_1_phosphate D20 0.0 0.4
1.0 CX4-CX3->C6-C5,CX2-CX1->C4-C5,CX1-CX3->C5-C5,
Network48 bmse000087 alpha_D_glucose_1_6_bisphosphate D20 0.0 0.2
0.5 CX2-CX1->C4-C5, CX4-CX3->?=?,CX1-CX3->C5-?,
Network48 bmse000099 D_mannitol D20 0.0 0.4 0.75 CX2-CX1->C7-
C9,CX1-CX3->C9-C3, CX4-CX3->?=?,
Network48 bmse000115 D_sorbitol D20 1.0 0.2 0.75 CX4-CX3->C6-
C5,CX1-CX3->C5-C5, CX2-CX1->?=?,
Network48 bmse000138 D_cellobiose D20 1.0 0.2 0.75 CX4-CX3-
>C6-C2,CX1-CX3->C8-C2, CX2-CX1->?=?,
Network48 bmse000151 alpha_D_galactose_1_phosphate D20 0.0 0.4
1.0 CX4-CX3->C6-C5,CX2-CX1->C4-C5,CX1-CX3->C5-C5,
Network48 bmse000163 N_acetyl_D_glucosamine_1_phosphate D20 0.625
0.333 0.75 CX4-CX3->C15-C10,CX1-CX3->C7-C10, CX2-CX1->?=?,
Network48 bmse000189 D_glucosamine_6_phosphate D20 0.0 0.2 0.5
CX2-CX1->C3-C4, CX4-CX3->?=?,CX1-CX3->C4-?,
Network48 bmse000233 melibiose D20 0.0 0.2 0.5 CX2-CX1->C9-
C7, CX4-CX3->?=?,CX1-CX3->C7-?,
Network48 bmse000235 L_gulonolactone D20 0.0 0.6 1.0 CX4-
CX3->C8-C4,CX2-CX1->C2-C5,CX1-CX3->C5-C4,
Network48 bmse000865 D_tagatose D20 0.833 0.4 0.75 CX4-CX3->C8-
C10,CX1-CX3->C6-C10, CX2-CX1->?=?,
Network48 bmse000898 D_xylose D20 1.0 0.25 0.75 CX4-CX3->C5-
C4,CX1-CX3->C4-C4, CX2-CX1->?=?,
Network48 bmse000903 D_sorbose D20 0.5 0.4 0.75 CX4-CX3->C8-
C10,CX1-CX3->C6-C10, CX2-CX1->?=?,
Network48 bmse000939 D_cellobiose D20 1.0 0.2 0.75 CX4-CX3-
>C6-C2,CX1-CX3->C8-C2, CX2-CX1->?=?,
Network48 bmse000946 D_maltose D20 0.917 0.2 0.75 CX4-CX3->C6-
C2,CX1-CX3->C8-C2, CX2-CX1->?=?,
Network48 bmse001006 D_galactose D20 1.0 0.2 0.75 CX4-CX3-
>C6-C5,CX1-CX3->C5-C5, CX2-CX1->?=?,
Network48 bmse001007 D_sorbitol D20 1.0 0.2 0.75 CX4-CX3->C6-
C5,CX1-CX3->C5-C5, CX2-CX1->?=?,
Network48 bmse001008 D_allose D20 0.667 0.4 0.75 CX4-CX3->C4-
C1,CX1-CX3->C9-C1, CX2-CX1->?=?,
Network50 bmse000008 D_allose D20 0.667 0.2 1.0 CX2-CX1->C4-
C1,
Network50 bmse000010 D_fructose D20 1.0 0.2 1.0 CX2-CX1->C11-
C8,
Network50 bmse000013 D_galactose D20 1.0 0.2 1.0 CX2-CX1-
>C6-C5,

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Network50	bmse000018	D_mannose	D2O	1.0	0.2	1.0	CX2-CX1->C7-C6,
Network50	bmse000022	D_sorbose	D2O	0.5	0.2	1.0	CX2-CX1->C8-C10,
Network50	bmse000023	D_tagatose	D2O	0.833	0.2	1.0	CX2-CX1->C8-C10,
Network50	bmse000062	adonitol	D2O	0.0	0.25	1.0	CX2-CX1->C9-C7,
Network50	bmse000068	L_arabitol	D2O	0.0	0.25	1.0	CX2-CX1->C7-C3,
Network50	bmse000084	gluconic_acid	D2O	0.0	1.0	1.0	CX2-CX1->C6-C5,
Network50	bmse000087	alpha_D_glucose_1_6_bisphosphate	D2O	0.0	0.2	1.0	CX2-CX1->C6-C5,
Network50	bmse000095	i_erythritol	D2O	0.0	0.333	1.0	CX2-CX1->C5-C4,
Network50	bmse000099	D_mannitol	D2O	0.0	0.2	1.0	CX2-CX1->C2-C3,
Network50	bmse000100	meso_erythritol	D2O	0.0	0.333	1.0	CX2-CX1->C5-C4,
Network50	bmse000115	D_sorbitol	D2O	1.0	0.2	1.0	CX2-CX1->C6-C5,
Network50	bmse000121	L_threitol	D2O	0.0	0.333	1.0	CX2-CX1->C5-C4,
Network50	bmse000129	xylitol	D2O	0.0	0.25	1.0	CX2-CX1->C5-C4,
Network50	bmse000151	alpha_D_galactose_1_phosphate	D2O	0.0	0.2	1.0	CX2-CX1->C6-C5,
Network50	bmse000184	glycerol	D2O	0.0	0.5	1.0	CX2-CX1->C6-C1,
Network50	bmse000193	DL_alpha_glycerol_phosphate	D2O	0.0	0.5	1.0	CX2-CX1->C10-C7,
Network50	bmse000230	D_glucono_1_5_lactone	D2O	0.0	1.0	1.0	CX2-CX1->C6-C5,
Network50	bmse000235	L_gulonolactone	D2O	0.0	0.2	1.0	CX2-CX1->C8-C4,
Network50	bmse000440	D_Glucosaminic_acid	D2O	0.0	0.2	1.0	CX2-CX1->C12-C10,
Network50	bmse000856	glycerol	D2O	0.0	0.5	1.0	CX2-CX1->C6-C1,
Network50	bmse000865	D_tagatose	D2O	0.833	0.2	1.0	CX2-CX1->C8-C10,
Network50	bmse000869	L_arabitol	D2O	0.0	0.25	1.0	CX2-CX1->C7-C3,
Network50	bmse000903	D_sorbose	D2O	0.5	0.2	1.0	CX2-CX1->C8-C10,
Network50	bmse001005	D_fructose	D2O	1.0	0.2	1.0	CX2-CX1->C11-

C8,  
Network50 bmse001006 D\_galactose D20 1.0 0.2 1.0 CX2-CX1->C6-C5,  
Network50 bmse001007 D\_sorbitol D20 1.0 0.2 1.0 CX2-CX1->C6-C5,  
Network50 bmse001008 D\_allose D20 0.667 0.2 1.0 CX2-CX1->C4-C1,  
Network52 bmse000204 D\_ribose\_5\_phosphate D20 1.0 0.25 1.0 CX1-CX2->C7-C6,  
Network53 bmse000018 D\_mannose D20 1.0 0.2 1.0 CX2-CX1->C7-C6,  
Network53 bmse000193 DL\_alpha\_glycerol\_phosphate D20 0.0 0.5 1.0 CX2-CX1->C4-C7,  
Network53 bmse000257 FMN D20 0.0 1.0 1.0 CX2-CX1->C18-C16,  
Network54 bmse000069 betaine D20 0.8 1.0 1.0 CX1-CX2->C4-C3,  
Network54 bmse000948 betaine D20 0.8 1.0 1.0 CX1-CX2->C4-C3,  
Network55 bmse000208 L\_lactic\_acid D20 0.0 0.5 1.0 CX2-CX1->C1-C2,  
Network55 bmse000269 R\_Lactate D20 0.0 0.5 1.0 CX2-CX1->C6-C4,  
Network55 bmse000979 L\_lactic\_acid D20 0.0 0.5 1.0 CX2-CX1->C1-C2,  
Network56 bmse000008 D\_allose D20 0.667 0.2 0.75 CX4-CX2->C9-C9,CX4-CX1->C9-C7, CX2-CX3->C9-?,  
Network56 bmse000013 D\_galactose D20 1.0 0.4 0.75 CX4-CX2->C2-C5,CX4-CX1->C2-C1, CX2-CX3->C5-?,  
Network56 bmse000015 D\_glucose D20 1.0 0.2 0.75 CX4-CX1->C2-C1, CX4-CX2->C2-?,CX2-CX3->?-?,  
Network56 bmse000018 D\_mannose D20 1.0 0.4 0.75 CX4-CX2->C3-C6,CX4-CX1->C3-C2, CX2-CX3->C6-?,  
Network56 bmse000026 D\_xylose D20 1.0 0.25 0.75 CX4-CX1->C2-C1, CX4-CX2->C2-?,CX2-CX3->?-?,  
Network56 bmse000086 alpha\_D\_glucose\_1\_phosphate D20 0.0 0.2 0.75 CX4-CX2->C2-C3,CX2-CX3->C3-C3, CX4-CX1->C2-?,  
Network56 bmse000125 D\_trehalose D20 0.833 0.2 0.75 CX4-CX2->C14-C5,CX4-CX1->C14-C13, CX2-CX3->C5-?,  
Network56 bmse000233 melibiose D20 0.0 0.2 0.5 CX2-CX3->C20-C17, CX4-CX2->?-?,CX4-CX1->?-?,  
Network56 bmse000239 D\_saccharate D20 0.0 0.2 0.5 CX2-CX3->C8-C6, CX4-CX2->?-?,CX4-CX1->?-?,  
Network56 bmse000855 D\_glucose D20 1.0 0.2 0.75 CX4-CX1->C2-C1, CX4-CX2->C2-?,CX2-CX3->?-?,  
Network56 bmse000876 D\_trehalose D20 0.833 0.2 0.75 CX4-CX2->C14-C5,CX4-CX1->C14-C13, CX2-CX3->C5-?,  
Network56 bmse000898 D\_xylose D20 1.0 0.25 0.75 CX4-CX1->C2-

```

C1,      CX4-CX2->C2-?,CX2-CX3->?-?,
Network56  bmse001006 D_galactose      D2O      1.0      0.4      0.75     CX4-CX2-
>C2-C5,CX4-CX1->C2-C1, CX2-CX3->C5-?,
Network56  bmse001008 D_allose      D2O      0.667    0.2      0.75     CX4-CX2->C9-
C9,CX4-CX1->C9-C7, CX2-CX3->C9-?,
Network57  bmse000140 D_glucuronate D2O      0.0      1.0      1.0      CX3-CX4-
>C2-C2,CX1-CX3->C1-C2,CX2-CX4->C3-C2,
Network57  bmse000313 beta_gentiobiose D2O      0.0      0.2      1.0      CX3-
CX4->C20-C20,CX1-CX3->C18-C20,CX2-CX4->C22-C20,
Network59  bmse000346 Barbituric_acid D2O      0.0      0.5      1.0      CX1-
CX2->C6-C8,
Network63  bmse000039 L_histidine    D2O      0.333    0.25     1.0      CX1-CX2-
>C1-C4,
Network63  bmse000077 coumarin      CDC13    0.0      0.25     1.0      CX1-CX2->C4-
C2,
Network63  bmse000246 L_carnosine    D2O      0.0      0.25     1.0      CX1-CX2-
>C1-C6,
Network63  bmse000448 L_Histidinol  D2O      0.0      0.25     1.0      CX1-CX2-
>C9-C7,
Network63  bmse000707 N_acetyl_histidine D2O      0.0      0.2      1.0      CX1-
CX2->C11-C9,
Network63  bmse000744 histamine     D2O      0.0      0.333    1.0      CX1-CX2->C7-
C5,
Network63  bmse000762 neostigmine    D2O      0.167    0.25     1.0      CX1-CX2-
>C11-C13,
Network65  bmse000281 Nicotinamide   D2O      0.0      0.2      1.0      CX1-CX2-
>C9-C7,
Network65  bmse000622 3_pyridinecarbonitrile D2O      0.0      0.2      1.0
CX1-CX2->C5-C4,
Network66  bmse000432 Pyridine      D2O      0.0      0.25     1.0      CX1-CX2->C6-
C4,

```

### Example of MatchJres in PyINETA

```
Network1
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/pyINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (27.03,40.91):1129341600000.0;CX2[Present] (13.72,41.01):688701440000.0;"

Network2
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/pyINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (38.58,56.17):772994430000.0;CX2[Present] (17.27,55.93):827546400000.0;"

Network3
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/pyINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (18.7,72.18):2646701000000.0;CX2[Present] (53.2,71.98):1644530800000.0;"

Network4
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/pyINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (31.73,51.09):1181092500000.0;CX2[Present] (19.2,50.97):1298547300000.0;"

Network5
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/pyINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (31.82,52.36):1180879700000.0;CX2[Present] (20.57,52.45):1632450800000.0;"

Network6
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/pyINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (68.52,90.83):1235223000000.0;CX2[Present] (22.24,90.78):832188700000.0;"

Network7
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/pyINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (22.87,94.29):5607700600000.0;CX3[Present] (70.98,94.12):2692579000000.0;"

Network8
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/pyINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (24.7,51.54):1726148600000.0;CX2[Present] (42.71,69.7):1706099900000.0;CX4[Present] (26.95,69.7):1141805800000.0;CX5[Present] (64.76,107.56):1144215200000.0;CX8[Present] (23.43,50.35):1606211000000.0;"

Network9
```



```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (24.13,53.52):815090300000.0
;CX2[Present] (29.14,53.52):447770000000.0;"
```

Network10

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (32.59,57.04):1074266700000.
0;CX2[Present] (24.3,56.98):834897440000.0;"
```

Network11

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (24.64,201.46):1738039100000
.0;CX2[Present] (176.75,201.46):912965600000.0;"
```

Network12

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (25.69,67.17):993319650000.0
;CX2[Present] (41.51,67.17):1120700700000.0;"
```

Network13

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (183.03,208.87):779128500000
.0;CX2[Present] (25.8,209.03):980739600000.0;"
```

Network14

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (26.42,86.41):766211300000.0
;CX2[Present] (60.42,86.62):740303960000.0;"
```

Network15

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (48.6,75.22):741234100000.0;
CX2[Present] (26.53,75.15):762627160000.0;"
```

Network16

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (182.24,243.22):820035850000
.0;CX2[Present] (28.07,89.06):1381270400000.0;CX3[Present] (60.97,243.13):10
34876350000.0;"
```

Network17

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (28.95,85.85):4472378500000.
0;CX2[Present] (56.68,85.82):7125908600000.0;"
```

Network18

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (29.03,70.84):4478046600000.
```

```
0;CX2[Present] (33.61,213.39):2013865600000.0;CX4[Present] (179.7,213.2):931228200000.0;CX6[Present] (41.66,71.0):1278682200000.0;"
```

Network19

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/pyINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (176.46,233.69):876906100000.0;CX2[Present] (29.51,86.95):1781145800000.0;CX4[Present] (57.33,86.94):1294262500000.0;"
```

Network20

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/pyINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (32.53,64.2):1074889560000.0;"
```

Network21

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/pyINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (32.4,216.14):997792700000.0;CX2[Present] (183.72,216.19):773462160000.0;"
```

Network22

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/pyINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (32.76,89.8):863568660000.0;CX2[Present] (57.01,89.9):3025168000000.0;"
```

Network23

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/pyINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (176.93,210.88):917193600000.0;CX2[Present] (34.1,210.98):1200629400000.0;"
```

Network24

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/pyINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (39.26,75.41):1042545840000.0;CX2[Present] (40.64,80.07):698567500000.0;CX4[Present] (35.96,75.41):1009721000000.0;"
```

Network25

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/pyINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (57.67,93.92):1002780160000.0;CX2[Present] (36.09,94.03):1026640250000.0;"
```

Network26

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/pyINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (183.33,219.95):736758100000.0;CX2[Present] (36.39,220.0):1143852100000.0;"
```

Network27

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p  
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (36.72,221.34):2123371900000  
.0;CX2[Present] (184.84,221.45):806637000000.0;"
```

Network28

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p  
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (52.6,90.24):1226222500000.0  
;CX2[Present] (37.61,90.11):831301160000.0;"
```

Network29

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p  
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (37.96,214.07):9147714000000  
.0;CX2[Present] (176.08,214.05):1828480900000.0;CX4[Present] (62.97,239.03):  
1783091400000.0;"
```

Network30

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p  
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (49.98,88.1):11130542000000.  
0;CX2[Present] (38.15,88.13):9924519000000.0;"
```

Network31

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p  
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (41.77,97.25):1237302600000.  
0;CX2[Present] (55.36,97.25):804044200000.0;"
```

Network32

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p  
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (56.02,98.68):1177642100000.  
0;CX2[Present] (42.53,98.68):1676964700000.0;"
```

Network33

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p  
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (43.35,106.54):991751600000.  
0;CX2[Present] (63.12,106.42):1745454000000.0;"
```

Network34

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p  
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (53.69,97.62):763946140000.0  
;CX2[Present] (43.76,97.62):797534300000.0;"
```

Network35

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p  
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (44.0,218.17):2609389000000.  
0;CX3[Present] (174.37,218.7):938311750000.0;CX6[Present] (59.26,233.4):8884  
47960000.0;"
```

Network36

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (60.23,104.39):729235260000.
0;CX2[Present] (44.13,104.39):262513470000.0;"
```

Network37

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (180.62,225.68):736136270000
.0;CX2[Present] (44.86,225.79):824767150000.0;"
```

Network38

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (178.15,223.78):181460910000
0.0;CX3[Present] (46.0,223.94):4829206300000.0;"
```

Network39

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (46.86,115.12):1034964760000
.0;CX2[Present] (49.38,96.25):714900200000.0;CX4[Present] (68.23,115.23):722
591150000.0;"
```

Network40

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (52.76,114.37):1196998100000
.0;CX2[Present] (61.89,114.52):1095730600000.0;"
```

Network41

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (53.12,230.86):1627207500000
.0;CX2[Present] (177.71,230.86):915893500000.0;"
```

Network42

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (55.28,229.4):813073200000.0
;CX2[Present] (173.87,229.56):1173121000000.0;"
```

Network43

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (177.5,233.48):787060900000.
0;CX2[Present] (55.83,233.72):1235917300000.0;"
```

Network44

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (58.28,128.55):702064950000.
0;CX2[Present] (70.17,128.58):787518400000.0;"
```

Network45

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p  
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (60.68,122.31):772474140000.  
0;"
```

Network46

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p  
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (68.97,130.51):958458230000.  
0;CX2[Present] (62.07,130.8):1200856200000.0;CX3[Present] (61.04,130.4):1019  
011200000.0;"
```

Network47

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p  
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (175.75,238.53):780957060000  
.0;CX2[Present] (63.04,238.26):1752197800000.0;"
```

Network48

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p  
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (74.1,146.86):2497356800000.  
0;CX2[Present] (72.59,146.83):4374824700000.0;CX4[Present] (63.49,137.64):36  
40010000000.0;"
```

Network49

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p  
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (63.56,142.6):3657581700000.  
0;CX2[Present] (78.81,142.35):5037370000000.0;"
```

Network50

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p  
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (73.62,138.44):806795100000.  
0;CX2[Present] (64.94,138.43):1153838100000.0;"
```

Network51

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p  
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (65.74,243.31):759493750000.  
0;CX2[Present] (177.61,243.39):912987100000.0;"
```

Network52

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p  
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (85.82,153.46):718851300000.  
0;CX2[Present] (67.59,153.56):8085820500000.0;"
```

Network53

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p  
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (73.8,142.03):982890840000.0  
;CX2[Present] (67.95,141.93):9694922700000.0;"
```

Network54

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p  
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (171.19,239.93):72217240000  
.0;CX2[Present] (68.8,239.96):978746540000.0;"
```

Network55

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p  
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (71.04,255.69):2677792300000  
.0;CX2[Present] (184.53,255.64):1155736800000.0;"
```

Network56

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p  
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (94.87,169.46):2254432000000  
.0;CX2[Present] (74.43,150.18):1827269200000.0;CX3[Present] (75.56,150.18):2  
142712600000.0;"
```

Network57

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p  
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (78.72,155.7):5029263600000.  
0;CX2[Present] (98.79,175.98):3195300700000.0;CX3[Present] (76.99,155.7):250  
6849500000.0;"
```

Network58

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p  
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (77.39,167.15):853489000000.  
0;CX2[Present] (89.73,167.02):741308200000.0;"
```

Network59

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p  
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (79.69,251.91):721369000000.  
0;CX2[Present] (172.08,251.96):731503600000.0;"
```

Network60

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p  
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (85.51,177.14):669664540000.  
0;CX2[Present] (91.48,177.14):719250200000.0;"
```

Network61

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p  
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (168.57,273.8):726629500000.  
0;CX2[Present] (105.15,273.37):786833700000.0;"
```

Network62

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p  
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (126.48,244.29):787265360000  
.0;CX2[Present] (117.68,244.4):738509000000.0;"
```

Network63

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p  
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (119.39,254.13):741678700000  
.0;CX2[Present] (134.66,254.13):770011300000.0;"
```

Network64

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p  
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (121.12,272.98):734866500000  
.0;CX2[Present] (151.69,272.98):850254230000.0;"
```

Network65

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p  
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (126.82,281.2):1024876700000  
.0;CX2[Present] (154.51,281.4):980925200000.0;"
```

Network66

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p  
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (151.08,278.03):750467500000  
.0;CX2[Present] (126.96,278.13):1001294860000.0;"
```

Network67

```
"/Users/rtaujale/Dropbox/Projects/collaborative/13C_mouse_INETA/raw_data/p  
yINETA_AlaRef_2023/3.tilt.sym.ft2=CX1[Present] (178.26,363.89):185129360000  
0.0;CX2[Present] (186.07,363.89):793620840000.0;"
```

### *Evaluation of the consistency of annotation between the original INETA and PyINETA*

We evaluated the performance of our new Python pipeline. For this evaluation, we used the same INADEQUATE spectra that were used in the original study of INETA<sup>5</sup> as input files for PyINETA. They are INADEQUATE spectra collected for endo- and exometabolites of <sup>13</sup>C-labeled *Caenorhabditis elegans*. The results were consistent (Supplementary Table 7); out of 29 metabolites annotated in the previous study, 26 were found in the new pipeline. They covered a range of compounds, including amino acids, an amino acid derivative (creatine), organic acids, a fatty acid (palmitic acid), a polyamine (putrescine), a pyrimidine derivative (uracil), and sugar derivatives. Three of them that were not included in our results are *N*-acetyl glycine, stearic acid, and glutathione oxidized.



Supplementary Table S1. List of adjustable parameters in PyINETA. Those parameters are contained in each configuration file.

Parameter	Function	Value used for the mouse study	Note
<b><i>For INADEQUATE</i></b>			
<i>Peak picking</i>			
PPmin	Peak intensity minimum threshold	5.7e10	Clendinen et al. (2015)
PPmax	Peak intensity minimum threshold	4e11	Clendinen et al. (2015)
steps	Number of iterations	10	This study
PPCS	Chemical shift threshold for clustering	1	Clendinen et al. (2015)
PPDQ	Double quantum threshold for clustering	2	Clendinen et al. (2015)
<i>Network finding</i>			
DQT	Double quantum tolerance	0.2 ppm	Clendinen et al. (2015)
SumXY	Sum tolerance	2	This study
SDT	Symmetrical/diagonal tolerance	0.5 ppm	Clendinen et al. (2015)
CST	Chemical shift tolerance for vertical connection	0.05 ppm	Clendinen et al. (2015)
<i>Database matching</i>			
Ambiguity	Ambiguity allowance	1	Clendinen et al. (2015)
CSMT	Chemical shift match tolerance	1 ppm	Clendinen et al. (2015)
NCMT	Peak match tolerance	2	Clendinen et al. (2015)
DQMT	Double quantum threshold	4	Clendinen et al. (2015)
<b><i>For JRES</i></b>			
Peak_Width_1D	JRES peak width to be used to calculate peak area	0.5 ppm	This study
Intensity_threshold_1D	Intensity threshold to define presence or absence of peak	10,000	This study

Supplementary Table S2. Composition of the diet that was fed to the three mice in this study. For details on the composition of bacterial <sup>13</sup>C protein hydrolysate, see Supplementary Table S3.

<b>Component</b>	<b>g/kg</b>
Sucrose	350.0
Bacterial <sup>13</sup> C protein hydrolysate	200.0
Maltodextrin	130.0
Corn Starch	100.0
Soybean Oil	80.0
Cellulose	72.2
Mineral Mix, AIN-93M-MX (94049)	45.0
Vitamin Mix, AIN-93-VX (94047)	14.0
L-Cystine	3.0
Calcium Phosphate, dibasic	3.0
Choline Bitartrate	2.5
Ferric Citrate	0.25
THBQ, antioxidant	0.024

Supplementary Table S3. Amino acid composition of the hydrolysate that was fed to the three mice in this study (See also for Supplementary Table S2). The data are from a product sheet from Silantes. \*: essential amino acid

<b>Amino acid</b>	<b>Percent</b>
Asp	20.56
Thr*	3.70
Ser	4.10
Glu	11.10
Gly	11.10
Ala	14.8
Val*	2.98
Met*	1.78
Ileu*	2.10
Leu*	5.26
Tyr	1.80
Phe*	2.40
His*	9.65
Lys*	3.38
Arg	2.30
Pro	2.99

Supplementary Table S4. Amount of the samples used for the mouse samples in this study. The weight is based on dry weight.

<b>Tissue</b>	<b>Mouse ID</b>	<b>Weight or volume</b>
Liver	1	1092 mg
	2	998.9 mg
	3	1000 mg
Adrenal Gland	1	32.5 mg
	2	22.3 mg
	3	36.6 mg
Lung – Only 75% of lung tissue	1	203.1 mg
	2	353.9 mg
	3	171.2 mg
Muscle – Only 50% of muscle tissue	1	121.1 mg
	2	137.6 mg
	3	82.7 mg
Pancreas	1	162.4 mg
	2	152.6 mg
	3	165.9 mg
Plasma	1	50 $\mu$ L
	2	50 $\mu$ L
	3	50 $\mu$ L
Brain	1	226.5 mg
	2	204.3 mg
	3	226.5 mg
Spleen	1	155.7 mg
	2	154.8 mg
	3	184.3 mg
Thymus	1	59.83 mg
	2	105.6 mg
	3	68.3 mg

Supplementary Table S5. Parameters used for NMR experiments. SW, spectral width; TD, size of FID; NS, number of scans.

Figure	Experiment	Pulse program	SW (ppm)		TD		NS	Note
			F2	F1	F2	F1		
2	INADEAUATE	inadphppsp	214	433	4096	2048	16	
2, 3	JRES	jresdcqf	214	0.66	32768	50	64	The original uniform 180° pulse was replaced with an adiabatic pulse

Supplementary Table S6. Summary of NMRPipe processing parameters used for INADEQUATE and JRES experiments. SP: adjustable sine window function, ZF: zero filling. Original NMRPipe scripts are also available in Metabolomics Workbench.

Figure	Experiment	SP F2 and F1	ZF F2 and F1	Baseline correction F2 and F1	Note
2	INADEQUATE	-off 0.0 -end 0.95 -pow 2	-auto	MED	–
2, 3	JRES	-off 0.0 -end 0.98 -pow 2	-auto	–	Magnitude calculation (MC) was additionally used; spectra were further tilted and symmetrized

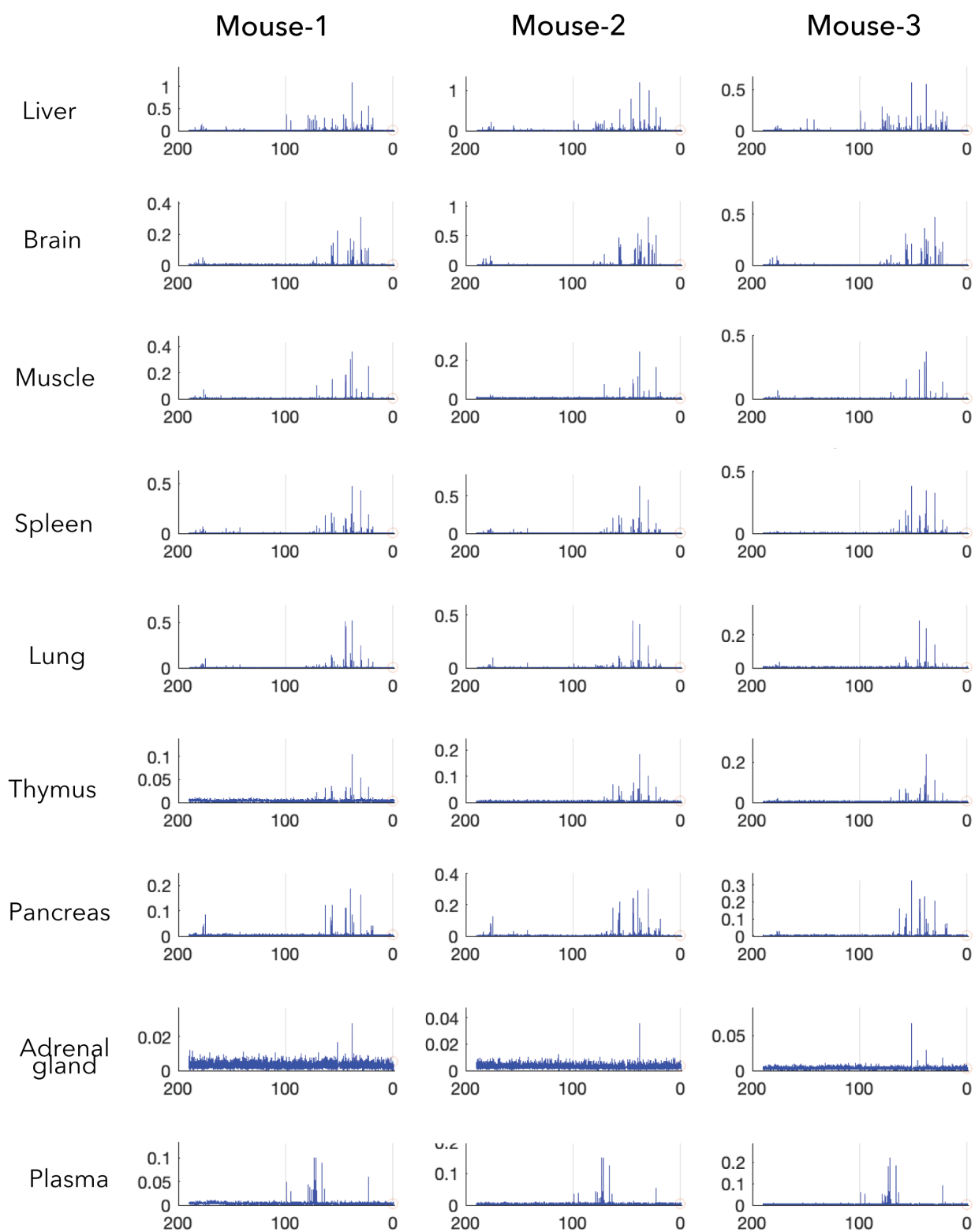
Supplementary Table S7. List of metabolites found by PyINETA for endo- and exometabolites from *Caenorhabditis elegans*. See also Experimental Section for the details on the confidence scores.

Compound name	Ambiguity score	Hit score	Coverage Score	Type
<b><i>Endometabolites</i></b>				
Proline	0	0.25	1	Found in both the previous and this studies
Valeric acid	0	0.25	1	Found in both the previous and this studies
Glucose 1,6 bisphosphate	0	0.2	1	Found in both the previous and this studies
Histidine	0.333	0.25	1	Found in both the previous and this studies
Threonine	0	0.333	1	Found in both the previous and this studies
Arabitol	0	0.25	1	Found in both the previous and this studies
Alanine	0	0.5	1	Found in both the previous and this studies
Isoleucine	0	0.2	1	Found in both the previous and this studies
Propionic acid	0	0.5	1	Found in both the previous and this studies
3-hydroxybutyrate	0	0.333	1	Found in both the previous and this studies
Lactic acid	0	0.5	1	Found in both the previous and this studies
Valine	0.4	0.25	1	Found in both the previous and this studies
Palmitic acid	0	0.2	0.625	Found in both the previous and this studies
<b><i>Exometabolites</i></b>				
Isoleucine	0	0.2	1	Found in both the previous and this studies
Alanine	0	0.5	1	Found in both the previous and this studies
Threonine	0	0.333	1	Found in both the previous and this studies
Glycine	0	1	1	Found in both the previous and this studies
Proline	0	0.25	1	Found in both the previous and this studies
Ornithine	0	0.25	1	Found in both the previous and this studies
Serine	0	0.5	1	Found in both the previous and this studies

Glucose 1,6 bisphosphate	0	0.2	1	Found in both the previous and this studies
Lysine	0	0.2	1	Found in both the previous and this studies
Glutamine	0	0.25	1	Found in both the previous and this studies
Lactic acid	0	0.5	1	Found in both the previous and this studies
Glucoronate	0	0.5	0.75	Found in both the previous and this studies
Putrescine	0	0.333	1	Found in both the previous and this studies
Acetic acid	0	1	1	Found in both the previous and this studies
Arabitol	0	0.25	1	Found in both the previous and this studies
Creatine	0	1	1	Found in both the previous and this studies
Valine	0.4	0.25	1	Found in both the previous and this studies
Allantoin	0	1	0.667	Found in both the previous and this studies
Propionic acid	0	0.5	1	Found in both the previous and this studies
Succinic acid	0	0.333	1	Found in both the previous and this studies
Methionine	0	0.333	0.5	Found in both the previous and this studies
Uracil	0	0.5	1	Found in both the previous and this studies

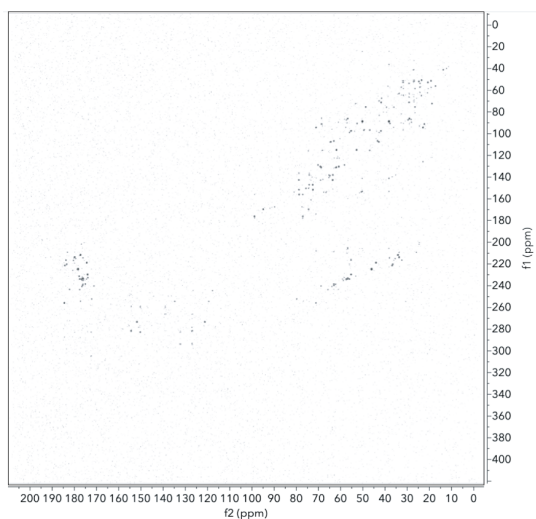
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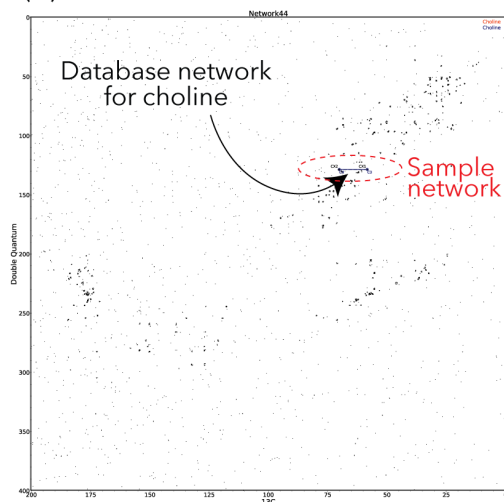


Supplementary Figure S1 JRES projection spectra for the mouse tissues. Solvent MeOH peaks around 50 ppm are removed and not shown.

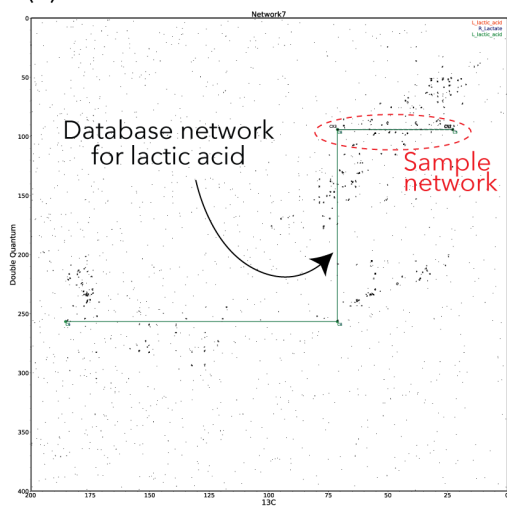
(a) Input INADEQUATE



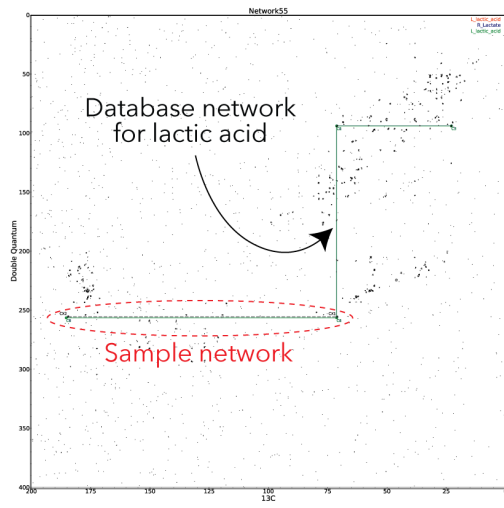
(b) Network 44



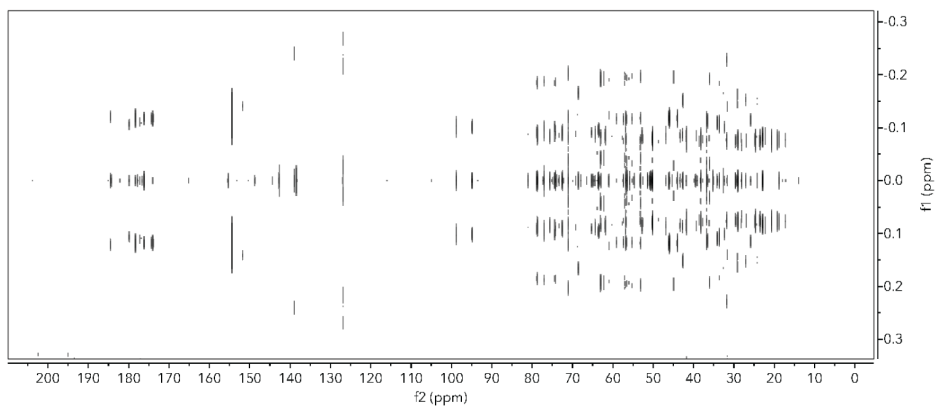
(c) Network 7



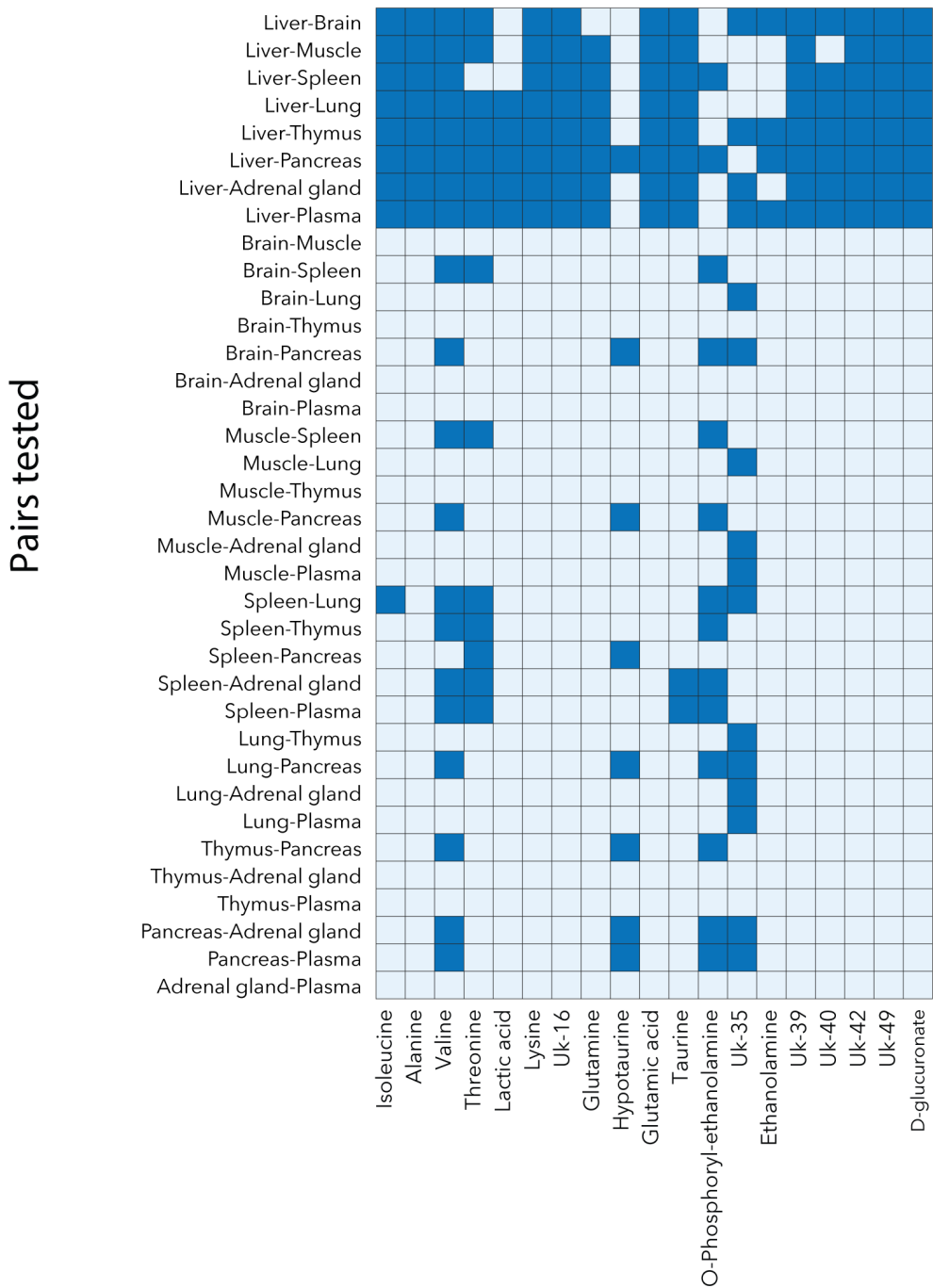
(d) Network 55



(e) Input JRES

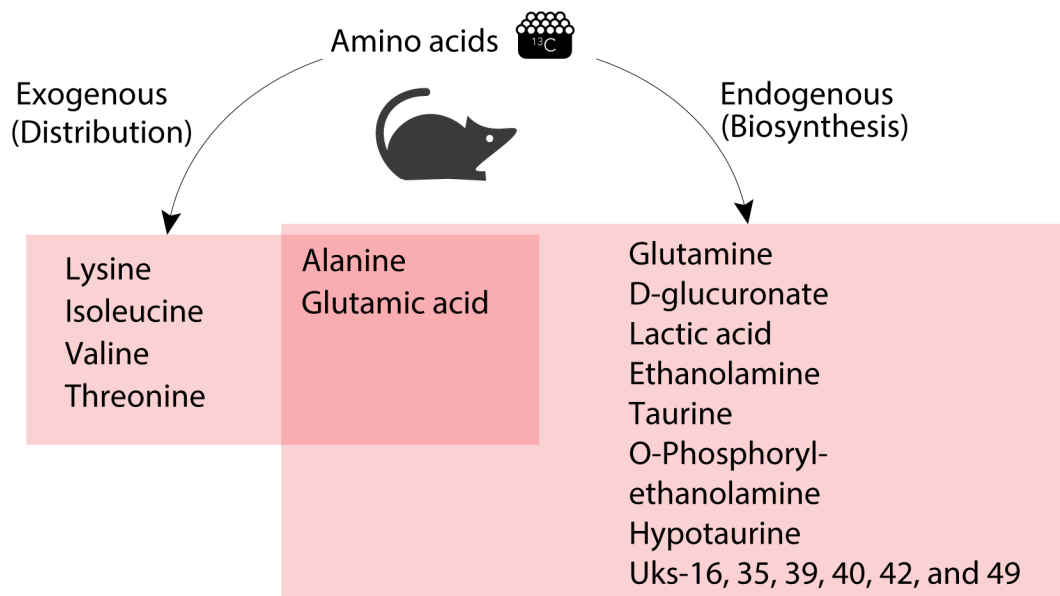


Supplementary Figure S2 (a) Input INADEQUATE spectrum for the liver sample; (b) Network 44 of PyINETA; (c) Network 7; (d) Network 55; (e) Input JRES spectrum for the liver sample. For a and e, the plots were produced by MestReNova v14.3.0. For b, c, and d, the plots are the original outputs from PyINETA, with a slight graphical modification.



### Metabolites

Supplementary Figure S3 Results of ANOVA and multiple comparison. Metabolites intensities were compared between tissues for the three mice. When there is a significant difference between tissues, they are highlighted in dark blue ( $p < 0.05$  with Bonfferoni Correction). This information is summarized in Figure 3.



Supplementary Figure 4 Possible sources of metabolites in this study

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1. Uchimiya, M.; Olofsson, M.; Powers, M. A.; Hopkinson, B. M.; Moran, M. A.; Edison, A. S.,  $^{13}\text{C}$  NMR metabolomics: J-resolved STOCSY meets INADEQUATE. *J Magn Reson* **2023**, *347*, 107365.
2. Delaglio, F.; Grzesiek, S.; Vuister, G. W.; Zhu, G.; Pfeifer, J.; Bax, A., NMRPipe - A multidimensional spectral processing system based on Unix pipes. *J Biomol Nmr* **1995**, *6* (3), 277-293.
3. Dieterle, F.; Ross, A.; Schlotterbeck, G.; Senn, H., Probabilistic quotient normalization as robust method to account for dilution of complex biological mixtures. Application in H-1 NMR metabonomics. *Anal Chem* **2006**, *78* (13), 4281-4290.
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