

Expanded View Figures

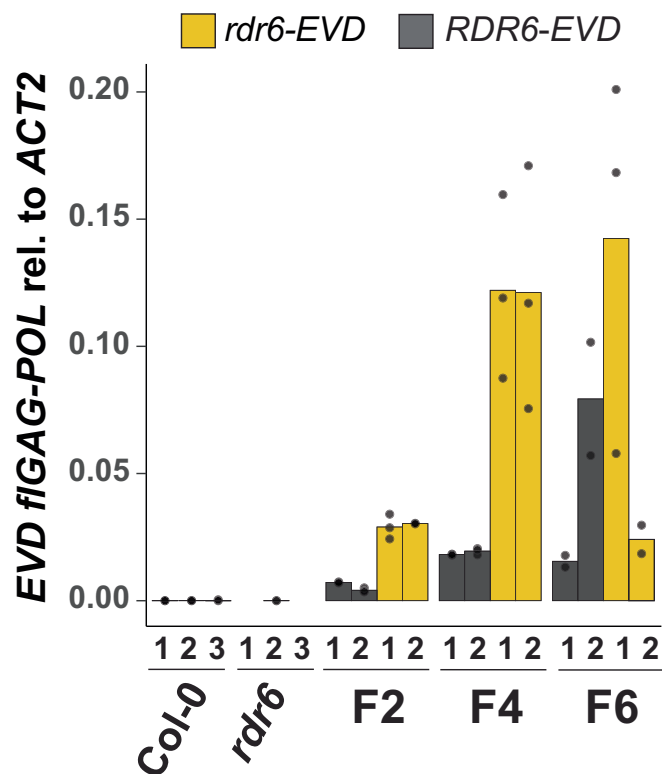


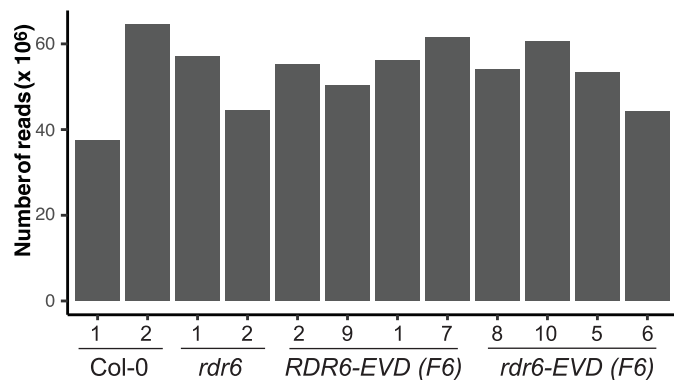
Figure EV1. EVD GAG-POL expression in RDR6 wild-type and mutant backgrounds.

(A) qPCR analysis of *flGAG-POL* expression normalized to *ACT2* in *EVD-RDR6* and *EVD-rdr6* lines at generations 2, 4, and 6 derived from two independent F1s (biological replicates). Each biological replicate, consistent of bulks of 8-10 plants, are represented for each genotype at each generation, dots show technical replicates.

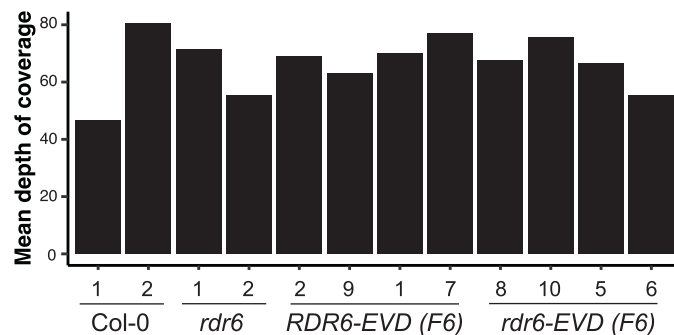
A

sample	conversion rate
Col-0 #1	99,857 %
Col-0 #2	99,866 %
<i>rdr6-15</i> #1	99,847 %
<i>rdr6-15</i> #2	99,861 %
<i>RDR6-EVD</i> #2	99,850 %
<i>RDR6-EVD</i> #9	99,870 %
<i>RDR6-EVD</i> #1	99,878 %
<i>RDR6-EVD</i> #7	99,862 %
<i>rdr6-EVD</i> #8	99,864 %
<i>rdr6-EVD</i> #10	99,830 %
<i>rdr6-EVD</i> #5	99,880 %
<i>rdr6-EVD</i> #6	99,865 %

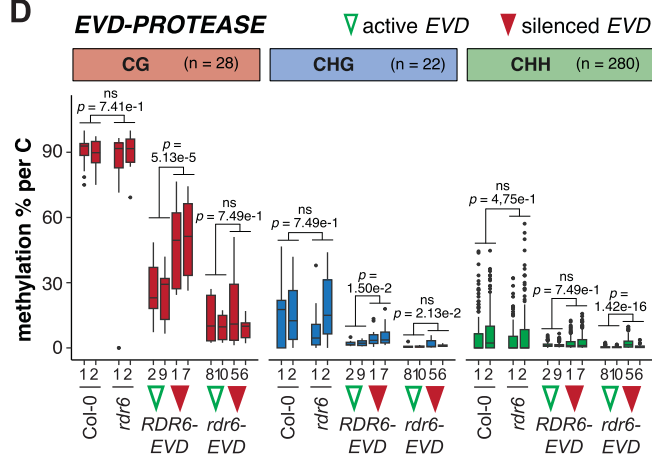
B



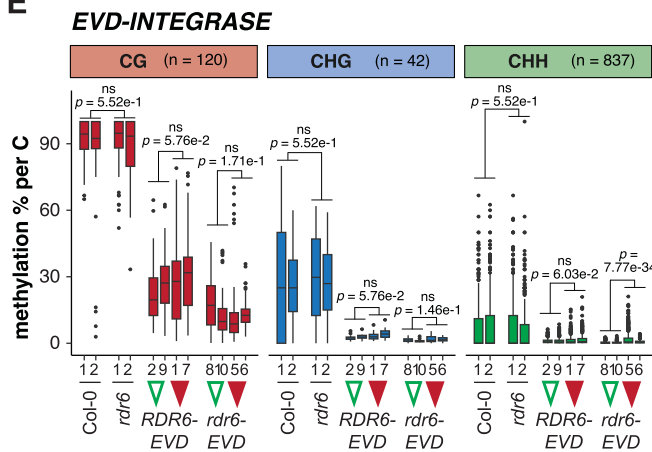
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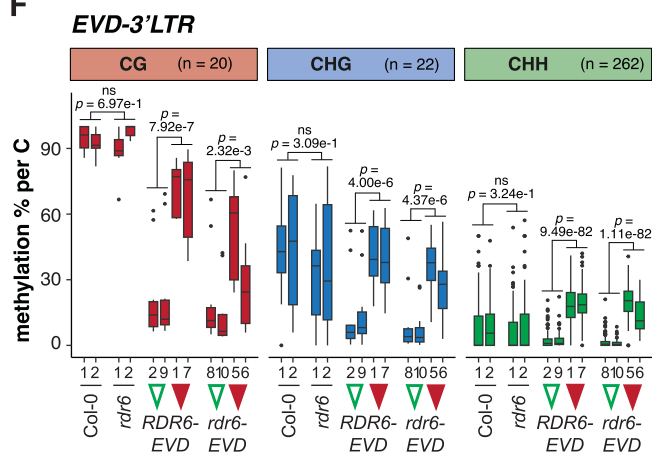
D



E

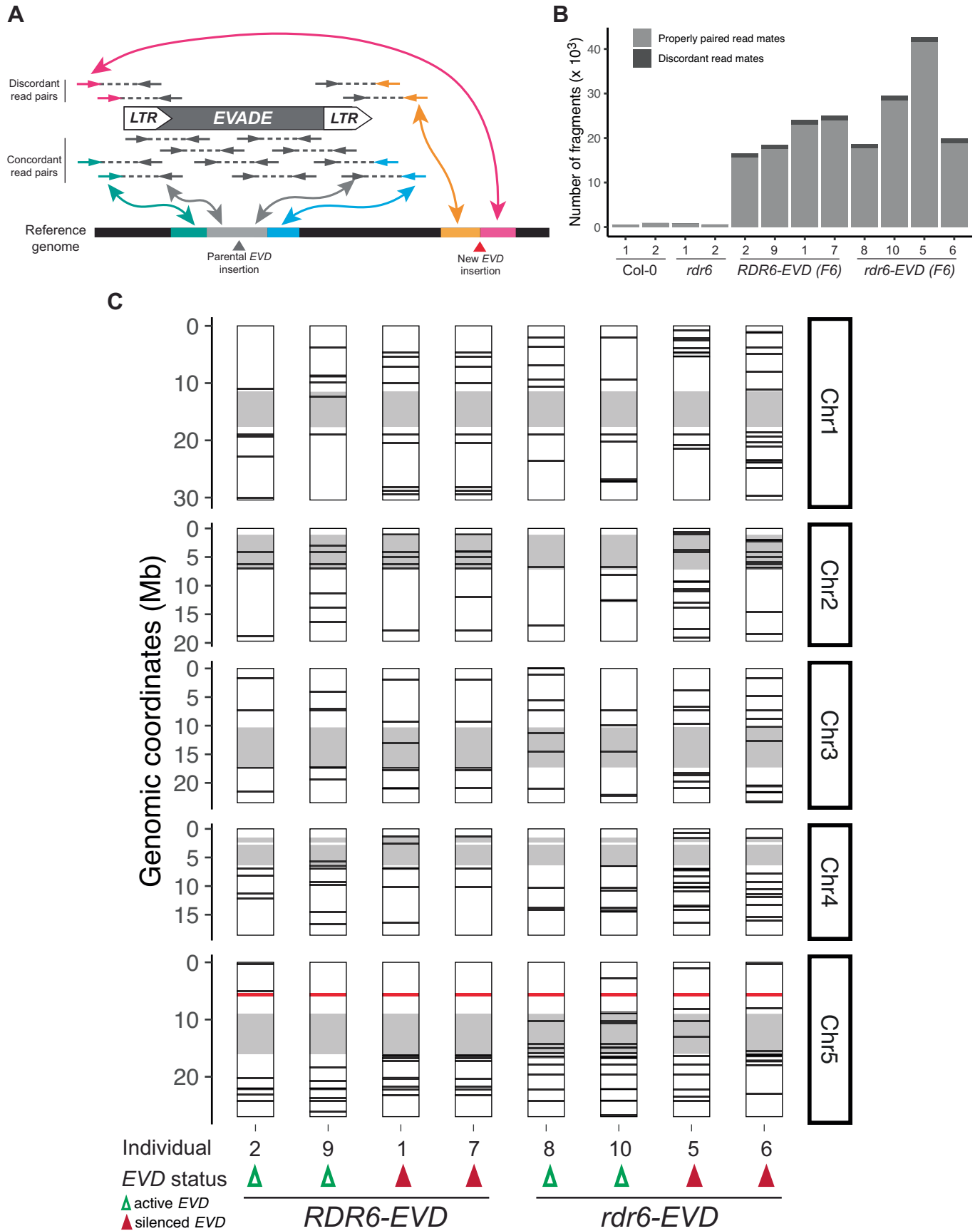


F



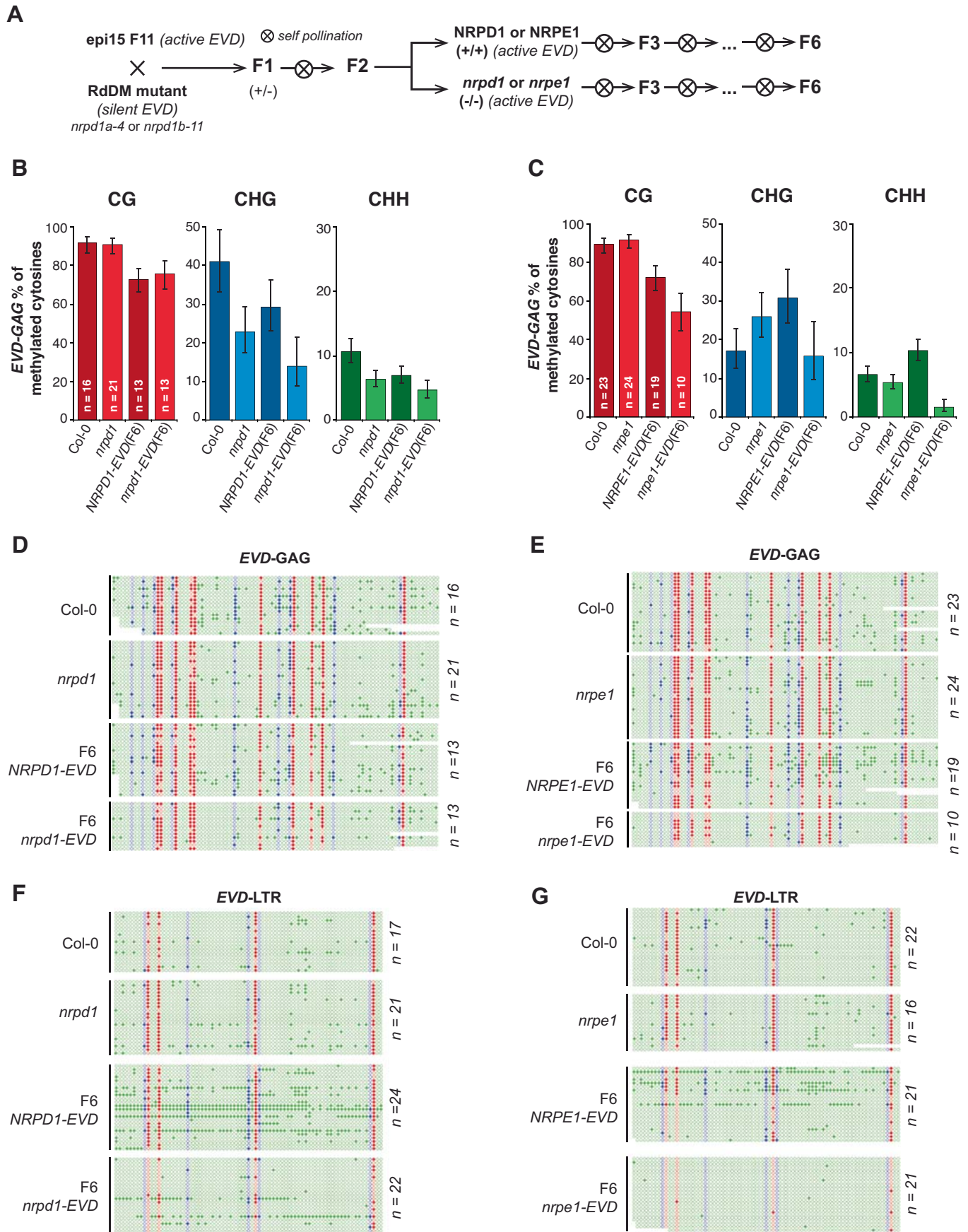
◀ Figure EV2. EM-seq libraries general stats and EVD POL and 3'LTR methylation in RDR6- and rdr6-EVD lines.

(A) Cytosine-to-thymine conversion rates of the unmethylated chloroplastic DNA for each EM-seq library (see materials and methods for further information). (B) Number of paired-end fragments obtained in each EM-seq library. (C) *Arabidopsis* genome coverage in each EM-seq library. (D-F) EM-seq analysis of DNA methylation (as % per of methylated cytosines) in CG, CHG, and CHH contexts in WT Col-0, *rdr6* and in *RDR6*- and *rdr6-EVD* F6 individuals with active and silenced *EVD* (marked with empty green and filled red arrowheads, respectively, numbers indicate same individuals as in Fig. 2) in: (D) *EVD-Protease*; (E) *EVD-Integrase*, and (F) *EVD-5'LTR*. In panels (D-F), *n* indicates the number of cytosines analyzed for each context per sample. In all boxplots: median is indicated by a solid bar, the boxes extend from the first to the third quartile and whiskers reach to the furthest values within 1.5 times the interquartile range. Dots indicate outliers, as data points outside of the above range. Wilcoxon rank sum test adjusted *p*-values between indicated groups of samples are shown. Differences are considered statistically significant if $p < 0.05$ ($5.00e-2$) or non-significant (ns) if $p \geq 0.05$.



◀ Figure EV3. Mapping of new EVD insertions in RDR6- and rdr6-EVD lines from EM-seq data.

(A) Schematic representation of the strategy used to map new EVD insertions using discordant read mates from EM-seq. (B) Number of fragments from concordant (properly paired) and discordant read mates mapping to EVD in each of the EM-seq libraries. (C) Genomic location of new EVD insertions mapped through discordant read pair mates in EM-seq data in the *Arabidopsis* genome. Parental EVD (AT5G17125) location is indicated with a red line. New EVD insertions are marked with black lines. Pericentromeric regions in each of *Arabidopsis* five chromosomes are marked in gray. See table provided in corresponding source data for precise chromosome coordinates of each new insertion.



◀ Figure EV4. BS-PCR analysis of EVD-GAG DNA methylation levels in RdDM mutants.

(A) Crossing scheme to generate *nripd1*- and *nripe1*-EVD lines. F2 plants were genotyped to select homozygous WT and mutant lines for each background and propagated through selfing until the F6 generation. (B, C) % methylated cytosines by bisulfite-PCR DNA methylation analysis at EVD-GAG sequences in the F6 generation of *NRPD1*-EVD (B) and *NRPE1*-EVD (C) lines, in both WT (darker shade) and mutant (lighter shade) backgrounds. Col-0, *nripd1* and *nripe1* were used as controls. Error bars represent 95% confidence Wilson score intervals of the % of methylated cytosines (C) in each context (CG, CHG, CHH). (D, E) Dot-plot representation of bisulfite-PCR sequencing data for the F6 generation of *NRPD1*-EVD (D) and *NRPE1*-EVD (E) lines, in both WT and mutant backgrounds, at EVD-GAG sequences. Col-0 and *nripd1* or *nripe1* were used as control for the endogenous parental EVD copies. (F, G) Dot-plot representation of bisulfite-PCR sequencing data for the F6 generation of *NRPD1*-EVD (F) and *NRPE1*-EVD (G) lines, in both WT and mutant backgrounds, at EVD-LTR sequences. Col-0 and *nripd1* or *nripe1* were used as control for the endogenous parental EVD copies. In all dot-plots, filled circle represent methylated, empty circles unmethylated cytosines in the CG (red), CHG (blue), and CHH (green) context.

INSERTION ORIGIN		STRUCTURE	HAIRPIN	OCCURENCE							
				RDR6-EVD (F6)				rdr6-EVD (F6)			
Tandem	Nested			2	9	1	7	8	10	5	6
Tail-to-Tail 3'-to-3' LTR	in Antisense 5'-to-3' LTR		YES	N	N	N	N	N	N	N	N
Head-to-Head 5'-to-5' LTR	in Antisense 5'-to-3' LTR		YES	N	N	N	N	N	N	N	N
Head-to-Tail 3'-to-5' LTR	in Sense 5'-to-5' LTR 3'-to-3' LTR		NO	N	N	N	N	N	N	N	N

Figure EV5. Summary table for the search of close proximity sense-antisense EVD-LTR events.

Putative origin of LTR hairpins as consequence of EVD transposition in tandem or nested configurations next to the scheme of the strategy used to find them using discordant read mates from EM-seq where both read mates map to EVD-LTR. The table indicates the occurrence and number of read mates found in each sample. Arbitrary threshold of at least three paired read mates was set to confidentially identify two LTRs in close proximity. N: non-occurrence, y: presence of at least one discordant paired read mate, Y: presence of discordant paired read mates and above threshold for confident calling.