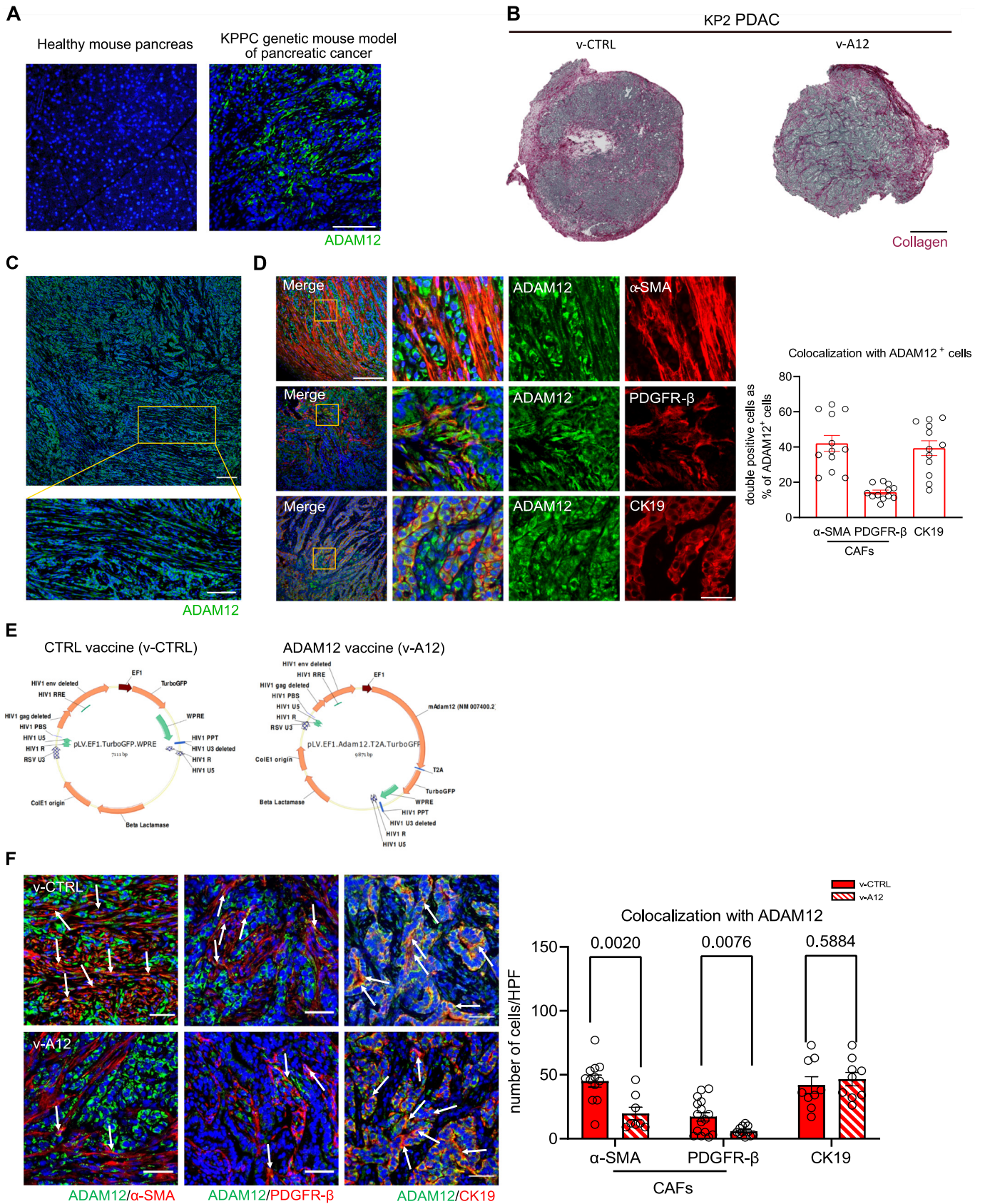
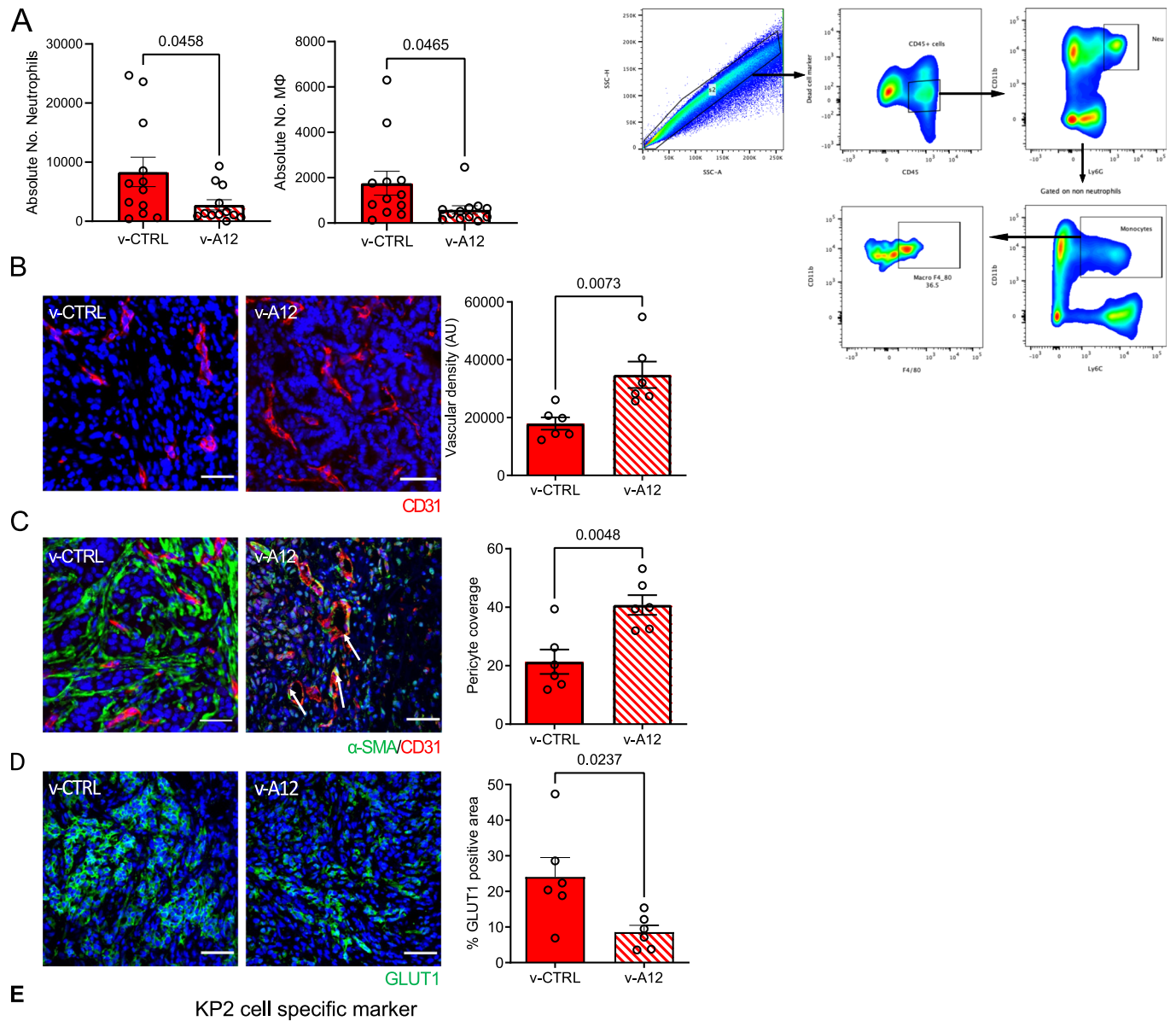


Expanded View Figures

Figure EV1. ADAM12 expression and characterization on KP2 PDAC tumor tissue.

(A) Representative image of ADAM12 immunofluorescence staining (ADAM12 antibody, Invitrogen, PA5-50594) on the healthy pancreas and PDAC tumor tissue from KPPC transgenic mice. (Scale bar 100 μm). (B) Representative images of Sirius Red/Fast Green staining on subcutaneous KP2 PDAC sections from v-CTRL mice and v-A12 mice. (Scale bar 100 μm). (C) Representative image of ADAM12 immunostaining (ADAM12 antibody, Invitrogen, PA5-50594) on PDAC tumor sections from subcutaneous KP2 PDAC grafts. (Scale bar 100 μm ; for the zoom-in picture, the scale bar is 50 μm). (D) Representative images (left) and quantitative analysis (right) of ADAM12⁺ cell characterization, ADAM12⁺ CAFs were indicated as ADAM12⁺/ α -SMA⁺ (upper, left) and ADAM12⁺/PDGFR- β ⁺ (middle, left), and ADAM12⁺ tumor cells were indicated as ADAM12⁺/CK19⁺ (bottom, left) co-expression on subcutaneous KP2 PDAC tumors. ($n = 12$ fields of view from four mice. Data were presented as mean \pm SEM. Scale bar is 200 μm , and the scale bar of the zoomed-in picture is 50 μm). (E) Vector maps of CTRL vaccine (v-CTRL) and ADAM12 vaccine (v-A12). (F) Quantitative analysis of ADAM12⁺/ α -SMA⁺ (left, left), ADAM12⁺/PDGFR- β ⁺ (left, middle), and ADAM12⁺/CK19⁺ (left, right) co-expression on PDAC tumors from control-vaccinated mice (v-CTRL) and ADAM12-vaccinated mice (v-A12). Data were represented as the number of cells per high power field (HPF). Representative double positive cells were indicated by white arrows respectively. ($n = 12-16$ in v-CTRL, $n = 8-9$ in v-A12. Data were presented as mean \pm SEM. Statistical test: multiple t -test. Scale bar 50 μm).





◀ Figure EV2. Prophylactic ADAM12 vaccination increased tumor vascular density and pericyte coverage.

(A) Quantitative analysis of the absolute number of neutrophils (left) and macrophages (middle) within PDAC tumor tissues from control-vaccinated mice (v-CTRL) and ADAM12-vaccinated mice (v-A12); corresponding flow cytometry gating strategy of myeloid cells (right): neutrophils were identified as CD45⁺CD11b⁺Ly6G⁺ cells; monocytes were gated based on non-neutrophils (negative gating), and then identified as CD11b⁺Ly6C⁺ population. Then, monocyte macrophages were further defined with an F4/80⁺ signal. (*n* = 12 mice in v-CTRL, *n* = 12 mice in v-A12. Data were presented as mean ± SEM. Statistical test: unpaired two-tailed Student's *t*-test). (B) Representative immunofluorescence staining images (left) and quantitative analysis of CD31-positive endothelial cells (right) on PDAC tumors from mice treated with either control vaccine (v-CTRL) or ADAM12 vaccine (v-A12). (*n* = 6 mice in v-CTRL, *n* = 6 mice in v-A12. Data were presented as mean ± SEM. Statistical test: unpaired two-tailed Student's *t*-test. Scale bar 50 μm). (C) Representative immunofluorescence staining images (left) and quantitative analysis of pericyte coverage assessed by α-SMA/CD31 co-localization on PDAC tumors with either control vaccine (v-CTRL) or ADAM12 vaccine (v-A12) and representative images of α-SMA/CD31 immunostaining. (*n* = 6 mice in v-CTRL, *n* = 6 mice in v-A12. Data were presented as mean ± SEM. Statistical test: unpaired two-tailed Student's *t*-test. Scale bar 50 μm). (D) Representative immunofluorescence staining images (left) and quantitative analysis of tumor hypoxia via GLUT1-positive cells on subcutaneous PDAC from v-CTRL mice and v-A12 mice together with representative images of GLUT1 immunostaining. (*n* = 6 mice in v-CTRL, *n* = 6 mice in v-A12. Data were presented as mean ± SEM. Statistical test: unpaired two-tailed Student's *t*-test. Scale bar 50 μm). (E) Gene expression analysis of the KP2-specific gene CK19 in peripheral blood at endpoint (day 28 post tumor inoculation) from subcutaneous PDAC mouse model with control vaccine (v-CTRL) or ADAM12 vaccine (v-A12). (*n* = 5 mice in v-CTRL, *n* = 5 mice in v-A12. Data were presented as mean ± SEM. Statistical test: unpaired two-tailed Student's *t*-test).

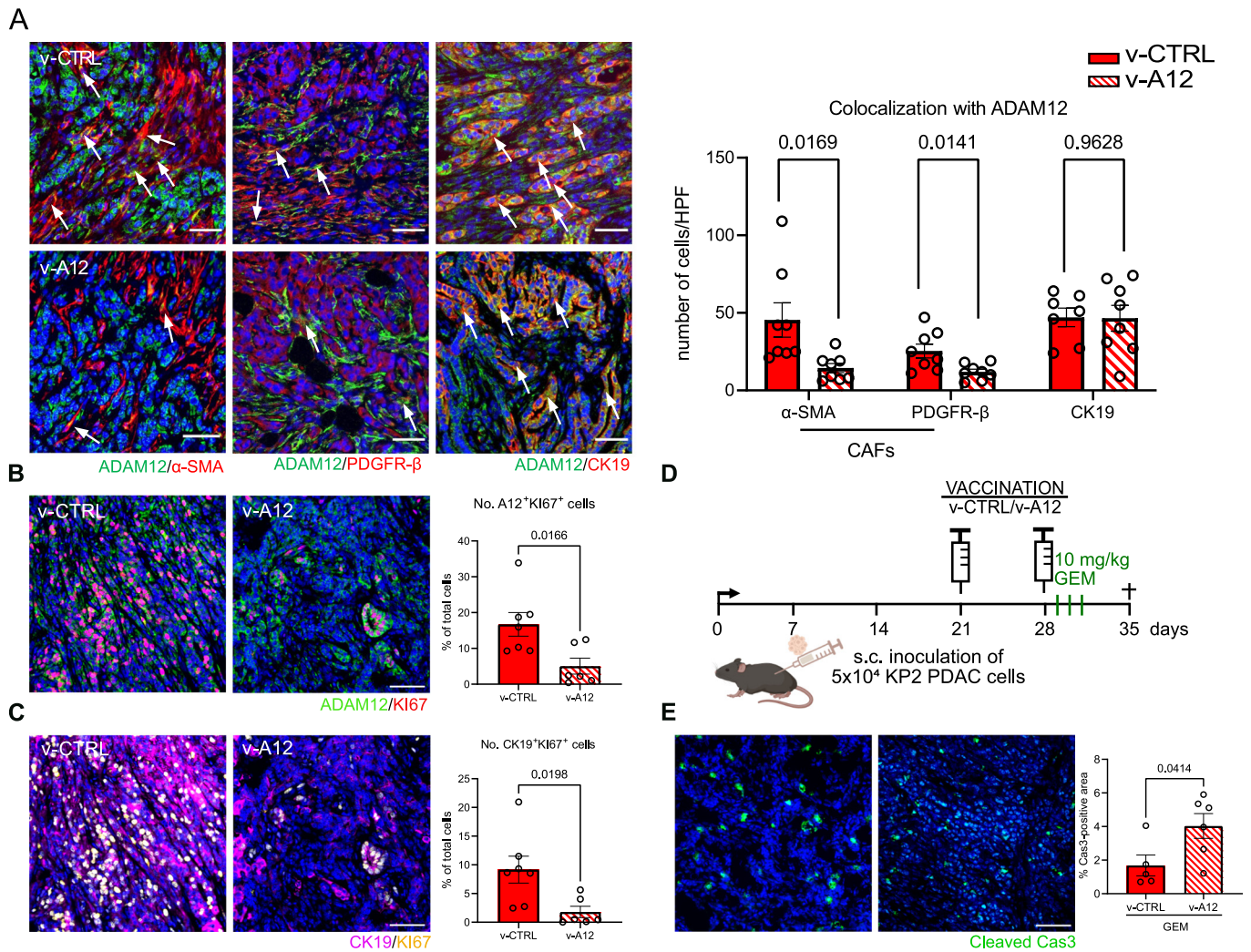
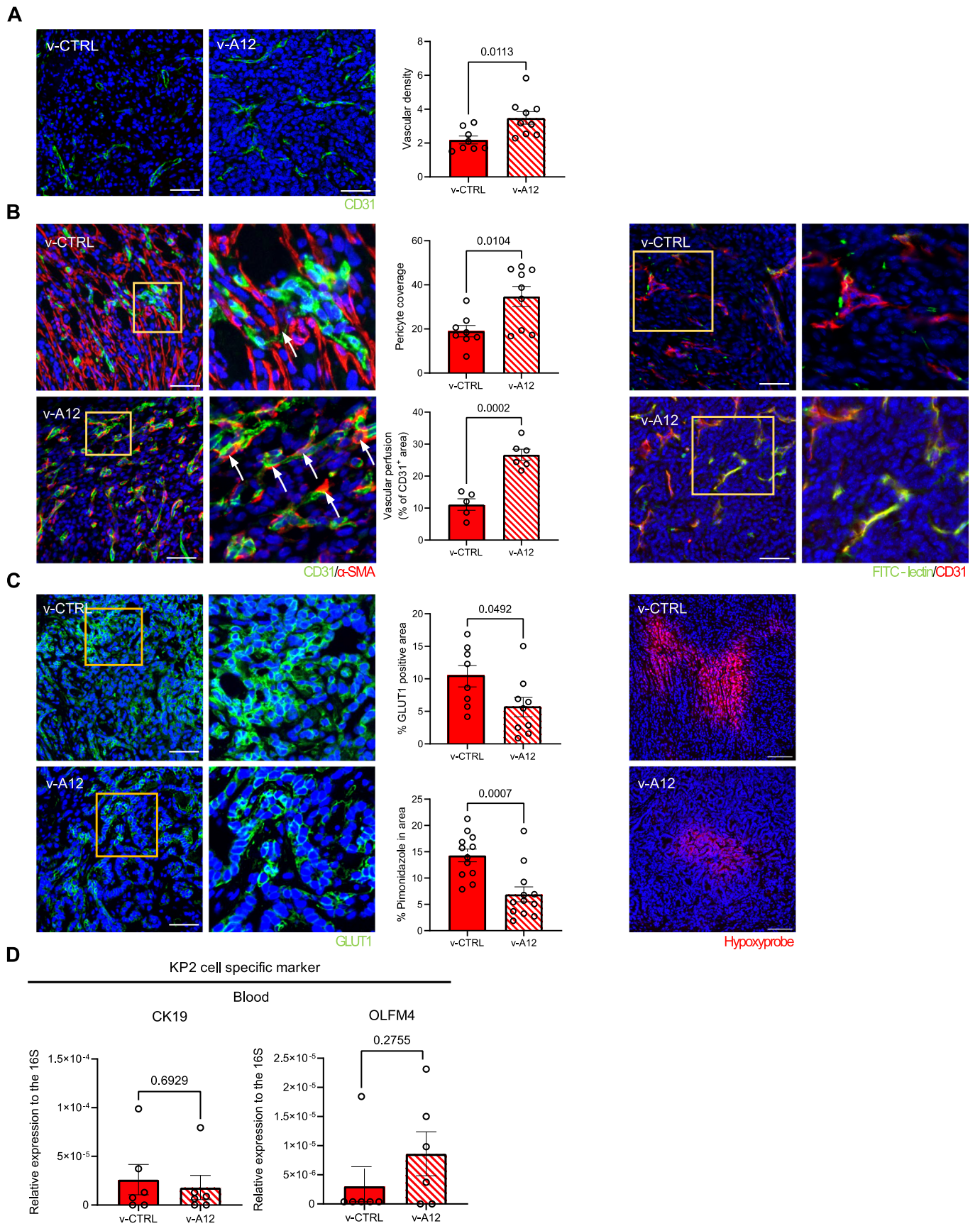


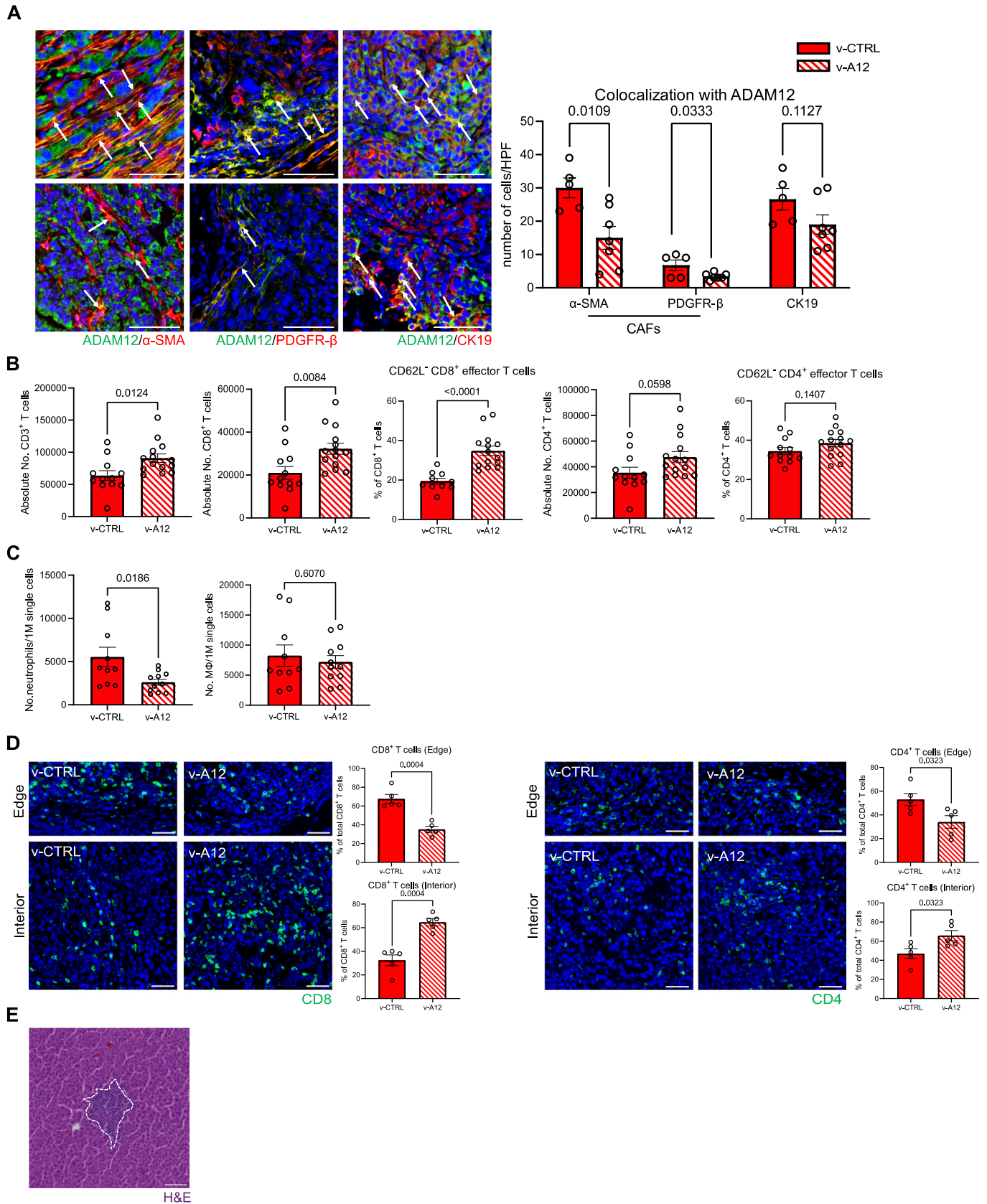
Figure EV3. Therapeutic ADAM12 vaccination targets ADAM12⁺ CAFs and inhibits cell proliferation in subcutaneous PDAC tumors.

(A) Representative immunofluorescence staining images (left) and quantitative analysis (right) of ADAM12⁺/α-SMA⁺ (left, left), ADAM12⁺/PDGFR-β⁺ (left, middle), and ADAM12⁺/CK19⁺ (left, right) co-expression on PDAC tumors from control-vaccinated mice (v-CTRL) and ADAM12-vaccinated mice (v-A12). Data were represented as the number of cells per high power field. Representative double positive cells were indicated by white arrows respectively. ($n = 7-8$ mice in v-CTRL, $n = 8$ mice in v-A12). Data were presented as mean \pm SEM. Statistical test: multiple t -test. Scale bar 50 μ m). (B) Representative immunofluorescence images (left) and quantitative analysis (right) of proliferating (Ki67⁺) ADAM12⁺ cells on PDAC tumors from v-CTRL mice and v-A12 mice. ($n = 7$ mice in v-CTRL, $n = 6$ mice in v-A12). Data were presented as mean \pm SEM. Statistical test: unpaired two-tailed Student's t -test. Scale bar 50 μ m). (C) Representative immunofluorescence staining images (left) and quantitative analysis (right) of proliferating (Ki67⁺) tumor (CK19⁺) cells on PDAC tumors from v-CTRL mice ($n = 7$) and v-A12 mice ($n = 6$). ($n = 7$ mice in v-CTRL, $n = 6$ mice in v-A12). Data were presented as mean \pm SEM. Statistical test: unpaired two-tailed Student's t -test. Scale bar 50 μ m). (D) Scheme of therapeutic vaccination with gemcitabine treatment on subcutaneous KP2 PDAC tumor-bearing mice. (E) Representative immunofluorescence images (left) and quantitative analysis (right) of apoptotic cells (cleaved caspase 3⁺) on PDAC tumors from v-CTRL mice and v-A12 mice. ($n = 5$ mice in v-CTRL, $n = 6$ mice in v-A12). Data were presented as mean \pm SEM. Statistical test: unpaired two-tailed Student's t -test. Scale bar 50 μ m).



◀ Figure EV4. Therapeutic ADAM12 vaccination increased tumor vascular density, pericyte coverage, and decreased tumor hypoxia.

(A) Representative images of immunofluorescence staining (left) and quantitative analysis of CD31-positive endothelial cells (right) on subcutaneous PDAC tumors treated with control vaccine (v-CTRL) and ADAM12 vaccine (v-A12) together with representative images of CD31 immunostaining. ($n = 8$ mice in v-CTRL, $n = 9$ mice in v-A12, Data were presented as mean \pm SEM. Statistical test: unpaired two-tailed Student's *t*-test. Scale bar 100 μ m). (B) Representative images of immunofluorescence staining (left, right) and quantitative analysis of pericyte coverage (middle, upper) assessed by α -SMA/CD31 co-localization (yellow) as well as vascular perfusion (middle, bottom) presented by FITC-lectin⁺/CD31⁺ cells on PDAC tumors treated with control vaccine (v-CTRL) and ADAM12 vaccine (v-A12). Representative pericytes were indicated by white arrows respectively. (Pericyte coverage: $n = 8$ mice in v-CTRL, $n = 9$ mice in v-A12; vascular perfusion: $n = 5$ mice in v-CTRL, $n = 6$ mice in v-A12. Data were presented as mean \pm SEM. Statistical test: unpaired two-tailed Student's *t*-test. Scale bar 50 μ m). (C) Representative images of immunofluorescence staining (left, right) and quantitative analysis of tumor hypoxia by GLUT1-positive cells (middle, upper) as well as by the coverage of Pimonidazole positive area (middle, bottom) on PDAC tumors from mice treated with control vaccine (v-CTRL, $n = 8$) or ADAM12 vaccine (v-A12, $n = 9$). (GLUT1 staining: $n = 8$ mice in v-CTRL, $n = 9$ mice in v-A12; Pimonidazole staining (hypoxyprobe): $n = 12$ mice in v-CTRL, $n = 12$ in v-A12. Data were presented as mean \pm SEM. Statistical test: unpaired two-tailed Student's *t*-test. Scale bar 100 μ m). (D) Gene expression analysis of the KP2-specific gene CK19 and OLFM4 in peripheral blood at endpoint (day 28 post tumor inoculation) from subcutaneous PDAC mouse model with control vaccine (v-CTRL) or ADAM12 vaccine (v-A12). ($n = 5$ mice in v-CTRL, $n = 5$ mice in v-A12. Data were presented as mean \pm SEM. Statistical test: unpaired two-tailed Student's *t*-test).



◀ **Figure EV5. Therapeutic ADAM12 vaccination in mice bearing orthotopic PDAC tumors decreased ADAM12⁺ CAFs, stimulated splenic CD8 T cell response, and promoted T cell relocalization within the PDAC tumor tissue.**

(A) Representative immunofluorescence staining images (left) and quantitative analysis (right) of ADAM12⁺/α-SMA⁺ (left, left), ADAM12⁺/PDGFR-β⁺ (left, middle) and ADAM12⁺/CK19⁺ (left, right) co-expression on PDAC tumors from control-vaccinated mice (v-CTRL, *n* = 5 mice) and ADAM12-vaccinated mice (v-A12, *n* = 7 mice). Data were represented as the number of cells per high power field (HPF). Representative double positive cells were indicated by white arrows respectively. (*n* = 5 in v-CTRL, *n* = 7 in v-A12. Data were presented as mean ± SEM. Statistical test: unpaired two-tailed Student's *t*-test. Scale bar 50 μm). (B) Quantitative FACS analysis of the absolute number of T cells, CD8⁺ T cells, CD62L:CD8⁺ effector T cells, CD4⁺ T cells, and CD62L:CD4⁺ effector T cells in spleens from control-vaccinated mice (v-CTRL) and ADAM12-vaccinated mice (v-A12). (*n* = 12 mice in v-CTRL, *n* = 14 mice in v-A12. **n* = 10 mice in v-CTRL in the CD62L:CD8⁺ effector T cell population because of two outliers. Data were presented as mean ± SEM. Statistical test: unpaired two-tailed Student's *t*-test.). (C) Quantitative analysis of the number of neutrophils (left) and macrophages (right) per 1 M single cells in PDAC tumors from control-vaccinated mice (v-CTRL) and ADAM12-vaccinated mice (v-A12). (*n* = 10 mice in v-CTRL, *n* = 11 mice in v-A12. Data were presented as mean ± SEM. Statistical test: unpaired two-tailed Student's *t*-test.). (D) Representative images of CD8⁺ T cell (left) and CD4⁺ T cell (right) localization, and quantitative analysis of CD8⁺ T cell and CD4⁺ T cell population at the edge of PDAC tumor tissue (right, upper) and the interior of PDAC tumor tissue (right, bottom). (*n* = 5 mice in v-CTRL, *n* = 5 mice in v-A12. Data were presented as mean ± SEM. Statistical test: unpaired two-tailed Student's *t*-test. Scale bar 50 μm). (E) Representative hepatic micrometastasis (area indicated within the white dotted outline) image with H&E staining, scale bar 50 μm.