

1 **Supplementary information to**

2

3 **The dual BCL-2 and BCL-XL inhibitor AZD4320 acts on-target and synergizes**
4 **with MCL-1 inhibition in B-cell precursor ALL**

5

6 **Materials and Methods:**

7 **Cell lines**

8 Seven different BCP-ALL cell lines were used: RS4;11, KOPN-8, REH, EU-3, RCH-
9 ACV and NALM-6 (purchased from DSMZ, Germany), UoCB6 (provided by J. Rowley,
10 USA). Cell lines were cultured in RPMI-1640, 20% fetal bovine serum, 1% L-Glutamine
11 and 1% Penicillin/Streptomycin. Cell lines were authenticated by short-tandem repeat
12 (STR)-profiling (GenePrint 10 System, Promega) and tested for negativity of
13 mycoplasma (MycoAlert Mycoplasma Detection Kit, Lonza).

14

15 **Patient-derived xenograft samples**

16 Samples were collected from patients with BCP-ALL after obtaining informed written
17 consent in accordance with institutional ethics review board guidelines. Patient-derived
18 xenograft (PDX) samples were established by intravenously transplanting ALL cells
19 into female NOD/SCID mice (NOD.CB17-Prkdcscid, Charles River), following
20 established protocols¹. Animal experiments received approval (RP Tübingen, No.
21 1260). Genetic alterations of the samples were assessed by Multiplex Ligation-
22 dependent Probe Amplification (MLPA) and RT-PCR, as previously described². For *ex*
23 *vivo* experiments analyzing EC₅₀ values, PDX samples were cultured in AIM V medium
24 (Gibco) without supplements.

25

26 **Cell death assays**

27 Cell death rates were assessed by propidium iodide (PI, P3566, Invitrogen) staining
28 following exposure to AZD4320 and AZD5991 for 48h in cell lines and for 24h in PDX
29 samples. To determine the half maximal effective concentration (EC_{50}), cells were
30 exposed to eleven different concentrations (cell lines: 0.1, 1, 5, 10, 50, 100, 250, 500,
31 1000, 5000 and 10000 nM; PDX samples: 0.1, 1, 2.5, 5, 10, 25, 50, 100, 500, 1000
32 and 5000 nM) of each inhibitor. The EC_{50} values of venetoclax, A-1331852 and S63845
33 have been published previously³. Combination effects in cell lines were analyzed using
34 dose-response matrix analyses upon treatment with seven different concentrations of
35 the inhibitors (0.1, 1, 2.5, 5, 10, 100 and 500 nM) for 48 hours. Samples were measured
36 on an Attune NxT Flow Cytometer (Thermo Fisher) using an autosampler and analyzed
37 using the FlowJo 10.7.1 software.

38

39 **ALL co-culture model with mesenchymal stroma cells (MSCs)**

40 Primary bone marrow hTERT-immortalized mesenchymal stroma cells (MSCs)⁴ were
41 kindly provided by Dario Campana. For co-culture experiments, MSCs were seeded
42 on day -1 at 5×10^3 per well in 96-well plates (REF83.3924, Sarstedt) in RPMI-1640
43 medium supplemented with 20% fetal bovine serum, 1% L-Glutamine and 1%
44 Penicillin/Streptomycin. After four hours, 1×10^5 ALL PDX cells or PBMCs stained with
45 1 μ M CellTrace Violet (C34557, Thermo Fisher) were added to the MSC cultures. On
46 day 0, samples were exposed to increasing concentrations of the inhibitors (0.1, 1, 2.5,
47 5, 10, 100 and 500 nM) in a drug matrix for 72 hours. Cell death rates were assessed
48 on an Attune NxT Flow Cytometer (Thermo Fisher) with an autosampler using
49 propidium iodide (PI) and CellTrace Violet staining to distinguish ALL PDX cells or
50 PBMCs from MSCs. Data analysis was performed using FlowJo 10.7.1 software.

51

52 **Dynamic BH3 profiling**

53 Dynamic BH3 profiling was carried out as previously described⁵⁻⁷. RCH-ACV cells were
54 exposed to AZD4320 or AZD5991 for two hours. Cells were then permeabilized using
55 digitonin, and exposed to the pro-apoptotic peptides indicated. Thereafter, cells were
56 stained using a viability marker (1:1000 Zombie Violet; 423113, BioLegend). Cells were
57 then fixed (formaldehyde) and neutralized in N2 buffer. Finally, cells were incubated in
58 an intracellular staining buffer (1:1000 Alexa Fluor 647 anti-Cytochrome c antibody;
59 612310, BioLegend). Results were analyzed on an Attune NxT Flow Cytometer
60 (Thermo Fisher) with an autosampler and processed with FlowJo 10.7.1.

61

62 **Western blot and immunoprecipitation**

63 Western blot analysis and immunoprecipitation were performed according to
64 established protocols³. Protein extraction was performed with lysis buffer containing
65 30 mM Tris-HCl, 1% Triton X-100 buffer, 10% glycerol. For immunoprecipitation, cell
66 lysates were co-incubated with BIM (C34C5) Rabbit mAb (2933, Cell Signaling) at 4°C
67 on a rotary overnight, followed by co-incubation with Protein A Agarose Beads (9863,
68 Cell Signaling) for 2-4 hours at 4°C. The beads were then washed five times with lysis
69 buffer and the precipitates were subjected to western blot analysis. For
70 immunoblotting, the following antibodies were used: Bim (C34C5) Rabbit mAb (2933,
71 Cell Signaling), Bcl-2 (124) Mouse mAb (15071, Cell Signaling), Bcl-xL (54H6) Rabbit
72 mAb (2764, Cell Signaling), Mcl-1 (D2W9E) Rabbit mAb (94296, Cell Signaling), α/β -
73 Tubulin Antibody (2148, Cell Signaling) followed by mouse anti-rabbit IgG-HRP (sc-
74 2357, Santa Cruz) or m-IgG_k BP-HRP (sc-516102, Santa Cruz).

75

76 **Statistics**

77 Statistical analyses were conducted using GraphPad Prism 10 software and Microsoft
78 Excel. Combination effects and synergy scores were analyzed using the Bliss
79 independence model with Synergyfinder⁸⁻¹⁰.

80 **Supplementary References:**

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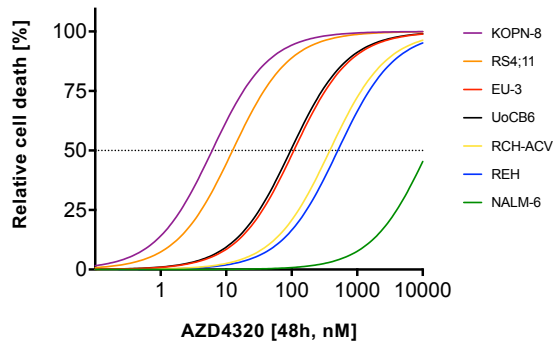
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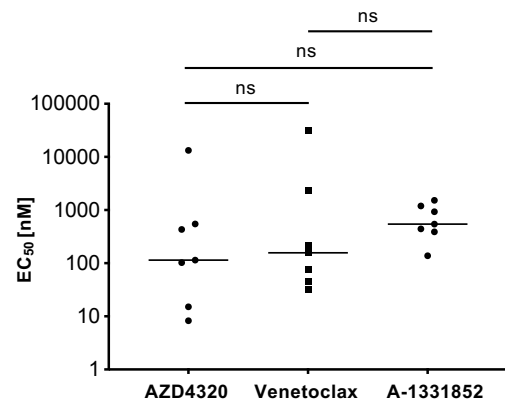
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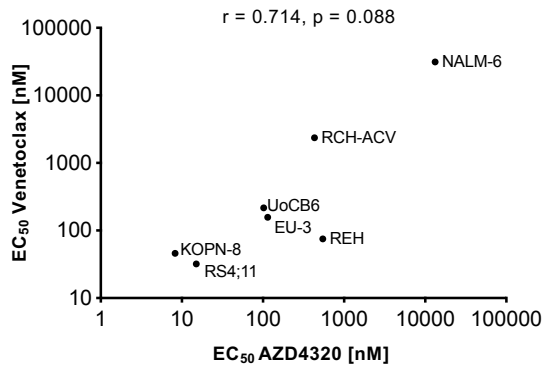
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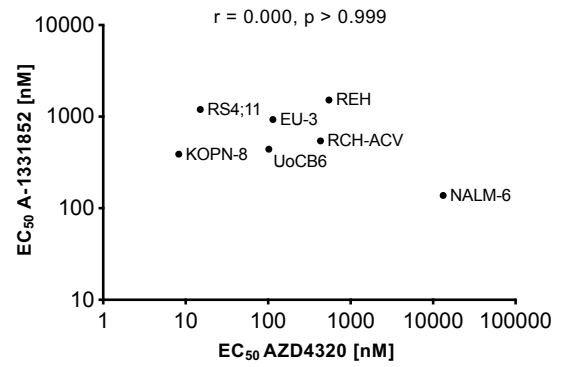
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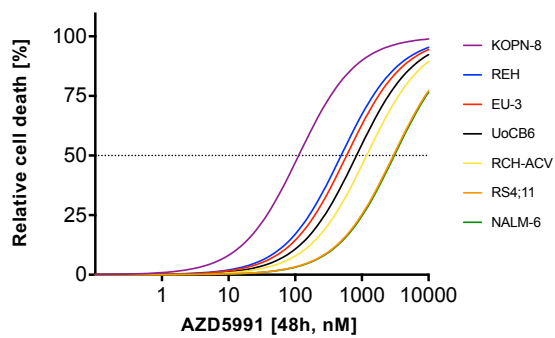
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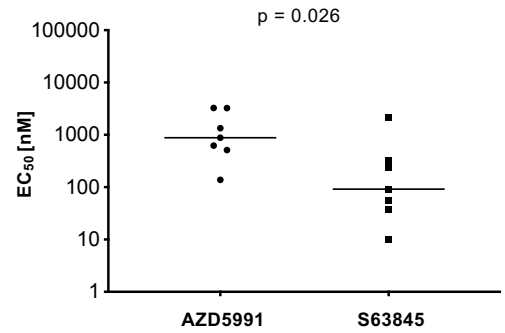
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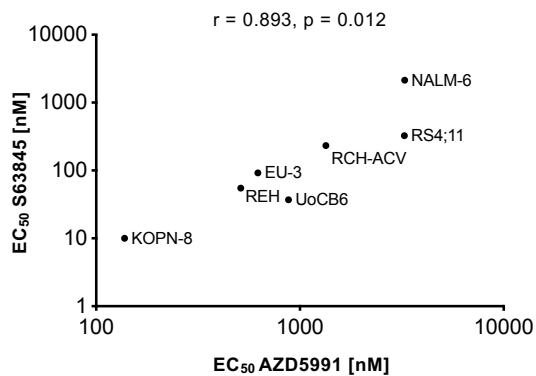
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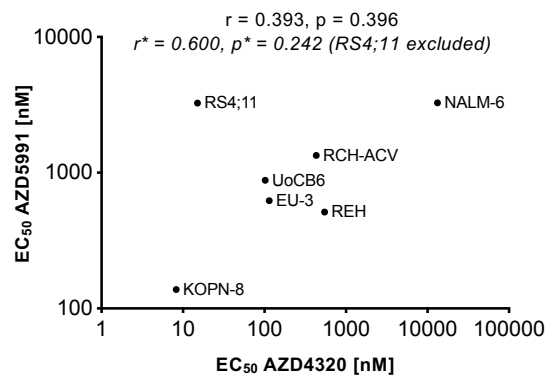
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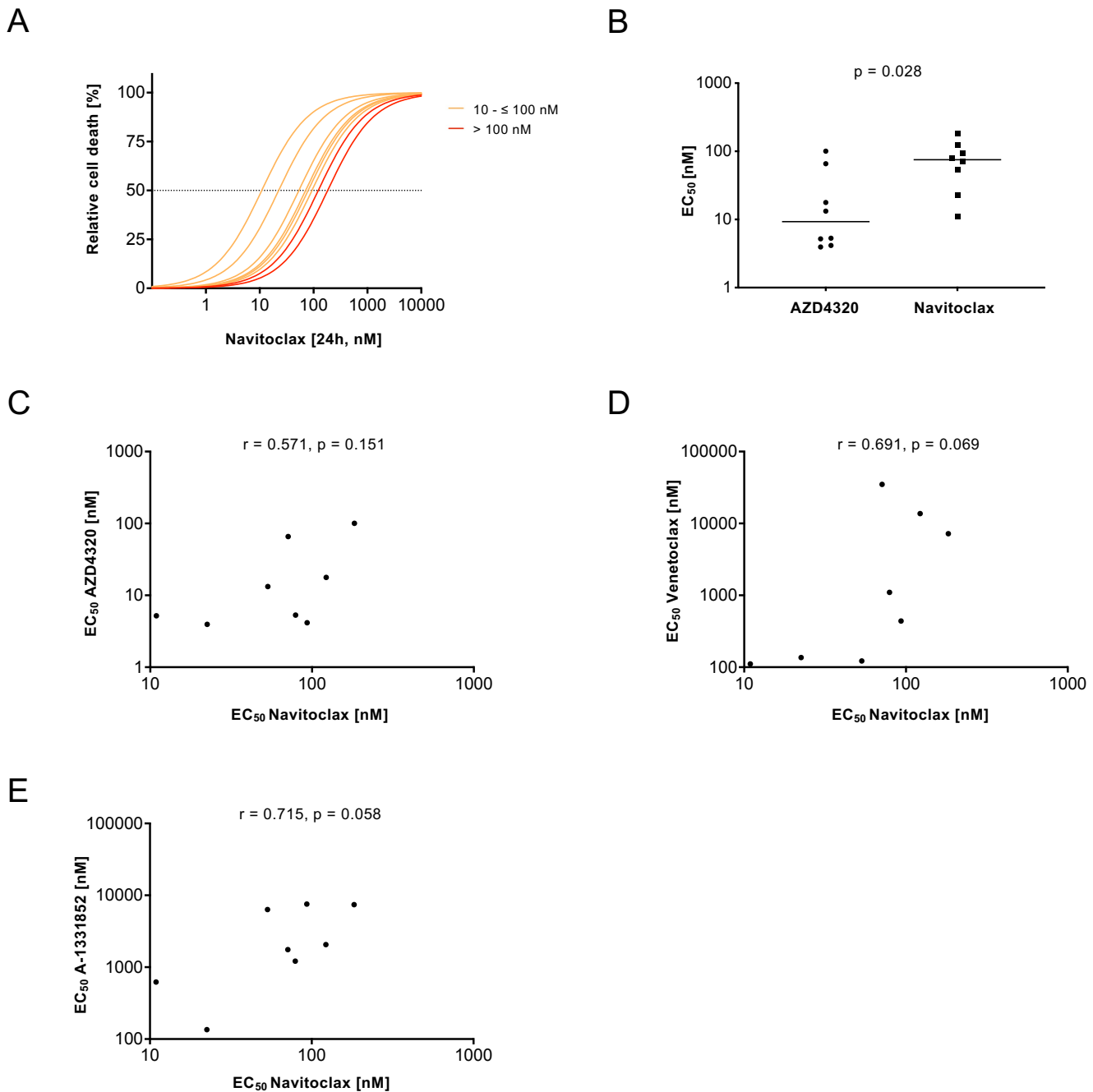


H



Legend to Supplementary Figure 1. Varying sensitivities towards BH3-mimetics of BCP-ALL cell lines

(A) Cell lines were exposed to eleven increasing concentrations (0.1, 1, 5, 10, 50, 100, 250, 500, 1000, 5000 and 10000 nM) of AZD4320 for 48 hours. Cell death rates were measured by propidium iodide (PI) staining. EC_{50} values were determined showing a range from 8.26 nM to 13.23 μ M and a median of 114.20 nM. **(B)** The sensitivity of AZD4320 (inhibiting BCL-2/BCL-XL) was compared with venetoclax (BCL-2) and A-1331852 (BCL-XL). The scatter plot shows individual EC_{50} values and the lines show medians. Mann-Whitney U-test; p, significance. **(C)** Correlation of the sensitivities of AZD4320 with venetoclax **(D)** and of AZD4320 with A-1331852. Spearman correlation; r, correlation coefficient; p, significance. **(E)** Cell death rates (PI) were assessed upon exposure of cell lines to eleven concentrations (0.1, 1, 5, 10, 50, 100, 250, 500, 1000, 5000 and 10000 nM) of AZD5991 for 48 hours. EC_{50} values ranged between 137.8 nM and 3.27 μ M with a median of 879.1 nM. **(F)** Comparison of sensitivities of two MCL-1 inhibitors. Mann-Whitney U-test; p, significance. **(G)** Spearman correlation of the EC_{50} values of both MCL-1 inhibitors **(H)** and of AZD4320 and AZD5991. Spearman correlation; r, correlation coefficient; p, significance; r^* and p^* indicate the correlation coefficient and significance, respectively, excluding the RS4;11 cell line. The EC_{50} values of venetoclax, A-1331852 and S63845 have been published previously (3).

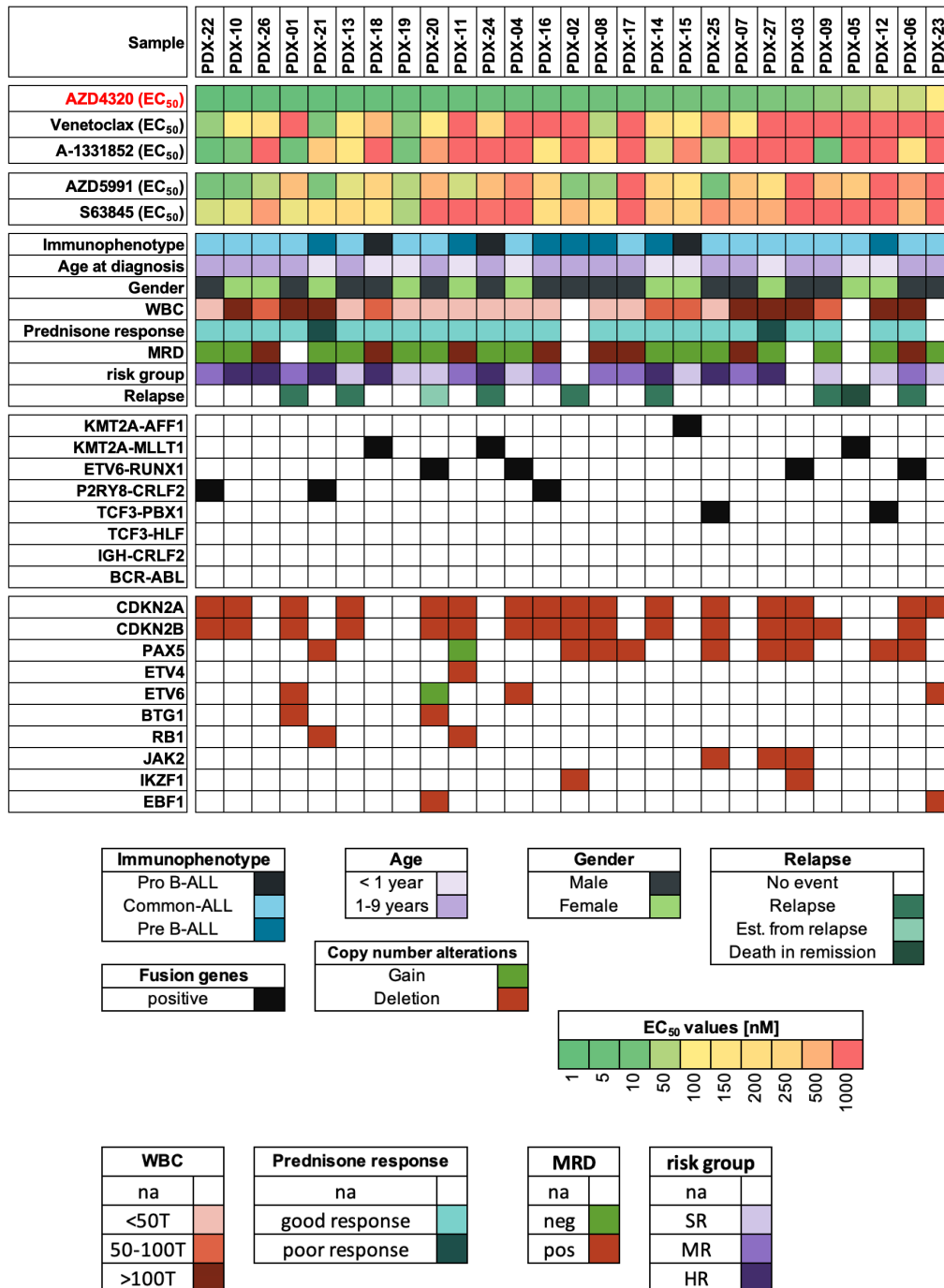


Legend to Supplementary Figure 2. Varying sensitivities towards navitoclax in BCP-ALL patient-derived xenograft samples

(A) B-cell precursor acute lymphoblastic leukemia (BCP-ALL) patient-derived xenograft (PDX) samples (N=8) were cultured for 24 hours and exposed to eleven increasing concentrations (0.1, 1, 2.5, 5, 10, 25, 50, 100, 500, 1000 and 5000 nM) of the BCL-2/BCL-XL inhibitor navitoclax. Cell death rates were measured by propidium iodide (PI) staining. EC₅₀ values were determined showing a range from 10.91 to 182.9 nM and a median of 75.18 nM. **(B)** The sensitivity of navitoclax was compared with AZD4320 (both inhibiting BCL-2/BCL-XL). The scatter plot shows individual EC₅₀ values and the lines show medians. Mann-Whitney U-test; p, significance. **(C-E)** Correlation of the sensitivities of navitoclax with **(C)** AZD4320, **(D)** venetoclax and **(E)** A-1331852. Spearman correlation; r, correlation coefficient; p, significance.

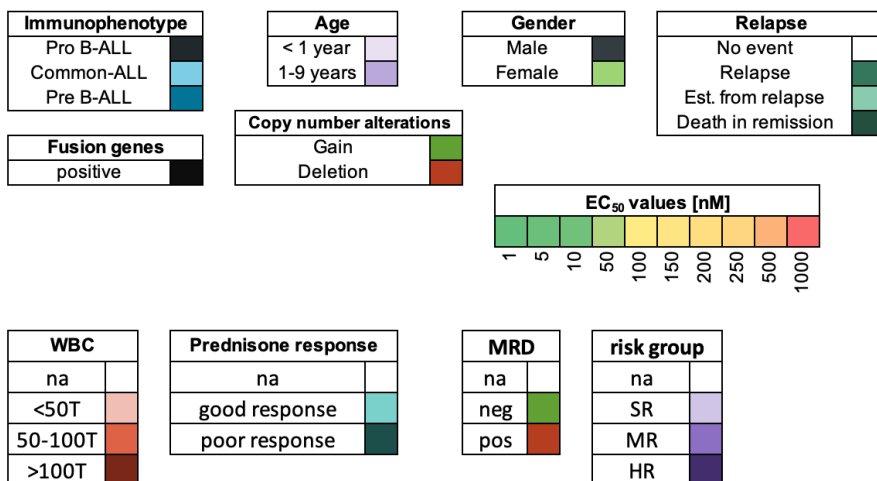
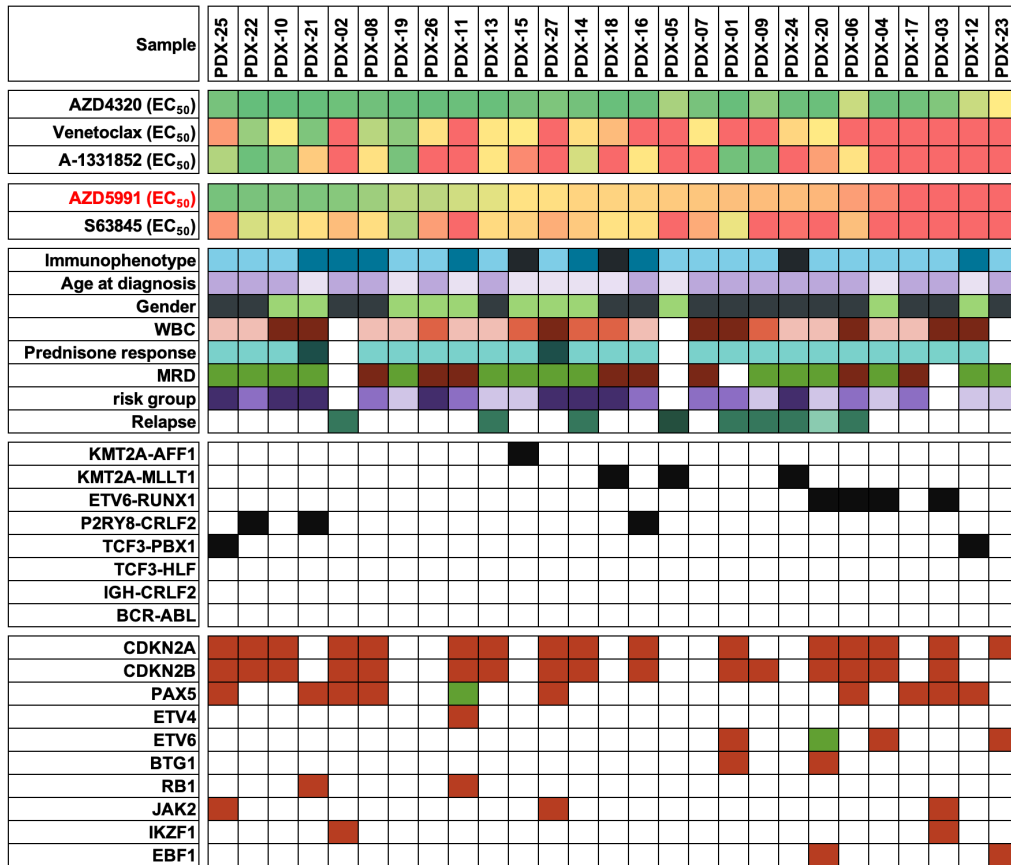
A

Supplementary Figure 3. Characteristics of patient-derived xenograft samples and corresponding patients. **(A)** The samples are sorted based on the EC₅₀ values of AZD4320. The EC₅₀ values of venetoclax, A-1331852 and S63845 have been published previously (3).



B

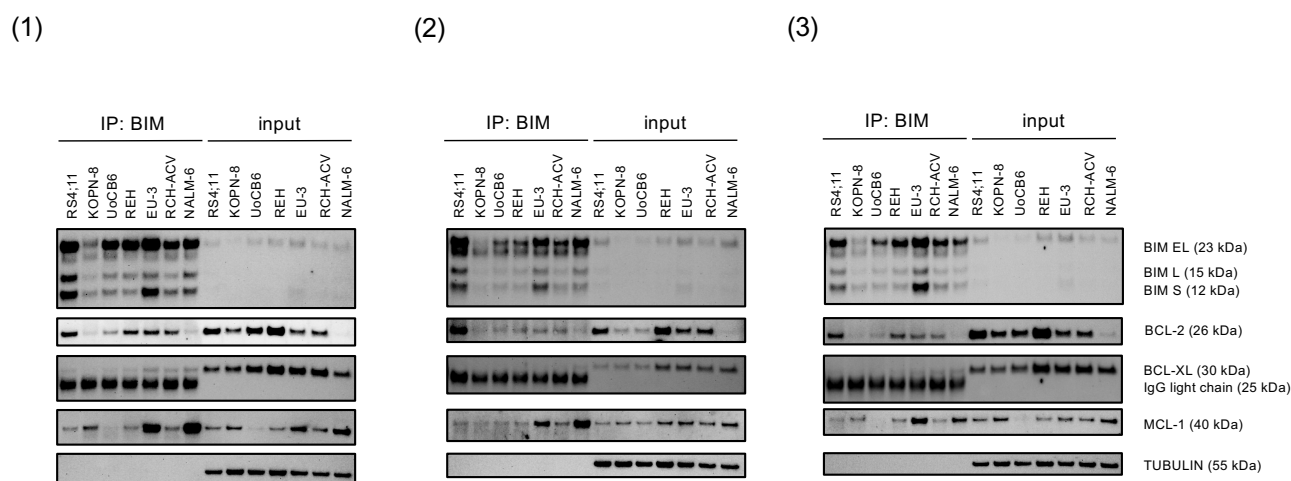
(B) The samples are sorted based on the EC₅₀ values of AZD5991.



Supplementary Figure 4. Co-immunoprecipitations of BIM BCP-ALL samples.

A

(A) Basal levels of proteins and protein complexes are shown in cell lines. N=3 independent experiments are shown.



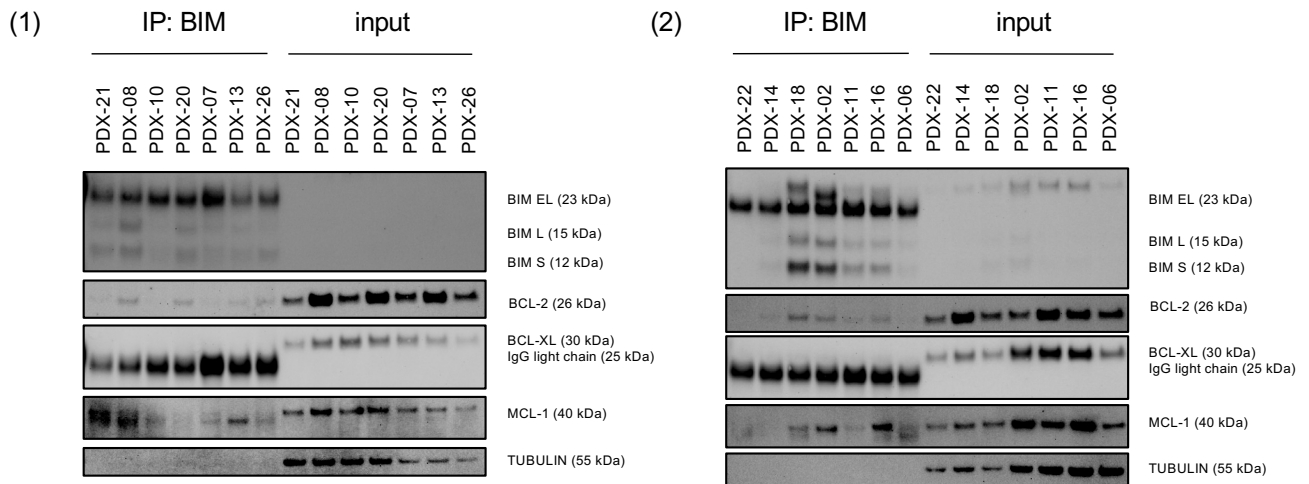
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(B) Densitometric quantification of (a) protein levels (b) protein ratios and (c) protein complexes in ALL cell lines from N=3 independent experiments. Spearman correlation of proteins with the EC_{50} values of the compounds. A negative Spearman rho would indicate that sensitivity (low EC_{50}) is correlated to a high level of protein or protein complex. N=7 cell lines; ρ , significance; significance level (Bonferroni-adjusted): $P \leq 0.05/45 = P \leq 0.001$.

Compounds		AZD4320		Venetoclax		A-1331852		S63845		AZD5991		
(a)	Protein levels	r_s	p	r_s	p	r_s	p	r_s	p	r_s	p	
Input	BCL-2	0.000	1.000	-0.429	0.345	0.893	0.012	-0.036	0.963	-0.107	0.840	
	BCL-XL	0.714	0.088	0.714	0.088	0.071	0.906	0.464	0.302	0.321	0.498	
	MCL-1	0.393	0.396	0.464	0.302	-0.429	0.354	0.571	0.200	0.357	0.444	
(b)	Protein ratios	r_s	p	r_s	p	r_s	p	r_s	p	r_s	p	
	Input	BCL-2/BCL-XL	-0.464	0.302	-0.750	0.066	0.643	0.139	-0.286	0.556	-0.214	0.662
	BCL-2/MCL-1	-0.143	0.783	-0.250	0.595	0.607	0.167	-0.321	0.498	-0.143	0.783	
BCL-2/(BCL-XL+MCL-1)	-0.429	0.354	-0.536	0.354	0.571	0.200	-0.179	0.713	0.000	1.000		
(b)	Protein complexes	r_s	p	r_s	p	r_s	p	r_s	p	r_s	p	
	IP: BIM	BCL-2-BIM	-0.429	0.345	-0.857	0.024	0.714	0.088	-0.143	0.783	-0.321	0.498
	BCL-XL-BIM	-0.143	0.783	0.250	0.595	-0.821	0.034	-0.071	0.906	0.107	0.840	
MCL-1-BIM	0.071	0.906	0.143	0.783	-0.536	0.236	0.000	1.000	-0.214	0.662		

C

(C) Basal levels of proteins and protein complexes are shown for PDX samples.



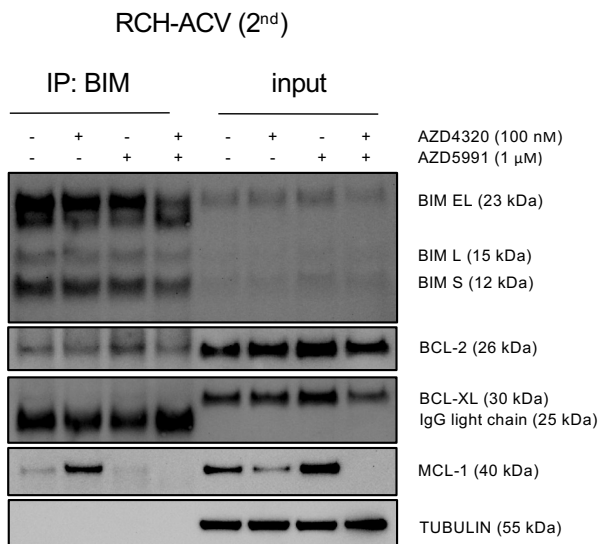
D

(D) Densitometric quantification of (a) protein levels, (b) protein ratios and (c) protein complexes in ALL PDX samples. Spearman correlation of proteins with the EC_{50} values of the compounds. A negative Spearman rho would indicate that sensitivity (low EC_{50}) is correlated to a high level of protein or protein complex. N=14 PDX samples; p, significance; significance level (Bonferroni-adjusted): $P \leq 0.05/45 = P \leq 0.001$.

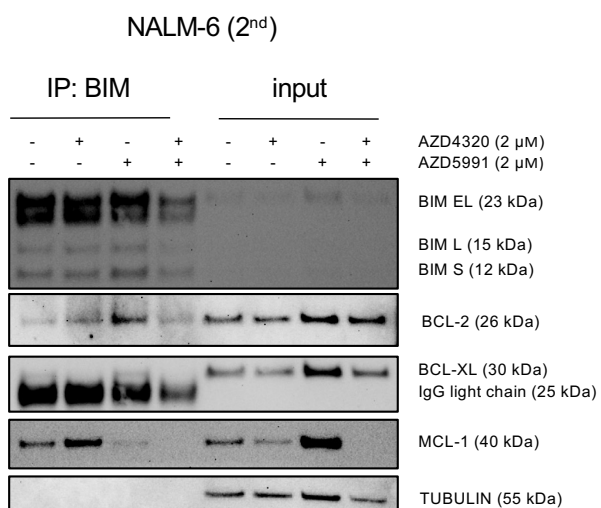
		AZD4320		Venetoclax		A-1331852		S63845		AZD5991		
(a)	Protein levels	r_s	p	r_s	p	r_s	p	r_s	p	r_s	p	
	Input	BCL-2	-0.024	0.940	-0.051	0.638	0.064	0.832	0.262	0.366	0.284	0.325
		BCL-XL	0.231	0.427	-0.165	0.445	0.332	0.246	0.319	0.267	0.152	0.605
		MCL-1	0.103	0.727	-0.077	0.739	0.543	0.048	0.196	0.502	0.253	0.382
(b)	Protein ratios	r_s	p	r_s	p	r_s	p	r_s	p	r_s	p	
	Input	BCL-2/BCL-XL	0.011	0.976	0.204	0.483	-0.112	0.704	0.095	0.750	0.297	0.302
		BCL-2/MCL-1	0.024	0.940	0.376	0.186	-0.376	0.616	0.244	0.400	0.288	0.318
		BCL-2/(BCL-XL+MCL-1)	-0.007	0.988	0.235	0.417	-0.152	0.605	0.130	0.659	0.288	0.318
(c)	Protein complexes	r_s	p	r_s	p	r_s	p	r_s	p	r_s	p	
	IP: BIM	BCL-2-BIM	-0.240	0.409	0.051	0.868	0.182	0.532	-0.042	0.892	-0.108	0.716
		BCL-XL-BIM	-0.323	0.260	-0.165	0.573	0.046	0.880	0.024	0.940	-0.086	0.773
		MCL-1-BIM	0.064	0.832	-0.077	0.797	-0.134	0.648	-0.402	0.155	-0.152	0.605

Supplementary Figure 5. Independent experiments of co-immunoprecipitations of BIM for **(A)** RCH-ACV corresponding to Figure 2A and for **(B)** NALM-6 corresponding to Figure 2B. Cells were exposed to the compounds at the concentrations indicated for four hours.

A



B



Supplementary Figure 6. Combination effects of AZD4320 and AZD5991 in BCP-ALL cell lines

Dose-response matrix analyses were conducted in three BCP-ALL cell lines that are less sensitive to AZD4320 and/or AZD5991 alone. The cell lines were exposed to seven increasing concentrations (5, 25, 50, 100, 250, 500 and 1000 nM) of AZD4320 and/or AZD5991 in a drug matrix for 48 hours. Cell death rates were determined by propidium iodide staining and flow cytometry measurement. The heatmaps show cell death rates after drug exposure. Efficacy scores were calculated as the arithmetic mean of normalized cell death rates across the matrix. Synergy effects were visualized using Synergyfinder and synergy scores were analyzed using the Bliss independence model. Dashed lines indicate the most synergistic area (MSA). Mean values of two independent experiments are shown.

