

#### Figure S1. The expression of NQO1 and SOD1 but not CAT and SOD2 corelates with poor patient survival of BC, related to Figure 1.

(A-C) High NQO1 expression corelates with low recurrence-free survival (RFS) of luminal A (A), luminal B (B) and HER2+(C) BC.

(**D-E**) High SOD1 expression corelates with low RFS of all BC subtypes (**D**) as well as basal BC (**E**).

(**F-G**) CAT expression does not corelate with low RFS of all BC subtypes (**F**) as well as basal BC (**G**).

(H-I) SOD2 expression has no corelation with low RFS of all BC subtypes (H) as well as basal BC (I).



Figure S2. IB-DNQ is more potent and specific than  $\beta$ -lap in killing TNBC cells and Dox-induced KD of SOD1 specifically enhances IB-DNQ's efficacy in killingTNBC cells, related to Figure 2.

(**A-B**) Comparison of LD<sub>50</sub> of IB-DNQ (A) vs.  $\beta$ -lap (B) in SUM159 BC cells treated with each compound alone or together with the NQO1 inhibitor DIC. Error bars denote SEM (n=6).

(**C**) SUM159 BC cells treated with Mock (DMSO), DIC (20  $\mu$ M), IB-DNQ (100 or 300nM) alone, or combination of IB-DNQ (100 or 300nM) and DIC (20 $\mu$ M) for 24h and examined by light microscopy.

(**D-E**) Comparison of LD<sub>50</sub> of IB-DNQ (D) vs.  $\beta$ -lap (E) in MDA-MB-157 BC cells treated with each compound alone or together with NQO1 inhibitor DIC. Error bars denote SEM (n=6).

(**F-G**) Comparison of LD<sub>50</sub> of IB-DNQ (F) vs.  $\beta$ -lap (G) in NQO1<sup>10</sup> HCC1806 BC cells treated with each compound alone or together with NQO1 inhibitor DIC. Error bars denote SEM (n=6).

(H) Relative survival of SUM159 BC cells treated with different doses of IB-DNQ measured by MTT vs. CCK8 assay. Error bars denote SEM (n=6).

(I) Relative survival of SUM159 BC cells treated with different doses of IB-DNQ in media containing high (25 mM) or low (5.5 mM) of glucose by CCK8 assay. Error bars denote SEM (n=6)

(J-L) Dox-inducible KD of NQO1, CAT, SOD1 and SOD2 in HCC70 BC cells validated by immunoblotting with respective antibodies (J), and KD of NQO1 renders HCC70 BC cells resistant to IB-DNQ treatment, while KD of SOD1, but not CAT or SOD2, in HCC70 significantly enhanced IB-DNQ-elicited lethality (K and L). \*\*\*\*: p< 0.0001 vs. shSCR (n=2, one-way ANOVA). Error bars denote SEM.

(**M-O**) Dox-inducible KD of CAT, SOD1 and SOD2 in NQO1<sup>IO</sup> SUM149 BC cells validated by immunoblotting with respective antibodies (**M**), and KD of SOD1 or SOD2 but not CAT in SUM149 significantly enhanced IB-DNQ-elicited lethality (**M** and **O**). \*\*, \*\*\*\*: p< 0.01 and 0.0001 vs. shSCR (n=2, one-way ANOVA). Error bars denote SEM.



## Figure S3. SOD1 inhibitor ATN224 synergistically enhances IB-DNQ's efficacy in killingTNBC but not nontumorigenic mammary epithelial cell MCF10A, related to Figure 2.

(**A**) Dox-induced KD of NQO1 in SUM159 BC cells with three different SMARTvector Dox-inducible lentiviral shRNA clones to validate the effect of knockdown of NQO1 expression in affecting IB-DNQ sensitivity, which is measured by MTT assay. \*\*\*\*: p< 0.0001 (shNQO1-2 or shNQO1-3 vs. shSCR, n=6, two-way ANOVA). Error bars denote SEM.

(**B**) Dox-induced KD of SOD1 in SUM159 BC cells with two SMARTvector Dox-inducible lentiviral shRNA clones to validate the effect of knockdown of SOD1 expression in affecting IB-DNQ sensitivity, measured by MTT assay. \*\*\*\*: p< 0.0001 (shSOD1-2 vs. shSCR or shSOD1-1, n=6, two-way ANOVA). Error bars denote SEM.

(**C-D**) Relative survival of NQO1<sup>Io</sup> HCC1806 BC cells treated with IB-DNQ alone or together with SOD1 inhibitor ATN224 (**C**, n=6) and combination index (CI) of ATN224 (at 5  $\mu$ M) with sublethal doses of IB-DNQ ranging from 12.5-200 nM in HCC1806 (**D**). Error bars denote SEM.

(E-F) Relative survival of NQO1<sup>Io</sup> MCF10A cells treated with with IB-DNQ alone or together with SOD1 inhibitor ATN224 (E, n=6) or LCS-1 (F, n=6). Error bars denote SEM.

(**G-I**) Relative survival of SUM149 (**G**, n=6) and SUM159 (**H**, n=6) TNBC cells treated with IB-DNQ alone or together with SOD1 inhibitor LCS-1, and combination index (CI) of LCS-1 (at 1  $\mu$ M) together with variousl doses of IB-DNQ ranging from 12.5-200 nM in SUM149 and SUM159 BC cells (**I**). Error bars denote SEM.



Figure S4. IB-DNQ preferentially inhibits CSC activity and genetic or phamacological inhibition of SOD1 potentiate the efficacy of IB-DNQ in supressing tumorsphere-forming capacity of TNBC cells, related to Figure 3.

(**A-B**) Tumorsphere formation of Vari068 (**A**) and SUM159 (**B**) BC cells sorted into 96-well ultralow-attachment plates at density of 20 cells/well. Bar: 200  $\mu$ m.

(**C**) Tumorsphere formation assay at clonal density (20 cells/well) was performed to examine the effects of Dox (0.5  $\mu$ g/ml) induced KD of CAT, SOD1 or SOD2 vs. SCR in affecting sphere-forming capacity of SUM149 BC cells and their impact on IB-DNQ-mediated suppression of tumorsphere formation. Bar: 200  $\mu$ m. \*\*\*\*: P < 0.0001 (n=3, one-way ANOVA). Error bars denote SEM.

(**D**) Tumorsphere formation of SUM149 BC cells treated with vehicle (DMSO), IB-DNQ (50nM), ATN224 (2.5  $\mu$ M), or IB-DNQ (50nM) plus ATN224 (2.5  $\mu$ M). Bar: 200  $\mu$ m. \*\*\*\*: P < 0.0001 (n=3, one-way ANOVA). Error bars denote SEM.

(E) Tumorsphere formation of SUM159 BC cells treated with vehicle (DMSO), ATN224 (1-4  $\mu$ M), IB-DNQ (10-40 nM) or IB-DNQ plus ATN224 at different doses. \*, \*\*, \*\*\*\*, \*\*\*\*: P <0.05, 0.01, 0.001 or 0.0001 (vs. Control, n=3, one-way ANOVA). Error bars denote SEM.





I

























Figure S5. IB-DNQ preferentially inhibits ALDH+ CSCs in a NQO1dependent manner and genetic or phamacological inhibition of SOD1 potentiate the efficacy of IB-DNQ in supressing both ALDH+ E- and CD24-CD44+ M-CSC-like cells, related to Figure 3.

(**A-D**) SUM159 BC cells were treated with Control (DMSO), DIC (20  $\mu$ M), IB-DNQ (50-200 nM) alone or IB-DNQ together DIC for 20h, and examined by flow cytometry to measure the content and absolute number of ALDH+ (**A** and **B**) and CD24-CD44+ (**C** and **D**) CSC-like cells. \*, \*\*, \*\*\*, \*\*\*\*: P< 0.05, 0.01, 0.001 or 0.0001 respectively (n=3). Error bars denote SEM.

(E-G) Vari068 BC cells expressing SCR or shNQO1 were pretreated with Dox (0.5  $\mu$ g/ml) for 48h and then treated with different doses of IB-DNQ (0-200nM) for 20h to examine the percentage and total number of ALDH+CSCs. \*\*, \*\*\*,\*\*\*: P< 0.01, 0.001, 0.0001 respectively (n=3). Error bars denote SEM.

(H-K) SUM149 BC cells expressing SCR or shSOD1 were pretreated with Dox (0.5  $\mu$ g/ml) for 48h, and then treated with different doses of IB-DNQ (50, 100, 200nM) for 20h to examine the percentage and total number of ALDH+ and CD24-CD44+ CSCs. \*, \*\*, \*\*\*, \*\*\*\*: P< 0.05, 0.01, 0.001, 0.0001 respectively (n=3). Error bars denote SEM.

(L-M) SUM159 BC cells were treated with Control (DMSO), DIC (10  $\mu$ M), IB-DNQ (50-200 nM), ATN224 (5  $\mu$ M) alone or IB-DNQ together with ATN224 and/or DIC for 20h, and examined by flow cytometry to measure the content (L) and absolute number (M) of ALDH+ CSCs. \*, \*\*, \*\*\*,\*\*\*\*: P< 0.05, 0.01, 0.001 or 0.0001 respectively vs. Control or indicated with brackets (n=3). Error bars denote SEM.



Figure S6. IB-DNQ treatment induces apoptotic death of TNBC cells overexpressing NQO1 and SOD1 inhibitor ATN224 potentiates the ability of IB-DNQ to elevate mitoROS in TNBC expressing different levels of NQO1, related to Figure 4 and Figure 5.

(**A**) IB-DNQ treatment dose-dependently induces apoptosis of NQO<sup>hi</sup> Vari068 BC cells as examined by Annexin V labeling and flow cytometry, related to Figure 4.

(**B-C**) Measurement of cellular superoxide levels in SUM159 (**B**) and SUM149 (**C**) BC cells treated with vehicle (DMSO) and different doses of IB-DNQ alone or together with ATN224. \*, \*\*, \*\*\*, \*\*\*\*: P< 0.05, 0.01, 0.001 and 0.0001 vs. Control or indicated with brackets (n=3). Error bars denote SEM.

(**D-F**) Vari068 (**D**), SUM159 (**E**) and SUM149 (**F**) BC cells treated with different doses of IB-DNQ alone or IB-DNQ plus ATN224 were stained with MitoSOX red (2.5  $\mu$ M) and mean fluorescence intensity (MFI) of MitoSOX red was analyzed in each condition by flow cytometry, related to Figure 5.



Β

	SUM159							
DMSO	+	-	-	-	-	-	-	-
ATN224 5 μM	-	+	-	-	-	+	+	+
IB-DNQ 50 nM	-	-	+	-	-	+	-	-
IB-DNQ 100 nM	-	-	-	+	-	-	+	-
IB-DNQ 200 nM	-	-	-	-	+	-	-	+
Casp.3	-	-	-	-	-	-	-	-
Clevd Casp.3		-	ALL ST	部分	-14	12.3		-
PARP	-	-	-	-	-	-	-	-
Clevd PARP	10000		Friend	-	-	-	-	-
BCL-2	-	-	-	-	-	-	-	-
					-		-	_
BCL-XL	-	-	-	-	-	-	-	
BIM	111	-	-	-	-	-	-	-
						_	-	-
β-Actin	-	-	-	-	-	-	-	-

Relative Survival to CTL ₅

С



D CTL IB 100 nM IB 200 nM +DEAB +DEAB \*+DEAB SS01-A :: SSC 488\_10-A SS01-A :: SSC 488\_10-/ SSC-A 0.03% 0.0<u>3%</u> 0.03% -DEAB -DEAB -DEAB SS01-A :: SSC 488\_10-A SS01-A :: SSC 488\_10-A SSC-A 100 8.63% 0.64% 28.9% BAAA BAAA BAAA FMK+IB200 FMK FMK+IB100 +DEAB +DEAB +DEAB SS01-A :: SSC 488\_10-A SS01-A :: SSC 488\_10-A 150) SSC-A 0.03% 0.03% 0.03% -DEAB -DEAB -DEAB 200K SS01-A :: SSC 488\_10-A SS01-A :: SSC 488\_10-A SSC-A 100 K 29.8% 18.1% 1.56% BAAA BAAA BAAA



## Figure S7. IB-DNQ induced cell death and abrogation of ALDH+ BCSCs in SUM159 is rescued by Casp 3 inhibitor Z-DEVD-FMK, related to Figure 6.

(A) SUM159 BC cells treated with 300 nM of IB-DNQ for 2h were examined by CCK8 assay (upper panel) and optical microscopy (lower panels) to examine the viability of SUM159 BC cells (n=3). ns: not significantly different.

(**B**) SUM159 BC cells were treated with vehicle, ATN224 (5  $\mu$ M), and 50-200 nM of IB-DNQ alone or in combination with ATN224 for 20h, and subjected to immunoblotting to examine the activation of Casp3 and the cleavage of its substrate PARP, as well as the expression of anti- and pro-apoptotic BCL2 family proteins.

(**C**) SUM159 BC cells treated with CTL(DMSO), Z-DEVD-FMK (50  $\mu$ M), Ferrostatin-1 (5  $\mu$ M) or IB-DNQ (100-300nM) alone, or combination of IB-DNQ with Z-DEVD-FMK or Ferrostatin-1 for 4h, and relative suvival of SUM159 BC were measured with MTT assay (n=6). \*\*, \*\*\*\*: P< 0.01 or 0.0001 (vs. CTL or indicated with brackets). Data are representatives of two independent experiments.

(**D**) SUM159 BC cells treated with Mock (DMSO), Z-DEVD-FMK (50 μM), IB-DNQ (100-200nM) alone, or combination of IB-DNQ and Z-DEVD-FMK for 3h and examined for ALDH+ CSCs by Aldefluor assay and flow cytometry (left panel). The proprotion of ALDH+ CSCs in each condition was plotted based on 3 independent experiments (right panel). \*\*\*, \*\*\*\*: P< 0.001 and 0.0001 respectively vs. CTL (DMSO). Error bars denote SD (stardard deviations). DEAB: the inhibitor of ALDH enzymatic activity. BAAA: the substrate of ALDH enzymatic activity.



# Figure S8. Dox-inducied KD of SOD1 suppresses tumor growth of SUM149 BC cells and IB-DNQ treatment further enhances tumor growth retardation of SOD1 KD cells, related to Figure 7.

(A) SUM149 BC cells expressing Dox-inducible KD (shSOD1) or scrambled control (SCR) were injected into the #4 MFP of 7-wk-old female SCID mice, and mice bearing mammary tumors (3-4 mm in diameter) expressing shSOD1 or SCR sequence were each randomized into two groups (6 mice/group) and subject to treatment with Dox (200  $\mu$ g/ml in water) plus vehicle (20% HP $\beta$ CD, I.V.) or IB-DNQ (10 mg/kg, I.V.) for 3 weeks.

(**B**) Tumor growth of SUM149 BC cells expressing SCR or shSOD1 after treated with Dox (200 µg/ml in water) plus vehicle or IB-DNQ. \*\*, \*\*\*\*: P< 0.01 or 0.0001 respectively (n=6, two-way ANOVA). Error bars denote SD.

#### **KEY RESOURCES TABLE**

REAGENT or RESOURCE	SOURCE	IDENTIFIER
Antibodies and Labeling Reagents		
Anti-ALDH1A1	Invitrogen	Cat# MA5-29023 RRID: AB 2784960
Anti-ALDH1A3	Invitrogen	Cat# PA5-29188 RRID: AB_2546664
Anti-BCL-2	Cell Signaling Technology	Cat# 15071S
Anti-BCL-xL	Cell Signaling Technology	Cat# 2764S
Anti-β-Actin	Cell Signaling Technology	Cat# 4970S
Anti-Bim	Cell Signaling Technology	Cat# 2933S
Anti-Caspase-3	Cell Signaling Technology	Cat# 9662S
Anti-Cleaved Caspase-3	Cell Signaling Technology	Cat# 9661S
Anti-Catalase	Cell Signaling Technology	Cat# 12980S
Anti-Cytochrome c	Cell Signaling Technology	Cat# 12963S
Anti-Ki67	Cell Signaling Technology	Cat# 12202S
Anti-NQO1	Cell Signaling Technology	Cat# 62262S
Anti NRF2	Cell Signaling Technology	Cat# 12721S
Anti-p-H2A.X	Cell Signaling Technology	Cat# 2577S
Anti-PARP	Cell Signaling Technology	Cat# 9542S
Anti-Cleaved PARP	Cell Signaling Technology	Cat# 5625S
Anti-SDHA	Cell Signaling Technology	Cat# 5839S
Anti-Skp1	Cell Signaling Technology	Cat# 2156S

Anti-SOD1	Cell Signaling	Cat# 37385S
	Call Signating	C-+# 42((S
Anti-SODI	Cell Signaling	Cat# 42005
	Call Signating	<u>C-+# 121410</u>
Anti-SOD2	Cell Signaling	Cat# 131418
	Call Signating	
Anti-vDAC	Cell Signaling	Cat# 40015
A set: D all it Is C UDD I is last A set it a last	Call Signating	C-+# 70749
Anti-Rabbit IgG, HRP Linked Antibody	Technology	Cal# /0/45
And Margare L.C. LIDD Linder 1 And her	Call Signating	0-+# 707(9
Anti-Mouse IgG, HRP Linked Antibody	Cell Signaling	Cat# /0/65
ADC many Anti langun CD44	DD Diagoing and	C-+# 550042
APC mouse Anti-numan CD44	BD Biosciences	Cat# 559942
		KRID:
ADC anti mana II 2V[4]	DisLagand	AB_398083
APC anti-mouse H-2K[d]	BioLegend	Cat# 110020
EITC Anti human CD24	DD Dissoisson	AB_10043328
FIIC Anti-numan CD24	BD Biosciences	Cal# 300992
		KKID:
EITC Anti human CD44	DD Dissoisson	AB_10302033
FIIC Anti-numan CD44	BD Biosciences	Cal# 3334/8
		RRID:AB_39387
DE anti mausa H 2V[d]	PD Diagoionago	0 Cot# 552566
FE anti-mouse H-2K[u]	BD Blosciences	
		$\begin{array}{c} \mathbf{K}\mathbf{K}\mathbf{I}\mathbf{D}, \\ \mathbf{A}\mathbf{D}, 204024 \end{array}$
DE/Cy/7 Anti human CD24	PD Piosoionoos	AD_394924
r E/Cy/ Anti-Indinan CD24	BD Biosciences	$\Delta P = 10802826$
DE/Cy/7 Anti human CD24	Piol agand	$\frac{AD_{10092020}}{C_{0}t\# 211120}$
FE/Cy/ Anti-numan CD24	BioLegend	AB 22508/3
	Sigma Aldrich	$AD_{2239043}$
ADC Approvin V	D Diagoionago	Cat# D9342
	BD Blosciences	$\Delta D 286885$
CallPOV® Oranga Paggant	Thormo Fisher Scientifie	$AD_{2000003}$
MitoSOVIM Red	Thermo Fisher Scientific	Cat# C10443
Chamicala Dantidas and Decembinant Dratains	Thermo Pisher Scientific	Cat# 14150000
Chemicals, Peptides, and Recombinant Proteins		
ATN224	Cayman Chemical	Cat# 23553
		CAS: 649749-10-0
3-AT	Sigma-Aldrich	Cat# A8056
		CAS: 61-82-5
Dicoumarol	Sigma-Aldrich	Cat# 2287897
		CAS: 66-76-2
Doxycycline	Sigma-Aldrich	Cat# D3072
		CAS: 10592-13-9

IB-DNQ	Provided by Dr. Paul J.	N/A
	Hergenrother	
β-Lapachone	MedChemExpress	Cat# HY-13555
	(MCE)	CAS: 4707-32-8
LCS-1	MedChemExpress	Cat# HY-115445
	(MCE)	CAS: <u>41931-13-9</u>
Z-DEVD-FMK (Caspase-3 Inhibitor)	Selleck Chemicals	Cat#: S7312
		CAS: 210344-95-9
Ferrostatin-I	Selleck Chemicals	Cat# : S7243
Materia 10 Decement Manufactor Materia		CAS: 34/1/4-05-4
Matrigel® Basement Membrane Matrix	Corning Life Sciences	Cat# 354234
Critical Commercial Assays		~ // 0.1 = 0.0
Aldefluor Assay kit	STEMCELL	Cat# 01700
	lechnologies	
In Situ Cell Death Detection Kit (TUNEL assay)	Roche	Cat#11684795910
Image-iT <sup>TM</sup> TMRM Kit	Invitrogen	Cat#I34361
Mitochondria Isolation Kit	Sigma-Aldrich	Cat# 89874
Extracellular Oxygen Consumption Assay kit	Abcam	ab197243
Cellular Superoxide Detection Assay kit	Abcam	ab139477
ATP Assay Kit	Beyotime Biotechnology	S0026
MammoCult Human Medium Kit (for human	STEMCELL	Cat# 05620
tumorsphere culture)	Technologies	
Experimental Models: Cell Lines		
Human: HCC1937	ATCC	CRL-2336
Human: HCC1806	ATCC	CRL-2335
Human: HCC70	ATCC	CDI 2215
		CRL-2315
Human: HCC38	ATCC	CRL-2315 CRL-2314
Human: HCC38 Human: BT20	ATCC ATCC	CRL-2315 CRL-2314 HTB-19
Human: HCC38 Human: BT20 Human: SUM149 and SUM159	ATCC ATCC Stephen P. Ethier, Ph.D.	CRL-2315 CRL-2314 HTB-19 Medical
Human: HCC38 Human: BT20 Human: SUM149 and SUM159	ATCC ATCC Stephen P. Ethier, Ph.D.	CRL-2315 CRL-2314 HTB-19 Medical University of
Human: HCC38 Human: BT20 Human: SUM149 and SUM159	ATCC ATCC Stephen P. Ethier, Ph.D.	CRL-2315 CRL-2314 HTB-19 Medical University of South Carolina
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Human: HCC38 Human: BT20 Human: SUM149 and SUM159 MDA-MB-231 NQO1 <sup>+</sup> and MDA-MB-231 NQO1 <sup>-</sup> isogenic TNBC cell lines	ATCC ATCC Stephen P. Ethier, Ph.D. Donated by the late Dr. David A. Boothman.	CRL-2313 CRL-2314 HTB-19 Medical University of South Carolina Indiana University School of
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Human: HCC38 Human: BT20 Human: SUM149 and SUM159 MDA-MB-231 NQO1 <sup>+</sup> and MDA-MB-231 NQO1 <sup>-</sup> isogenic TNBC cell lines	ATCC ATCC Stephen P. Ethier, Ph.D. Donated by the late Dr. David A. Boothman.	CRL-2313 CRL-2314 HTB-19 Medical University of South Carolina Indiana University School of Medicine, Indianapolis, IN 46202, USA
Human: HCC38 Human: BT20 Human: SUM149 and SUM159 MDA-MB-231 NQO1 <sup>+</sup> and MDA-MB-231 NQO1 <sup>-</sup> isogenic TNBC cell lines Vari068 TNBC cell line, derived from patient-	ATCC ATCC Stephen P. Ethier, Ph.D. Donated by the late Dr. David A. Boothman.	CRL-2313 CRL-2314 HTB-19 Medical University of South Carolina Indiana University School of Medicine, Indianapolis, IN 46202, USA N/A
Human: HCC38 Human: BT20 Human: SUM149 and SUM159 MDA-MB-231 <i>NQO1</i> <sup>+</sup> and MDA-MB-231 <i>NQO1</i> <sup>-</sup> isogenic TNBC cell lines Vari068 TNBC cell line, derived from patient- derived xenograft tumors.	ATCC ATCC Stephen P. Ethier, Ph.D. Donated by the late Dr. David A. Boothman.	CRL-2313 CRL-2314 HTB-19 Medical University of South Carolina Indiana University School of Medicine, Indianapolis, IN 46202, USA N/A
Human: HCC38   Human: BT20   Human: SUM149 and SUM159   MDA-MB-231 NQO1 <sup>+</sup> and MDA-MB-231 NQO1 <sup>-</sup> isogenic TNBC cell lines   Vari068 TNBC cell line, derived from patient-derived xenograft tumors.   Human: Hs578t	ATCC ATCC Stephen P. Ethier, Ph.D. Donated by the late Dr. David A. Boothman. University of Michigan	CRL-2313 CRL-2314 HTB-19 Medical University of South Carolina Indiana University School of Medicine, Indianapolis, IN 46202, USA N/A HTB-126
Human: HCC38   Human: BT20   Human: SUM149 and SUM159   MDA-MB-231 NQO1 <sup>+</sup> and MDA-MB-231 NQO1 <sup>-</sup> isogenic TNBC cell lines   Vari068 TNBC cell line, derived from patient-derived xenograft tumors.   Human: Hs578t   Human: MDA-MB-157	ATCC ATCC Stephen P. Ethier, Ph.D. Donated by the late Dr. David A. Boothman. University of Michigan ATCC ATCC	CRL-2313 CRL-2314 HTB-19 Medical University of South Carolina Indiana University School of Medicine, Indianapolis, IN 46202, USA N/A HTB-126 HTB-24
Human: HCC38   Human: BT20   Human: SUM149 and SUM159   MDA-MB-231 NQO1 <sup>+</sup> and MDA-MB-231 NQO1 <sup>-</sup> isogenic TNBC cell lines   Vari068 TNBC cell line, derived from patient-derived xenograft tumors.   Human: Hs578t   Human: MDA-MB-157   Human: MDA-MB-231	ATCC ATCC Stephen P. Ethier, Ph.D. Donated by the late Dr. David A. Boothman. University of Michigan ATCC ATCC ATCC	CRL-2313 CRL-2314 HTB-19 Medical University of South Carolina Indiana University School of Medicine, Indianapolis, IN 46202, USA N/A HTB-126 HTB-24 CRM-HTB-26

Human: MDA-MB-468	ATCC	HTB-132
Human: MCF7	ATCC	HTB-22
Human: ZR-75-1	ATCC	CRL-1500
Human: MCF10A	ATCC	CRL-10317
Experimental Models: Organisms/Strains		
Mouse strain: Fox Chase SCID	Charles River	Strain Code: 236
	Laboratories	
Recombinant DNA		
TRIPZ Inducible Lentiviral Human NQO1 shRNA	Horizon Discovery	V2THS_235287
TRIPZ Inducible Lentiviral Human SOD1 shRNA	Horizon Discovery	V3THS_369718
TRIPZ Inducible Lentiviral Human SOD2 shRNA	Horizon Discovery	V3THS_307804
TRIPZ Inducible Lentiviral Human CAT shRNA	Horizon Discovery	V2THS_150247
SMARTvector Inducible Lentiviral Human NQO1 shRNA	Horizon Discovery	V3IHSHER_1077 1835
SMARTvector Inducible Lentiviral Human NQO1 shRNA	Horizon Discovery	V3IHSHER_7177 739
SMARTvector Inducible Lentiviral Human NQO1 shRNA	Horizon Discovery	V3IHSHER_8909 942
SMARTvector Inducible Lentiviral Human SOD1 shRNA	Horizon Discovery	V3IHSHER_9961 124
SMARTvector Inducible Lentiviral Human SOD1 shRNA	Horizon Discovery	V3IHSHER_9099 989
TRIPZ Inducible Lentiviral shRNA Scrambled Control	University of Michigan vector core	N/A
Software and Algorithms	-	-
ELDA	WEHI Bioinformatics Resources	http://bioinf.wehi. edu.au/software/el da/index.html
FlowJo	FlowJo, LLC	https://www.flowj o.com/solutions/fl owjo/
GraphPad Prism	GraphPad Software Inc	http://www.graph pad.com/scientific -software/prism/
ImageJ	NIH	https://imagej.nih. gov/ij/
Compusyn	ComboSyn, Inc.	https://www.comb osyn.com