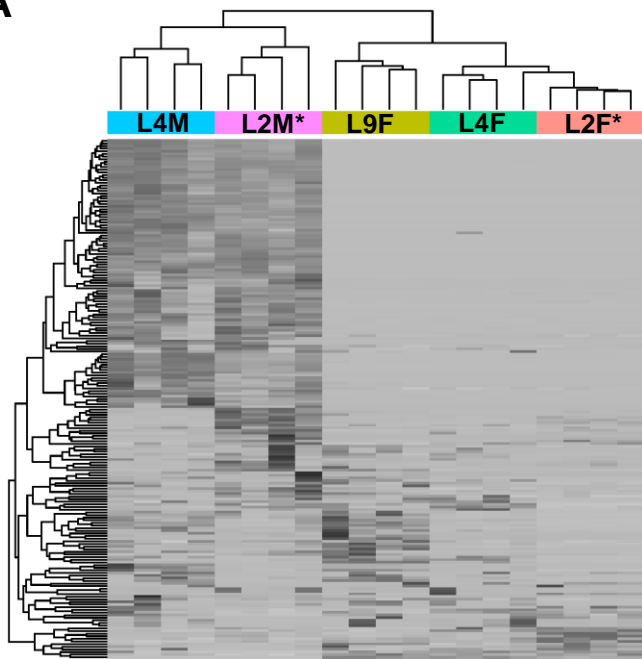
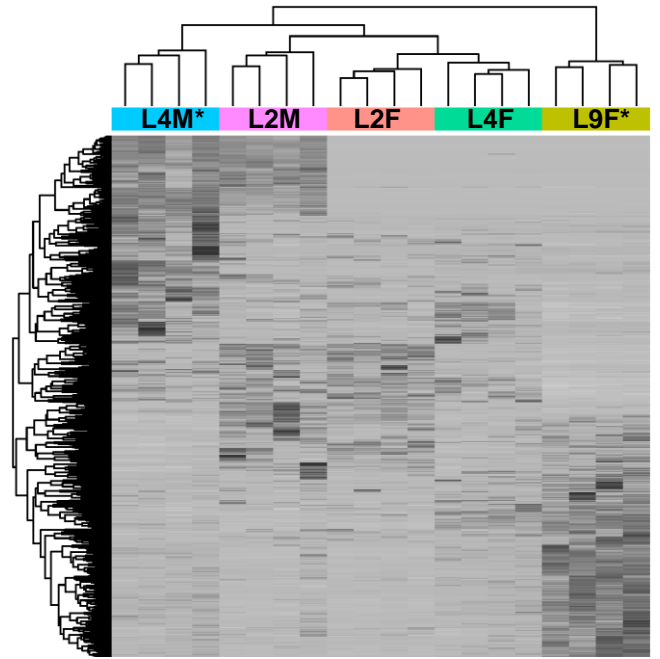


**Supplementary Fig. S1. Sequence comparison of ChrY transcripts to ‘CBDRx’ and ‘FINOLA’ genomes.** Putative ChrY transcripts were compared against the female *C. sativa* reference genome ‘CBDRx’ (GCF\_900626175.2) (Grassa et al., 2021) using BLAST, with sequence identities as fractions (A) and the match lengths (B) in base pairs are displayed as bar charts. Subsequently, putative ChrY transcripts were compared against the male genome of the *C. sativa* hemp cultivar ‘FINOLA’ (ASM341772v2) (Lavery et al., 2019), and sequence identity as fractions (C) and match lengths in base pairs (D) are presented as bar charts. To establish a baseline for similarity between ‘CBDRx’ and ‘FINOLA’ genomes, a set of randomised ‘CBDRx’ transcripts was BLASTed against the ‘FINOLA’ genome, with identity as fractions (E) and match lengths in base pairs (F) being displayed as bar charts.

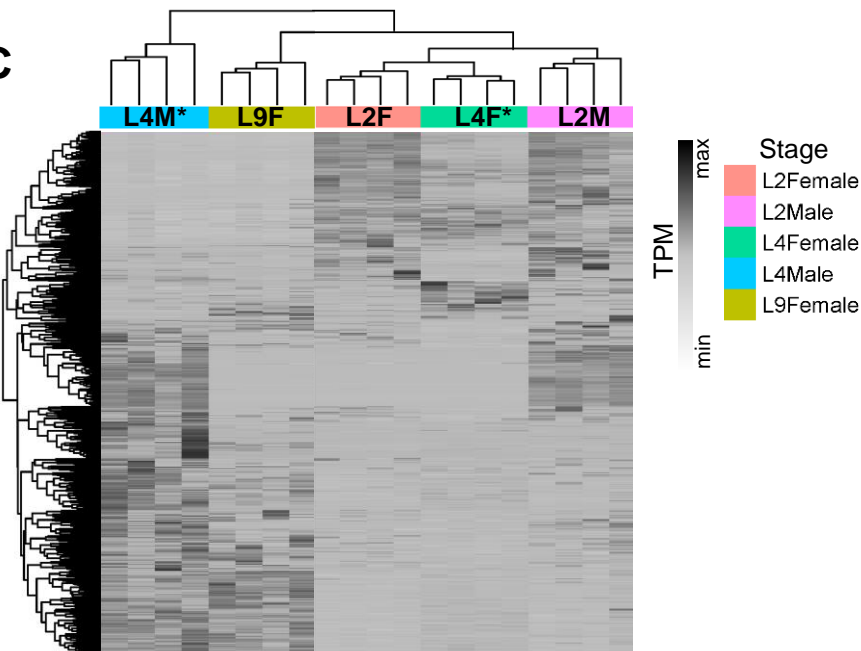
A



B

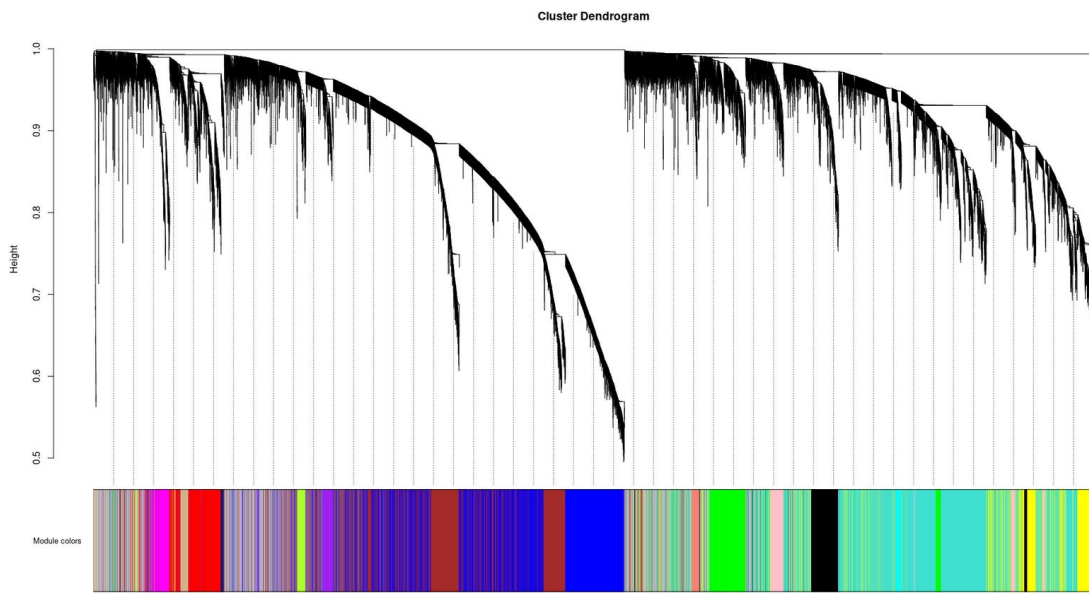


C



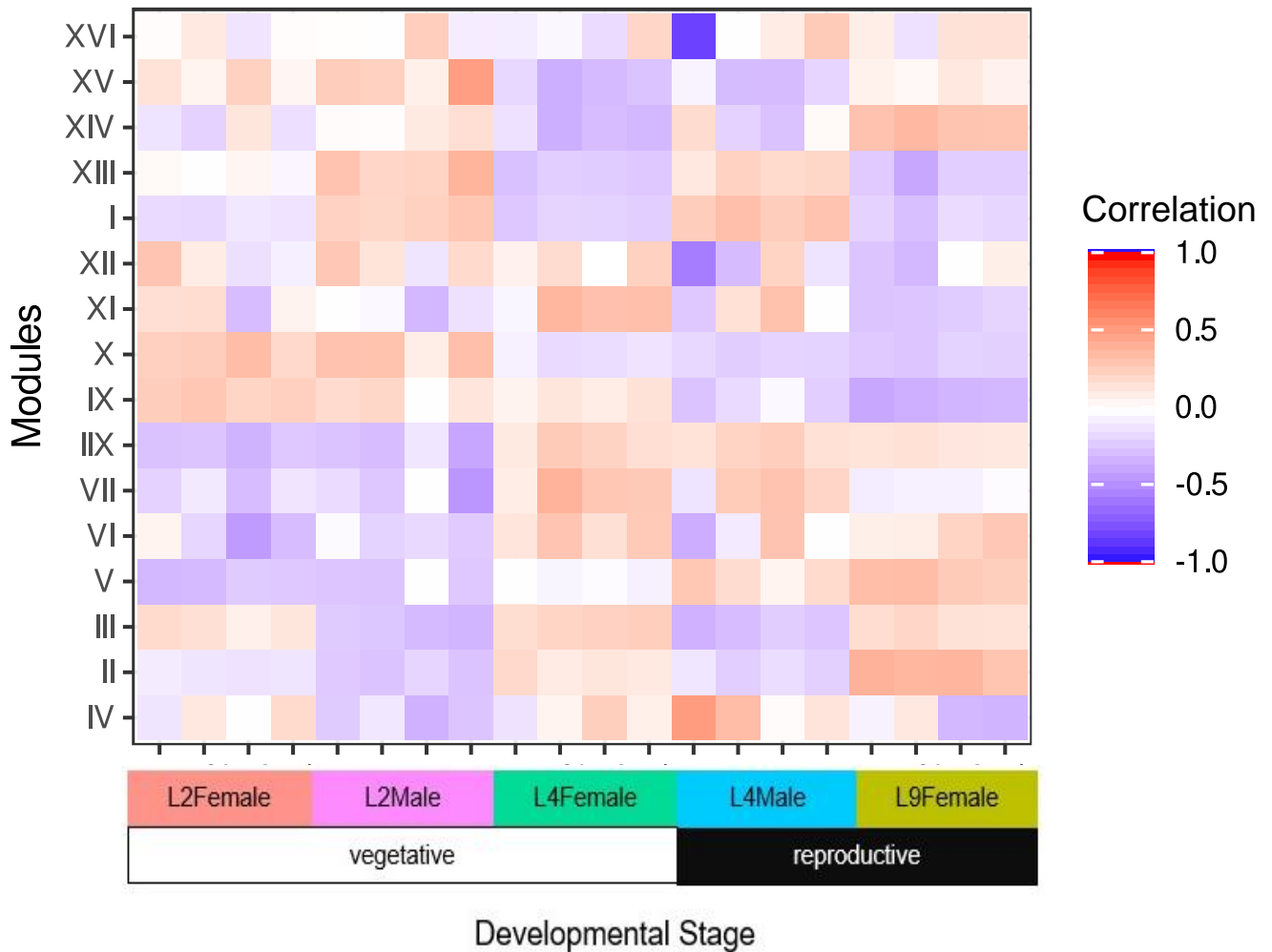
**Supplementary Fig. S2. Hierarchical clustering of differentially expressed genes (DEGs) in hemp.** DEGs identified in a comparison of L2M vs L2F (A), L4M vs L9F (B), L4M vs L4F (C), with bi-directional clustering based on the complete method performed according to developmental stage and expression level. Asterisks (\*) indicate the stages used to determine DEGs. Heatmaps present the relative expression levels of each transcript (row) in each plant sample (column). The expression values were normalised using TPM values. The grey scale, based on a relative color scheme, represents high expression (dark black) and low expression (light grey) of the transcripts across samples. The lightest and darkest colours are converted using the minimum and maximum values in each row.

A

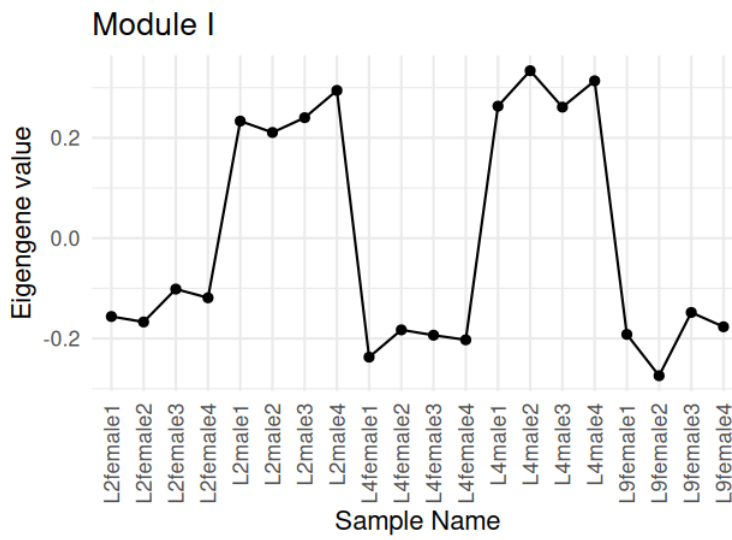
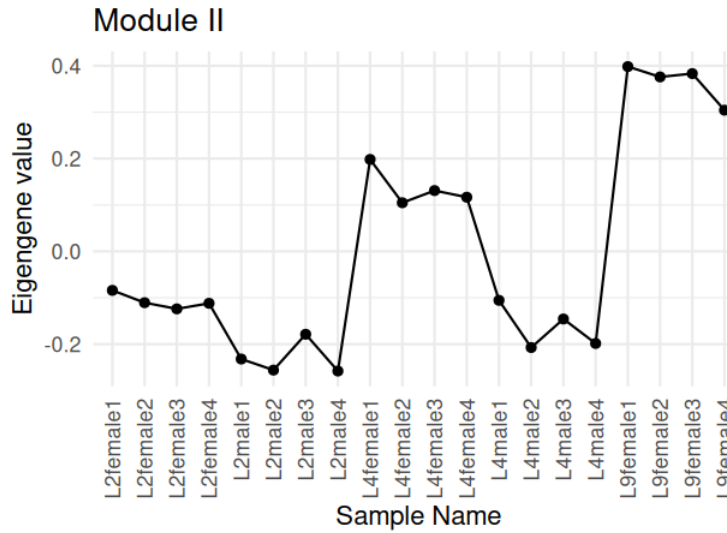
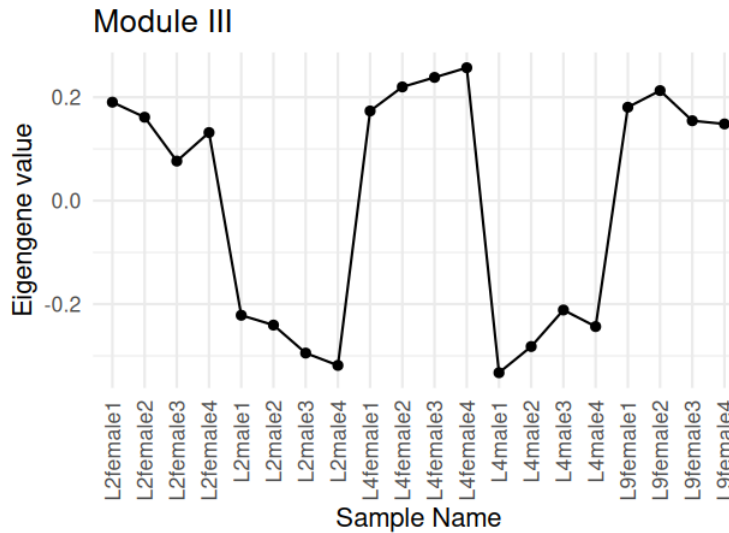


## Module-trait Relationships

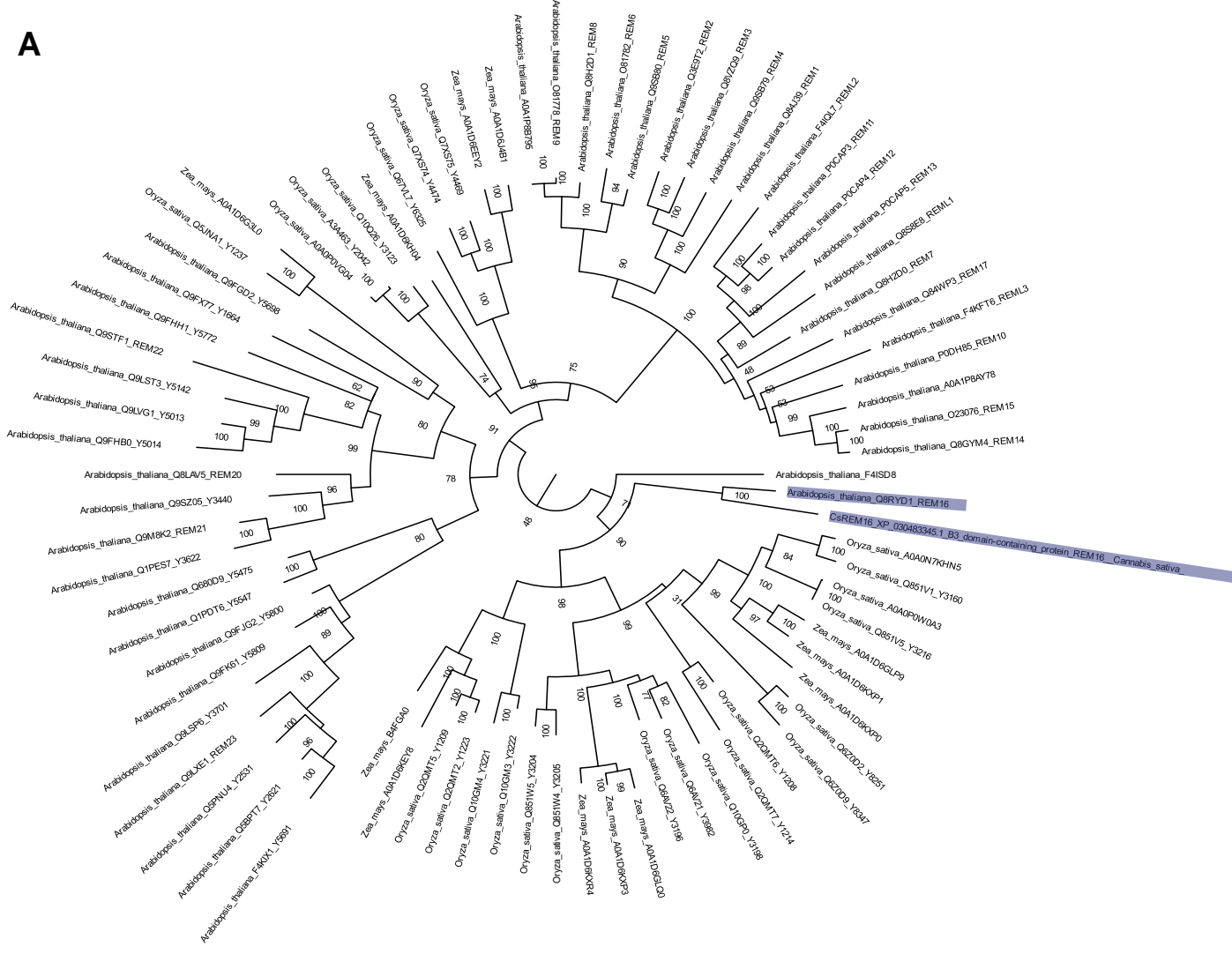
B



**Supplementary Fig. S3. Modules of co-expressed genes identified by Weighted Gene Co-expression Network Analysis (WGCNA).** A cluster dendrogram of all modules identified with WGCNA (A). The colour row underneath the dendrogram shows the determined module assignment. A heatmap of average expression per sample in all modules (B). The blue colour represents a negative correlation, while the red represents a positive correlation. A complete list of all genes can be found in Supplementary Table S5.

**A****B****C**

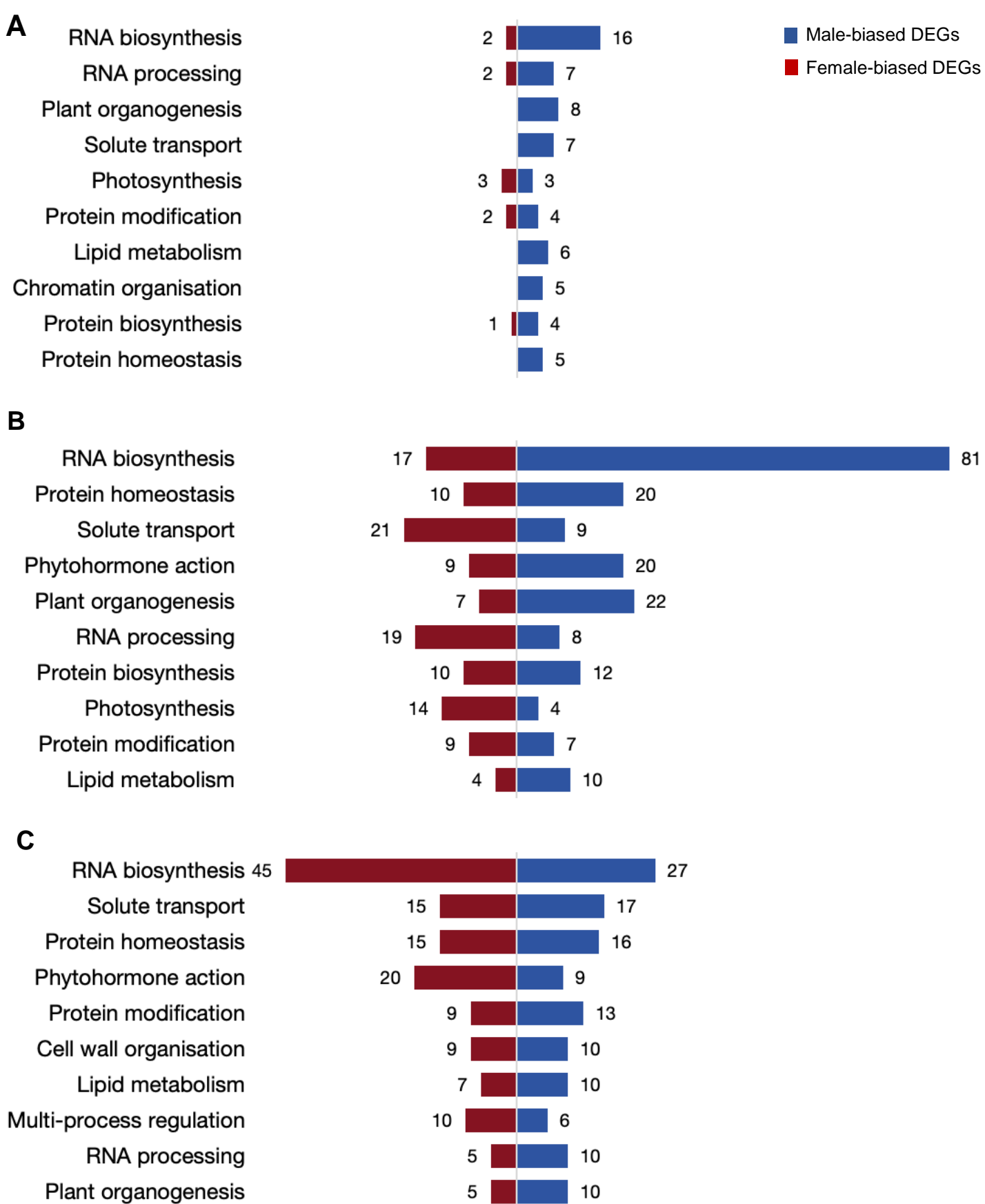
**Supplementary Fig. S4. The eigengene values for the sex-related modules.** Plot of eigengene values summarising the activity of the three modules with clear sex correlation, obtained by taking the first principal component of expression level of genes in that module. Module I (A) shows clear activity in male samples. While modules II (B) and III (C) show correlation with late stage and all stages female samples respectively.

**A****B**

**Supplementary Fig. S5. SHOOT.bio homology association.** Phylogeny of *CsREM16* (LOC115699937), shows that *CsREM16* putative homolog in *A. thaliana* is Q8RYD1 (At4g33280) named *AtREM16* (A). Phylogeny of FE.chrY.t58, shows that its putative homolog in *A. thaliana* is AT5G42910, a gene in the same bZIP group A as *FD* (B).

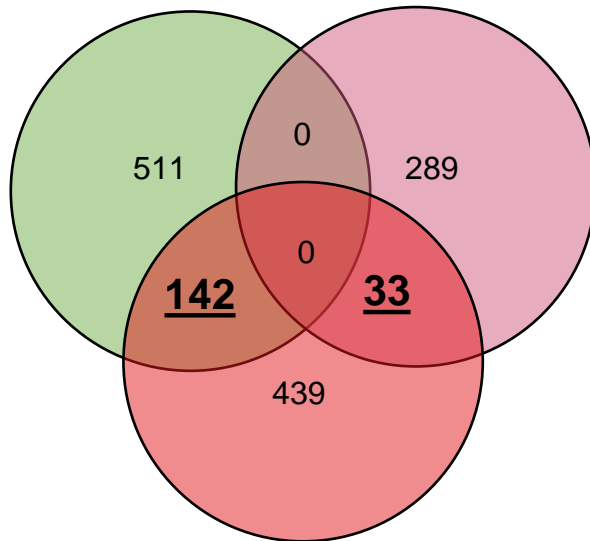


**Supplementary Fig. S6. Summary of MapMan metabolic pathways from the DEGs.** Differentially expressed genes identified through comparison of L2M vs L2F, L4M vs L4F and L4M vs L9F were analysed using MapMan and assigned to metabolic pathway bins: 1- photosynthesis, 2- cellular respiration, 3- carbohydrate metabolism, 4- amino acid metabolism, 5- lipid metabolism, 6- nucleotide metabolism, 7- coenzyme metabolism, 8- polyamine metabolism, 9- secondary metabolism, 10- redox homeostasis, 11- phytohormone action, 12- chromatin organization, 13- cell cycle organization, 14- DNA damage response, 15- RNA biosynthesis, 16- RNA processing, 17- protein biosynthesis, 18- protein modification, 19- protein homeostasis, 20- cytoskeleton organization, 21- cell wall organization, 22- vesicle trafficking, 23- protein translocation, 24- solute transport, 25- nutrient uptake, 26- external stimuli response, 27- multi-process regulation, 28- plant reproduction, 29- plant organogenesis, 30- clade-specific metabolism. Expression values are log2fold changes (log2FC) with colour scales of red (female-biased), blue (male-biased), and white indicating log2fold change values close to 0.



**Supplementary Fig. S7. The top 10 MapMan metabolic pathways from the DEGs in hemp.** To better identify the functional pathway response to sex determination, the top ten metabolic pathway bins, which contain the largest number of genes, are presented in butterfly plots for DEGs from L2M vs L2F (A), L4M vs L4F (B), and L4M vs L9F (C), respectively.

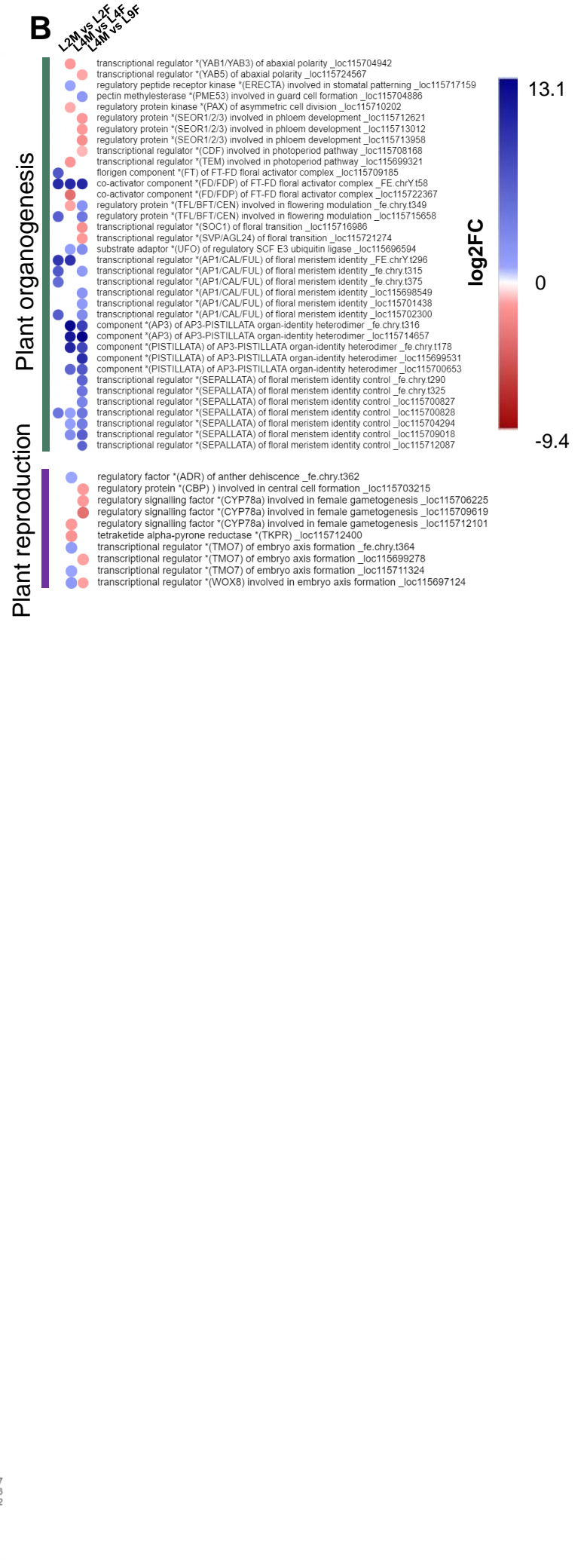
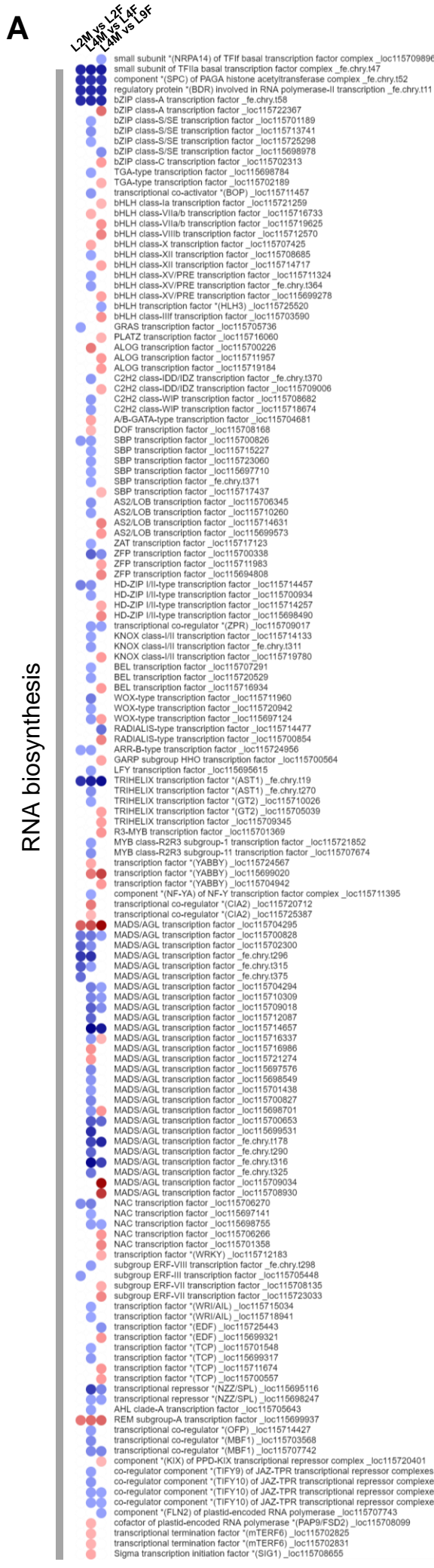
Male specific module I Female module II and III



DEGs (L4M vs L9F)

**Supplementary Fig. S8. Identification of putative candidate genes for flower development in hemp.** The Venn diagram illustrates the number of overlapping genes between DEGs from the L4M vs L9F comparison and the male-specific or female-specific module from WGCNA results.





**Supplementary Fig. S9. Differentially expressed genes potentially involved in sex determination and floral development.** Differential expression between L2F vs L2F, L4M vs L4F and L4MF vs L9F is indicated for genes linked to RNA biosynthesis (A), or and plant organogenesis and plant reproduction (B) as identified through MapMan analysis. The relative expression of each gene (row) in each comparison (column) is shown. Expression values are log<sub>2</sub>fold changes (log<sub>2</sub>FC) with colour scales of red (female-biased) and blue (male-biased).

# A Module I

- component \*(NRPD2) of RNA polymerase IV complex \_fe.chry.t126
- component \*(NRPE2) of RNA polymerase V complex \_fe.chry.t126
- small subunit of TFIIa basal transcription factor complex \_fe.chry.t47
- subunit beta of TFIIe basal transcription factor complex \_loc115709141
- component \*(SCS1/SCS2) of HAT histone acetyltransferase module \_loc115721375
- component \*(ADA1) of SAGA transcription co-activator complex \_fe.chry.t40
- component \*(ADA1) of SAGA transcription co-activator complex \_loc115715332
- component \*(SPC) of PAGA histone acetyltransferase complex \_fe.chry.t52
- component \*(MED18) of head module of MEDIATOR transcription co-activator complex \_loc115700285
- regulatory protein \*(BDR) involved in RNA polymerase-II transcription \_fe.chry.t11
- bZIP class-A transcription factor \_fe.chry.t58
- bZIP class-A transcription factor \_loc115702933
- bZIP class-S/SE transcription factor \_loc115698978
- bZIP class-S/SE transcription factor \_loc115725298
- bHLH class-IVa transcription factor \_loc115712227
- bHLH class-IIIa transcription factor \_loc115718651
- BSD-type transcription factor \_loc115712016
- C2H2 class-WIP transcription factor \_loc115709101
- C2H2 class-WIP transcription factor \_loc115721385
- DOF transcription factor \_loc115697076
- AS2/LOB transcription factor \_loc115705047
- AS2/LOB transcription factor \_loc115705389
- zf-HD-type transcription factor \_loc115700466
- HSF transcription factor \_loc115716259
- HSF transcription factor \_loc115722192
- TRIHELIX transcription factor \*(AST1) \_fe.chry.t19
- MYB class-R2R3 subgroup-1 transcription factor \_loc115710324
- MYB class-R2R3 subgroup-13 transcription factor \_loc115711659
- MADS/AGL transcription factor \_loc115700653
- MADS/AGL transcription factor \_loc115714657
- NAC transcription factor \_loc115709772
- NAC transcription factor \_loc115723252
- VOZ transcription factor \_loc115699585
- subgroup ERF-VIII transcription factor \_loc115723530
- subgroup ERF-III transcription factor \_loc115708230
- transcription factor \*(WRI/AIL) \_loc115718941
- transcriptional repressor \*(NZZ/SPL) \_loc115695116
- transcriptional repressor \*(NZZ/SPL) \_loc115698247
- AHL clade-A transcription factor \_loc115704801
- transcriptional co-regulator \*(MBF1) \_loc115707742
- co-regulator component \*(TIFY10) of JAZ-TPR transcriptional repressor complexes \_loc115707122
- regulatory protein \*(LAP3) of exine formation \_loc115724889
- fatty acyl-CoA reductase \*(MS2) \_loc115699209
- regulatory protein \*(ASL19/20) of xylem differentiation \_loc115705047
- cleavage and polyadenylation specificity factor \*(FY) \_fe.chry.t69
- florigen component \*(FT) of FT-FD floral activator complex \_loc115700781
- co-activator component \*(FD/FDP) of FT-FD floral activator complex \_fe.chry.t58
- regulatory protein \*(TFL/BFT/CEN) involved in flowering modulation \_loc115700781
- component \*(AP3) of AP3-PISTILLATA organ-identity heterodimer \_loc115714657
- component \*(PISTILLATA) of AP3-PISTILLATA organ-identity heterodimer \_loc115700653

■ RNA biosynthesis

■ Plant reproduction

■ Plant organogenesis

kWithin

1.2

129.2

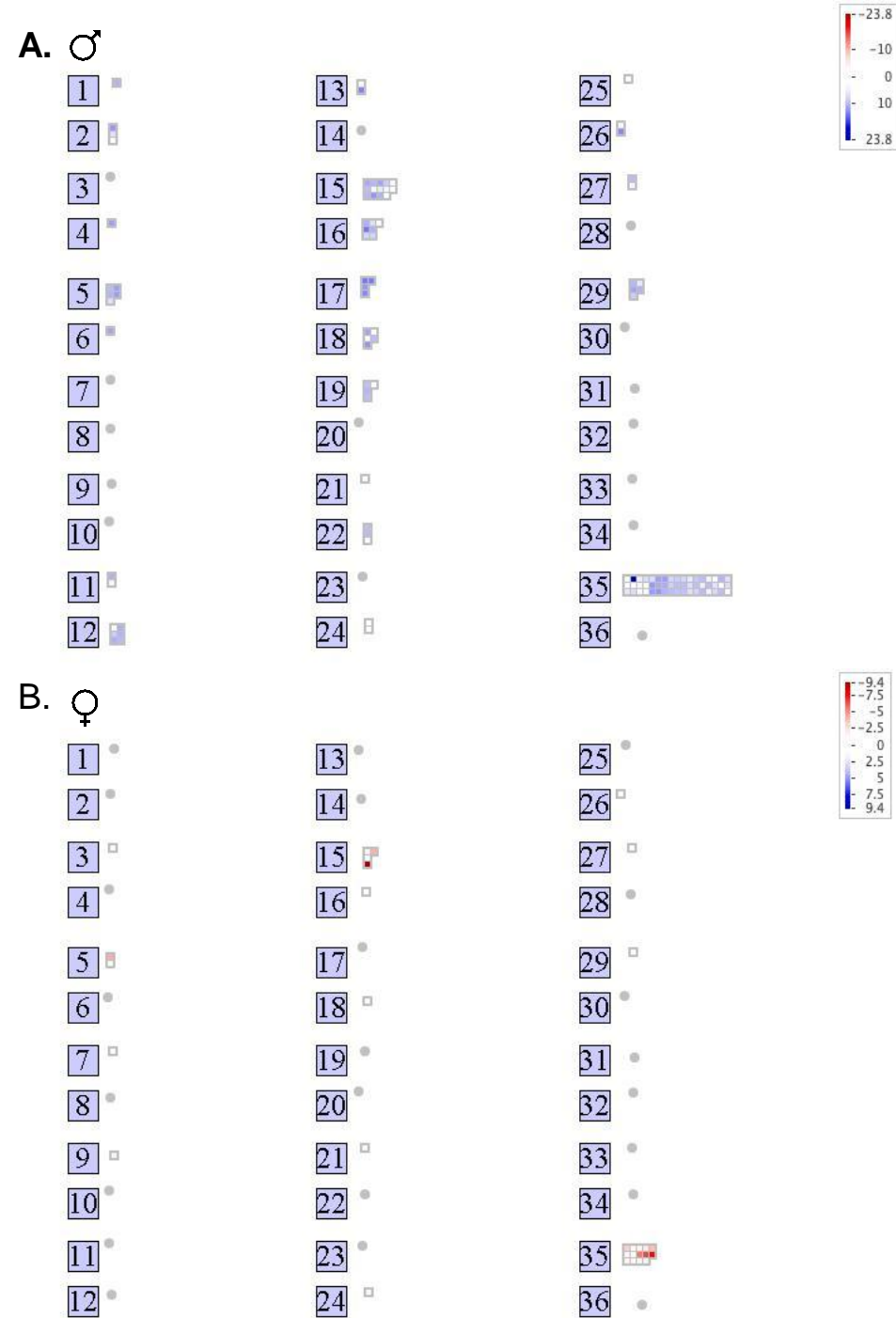
# B Module II

- bZIP class-C transcription factor \_loc115702313
- TGA-type transcription factor \_loc115699157
- bHLH class-X transcription factor \_loc115702403
- bHLH class-XII transcription factor \_loc115712856
- DBP phosphatase-domain transcription factor \_loc115724355
- ALOG transcription factor \_loc115697612
- C2H2 class-IDD/IDZ transcription factor \_loc115709006
- A/B-GATA-type transcription factor \_loc115704484
- ZFP transcription factor \_loc115722966
- HD-ZIP I/II-type transcription factor \_loc115700801
- zf-HD-type transcription factor \_loc115700881
- transcription factor \*(ARID3/4/6) \_loc115711694
- MADS/AGL transcription factor \_loc115704295
- FRS/FRF transcription factor \_loc115704634
- transcription factor \*(WRKY) \_loc115700154
- subgroup ERF-V transcription factor \_loc115703125
- transcription factor \*(TCP) \_loc115706624
- REM subgroup-A transcription factor \_loc115699925
- transcriptional co-regulator \*(TIFY) \_loc115699142
- component \*(LUG/LUH) of transcriptional co-repressor complex \_loc115702276
- adapter component \*(SEU/SLK) of transcriptional co-repressor complex \_loc115710310
- regulatory protein kinase \*(PAX) of asymmetric cell division \_loc115710202

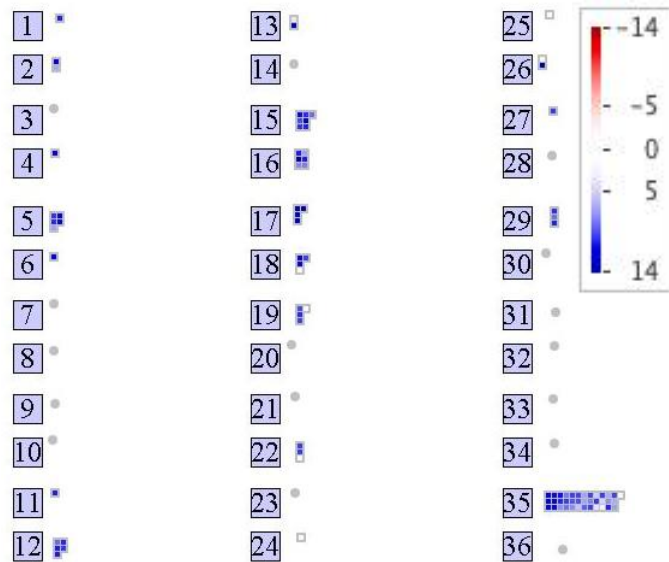
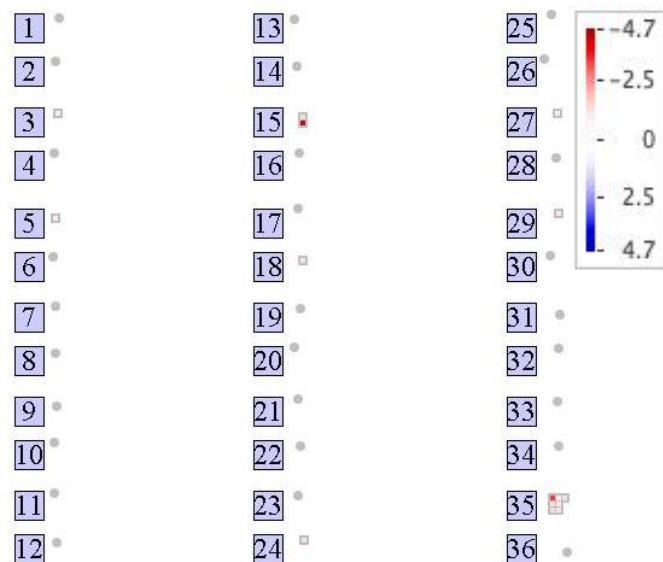
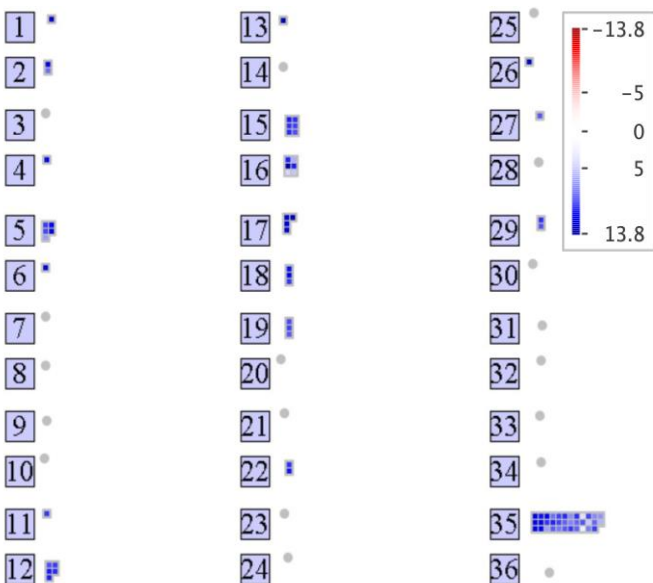
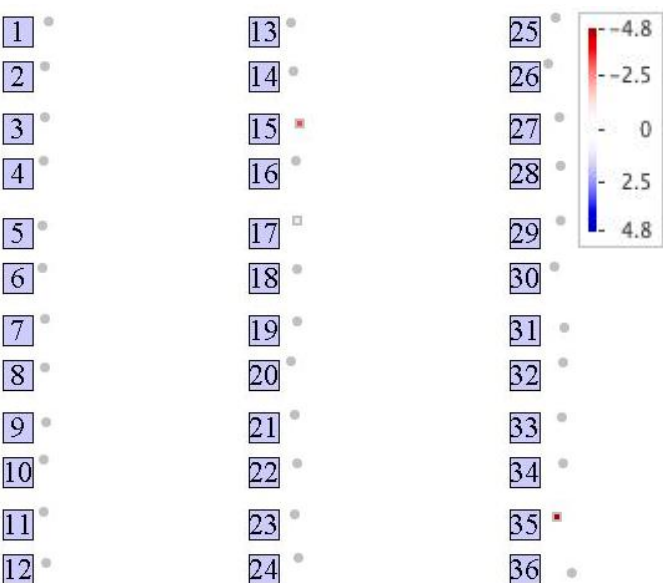
# C Module III

- component \*(SSL2/XPB) of TFIIh basal transcription factor complex \_loc115703047
- component \*(ADA1) of SAGA transcription co-activator complex \_loc115700092
- C-GATA-type transcription factor \_loc115697696
- C-GATA-type transcription factor \_loc115697707
- TRIHELIX transcription factor \*(AST1) \_loc115710018
- transcription factor \*(WRKY) \_loc115706387
- REM subgroup-A transcription factor \_loc115699937
- cleavage and polyadenylation specificity factor \*(FY) \_loc115724500

**Supplementary Fig. S10. Sex-related genes identified by WGCNA with potential function in RNA biosynthesis, plant organogenesis, and plant reproduction.** Genes included in module I (A), module II (B) and module III (C) were assigned to either RNA biosynthesis, plant organogenesis, and plant reproduction using MapMan. The colour indicates the kWithin value derived from the WGCNA. Light grey presents the lower value, and dark green presents the higher value which means a stronger correlation in the module.



**Supplementary Fig. S11. Functional annotation of putative candidate genes for flower development.** The MapMan functional annotation of the putative candidate genes for flower development. Genes that were significantly differentially expressed between L4M vs LF9, and highly correlated in WGCNA were considered putative candidates (see Figs. 6, 7). Candidate genes for flower development in male plants(A), candidate genes for flower development in female plants (B) were sorted into MapMan function bins. The colour bar represents the expression values as log<sub>2</sub> fold changes (log<sub>2</sub>FC) of female-related genes (red) and male-related genes (blue).

**A.****B.****C.****D.**

**Supplementary Fig. S12. Identification of putative candidate genes for early sex determination.** Genes that were significantly differentially expressed between L2M vs L2F, highly correlated in WGCNA and located on sex chromosomes were considered putative candidates (refer to Fig. 7). Putative gene for male sex identity in the flowering stage (A). Putative gene for female sex identity in the flowering stage (B). Putative gene for male sex identity in the vegetative stage (C). Putative genes for female sex identity in the vegetative stage (D). The colour bar represents the expression values as log<sub>2</sub> fold changes (log<sub>2</sub>FC) of female-related genes (red) and male-related genes (blue).