Supplementary Material

Cardiomyocyte proliferation and heart regeneration in adult *Xenopus tropicalis* evidenced by a transgenic reporter line

Xiao-Lin Lin, Jin-Hua Lin, Yan Cao, Han Zhang, Si-Yi He, Hai-Yan Wu, Ze-Bing Ye, Li Zheng, Xu-Feng Qi

Supplementary figures 1-4

Supplementary table 1

Supplementary video legends 1-2

Supplementary figure and legend



Supplementary Figure 1. Effects of reporter gene on heart development and function in transgenic reporter line Tg(mlc2:H2C). a Whole-mount bright-field (upper) and epifluorescence (lower) images showing mCherry expression in Wild-type (WT, left) and F1 Tg(mlc2:H2C) (right) tadpoles. Arrows indicate heart expressing mCherry. b qPCR validation of heart development-related genes in WT and Tg(mlc2:H2C) tadpoles. Data are presented as mean \pm SEM (n = 4 independent experiments). Ns, no significant difference (one-way ANOVA test). c Quantification of HW/BW ratio in adult frogs. Data are presented as mean \pm SEM ($n = 9 \sim 11$ frogs). Ns, no significant difference (Student's *t* test). This figure was created by Photoshop Image 12 software using our own data in the work.

Supplementary Figure 2. Accuracy evaluation of nuclear staining of cardiomyocytes in adult X. tropicalis. a Immunostaining for mCherry (red) and Mef2c (green) expression in a whole cardiac section from the adult heart of F1 Tg(mlc2:H2C) frog. DAPI was used as a nuclear stain (blue). **b** Magnified immunostaining image of the non-apical region white boxed in the whole cardiac section. c Magnified immunostaining image of the white boxed region in figure B showing that most mCherry-positive nuclei are co-labeled by Mef2c (mCherry⁺Mef2c⁺). Arrow denotes the mCherry⁺Mef2c⁻ cells. Arrowhead denotes mCherry⁻Mef2c⁺ cells with intranuclear expression of Mef2c. Triangle denotes mCherry⁻Mef2c⁺ cells with extranuclear expression of Mef2c. d Single- and double-channel fluorescence images of figure c for mCherry/DAPI (pink), mCherry (red), Mef2c (green), and mCherry/Mef2c (yellow) expression. eg Representative Z-stack confocal images of mCherry⁺Mef2c⁺ (e), mCherry⁺Mef2c⁻ (f), and mCherry⁻ Mef2c⁺ (g) cells in figure c. h and i Quantification of mCherry⁺ and mCherry⁺Mef2c⁺ cell numbers in adult hearts of F1 Tg(mlc2:H2C) frogs. Data are presented as mean \pm SEM ($n = \sim 150$ sections from 11 hearts for h, n=11 hearts for i). Ns, no significant difference (Student's t test). j Quantification of mCherry⁺Mef2c⁺ and mCherry⁺Mef2c⁻ cells percentages in adult hearts of F1 Tg(mlc2:H2C) frogs. Data are presented as mean \pm SEM (n = 11 hearts). k Quantification and comparation of mCherry⁺ α -actinin⁺ and mCherry⁺Mef2c⁺ cells percentages in adult hearts of F1 Tg(mlc2:H2C) frogs. Data are presented as mean \pm SEM (n = 16 and 11 hearts, respectively). ****p < 0.0001 (Student's t test).

Supplementary Figure 3. Evaluation of cardiomyocyte hypertrophy in adult reporter line during heart regeneration. a and b Representative WGA staining images (a) and quantification (b) of cardiomyocyte size located in the ventricular apex at the indicated time points. Data are presented as mean \pm SEM (n = 8 hearts, one-way ANOVA test). c qPCR validation of hypertrophic markers including *nppa*, *nppb*, and *myh7* in ventricles at the indicated time points. Data are presented as mean \pm SEM (n = 3 hearts, two-way ANOVA test).

Supplementary Figure 4. Determination of cardiomyocyte proliferation in adult Tg(mlc2:H2C) transgenic X. tropicalis line using antibody against pH3 (CST, #3377). a Immunostaining for mCherry (red) and pH3 (green, antibody from CST, #3377) expression in the apex of adult hearts from F1 Tg(mlc2:H2C) frogs at the indicated time points after resection. DAPI was used as a nuclear stain (blue). b Quantification of mCherry⁺pH3⁺ cells in ventricular apex during heart regeneration within 30 days. Data are presented as mean \pm SEM (n = 3 hearts for sham, $5 \sim 8$ hearts for injured groups). ****p < 0.0001 versus sham, ####p < 0.0001 (one-way ANOVA test). c Representative images of mCherry⁺pH3⁺ cells in the whole apical regions at 7 and 30 dpr. d Magnified Z-stack confocal images of mCherry⁺pH3⁺ cells indicated by dashed line in Figure c are shown.

Genes	Primer sequence (5'-3')	
	Forward primer	Reverse primer
nppa	CAGTCCTGCATACAGCTC	AAGCATCGGCAACATCA
nppb	CAGCCCACCCAGTTATGGA	ATTTGTAGCATCTGCGTCC
myh7	TTCATTGACTTCGGCATGGA	AATGAAGTGTCTGTTGCCTT
tbx5	GGGTCAGTAGCACCTCC	TCCAGAACGATAGTAAGGGT
tbx20	GCTATGGGACAAATTCCATG	TTTTCCAGCCACCAACC
tnni3	TGTCCGGCTTGTCCCTA	ACTTTAGCCTCCATGTCGT
tnnt2	AAAAGATCTTACTGAACTGC	GTTCAGCTCGTCTCTTC
hand1	AAGACCCTAAGACTAGCCAC	CCGCACCCCAATATCCTTC
hand2	ACCTGCTGGCCAAGGAC	CCAGACATGTTGCGGCCAA
bmp4	TAACACCGTGAGGAGCTTC	ATAGAGTCTTAGTTCTGCT
fbrsl1	AAAGAAGGACCTTTTGCT	CAATGTCTTTGGAGCGAT
Odc	AGGCCACACTGGCAACTCA	TGCGCTCAGTTCTGGTACTTCA

Supplementary Table 1. Primer sequences for real-time PCR analysis in *Xenopus tropicalis*.

Supplementary video legends

Supplementary Video 1. Characterization of cardiac expression of mCherry in Tg(mlc2:H2C) tadpole. Beating heart of Tg(mlc2:H2C) tadpole was captured by Leica M205FA stereo fluorescence microscope in bright- and epifluorescence-field vison. Red fluorescence indicates mCherry expression in heart. Scale bar, 250 µm.

Supplementary Video 2. Characterization of nuclear expression of mCherry in cardiomyocytes of Tg(mlc2:H2C) tadpole. Beating heart of Tg(mlc2:H2C) tadpole was captured by the SpinSR10 spinning disk confocal super resolution microscope (Olympus). Red fluorescence indicates nuclear expression of mCherry in cardiomyocytes. Scale bar, 100 µm.