

Fig. S1. Computational screen of AHs in nuclear envelope proteins.

(A) Detailed flow chart of data mining for putative NE proteins and identification of predicted AHs. Numbers in

parenthesis indicate the number of proteins. (B) Breakdown of the 410 NE proteins by the number of the NE proteome

papers. NE proteins found in only a single NE proteome paper were further categorized by the specific NE proteome

paper. (C) Breakdown of NE proteins with or without a predicted AH by the number of NE proteome papers.



Fig. S2. Overlap of AHs in this study and ALPS-like motifs in Drin et al. 2007.

Venn diagram representing the NE proteins, those with or without a MemBrain-predicted AH, and proteins with a putative

ALPS-like motifs (Drin et al. 2007).



z = 5

z = 5

No membrane-bound organelle

Fig. S3. Subcellular localization patterns of AH candidates tagged with mNG that did not localize to membrane-

bound organelles.

Representative spinning disk confocal images of a single section in living U2OS cells expressing the indicated AH-mNG protein are shown. Schematic representations of corresponding helical wheel projections of the predicted AHs with mean hydrophobic moment $\langle \mu H \rangle$ and net charge z is shown. AH-mNG proteins with a nuclear body-like pattern are highlighted at the bottom right. Scale bars, 10 µm.



Fig. S4. AH-mNG without localization to membrane-bound organelles or nuclear bodies is expressed without

unexpected cleavage of the AH.

Immunoblot of mNG alone or the indicated AH-mNG proteins transiently expressed in U2OS cells. Among the AH-mNG proteins without membrane-bound organelle localization (Fig. S2), we tested AH-mNG proteins that did not localize to nuclear bodies, because the nuclear body localization indicated the presence of AH in the expressed protein.















Fig. S5. Subcellular localization patterns of AH candidates tagged with mNG that displayed non-mitochondrial

patterns.

Representative spinning disk confocal images of a single section in living U2OS cells expressing the indicated AH-mNG protein are shown. Schematic representations of corresponding helical wheel projections of the predicted AHs with mean hydrophobic moment $\langle \mu H \rangle$ and net charge z is shown. Scale bars, 10 μ m.



AH-APMAP

































<µH> = 0.455 z = 0

\H-	TME	M1	60	
(ja)	Mar I	E.	-	_
	-/1			1
	Tak			. A
	AL.	4		
	my By			

AH1-TMEM109

AH-TMEM245





AH-GGCX







AH1-WFS1





<µH> = 0.357 z = 3

z = 0

AH-TMEM192 <µH> = 0.513

AH-TMEM68





z = 5

<uH> = 0.489 z = 0



Mitochondria





<µH> = 0.608 z = 1

<µH> = 0.598 z = 0



AH-NDUFB5

<µH> = 0.374

z = 2 AH-SGPP1



z = 3

Fig. S6. Subcellular localization patterns of AH candidates tagged with mNG that displayed mitochondrial patterns.

Representative spinning disk confocal images of a single section in living U2OS cells expressing the indicated AH-mNG protein are shown. Schematic representations of corresponding helical wheel projections of the predicted AHs with mean hydrophobic moment $\langle \mu H \rangle$ and net charge z is shown. Scale bars, 10 µm.



Fig. S7. Assessment of the organelle type to which that AH-mNG constructs localized.

Representative spinning disk confocal images of a single section in living U2OS cells co-expressing the indicated AH-mNG

and the indicated organelle marker are shown. Scale bars, $10 \,\mu m$.



Fig. S8. Sequence codes in hydrophobic face that discriminate AHs with non-mitochondrial and mitochondria-like

patterns.

(A) Heat map representation of the fraction of each amino acid (columns) present within the hydrophobic face of each

amphipathic helix (row) categorized by indicated localization patterns. (B-D) Scatter plots of the indicated metrices of AH

candidates categorized and color-coded by their localization patterns as determined from experiments in Fig. 1B.





```
<µH> = 0.399
z = 2
```

<µH> = 0.456

z = 0

AH(ALPS1-ALPS2)-mNG-NLS



Fig. S9. Subcellular localization patterns and INM association of AHs from human Nup153, Nup133 and ArfGAP1.

Representative spinning disk confocal images of a single section in living U2OS cells expressing the indicated AH-mNG (A) and AH-mNG-NLS protein with magnified zoom of the nuclear rim area (A and B) are shown. Schematic representations of corresponding helical wheel projections of the AHs with mean hydrophobic moment $\langle \mu H \rangle$ and net charge z is shown. Scale bars, 10 µm or 2 µm (zoom).



Fig. S10. Purification of AH-GFP.

Coomassie staining of the indicated AH-GFP recombinant proteins expressed and purified from bacterial cells.



В



Fig. S11. Putative AH in TMEM214 protein.

(A) The putative AH is highlighted in the 3D structure of TMEM214 protein, predicted by AlphaFold2 and shown by PyMol. A putative transmembrane (TM) domain predicted by TMHMM is shown in brown. The N and C termini are predicted to face cytoplasm/nucleoplasm and NE/ER lumen, respectively, according to InterPro (ID: Q6NUQ4). (B) PyMol presentation

of the TMEM214 AH. Amino acid residues are color-coded according to the wheel projection generated by HeliQuest.



Zoom

Fig. S12. Hypotonic shock promotes AH binding to the inner nuclear membrane.

 $Single \ z\text{-section spinning disk confocal images of living U2OS \ cells \ expressing \ with \ the \ indicated \ AH-mNG-NLS \ constructs$

before and after the hypotonic shock. An arrowhead indicates the deformation of the nuclear rim wrinkle that is no longer

present after hypotonic shock. Scale bars, 10 μ m or 2 μ m (zoom).



Fig. S13. Flow chart of the NE enrichment score quantification and schematic of ROIs in AH-GUV binding

quantification.

(A) Flow chart of the NE enrichment score quantification. (B) Schematic illustration of ROIs in AH-GUV binding

quantification. See methods for more detail.