Supporting Information

Synthesis of Benzopyran-Phenylpropanoid Hybrids via Matsuda-Heck-Arylation and Allylic Oxidation

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A Single crystal X-ray structure analyses of compounds 16cf

1 General details of X-ray structure analysis

The crystal structures were determined by single crystal structure analysis. Suitable single crystals were selected using a Leica M205C light microscope and separated with oil. **Figure S1** shows a single crystal of **16cf**. The X-ray crystal structure analysis was performed on a Stadivari diffractometer (Stoe) with monochromated Mo-*K* α radiation ($\lambda = 0.71073$ Å). The data correction was performed using the program X-Area.¹ The structure was solved by direct methods and refined against *F*² on all data by full-matrix least-squares using the SHELX suite of programs.^{2,3} All non-hydrogen atoms were refined anisotropically; the hydrogen atoms were placed on calculated positions. **Table S1** was created using FinalCif.⁴ The crystal structure was visualized with Diamond.⁵ The data (**16cf**: CCCD 2156920) can be obtained free of charge from The Cambridge Crystallographic Data Centre, http://www.ccdc.cam.ac.uk.



Figure S1: Microscope image of a single crystal of 16cf.

2 Crystallographic Data:

Compound	16cf
CCDC number	2156920
Empirical formula	$C_{28}H_{28}O_{6}$
Formula weight	460.50
Temperature [K]	210
Crystal system	triclinic
Space group	$P\overline{1}(2)$
a [Å]	10.0329(5)
<i>b</i> [Å]	12.4721(6)
c [Å]	20.1848(9)
α [°]	107.193(4)
β[°]	99.409(4)
γ [°]	91.739(4)
Volume [Å ³]	2372.1(2)
Ζ	4
$\rho_{\rm calc} [\rm g cm^{-3}]$	1.289
$\mu \text{ [mm^{-1}]}$	0.090
F(000)	976
Crystal size [mm ³]	0.400×0.245×0.095
Crystal color	colorless
Crystal shape	needle
Radiation	Mo K_{α} (λ=0.71073 Å)
2θ range [°]	4.30 to 59.98
	(0.71 Å)
Index ranges	$-14 \le h \le 12$
	$-17 \le k \le 17$
	$-28 \le l \le 28$
Reflections	59370
collected	
Independent	13808
reflections	$R_{\rm int} = 0.0392$
	$R_{\rm sigma} = 0.0374$
Completeness to	99.8 %
$\theta = 25^{\circ}$	
Data / Restraints /	13808/92/778
Parameters	
Goodness-of-fit on	1.067
F^2	
Final <i>R</i> indexes	$R_1 = 0.0556$
$[I \ge 2\sigma(I)]$	$wR_2 = 0.1528$
Final <i>R</i> indexes	$R_1 = 0.1043$
[all data]	$wR_2 = 0.1735$
Largest peak/hole [eÅ ⁻³]	0.43/-0.35

Table S1. Crystal data and details of structure refinement for 16cf.

3 Visualization of the crystal structure and molecular structure for compound 16cf



Figure S2: Molecular structure with atom labeling of the symmetry-independent molecule A in 16cf. Displacement ellipsoids are shown at the 50% probability level.



Figure S3: Disorder in the asymmetric molecule A in 16cf with the main part shown in orange and the minor part shown in green color (hydrogens are omitted).



Figure S4: Molecular structure with atom labeling of the symmetry-independent molecule B in 16cf. Displacement ellipsoids are shown at the 50% probability level.



Figure S5: Disorder in the asymmetric molecule B in 16cf with the main part shown in orange and the minor part shown in green color (hydrogens are omitted).



Figure S6: C-H…O hydrogen bonding (red dashed lines) in 16cf.



Figure S7: Stacking interactions (purple dotted lines) between the molecules in **16cf**. (X1, X2 and X3 mark the centers of the aromatics in molecule A, X4, X5 and X6 mark the centers in molecule B, hydrogens are omitted).



Figure S8: Cell view of 16cf with the asymmetric molecule A colored in orange and the asymmetric molecule B colored in green looking along the crystallographic b axis (hydrogens are omitted).



Figure S9: Cell view of 16cf with the asymmetric molecule A colored in orange and the asymmetric molecule B colored in green looking along the crystallographic c axis (hydrogens are omitted).



Figure S10: Cell view of 16cf with the asymmetric molecule A colored in orange and the asymmetric molecule B colored in green looking along the crystallographic a axis (hydrogens are omitted).

B Syntheses of compounds 11d and 15b

1 Synthesis of 5-(prop-2-en-1-yl)-1,2,3,4-tetrahydronaphthalene (11d)



Scheme S1 Synthesis of test substrate 11d

5,6,7,8-Tetrahydronaphthalen-1-ylacetic acid (SI-2).⁶ The title compound was synthesized according to a modified literature procedure:⁶ to a solution of SI-1 (1.86 g, 10.0 mmol) in ethyl acetate (15 mL) was added PtO₂ (180 mg, 0.8 mmol, 8 mol-%) and the solution was purged with hydrogen. The mixture was then stirred under an atmosphere of hydrogen (2 bar) for 3

days. The mixture was filtered through a short pad of celite, washed with ethyl acetate (10 mL), and the combined ethyl acetate solution was evaporated. The residue was recrystallized from hexane to give **SI-2** as colourless crystals; yield: 1.31 g (6.9 mmol, 69%); mp 128 – 130 °C; IR (ATR) ν 2941 (w), 1705 (s), 1406 (m), 1257 (m), 1210 (s), 780 (m) cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.12-7.02 (m, 3H), 3.65 (s, 2H), 2.81 (t, J = 6.0 Hz, 2H), 2.70 (t, J = 6.0 Hz, 2H), 1.89-1.74 (m, 4H); ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 178.4, 138.0, 136.0, 132.1, 129.0, 128.0, 125.6, 38.7, 30.2, 26.5, 23.4, 22.8; HRMS (EI) m/z [M+H]⁺ calcd for C₁₂H₁₅O₂ 191.1067, found 191.1072.

2-(5,6,7,8-Tetrahydronaphthalen-1-yl)ethanol (SI-3). Conditions adapted from a literature procedure:⁷ under an atmosphere of dry nitrogen, LiAlH₄ (0.19 g, 5.3 mmol) was suspended in dry THF (40 mL) and cooled to 0 °C. A solution of SI-2 (1.00 g, 5.3 mmol) in dry THF (40 mL) was added dropwise, the mixture was warmed to ambient temperature and then heated at 65 °C for 24 h. It was then cooled to ambient temperature, poured onto ice-water mixture (35 mL), and the resulting precipitate was dissolved with aq. H₂SO₄ (1 M). MTBE (25 mL) was added to the mixture, the organic layer was separated, and the aqueous layer was extracted with MTBE (3 times 10 mL). The combined organic extracts were dried with MgSO₄, filtered and evaporated. The product was sufficiently pure to be used in the next step without further purification; colourless oil; yield: 0.84 g (4.8 mmol, 90%); IR (ATR) v 3327 (s, br.), 2927 (s), 2837 (m), 1587 (m), 1458 (m), 1436 (m), 1036 (s), 771 (s), 719 (s) cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.11 (t, J = 7.5 Hz, 1H), 7.04 (d, J = 7.5 Hz, 1H), 7.01 (d, J = 7.5 Hz), 3.85 (t, J = 7.0 Hz, 2H), 2.89 (t, J = 7.0 Hz, 2H), 2.83 (t, J = 6.2 Hz, 2H), 2.78 (t, J = 6.2 Hz, 2H), 2.37 (s(br.), 1H), 1.92-1.77 (m, 4H); ${}^{13}C{}^{1}H$ NMR (100 MHz, CDCl₃) δ 137.7, 136.4, 135.6, 127.9, 127.0, 125.4, 62.6, 35.9, 30.2, 26.3, 23.5, 22.9; HRMS (EI) m/z [M⁺] calcd for C₁₂H₁₆O 176.1196, found 176.1193.

5,6,7,8-Tetrahydronaphthalen-1-ylacetaldehyde (SI-4). Conditions adapted from a literature procedure:⁸ to a solution of **SI-3** (400 mg, 2.3 mmol) in CH₂Cl₂ (19 mL) was added Dess-Martin-periodinane (DMP, 1060 mg, 2.5 mmol) in portions at ambient temperature. After stirring for two hours, another portion of DMP (530 mg, 1.3 mmol) was added and stirring was continued for 12 h. The mixture was diluted with diethylether (20 mL), and aq. NaOH (1 M, 20 mL) was added. The organic layer was separated, washed with aq. NaOH (1 M, 10 mL) followed by water (20 mL), and then dried with MgSO₄. It was filtered and evaporated to furnish crude **SI-4** (400 mg, 2.3 mmol, quant.). All attempts to purify aldehyde **SI-4** resulted in decomposition; therefore, **SI-4** was used in the following reaction without further purification; ¹H NMR (400 MHz, CDCl₃) δ 9.72 (t, *J* = 2.3 Hz, 1H), 7.15 (t, *J* = 7.5 Hz, 1H), 7.09 (d, *J* = 7.5 Hz, 1H), 7.09 (d,

Hz, 1H), 7.04 (d, J = 7.5 Hz, 1H), 3.69 (d, J = 2.3 Hz, 2H), 2.84 (t, J = 6.2 Hz, 2H), 2.64 (t, J = 6.2 Hz, 2H), 1.94-1.72 (m, 4H); ¹³C{¹H} NMR (100 MHz, CDCl₃): $\delta = 199.7$, 138.2, 136.1, 130.7, 129.0, 128.1, 125.8, 48.4, 30.1, 26.8, 23.3, 22.7.

5-(Prop-2-en-1-yl)-1,2,3,4-tetrahydronaphthalene (11d). Conditions adapted from a literature procedure:⁹ a solution of methyltriphenylphosphonium bromide (986 mg, 2.8 mmol) in THF (7 mL) was cooled to 0 °C. A solution of BuLi in hexane (2.5 M, 1.01 mL, 2.5 mmol) was slowly added, and the mixture was warmed to ambient temperature and stirred for 0.5 h. The mixture was cooled to 0 °C, a solution of SI-4 (400 mg, 2.3 mmol) in THF (2.5 mL) was slowly added, and it was warmed to ambient temperature and stirred at ambient temperature for 2 h. The reaction was quenched by addition of methanol (10 mL), and all volatiles were removed in vacuo. The residue was extracted with pentane (three times, 10 mL each), filtered, and evaporated. The residue was purified by column chromatography on silica, using hexanes as eluent; colourless liquid; yield: 350 mg (2.0 mmol, 88%); IR (ATR): 2926 (s), 1637 (m), 1586 (w), 1458 (s), 1436 (s), 993 (s), 909 (s), 772 (s), 728 (s) cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.10 (t, J = 7.5 Hz, 1H), 7.05-6.98 (m, 2H), 5.99 (ddt, J = 16.9, 10.1 6.5 Hz, 1H), 5.10 (dq, J = 10.1, 1.6 Hz, 1H), 5.05 (dq, J = 16.9, 1.7 Hz, 1H), 3.77 (dm, J = 6.5 Hz, 2H), 2.83 (t, J = 6.3 Hz, 1H), 2.73 (t, J = 6.3 Hz, 1H), 1.91-1.76 (m, 4H); ${}^{13}C{}^{1}H$ NMR (100 MHz, CDCl₃) δ 138.3, 137.5, 136.9, 135.4, 127.6, 126.6, 125.4, 115.7, 37.4, 30.3, 26.2, 23.5, 23.0. No matching HRMS data could be obtained using either EI or ESI.













 Figure S15: ¹H NMR (400 MHz, CDCl₃) of SI-4 (crude reaction mixture, due to decomposition upon purification attempts)

 NEO400_2024-0617_mak.10.fid

 718roh







2 Synthesis of 7-methoxy-2-[4-(methoxymethoxy)phenyl]-8-(prop-2-en-1-yl)-2,3dihydro-4*H*-chromen-4-one (15b)



Scheme S2 Synthesis of allylflavanone 15b

(2*E*)-3-[4-(Methoxymethoxy)phenyl]-1-[4-methoxy-2-(prop-2-en-1-vloxy)phenyl]prop-2en-1-one (SI-7). A solution of KOH in methanol (60 wt %, 40 mL) was added dropwise to a well stirred solution of acetophenone SI-5 (2.06 g, 10.0 mmol) and 4-OMOM-substituted benzaldehyde SI-6 (1.66 g, 10.0 mmol) in methanol (20 mL) at 20 °C. The reaction mixture was stirred at 20 °C for 48 h. It was then poured into ice water (100 mL) and neutralized with aqueous HCl (4 M). The aqueous solution was extracted with EtOAc (three times 60 mL). The combined organic extracts were washed with brine, dried with anhydrous MgSO₄, and filtered. The solvent was evaporated under reduced pressure, and the residue was purified by column chromatography on silica using hexane – EtOAc mixture (5:1 (v/v)) as eluent to afford SI-7 as a pale yellow solid; yield: 2.30 g (6.5 mmol, 65%); mp 81 – 83 °C; IR (ATR) v 2903 (w), 1649 (m), 1584 (s), 1244 (m), 1149 (s) cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.77 (d, J = 8.6 Hz, 1H). 7.66 (d, J = 15.7 Hz, 1H), 7.53 (d, J = 8.6 Hz, 2H), 7.50 (d, J = 15.7 Hz, 1H), 7.04 (d, J = 8.6 Hz, 2H), 6.57 (dd, J = 8.6, 2.3 Hz, 1H), 6.48 (d, J = 2.3 Hz, 1H), 6.06 (ddt, J = 17.2, 10.6, 5.1 Hz, 1H), 5.45 (dm, J = 17.2 Hz, 1H), 5.28 (dm, J = 10.6 Hz, 1H), 5.21 (s, 2H), 4.62 (dt, J = 5.1, 1.6 Hz, 2H), 3.86 (s, 3H), 3.49 (s, 3H); ${}^{13}C{}^{1}H$ NMR (100 MHz, CDCl₃) δ 190.7, 164.1, 159.4, 158.9, 141.7, 133.1, 132.6, 130.0, 129.4, 125.7, 122.8, 118.1, 116.5, 105.7, 99.9, 94.4, 69.5, 56.3, 55.7; HRMS (EI) m/z [M⁺] calcd for C₂₁H₂₂O₅ 354.1467, found 354.1472.

7-Methoxy-2-[4-(methoxymethoxy)phenyl]-8-(prop-2-en-1-yl)-2,3-dihydro-4*H***-chromen-4-one (15b)**. A solution of the **SI-7** (710 mg, 2.0 mmol) in toluene (10 mL) was placed in a S19 vessel suited for microwave irradiation. The vessel was sealed and irradiated in a microwave reactor at 250 °C for 1.5 h. The solvent was evaporated, and the residue was redissolved in methanol (20 mL) in a vessel suited for microwave irradiation. NaOAc (1.68 g, 20.00 mmol) was added to the solution, the vessel was sealed again, and the mixture was further irradiated in a microwave reactor at 100 °C for 2 h. The solvent was then evaporated, and water (50 mL) was added to the residue. The mixture was extracted with EtOAc (three times 30 mL). The combined organic extracts were washed with brine, dried with anhydrous MgSO₄, and filtered. The solvent was evaporated under reduced pressure and the residue was purified by column chromatography on silica using hexanes – MTBE mixture (4:1 (v/v)) to afford the **15b** as a pale yellow solid; yield: 320 mg (0.90 mmol, 45%); mp 87 - 89 °C; IR (ATR) v 2901 (w), 2884 (w), 1667 (s), 1601 (s), 1234 (m), 1022 (s) cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.85 (d, J = 8.8 Hz, 1H), 7.39 (d, J = 8.7 Hz, 2H), 7.08 (d, J = 8.7 Hz, 2H), 6.63 (d, J = 8.8 Hz, 1H), 5.91 (ddt, J = 17.1, 10.0, 6.3 Hz, 1H), 5.39 (dd, J = 12.8, 3.0 Hz, 1H), 5.18 (s, 2H), 4.98 (dm, J = 17.1 Hz, 1H), 4.94 (dm, J = 10.0 Hz, 1H), 3.87 (s, 3H), 3.47 (s, 3H), 3.41 (d, J = 6.3, Hz, 2H), 2.97 (dd, J = 16.8, 12.8 Hz, 1H), 2.82 (dd, J = 16.8, 3.0 Hz, 1H); ${}^{13}C{}^{1}H$ NMR (100 MHz, CDCl₃) δ 191.5, 163.3, 160.3, 157.3, 135.8, 132.6, 127.3, 126.7, 116.3, 115.9, 115.3, 114.7, 104.9, 94.3, 79.0, 56.0, 55.9, 44.2, 27.1; HRMS (EI) m/z [M⁺] calcd for C₂₁H₂₂O₅ 354.1467, found 354.1481.





Figure S20: ¹³C{¹H} NMR (100 MHz, CDCl₃) of **SI-7**





C Copies of NMR-spectra

compound	page	compound	page	compound	page
12aa	S26	12ar	S75	13aq	S135
12ab	S28	12at	S77	14ar	S141
12ac	S30	12au	S83	13at	S148
12ad	S 32	13aa	S85	14au	S155
12ae	S34	13ab	S91	16af	S162
12af	S 36	13ac	S93	16bf	S167
12ag	S43	13ad	S95	16cf	S171
12ah	S45	13ae	S97	17af	S175
12ai	S51	14ae	S103	17bf	S180
12ak	S57	13af	S109	17cf	S184
12al	S59	13ai	S116	18af	S188
12am	S61	13ak	S118	18bf	S190
12an	S63	13al	S120	18cf	S192
12ao	S65	14am	S126	20	S194
12ap	S67	13an	S131	21	S199
12aq	S 73	13ap	S133	23	S204































Figure S33: NMR-Signal assignment and selected HMBC-couplings for compound 12af


Figure S35: ¹³C{¹H} NMR (100 MHz, CDCl₃) of 12af





Figure S36: COSY (400 MHz, CDCl₃) of 12af



Figure S37: HSQC (400/100 MHz, CDCl₃) of 12af



Figure S38: HMBC (400/100 MHz, CDCl₃) of 12af



Figure S39: NOESY (400 MHz, CDCl₃) of 12af













Figure S45: HSQC (400/100 MHz, CDCl₃) of 12ah





Figure S46: HMBC (400/100 MHz, CDCl₃) of 12ah

Figure S47: NOESY (400 MHz, CDCl₃) of 12ah















Figure S52: HMBC (400/100 MHz, CDCl₃) of 12ai



Figure S53: NOESY (400 MHz, CDCl₃) of 12ai



























Figure S66: COSY (400 MHz, CDCl₃) of 12ap



Figure S67: HSQC (400/100 MHz, CDCl₃) of 12ap





Figure S69: NOESY (400 MHz, CDCl₃) of 12ap










Figure S75: ¹³C{¹H} NMR (100 MHz, CDCl₃) of 12at





Figure S76: COSY (400 MHz, CDCl₃) of 12at



Figure S77: HSQC (400/100 MHz, CDCl₃) of 12at





Figure S79: NOESY (400 MHz, CDCl₃) of 12at









Figure S84: COSY (400 MHz, CDCl₃) of 13aa





Figure S85: HSQC (400/100 MHz, CDCl₃) of 13aa



Figure S86: HMBC (400/100 MHz, CDCl₃) of 13aa























Figure S96: COSY (400 MHz, CDCl₃) of 13ae



Figure S97: HSQC (400/100 MHz, CDCl₃) of 13ae



Figure S98: HMBC (400/100 MHz, CDCl₃) of 13ae













Figure S102: COSY (400 MHz, CDCl₃) of 14ae



Figure S103: HSQC (400/100 MHz, CDCl₃) of 14ae



Figure S104: HMBC (400/100 MHz, CDCl₃) of 14ae



Figure S105: NOESY (400 MHz, CDCl₃) of 14ae


Figure S106: NMR-Signal assignment and selected HMBC-couplings for compound 13af





Figure S109: COSY (400 MHz, CDCl₃) of 13af





Figure S110: HSQC (400/100 MHz, CDCl₃) of 13af



Figure S111: HMBC (400/100 MHz, CDCl₃) of 13af



Figure S112: NOESY (400 MHz, CDCl₃) of 13af

























Figure S122: HSQC (400/100 MHz, CDCl₃) of 13am

Figure S123: HMBC (400/100 MHz, CDCl₃) of 13am









Figure S125: NMR-Signal assignment and selected HMBC-couplings for compound 14an







Figure S128: HSQC (400/100 MHz, CDCl₃) of 14an



Figure S129: HMBC (400/100 MHz, CDCl₃) of 14an







Figure S133: ${}^{13}C{}^{1}H$ NMR (100 MHz, CDCl₃) of 13aq_NE0400_2023-1117_mak.71.fid



Figure S134: COSY (400 MHz, CDCl₃) of 13aq





Figure S135: HSQC (400/100 MHz, CDCl₃) of 13aq











Figure S138: NMR-Signal assignment and selected HMBC-couplings for compound 14ar










Figure S142: HSQC (400/100 MHz, CDCl₃) of 14ar









Figure S145: NMR-Signal assignment and selected HMBC-couplings for compound 13at









Figure S148: COSY (400 MHz, CDCl₃) of 13at





Figure S150: HMBC (400/100 MHz, CDCl₃) of 13at



8.6 8.4 8.2 8.0 7.8 7.6 7.4 7.2 7.0 6.8 6.6 6.4 6.2 6.0 5.8 5.6 5.4 5.2 5.0 4.8 4.6 4.4 4.2 4.0 3.8 3.6 3.4 f2 (ppm)

Figure S151: NOESY (400 MHz, CDCl₃) of 13at

3.5

-4.0

-4.5

-5.0

-5.5

-6.0

-6.5

-7.0

-7.5

-8.0

-8.5

f1 (ppm)

١

1

.

4

1



Figure S152: NMR-Signal assignment and selected HMBC-couplings and NOE-interaction for compound 14au







Figure S155: COSY (400 MHz, CDCl₃) of 14au









Figure S157: HMBC (400/100 MHz, CDCl₃) of 14au



Figure S158: NOESY (400 MHz, CDCl₃) of 14au



Figure S159: NMR-Signal assignment and selected HMBC-couplings for compound 16af





Figure S161: $^{13}C\{^1H\}$ NMR (100 MHz, CDCl₃) of 16af











Figure S165: ¹³C{¹H} NMR (100 MHz, CDCl₃) of **16bf**







Figure S168: ¹H NMR (400 MHz, CDCl₃) of 16cf



Figure S169: $^{13}C\{^1H\}$ NMR (100 MHz, CDCl₃) of 16cf



Figure S170: HSQC (400/100 MHz, CDCl₃) of 16cf



Figure S171: HMBC (400/100 MHz, CDCl₃) of 16cf



Figure S172: NMR-Signal assignment and selected HMBC-couplings for compound 17af












Figure S178: ¹³C{¹H} NMR (100 MHz, CDCl₃) of 17bf











Figure S181: ¹H NMR (400 MHz, CDCl₃) of 17cf



Figure S182: ¹³C{¹H} NMR (100 MHz, CDCl₃) of 17cf

Figure S183: HSQC (400/100 MHz, CDCl₃) of 17cf













Figure S187: ¹H NMR (400 MHz, CDCl₃) of **18bf**









Figure S191: NMR-Signal assignment and selected HMBC-couplings for compound 20









Figure S195: HMBC (400/100 MHz, CDCl₃) of 20



Figure S196: NMR-Signal assignment and selected HMBC-couplings for compound 21































D References

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