### Supplementary figure legends

Figure S1. IL-2 signaling enhances the effects of TCR/CD28 co-stimulation on AMBRA1 expression. A. Immunoblot analysis of AMBRA1 protein levels in CD8<sup>+</sup> T cells collected at days 0, 2, 5 and 7 after stimulation with anti-CD3/anti-CD28 coated beads and cultured in the presence or absence of 50 U/ml IL-2. Actin was used as loading control. The migration of molecular mass markers is indicated. The histograms show the quantification of AMBRA1 expression during CTL differentiation ( $n_{donor}$ =3, one-way ANOVA test, day 0 value=1). B. Flow cytometry analysis of IL-2 receptor (CD25) expression in CD8<sup>+</sup> T cells collected at days 0, 2, 5 and 7 after stimulation in the presence or absence of 50 U/ml IL-2. The histograms show the percentage (%) of CD25<sup>+</sup> cells ( $n_{donor}$ =5, one-way ANOVA test). Data are shown as mean fold ± SD. \* P≤ 0.05; \*\* P ≤ 0.01; \*\*\* P ≤ 0.001; \*\*\*\* P ≤ 0.0001.

#### Figure S2. Neither CTL expansion nor IL-2R expression and signaling are affected by

**AMBRA1 depletion. A.** Flow cytometry analysis of live cell number (propidium iodide<sup>-</sup> cells) in control (ctr, scramble RNAi) and AMBRA1 KD CTLs 48h after transfection. The histograms show the quantification of live cells in ctr and AMBRA1 KD CTLs ( $n_{donor}$ =3, one-sample t test, ctr value=1). **B.** Flow cytometry analysis of surface CD25 expression in control and AMBRA1 KD CTLs. The histograms show the percentage (%) of CD25<sup>+</sup> cells ( $n_{donor}$ =5, Student's t test). **C.** Immunoblot analysis of STAT3 phosphorylation (Tyr705) in response to IL-2 stimulation for 15 minutes. Actin was used as loading control. The migration of molecular mass markers is indicated. The histograms show the quantification of STAT3 phosphorylation in control and AMBRA1 KD CTLs ( $n_{donor}$ =3, one-way ANOVA test, day 0 value=1). Data are shown as mean fold ± SD. \* P≤ 0.05; \*\* P ≤ 0.01; \*\*\* P ≤ 0.001; \*\*\*\* P ≤ 0.001.

Figure S3. Control of SBI-0206965 and LB-100 activity and effect on cell viability in CTLs. A. Immunoblot analysis of the autophagy marker LC3B in lysates of CTLs untreated or treated with SBI-0206965 or LB-100 for 48 h, as described in Fig.3A. Actin was used as loading control. The migration of molecular mass markers is indicated. The histogram shows the quantification of LC3II in treated vs untreated CTLs. Data are shown as mean fold ± SD (n=3, one sample t test, vehicle value in CTLs set as 1). B. Immunoblot analysis of AKT (S473) and ERK1/2 phosphorylation in lysates of CTLs untreated or treated with SBI-0206965 or LB-100 for 48 h, as described in Fig.3A. Actin was used as loading control. The migration of molecular mass markers is indicated. The histogram shows the quantification of phospho-proteins in treated vs untreated CTLs. Data are shown as mean fold ± SD (n=3, one sample t test, vehicle value in CTLs set as 1). C. Quantification of cell viability of CTLs treated for 48h with LB-100 or SBI-0206965, assessed with trypan blue dye staining. Data are shown as mean fold ± SD (n>3, one-way ANOVA test). D. Immunoblot analysis of pFOXO3A S253 and FOXO3A in CTLs untreated or treated with LB-100 or SBI-0206965 for 48 h, as described in Fig.4A. Actin was used as loading control. The migration of molecular mass markers is indicated. The histograms show the quantification of pFOXO3A S253/FOXO3A and FOXO3A/actin in LB-100 and SBI-0206965 treated CTLs related to untreated CTLs. Data are shown as mean fold ± SD (n=3, one sample t test, vehicle value=1). \* P≤ 0.05; \*\* P ≤ 0.01; \*\*\* P ≤ 0.001; \*\*\*\* P ≤ 0.0001.

**Figure S4. AMBRA1 depletion, ULK1 or PP2A inhibition enhance the mTORC1 pathway. A.** Immunoblot analysis of P-p70S6K T389 in CTLs untreated or treated with LB-100 or SBI-0206965 for 48 h, as described in Fig.4A. Actin was used as loading control. The migration of molecular mass markers is indicated. The histogram shows the quantification of P-p70S6K in treated vs untreated CTLs. Data are shown as mean fold ± SD (n=3, one sample t test, vehicle value=1). **B.** Immunoblot analysis of P-p70S6K T389 in lysates from control and AMBRA1 KD CTLs. Actin was used as loading control. The migration of molecular mass markers is indicated. The histogram shows the quantification of P-p70S6K in control and AMBRA1 KD CTLs. Data are shown as mean fold  $\pm$  SD (n=3, one sample t test, vehicle value=1). \* P≤ 0.05; \*\* P ≤ 0.01; \*\*\* P ≤ 0.001; \*\*\*\* P ≤ 0.0001.



Supplementary figure 1







### Supplementary Tables

Antibody anti-	Supplier	Identifier	Host	Diluition	Application
AMBRA1	Merck	ABC131	Rabbit	1:1000	WB
β-actin	Sigma-Aldrich	MAB1501	Mouse	1:10000	WB
Granzyme B	Cell Signaling	17215S	Rabbit	1:2000	WB
Perforin 1	Santa Cruz	sc-136994	Mouse	1:500	WB
p-FoxO3a (Ser253)	Cell Signaling	9466	Rabbit	1:1000	WB
FoxO3a	Cell Signaling	2497	Rabbit	1:1000	WB
LC3B	Cell Signaling	3868	Rabbit	1:500	WB
GFP	Invitrogen	A11120	Mouse	1:100	IF
β-tubulin	Cell Signaling	15115	Rabbit	1:2000	WB
TFEB	Cell Signaling	37785	Rabbit	1:1000	WB
P-p70S6K (T389)	Cell Signaling	9234S	Rabbit	1:500	WB
P-AKT (S473)	Cell Signaling	9271S	Rabbit	1:500	WB
P-ERK1/2	Cell Signaling	9101	Rabbit	1:1000	WB
P-STAT3 (Tyr705)	Cell Signaling	9145S	Rabbit	1:1000	WB
CD25 (PE)	BioLegend	302606	Mouse	1:40	FC

# Supplementary Table 1. List of the primary antibodies used in this work

Gene	Forward 5'-3'	Reverse 5'-3'		
18S	CGCCGCTAGAGGTGAAATT	CTTGGCAAATGCTTTCGC		
AMBRA1	AACCCTCCACTGCGAGTTGA	TCTACCTGTTCCGTGGTTCTCC		
TFEB	GGAGTACCTGTCCGAGACCT	GGGCTATTGGGAGCACTGTT		
TBET	TAATAACCCCTTTGCCAAAG	TCCCCCAAGGAATTGACAGT		
RUNX3	CCCCTCCGTTCCTAACTGTT	ACAGAGAGTGGATGCGTTGA		
GZMB	TCAAAGAACAGGAGCCGACC	TTGGCCTTTCTCTCCAGCTG		
GZMA	AACCAGGAACCATGTGCCAA	GGCTTCCAGAATCTCCATTGC		
PRF1	CCTGCAGTCACAGCTACACA	GGGGCTCCAGTTAAGGCAAT		
SGLN	GACGAGAATCCAGGACTTGAA	GGGCAGATTCCTGTCAAGAG		
GNLY	GGATAAGCCCACCCAGAGAAG	ACAGATCTGCTGGGCAGTTT		
EOMES	TTGATTCCTTAACCCCCGGC	TCTCTCCTGAGTCCCACTGG		
CTSD (CHIP)	GCAGCTTCCAGGTCATAGGG	CTGACCTCAGGTGATCTGCC		
TBET (CHIP)	ATCGTCCTATCTCCGAGGCA	GAGAGGGTGTACGTGTGCAG		
RUNX3	CTGGTGAGTCAGAGGCTGAA	GGGAAAGGTGGCGGCTTTTA		
(CHIP)				

## Supplementary Table 2. List of the primers used in this study