

Reporting Summary

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our [Editorial Policies](#) and the [Editorial Policy Checklist](#).

Statistics

For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.

n/a Confirmed

- The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
- A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
- The statistical test(s) used AND whether they are one- or two-sided
Only common tests should be described solely by name; describe more complex techniques in the Methods section.
- A description of all covariates tested
- A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
- A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
- For null hypothesis testing, the test statistic (e.g. F , t , r) with confidence intervals, effect sizes, degrees of freedom and P value noted
Give P values as exact values whenever suitable.
- For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
- For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
- Estimates of effect sizes (e.g. Cohen's d , Pearson's r), indicating how they were calculated

Our web collection on [statistics for biologists](#) contains articles on many of the points above.

Software and code

Policy information about [availability of computer code](#)

Data collection

Data analysis

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio [guidelines for submitting code & software](#) for further information.

Data

Policy information about [availability of data](#)

All manuscripts must include a [data availability statement](#). This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our [policy](#)

The raw data of single-cell RNA transcriptome data has been deposited in the GEO database with the accession code GSE254132. Proteome data is available from MassIVE with accession code MSV000093736. Requests for custom code can be directed to the corresponding authors, and the code will be provided within 30 days and without restrictions.

Research involving human participants, their data, or biological material

Policy information about studies with [human participants or human data](#). See also policy information about [sex, gender \(identity/presentation\), and sexual orientation](#) and [race, ethnicity and racism](#).

Reporting on sex and gender	Our study included both male and female subjects. We ensured balanced representation of sexes in our experimental design and accounted for potential sex-based differences in our analysis.
Reporting on race, ethnicity, or other socially relevant groupings	Our study included only Asian subjects. As a result, the findings may be most applicable to Asian populations, and caution should be exercised when generalizing to other racial or ethnic groups.
Population characteristics	Our study population consisted of Asian subjects, including both male and female participants. The age range of the subjects was 46-57 year-old. Patients were included if they were diagnosed with complicated type B AD, which was confirmed through computed tomography angiography (CTA) and had signed informed consent for TEVAR. Detailed demographic and clinical characteristics are provided in Supplementary Tables.
Recruitment	We prospectively collected and analyzed the clinical data of patients with type B aortic dissection who underwent endovascular therapy in the Zhongshan Hospital of Fudan University between November 2016 and November 2020. Patients were included if they were diagnosed with complicated type B AD, which was confirmed through computed tomography angiography (CTA) and had signed informed consent for TEVAR. Patient demographics and baseline information were recorded by medical record. The blood samples were collected before TEVAR.
Ethics oversight	This study was approved by the Ethics Committee of Zhongshan Hospital of Fudan University (IRB number B2019-231R). Healthy normal aortas and dissected aortas were collected from participants with informed written consent, and under approval of local medical ethnics from Zhongshan Hospital Fudan University.

Note that full information on the approval of the study protocol must also be provided in the manuscript.

Field-specific reporting

Please select the one below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.

Life sciences Behavioural & social sciences Ecological, evolutionary & environmental sciences

For a reference copy of the document with all sections, see nature.com/documents/nr-reporting-summary-flat.pdf

Life sciences study design

All studies must disclose on these points even when the disclosure is negative.

Sample size	The sample size of $n = 12$ per group was determined based on a power analysis conducted using GPower. The analysis was designed to detect a disease incidence with a power of 0.8 and a two-sided significance level of 0.05.
Data exclusions	No data were excluded from analyses.
Replication	All experiments were conducted using standardized protocols to ensure consistency across all groups. Detailed protocols were followed for animal handling, treatment administration, data collection, and analysis.
Randomization	Animals were randomly assigned to experimental groups using a computer-generated random number sequence to ensure unbiased allocation. The randomization was stratified by weight to balance this variable across groups.
Blinding	To reduce bias, experiments were conducted in a blinded manner. Researchers responsible for data collection and analysis were unaware of the group allocations during the study.

Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Materials & experimental systems

- n/a Involved in the study
- Antibodies
- Eukaryotic cell lines
- Palaeontology and archaeology
- Animals and other organisms
- Clinical data
- Dual use research of concern
- Plants

Methods

- n/a Involved in the study
- ChIP-seq
- Flow cytometry
- MRI-based neuroimaging

Antibodies

Antibodies used

Human aortic tissue samples were then permeabilized and stained with rabbit MPO antibody (ab208670, Abcam, Ltd., Cambridge, UK) at 1 mg/mL, polyclonal rabbit CitH3 antibody (ab5103, Abcam, Ltd., Cambridge, UK) at 1 mg/mL, rabbit CXCL3 antibody (AV07037, Sigma-Aldrich Co., St. Louis, MO, USA) at 1 mg/mL, and with rabbit CXCR2 antibody (ab225732, Abcam, Ltd., Cambridge, UK) at 1 mg/mL. Mouse aortic tissue samples were then permeabilized and stained with rabbit Ly6G antibody (ab238132, Abcam, Ltd., Cambridge, UK) at 2 mg/mL, rabbit CitH3 antibody (ab5103, Abcam, Ltd., Cambridge, UK) at 1 mg/mL, rabbit CXCL3 antibody (ab220431, Abcam, Ltd., Cambridge, UK) at 1 mg/mL, and with rabbit CXCR2 antibody (bs-12257R, Bioss Inc. Woburn, Massachusetts, USA) at 1 mg/mL.

As for in vivo study, the challenged mice were treated with either isotype control antibody (n = 12) or rabbit CXCL3 antibody (2 mg/kg, AF5568, R&D systems, Minneapolis, MN, USA) (n = 12) or CXCR2 antibody (2 mg/kg, MAB2164-100, R&D systems, Minneapolis, MN, USA) (n = 12)

Validation

The primary antibody used in this study was validated for its specificity in mouse through western blot analysis, which confirmed specific binding and no cross-reactivity with other proteins. The antibody was validated for use in immunohistochemistry by testing on tissue sections known to express the target protein and by including appropriate positive and negative controls.

Eukaryotic cell lines

Policy information about [cell lines and Sex and Gender in Research](#)

Cell line source(s)

This study did not utilize any cell lines. All experiments were conducted using animal models and patient samples.

Authentication

This study did not utilize any cell lines.

Mycoplasma contamination

This study did not utilize any cell lines.

Commonly misidentified lines
(See [ICLAC](#) register)

This study did not utilize any cell lines.

Palaeontology and Archaeology

Specimen provenance

This study did not involve the use of biological specimens that require provenance documentation.

Specimen deposition

This study did not involve the use of biological specimens that require provenance documentation.

Dating methods

This study did not involve the use of biological specimens that require provenance documentation.

Tick this box to confirm that the raw and calibrated dates are available in the paper or in Supplementary Information.

Ethics oversight

This study did not involve the use of biological specimens that require provenance documentation.

Note that full information on the approval of the study protocol must also be provided in the manuscript.

Animals and other research organisms

Policy information about [studies involving animals](#); [ARRIVE guidelines](#) recommended for reporting animal research, and [Sex and Gender in Research](#)

Laboratory animals

Wild-type C57BL/6J male 3-week-old mice were used in this study and were purchased from the Beijing Vital River Laboratory Animal Technology.

Wild animals

The study did not involve wild animals.

Reporting on sex

We focused on male mice since sexual dimorphism has been reported in aortic dissection (AD) formation in humans and mice. In humans, AD has been more prevalent in men than in women. Likewise, in several mouse models, the incidence and severity of aortic

pathologies are sexually dimorphic, with greater severity in male mice. While no studies have reported sexual dimorphism when β -Aminopropionitrile (BAPN) is administered alone, a few studies have indicated a lower incidence of AD in female mice co-administered BAPN and Angiotensin-II. Taking these factors into consideration, similar to other studies published in top-tier cardiovascular journals, we chose to use male mice for our experiments.

Field-collected samples

The study did not involve samples collected from field.

Ethics oversight

All animal experiments were approved by the Institutional Animal Care and Use Committee at Zhongshan Hospital.

Note that full information on the approval of the study protocol must also be provided in the manuscript.

Clinical data

Policy information about [clinical studies](#)

All manuscripts should comply with the ICMJE [guidelines for publication of clinical research](#) and a completed [CONSORT checklist](#) must be included with all submissions.

Clinical trial registration

This study did not involve a clinical trial. The research was conducted using observational studies.

Study protocol

This study did not involve a clinical trial. The research was conducted using observational studies.

Data collection

This study did not involve a clinical trial. The research was conducted using observational studies.

Outcomes

This study did not involve a clinical trial. The research was conducted using observational studies.

Dual use research of concern

Policy information about [dual use research of concern](#)

Hazards

Could the accidental, deliberate or reckless misuse of agents or technologies generated in the work, or the application of information presented in the manuscript, pose a threat to:

- | No | Yes |
|-------------------------------------|---|
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Public health |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> National security |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Crops and/or livestock |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Ecosystems |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Any other significant area |

Experiments of concern

Does the work involve any of these experiments of concern:

- | No | Yes |
|-------------------------------------|--|
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Demonstrate how to render a vaccine ineffective |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Confer resistance to therapeutically useful antibiotics or antiviral agents |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Enhance the virulence of a pathogen or render a nonpathogen virulent |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Increase transmissibility of a pathogen |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Alter the host range of a pathogen |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Enable evasion of diagnostic/detection modalities |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Enable the weaponization of a biological agent or toxin |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Any other potentially harmful combination of experiments and agents |

Plants

Seed stocks

This study did not involve the use of seed stocks.

Novel plant genotypes

This study did not involve the use of seed stocks.

Authentication

This study did not involve the use of seed stocks.

ChIP-seq

Data deposition

Confirm that both raw and final processed data have been deposited in a public database such as [GEO](#).

Confirm that you have deposited or provided access to graph files (e.g. BED files) for the called peaks.

Data access links

May remain private before publication.

This study did not involve the use of CHIP-seq.

Files in database submission

This study did not involve the use of CHIP-seq.

Genome browser session

(e.g. [UCSC](#))

This study did not involve the use of CHIP-seq.

Methodology

Replicates

This study did not involve the use of CHIP-seq.

Sequencing depth

This study did not involve the use of CHIP-seq.

Antibodies

This study did not involve the use of CHIP-seq.

Peak calling parameters

This study did not involve the use of CHIP-seq.

Data quality

This study did not involve the use of CHIP-seq.

Software

This study did not involve the use of CHIP-seq.

Flow Cytometry

Plots

Confirm that:

The axis labels state the marker and fluorochrome used (e.g. CD4-FITC).

The axis scales are clearly visible. Include numbers along axes only for bottom left plot of group (a 'group' is an analysis of identical markers).

All plots are contour plots with outliers or pseudocolor plots.

A numerical value for number of cells or percentage (with statistics) is provided.

Methodology

Sample preparation

This study did not involve the use of flow cytometry.

Instrument

This study did not involve the use of flow cytometry.

Software

This study did not involve the use of flow cytometry.

Cell population abundance

This study did not involve the use of flow cytometry.

Gating strategy

This study did not involve the use of flow cytometry.

Tick this box to confirm that a figure exemplifying the gating strategy is provided in the Supplementary Information.

Magnetic resonance imaging

Experimental design

- Design type
- Design specifications
- Behavioral performance measures

Acquisition

- Imaging type(s)
- Field strength
- Sequence & imaging parameters
- Area of acquisition
- Diffusion MRI Used Not used

Preprocessing

- Preprocessing software
- Normalization
- Normalization template
- Noise and artifact removal
- Volume censoring

Statistical modeling & inference

- Model type and settings
- Effect(s) tested
- Specify type of analysis: Whole brain ROI-based Both
- Statistic type for inference
- (See [Eklund et al. 2016](#))
- Correction

Models & analysis

- n/a Involved in the study
 - Functional and/or effective connectivity
 - Graph analysis
 - Multivariate modeling or predictive analysis
- Functional and/or effective connectivity
- Graph analysis
- Multivariate modeling and predictive analysis