

# **Synaptic Signatures and Disease Vulnerabilities of Layer 5 Pyramidal Neurons**

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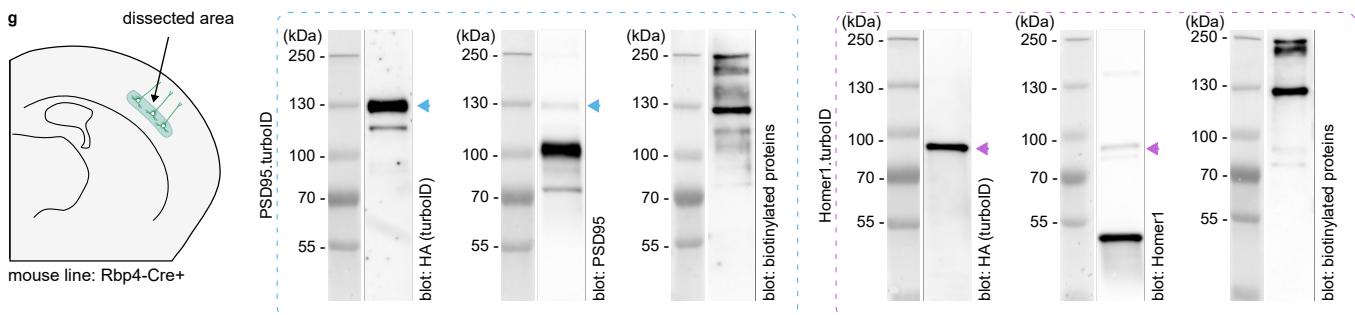
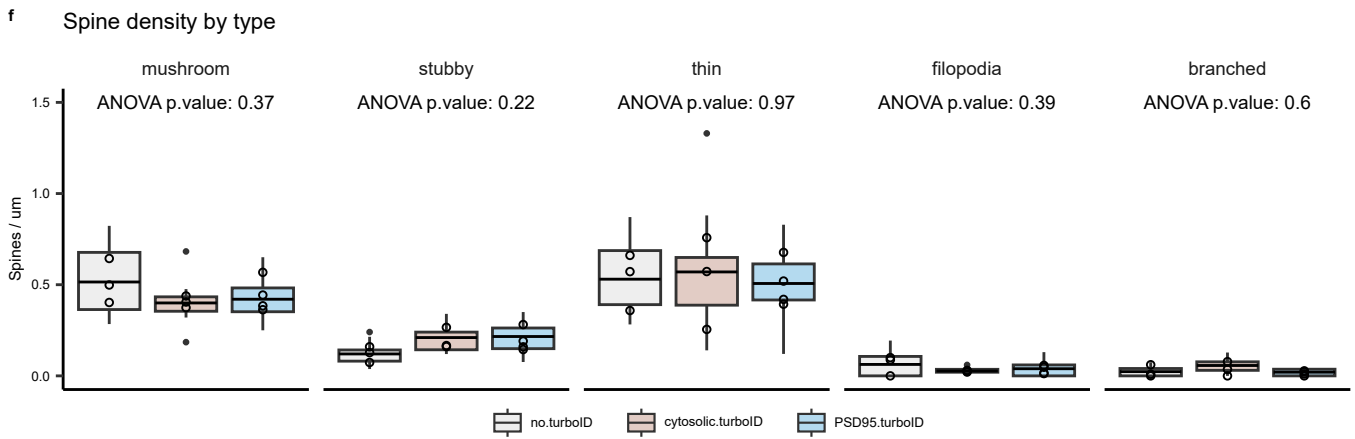
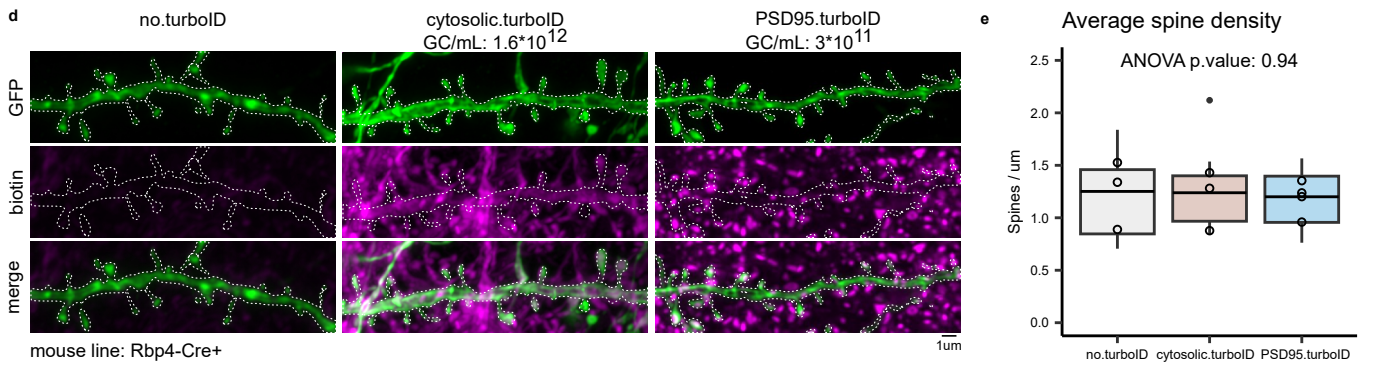
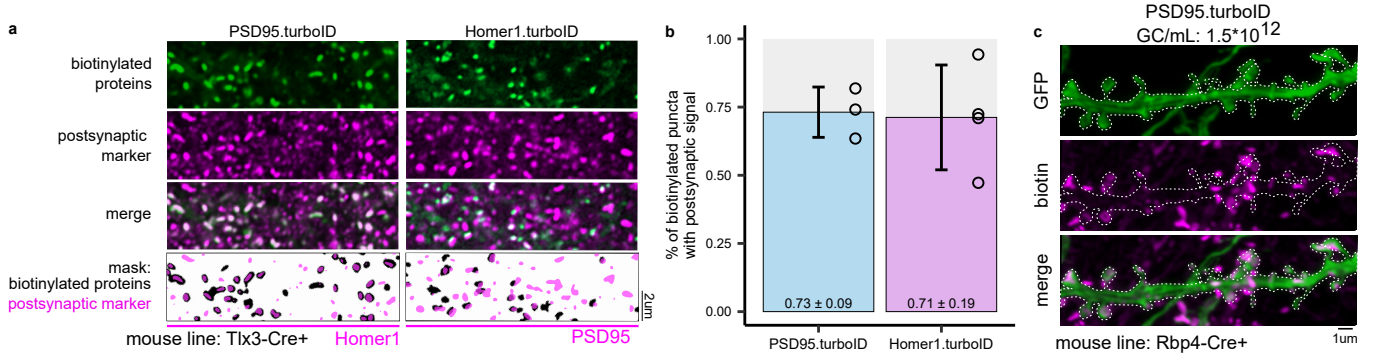
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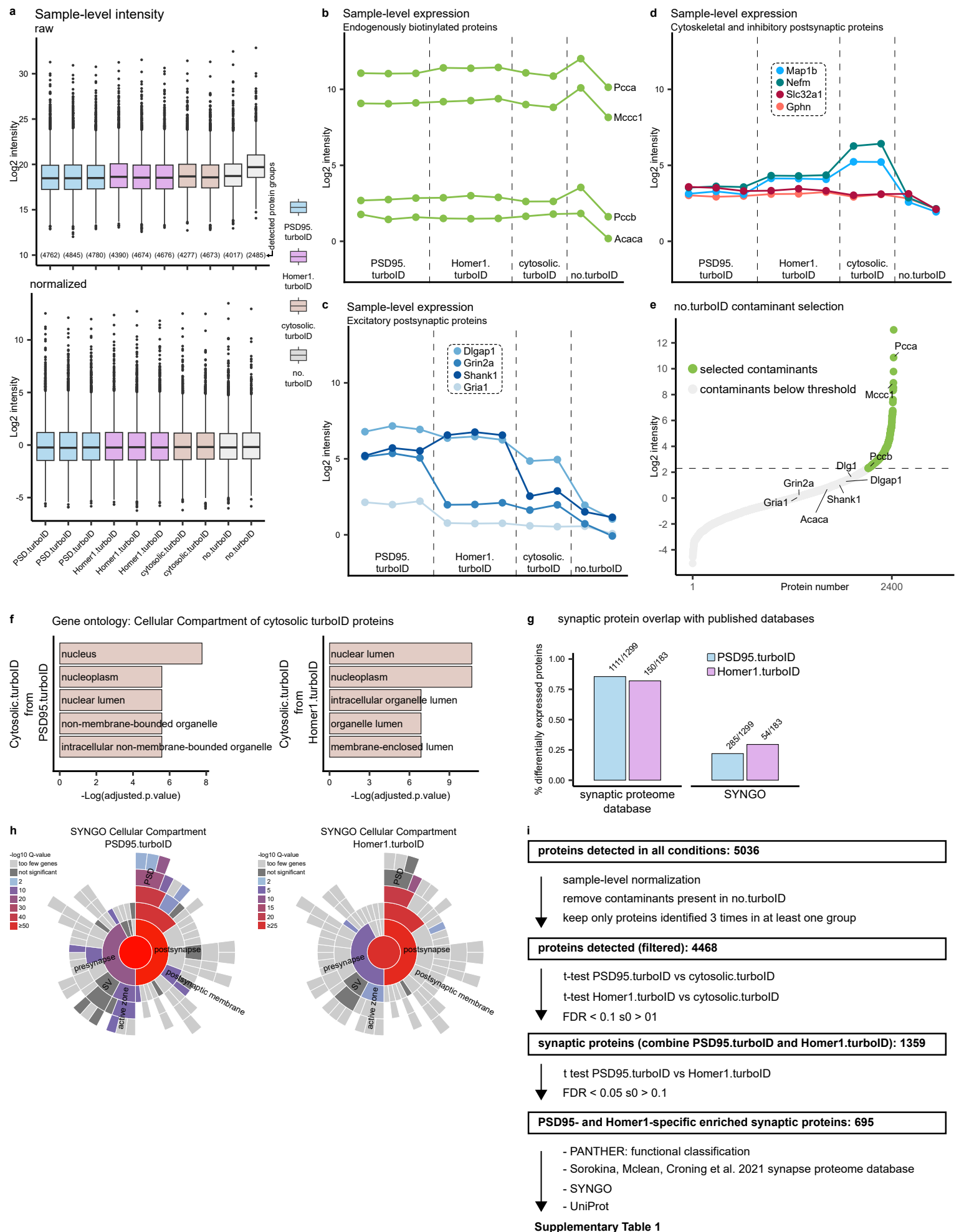
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**Supplementary Figure 1. TurboID expression has no effect on dendritic spine density or morphology.**

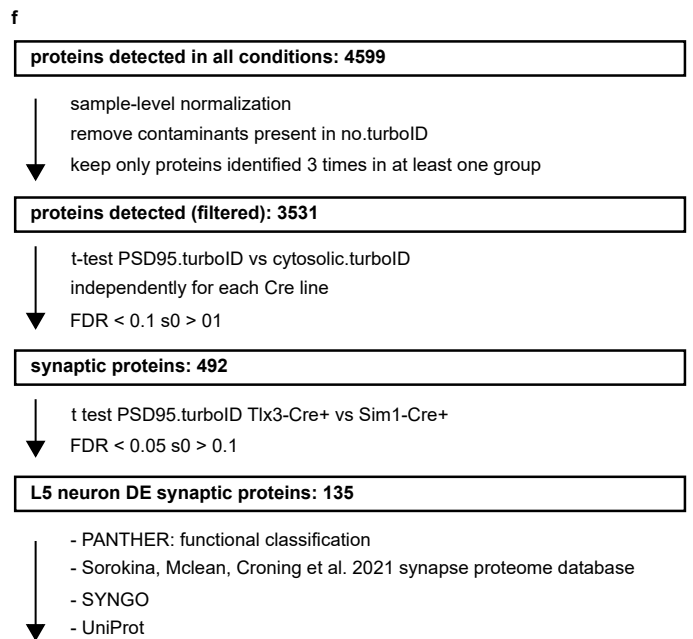
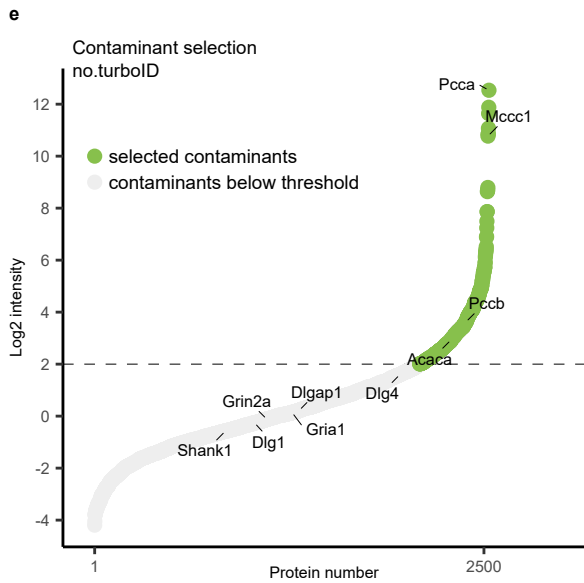
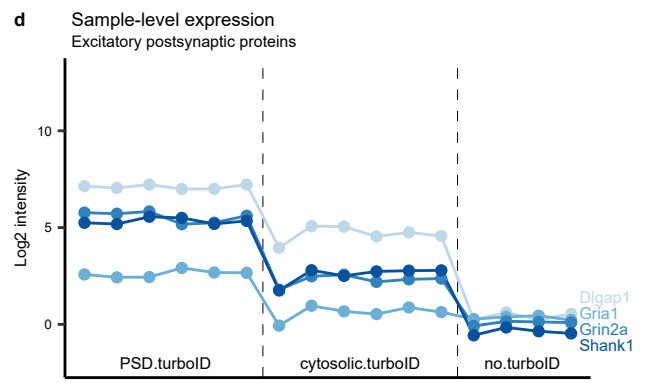
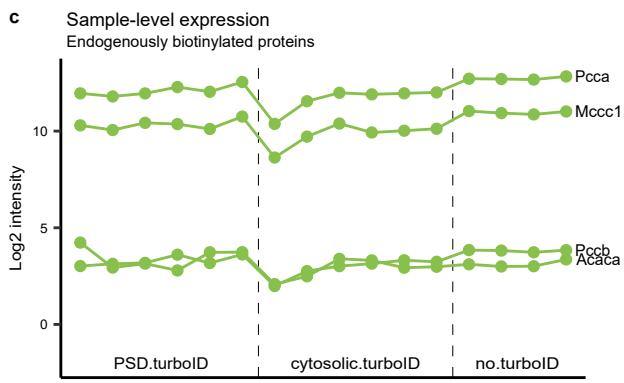
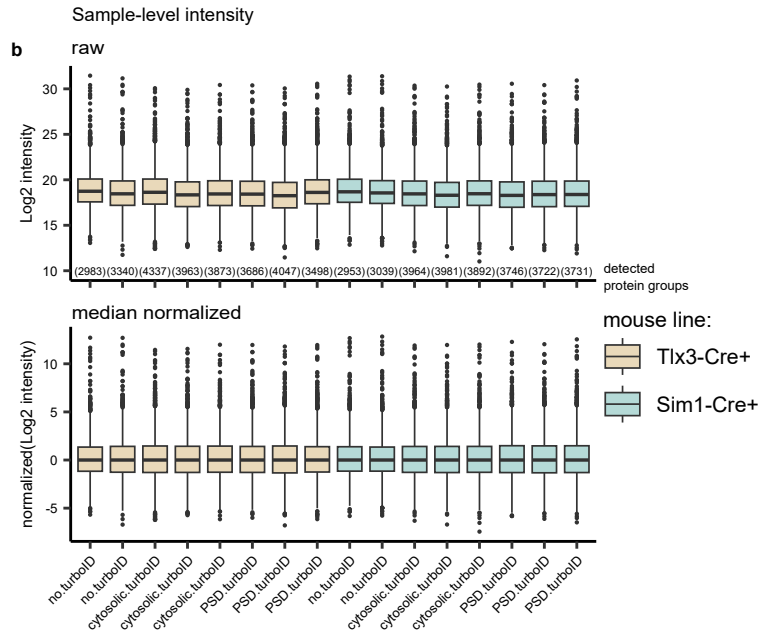
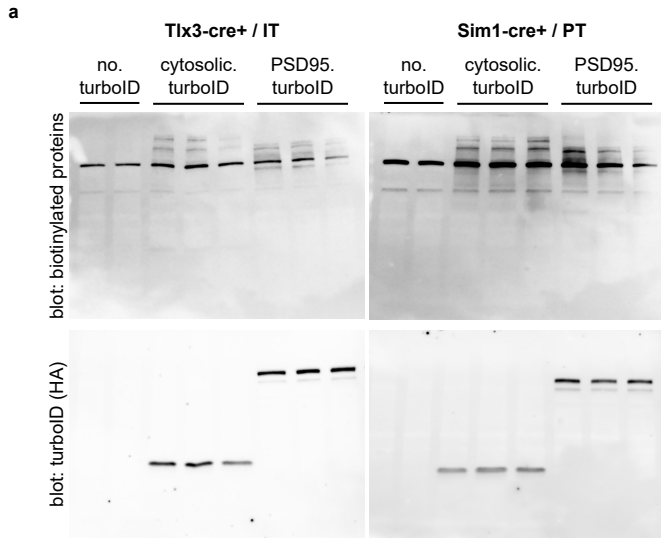
**a**, High resolution images of biotinylated proteins produced by synaptic TurboID constructs, corresponding postsynaptic markers and threshold masks used for quantification in (b). Scale bar is 2  $\mu\text{m}$ . **b**, Fraction of biotinylated objects that contain postsynaptic marker signal for synaptic TurboID constructs. Error bars show standard deviation. Mean and standard deviation are indicated on the graph. N = 3 mice per condition (PSD95.turboID) or 4 mice per condition (Homer1.turboID), 1 single-plane field of view per mouse is quantified as a replicate. Source data are provided as a Source Data file. **c**, Representative image of Supernova-labeled L5 neuron dendrite of Rbp4-Cre mice injected with high titer AAV-PSD95.turboID. Viral titer is indicated as Genome Copies (GC) per mL. Scalebar is 1  $\mu\text{m}$ . **d**, Representative images of Supernova-labeled dendrites of control L5 neurons and L5 neurons expressing different TurboID constructs. Viral titer is indicated as Genome Copies (GC) per mL. Scalebar is 1  $\mu\text{m}$ . **e, f**, Quantification of spine density (e) and quantification of spine density by spine type (f). N = 3 mice per replicate except PSD95.turboID = 4 mice per replicate, average of 2 dendrites per mouse. Boxplots show spine density distribution of individual dendrites, empty dots display the average spine density per replicate. The line across the box describes the mean. The lower and upper hinges correspond to the first and third quartiles. The upper whisker extends from the hinge to the largest value no further than  $1.5 \times$  inter-quartile range. The lower whisker extends from the hinge to the smallest value at most  $1.5 \times$  inter-quartile range of the hinge. Data beyond the end of the whiskers (outliers) are plotted individually as a small, solid dot. ANOVA statistical analysis is performed on the replicate means. Source data are provided as a Source Data file. **g**, AAV expressing Cre-dependent GFP was co-injected with AAV-PSD95.TurboID or AAV-Homer1.TurboID in the somatosensory cortex of Rbp4-Cre mice. Tissue was dissected as close as possible to the cell bodies to minimize inclusion of non-transduced material. Western blot analysis of the TurboID tag (HA) shows the molecular weight of the PSD95.turboID (blue arrow) and Homer1.turboID (purple arrow) constructs. Molecular weight markers in kDa are indicated on the left. Western blot analysis with an antibody against endogenous PSD95 and Homer1 proteins shows low expression levels of TurboID-fusion proteins compared to endogenous proteins levels. Biotinylated protein analysis confirms successful transduction and TurboID activity. These experiments were independently repeated more than 2 times with similar results.





**Supplementary Figure 2. Quality control and analysis for PSD95.turboID and Homer1.turboID synaptic proteome workflow.**

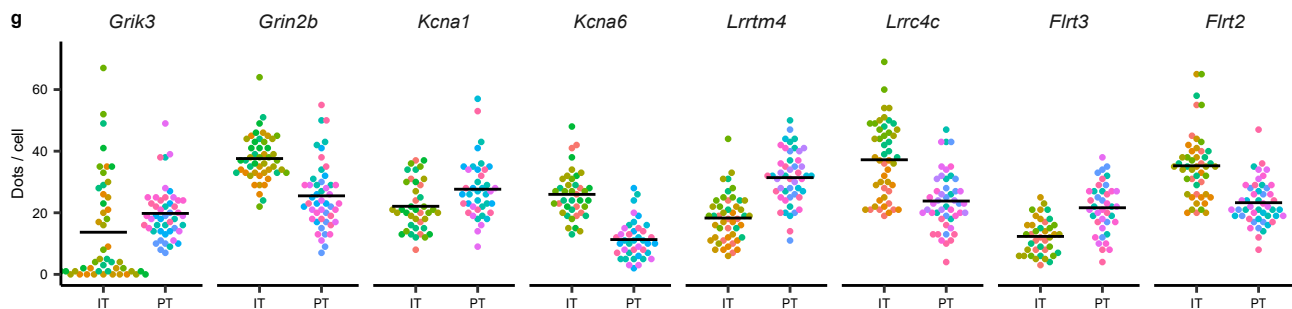
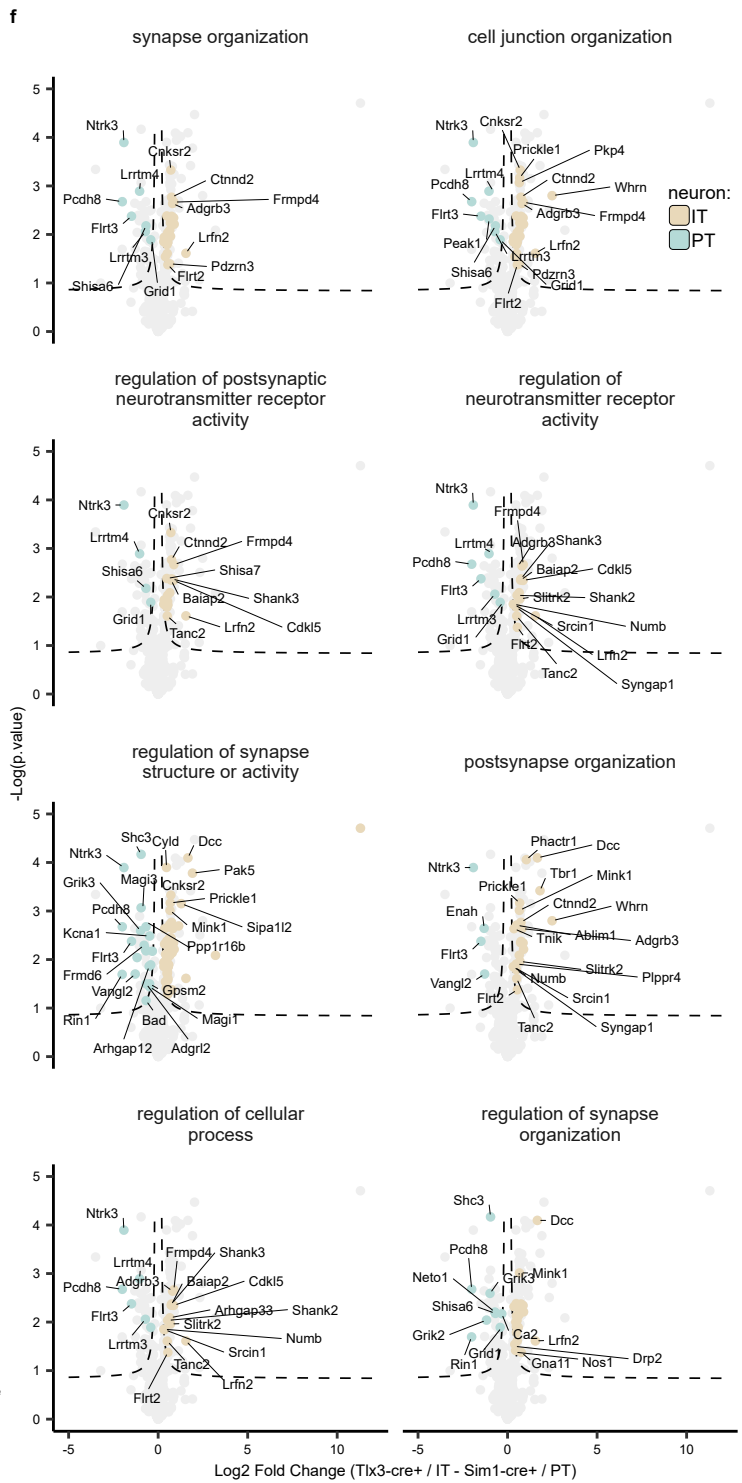
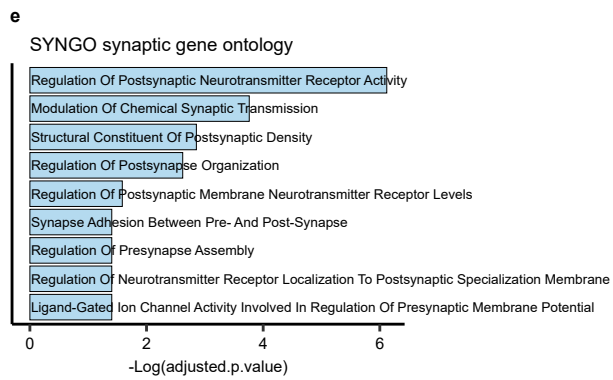
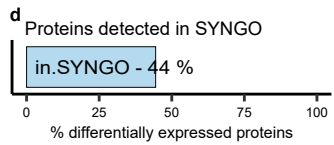
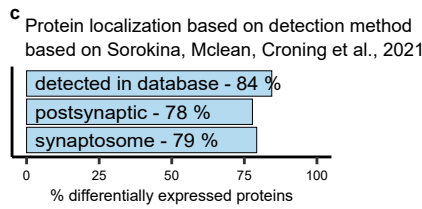
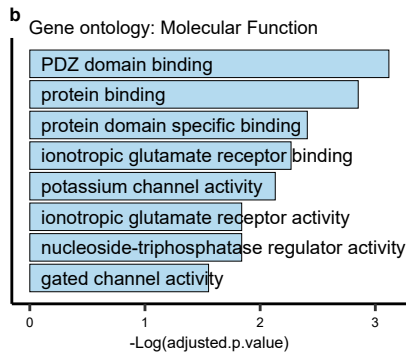
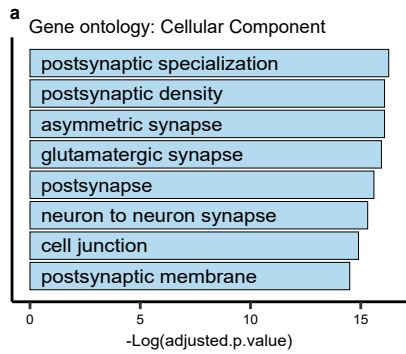
**a**, Sample-level intensity values for each replicate before and after median normalization. Protein groups detected in each replicate are reported above the x-axis on the top graph. **b**, Intensity levels in each sample for 4 endogenously biotinylated proteins. **c**, Intensity levels in each sample for 4 known PSD proteins. **d**, Intensity levels in each sample for 2 known inhibitory postsynapse proteins and 2 cytoskeletal proteins. **e**, Proteins detected in all 4 no.turboID replicates are ranked by intensity. Endogenously biotinylated proteins and known PSD proteins are labelled. Threshold for selecting contaminants is based on the expression of endogenously biotinylated proteins and known PSD components. **f**, Gene ontology analysis of cytosolic proteins in PSD95.turboID and Homer1.turboID samples (refers to Figure 2f). **g**, Overlap of the enriched proteins with the synaptic proteome database and SYNGO. **h**, SYNGO cellular compartment enrichment analysis plots of synaptic proteins in PSD95.turboID and Homer1.turboID samples showing distribution between pre- and postsynapse. **i**, Proteomic analysis workflow to enrich for synaptic proteins. See Supplementary Data 1 for complete annotation.



**Supplementary Table 2**

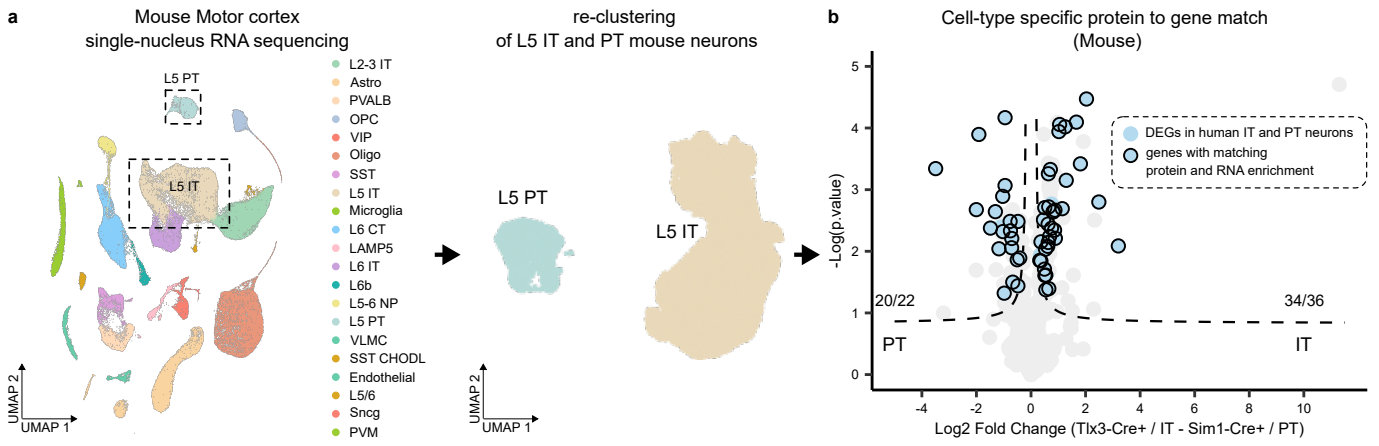
**Supplementary Figure 3. Quality control and analysis for cell type-specific TurboID synaptic proteome workflow.**

**a**, Western blots for biotinylated proteins and TurboID (HA) on replicates confirm successful biotinylation in every sample and correct TurboID construct expression. **b**, Sample-level intensity values for each replicate before and after median normalization. Protein groups detected in each replicate are reported on the top graph. **c**, Intensity levels in each sample for 4 endogenously biotinylated proteins. **d**, Intensity levels in each sample for 4 known PSD proteins. **e**, Proteins detected in all 4 no.turboID replicates are ranked by intensity. Endogenously biotinylated proteins and known PSD proteins are labelled. Threshold for selecting contaminants is based on the expression of endogenously biotinylated proteins and known PSD components. **f**, Proteomic analysis workflow to enrich for synaptic proteins and select candidates from differentially expressed proteins. See Supplementary Data 2 for complete annotation.

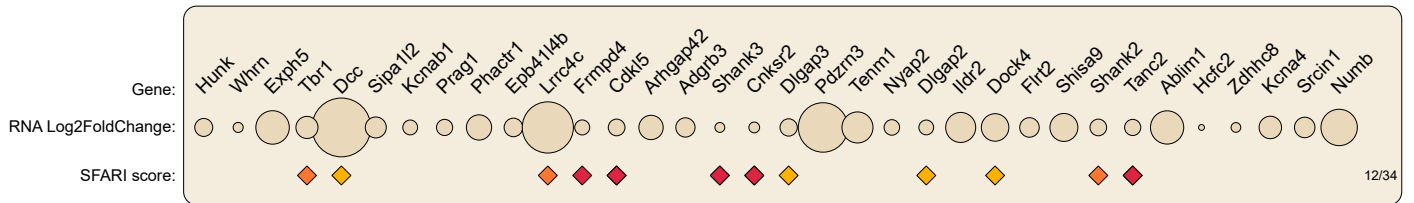


**Supplementary Figure 4. Gene ontology analysis of differentially expressed proteins in layer 5 neurons.**

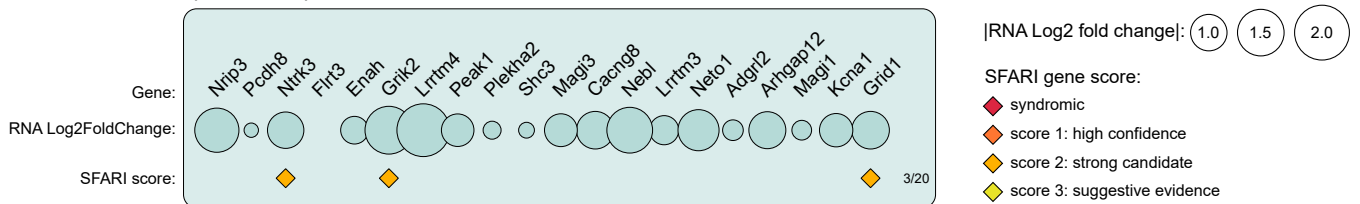
**a**, Gene ontology for Cellular Compartment and **b**, Molecular Function categories using differentially expressed proteins as input. **c**, Analysis of differentially expressed proteins using the synapse proteome database<sup>63</sup>. 85% of differentially expressed proteins were detected in the database; 79% and 82% were identified in postsynaptic fractionation studies and synaptosome fractionation studies, respectively. **d**, 44% of differentially expressed proteins were detected in the SYNGO database<sup>46</sup>. **e**, SYNGO gene ontologies. **f**, Annotation of proteins found in the Biological Process gene ontology terms (refers to Figure 4b). Not all protein names are displayed due to space limitations, see Supplementary Data 2 for complete annotation. **g**, Plots from Figure 4f and 4h showing distribution of the analyzed cells. Each color represents a different mouse, each dot represents a single cell.



**c** RNA expression of proteins enriched in mouse IT neurons



RNA expression of proteins enriched in mouse PT neurons



**Supplementary Figure 5. Comparison of differentially expressed synaptic proteins with mouse transcriptomic data.**

**a**, L5 IT and PT neurons were extracted from a published mouse motor cortex snRNA-Seq dataset<sup>61</sup> and re-clustered. **b**, Differentially expressed genes (DEGs) between mouse IT and PT neurons (adjusted p-value < 0.01 and log fold change > 0.25) were plotted on top of mouse synaptic proteins enriched in IT and PT neurons (two-sided, unpaired t-test with permutation-based FDR correction at 5%,  $s_0 > 0.1$ ) to evaluate their overlap. 20/22 DEGs matched PT neuron-enriched synaptic proteins, 34/36 DEGs matched IT neuron-enriched synaptic proteins. **c**, Synaptic proteins from mouse L5 IT and PT neurons that show matching differential gene expression in mouse motor cortex L5 IT and PT neurons are shown. SFARI scores are annotated.