Supporting Information for

Original article

Pretheranostic agents with extraordinary NIRF/photoacoustic imaging performance and photothermal oncotherapy efficacy

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1. Materials and apparatus

The raw materials used in organic synthesis are purchased from Macklin, Bidepharmatech and Aladdin, and all raw materials do not require further purification. amino acid and metal-ion reagents were purchased from Macklin. The pH of the solution was determined with the pH detector (ST 3100, OHAUS). Glass bottom cell culture dish ($\Phi 20$ mm, TC) was purchased from Nest. N-Ethylmaleimide was purchased from Shanghai Taoshu Biotechnology Co., Ltd. SuperKind TM Hypersensitive cell proliferation detection reagent (CCK-8) was purchased from Guangzhou Sishijia Biotechnology Co., Ltd. nude mice (6-week-old) were purchased from Guangzhou Southern Medical University Laboratory Animal Technology Development Co., Ltd. (Guangzhou, China), and used according to the regulation of the Local Institutional Animal Care and Use Committee (IACUC). ¹H NMR and ¹³C NMR spectra were obtained on a 400 MHz nuclear magnetic resonance instrument (AVANCE III 400, Bruker), and the mass spectra and liquid phase spectra were obtained on triple quadrupole liquid mass spectrometry (Prelude SPLC+TSQ Quantiva LC-MS/MS, Thermo Fisher Scientific). The absorption spectrum was measured with the UV-visible spectrophotometer (Hitachi U-3010, Japan), while the emission spectrum was obtained from the fluorescence spectrophotometer (FLS 980, Edinburgh), and the photoacoustic signal and intensity were measure from MSOT in Vision 128, iThera Medical Germany. The cytotoxicity was measured in a microplate reader (Multiskan FC, Thermo Fisher Scientific). Cell imaging was obtained on a 63X lens of the inverted confocal laser microscope (LSM 880 with AiryScan, Carl Zeiss). Animal imaging was obtained on the multi-mode small animal live imager (FX PRO, Bruker). Photothermal imaging were performed on 660 nm laser (RAL660T1-2.0W, Beijing Leiyuan Technology Co., Ltd) and photothermal images were recorded by an IR thermal camera (FLUKE-VT08/CN, Fluke Testing Instruments (Shanghai) Co., Ltd).

2. Synthesis of S-NBD

Synthesis of compound 2. Put 11.2 mL of DMF and 50 mL of CHCl₃ into a flask, slowly drip 12.4 mL of PBr₃ (130.5 mmol) at 0 °C, stir for 45 min, and then add 5 mL of Cyclohexanone (48.4 mmol). After stirring at room temperature for 8 hours, the reaction solution was poured into 200 mL of ice water, and NaHCO₃ solid was added. The pH was adjusted to around 7. The mixture was extracted with CH₂Cl₂, and the organic phase was dried with anhydrous sodium sulfate. A silica gel chromatographic column was used to obtain 5.6 g of a yellow oil, with a yield of 61%. ¹H NMR (400 MHz, Chloroform-*d*) δ 9.91 (s, 1H), 2.66 (t, *J* = 6.2 Hz, 2H), 2.17 (t, *J* = 6.1 Hz, 2H), 1.68 (dd, *J* = 11.4, 5.9 Hz, 2H), 1.60 (q, *J* = 5.7 Hz, 2H). ¹³C NMR (101 MHz, CDCl₃) δ 193.32, 143.46, 135.06, 38.66, 24.86, 24.13, 20.95. MS: m/z calculated for C₇H₉BrO₃ ([M-H]⁺):188.9; found:188.9.

Synthesis of compound 3. 204 mg of compound 2 (1.08 mmol) was dissolved in 6 mL of DMF, 180 mg of 4-bromo-2-hydroxybenzaldehyde (0.90 mmol) and 880 mg of Cs₂CO₃ (2.70 mmol) were weighed into the solution, stirred at room temperature for 16 h, the reaction solution was filtered, extracted and passed through a silica gel chromatographic column, and 132 mg of the compound was purified, with a yield of 42%.¹H NMR (400 MHz, Chloroform-*d*) δ 10.30 (s, 1H), 7.27 (s, 1H), 7.20 (d, *J* = 8.1 Hz, 1H), 7.01 (d, *J* = 8.1 Hz, 1H), 6.62 (s, 1H), 2.60 – 2.54 (m, 2H), 2.44 (t, *J* = 6.0 Hz, 2H), 1.72 (p, *J* = 6.1 Hz, 2H). ¹³C NMR (101 MHz, CDCl₃) δ 187.75, 159.48, 152.23, 130.20, 127.46, 126.90, 125.62, 122.89, 120.17, 118.69, 113.73, 30.07, 21.37, 20.11. MS: m/z calculated for C₁₄H₁₁BrO₂ ([M-H]⁺):290.9; found:290.9.

Synthesis of compound 4. Dissolve 100 mg of compound 3 (0.34 mmol) in 20 mL of 1,4-dioxane, weigh 4-methoxy-N-phenylaniline (100 mg, 0.5 mmol), $Pd_2(dba)_3$ (46 mg, 0.05 mmol), x-phos (74 mg) and Cs_2CO_3 (277 mg, 0.85 mmol) into the solution, heat and reflux at 100 °C for 16 h under nitrogen protection. After filtering the reaction solution, a silica gel chromatographic column was used to purify 82 mg of the compound, with a yield of 58%.¹H NMR (400 MHz, Chloroform-*d*) δ 10.17 (s, 1H),

7.30 (d, J = 7.4 Hz, 1H), 7.09 (q, J = 9.7, 8.8 Hz, 6H), 6.95 (d, J = 8.1 Hz, 1H), 6.88 (d, J = 8.4 Hz, 2H), 6.65 (t, J = 10.3 Hz, 3H), 3.82 (s, 3H), 2.58 – 2.53 (m, 2H), 2.42 (t, J = 5.6 Hz, 2H), 1.69 (p, J = 6.4 Hz, 2H). ¹³C NMR (101 MHz, CDCl₃) δ 187.46, 160.85, 156.97, 153.10, 150.14, 146.75, 139.40, 129.39, 127.93, 126.94, 126.88, 126.14, 124.70, 123.73, 116.07, 114.98, 114.62, 112.26, 106.44, 55.44, 29.94, 21.49, 20.47. MS: m/z calculated for C₂₇H₂₃NO₃ ([M-H]⁺):410.1;found:410.1.

Synthesis of compound 5. 500 mg of compound 4 (1.22 mmol) was dissolved in 20 mL of dichloromethane solution, PBr₃ 2.27 mL (24.4 mmol) was slowly added dropwise in an ice bath, stirred for 4 h, and finally 454 mg of compound was obtained by silica gel chromatography column, with a yield of 94%.¹H NMR (400 MHz, DMSO- d_6) δ 10.09 (s, 1H), 9.55 (s, 1H), 7.34 (t, J = 7.8 Hz, 2H), 7.17 (d, J = 9.1 Hz, 1H), 7.11 (d, J = 5.2 Hz, 2H), 7.08 (s, 1H), 7.00 (d, J = 8.3 Hz, 2H), 6.91 (s, 1H), 6.81 (d, J = 8.4 Hz, 2H), 6.55 (s, 2H), 2.54 (s, 2H), 2.27 (t, J = 5.5 Hz, 2H), 1.61 (s, 2H). ¹³C NMR (101 MHz, DMSO- d_6) δ 186.52, 160.53, 155.72, 152.84, 150.37, 146.75, 137.45, 130.02, 128.91, 128.07, 127.64, 125.62, 124.96, 124.26, 117.00, 115.39, 114.20, 111.76, 105.07, 29.44, 21.67, 20.44. MS: m/z calculated for C₂₆H₂₁NO₃ ([M-H]⁺): 396.1;found:396.1.

Synthesis of compound 7. Dissolve 8 mL (50 mmol) of isophorone, 3.96 g (60 mmol) of Malononitrile and 0.54 mL (5 mmol) of piperidine in 50 mL of anhydrous acetonitrile. Heat and reflux at 81°C for 12 hours under nitrogen protection conditions. After passing through a silica gel chromatographic column, the compound was recrystallized with n-hexane to obtain 4.9 g of compound with a yield of 74%.¹H NMR (400 MHz, Chloroform-*d*) δ 6.61 (s, 1H), 2.51 (s, 2H), 2.17 (s, 2H), 2.03 (s, 3H), 1.01 (s, 6H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 170.34, 159.74, 120.51, 113.13, 112.35, 45.60, 42.56, 32.31, 27.75, 25.27. MS: m/z calculated for C₁₂H₁₄N₂ ([M-H]⁺): 187.1;found:187.1.

Synthesis of Compound 8 (S-OH). Compoud 5 (200 mg, 0.5 mmol), compund 7 (113 mg, 0.6 mmol) and 100 μ L piperidine were dissolved in 15 ml acetonitrile, heated at 81°C in the nitrogen atmosphere and refluxed for 10 hours, the organic phase

was concentrated to obtain 103 mg of purple product with a yield of 36%. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.56 (s, 1H), 7.49 (d, *J* = 15.5 Hz, 1H), 7.29 (t, *J* = 7.6 Hz, 2H), 7.07 (d, *J* = 8.4 Hz, 1H), 7.05 (s, 1H), 7.01 (d, *J* = 8.0 Hz, 2H), 6.96 (d, *J* = 8.5 Hz, 2H), 6.79 (d, *J* = 8.5 Hz, 2H), 6.65 (d, *J* = 11.6 Hz, 3H), 6.59 – 6.47 (m, 2H), 2.51 (s, 4H), 2.43 (s, 4H), 1.79 – 1.60 (m, 2H), 0.94 (s, 6H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 169.49, 157.41, 155.49, 153.58, 151.75, 149.66, 147.21, 137.76, 132.80, 129.86, 128.52, 127.71, 127.50, 126.07, 123.82, 123.40, 120.93, 116.91, 115.87, 115.15, 112.10, 106.71, 72.44, 55.32, 42.70, 38.51, 32.00, 29.39, 27.74, 24.69, 20.82. MS: m/z calculated for C₃₈H₃₃N₃O₂ ([M-H]⁻):562.2; found:562.2.

Synthesis of probe 9 (S-NBD). 200 mg (0.36 mmol) of S-OH was dissolved in 10 mL of dichloromethane solution. 85 mg (0.43 mmol) of 4-chloro-7-nitro-2,1,3benzoxadiazole was added under 0 °C ice bath conditions, followed by 3-5 drops of triethylamine. Stirring for 2 hours, 121 mg of product was obtained through a silica gel column with a yield of 47%. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.62 (d, *J* = 8.4 Hz, 1H), 7.53 – 7.47 (m, 1H), 7.40 – 7.31 (m, 4H), 7.16 (dq, *J* = 21.1, 8.1, 6.5 Hz, 6H), 6.85 (d, *J* = 8.6 Hz, 2H), 6.67 (d, *J* = 11.9 Hz, 3H), 6.58 (s, 1H), 2.51(s,4H), 2.42 (s, 4H), 1.69 (s, 2H), 0.94 (s, 6H). ¹³C NMR (100 MHz, DMSO-*d*₆) δ 169.61, 157.26, 153.93, 153.61, 151.28, 148.59, 148.05, 146.71, 145.75, 145.50, 144.72, 135.85, 132.95, 130.55, 130.26, 129.25, 127.79, 126.14, 125.18, 124.67, 122.62, 122.52, 121.30, 118.90, 117.60, 115.05, 114.19, 112.48, 109.81, 109.31, 42.72, 42.70, 38.49, 31.99, 29.49, 27.74, 20.76. MS: m/z calculated for C₄₄H₃₄N₆O₅ ([M-H]⁻):725.3; found: 725.3.







Figure S2¹³C NMR spectra of Compound 2 in CDCl₃.



Figure S4¹H NMR spectra of Compound 3 in CDCl₃.



Figure S6 Mass spectra of Compound 3.



Figure S7¹H NMR spectra of Compound 4 in CDCl₃.



Figure S8¹³C NMR spectra of Compound 4 in CDCl₃.



Figure S9 Mass spectra of Compound 4.



Figure S10 ¹H NMR spectra of Compound 5 in DMSO-*d*₆.











Figure S13 ¹H NMR spectra of Compound 7 in CDCl₃.



Figure S14 ¹³C NMR spectra of Compound 7 in CDCl₃.







Figure S16¹H NMR spectra of S-OH in DMSO-*d*₆.



Figure S17¹³C NMR spectra of S-OH in DMSO-*d*₆.



Figure S18¹H NMR spectra of probe S-NBD in DMSO-*d*₆.



Figure S19¹³C NMR spectra of probe S-NBD in DMSO-*d*₆.



Figure S20 Mass spectra of Probe S-NBD, S-OH and Probe S-NBD + Cys.

3. Spectrometric Studies of S-NBD



Figure S21 Fluorescence intensity at 838 nm of S-NBD in the absence or presence of Hcy in different ratios of EtOH/PBS solutions (v/v, 1/99, 1/9, 2/8, 3/7, 4/6, 5/5, 6/4, 7/3, 8/2, 9/1, 99/1).

Figure S22 Time-dependent absorption spectra of S-NBD (10 μ mol/L) in the presence of (A)10 equiv. of Cys, (B) 10 equiv. of Hcy, (C) 10 equiv. of GSH and (D)10 equiv. of NaHS in EtOH-PBS (pH 7.4, v/v, 5/5).

Figure S23 Time-dependent fluorescence spectra of S-NBD (10 μ mol/L) in the presence of (A)10 equiv. of Cys, (B) 10 equiv. of Hcy, (C) 10 equiv. of GSH and (D)10 equiv. of NaHS in EtOH-PBS (pH 7.4, v/v, 5/5).

Figure S24 Absorption spectra of S-NBD (10 μ mol/L) reacted with Cys /Hcy/GSH and NaHS (0, 1, 2, 4, 6, 8, 10, 20, 40, 60, 80,100 and 200 μ mol/L).

Figure S25 Fluorescence spectra of S-NBD. (A) Fluorescence spectra of S-NBD (10 μ mol/L) upon the addition of Cys, (B) Hcy, (C) GSH and (D) NaHS (0–200 μ mol/L). Relationship between fluorescence intensity and concentrations of (E) Cys, (F) Hcy, (G) GSH and (H) NaHS at 838 nm ($\lambda_{ex} = 660$ nm); The linear relationship between fluorescence intensity and low concentration of biothiols. The plot of fluorescence intensity against (I) Cys's and (J) Hcy's at 550 nm ($\lambda_{ex} = 470$ nm) concentration; linearity plot of the fluorescence intensity versus low concentrations of (I) Cys and (J) Hcy.

Figure S26 Photo-stability of Probe S-NBD.

Figure S27 Fluorescence spectra of S-NBD and S-NBD NPs. Fluorescence spectra of (A) S-NBD (10 μ mol/L) after Cys (0–200 μ mol/L) was added. Relationship between Cys concentrations and fluorescence intensity (B) at 550 nm ($\lambda_{ex} = 470$ nm) and (C) at 838 nm ($\lambda_{ex} = 660$ nm). Fluorescence spectra of (D) S-NBD NPs (10 μ mol/L) upon the addition of Cys (0–200 μ mol/L). Relationship between fluorescence intensity and concentrations of Cys (E) at 550 nm ($\lambda_{ex} = 470$ nm) and (F) at 838 nm ($\lambda_{ex} = 660$ nm). The linear relationship between fluorescence intensity and low concentration of Cys.

Figure S28 Time-dependent fluorescence spectra of (A) S-NBD (10 μ mol/L) and (B) S-NBD NPs (10 μ mol/L) in the presence of 10 equiv. of Cys in EtOH-PBS (pH 7.4, v/v, 5/5). Time-dependent fluorescence spectra of S-NBD and S-NBD NPs (10 μ mol/L) in the presence of 10 equiv. of of Cys (C) at 550 nm ($\lambda_{ex} = 470$ nm) and (D) at 838 nm ($\lambda_{ex} = 660$ nm).

4. Photoacoustic Imaging for Biothiols in Vitro

Figure S29 PA intensity of S-NBD (10 μ mol/L) reacted with Cys (100 μ mol/L) in EtOH/PBS (pH 7.4, v/v, 5/5) at different wavelength (680–850 nm).

Figure S30 (A) PA images of S-NBD and S-NBD NPs reacted with different concentrations of Cys (0, 10, 20, 40, 60, 80 and 100 μ mol/L) in EtOH/PBS (pH 7.4, v/v, 5/5). (B) PA signal intensity of images in (A).

5. Photothermal Imaging for Biothiols in Vitro

Figure S31 Stability of Probe S-NBD NPs.

Figure S32 The UV absorption curve as the concentration of S-NBD changes in THF.(B) Standard curve of S-NBD.

Figure S33 The temperature changes of S-NBD NPs and S-OH NPs (100 μ mol/L, 1 W/cm², 10 min) irradiated by 660 nm laser.

6. Cell Cytotoxicity by CCK-8 Assay

Figure S34 CCK-8 assay for estimating cell viability (%) of HeLa cells treated with various concentrations of S-NBD and S-NBD NPs (0–80 µmol/L) after 24 h incubation.

7. Fluorescence Microscopic Imaging

Figure S35 Fluorescence images of HeLa cells incubated with S-NBD ($10 \mu mol/L$) for different times (0.5, 1, 1.5, 2 hours). Scale bar = $10 \mu m$.

8. Photothermal Therapy in Cells

Figure S36 CCK-8 assay for estimating cell viability (%) of HeLa cells treated with various concentrations of S-NBD NPs (0–80 μ mol/L) after 24 h incubation and then treated with S-NBD NPs with 660 nm laser irradiation (0.2 W/cm², 20 min).