The Intermediary Role of 5-Pregnene-3β,20β-diol in the Biosynthesis of 16-Unsaturated C₁₉ Steroids in Boar Testis

By K. H. LOKE* and D. B. GOWER

Department of Biochemistry, Guy's Hospital Medical School, London SE1 9RT, U.K.

(Received 9 November 1971)

1. The possible involvement of 5-pregnene- 3β , 20β -diol in 16-unsaturated C₁₉ steroid biosynthesis has been investigated. 2. 5,16-Androstadien-3 β -ol (andien- β) formation from $[4^{14}C]$ pregnenolone (3 β -hydroxy-5-pregnen-20-one), 5-pregnene-3 β , 20 α -diol and 5pregnene-3 β ,20 β -diol was studied in homogenates of boar testis and the mean yields obtained were 25.6, 2.7 and 16.0% respectively. 3. Short-term kinetic studies with pregnenolone and 5-pregnene- 3β , 20β -diol separately and together suggested that the latter compound might be an intermediate in the biosynthesis of and ien- β . 4. In agreement with this interpretation, radioactive 5-pregnene- 3β , 20β -diol has been isolated during and β biosynthesis from [4-14C] pregnenolone in the presence of NADPH, more radioactivity being trapped under limiting conditions of and β formation with NADH present as cofactor. 5. Further, 5-pregnene- 3β , 20β -diol and and ien- β have been shown to inhibit the formation of the 16-unsaturated C₁₉ steroid from [4-14C]pregnenolone, the yield of radioactive 5-pregnene- 3β , 20β -diol increasing in the presence of added unlabelled and ien- β . 6. It is concluded that there may be two pathways leading to 16unsaturated C₁₉ steroid formation from pregnenolone, one of these involving 5pregnene- 3β , 20β -diol as an intermediate. Possible mechanisms are presented and discussed.

The biosynthesis in vitro of 16-unsaturated C_{19} steroids from progesterone and pregnenolone (3β) hydroxy-5-pregnen-20-one) by boar testis tissue has been described in earlier papers (Gower & Ahmad, 1967; Ahmad & Gower, 1968; Katkov & Gower, 1968; Gower & Katkov, 1969; Katkov & Gower, 1970). Various other steroidal substrates have been tested as intermediates in 16-unsaturated C₁₉ steroid biosynthesis from these two C21 precursors but all were shown to be ineffective. In continuing these earlier studies the possible involvement of a 20-hydroxy- C_{21} steroid in 16-dehydrosteroid biosynthesis was considered. To investigate this possibility the conversion of pregnenolone into and (5,16-and rostadien- 3β -ol) has been studied in detail since the latter is formed in high yields and may be the first 16unsaturated C_{19} steroid formed from pregnenolone (Katkov & Gower, 1970). Some of these results have appeared in a preliminary communication (Loke & Gower, 1971).

Materials and Methods

Materials

Authentic Δ^{16} -steroids, solvents and materials for radioautography, t.l.c. and g.l.c. were as described

* Present address: Department of Biochemistry, University of Malaya.

Vol. 127

by Gower & Ahmad (1967). Dry hexane (laboratory grade; Hopkin and Williams Ltd., Chadwell Heath, Essex, U.K.) was redistilled and dried over sodium. All reagents, unless otherwise stated, were analytical grade. [4-¹⁴C]Pregnenolone (specific radioactivity 55.7 mCi/mmol) was purchased from The Radiochemical Centre, Amersham, Bucks., U.K. Radioactivity was measured by liquid-scintillation counting as described by Gower & Ahmad (1967) and the weight of andien- β by g.l.c. as described by Gower & Thomas (1968).

Methods

Preparation of the C-20 epimeric 5-pregnene- 3β ,20 β diols. These were prepared by treating [4-1⁴C]pregnenolone (0.25–0.5 μ Ci) in methanol (0.1ml) with potassium borohydride (5–10mg) at room temperature. After at least 30min the reaction mixture was diluted with 0.1M-acetic acid and extracted with ethyl acetate. The dried extract was evaporated and the residue subjected to t.l.c. in benzene – acetone (4:1, v/v). After radioautography, two radioactive spots corresponding to authentic 5-pregnene- 3β ,20 α -diol (R_F 0.22–0.27) and 5-pregnene- 3β ,20 β -diol (R_F 0.26–0.31) were seen and these were eluted for use in subsequent incubations. The ratio of the yield of the 20 α - to 20 β -epimers was approximately 1 to 5. Pregnenolone had $R_F 0.35-0.45$.

Determination of protein. The protein content of tissue homogenates was determined by the method of Lowry et al. (1951) with crystalline bovine plasma albumin (Armour Pharmaceuticals Ltd., Eastbourne, Sussex, U.K.) as standard.

Preparation of tissue and incubation conditions. Boar testis that had been stored at -20° C was used throughout. The preparation and incubation of tissue homogenates were carried out as described by Katkov & Gower (1970). Unless otherwise stated, 0.2ml portions of a 10% (w/v) homogenate in 0.25 M-sucrose were used in the incubations and these were found to contain 1200-1400 μ g of protein. The incubations were terminated by the addition of ethyl acetate (2ml) and carrier steroids.

Isolation and purification of metabolites. Andien- β was isolated and purified as described by Katkov & Gower (1970) and the yield was corrected for analytical losses. Pregnenolone and dehydroepiandrosterone were purified by t.l.c. in benzene-acetone (4:1, v/v). For the isolation of the metabolites 17-hydroxypregnenolone (3 β ,17-dihydroxy-5-pregnen-20-one) and 5-pregnene-3 β ,20 β -diol, however, which are not separated by t.l.c. in this system, a Girard separation (see below) was first performed on the mixture. Carrier steroids were then added to the appropriate fractions obtained and recrystallized to constant specific radioactivity.

In a typical experiment with [4-14C]pregnenolone as substrate, radioactive 17-hydroxypregnenolone was isolated as above. Carrier (10mg) was added and the specific radioactivities of crystals after successive recrystallizations in ethanol were 3880, 3790 and 3940d.p.m./mg.

Girard separation. The procedure described by Bush (1961) was used except that the reaction with the Girard T reagent was carried out at room temperature.

Results

Formation of andien- β from pregnenolone, 5-pregnene-3 β ,20 α -diol and 5-pregnene-3 β ,20 β -diol in boar testis homogenate

[4-¹⁴C]Pregnenolone, 5-[4-¹⁴C]pregnene- 3β ,20 α diol and 5-[4-¹⁴C]pregnene- 3β ,20 β -diol (each 9.4-11.0×10⁴ d.p.m.) were separately incubated at 37°C for 20min with portions (0.1 ml) of boar testis homogenate in 0.05*m*-tris-HCl buffer, pH 7.4, containing 0.4mm-NADPH. After addition of carrier andien- β (50µg), radioactive andien- β was isolated and assayed. The mean yields (four experiments) of andien- β formed from pregnenolone, 5-pregnene- 3β ,20 α -diol and 5-pregnene- 3β ,20 β -diol were 25.6, 2.7 and 16.0% respectively. A control experiment in

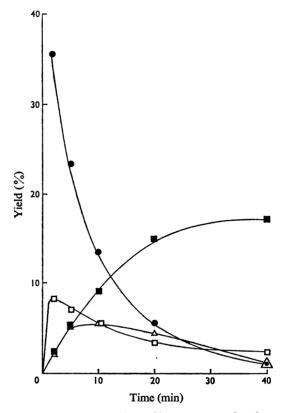


Fig. 1. Metabolism of 5-[4-¹⁴C]pregnene-3β,20β-diol in boar testis homogenate

Portions (0.1 ml) of boar testis homogenate were incubated with 5-[4-14C]pregnene- 3β ,20 β -diol (816000 d.p.m.) at 37°C in 1.2 ml of 0.05 M-tris – HCl buffer, pH7.4, containing 0.4 mM-NADPH. The reactions were terminated at various times, carrier steroids were added and metabolites extracted and isolated as described in the Materials and Methods section. •, 5-Pregnene- 3β ,20 β -diol; \Box , pregnenolone; \triangle , 17-hydroxypregnenolone; \blacksquare , andien- β .

which boiled homogenate was used did not yield any andien- β from 5-pregnene- 3β ,20 β -diol. The pregnenolone isolated from the incubation of 5-[4-¹⁴C]pregnene- 3β ,20 α -diol contained very small amounts of radioactivity although the 20 β -isomer under the same conditions yielded pregnenolone containing approximately 3% of the initial radioactivity (but see also below and Fig. 1).

Kinetic studies of 5-pregnene- 3β ,20 β -diol and pregnenolone metabolism

With 5-pregnene- 3β , 20β -diol. Portions of boar testis homogenate (0.1 ml) were incubated in separate

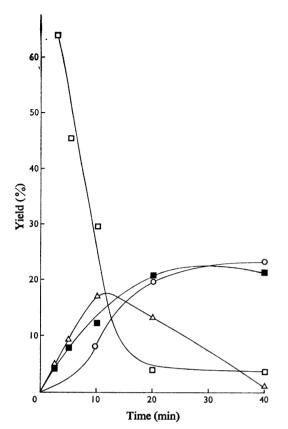


Fig. 2. Metabolism of [4-14C]pregnenolone in boar testis homogenate

[4-14C]Pregnenolone (932000d.p.m.) was incubated with portions of boar testis homogenate (0.1 ml) under conditions described in the legend to Fig. 1. \Box , Pregnenolone; \triangle , 17-hydroxypregnenolone; o, dehydroepiandrosterone; \blacksquare , andien- β .

flasks with 5-[4-¹⁴C]pregnene-3 β ,20 β -diol (81600 d.p.m.) dissolved in buffer (1.2ml) containing 8mm-NADPH. After 2, 5, 10, 20 and 40min respectively carrier andien- β (50 μ g) was added to each flask and the metabolites were extracted and separated as described in the Materials and Methods section. Fig. 1 shows the yields of metabolites plotted against time of incubation.

With pregnenolone. The metabolism of [4-¹⁴C]pregnenolone by boar testis homogenate was studied over short periods of time in a manner similar to that described for 5-pregnene- 3β ,20 β -diol. The percentage yields of andien- β , pregnenolone, dehydroepiandrosterone and 17-hydroxypregnenolone were plotted against time of incubation (Fig. 2). Radioactive 5-pregnene- 3β ,20 β -diol was not isolated from this experiment, but in further studies it was possible to demonstrate its formation from [¹⁴C]-pregnenolone under the following conditions of andien- β biosynthesis.

(a) In NADPH-supported incubations. A portion of boar testis homogenate (0.4ml) was incubated at 37° C with [4-¹⁴C]pregnenolone (1670000d.p.m.) and 0.4mM-NADPH for 20min. Radioactive and ien- β and 5-pregnene- 3β ,20 β -diol were isolated and purified as described above, the corrected yield of andien- β being 21.0%, whereas that for the 5pregnene- 3β ,20 β -diol, isolated and characterized by reverse isotope dilution (Table 1), was 0.23%.

(b) In NADH-supported incubations. Since Katkov & Gower (1970) have found that NADH was less active than NADPH in promoting and β biosynthesis from pregnenolone, two experiments were performed to determine if there was a tendency for the postulated intermediary diol to accumulate under such limiting conditions of and β formation. Boar testis homogenate was incubated at 37°C for 20 min with [4-14C]pregnenolone, as in the NADPHsupported experiment above, but 0.6mm-NADH was added as cofactor instead of NADPH. The yields of and ien- β and 5-pregnene-3 β ,20 β -diol isolated were 5.7 and 0.33% (0.34% in a second experiment) respectively. The use of NADH therefore resulted in only 27% of the andien- β obtained when NADPH was present.

Kinetic study of $[7\alpha^{-3}H]$ pregnenolone plus 5- $[4^{-14}C]$ pregnene- 3β ,20 β -diol

Portions of boar testis homogenate (0.1 ml) were incubated at 37°C as described above with mixtures of $[7\alpha^{-3}H]$ pregnenolone (98000d.p.m.) and 5-[4-¹⁴C]pregnene-3 β ,20 β -diol (66200d.p.m.) for 2, 5, 10, 20 and 40min. NADPH (0.3 mM) was included in each tube. Table 2 shows that the yield of [³H]andien- β and [¹⁴C]andien- β isolated (corrected for losses) increased progressively up to 30min, comparable with increases depicted in Figs. 1 and 2 when pregnenolone and 5-pregnene-3 β ,20 β -diol were incubated separately. The ³H/¹⁴C ratios of the andien- β samples, however, decreased as the time of incubation increased (Table 2).

Studies on the inhibition of and ien- β biosynthesis

These experiments were designed to investigate whether andien- β formation from pregnenolone was affected by preincubation of boar testis homogenate with the postulated intermediate, 5-pregnene- 3β ,20 β diol, the product andien- β or the well-known intermediate in androgen biosynthesis, 17-hydroxypregnenolone. Three tubes containing tris-HCl buffer, pH7.4 (1.2ml), boar testis homogenate

Table 1. Purification by recrystallization to constant specific radioactivity (d.p.m./mg) of 5-pregnene-3β,20β-diol obtained after incubating [4-14C]pregnenolone with boar testis homogenates

Values in parentheses are the weights (mg) of crystals. Solvents were: A, ethyl acetate; B, hexane; C, acetone; D, benzene. Boar testis homogenate (0.4ml) was incubated with [4-¹⁴C]pregnenolone (1670000d.p.m.) for 20 min at 37°C in the presence of 0.4mm-NADPH or 0.6mm-NADH. After the addition of carrier, radioactive andien- β was purified by conventional t.l.c. and by t.l.c. on plates impregnated with AgNO₃ (see the text), whereas 5-pregnene-3 β ,20 β -diol was isolated by t.l.c. and by recrystallization to constant specific radioactivity.

| In NADPH-supported incubations | | | Expt. 1 | | | Expt. 2 | | |
|---|---------|-------------------------|---|-------------|-------------------------|---|---------|-------------------------|
| Number of recrystal- lizations | Solvent | Sp. radioactivity | Number of recrystal- lizations | Solvent | Sp. radioactivity | Number of recrystal- lizations | Solvent | Sp. radioactivity |
| 0 | | 1330 (10.0) | 0 | | 915 (10.8) | 0 | | 1138 (8.5) |
| 1 | A,B | 638 (4.2) | ĩ | Α | 637 (2.6) | ı 1 | D | 559 () |
| 2 | C,B | 384 (3.5) 199 (5.8)* | 2 | D | 513 (1.0) 149 (2.5)* | 2 | Α | 559 (2.6) 229 (3.9)* |
| 3 | D | 208 (4.3) | 3 | C,B | 156 (2.0) | 3 | C,B | 324 (—) |
| 4 | A,B | 191 (1.9) | | • | | | | • • |
| | · | | * After | addition of | more carrier. | | | |

In NADH-supported incubations

Table 2. Kinetic study of the formation of 5,16-androstadien-3 β -ol from $[7\alpha^{-3}H]$ pregnenolone and 5- $[4^{-14}C]$ -pregnene-3 β ,20 β -diol in boar testis in vitro

 $[7\alpha^{-3}H]$ Pregnenolone (98000d.p.m.) and 5-[4-¹⁴C]pregnene-3 β ,20 β -diol (66200d.p.m.) were incubated with portions of boar testis homogenate (0.2ml) in the presence of 0.4mM-NADPH for various times. Labelled and ien- β was then isolated, purified and the ³H and ¹⁴C contents of each sample were measured. All percentages are corrected for analytical losses.

Time of incubation

| (min) | 2 | 5 | 10 | 20 | 40 |
|---|------|------|------|------|------|
| ³ H in and ien- β (%) | 5.8 | 10.9 | 16.1 | 20.7 | 23.8 |
| ¹⁴ C in and ien- β (%) | 2.2 | 5.0 | 7.8 | 14.6 | |
| ³ H/ ¹⁴ C ratio | 2.64 | 2.18 | 2.07 | 1.42 | 1.23 |

(0.2 ml containing $1250\,\mu g$ of protein N), NADPH (0.4 mM) and 5-pregnene- $3\beta,20\beta$ -diol (200 μg), andien- β (210 μg) or 17-hydroxypregnenolone (200 μg) respectively were preincubated at 37°C for 5 min. A control tube containing buffer, NADPH and homogenate was likewise incubated. [4-14C]Pregnenolone (405000 d.p.m.) was then added to each tube and incubation continued for a further 20 min. Table 3 shows that andien- β formation was inhibited by preincubation with 5-pregnene- $3\beta,20\beta$ -diol (74.9%) and by andien- β (44.3%). Preincubation with 17-hydroxypregnenolone, however, had no effect on the yield of andien- β obtained from pregnenolone. After isolating and recrystallizing the 5-pregnene-3 β ,20 β -diol to constant specific radioactivity, it was calculated that 0.12 and 0.24% of substrate radioactivity was trapped in tubes preincubated with 5-pregnene-3 β ,20 β -diol and andien- β respectively.

The effect of pregnenolone on the metabolism of 5-pregnene- 3β ,20 β -diol to andien- β was studied as follows. Boar testis homogenate (0.2ml) was preincubated (5min) with pregnenolone (200 μ g) in tris-HCl buffer, pH7.4 (1.2ml), containing 0.4mM-NADPH. 5-[4-¹⁴C]Pregnene- 3β ,20 β -diol (66200 d.p.m.) was then added and the incubation continued for a further 20min. The yield of [¹⁴C]andien- β isolated was only 0.1% compared with 14.6% (cf. Table 2) in a control incubation without pregnenolone. Moreover, the pregnenolone subsequently isolated was devoid of radioactivity.

Discussion

In the studies testing the capacity of the epimeric compounds 5-pregnene- 3β ,20 α -diol and 5-pregnene- 3β ,20 β -diol to serve as precursors of andien- β in boar testis, and comparing their efficiency in this respect with that of pregnenolone, it was found that 5-pregnene- 3β ,20 β -diol was six to seven times more effective in forming andien- β than the 20 α -epimer. This result, although in agreement with the working hypothesis, is by itself not proof for the intermediary

Table 3. Effect of various steroids on the formation of 5,16-androstadien- 3β -ol from pregnenolone in boar testis in vitro

Boar testis homogenate (0.2ml containing $1250 \mu g$ of protein N) was incubated for 5min with the unlabelled steroids as indicated. [4-14C]Pregnenolone (405000d.p.m.) was then added and the incubation continued for a further 20min, after which and ien- β and 5-pregnene- 3β ,20 β -diol were isolated as described in the text. N.D., not determined.

| Steroid added | None | 5-Pregnene-3 β ,20 β -diol (200 μ g) | Andien- β (210 μ g) | 17-Hydroxy- pregnenolone (200 μ g) |
|--|------|---|-------------------------------|---|
| Yield of and ien- β (%) | 25.1 | 6.3 | 13.9 | 26.4 |
| Inhibition compared with yield in control tube (%) | 0 | 74.9 | 44.3 | 0 |
| Substrate radioactivity trapped in 5-pregnene-3β,20β-diol | | 0.12 | 0.21 | N.D. |
| | | | | |

role of the 20β -hydroxy steroid in andien- β biosynthesis, since it is possible that by its preferential oxidation to pregnenolone it could give rise to andien- β .

The subsequent short-term kinetic studies on pregnenolone and 5-pregnene-3 β ,20 β -diol metabolism by boar testis were therefore carried out to shed some light on this problem. As can be seen from Fig. 1 both pregnenolone and and β were formed rapidly in the early stages of the metabolism of 5-pregnene- 3β , 20 β -diol, the yields of pregnenolone being greater than those of and β . However, after increasing to a maximum, the amount of pregnenolone declined to low values with increasing time of incubation whereas the andien- β in the medium progressively increased and then levelled off with depletion of the substrate. Moreover, it appeared from the kinetic results that the pregnenolone formed was mainly channelled into the biosynthesis of 17-hydroxypregnenolone. When considered together, the results are not inconsistent with the existence of a pathway leading directly from 5pregnene-3 β ,20 β -diol to and ien- β .

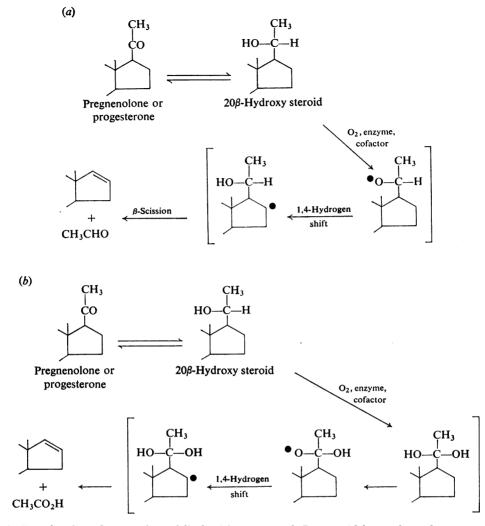
In the kinetic study with pregnenolone as substrate (Fig. 2), 17-hydroxypregnenolone was formed that bore a precursor-product relationship with pregnenolone and, moreover, was almost certain to be a precursor of dehydroepiandrosterone in boar testis.

The case for the intermediacy of 5-pregnene- $3\beta,20\beta$ -diol in andien- β biosynthesis would be considerably strengthened if this compound could be isolated under the conditions of andien- β biosynthesis from pregnenolone. As reported in the Results section, attempts to isolate the intermediate from NADPH-supported incubations, which have been shown earlier by Katkov & Gower (1970) to result in optimum yields of andien- β , were successful. The yields obtained were generally low (0.2%) and this fact would seem to indicate a rapid utilization of the postulated intermediate as soon as it was formed. This could also be isolated from the incubations with NADH as cofactor and it may be significant that there was a greater yield of the 20β -hydroxy steroid under these experimental conditions than when andien- β synthesis was occurring in optimum amounts. The finding that only 27% of andien- β was formed in the presence of NADH compared with the yield in the presence of NADPH confirms the results of Katkov & Gower (1970), who obtained a value of 21.3%.

If the 20 β -ol were involved in andien- β biosynthesis, it seems reasonable to expect that preincubation of the enzyme system with 5-pregnene-3 β ,20 β -diol would result in an inhibition of the formation of andien- β from pregnenolone. This was indeed found to be the case (Table 3) and, moreover, consistent with its intermediary role, substrate radioactivity was trapped by the added inhibitor. Further, the presence of excess of andien- β also caused inhibition, and twice as much radioactivity was trapped in the postulated intermediate (Table 3).

The cumulative evidence therefore suggests an intermediary role for 5-pregnene- 3β , 20β -diol in and β biosynthesis. However, at present it is uncertain if the formation of and β from pregnenolone must compulsorily involve the 208hydroxy derivative. The finding that the yield of and ien- β from the 5-pregnene-3 β ,20 β -diol was about 65% that from pregnenolone would seem to indicate the existence of another pathway by-passing the 20β -hydroxy derivative. The fact that preincubation with unlabelled pregnenolone almost eliminates and β synthesis, even though the pregnenolone subsequently isolated was devoid of radioactivity, suggests that and β formation is not occurring from 5-pregnene-3 β ,20 β -diol via pregnenolone. This view is supported by the kinetic results shown in Figs. 1 and 2.

Taking into consideration the results obtained from the double-isotope experiment (Table 2), it seems likely that, since the ${}^{3}H/{}^{14}C$ ratios fell progressively as incubation time increased, the con-



Scheme 1. Postulated mechanisms (a and b) for 16-unsaturated C_{19} steroid biosynthesis from pregnenolone or progesterone involving 20β -hydroxy- C_{21} steroid intermediates

version of pregnenolone into andien- β is faster than that of 5-pregnene- 3β ,20 β -diol into andien- β . The intermediary role of a 20 β -hydroxy- C_{21} steroid in 16-unsaturated C_{19} steroid synthesis may be rationalized by one or both of the mechanisms shown in Scheme 1. Both mechanisms are based on the postulate of Lieberman *et al.* (1969), which stated that, in those steroid biosyntheses utilizing oxygen, freeradical intermediates might be involved. In agreement with this postulate Lippman & Lieberman (1970) have reported the conversion of deoxycorticosterone (21-hydroxy-4-pregnene-3,20-dione) nto low yields of 4,16-androstadien-3-one by boar testis homogenate and by the action of free-radicalgenerating reagents. On the basis of their findings these workers have suggested a possible mechanism for 16-unsaturated C_{19} steroid biosynthesis involving a C-21 alkoxy radical.

The reaction of 20β -hydroxy steroids with lead tetra-acetate (a free-radical-generating reagent) has been extensively used in the introduction of functionality into angular methyl groups and is well documented in chemical literature (Erikson & Forbess, 1963). However, the finding of Cainelli *et al.* (1961) that a ring D-saturated C₁₉ steroid was among the products of the reaction between the 3-ethylene ketal derivative of 20β -hydroxy-4-pregnen-3-one and lead tetra-acetate is particularly noteworthy. Thus by analogy it is conceivable that, under the influence of NADPH, oxygen and the enzyme system, the 20β - hydroxy steroid would give rise to a C-20 alkoxy radical, that would then undergo a 1,4-hydrogen shift to the C-16 radical. Subsequent β -scission of the C-16 radical would result in the formation of a 16-unsaturated C₁₉ steroid and acetaldehyde (Scheme 1*a*).

Mechanism (b) is a variant of (a) and its key feature is the hypothetical C-20 dihydroxylated intermediate, which would either form the 20-ketone or undergo fragmentation to a 16-unsaturated C₁₉ steroid and acetic acid as shown. As observed in the kinetic experiment with 5-pregnene- 3β ,20 β -diol, pregnenolone was formed early in the incubation and, since its formation occurred in the presence of NADPH and oxygen, the mechanism may be considered as analogous to that proposed by Skinner & Akhtar (1968) to account for the conversion under similar conditions of 19-hydroxy-4-androstene-3,17dione by placental microsomes into 19-oxo-4androstene-3,17-dione.

Skinner & Akhtar (1969) have emphasized that the removal of C-19 in oestrogen biosynthesis must occur at the oxidation state of a 19-aldehyde with the liberation of formic acid. By analogy it is possible that the 20-ketone or its probable equivalent, the C-20 dihydroxylated intermediate, may undergo fragmentation to the 16-dehydrosteroid and acetic acid.

It is noteworthy that 16-unsaturated C_{19} steroids have been chemically synthesized from the mesylates or tosylates of pregnane-16,20-diols (Matsui & Fukushima, 1970). The possibility that such transformations may occur in boar testis tissue has yet to be investigated.

It should be possible to distinguish the mechanisms from one another by a study of the intermediates involved or the nature of the C_2 fragment liberated or both; preliminary results, obtained by using $[21-^{14}C]$ -progesterone with boar testis homogenates, indicate that acetaldehyde may be liberated during the reaction.

We are grateful to Professor G. A. D. Haslewood for continued interest and helpful criticism and to Dr. S. Lieberman for allowing us to read a manuscript before publication. K. H. L. thanks Professor Haslewood for the hospitality shown him on his study leave from the University of Malaya.

References

- Ahmad, N. & Gower, D. B. (1968) Biochem. J. 108, 233
- Bush, I. E. (1961) *The Chromatography of Steroids*, p. 362, Pergamon Press, London
- Cainelli, G., Kamber, B., Keller, J., Mikailovic, M. Lj., Arigoni, D. & Jeger, O. (1961) Helv. Chim. Acta 44, 518
- Erikson, J. M. & Forbess, D. L. (1963) in *Steroid Reactions* (Djerassi, C., ed.), p. 327, Holden-Day Inc., San Francisco
- Gower, D. B. & Ahmad, N. (1967) Biochem. J. 104, 550
- Gower, D. B. & Katkov, T. (1969) Biochem. J. 115, 16P
- Gower, D. B. & Thomas, B. S. (1968) J. Chromatogr. 36, 338
- Katkov, T. & Gower, D. B. (1968) Biochim. Biophys. Acta 164, 134
- Katkov, T. & Gower, D. B. (1970) Biochem. J. 117, 533
- Lieberman, S., Bondy, L., Lippman, V. & Roberts, K. D. (1969) Biochem. Biophys. Res. Commun. 34, 367
- Lippman, V. & Lieberman, S. (1970) Proc. Nat. Acad. Sci. U.S. 67, 1754
- Loke, K. H. & Gower, D. B. (1971) Biochem. J. 122, 27 P
- Lowry, O. H., Rosebrough, N. J., Farr, A. L. & Randall, R. J. (1951) J. Biol. Chem. 193, 265
- Matsui, M. R. & Fukushima, D. K. (1970) J. Org. Chem. 35, 561
- Skinner, S. J. M. & Akhtar, M. (1968) Biochem. J. 109, 318
- Skinner, S. J. M. & Akhtar, M. (1969) Biochem. J. 114, 75