

A Munc13/RIM/Rab3 Tripartite Complex: From Priming to Plasticity?

Irina Dulubova¹, Xuelin Lou², Jun Lu¹, Iryna Huryeva¹, Amer Alam¹, Ralf

Schneggenburger², Thomas C. Südhof³ and Josep Rizo^{1,4}

¹*Departments of Biochemistry and Pharmacology, ³Center for Basic Neuroscience, Department of Molecular Genetics, and Howard Hughes Medical Institute, University of Texas Southwestern Medical Center, 5323 Harry Hines Blvd., Dallas, Texas 75390, USA*

²*Max Planck Institute for Biophysical Chemistry, AG Synaptic Dynamics & Modulation and Dept. of Membrane Biophysics, Am Fassberg 11, 37077 Germany*

⁴ Corresponding author. E-mail: jose@arnie.swmed.edu. Phone: 214-645-6360. FAX: 214-645-6353

Bacterial expression vectors.

Plasmids for bacterial expression of RIM2 α , Munc13-1 and Rab3A fragments were generated by polymerase chain reaction (PCR) with custom-designed primers and subcloned into pGEX-KG (Guan and Dixon, 1991), pGEX-KT (Hakes and Dixon, 1992) or pET-21 (Novagen) vectors. The longest RIM2 α fragment (residues 1-165) in pGEX-KG vector was flanked by *Bam*HI and *Hind*III restriction sites. RIM2 α fragments (residues 22-81, 22-155, 82-142 and 82-155) were amplified on a RIM2 α ₁₋₁₆₅ template using corresponding pairs of the following primers that incorporated either *Bam*HI or *Eco*RI restriction sites: (1) 5'-AGGGATCCCCGCAGCCCGAGATGCCG-3', (2) 5'-CAGGATCCCAGCAAGAGCAGAAGGGCGA-3', (3) 5'-GCGAATTCTACTGTTGTGATTCCTCTCCCA-3', (4) 5'-GTGAATTCTACCCACTATTGTAAAACCACG-3' and subcloned into pGEX-KT vector. Site-directed mutagenesis of RIM2 α ₈₂₋₁₄₂ fragment was performed using QuickChangeTM Site-Directed Mutagenesis Kit (Stratagene).

The plasmid for bacterial expression of the recombinant rat Munc13-1 fragment (residues 3-317) in pGEX-KG vector was described earlier (Betz *et al*, 2001). The shorter Munc13-1 N-terminal fragments (residues 3-132, 3-150, 3-209, 3-228 and 3-252) were generated by PCR on the Munc13-1₃₋₃₁₇ template using sense primer 5'-GGGCTGGCAAGCCACGTTTGGT-3' that initiates in the pGEX vector sequence 5'- to the MCS region and the following antisense primers introducing a *Hind*III sequence: (1) 5'-GTAAGCTTTCAGTCCAAAGGCAGCTCAAAA-3', (2) 5'-CTAAGCTTAGGCATTTCAGCTGCTCCA-3', (3) 5'-GTAAGCTTATGGGATGCTGTTGCTCGTCT-3', (4) 5'-AGAAGCTTACCG

CACGGAGTACTGGTG-3', (5) 5'-ACAAGCTTACCGCGGCTCAGAGAACTC-3', and subcloned in pGEX-KG vector using the *EcoRI* and *HindIII* cloning sites. To produce T7-tagged Munc13-1 fragments, the corresponding *EcoRI/HindIII* fragments were subcloned from pGEX-KG into the pET-21 bacterial expression vector (Novagen).

Plasmids for bacterial expression of Rab3A fragments (residues 15-217) were generated by PCR with the following primers: 5'-AGGGATCCGACCAGAACTTCGACTATATG-3' and 5'-GCGAATTCAATCCTGATGAGGTGGCG-3' using earlier described constructs (Schluter *et al*, 2002) encoding mouse full-length Rab3A (either Q81L or T36N mutant) as template. PCR fragments were subcloned in pGEX-KT vector using *BamHI* and *EcoRI* restriction sites. All constructs were verified by sequencing.

Betz A, Thakur P, Junge HJ, Ashery U, Rhee JS, Scheuss V, Rosenmund C, Rettig J, Brose N (2001) Functional interaction of the active zone proteins Munc13-1 and RIM1 in synaptic vesicle priming. *Neuron* **30**: 183-96

Guan, KL and Dixon JE (1991) Eukaryotic proteins expressed in *Escherichia coli*: an improved thrombin cleavage and purification procedure of fusion proteins with glutathione S-transferase. *Anal. Biochem.*, **192**, 262-267.

Hakes DJ and Dixon JE (1992) New vectors for high level expression of recombinant proteins in bacteria. *Anal. Biochem.*, **202**, 293-298.

Schluter OM, Khvotchev M, Jahn R, Sudhof TC (2002) Localization versus function of Rab3 proteins. Evidence for a common regulatory role in controlling fusion. *J Biol Chem.*, **277**:40919-29