

## CONTRACTILE PROPERTIES OF SKELETAL MUSCLES FROM YOUNG, ADULT AND AGED MICE

BY SUSAN V. BROOKS AND JOHN A. FAULKNER\*

*From the Department of Physiology, Bioengineering Program, and the Institute of Gerontology, University of Michigan Medical School, Ann Arbor, MI 48109, U.S.A.*

(Received 6 July 1987)

### SUMMARY

1. Comparisons were made *in vitro* at 25 °C among soleus and extensor digitorum longus (EDL) muscles from young (2–3 months), adult (9–10 months), and aged (26–27 months) male mice. We tested the hypotheses that, compared with soleus and EDL muscles of young and adult mice, those from aged mice develop decreased maximum tetanic force ( $P_0$ , mN) and specific  $P_0$  (N/cm<sup>2</sup>), and that no significant differences occur for contraction time, half-relaxation time, or force–velocity relationship.

2. For the aged mice, the  $P_0$  of the soleus muscles and EDL muscles were 78 and 73% respectively of the values for adult mice. The specific  $P_0$  of EDL muscles of aged mice was 78% of the value of 23 N/cm<sup>2</sup> obtained for young and adult mice. For soleus muscles, the specific  $P_0$  of 21 N/cm<sup>2</sup> did not change with age.

3. Compared to values for young and adult mice, the contraction and half-relaxation times of soleus muscles from aged mice were increased, but the overall force–velocity relationships of soleus and EDL muscles did not change. The pooled values for the maximum velocity of unloaded shortening extrapolated from the force–velocity relationship of soleus and EDL muscles were 4.6 and 10.1 fibre lengths/s, respectively.

4. The decrease in the specific  $P_0$  of the EDL muscle with ageing must result from either a decrease in the number of cross-bridges in the driving stroke or a decrease in the force developed by each cross-bridge.

### INTRODUCTION

During the past 150 years, investigators have studied many different muscles, but the basic observation of Quetelet (1835) remains that between 30 and 80 years of age muscle strength decreases 30–40%, with men showing a greater loss than women (Larsson, Grimby & Karlsson, 1979; Grimby & Saltin, 1983; Young, Stokes & Crowe, 1984, 1985). The limited observations on the maximum isometric tetanic force ( $P_0$ , mN) developed by small whole muscles of rodents studied *in vitro* (Gutmann & Carlson, 1976; Fitts, Troup, Witzmann & Holloszy 1984; Larsson & Edström, 1986)

\* To whom correspondence should be sent at the Department of Physiology, University of Michigan, M7774 Medical Science II, Box 0622, Ann Arbor, MI 48109-0010, U.S.A.

support the premise that a decrease in  $P_0$  with age is not unique to humans. In a 1983 review, Grimby & Saltin concluded that the decrease in muscle strength with age correlated highly with the decrease in muscle mass and there was no need to propose changes in the composition of skeletal muscle with ageing. More recent estimates of maximum specific strength suggest a decrease of maximum specific strength with age for men (Young *et al.* 1985) but no change for women (Young *et al.* 1984). For muscles from rats, Fitts *et al.* (1984) and McCarter & McGee (1987) reported no significant differences in the specific  $P_0$  (N/cm<sup>2</sup>) with age. Consequently, the issue of a decrease in the specific  $P_0$  of muscle in aged compared to young animals remains controversial.

The effect of age on the velocity of shortening of human muscles has not been studied extensively. Larsson *et al.* (1979) initially reported that the maximum knee extension velocity of 65 year old men was 66% that of young men, but Grimby & Saltin (1983) reassessed the data of this and other studies and concluded that 'the steepness of the force-velocity curve did not differ markedly' between young and old subjects. In a study of soleus and EDL muscles from 9- and 28-month-old rats, Fitts *et al.* (1984) found no change with age for the maximum velocity of unloaded shortening ( $V_{\max}$ ) extrapolated from the force-velocity relationship. For small bundles of fibres and single skinned fibre segments obtained from 9-month- and 29- to 30-month-old male rats, Eddinger, Cassens & Moss (1986) concluded that soleus muscles became 'faster and stronger'. No significant difference was observed with ageing for the  $P_0$  or  $V_{\max}$  of bundles from EDL muscles, but the velocities at intermediate after-loads were lower.

The absence of adequate data on the contractile properties of whole skeletal muscles from young, adult and aged rodents measured *in vitro* prevents an unequivocal statement as to the deficit for  $P_0$  and specific  $P_0$  and a resolution of the conflicting data on  $V_{\max}$  and the force-velocity relationship of slow and fast muscles. To resolve these issues, data were collected on the soleus and EDL muscles from 3-4 month (young), 9-10 month (adult), and 26-27 month (aged) male mice. We tested the hypotheses that: (1) muscles from aged mice develop a lower  $P_0$  and specific  $P_0$  than muscles from young or adult mice; and (2) with ageing, no significant differences occur in contraction or relaxation times or force-velocity relationships. Our results support both hypotheses, with the exceptions that for soleus muscles of aged mice, compared to those from young or adult mice, the specific  $P_0$  did not change, the contraction and relaxation times were prolonged and the frequency-force relationship was shifted upwards and to the left. A brief report of these results was presented to the American Biophysical Society (Brooks & Faulkner, 1987).

#### METHODS

Experiments were performed on fifty-five male mice of the C57BL/6 strain. All experiments were conducted in accordance with the policy statement outlining the care and use of animals published by the American Physiological Society. Mice were anaesthetized with an initial intraperitoneal injection of sodium pentobarbitone (40 mg/kg). Supplemental injections were given as needed. Contractile properties were measured *in vitro* and in most cases on two soleus and two EDL muscles from each young, adult or aged mouse. The small 8-11 mg muscles of mice met all of the criteria chosen previously for a viable and stable muscle preparation (Segal & Faulkner, 1985). No muscle showed a decrease in  $P_0$  greater than 15%.

Following the removal of the muscles, the anaesthetized mice were killed by an overdose of sodium pentobarbitone. The muscles were isolated and 5-0 silk suture was tied securely to the proximal and distal tendons. Each muscle was then removed and immersed in a horizontal bath containing buffered physiological salt solution (composition in mM: NaCl, 137; NaHCO<sub>3</sub>, 24; glucose, 11; KCl, 5; CaCl<sub>2</sub>, 2; MgSO<sub>4</sub>, 1; NaH<sub>2</sub>PO<sub>4</sub>, 1; and tubocurarine chloride, 0.025). The solution was maintained at 25 °C, and the pH was maintained at approximately 7.4 by buffering with a gas mixture of 95% O<sub>2</sub> and 5% CO<sub>2</sub>. The distal tendon of the muscle was attached to an inflexible post and the proximal end was tied directly to the lever arm of a position feed-back servomotor (Cambridge Technology Inc., Model 300H). The motor and associated electronics provided position and force information as well as variable electromagnetic after-loads.

Muscles were stimulated directly by an electrical field generated between two platinum electrodes. All muscle fibres in a muscle were activated simultaneously by stimulation with 200  $\mu$ s pulses of supramaximal intensity. Muscle length was adjusted until a single stimulus pulse elicited maximum force during an isometric twitch. The twitch force ( $P_t$ ) and contraction and half-relaxation times were measured during each isometric twitch for each muscle. Although for individual soleus and EDL muscles of mice the optimal length for  $P_t$  and  $P_0$  might vary by a maximum of  $\pm 2\%$  of muscle length, the mean optimal lengths for  $P_t$  and  $P_0$  of each of these muscles were not significantly different. Furthermore, a change of this magnitude for muscle length had no significant effect on mean  $P_0$  of either muscle for any of the three age groups. Other investigators (Larsson & Edström, 1986) have reported no significant differences in optimal length for  $P_t$  and  $P_0$  for slow and fast muscles of adult and old rats. The determination of optimal length by twitches is energetically less demanding than determination by tetanic contractions, consequently, the optimal muscle length ( $L_0$ ) for the development of twitch force was determined and then held constant for all subsequent isometric measurements.

A frequency-force curve was constructed for each muscle from records of the force exerted during periods of stimulation at increasing frequencies. The frequency-force curves for soleus and EDL muscles from young, adult and aged mice are shown in Fig. 1.  $P_0$  was defined as the maximum isometric tetanic force on the frequency-force curve. The twitch/tetanus ratio was calculated ( $P_t/P_0$ ). The stimulus frequency that produced the maximum rate of force development ( $dP/dt_0$ ) was determined by increasing the stimulus frequency from that necessary to elicit  $P_0$  until a plateau was observed in the frequency- $dP/dt$  relationship (Buller & Lewis, 1965). A stimulus frequency of 350 Hz was usually required to elicit a maximum  $dP/dt_0$ .

Shortening velocities were measured during twelve different after-loaded isotonic contractions ( $P/P_0 = 0.05-0.50$ ). The same relative after-loads were used for each muscle tested. Shortening velocities were measured in order of increasing after-loads and were determined from the straight, maximum velocity segment of the displacement signals (Fig. 2). The stimulation frequency that resulted in the maximum  $dP/dt_0$  was used for measuring velocities and was held constant during the experiment. As reported by others (Jewell & Wilkie, 1958; Lannergren, 1978; Ranatunga, 1982) the velocity observed with an after-loaded isotonic contraction was not significantly different from the velocity after a quick release from  $P_0$  to a given after-load (D. R. Clafin & J. A. Faulkner, unpublished observation). Consequently, the conclusion of Jewell & Wilkie (1958) that 'this presumably indicates that sufficient time had elapsed before the load was lifted for the muscle to become fully active' is valid. The  $P_0$  was monitored throughout the procedure, and after-loads were calculated based on the most recently measured  $P_0$ . After determination of shortening velocities, a final measurement of  $P_0$  was made. The  $P_0$  measured after the determination of the force-velocity relationship averaged 102% of pre-test value for the soleus muscle and 87% for the EDL muscle.

Following the measurements of force and velocity, the tendons were removed from the muscle, the muscle was blotted, and immediately weighed. The muscle was pinned at  $L_0$  and viewed under a dissection microscope. With microdissection, five to ten small bundles of fibres were isolated from their origin on the proximal fascial sheath to their insertion on the distal fascial sheath. The lengths of the bundles of fibres were measured and a mean length for the bundles of fibres was obtained for each muscle.

The fibre lengths of soleus and EDL muscles from young, adult and aged mice were determined using the nitric acid digestion technique (Close, 1964; Maxwell, Faulkner & Hyatt, 1974). For these experiments, the muscle was removed and isometric contractile properties were measured as described previously. The muscle was pinned at  $L_0$  and the lengths of five to ten fibres were

measured under a dissecting microscope. Still pinned at  $L_0$ , the muscle was fixed overnight in 10% formaldehyde solution in 0.9% saline. After fixation, the muscle was immersed in 20%  $\text{HNO}_3$  for 8–12 h to dissolve the connective tissue. The acid was replaced by 50% glycerine solution. For each digested muscle, the resting length was measured and the muscle was separated into several bundles. Bundles were selected from the proximal, distal and middle portions of muscles. Single fibres were teased from the bundles, and the lengths of approximately five fibres from each bundle were measured at  $100\times$  magnification. A total of eighteen to twenty fibres from each muscle were measured, and the mean fibre length was determined. The fibre length/muscle length ratio was determined by dividing the mean fibre length by the resting length of the digested muscle.

The velocities of shortening were converted from mm/s to fibre lengths/s. For each muscle, the data on force ( $P/P_0$ ) and velocity ( $L_t/s$ ) were fitted by a rectangular hyperbola of the form  $(V+b)(P/P_0 \pm a/P_0) = b(1+a/P_0)$  where  $V$  is the velocity of shortening (Hill, 1938). The constants  $a/P_0$  and  $b$  were determined from an algorithm which minimized the sum of the squares of the differences between the resultant hyperbola and the measured velocities (Clafin & Faulkner, 1985).  $V_{\max}$  was calculated from  $b/(a/P_0)$ . For each of the two muscles and each of the three age groups, the mean  $\pm 1$  s.e.m. was calculated for the velocities at each of the twelve after-loads. The force-velocity relationship was then plotted for soleus and EDL muscles of young, adult and aged mice (Fig. 2).

The dry masses of four to six soleus and EDL muscles for each age group were determined. The muscles were dehydrated at 80 °C for approximately 30 min. After 30 min, the muscles were weighed at 5 min intervals until the masses stabilized. Dry mass/wet mass ratios were calculated. The mean cross-sectional area of each muscle was estimated by dividing muscle wet mass by the product of fibre length and 1.06 g/cm<sup>3</sup>, the density of mammalian skeletal muscle (Méndez & Keys, 1960).

For each variable, significant differences among groups were determined by a univariate one-way analysis of variance. If the  $F$  statistic of the analysis of variance was significant, differences between groups were assessed by a Scheffe comparison. A paired  $t$  test was used to determine the difference between the two techniques for measuring fibre lengths. The accepted level of significance for all statistical tests was set at  $P \leq 0.05$ . All data are reported as a mean  $\pm 1$  s.e.m.

## RESULTS

The body mass of the mice increased 25% from 3 to 10 months of age and decreased by 13% from 10 to 27 months of age (Table 1). Compared to soleus and EDL muscles from adult mice, significant decreases of 20 and 13% respectively were observed for the muscle wet masses of soleus and EDL muscles from aged mice (Table 1). The dry mass/wet mass ratio did not differ among the three age groups for either muscle nor was the ratio different between the muscles. The pooled dry mass/wet mass ratio was  $0.25 \pm 0.005$ .

As reported previously by Segal, White & Faulkner (1986), no significant difference was observed between the fibre lengths measured on bundles of fibres *in situ* and on single fibres after nitric acid digestion. Furthermore, the fibre lengths from soleus and EDL muscles did not change significantly with age (Table 2). The fibre length/muscle length ratio determined on intact soleus muscles was not different among the three age groups and the pooled mean equalled  $0.69 \pm 0.006$ . With the nitric acid digestion technique, in spite of no differences in the mean fibre length with age, the value of 0.68 for the fibre length/muscle length ratio of the soleus muscles of adult mice was significantly smaller than the values of 0.71 and 0.70 for young and aged mice, respectively. This difference of 3–4% is not likely to be of physiological significance. For EDL muscles, the fibre length/muscle length ratio did not change with age and the pooled ratio was  $0.45 \pm 0.004$ . Although muscle wet mass decreased with age, small differences in individual fibre lengths and muscle wet

TABLE 1. Data are given (mean  $\pm$  1 s.e.m.) for body mass, muscle wet mass, dry mass/wet mass ratio, fibre length and total cross-sectional area of muscle fibres, as estimated by dividing muscle mass by the product of fibre length and 1.06 g/cm<sup>3</sup>, the density of mammalian skeletal muscle, of soleus and extensor digitorum longus (EDL) muscles of young (2–3 months), adult (9–10 months), and aged (26–27 months) mice

	Soleus muscle			EDL muscle		
	Young (n = 11)	Adult (n = 14)	Aged (n = 14)	Young (n = 15)	Adult (n = 18)	Aged (n = 18)
Body mass (g)	27.0 $\pm$ 1.04*	35.4 $\pm$ 1.44	29.7 $\pm$ 1.09*	27.5 $\pm$ 0.75*	33.3 $\pm$ 1.02	30.0 $\pm$ 0.59*
Muscle wet mass (mg)	8.75 $\pm$ 0.61	10.3 $\pm$ 0.48	8.27 $\pm$ 0.50*	10.74 $\pm$ 0.22	10.66 $\pm$ 0.40	9.28 $\pm$ 0.37*†
Dry mass/wet mass	0.24 $\pm$ 0.003	0.25 $\pm$ 0.02	0.24 $\pm$ 0.01	0.26 $\pm$ 0.01	0.24 $\pm$ 0.01	0.25 $\pm$ 0.02
Fibre length (mm)	7.84 $\pm$ 0.22	8.15 $\pm$ 0.22	7.45 $\pm$ 0.17	5.44 $\pm$ 0.12	5.76 $\pm$ 0.15	5.37 $\pm$ 0.08
Fibre area (mm <sup>2</sup> )	1.05 $\pm$ 0.06	1.17 $\pm$ 0.03	1.03 $\pm$ 0.06	1.82 $\pm$ 0.06	1.74 $\pm$ 0.04	1.63 $\pm$ 0.08

\* Value for muscle of young or aged mice significantly different from value for adult mice ( $P \leq 0.05$ ).

† Value for muscle of aged mice significantly different from young mice ( $P \leq 0.05$ ).

TABLE 2. Fibre lengths and fibre length/muscle length ratios ( $L_i/L_o$ ) as measured from intact muscles (intact) and following nitric acid digestion (digested), are given for soleus and EDL muscles from young (2–3 months), adult (9–10 months) and aged (26–27 months) mice

	Soleus muscle			EDL muscle		
	Young (n = 3)	Adult (n = 6)	Aged (n = 5)	Young (n = 3)	Adult (n = 5)	Aged (n = 5)
$L_i$ intact (mm)	8.00 $\pm$ 0.15	7.37 $\pm$ 0.20	7.11 $\pm$ 0.21	5.46 $\pm$ 0.07	5.67 $\pm$ 0.07	5.70 $\pm$ 0.14
$L_i/L_o$ intact	0.71 $\pm$ 0.02	0.69 $\pm$ 0.01	0.69 $\pm$ 0.01	0.45 $\pm$ 0.003	0.44 $\pm$ 0.004	0.45 $\pm$ 0.01
$L_i$ digested (mm)	8.08 $\pm$ 0.27	7.28 $\pm$ 0.20	7.21 $\pm$ 0.21	5.52 $\pm$ 0.19	6.03 $\pm$ 0.21	5.61 $\pm$ 0.17
$L_i/L_o$ digested	0.71 $\pm$ 0.01*	0.68 $\pm$ 0.002	0.70 $\pm$ 0.003*	0.45 $\pm$ 0.01	0.47 $\pm$ 0.02	0.44 $\pm$ 0.004

\* Value for muscle of young or aged mice significantly different from value for adult mice ( $P \leq 0.05$ ).

masses resulted in no significant differences in the cross-sectional area of either muscle with ageing (Table 1).

Between 3 and 10 months of age, the soleus showed an increase in  $P_0$ , whereas the EDL muscle showed no change in  $P_0$  (Table 3). The  $P_0$  of both muscles decreased significantly with ageing, the soleus muscles to 78% of the adult value and that of the EDL muscles to 73%. The specific  $P_0$  of 21 N/cm<sup>2</sup> for soleus muscles did not differ among the three age groups. For EDL muscles, the specific  $P_0$  of 23 N/cm<sup>2</sup> did not change between 3 and 10 months of age, but that of the aged mice decreased significantly to 78% of the value for adult mice (Table 3).

For the soleus muscles, the contraction and half-relaxation times were prolonged significantly in adult compared to young and aged compared to young mice. The

TABLE 3. *In vitro* contractile properties at 25 °C are presented for young (2–3 months), adult (9–10 months) and aged (26–27 months) mice. Data (mean  $\pm$  1 s.e.m.) include: time to peak twitch tension (TPT), half-relaxation time ( $RT_{1/2}$ ), maximum isometric tetanic force ( $P_0$ ), maximum specific tetanic force (specific  $P_0$ ), and twitch tetanus ratio ( $P_t/P_0$ ), extrapolated maximum velocity of unloaded shortening ( $V_{max}$ ), and the parameter  $a/P_0$  that describes the force–velocity relationship

	Soleus muscle			EDL muscle		
	Young ( $n = 11$ )	Adult ( $n = 14$ )	Aged ( $n = 14$ )	Young ( $n = 15$ )	Adult ( $n = 18$ )	Aged ( $n = 18$ )
TPT(ms)	28 $\pm$ 1*	34 $\pm$ 1	39 $\pm$ 2†	16 $\pm$ 1	15 $\pm$ 1	16 $\pm$ 1
$RT_{1/2}$ (ms)	35 $\pm$ 2*	45 $\pm$ 2	50 $\pm$ 2†	18 $\pm$ 1	17 $\pm$ 1	19 $\pm$ 1
$P_0$ (mN)	213 $\pm$ 6*	259 $\pm$ 11	202 $\pm$ 9*	413 $\pm$ 11	411 $\pm$ 13	299 $\pm$ 14*†
Specific $P_0$ (N/cm <sup>2</sup> )	20.6 $\pm$ 0.67	22.1 $\pm$ 0.65	18.7 $\pm$ 1.05	23 $\pm$ 0.80	23.8 $\pm$ 0.67	18.6 $\pm$ 0.86*†
$P/P_0$	0.14 $\pm$ 0.01	0.15 $\pm$ 0.01	0.19 $\pm$ 0.02	0.23 $\pm$ 0.01	0.26 $\pm$ 0.01	0.28 $\pm$ 0.01†
$V_{max}$ (L <sub>t</sub> /s)	4.8 $\pm$ 0.19	4.2 $\pm$ 0.26	4.6 $\pm$ 0.18	10.4 $\pm$ 0.25	9.8 $\pm$ 0.2	10.0 $\pm$ 0.39
$a/P_0$	0.22 $\pm$ 0.02	0.20 $\pm$ 0.01	0.19 $\pm$ 0.02	0.34 $\pm$ 0.02	0.38 $\pm$ 0.01	0.34 $\pm$ 0.02

\* Value for muscle of young or aged mice significantly different from value for adult mice ( $P \leq 0.05$ ).

† Value for muscle of aged mice significantly different from young mice ( $P \leq 0.05$ ).

significant prolongations of the contraction and half-relaxation times of the soleus muscle with ageing were supported by significant increases in the tetanic force developed at submaximum frequencies of stimulation by soleus muscles of aged mice compared to the forces developed by soleus muscles of young and adult mice (Fig. 1). In contrast, the EDL muscles showed no significant differences in any of these variables with ageing (Table 3), except for one significant Scheffe comparison between young and aged mice in the frequency–force relationship at 50 Hz.

Neither the force–velocity relationships (Fig. 2) nor the  $V_{max}$  (Table 3) of soleus or EDL muscles showed any significant change with age. The lack of a significant change in the force–velocity relationships of soleus and EDL muscles with ageing was supported by no evidence of significant differences in the values for  $a/P_0$  (Table 3).

#### DISCUSSION

The conclusion that human beings lose a significant amount of muscle mass with ageing is based primarily on cross-sectional studies and indirect estimates of muscle mass, but the magnitude of the estimated loss, greater than one-third of the muscle mass, attests to the validity of the observation (Grimby & Saltin, 1983). For rats, decreases in mass of 13–26% have been reported for soleus muscles and 15–22% for EDL muscles (Gutmann & Carlson, 1976; Fitts *et al.* 1984; Larsson & Edström, 1986). The aged mice in our study had a decrease of 20% in the mass of the soleus muscle and of 13% in the mass of the EDL muscle. These decreases in muscle mass with ageing are similar to the values of 10 and 18% for soleus and EDL muscles, respectively, reported by White, Clarke & Kandarian (1986) for mice of the same strain. The lack of any change in the dry mass/wet mass ratio with ageing is consistent with a previous report on soleus and omohyoid muscles of rats (McCarter & McGee, 1987). The stable value of approximately 0.25 in both muscles at all ages

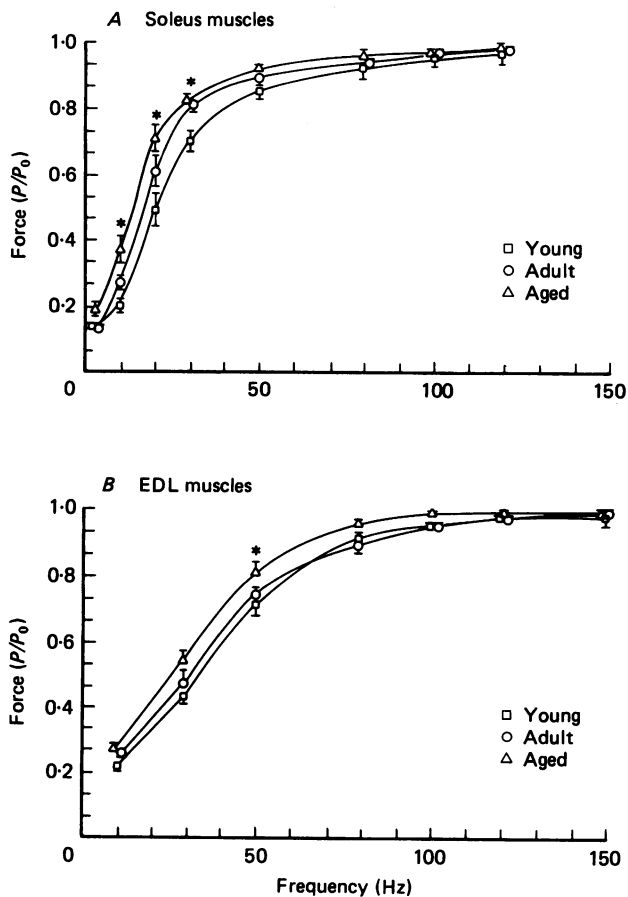


Fig. 1. Frequency-force relationships for soleus muscles (A) and extensor digitorum longus (EDL) muscles (B) from young, adult and aged mice. The values on the ordinate are normalized as the force at each frequency ( $P$ ) divided by the maximum isometric tetanic force ( $P_0$ ). Plots are for means  $\pm$  1 S.E.M. Error bars are omitted when they fall within the symbol. The asterisks indicate significant differences in the  $F$  ratio of the univariate one-way analysis of variance among the three groups and a significant Scheffe comparison between the muscles from the young and aged mice ( $P \leq 0.05$ ).

in spite of significant decreases in mass with age indicates that losses in protein and water were proportional.

For male Wistar rats 26 months of age, the EDL muscle developed a  $P_0$  24% less than that of EDL muscles from 5-month-old rats (Gutmann & Carlson, 1976). In a similar comparison of soleus muscles in 6-month- and 20- to 24-month-old rats, the deficit in  $P_0$  was 19% (Larsson & Edström, 1986). Fitts and his associates (1984) did not present data on the  $P_0$  of muscles in their study of ageing of Long-Evans rats, so values for  $P_0$  were calculated from their data on specific  $P_0$  using predetermined fibre length/muscle length ratios of 0.60 for soleus and 0.40 for EDL muscles (Segal & Faulkner, 1985). Compared to data for 9-month-old rats, the deficit in  $P_0$  for 28-month-old rats was 14% for soleus muscles and 21% for EDL muscles. In the

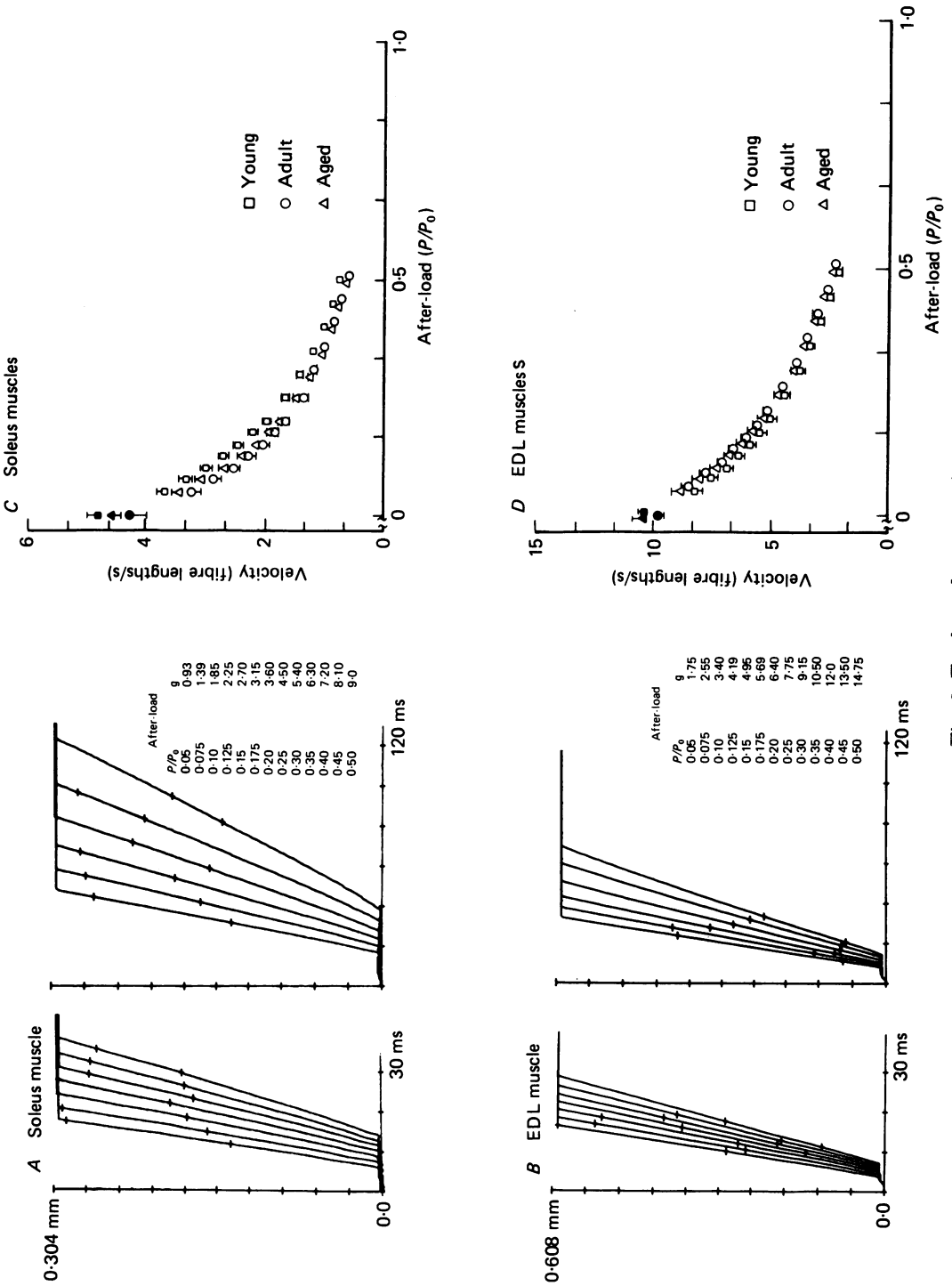


Fig. 2. For legend see opposite.



present study, soleus and EDL muscles from aged male mice showed a deficit in  $P_0$  of 22 and 27% respectively. Although the deficits for the  $P_0$  of soleus and EDL muscles of aged compared to young rodents are lower than the deficits of 30–40% reported for the maximum strength developed by aged compared to young men and women (Grimby & Saltin, 1983), these data support the hypothesis of a significant deficit in the development of maximum force by whole skeletal muscles of aged animals.

The omohyoideus of the rat, a parallel-fibred muscle with the fibres parallel to the long axis of the muscle (McCarter, Radicke & Yu, 1977), has a specific  $P_0$  of approximately 28 N/cm<sup>2</sup> (McCarter & McGee, 1987). In contrast, in the present study of mice the soleus muscle had a specific  $P_0$  of 22 N/cm<sup>2</sup> and the EDL muscle of 24 N/cm<sup>2</sup> with similar values reported previously for rats (Close, 1972). The hierarchy in the values for specific  $P_0$  from soleus to EDL to omohyoideus is explained in part by the angle of pennation of 5 deg for the soleus, 3.5 deg for the EDL muscles (Close, 1964), and 0 deg for the omohyoideus muscle (McCarter *et al.* 1977). Although Close (1964) calculated that the effect of the angles of pennation on the force developed by the soleus and EDL muscles was less than 0.1%, our data indicate the effect is between 15 and 20%. Since the fibre length/muscle length ratio does not change with ageing for either the soleus or the EDL muscles, the pennation remains constant and therefore does not contribute to the decrease in the development of force that occurs with ageing.

Ageing does not appear to influence the specific  $P_0$  of the anterior tibial, soleus, or omohyoideus muscles of rats (Fitts *et al.* 1984; Larsson & Edström, 1986; McCarter & McGee, 1987). In the present study, the specific  $P_0$  for soleus muscles of all ages and EDL muscles of young and adult mice were within the normal range for the specific  $P_0$  of young animals (Close, 1972). In contrast, the specific  $P_0$  of EDL muscles from aged mice was significantly lower than the values for young and adult mice. Fitts *et al.* (1984) reported no difference in the specific  $P_0$  of EDL muscles of rats, but an adequate description of the method for normalization of the  $P_0$  was not presented. The deficit in the specific  $P_0$  of 22% for the EDL muscles from aged male mice is in reasonable agreement with the 20% deficit observed in the maximum voluntary contraction/cross-sectional area of aged men (Young *et al.* 1985), although the mechanisms responsible are potentially quite different.

---

Fig. 2. A sample of the displacement records at different after-loads for one soleus muscle (A) and one extensor digitorum longus (EDL) muscle (B) from an aged mouse, and the mean force-velocity relationship for soleus ( $n = 11, 14, 14$ ) (C) and EDL ( $n = 15, 18, 18$ ) (D) muscles from young, adult and aged C57BL/6 male mice, respectively. On the abscissa of the displacement curves (panels A and B), two different time scales are used to accommodate the variations in velocity at different after-loads. The velocity for each after-load was measured between the plus signs on each displacement curve based on a computer program that selected the section of the displacement record with the highest velocity. Open symbols (panels C and D) are velocities measured at each of the twelve after-loads, and the maximum velocity of unloaded shortening ( $V_{\max}$ : closed symbols) is extrapolated from the hyperbola which best fits the data. No significant differences were observed for the overall relationship, for  $V_{\max}$ , or for  $a/P_0$  (see Table 3 for  $V_{\max}$  and  $a/P_0$ ). Plots are for mean  $\pm 1$  s.e.m. Error bars are omitted when they fall within a symbol.

Single skinned fibres from soleus and EDL skeletal muscles of aged rats do not show significant deficits in specific  $P_0$  (Eddinger *et al.* 1986). During measurements of fibre length after nitric acid digestion, most of the fibres in muscles of aged mice appeared to be of normal size, but a significant population of small, atrophic fibres was observed in each soleus and EDL muscle. White *et al.* (1986) have noted such a population of fibres in their frequency histogram for fibre cross-sectional areas of soleus and EDL muscles of mice. These observations suggest that the deficit of 22% observed for the specific  $P_0$  of EDL muscles of aged mice results from impairments in the force development of select fibres, but the possibility of differences between species and observed differences between soleus and EDL muscles raise issues that preclude a clear resolution of the problem.

Regardless of whether some or all of the fibres are involved, the significant decrease in the force-generating capacity per unit cross-sectional area of whole EDL muscles from aged male mice must result from a decrease in the number of cross-bridges per unit area, fewer cross-bridges in the driving stroke at any one time, or a decrease in the force developed by a cross-bridge during the driving stroke. Fragments of the  $S_1$  and  $S_2$  portions of the myosin molecule have been observed in rested muscles of young chickens (Ball, Krus & Alizadeh, 1987). An increased susceptibility to the disruption of cross-bridges in muscles of aged animals would reduce the total number of cross-bridges. A decrease in the number of cross-bridges in the driving stroke occurs in the presence of high concentrations of inorganic phosphate (Hibberd, Dantzig, Trentham & Goldman, 1985) or hydrogen ions (Metzger & Moss, 1987). Consequently, an increased concentration of the end-products of metabolism could reduce the force per  $\text{cm}^2$  developed by skeletal muscle fibres. A possible mechanism unsupported by evidence is a decrease in the force developed by a cross-bridge during the driving stroke. Conformational changes in proteins with ageing could impair movement of the cross-bridge during the driving stroke.

Our observation of no significant difference with age for the  $V_{\max}$  of either soleus or EDL muscles of male mice is consistent with previous data for whole muscles of rats (Fitts *et al.* 1984). The lack of any significant change in the whole force-velocity relationship of either soleus or EDL muscles, supports the contention of Grimby & Saltin (1983) that the force-velocity relationship of humans was not influenced by age. In a muscle composed of heterogeneous fibre types, the force-velocity relationship at low velocities is a function of all of the fibres, but at high velocities of shortening, velocities exceed the  $V_{\max}$  of the slower fibres and only the faster fibres contribute to the relationship (Claffin & Faulkner, 1985). The stability of the total force-velocity curve suggests that no significant changes occur in the distribution of the fibre types in either soleus or EDL muscles.

In the present study, the prolongation of the time characteristics of isometric twitches of soleus muscles appeared to be of physiological significance. Fitts *et al.* (1984) and Larsson & Edström (1986) also report significant increases in the contraction and relaxation times of soleus muscles of aged compared to adult rats. The changes observed with ageing in the soleus muscle are consistent with a decrease in the rate of uptake of calcium by the sarcoplasmic reticulum, whereas the lack of change with age for the overall force-velocity relationship and  $V_{\max}$ , indicate no

significant modifications in myosin ATPase activity (Barany, 1967), or in the light or heavy chains of myosin (Hoh, McGrath & White, 1976; Reiser, Moss, Giulian & Greaser, 1985; Sweeney, Kushmerick, Mabuchi, Gergely & Stréter, 1986). Close (1965, 1972) considered the product of  $V_{\max}$  and contraction time to be a constant for slow and fast muscles from a wide variety of species and the correlation coefficient for the means of these two variables for eight mammalian species (Close, 1965) was 0.99. For soleus muscles from young and adult mice, the correlation coefficient between  $V_{\max}$  and  $1/\text{contraction time}$  is 0.76. The significant change in the contraction time of the soleus muscle without a change in the force-velocity relationship results in a decrease in the correlation to 0.17 which is not significantly different from zero. Apparently, with ageing the biochemical events underlying  $V_{\max}$  and contraction time vary independently of one another.

Eddinger and his associates (1986) conclude that the decrease in strength observed in old age does not reside in changes intrinsic to skeletal muscles. Similarly, McCarter & McGee (1987) report that their major finding was the 'age-related decline in motor performance is most likely unrelated to deficits in the functional properties of fibers'. In contrast, for whole skeletal muscles from mice studied *in vitro*, we found significant decreases with ageing in skeletal muscle mass,  $P_0$ , and for the EDL muscle in the specific  $P_0$ . A species difference is not a likely explanation of the differences since muscles of rats and mice show the same magnitude of deficit in mass and  $P_0$ . The studies of bundles of fibres (Eddinger *et al.* 1986; McCarter & McGee, 1987) and single skinned fibres (Eddinger *et al.* 1986) show considerable variability in the values for specific  $P_0$  within and between groups which precludes the recognition of any age-associated change. Under these circumstances, comparisons between values for specific  $P_0$  of whole muscles with those of bundles and single fibres are difficult. The measurements of time characteristics and force-velocity relationships of bundles and single skinned fibres show little change with age and our observations are consistent with these data.

The authors thank James Kneebone for his assistance in data collection. This research was supported by a grant from the National Institute on Aging, AG 06157.

#### REFERENCES

- BALL, R. D., KRUSS, D. L. & ALIZADEH, B. (1987). Myosin degradation fragments in skeletal muscle. *Journal of Molecular Biology* **193**, 47-56.
- BARANY, M. (1967). ATPase activity of myosin correlated with speed of muscle shortening. *Journal of General Physiology* **50**, 197-216.
- BROOKS, S. V. & FAULKNER, J. A. (1987). Contractile properties of skeletal muscles from young, adult, and aged mice. *Biophysical Journal* **51**, 222a.
- BULLER, A. J. & LEWIS, D. M. (1965). The rate of tension development in isometric tetanic contractions of mammalian fast and slow skeletal muscle. *Journal of Physiology* **176**, 337-354.
- CLAFLIN, D. R. & FAULKNER, J. A. (1985). Shortening velocity extrapolated to zero load and unloaded shortening velocity of whole rat skeletal muscle. *Journal of Physiology* **359**, 357-363.
- CLOSE, R. (1964). Dynamic properties of fast and slow skeletal muscles of the rat during development. *Journal of Physiology* **173**, 74-95.
- CLOSE, R. (1965). The relation between intrinsic speed of shortening and duration of the active state of muscle. *Journal of Physiology* **180**, 542-559.

- CLOSE, R. I. (1972). Dynamic properties of mammalian skeletal muscles *Physiological Reviews* **52**, 129–197.
- EDDINGER, T. J., CASSENS, R. G. & MOSS, R. L. (1986). Mechanical and histochemical characterization of skeletal muscles from senescent rats. *American Journal of Physiology* **251**, C421–430.
- FITTS, R. H., TROUP, J. P., WITZMANN, F. A. & HOLLOSZY, J. O. (1984). The effect of ageing and exercise on skeletal muscle function. *Mechanisms of Ageing and Development* **27**, 161–172.
- GRIMBY, G. & SALTIN, B. (1983). The aging muscle. *Clinical Physiology* **3**, 209–218.
- GUTMANN, E. & CARLSON, B. M. (1976). Regeneration and transplantation of muscles in old rats and between young and old rats. *Life Sciences* **18**, 109–114.
- HIBBERD, M. G., DANTZIG, J. A., TRENTHAM, D. R. & GOLDMAN, Y. E. (1985). Phosphate release and force generation in skeletal muscle fibres. *Science* **228**, 1317–1319.
- HILL, A. V. (1938). The heat of shortening and the dynamic constants of muscle. *Proceedings of the Royal Society B* **126**, 136–195.
- HOH, J. F. H., MCGRATH, P. A. & WHITE, R. I. (1976). Electrophoretic analysis of multiple forms of myosin in fast-twitch and slow-twitch muscles of the chick. *Biochemical Journal* **157**, 87–95.
- JEWELL, B. R. & WILKIE, D. R. (1958). An analysis of the mechanical components in frog's striated muscle. *Journal of Physiology* **143**, 515–540.
- LANNERGREN, J. (1978). The force-velocity relation of isolated twitch and slow muscle fibres of *Xenopus laevis*. *Journal of Physiology* **283**, 501–521.
- LARSSON, L. & EDSTRÖM, L. (1986). Effects of age on enzyme-histochemical fibre spectra and contractile properties of fast- and slow-twitch skeletal muscles in the rat. *Journal of the Neurological Sciences* **76**, 69–89.
- LARSSON, L., GRIMBY, G. & KARLSSON, J. (1979). Muscle strength and speed of movement in relation to age and muscle morphology. *Journal of Applied Physiology* **46**, 451–456.
- MCCARTER, R. & MCGEE, J. (1987). Influence of nutrition and ageing on the composition and function of rat skeletal muscle. *Journal of Gerontology* **42**, 432–441.
- MCCARTER, R., RADICKE, D. & YU, B. P. (1977). A model preparation for studying fast mammalian skeletal muscles. *Proceedings of the Society for Experimental Biology and Medicine* **156**, 40–45.
- MAXWELL, L. C., FAULKNER, J. A. & HYATT, G. J. (1974). Estimation of the number of fibers in guinea pig skeletal muscle. *Journal of Applied Physiology* **37**, 259–264.
- MENDEZ, J. & KEYS, A. (1960). Density and composition of mammalian muscle. *Metabolism* **9**, 184–188.
- METZGER, J. M. & MOSS, R. L. (1987). Greater hydrogen ion-induced depression of tension and velocity in skinned single fibres of rat fast than slow muscles. *Journal of Physiology* **393**, 727–742.
- QUETELET, A. (1835). *Sur l'homme et le développement de ses facultés*, vol. 2. Paris: Bachelier, Imprimeur-Libraire.
- RANATUNGA, K. W. (1982). Temperature-dependence of shortening velocity and rate of isometric tension development in rat skeletal muscle. *Journal of Physiology* **329**, 465–483.
- REISSER, P. J., MOSS, R. L., GIULIAN, G. G. & GREASER, M. L. (1985). Shortening velocity and myosin heavy chains of developing rabbit muscle fibers. *Journal of Biological Chemistry* **27**, 14403–14405.
- SEGAL, S. S. & FAULKNER, J. A. (1985). Temperature-dependent physiological stability of rat skeletal muscle *in vitro*. *American Journal of Physiology* **248**, C265–270.
- SEGAL, S. S., WHITE, T. P. & FAULKNER, J. A. (1986). Architecture, composition and contractile properties of rat soleus muscle grafts. *American Journal of Physiology* **250**, C474–479.
- SWEENEY, H. L., KUSHMERICK, M. J., MABUCHI, K., GERGELY, J. & STRÉTER, F. A. (1986). Velocity of shortening and myosin isozymes in two types of rabbit fast-twitch muscle fibers. *American Journal of Physiology* **251**, C431–434.
- WHITE, T. P., CLARK, K. I. & KANDARIAN, S. C. (1986). Mass, fiber number, and cross-sectional area of hindlimb skeletal muscles from C57BL/6 mice at 12 and 28 months of age. *Physiologist* **29**, 151.
- YOUNG, A., STOKES, M. & CROWE, M. (1984). Size and strength of the quadriceps muscle of old and young women. *European Journal of Clinical Investigation* **14**, 282–287.
- YOUNG, A., STOKES, M. & CROWE, M. (1985). The size and strength of the quadriceps muscle of old and young men. *Clinical Physiology* **5**, 145–154.