

Supplementary information

Supplementary Methods

Isolation of conditional lethal mutants with polarity defects

Mutants that showed an altered morphology were selected visually by fluorescence microscopy after staining with Calcofluor white, as described previously (Hirata *et al.*, 1998; Radcliffe *et al.*, 1998). The isolated round mutants were further examined for sensitivity to the protein kinase inhibitor staurosporine (1.5 $\mu\text{g/ml}$). The temperature-sensitive (ts) *pmo25* mutant was created by error-prone PCR mutagenesis as follows:

The DNA fragments containing the *pmo25⁺:GFP:kan^r* gene amplified by PCR from the *pmo25⁺:GFP:kan^r* strain were integrated into the *pmo25⁺* locus in wild-type cells; and from the resultant *kan^r* cells, the ts *pmo25-35* mutant was isolated at 36°C.

Immunochemical and kinase assays

Preparation of cell extracts, immunoprecipitation, immunodetection, and kinase assays were performed as previously described (Matsusaka *et al.*, 1995; Bähler and Nurse, 2001; Huang *et al.*, 2003; Wiley *et al.*, 2003). Immunoprecipitation was done by using anti-HA antibody (HA.11, BabCO), anti-GFP antibody (8362-1, Clontech), anti-Myc antibody (9E10, Calbiochem), and magnetizable beads conjugated to protein A or G (Dynabeads, DYNAL).

Microscopy techniques

For the observation of Pmo25-GFP, the cells expressing Pmo25-GFP were fixed with methanol (-20°C) for 10 min and washed three times with PEM buffer (Alfa *et al.* 1993). Fixed-cell images were collected with an Axiophot 2 MOT (ZEISS), the ApoTome sectioning system, and AxioCam MRm CCD camera; and the images were further processed with AxioVision software. For time-lapse microscopy, a 35 mm glass-bottomed culture dish (MatTek Corporation, P35G-1.5-10-C) was coated with 100 µg/ml concanavalin A. The culture of logarithmically growing cells (50 µl) was deposited in the well for a couple of minutes and then removed. The dish was filled with 3 ml of EMM medium and the cells that were attached to the bottom of the well were subjected to microscopic analysis. Live-cell images were collected with an IX70 (OLYMPUS) and DeltaVision sectioning system. Cytological techniques were performed according to Matsusaka *et al.* (1995).

In vitro binding assay

The expression plasmids for HA-Pmo25 or Myc-Nak1 proteins were generated as follow: An HA-tagged *pmo25* cDNA or Myc-tagged *nak1* cDNA was amplified from pACT2-HA-Pmo25 or pGBKT7-Myc-Nak1, respectively, by PCR. The PCR products were inserted into the *NotI* and *XhoI* sites of the plasmid pTWIN1. The HA-Pmo25 and Myc-Nak1 proteins were synthesized using IMPACT-TWIN System (New England Biolabs). The HA-Pmo25 protein (10 µg/ml) was incubated with Myc-Nak1 protein (10

$\mu\text{g/ml}$) in Buffer A (Boudeau *et al.*, 2004), which contained 50 mM Tris-HCl (pH7.5), 0.27 M Sucrose, 0.1 mM EDTA (pH8.0), 0.1%(v/v) mercaptoethanol and protease inhibitors, for 1 h at 4°C. The mixture was incubated in 1:1000 dilution of a mouse anti-HA antibody (HA.11, Babco) and magnetizable beads conjugated to protein A (Dynabeads, DYNAL) for 1.5 h at 4°C. The beads were washed three times with Buffer A, and the bound proteins were subjected to an immunoblot analysis.

Supplementary References

Alfa, C., Fantes, P., Hyams, J., McLeod, M. and Warbrick, E. (1993) *Experiments with Fission Yeast: A Laboratory Course Manual*. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY.

Bähler, J. and Nurse, P. (2001) Fission yeast Pom1p kinase activity is cell cycle regulated and essential for cellular symmetry during growth and division. *EMBO J.* **20**, 1064-1073.

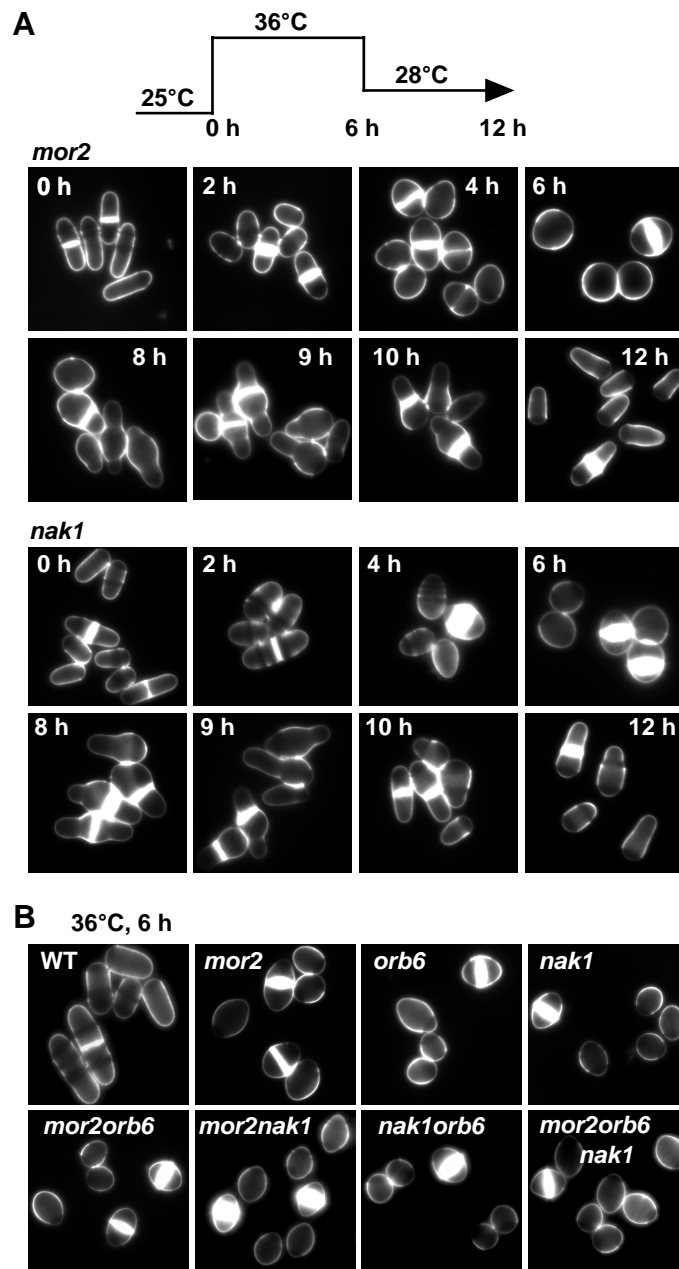
Matsusaka, T., Hirata, D., Yanagida, M. and Toda, T. (1995) A novel protein kinase *ssp1⁺* is required for alteration of growth polarity and actin localization in fission yeast. *EMBO J.*, **14**, 3325-3338.

Supplementary Table 1. Yeast strains used in this study

Strain	Genotype	Source
L972	<i>leu1 h⁻</i>	Lab Stock
KSP4-25	<i>pmo25::ura4⁺/pmo25⁺ leu1/leu1 ura4/ura4</i> <i>ade6-M216/ade6-M210 h⁻/h⁺ pREP41-Pmo25</i>	This study
KSP1-1	<i>pmo25⁺:GFP:kan^r h⁻</i>	This study
VS2933	<i>cdc7⁺:GFP:kan^r leu1 ura4 ade6 h⁻</i>	V. Simanis
MS192	<i>sad1⁺:dsRED:LEU2 leu1 h⁻</i>	O. Niwa
DH797-3B	<i>pmo25⁺:GFP:kan^r sad1⁺:dsRED:LEU2 leu1 h⁻</i>	This study
DH820-2D	<i>pmo25⁺:GFP:kan^r cdc7⁺:GFP:kan^r ura4 ade6 h⁻</i>	This study
DH614-6A	<i>nak1⁺:HA:kan^r leu1 h⁻</i>	This study
DH841-1D	<i>pmo25⁺:GFP:kan^r nak1⁺:HA:kan^r leu1</i>	This study
YS78-1	<i>mor2⁺:HA:kan^r h⁻</i>	Hirata <i>et al.</i> 2002
KK6-1	<i>orb6⁺:HA:kan^r h⁻</i>	This study
DH445-10D	<i>mor2⁺:HA:kan^r orb6⁺:HA:kan^r</i>	This study
DH107-4C	<i>mor2-786 leu1 h⁻</i>	Hirata <i>et al.</i> 2002
DH630-1	<i>pmo25-35:GFP:kan^r h⁻</i>	This study
DH842-1D	<i>pmo25-35:ura4⁺ ura4 h⁻</i>	This study
KP1-6D	<i>nak1-125/mor4-125 leu1 h⁻</i>	This study
Cdc10	<i>cdc10-129 h⁻</i>	P. Nurse
Cdc25	<i>cdc25-22 h⁺</i>	P. Nurse
DH92-2B	<i>cdc10-129 mor2-786 h⁻</i>	This study
DH93-2A	<i>cdc25-22 mor2-786</i>	This study
DH907-1C	<i>cdc10-129 pmo25-35:ura4⁺ ura4 h⁻</i>	This study
DH908-3A	<i>cdc25-22 pmo25-35:ura4⁺ ura4 h⁻</i>	This study
DH947-2A	<i>cdc10-129 nak1-125 leu1 his2 h⁺</i>	This study
DH910-2B	<i>cdc25-22 nak1-125 leu1 his2 h⁺</i>	This study

Orb6	<i>orb6-25 leu1 ade6 h⁻</i>	Verde <i>et al.</i> 1995
DH433-12C	<i>orb6-25 leu1 h⁻</i>	This study
YS85	<i>kan^r:nmt1:GFP:mor2⁺ h⁻</i>	Hirata <i>et al.</i> 2002
DH839-2	<i>kan^r:nmt1:GFP:mor2⁺ pmo25-23:ura4⁺ ura4</i>	This study
DH807-1A	<i>kan^r:nmt1:GFP:mor2⁺ nak1-125 leu1 h⁻</i>	This study
DH475-1C	<i>kan^r:nmt1:GFP:mor2⁺ orb6-23 leu1 h⁻</i>	This study
DH847-3C	<i>kan^r:nmt1:GFP:mor2⁺ cdc7-24 h⁻</i>	This study
DH789-1B	<i>pmo25⁺:GFP:kan^r mor2-786 h⁻</i>	This study
DH787-4A	<i>pmo25⁺:GFP:kan^r nak1-125 leu1 his2 h⁺</i>	This study
DH783-2C	<i>pmo25⁺:GFP:kan^r orb6-25 h⁻</i>	This study
DH852-7A	<i>nak1⁺:HA:kan^r pmo25-35:GFP:kan^r leu1 ura4 ade6</i>	This study
DH853-6D	<i>nak1⁺:HA:kan^r mor2-786 leu1 ade6</i>	This study
DH850-3D	<i>nak1⁺:HA:kan^r orb6-25 leu1</i>	This study
YDM975	<i>mob2⁺:Myc:kan^r leu1 ura4 ade6 h⁻</i>	D. McCollum
FV521	<i>mob2⁺:Myc:kan^r orb6⁺:HA:sup3-5 leu1 ura4 ade6-704</i>	Wiley <i>et al.</i> 2003
DH957-8A	<i>mob2⁺:Myc:kan^r orb6⁺:HA:sup3-5 pmo25-35:ura4⁺ ura4</i>	This study
DH990-2C	<i>mob2⁺:Myc:kan^r orb6⁺:HA:sup3-5 nak1-125 leu1 ura4 ade6-704</i>	This study
DH989-2B	<i>mob2⁺:Myc:kan^r orb6⁺:HA:sup3-5 mor2-786 leu1 ade6-704</i>	This study
KL284	<i>orb3⁺/nak1⁺:GFP:kan^r leu1 ura4 ade6 h⁻</i>	Leonhard and Nurse 2005
DH866-4C	<i>nak1⁺:GFP:kan^r leu1 h⁻</i>	This study
DH867-2A	<i>nak1⁺:GFP:kan^r pmo25-35:ura4⁺ leu1 ura4 his2 h⁺</i>	This study
DH967-1A	<i>pmo25⁺:mRFP:kan^r nak1⁺:GFP:kan^r leu1 ura4 his2 h⁺</i>	This study
DH821-1A	<i>pmo25-35:ura4⁺ cdc7⁺:GFP:kan^r leu1 ura4 h⁻</i>	This study
IH1297	<i>cdc16-116 ura4 h⁻</i>	I. Hagan
Cdc7	<i>cdc7-24 h⁺</i>	P. Nurse

KP553	<i>cdc7-i10 leu1 h⁻</i>	T. Kuno
IH1469	<i>sid1-239 leu1 ura4 ade6 h⁺</i>	I. Hagan
IH1470	<i>sid2-250 leu1 ura4 ade6 h⁺</i>	I. Hagan
DH888-6C	<i>pmo25⁺:GFP:kan^r cdc16-116 h⁻</i>	This study
DH798-6C	<i>pmo25⁺:GFP:kan^r cdc7-24 h⁻</i>	This study
DH1052-1A	<i>pmo25⁺:GFP:kan^r cdc7-i10 leu1</i>	This study
DH890-1B	<i>pmo25⁺:GFP:kan^r sid1-239</i>	This study
DH891-2B	<i>pmo25⁺:GFP:kan^r sid2-250 leu1</i>	This study
Cps1	<i>cps1-12 h⁺</i>	J. Ishiguro
DH1085-4B	<i>pmo25⁺:GFP:kan^r cps1</i>	This study
DH851-1A	<i>nak1⁺:HA:kan^r cdc7-24 leu1</i>	This study
DH922-2C	<i>nak1⁺:HA:kan^r sid1-239 leu1 ade6</i>	This study
DH923-2D	<i>nak1⁺:HA:kan^r sid2-250 leu1</i>	This study
KS1158	<i>mto1/mod20::kan^r leu1 ura4 ade6 h⁻</i>	K. Sawin
KGY3251	<i>mto1/mbol::ura4⁺ leu1 ura4 ade6 h⁻</i>	K. Gould
DH936-4A	<i>mob2⁺:Myc:kan^r leu1</i>	This study
DH948-8D	<i>mob2⁺:Myc:kan^r cdc7-24 leu1 ura4 h⁻</i>	This study
DH949-3A	<i>mob2⁺:Myc:kan^r sid1-239 leu1 ura4 ade6</i>	This study
DH939-2B	<i>mob2⁺:Myc:kan^r sid2-250 leu1 ura4 ade6</i>	This study
DH1080-2C	<i>mob2⁺:Myc:kan^r Δmto1::ura4⁺ leu1 ura4 ade6</i>	This study



Supplementary
Figure 1