Mutations in the *sup-38* Gene of *Caenorhabditis elegans* Suppress Muscle-Attachment Defects in *unc-52* Mutants

Erin J. Gilchrist and Donald G. Moerman

Department of Zoology, University of British Columbia, Vancouver, British Columbia, Canada V6T 1Z4 Manuscript received April 10, 1992 Accepted for publication July 2, 1992

ABSTRACT

Mutations in the *unc-52* locus of *Caenorhabditis elegans* have been classified into three different groups based on their complex pattern of complementation. These mutations result in progressive paralysis (class 1 mutations) or in lethality (class 2 and 3 mutations). The paralysis exhibited by animals carrying class 1 mutations is caused by disruption of the myofilaments at their points of attachment to the cell membrane in the body wall muscle cells. We have determined that mutations of this class also have an effect on the somatic gonad, and this may be due to a similar disruption in the myoepithelial sheath cells of the uterus, or in the uterine muscle cells. Mutations that suppress the body wall muscle defects of the class 1 *unc-52* mutations have been isolated, and they define a new locus, *sup-38*. Only the muscle disorganization of the Unc-52 mutants is suppressed; the gonad abnormalities are not, and the suppressors do not rescue the lethal phenotype of the class 2 and class 3 mutations. The suppressor mutations on their own exhibit a variable degree of gonad and muscle disorganization. Putative null *sup-38* mutations cause maternal-effect lethality which is rescued by a wild-type copy of the locus in the zygote. These loss-of-function mutations have no effect on the body wall muscle structure.

CUPPRESSION analysis has proven very useful in D bacteria and viruses for examining gene interactions in vivo (HARTMAN and ROTH 1973). Suppressor mutations have proved more difficult to isolate in multicellular organisms largely because of the lack of simple methods for examining sufficient numbers of animals. However, this powerful type of analysis has been possible with the nematode Caenorhabditis elegans. Large numbers of these worms can be grown in a very short period of time, and many specific mutations have already been described which permit easy detection of revertant phenotypes. Through this type of analysis several different mechanisms of suppression have been observed in C. elegans including intragenic rearrangement (WATERSTON, HIRSH and LANE 1984; ANDERSON and BRENNER 1984), gene duplication (RIDDLE and BRENNER 1978; MARUYAMA, MILLER and BRENNER 1989), epistasis (MOERMAN et al. 1982; GREENWALD and HORVITZ 1982) and informational suppression (WATERSTON and BRENNER 1978; WATERSTON 1981; HODGKIN, KONDO and WA-TERSTON 1987; HODGKIN et al. 1989). We have used suppression analysis here to identify a new gene involved in maintaining the attachment of myofilaments to the cell membrane and the extracellular matrix of the body wall muscle cells.

In C. elegans, as in other nematodes, the muscles are obliquely striated as a result of the positioning of the dense bodies relative to one another along the length of the muscle cells (ROSENBLUTH, 1965; FRAN-CIS and WATERSTON 1985; WATERSTON 1988). Dense

Genetics 132: 431-442 (October, 1992)

bodies perform a function analogous to the Z-line in vertebrate muscle and to the dense plaques or focal adhesions in many types of cultured cells. Like the Zline, they are responsible for anchoring the muscle cell thin filaments to the cell membrane and underlying extracellular matrix, and for maintaining the contractile units in register. The nematode dense body is also required to transmit force from the contractile apparatus to the hypodermis and outer cuticle, and in this function resembles the focal adhesions of cultured cells.

One class of mutation in the unc-52 gene of C. elegans leads to disruption of the muscle cell dense bodies (see WATERSTON 1988). Animals homozygous for these class 1 mutations develop normally as young larvae, but adults are thin, paralyzed, and partially egg-laying defective (BRENNER 1974). The paralysis is correlated with a gradual disruption of the muscle cell structure which begins at the third or fourth larval stage, and affects only the body wall muscle cells posterior to the pharynx (MACKENZIE et al. 1978; WATERSTON, THOMSON and BRENNER 1980). Electron microscopy has revealed that the muscle cell dense bodies are fractured, so that the apical portion of the structure is no longer associated with the cell membrane (reviewed in WATERSTON 1988). The phenotype of these mutants indicates that the unc-52 gene product is required for maintaining normal muscle structure.

Other classes of *unc-52* mutations result in embryonic lethality (B. WILLIAMS, personal communication; K. KONDO and I. KATSURA, personal communication), suggesting that the gene also plays an essential role in early development. In class 2 and class 3 mutant embryos, elongation arrests at the two-fold stage and body wall muscle is severely disorganized. This phenotype is similar to that produced by mutations in the muscle-affecting genes myo-3 (WATERSTON 1989), unc-45 (VENOLIA and WATERSTON 1990), deb-1 (BARSTEAD and WATERSTON 1991) and the *pat* genes (B. WIL-LIAMS and R. H. WATERSTON, personal communication).

The sequence of the *unc-52* gene has been determined (T. ROGALSKI and D. MOERMAN, unpublished results), and it resembles that of the mouse basement membrane heparan sulfate proteoglycan, perlecan (NOONAN *et al.* 1991). *In vitro*, proteoglycans have been shown to stabilize focal adhesions in cell cultures (reviewed in BURRIDGE *et al.* 1988). The sequence of *unc-52*, and its mutant phenotypes, suggest that this gene encodes a proteoglycan-like component which is important *in vivo* for stabilizing the interactions between muscle cells and the extracellular matrix.

In this paper we describe the isolation and characterization of intergenic suppressors of *unc-52*. All of these new mutations map to a single genetic locus, *sup-38*. Hermaphrodites homozygous for suppressor alleles of *sup-38* have defects in both muscle and gonad which are independent of the *unc-52* mutations. Putative null alleles of the *sup-38* locus have been isolated by selecting for loss of suppressor activity, and the terminal phenotype of animals homozygous for these mutations is maternal-effect lethal. Mutants homozygous for these *sup-38* loss-of-function alleles have wildtype muscle structure, suggesting that *sup-38* is not essential for normal muscle development.

MATERIALS AND METHODS

Nematode strains and culture conditions: Nematodes were grown and maintained on NGM plates, streaked with *Escherichia coli* strain OP50, according to the standard techniques outlined by BRENNER (1974). Some strains used in this work were obtained from D. L. BAILLIE (Simon Fraser University, Burnaby, British Columbia), J. HODGKIN (MRC, Laboratory of Molecular Biology, Cambridge, United Kingdom), I. KATSURA (University of Tokyo, Tokyo, Japan), A. M. ROSE (University of British Columbia, Vancouver), and B. WILLIAMS and R. H. WATERSTON (Washington University, St. Louis, Missouri). Genetic nomenclature follows the recommendations of HORVITZ et al. (1979).

The class 1 unc-52 alleles used in this study, e444, e998, e669am, st196::Tc1, e1012, e1421 and e699su250ts are maintained as homozygous strains. The lethal, class 2 allele, st549, is maintained in a strain carrying a duplication for the unc-52 region of linkage group (LG) II (st549/st549/mnDp34, B. WILLIAMS and R. H. WATERSTON, personal communication). The class 3 allele, ut111, is maintained in a heteroallelic strain which also carries the class 1 allele e444 (K. KONDO and I. KATSURA, personal communication). The following chromosomal rearrangements were also used: sDf21, sDf23, nDf27, nT1(IV, V) (FERGUSON and HORVITZ

1985), and *nT1*[(*m435*)*IV*, *V*] (ROGALSKI and RIDDLE, 1988).

Microscopy: Living worms were examined using both polarized light microscopy and Nomarski differential interference contrast optics. Worms were removed from NGM plates, placed in a drop of M9 buffer on a slide, and then immobilized by gently placing a coverslip over them. Pictures were taken on KODAK TMAX400 film using a Zeiss Axiophot Photomicroscope (Carl Zeiss D-7082 Oberkochen).

Characterization of unc-52 alleles: Heteroallelic class 1/ class 1 unc-52 animals (e444/e1012, e444/e1421, e444/ e669su250, e669/e1012, e669/e1421, e669/e669su250, e998/ e1012, e998/e1421, e998/e669su250) were constructed by crossing heterozygous males with hermaphrodites homozygous for another class 1 allele. Reciprocal crosses were done for all strains involving e1421. Heteroallelic class 1/class 2 unc-52 animals (st549/e998, st549/e444, st549/e669, st549/ st196, st549/e1012 and st549/e1421) were constructed by: (1) mating heterozygous class 1 males with st549/st549/ mnDp34 hermaphrodites and (2) mating heterozygous st549/+ males with homozygous class 1 hermaphrodites. Reciprocal crosses were done for all combinations. Class 1/ class 3 complementation tests were originally done by K. KONDO and I. KATSURA at the University of Tokyo, Tokyo, Japan. In our laboratory, class 1 Unc-52 mutants were mated to males heterozygous for the ut111 allele, and well moving F_1 progeny were selected and scored for the presence of class 3 lethals in the F_2 generation. The ut111 allele was tested with e998, e444, e669, st196, e1012, e1421 and e669su250. Class 2/class 3 complementation tests were done by mating ut111/+ males to st549/st549/mnDp34 hermaphrodites and screening for the presence of the ut111 phenotype in the F_1 generation.

Brood sizes for homozygous and heteroallelic *unc-52* animals were determined by counting the total number of progeny from 30 hermaphrodites, and calculating the mean and the 95% confidence intervals for each strain.

Isolation of revertants: Mutagenesis was done using 0.05 or 0.025 M ethyl methanesulfonate (EMS) in M9 buffer as described by SULSTON and HODGKIN (1988). Dominant or semidominant revertants were obtained by screening the F1 progeny of mutagenized Unc-52 hermaphrodites for animals that moved as adults. Briefly, worms were washed off 10 small (60 mm) plates into M9 buffer containing EMS, and gently agitated for 4-5 hr at room temperature. Then 20-30 worms per plate were transferred onto approximately 100 large (100 mm) NGM plates seeded with E. coli, and allowed to grow for 3-6 days before screening. In most cases, screening continued until the worms were near starvation. To reduce the probability of isolating tRNA amber suppressors when isolating revertants of e669, a strain was constructed which also carried an amber mutation in the dpy-20(IV) gene. The unc-52(e669am); dpy-20(e2017am) double mutants were mutagenized, and their F_1 progeny screened for animals that were able to move but were still Dpy in phenotype. Intergenic suppressors were maintained as homozygous unc-52; sup-38, unc-52; dpy-20sup-38, or sup-38 strains after outcrossing.

Mapping of suppressors: Intergenic suppressor loci were mapped using the following dpy markers: dpy-5(e61)I, dpy-10(e128)II, dpy-18(e364)III, dpy-13(e184)IV, dpy-4(e1166)IV, dpy-11(e224)V, and dpy-6(e14)X. Males of genotype unc-52/+;dpy/+ were mated to hermaphrodites from an unc-52;sup-38 strain and unc-52/unc-52;sup-38/+;dpy/+ F₁ progeny selected. The presence of $\frac{1}{16}$ DpyUncs in the F₂ generation indicated that the suppressor was not on the same LG as the dpy marker. A frequency of greater than $\frac{1}{16}$ DpyUncs indicated linkage of the suppressor locus to the dpy marker. To obtain sup-38 dpy-4 strains, DpySup segregants were selected from mapping experiments using dpy-4 with the suppressor alleles ra5, ra14, ra18, ra20 and ra21. These Dpy hermaphrodites were outcrossed to determine whether they still carried unc-52, and those which did were maintained as unc-52; sup-38 dpy-4 strains. Strains homozygous for the suppressor locus on its own were constructed for ra5, ra14, ra18, ra20 and ra21. This was done by crossing dpy-4/+ males with unc-52; sup-38 hermaphrodites, and selecting non-Dpy, well moving F2 progeny from semi-Dpy F1 hermaphrodites. The strains were then outcrossed to ensure that the unc-52 locus was no longer present, and heterozygous males were crossed to unc-52 hermaphrodites to ensure that the strain still carried the suppressor mutation. The presence of 1/16 instead of 1/4 Uncs in the F2 generation of this last cross indicated that the suppressor was present.

Three factor mapping of the suppressor alleles ra20 and ra21 was done by mating heterozygous unc-26(e345) dpy-4(e1166)/++ males with unc-52(e1421); sup-38 worms to obtain unc-52/+;unc-26 + dpy-4/+ sup-38 + hermaphrodites, and then selecting Dpy and Unc recombinants from their progeny. These animals were heterozygous for the recombinant chromosome (unc-26 + dpy-4/+ sup-38 dpy-4, or unc-26 + dpy-4/unc-26 sup-38 +) and to test for the presence of the sup-38 allele it was necessary to obtain homozygous strains (sup-38 dpy-4, or unc-26 sup-38). This was achieved by allowing the recombinant hermaphrodites to reproduce for one or two generations and then outcrossing these worms to wild-type males to determine whether they still carried a nonrecombinant chromosome or the unc-52 mutation. If the worms were homozygous for one of the recombinant chromosomes but still carried the unc-52 mutation then it was inferred that these strains were also carrying the suppressor mutation since none of the selected animals were paralyzed. Heterozygous males from strains which had lost unc-52 but were homozygous for one of the recombinant chromosomes were mated to unc-52(e1421) hermaphrodites. The presence of DpyUnc-52, or Unc-26Unc-52 progeny in the F2 generation indicated that the suppressor was absent, while the absence of these phenotypes indicated that it must be present. Strains carrying unc-26 or dpy-4 in cis with the suppressor have been maintained for both of these alleles.

Two-factor mapping of sup-38(ra5) and sup-38(ra14) relative to dpy-4 was done as follows. Heterozygous unc-52/+;dpy-4/+ males were crossed to homozygous unc-52;sup-38 hermaphrodites, and unc-52;dpy-4 +/+ sup-38 progeny were selected. These hermaphrodites were then allowed to reproduce, and the proportion of Unc and Dpy (non-Unc) recombinant worms in the F₂ progeny was used to determine the distance of the suppressor locus from dpy-4 using the formula $p = 1 - \sqrt{1 - 2R}$ (BRENNER 1974).

Characterization of suppressors: Complementation testing between the five sup-38 alleles, ra5, ra14, ra18, ra20 and ra21, was accomplished by mating heterozygous dpy-4sup-38/+ males with homozygous sup-38 hermaphrodites carrying a different sup-38 allele. The total number of progeny from each dpy-4 sup-38/+ sup-38 worm was determined and compared to the number of progeny produced by sup-38/+ animals. If the mean brood size of the heteroallelic mutants was less than that produced by heterozygous sup-38/+ animals the two alleles were said to fail to complement.

To test the allele specificity of the suppressors, class 1 unc-52/+;sup-38 dpy-4/++ hermaphrodites were obtained by mating sup-38 dpy-4/++ males to e444, e669, e998, st196, e1012, e1421 or e669su250 hermaphrodites. If DpyUncs were seen in the progeny of these worms, it was assumed

that the allele being tested was not suppressed. The sup-38 alleles were also tested against a lethal unc-52 allele, st549, by the following procedure. Hermaphrodites of genotype unc-52 (class 1)/unc-52 (class 2); sup-38 dpy-4/++ were obtained by mating unc-52 (class 1)/+;sup-38 dpy-4/++ males to unc-52(st549)/unc-52(st549)/mnDp34 hermaphrodites. Approximately 18-24 Dpy segregants were selected in the F_2 generation, and these Dpy worms were then outcrossed to determine their genotype. Nine or 10 outcrossed hermaphrodites from each mating were selected and their progeny screened for homozygous class 1 and class 2 progeny. If the Dpy animals being tested had been homozygous for st549, then class 1 animals should not be present. The absence of animals homozygous for the class 2 allele, st549, indicated that this allele was never suppressed. The ra5 and ra14 alleles were also tested for their ability to suppress the class 3 lethal allele, ut111. In this case, heterozygous unc-52(ut111)/+ males were mated with homozygous sup-38 dpy-4 hermaphrodites to produce unc-52(ut111)/+;sup-38 dpy-4/++ hermaphrodites. The Dpy progeny of these animals were then outcrossed and their F₂ progeny screened to determine whether any of these Dpy animals was homozygous for the ut111 mutation. If the original Dpy animal had been homozygous for *ut111*, then all of the outcrossed hermaphrodites would be expected to segregate homozygous ut111 larvae. The absence of ut111/ut111; sup-38 dpy-4/sup-38 dpy-4 animals indicated that the sup-38 mutation did not suppress *ut111*.

The five suppressor alleles, ra5, ra14, ra18, ra20 and ra21, were also tested against three other muscle-affecting genes to determine whether they were capable of suppressing different aspects of muscle disorganization. Male sup-38 dpy-4/++ animals were mated with hermaphrodites homozygous for unc-23(e25)V, unc-54(st1008)I, or unc-112(r367)V. The F₂ progeny of these crosses were screened for the presence of DpyUnc hermaphrodites and a ratio of $\frac{1}{16}$ DpyUncs was taken to indicate lack of suppression.

Elimination of suppressor activity: Reversion of suppression was achieved by mutagenizing unc-52;sup-38 dpy-4/nT1[let(m435)IV,V] worms with 0.025 M EMS, and selecting F1 offspring which were paralyzed. A total of approximately 240,000 worms of this genotype were screened. Six mutagenesis runs allowed us to screen 600 plates carrying approximately 400 worms each (of the correct genotype), 4-6 days after mutagenesis. All but one of the nine strains isolated in these screens carried a lethal mutation balanced by the nT1 translocation. Hermaphrodites from all of the strains carrying lethal mutations were mated to wild-type males, and the F2 progeny were counted and scored to determine whether or not the lethal mutation was closely linked to the dpy-4 locus. The proportion of viable Dpy recombinants was used to calculate the approximate map distance between the lethal mutation and dpy-4 using the formula $p = 1 - \sqrt{1 - 2R}$. The frequency of Unc-52 progeny was used to determine if suppressor activity was still present in the strain. A ratio of 3 wild-type:1 Unc indicated that suppression had been eliminated, while a ratio of 11 wild-type: I Unc was seen when suppressor activity was present. The lethal mutations which did not map to the sup-38 locus were eliminated by rebalancing the new sup-38 mutations in cis with dpy-4, over the nT1 chromosome. These were then maintained as heterozygous strains.

Three factor mapping of the maternal-effect lethal (Mel) mutation ra60 was done by mating ra5ra60 dpy-4/++ males to homozygous unc-22(s8) hermaphrodites. Recombinant DpyUnc progeny (unc-22? dpy-4/unc-22? dpy-4) were selected in the F₂ generation and scored for the presence of the Mel phenotype. If ra60 was to the right of dpy-4 then

all of the recombinants would be homozygous for ra60 and exhibit the Mel phenotype, whereas if it was between dpy-4 and *unc-22* only a certain proportion of the recombinants should be Mel.

Complementation testing of the null suppressor alleles ra61 and ra65 was done with two essential genes which map to the same region, *let-323* and *let-324* (CHAREST *et al.* 1990). Males heterozygous for *let-323(s1719)* or *let-324(s1727)* linked to *unc-22(s7)* were mated to hermaphrodites of genotype *unc-52;sup-38(ra5ra61) dpy-4/nT1* or *unc-52;sup-38(ra14ra65) dpy-4/nT1*. Well moving F₁ progeny were selected and plated individually to allow self-fertilization. The F₂ progeny of this cross were then screened for the presence of the *dpy-4* marker and the *unc-22-*linked lethal mutation. The presence of both of these markers on the same plate indicated the ability of the *sup-38* allele to complement the lethal mutations.

To determine whether the maternal-effect lethal phenotype could be zygotically rescued, heterozygous dpy-4 sup-38++ hermaphrodites were allowed to reproduce, and their Dpy progeny were mated to wild-type males. The semi-Dpy progeny of this cross were then scored for their ability to produce offspring. The presence of fertile F₂ hermaphrodites in this experiment indicated that a wild-type copy of the sup-38 locus in the zygote was sufficient to rescue the maternal-effect lethal phenotype produced by the null mutations.

RESULTS

Characterization of unc-52: Seven of the class 1 unc-52 alleles have been ranked here, based on the severity of the paralyzed phenotype (Figure 1; Table 1). The e998 and e444 alleles were the most severe, causing paralysis to begin early in the fourth larval (L4) stage. As adults, animals homozygous for these mutations were completely paralyzed, except for their heads, and were very thin and smaller than wild-type worms. The phenotype produced by the e669 allele was milder than that of the two above alleles, and mutants carrying this allele did not become paralyzed until later in the L4 stage. They also grew to a larger overall size, and were not as completely paralyzed as e998 or e444 worms. The e669 allele has been shown to be well suppressed by the amber suppressors, sup-5 III and sup-7 X (WATERSTON 1981). The st196, e1012 and e1421 alleles did not cause paralysis until early in adulthood although movement was slower than normal by the L4 stage. Mutants carrying these alleles were also larger, as mature adults, than the more severe unc-52 alleles. The st196 allele carries the transposon, Tc1, inserted in the unc-52 locus (D. MOERMAN, unpublished results). The other class 1 unc-52 allele which we ranked here was the temperature-sensitive mutation, e669su250 (S. EMMONS and L. JACOBSON, personal communication). Animals carrying this mutation exhibited the mildest phenotype of all. Even at the restrictive temperature (25°) paralysis did not begin until adulthood, and the worms rarely became totally paralyzed.

We have observed defects in the somatic gonads of the various Unc-52 mutants which had not been pre-

TABLE 1

Phenotypes of Unc-52 and Sup-38 mutants

unc-52 allele	sup-38 allele	Muscle defects ^a	Gonad, defects	Mean brood size
e998	+	Severe	Severe	20
e998	ra5	Moderate	Severe	22
e444	+	Severe	Severe	18
e444	ra14	Moderate	Severe	21
e669	+	Severe	Severe	19
st196	+	Severe	Severe	27
e1012	+	Severe	Severe	57
e1421	+	Severe	Severe	31
e1421	ra 18	Very mild	Mild	158
e1421	ra21	Very mild	Mild	143
e669su250	+	Moderate	Mild	133
e669su250	ra5	Mild	Severe	8
+	ra5	Mild	Severe	29
+	ra14	Mild	Severe	6
+	ra18	None	Mild	79
+	ra21	None	Mild	207
+	+	None	None	258

^a Severe: all adult animals exhibit severe disorganization in all body wall muscle cells posterior to the pharynx; moderate: most animals exhibit some muscle disorganization, but not in all body wall muscle cells; mild: less than 50% of animals exhibit some muscle disorganization; very mild: less than 90% of animals exhibit any muscle disorganization, and only in 1–2 cells per animal; none: wild-type muscle structure in all animals examined.

⁶ Severe: gonad structure always appears abnormal in some way, and animals are frequently sterile; mild: only mild abnormalities are present in the gonad, and animals are rarely sterile; none: wildtype gonad structure in all animals examined. ⁶ n = 30.

viously described (Figure 2). These defects were primarily limited to the proximal arm of the gonad which was sometimes enlarged (Figure 2b) or, in older adults, completely disorganized. This aspect of the phenotype was not fully penetrant and in some worms the gonad was not noticeably disorganized. In some animals, the anterior arm of the gonad was significantly shorter than the posterior arm, although this did not appear to be correlated with the overall degree of disorganization. The brood size of Unc-52 animals was very variable for any given allele (Table 1) and was correlated with the degree of disorganization of the gonad. In worms heteroallelic for a class 1 and a class 2 allele this defect was more severe, and in some worms the basement membrane, or the sheath cells surrounding the gonad appeared to have disappeared in places, allowing the oocytes to leak out into the body cavity of the worm (data not shown). Presumably, this contributed to the near-sterility of these animals. In worms heteroallelic for a class 1 and the class 3 allele the gonad appeared normal indicating that the ut111 allele complemented the class 1 alleles in this function.

All of the *unc-52* mutations used here were completely recessive. Complementation testing between the different class 1 alleles, and between class 1 alleles and the class 2 allele, *st549*, indicated a failure to



FIGURE 1.—Polarized light micrograph of body wall muscle cells of adult hermaphrodites grown at 20°: (a) N2, (b) unc-52(e998), (c) unc-52(e998);sup-38(ra5), (d) sup-38(ra5). Straight arrows point to dense bodies, curved arrows show thick filaments. Bar is 5 μ m.

complement. Animals which were heteroallelic for this class 2 allele and a class 1 allele became paralyzed by about the second larval stage and had very small brood sizes (0–15 progeny). The e1421 allele of unc-52, although similar in phenotype to the other class 1 alleles, behaved somewhat differently in these crosses. When st549/e1421 animals were obtained by mating homozygous e1421 hermaphrodites to males hetero-

zygous for the st549 allele, they became paralyzed by about the L2 stage, but were still viable and produced some offspring. These worms were similar in phenotype to those of genotype st549/e998 or st549/e669. However, when the reciprocal cross was done, the heteroallelic *unc-52* progeny were early larval lethals which resembled class 3, ut111, animals in phenotype. Similar crosses with any of the other class 1 alleles



FIGURE 2.—Nomarski micrograph showing one arm of the gonad of young adult hermaphrodites grown at 20°: (a) N2, (b) *unc*-52(e444), (c) *sup*-38(ra5). Arrows point to abnormalities in the structure. Bar is 5 μ m.

produced only the viable, paralyzed, heteroallelic animals described above.

When complementation tests were done with the class 1 alleles and the class 3 allele, ut111, the heteroallelic progeny appeared much healthier than either of the parental alleles. This indicated that alleles of these two classes complement one another at least partially. Complementation testing of ut111 with e444 was originally done by K. KONDO and I. KATSURA (personal communication). We tested ut111 with e998, e669, st196, e1012, e1421 and e669su250 by mating ut111/+ males with class 1 hermaphrodites as described in MATERIALS AND METHODS. Heteroallelic class 1/class 3 animals appeared nearly wild-type in size and ability to move, but muscle disorganization was consistently apparent when the body wall muscle cells were examined using polarized light microscopy (data not shown). The same type of disorganization was seen when strains heteroallelic for ut111 and any of the class 1 alleles were constructed, but was never seen in *ut111/+* worms. Reciprocal crosses were done for e1421, and the ut111/e1421 heteroallelic progeny were found to be larval lethals. The lethal stage of arrest was later in ut111/e1421 animals (L3–L4) than in st549/e1421 animals (L1–L2). Complementation testing between the class 2 allele, st549, and ut111 was also carried out by crossing ut111/+ males to st549/st549/+ hermaphrodites. These two alleles failed to complement one another and heteroallelic st549/ut111 animals arrested at the same stage and had the same general appearance as homozygous ut111 animals.

Isolation of revertants: We have isolated revertants of class 1 Unc-52 mutants in order to identify other genes which normally interact with this locus. Fortyfour *unc-52* revertants were isolated after screening approximately 1.3×10^8 animals homozygous for a class 1 allele (Table 2). Since screens were done in the F₁ generation, all of the animals selected were heterozygous for the new mutation. Revertant worms were picked and allowed to reproduce for several generations until they were homozygous (*i.e.*, no longer

TABLE 2Screen for suppressors of Unc-52

unc-52 allele	No. of revertants isolated	Total No. screened	Intra- genics	Inter- genics	Untested	Suppressors mapped to LGIV(R)
e998	12	3.5×10^{7}	9	2	1	ra5
						ra 40
e444	2	1×10^{7}	0	2	0	ra13
						ral4
e669	6	2.5×10^{7}	1	5	0	ra10
						ra17
e1012	2	1×10^{7}	2	0	0	
e1421	22	2.5×10^{7}	10	11	1	ra18
						ra20
						ra21
Total	44	13.5×10^{7}	22	20	2	

segregated Unc progeny), and then were outcrossed to wild-type males to determine whether the revertant phenotype was due to a second mutation within the unc-52 locus or to a mutation at a separate locus which suppressed the original unc-52 mutation. The results are presented in Table 2. Twenty-two of the revertants produced no Unc-52 worms after being outcrossed, indicating that they now carry a second mutation within, or very close to, the unc-52 locus. Twenty of the revertants did segregate Unc-52 progeny when outcrossed and were presumed to carry unlinked suppressors of unc-52. The remaining two revertants were not outcrossed because they showed only partial suppression of the unc-52 mutation and, as a result, were difficult to work with. Both intragenic revertants and intergenic suppressors were obtained for three unc-52 alleles, but the screen using the e444 allele did not produce any intragenic revertants, and we did not obtain any intergenic suppressors of the e1012 allele.

Mapping of intergenic suppressors: Seven intergenic suppressor mutations (ra5, ra13, ra14, ra18, ra20, ra21 and ra40) were positioned near dpy-4 on the right arm of chromosome IV (Figure 3). Genetic mapping was performed using Dpy markers from each of the *C. elegans* linkage groups as described in MA-TERIALS AND METHODS. In each case, when dpy-4 on LG IV was used, approximately one quarter of the F₂ progeny were DpyUnc, indicating linkage of the suppressor mutations. In contrast, only $\frac{1}{16}$ DpyUnc progeny were seen with the dpy genes on the other chromosomes.

The ra10 and ra17 suppressors were isolated in strains carrying dpy-20(e2017)IV (see MATERIALS AND METHODS). When these strains were outcrossed to wild-type males we observed that the suppressor locus segregated with dpy-20 in the F₂ generation, indicating that these suppressors were also on LG IVR.

Two of the suppressor mutations, ra20 and ra21, were positioned to the left of dpy-4 by three-factor



FIGURE 3.—Approximate map location of sup-38 on LG IV(R).

mapping. Approximately one third (5/16) of the Dpy-4 and two thirds (8/14) of the Unc-26 recombinants obtained from unc-52;+ sup+/unc-26 + dpy-4 hermaphrodites carried the suppressor, suggesting that both mutations lie within the three map unit interval between unc-26 and dpy-4, slightly more than one map unit from dpy-4. Two-factor mapping of ra5 and ra14positioned these alleles approximately 1.1 map units away from the dpy-4 locus.

Characterization of intergenic suppressors: The five suppressor alleles we have analyzed confer an essentially wild-type phenotype in terms of appearance and movement even when the unc-52 mutation is still present in the strain. Suppressor strains free of the unc-52 mutations were obtained for the ra5, ra14, ra18, ra20 and ra21 alleles as described in MATERIALS AND METHODS. When the muscle cells of animals carrying a suppressor mutation were observed using polarized light microscopy, most of the cells exhibited an almost wild-type pattern of birefringent filaments and dense bodies (Figure 1c). However, very rarely an occasional cell in an otherwise wild-type worm was disrupted (Figure 1d). This disruption appears to be due to fragility in the muscle structure of worms carrying the suppressor mutation since it appeared more often after the worms had been rolled under the coverslip to expose the different muscle quadrants. This fragility is not apparent in wild-type animals.

Worms carrying any of the suppressor mutations also exhibited some degree of gonad disorganization when observed using Nomarski microscopy (Figure 2c). The ra5 and ra14 alleles consistently exhibited more severe defects which were reflected in the greatly reduced brood sizes of homozygous animals. The ra18, ra20 and ra21 mutants had only slightly reduced brood sizes compared to wild-type worms (Table 1), and showed a variable amount of gonadal disorganization under Nomarski optics.

TABLE 3

Complementation testing between sup-38 alleles (n > 10) (mean brood size)

	+	ra5	ra 14	ra18	ra20	ra21
+	259	187	144	156	200	228
ra5		29	20	108	119	166
ra 14			6	84	ND	ND
ra 18				79	ND	ND
ra20					79	ND
ra21						207

ND = not determined.

Strains carrying ra5, ra14 and ra18 also exhibited a slight Dpy phenotype, and those carrying the ra5and ra14 alleles were Him, producing approximately 5% male progeny. To determine whether the Him phenotype was due to a mutation in the tra-3 locus (HODGKIN and BRENNER 1977) which maps near the suppressor alleles, these males were mated to wildtype hermaphrodites to determine whether they were fertile. Although their fertility was well below that of wild-type males, they did produce offspring, indicating that they were not carrying a tra-3 mutation. Like their hermaphrodite siblings, the gonads of these males appeared variably abnormal when observed under Nomarski microscopy (data not shown).

The fact that all of the suppressor mutations mapped to the same location on LG IV suggested that they might be alleles of the same gene. The low fecundity of the ra5 and ra14 alleles was found to be semidominant. Homozygous ra5 and ra14 hermaphrodites had, on average, less than 20 progeny each and exhibited a high incidence of sterility (>10%). Heterozygous animals, on the other hand, produced more than 140 progeny each (Table 3) and were rarely sterile. Using this phenotype, we determined that ra5 and ra14 failed to complement one another since hermaphrodites heteroallelic for these two mutations produced less than 20 progeny each and were frequently sterile. The ra21 mutation was also found to be allelic with ra5 and ra14 based on the fact that heteroallelic ra21/ra5 mutants produced fewer progeny than homozygous or heterozygous ra21 animals (Table 3). The ra18 and ra20 alleles can also be said to fail to complement ra5 since ra18/ra5 and ra20/ra5 animals had smaller brood sizes than ra18/+ or ra20/+ animals. However, unlike the ra21 allele, heteroallelic ra18/ra5 and ra20/ra5 animals had greater brood sizes than homozygous ra18 or ra20 mutants. The locus defined by these mutations has been designated sup-38.

Allele specificity of suppressors: The allele specificity of the five suppressor mutations described above was determined as described in MATERIALS AND METH-ODS. The ra5 and ra14 mutations suppress the e998, e444, e669am, st196::Tc1, e1012, e1421 and

e669su250ts alleles of unc-52. These results are based on the absence of DpyUnc progeny produced in the F_2 generation of unc-52/+;sup-38 dpy-4/++ hermaphrodites. In all cases, suppression by these two alleles was semidominant. Homozygous unc-52 hermaphrodites carrying two copies of ra5 or ra14 were able to move better than those with a single copy of the suppressor mutation, but movement was significantly improved even with one copy of the suppressor. The three suppressors derived from e1421 screens (ra18, ra20 and ra21) appear to be specific to that allele, since DpyUnc animals were always seen in the F_{2} progeny at the expected ratio when these suppressors were tested with the e998, e444, e669am, st196::Tcl, e1012 and e669su250 alleles. These three suppressor alleles were completely dominant in the e1421 background, and there was no visible difference in phenotype when one or two copies of the suppressor alleles were present.

The ra5, ra14, ra18, ra20 and ra21 alleles were also tested for their ability to suppress a class 2 allele, unc-52(st549), by crossing class 1 unc-52/+;sup-38 dpy-4/ ++ males with hermaphrodites carrying st549 (unc-52(st549)/unc-52(st549)/mnDp34). Dpy hermaphrodites were selected in the F2 generation and then outcrossed to determine whether any were homozygous for the st549 mutation. It was expected that one quarter of these Dpy worms would be homozygous for the lethal mutation if suppression was occurring. None of the alleles tested appeared to suppress the lethal phenotype of st549 since unc-52(st549);sup-38 dpy-4 adult animals were never found in the F₂ generation. Similar crosses were also done to test the ra5 and ra14 suppressor alleles with the class 3 mutation, unc-52(ut111). In this case heterozygous unc-52(ut111)/+ males were mated to homozygous sup-38dpy-4 hermaphrodites. No unc-52(ut111);sup-38 dpy-4 progeny were seen in the F₂ generation of these crosses either, indicating lack of suppression.

The same five suppressors were also tested against three other muscle-affecting genes to determine whether they were capable of suppressing different aspects of muscle disorganization. To do this, sup-38dpy-4/++ males were mated with hermaphrodites homozygous for unc-23(e25)V, unc-54(st1008)I, or unc-112(r367)V. The F₂ progeny of these crosses were screened for the presence of DpyUnc hermaphrodites and, in each case, the expected ratio of $^{1}/_{16}$ DpyUncs was observed, indicating lack of suppression.

Elimination of suppressor activity: Since the suppressors arose at such a low frequency ($\sim 1.3 \times 10^{-7}$) in our *unc-52* reversion screens, it was not likely that any of these alleles were the null state of the gene. We attempted to isolate loss-of-function alleles in order to determine the normal function of the suppressor locus. We reasoned that the induction of a null



FIGURE 4.—Protocol used to isolate loss-of-function alleles of *sup-38*. Asterisk indicates loss of suppressor activity.

mutation in a gene already carrying a suppressor allele should eliminate the dominant suppressor phenotype. The strain constructed for this experiment was homozygous for an unc-52 mutation and heterozygous for the suppressor in cis with dpy-4, over a recessive lethal balancer chromosome (unc-52/unc-52;sup-38 dpy-4/ nT1[let(m435)IV;V]) (see Figure 4). Because the suppressor mutations were dominant, these worms were well moving and semi-Dpy, and they segregated both semi-Dpy and Dpy progeny. After mutagenesis, the F_1 progeny of these hermaphrodites were screened for the return of the paralyzed phenotype which indicated loss of the suppressor function. This screen allowed the recovery of putative null sup-38 alleles regardless of whether the phenotype of these mutations was Wild, Unc or Let.

Approximately 240,000 animals were screened as described above, and nine strains of paralyzed worms were isolated (Table 4). The initial analysis was complicated by the fact that most of the strains isolated carried more than one new mutation, presumably because the nT1 translocation allowed the recovery of mutations anywhere in the balanced region (covering approximately 50 map units). Mutations that did not affect *sup-38* were removed from the strains

as described in MATERIALS AND METHODS, and the *sup-38 dpy-4* chromosomes carrying new *sup-38* mutations were then rebalanced over nT1.

Four of the nine strains regained suppressor activity after outcrossing and were not analyzed further (see MATERIALS AND METHODS and Table 4). Four strains appeared to have lost suppressor activity completely, suggesting that second-site mutations had been induced in the sup-38 locus. Suppressor activity in the remaining strain (ra5ra55) appeared to have been reduced, but not eliminated since suppression became recessive instead of dominant (see Table 4). The phenotypes of these five putative sup-38 alleles were very similar. Homozygous sup-38 dpy-4 hermaphrodites produced by heterozygous parents were well moving, large Dpys. These animals produced only very small, Dpy progeny which died at the L4 stage or, if they reached adulthood, produced few or no progeny. Thus, these strains exhibited a maternal effect lethal (Mel) phenotype. Strains carrying ra5ra60 or ra5ra61 mutations were quite strict Mels, while the ra5ra55, ral4ra65 and ral4ra66 mutations were leaky and caused only a significant reduction in brood size. Dpy progeny obtained from dpy-4 sup-38(ra5ra61)/nT1 or dpy-4 sup-38(ra14ra65)/nT1 hermaphrodites were mated to wild-type males to determine whether the lethal phenotype exhibited by these mutants was a strict maternal-effect. The heterozygous progeny of these animals were not Mel, indicating that a wildtype copy of *sup-38* in the zygote rescued the maternal phenotype. Another characteristic common to all of the sup-38(Mel) alleles was the maternal effect enhancement of the Dpy-4 phenotype. Animals carrying a sup-38 mutation in cis with dpy-4 were much shorter than mutants carrying the dpy-4 mutation on its own. Sup-38 Mel mutants displayed no gross abnormalities when observed using Nomarski microscopy, and had completely wild-type muscle structure when examined under polarized light.

Based on the reduced brood sizes of these strains, complementation tests were done as described in MA-TERIALS AND METHODS, and the results obtained indicated that all five mutations were alleles of a single locus. Animals heteroallelic for any of the putative null alleles were very small Dpys which exhibited the Mel phenotype typical of homozygous loss-of-function mutants.

The maternal effect lethal phenotype of the null suppressor mutations was used to determine their map position relative to dpy-4. Homozygous DpyUnc-22 recombinants obtained from + sup-38(Mel) dpy-4/unc-22++ hermaphrodites were tested to determine whether any of them exhibited a maternal effect lethal phenotype. If the suppressor locus was to the right of dpy-4 then all of the DpyUnc recombinants should have been Mel. However, five out of the 29 recom-

E. J. Gilchrist and D. G. Moerman

TABLE	4
-------	---

Screen for Sup-38 loss-of-function mutants

Suppressor allele	No. of chromosomes screened	No. of Uncs isolated	Phenotype	No. of Sup-38 nulls or hypomorphs
ra5	80,000	1	Carries lethal near $dpy-4$ still has suppressor	3
		1	Suppressor is now recessive (and Mel)	
		2	Carries lethal on LGV lost suppressor (now Mel)	
ra 14	40,000	1	Carries lethal on LGV still has suppressor	2
		2	Carries lethal near <i>dpy-4</i> lost suppressor (now Mel)	
ra18	40,000	1	Carries lethal on LGV still has suppressor	0
ra21	80,000	1	Carries lethal near <i>dpy-4</i> still has suppressor	0

binants were not, confirming that the Mel mutations lie to the left of dpy-4, in the same region as the Sup mutations.

The ability of ra61 and ra65 to complement two essential genes in this region, *let-323* and *let-324* indicated that the *sup-38* alleles represent a newly defined genetic locus (see MATERIALS AND METHODS).

DISCUSSION

Mutations in the unc-52 locus of C. elegans have been divided into three groups based on their mutant phenotypes and complementation patterns. Class 1 mutations result in animals which become progressively paralyzed as they mature (BRENNER 1974). The degree of final paralysis and the stage of its onset vary depending upon the allele and are correlated with a disorganization of the muscle sarcomeres at the cellular level. We have observed that worms carrying class 1 alleles also exhibit variably disorganized somatic gonads and have greatly reduced brood sizes. This may be due to the fact that the sheath cells which surround the proximal arm of the gonad are myoepithelial cells (HIRSH, OPPENHEIM and KLASS 1976) which contain some of the same components as the body wall muscle cells (FRANCIS and WATERSTON 1991). The unc-52 gene product may, therefore, be important to both of these types of contractile cells.

A second class of mutation at the unc-52 locus, represented by the allele st549, causes homozygous animals to arrest at the twofold stage of embryogenesis before elongation is complete. Mutants of this class show an almost complete lack of movement even before elongation ceases (B. WILLIAMS, personal communication). A class 3 mutation, ut111, also results in embryonic lethality, and embryos homozygous for this mutation also fail to elongate beyond the twofold stage, although they do hatch and are capable of some movement (K. KONDO and I. KATSURA, personal communication). PRIESS and HIRSH (1986) have shown that cytoskeletal elements of the hypodermis are responsible for generating the circumferential tension necessary for elongation of the C. elegans embryo. The phenotype exhibited by animals homozygous for

lethal mutations in many muscle-affecting genes is consistent with the idea that muscle function is also necessary for elongation beyond the twofold stage (BARSTEAD and WATERSTON 1991; B. WILLIAMS and R. H. WATERSTON, personal communication). It is not surprising, therefore, that mutants homozygous for lethal mutations in *unc-52* fail to elongate since the product of this gene appears to be involved in the interaction between muscle cells and the extracellular matrix which connects them to the hypodermis.

The results of complementation tests with the various unc-52 mutations have revealed that class 1 and class 3 alleles complement each other almost fully. Class 1/ut111 animals exhibited a slight disorganization of the body-wall muscles, but young animals were essentially wild-type in size and ability to move. Analysis of unc-52 cDNAs indicates that several different transcripts are produced from this gene (T. ROGALSKI and D. MOERMAN, unpublished results) and this may explain the complementation pattern we have observed. Since the st549 allele produced the most severe phenotype and failed to complement all other alleles, it may represent the null state of unc-52. The class 1 and class 3 mutations, on the other hand, may affect either different, specific gene transcripts or different domains within a single Unc-52 polypeptide. We have identified over 20 intragenic revertants of the class 1 alleles in screens for suppressors of these mutations. Further analysis of these revertants should contribute to our understanding of the unc-52 locus.

The e1421 allele behaved differently from other class 1 alleles in complementation tests with the st549and ut111 alleles. When e1421 was maternally inherited in these complementation tests, the results were similar to those obtained using other class 1 alleles. However, when e1421 was inherited from the male, st549/e1421 animals were early larval lethals which resembled ut111 homozygotes in phenotype. The ut111/e1421 heteroallelic animals from analogous crosses were also lethal, but these animals did not die until the late larval stages or early in adulthood. This paternal effect was not seen with any of the other alleles and the cause of it is unclear at this point. The results of reversion studied revealed that there are also other differences between *e1421* and the other class 1 alleles (see below).

The unc-52 protein must play an essential role in development since some mutations at this locus result in embryonic lethality. It must also be involved in maintaining the normal structure of the muscles and the somatic gonad in adult worms since class 1 mutations in the gene result in disruption of these tissues. Sequence analysis (T. ROGALSKI and D. MOERMAN, unpublished results) indicates that unc-52 encodes a protein with considerable similarity to the mouse basement membrane heparan sulfate proteoglycan, perlecan (NOONAN et al. 1991). The basement membrane appears very early during embryogenesis in all higher organisms, and is known to be important in the development and maintenance of most tissues (MARTIN and TIMPL 1987). The fact that the Unc-52 protein in C. elegans is localized to the basement membrane but affects the integrity of the internal structure of the muscle cells allows us to examine the role of the basement membrane in vivo in determining and maintaining the internal structure of the cells or tissues it underlies.

The conservation of extracellular matrix proteins between higher organisms and a genetically manipulable system like C. elegans has provided a system for identifying other molecules which are involved in the formation and maintenance of this structure. Mutations in a previously unidentified locus, sup-38, have been isolated and shown to suppress the paralysis typical of class 1 mutations in unc-52. The sup-38 mutations suppressed the disorganized muscle phenotype of the class 1 mutations, but did not suppress the gonad disorganization associated with these alleles, nor did they suppress the lethal alleles of unc-52. A total of nine suppressor mutations have been mapped to LG IV(R). Three of these, ra5, ra14 and ra21, have been shown to be allelic, and two other mutations, ra18 and ra20 are most likely alleles of sup-38 as well. Three of the five intergenic suppressors, ra18, ra20 and ra21, are allele-specific and will only suppress unc-52(e1421), whereas ra5 and ra14 are able to suppress all of the class 1 alleles. The specificity of the e1421-specific suppressors is not just a reflection of the fact that e1421 is one of the mildest unc-52 alleles because they failed to suppress e669su250 which is even milder. All five suppressor alleles confer a similar phenotype, with mutant hermaphrodites exhibiting a slightly fragile muscle structure and variable gonad disorganization. The ra18, ra20 and ra21 alleles result in a milder phenotype than the other two alleles and are dominant suppressors, whereas ra5 and ra14 are semidominant. Mutants of genotype ra5/+ or ra14/+ are healthier and have larger brood sizes than homozygous mutant worms (Table 3), but still exhibit some gonad and muscle disorganization, and have a more severe phenotype than *ra18*, *ra20* or *ra21* homozygous animals (data not shown).

The suppressor alleles of sup-38 are not likely to represent the null state of the gene because they arose at an extremely low frequency (1.3×10^{-7}) . We have isolated four loss-of-function alleles of sup-38 by selecting for loss of suppressor activity. The frequency of this type of mutation was approximately one in 30,000 (excluding the e1421-specific suppressor screens), indicating either that sup-38 is a relatively small target for EMS mutagenesis (BRENNER 1974), or that these are not null alleles of the locus. Alleles of this type show a maternal effect since the late larval lethal phenotype was only seen in progeny from homozygous sup-38 hermaphrodites. A fifth mutation isolated in these screens results in loss of the dominant suppressor function, so that worms carrying this new mutation are now only capable of suppressing the class 1 unc-52 mutant phenotype when they are homozygous. Mutants homozygous for this recessive suppressor mutation also display the Mel phenotype typical of the putative sup-38 null mutants, but occasionally homozygous worms mature and produce offspring. Two of the other strains also exhibited this leaky Mel phenotype and may, therefore, also be hypomorphs rather than complete loss-of-function mutants. The lack of stable deficiencies for this region has prevented a thorough genetic analysis of these mutations. Surprisingly, the body wall muscles of worms homozygous for the null, or loss-of-function mutations were completely wild type, indicating that the sup-38 gene product is not essential for normal muscle structure. The fragile sarcomere organization seen in the suppressor mutants may, therefore, be due to abnormal expression of sup-38. It is unclear, at this point, whether suppression is due to a sup-38 gene product interacting with, or simply replacing a mutant unc-52 gene product. Molecular analysis of the suppressor locus should provide insight into its normal function and its mechanism of suppression of unc-52.

We would like to thank TERESA ROGALSKI for her support and encouragement, and for both scientific and literary advice. We wish to thank BEN WILLIAMS, BOB WATERSTON and an anonymous reviewer for constructive criticism of this manuscript. We would also like to thank BEN WILLIAMS for sending us the strain carrying the st549 allele of unc-52, and KAZUNORI KONDO and I. KATSURA for providing us with strains carrying the ut111 allele. We are grateful to D. L. BAILLIE, J. HODGKIN, A. M. ROSE, R. H. WATER-STON, and the people in their labs for strains obtained from them. Some strains used in this work were provided by the Caenorhabditis Genetics Center (University of Missouri, Columbia), which is funded by the National Institutes of Health National Center for Research Resources. This project was supported by grants from the Muscular Dystrophy Association of Canada, the National Science and Engineering Research Council, and Medical Research Council of Canada.

LITERATURE CITED

- ANDERSON, P., and S. BRENNER, 1984 A selection for myosin heavy chain mutants in the nematode, *Caenorhabditis elegans*. Proc. Natl. Acad. Sci. 81: 4470-4471.
- BARSTEAD, R. J., and R. H. WATERSTON, 1991 Vinculin is essential for muscle function in the nematode. J. Cell Biol. 114: 715– 724.
- BRENNER, S., 1974 The genetics of *Caenorhabditis elegans*. Genetics 77: 71-94.
- BURRIDGE, K., K. FATH, T. KELLY, G. NUCKOLLS and C. TURNER, 1988 Focal adhesions: transmembrane junctions between the extracellular matrix and the cytoskeleton. Annu. Rev. Cell Biol. 4: 487–525.
- CHAREST, D. L., D. V. CLARK, M. E. GREEN and D. L. BAILLIE, 1990 Genetic and fine structure analysis of *unc-26(IV)* and adjacent regions in *Caenorhabditis elegans*. Mol. Gen. Genet. **221:** 466-474.
- FERGUSON, E. L., and H. R. HORVITZ, 1985 Identification and characterization of 22 genes that affect the vulval cell lineages of *Caenorhabditis elegans*. Genetics **110**: 17–72.
- FRANCIS, R., and R. H. WATERSTON, 1985 Muscle organization in *Caenorhabditis elegans*: localization of proteins implicated in thin filament attachment and I-band organization. J. Cell Biol. 101: 1532-1549.
- FRANCIS, R., and R. H. WATERSTON, 1991 Muscle cell attachment in *Caenorhabditis elegans*. J. Cell Biol. **114**: 465–479.
- GREENWALD, I., and H. R. HORVITZ, 1982 Dominant suppressors of a muscle mutant define an essential gene of *Caenorhabditis elegans*. Genetics **101**: 211–225.
- HARTMAN, P. E., and J. R. ROTH, 1973 Mechanisms of suppression. Adv. Genet. 17: 1-105.
- HIRSH, D., D. OPPENHEIM and M. KLASS, 1976 Development of the reproductive system of *Caenorhabditis elegans*. Dev. Biol. 49: 200-219.
- HODGKIN, J., and S. BRENNER, 1977 Mutations causing transformation of sexual phenotype in the nematode *Caenorhabditis elegans*. Genetics **86**: 275–287.
- HODGKIN, J., K. KONDO and R. H. WATERSTON, 1987 Suppression in the nematode *Caenorhabditis elegans*. Trends Genet. **3:** 325– 329.
- HODGKIN, J., A. PAPP, R. PULAK, V. AMBROS and P. ANDERSON, 1989 A new kind of informational suppression in the nematode *Caenorhabditis elegans*. Genetics **123**: 301-313.
- HORVITZ, H. R., S. BRENNER, J. HODGKIN and R. K. HERMAN, 1979 A uniform genetic nomenclature for the nematode *Caenorhabditis elegans*. Mol. Gen. Genet. **175**: 129–133.
- MACKENZIE, J. M., R. L. GARCEA, J. M. ZENGEL and H. F. EPSTEIN, 1978 Muscle development in *Caenorhabditis elegans*: mutants exhibiting retarded sarcomere construction. Cell 15: 751–762.
- MARTIN, G. R., and R. TIMPL, 1987 Laminin and other basement membrane components. Annu. Rev. Cell Biol. **3:** 57–85.

- MARUYAMA, I. M., D. M. MILLER and S. BRENNER, 1989 Myosin heavy chain gene amplification as a suppressor mutation in *Caenorhabditis elegans*. Mol. Gen. Genet. **219**: 113–118.
- MOERMAN, D. G., S. PLURAD, R. H. WATERSTON and D. L. BAILLIE, 1982 Mutations in the *unc-54* myosin heavy chain of *Caenor-habditis elegans* that alter contractility but not muscle structure. Cell **29:** 773–781.
- NOONAN, D. M., A. FULLE, P. VALENTE, S. CAI, E. HORIGAN, M. SASAKI, Y. YAMADA and J. R. HASSELL, 1991 The complete sequence of perlecan, a basement membrane heparan sulfate proteoglycan, reveals extensive similarity with laminin A chain, low density lipoprotein-receptor, and the neural cell adhesion molecule. J. Biol. Chem. 266: 22939–22947.
- PRIESS, J. R., and D. HIRSH, 1986 Caenorhabditis elegans morphogenesis: the role of the cytoskeleton in elongation of the embryo. Dev. Biol. 117: 156–173.
- RIDDLE, D., and S. BRENNER, 1978 Indirect suppression in Caenorhabditis elegans. Genetics 89: 299–314.
- ROGALSKI, T., and D. RIDDLE, 1988 A Caenorhabditis elegans RNA polymerase II gene, *ama-1 IV*, and nearby essential genes. Genetics **118**: 61–74.
- ROSENBLUTH, J., 1965 Structural organization of obliquely striated muscle fibers in Ascaris lumbricoides. J. Cell Biol. 25: 495-515.
- SULSTON, J., and J. HODGKIN, 1988 Methods, in *The Nematode*, *Caenorhabditis elegans*, edited by W. B. WOOD. Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y.
- VENOLIA, L., and R. H. WATERSTON, 1990 The *unc-45* gene of *Caenorhabditis elegans* is an essential muscle-affecting gene with maternal expression. Genetics **126**: 345–353.
- WATERSTON, R. H., 1981 A second informational suppressor, sup-7 X, in Caenorhabditis elegans. Genetics 97: 307-325.
- WATERSTON, R. H., 1988 Muscle, pp 281-336 in The Nematode, Caenorhabditis elegans, edited by W. B. WOOD. Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y.
- WATERSTON, R. H., 1989 The minor myosin heavy chain, mhcA, of *Caenorhabditis elegans* is necessary for the initiation of thick filament assembly. EMBO J. 8: 3429–3436.
- WATERSTON, R. H., and S. BRENNER, 1978 A suppressor mutation in the nematode acting on specific alleles of many genes. Nature 275: 715–719.
- WATERSTON, R. H., D. HIRSH and T. R. LANE, 1984 Dominant mutations affecting muscle structure in *Caenorhabditis elegans* that map near the actin gene cluster. J. Mol. Biol. 180: 1473– 1496.
- WATERSTON, R. H., J. N. THOMSON and S. BRENNER, 1980 Mutants with altered muscle structure in *C. elegans*. Dev. Biol. **77**: 271-302.

Communicating editor: R. K. HERMAN