# *Age-related changes in the sulphation of the chondroitin sulphate linkage region from human articular cartilage aggrecan*

Robert M. LAUDER<sup>\*1</sup>, Thomas N. HUCKERBY<sup>\*</sup>, Gavin M. BROWN<sup>\*</sup>, Michael T. BAYLISS<sup>+</sup> and Ian A. NIEDUSZYNSKI<sup>\*</sup> \*Department of Biological Sciences, Lancaster University, Bailrigg, Lancaster LA1 4YQ, U.K., and †The Royal Veterinary College, Royal College Street, London NW1 0TU, U.K.

The chondroitin sulphate (CS) linkage regions have been isolated from human articular cartilage aggrecan (from 10- to 72-yearolds) by chondroitin ABC endolyase digestion and size-exclusion chromatography. Linkage region hexasaccharides have been characterized and their abundance estimated by high-pH anionexchange chromatography. The basic structure for the CS linkage region oligosaccharides identified from human aggrecan is as follows:  $\Delta UA(\beta 1-3)GalNAc[0S/4S/6S](\beta 1-4)GlcA(\beta 1-3)Gal [0S/6S](\beta1-3)Gal(\beta1-4)Xyl$ , where  $\Delta UA$  represents 4,5-unsaturated hexuronic acid, 4S and 6S represent an O-ester sulphate group on C-4 and C-6 respectively, and 0S represents zero sulphation. There are significant age-related changes in the abundance of the various *N*-acetylgalactosamine (GalNAc) sulphation forms identified, occurring up to approx. 20 years old. During the period from 10 to 20 years old the level of GalNAc 6-sulphation at the linkage region increases from approx.  $43\%$ to approx.  $75\%$ , while there is a corresponding reduction in

#### *INTRODUCTION*

The sulphated glycosaminoglycan (GAG), chondroitin sulphate (CS), is an abundant component of the large aggregating proteoglycan aggrecan, and it is known to contribute to the loadbearing properties of articular cartilage. The specific sulphation patterns in CS are important in macromolecular interactions and often the presence of 4-sulphated *N*-acetylgalactosamine (GalNAc) and the absence of 6-sulphation governs the interaction [1]. Such interactions include cytoadherence of malaria-infected red blood cells [2], the adherence of the malaria parasite to human placenta [3,4], and the regulation of neurite outgrowth [5]. However, it is a chondroitin 6-sulphate-containing epitope of the proteoglycan DSD-1, which is reported to influence neurite outgrowth [6]. In addition, the expression of some CS epitopes in the rodent are known to be developmentally regulated [7,8].

CS chains from articular cartilage comprise a linkage region, a repeat region and a chain cap. The repeat region consists of a repeating disaccharide  $[-4)$ GlcA( $\beta$ 1-3)GalNAc( $\beta$ 1-]<sub>n</sub>, which may be sulphated at C-4 and/or C-6 of GalNAc and at C-2 of glucuronic acid (GlcA). In adult articular cartilage CS 6 sulphation dominates (approx.  $95\%$ ) over both unsulphated (approx.  $\langle 1\% \rangle$  and 4-sulphated (approx. 5%) GalNAc residues, and the level of GlcA sulphation is also low [9]. The linkage region has the general structure  $Gal(\beta1-3)Gal(\beta1-4)Xyl(\beta1-4)$ O-)Ser [10]; a variety of CS linkage region structures with various sulphation patterns have been identified in articular cartilage [11], bovine nasal septum cartilage [12,13], rat chondrosarcoma

unsulphated (approx.  $30\%$  to approx.  $20\%$ ) and 4-sulphated (approx. 25% to approx.  $6\%$ ) GalNAc residues. There is also an increase in the incidence of linkage region galactose 6-sulphation (approx.  $2\%$  to approx.  $10\%$ ) which was only observed in linkage regions with GalNAc 6-sulphation. Beyond 20 years old there are few changes in the relative abundance of these GalNAc sulphation variants; however, there is a slight increase in the abundance of 6-sulphation between approx. 20 years old and approx. 40 years old and a slight decrease in its abundance beyond approx. 40 years old. Our data show that in the majority of chains from tissues of all ages the GalNAc residue closest to the linkage region is 6-sulphated, but the level of GalNAc 6 sulphation within the linkage region is lower than the average level observed within the repeat region.

Key words: ABC lyase, chondroitin sulphate, endolyase, glycosaminoglycan, osteoarthritis.

[14] and whale cartilage [15]. In addition, there are reports of galactose (Gal) 4- and 6-sulphation in CS isolated from aggrecan of shark cartilage [16,17], bovine nasal septum [12,13] and bovine articular cartilage [11]. For a review of CS linkage region structures see http://www.glycoforum.gr.jp/science/word/ proteoglycan/PGA06E.html. Clearly, further studies are required in order to elucidate the reasons for these, possibly tissuerelated, variations in linkage region sulphation.

The linkage region described above is common to CS, dermatan sulphate, heparan sulphate (HS) and heparin. Several studies have shown that the amino acid sequence of the protein core may have an effect upon the GAG synthesized [19,20]; however, the detailed structure of the priming oligosaccharide is also important [21,22]. The structure of the repeat region close to the linkage region of CS from adult bovine articular cartilage has been shown to differ from the bulk of the repeat region; it contains a higher than average level of unsulphated and 4 sulphated GalNAc residues and a concomitantly lower level of 6-sulphation [11,23,24]. In contrast, the CS chain cap has been found to have a high sulphation density with a 4,6-disulphated GalNAc being abundant [25,26].

There is mounting evidence for age-related changes in the structure of GAGs. Bayliss et al. [27] reported an age-related increase in the ratio of 6- to 4-sulphated GalNAc residues in human articular cartilage CS, while Plaas et al. [25,26] reported an age-related increase in the abundance of 4,6-disulphated GalNAc at the caps of such chains. The capping structure of articular cartilage keratan sulphates (KSs) also shows an age-

Abbreviations used: CS, chondroitin sulphate; GAG, glycosaminoglycan; Gal, galactose; GalNAc, *N*-acetylgalactosamine; GlcA, glucuronic acid; HPAEC, high-pH anion-exchange chromatography; HS, heparan sulphate; KS, keratan sulphate; ΔUA, 4,5-unsaturated hexuronic acid (4-deoxy-α-<br>L-threo-hex-4-enepyranosyluronic acid).

<sup>&</sup>lt;sup>1</sup> To whom correspondence should be addressed (e-mail r.lauder@lancaster.ac.uk).

related change [28,29], with an increasing level of  $\alpha$ (2-6)-linked *N*-acetylneuraminic acid [28,29], while the repeat region shows an increasing chain length, level of Gal sulphation and fucosylation [29]. Each of these studies found that in the human the greatest changes in the abundance of these structures occurred in the period up to 20 years old.

The sulphation profile of CS, both at different positions along a chain and at different ages, is of considerable interest, as it has been proposed that determination of average 4- or 6-sulphation levels may be an appropriate marker of articular cartilage turnover and of osteoarthritis [30]. In the present study the structure and age-related changes in the CS linkage regions from human aggrecan are examined.

#### *MATERIALS AND METHODS*

Sepharose CL-6B was purchased from Pharmacia (Uppsala, Sweden), Toyapearl HW-40s was obtained from Tosohaas via Anachem (Luton, Herts., U.K.), and Bio-Gel P2 was purchased from Bio-Rad (Watford, Herts., U.K.). Diphenylcarbamyl chloride-treated trypsin (bovine pancreas; EC 3.4.21.4), guanidinium chloride (practical grade) and mixed whale and shark CS were purchased from the Sigma Chemical Co. (Poole, Dorset, U.K.). Chondroitin ABC endolyase (protease-free) (*Proteus ulgaris*; EC 4.2.2.4), keratanase II and chondroitin ACII lyase (*Arthrobacter Aurescens*; EC 4.2.2.5) were obtained from Seikagaku Corp. (Chuo-Ku, Tokyo, Japan). CsCl was purchased from Fluka chemicals (Gillingham, Dorset, U.K.), and NaOH (A. R. 46/48 $\%$ ) was obtained from Fisons Scientific Equipment (Loughborough, Leics., U.K.). All other chemicals were of analytical grade.

#### *Cartilage source*

Each linkage region sample, with one exception, was derived from a single human femoral head. The exception was pooled material (mean age =  $72 \pm 12$  years old, *n* = 7) for which fibromodulin structure has already been reported [31].



#### *Figure 1 HW-40s size-exclusion chromatography of linkage region oligosaccharides*

The linkage region oligosaccharides were isolated for subsequent HPAEC analysis by sizeexclusion chromatography on a column (50 cm  $\times$  1 cm) of Toyapearl HW-40s eluted with 0.5 M ammonium acetate at 0.4 ml/min. The eluate was monitored on-line by measuring the refractive index (continuous line) and absorbance at 232 nm (broken line), and the hexasaccharide linkage regions were pooled as shown. The elution positions of known hexa-, tetra- and di-saccharides (6, 4 and 2 respectively) are shown. The hexasaccharides represent  $> 98\%$  of the CS oligosaccharides present by estimation of 232 nm absorbance.  $V_0$ , void volume;  $V_t$ , total volume.

## *Isolation of linkage region fragments*

The large aggregating proteoglycan, aggrecan, was isolated from human femoral head cartilage by standard methods, and the CS linkage regions were isolated as previously described for bovine and equine materials [11]. Briefly, following extraction of the comminuted cartilage with 4 M guanidinium chloride in the presence of a cocktail of protease inhibitors, the extract was subjected to associative CsCl density gradient centrifugation. CS-rich material (6B2 fraction) was obtained from the A1 fraction by chondroitin ABC endolyase and trypsin digestions followed by size-exclusion chromatography on a column of Sepharose CL-6B as previously described [11,32,33].

The CS linkage regions in the 6B2 fraction were released from the peptide by  $\beta$ -elimination with 1 M sodium borohydride in  $0.05$  M NaOH at 45 °C for 48 h [34], the reaction being terminated by the careful addition of 2 M acetic acid. The released CS oligosaccharides were separated from other materials by sizeexclusion chromatography on a column (50 cm  $\times$  1 cm) of Toyapearl HW-40s eluted with 0.5 M ammonium acetate at  $0.4$  ml/min [9,11,23]. The eluate was monitored by measuring the absorbance at 232 nm and the refractive index. The hexasaccharide-sized CS oligosaccharides (see Figure 1) were pooled and the volatile ammonium acetate was removed by repeated freeze-drying and resuspension in distilled water prior to examination by high-pH anion-exchange chromatography (HPAEC) fingerprinting [9,11,23].

#### *HPAEC*

The reduced linkage region oligosaccharides were examined by HPAEC on a calibrated Dionex chromatography system using a method previously described for CS linkage regions [9,11] and for repeat region di-, tetra- and hexa-saccharides [23,35]. The Carbopac PA1 column  $(4.6 \text{ mm} \times 250 \text{ mm})$  was calibrated with reduced repeat region di- and tetra-saccharides along with linkage region standards [9,11]. The relative abundance of each oligosaccharide identified was determined by estimation of the peak area and the application of a response factor [23,36].

## *NMR Spectroscopy*

Heterogeneous linkage region samples (75–145  $\mu$ g) isolated following HW-40s size-exclusion chromatography were resuspended in 0.5 ml of 99.8  $\frac{8}{2}$   $\text{H}_{2}$ O, buffered to pH 7 with 10 mM sodium phosphate and referenced with sodium 3-trimethylsilyl soutum phosphate and referenced with soutum 3-trimethyishyis<br>[<sup>2</sup>H<sub>4</sub>]propionate as an internal standard. After micro-filtration through 0.45  $\mu$ m nylon filters, samples were freeze-dried and exchanged several times with 0.5 ml of  $99.8\%$   $^{2}H_{2}O$  and once exchanged several times with 0.5 ml of 99.6  $\%$   $H_2O$  and once<br>with 0.5 ml of 99.96  $\%$   ${}^2H_2O$  before final dissolution into 0.7 ml with 0.5 lift of 99.<br>of 99.96%  ${}^{2}H_{2}O$ .

99.90%  $\sim$  - $H_2O$ .<br>High-field <sup>1</sup>H-NMR spectra at 600 MHz were performed at 43 °C using a Varian Unity INOVA spectrometer equipped with a 5 mm probe capable of field-gradient experiments. All chemical shifts are quoted relative to internal sodium 3-trimethylsilyl $[{}^{2}H_{4}]$ propionate at 0.0 p.p.m. The spectra were acquired using a 7000 Hz spectral width and approx. 28672 complex points, and were processed by application of a 0.2 Hz exponential window function and Fourier transformation into 65536 complex points.

The abundance of each form of the repeat region GalNAc present, i.e. 4-, 6- or un-sulphated, was estimated by integration of the responses for H-4 within the terminal 4,5-unsaturated hexuronic acid (∆UA), which is known to be sensitive to the sulphation of the adjacent GalNAc [24,37].

## *RESULTS*

A previously developed CS linkage region fingerprinting method [9,11,23] has been employed for the analysis of the structure of linkage regions from human articular cartilage aggrecan of various ages. The use of chondroitin ABC endolyase to remove the chain cap and repeat region, while the chain remains attached to the protein core, allows the subsequent isolation of hexasaccharide CS linkage regions [23]. A trypsin digest followed by size-exclusion chromatography using CL-6B allows the separation of the CS-rich 6B2 fraction from the KS-rich 6B1 fraction, the void volume  $(V_0)$  fraction containing hyaluronan along with the hyaluronan binding region of aggrecan and from CS di- and tetra-saccharides which elute in the total volume of the column  $(V_t)$ . Following  $\beta$ -elimination to release the CS linkage regions they were easily identified and resolved from



*Figure 2 HPAEC analysis of the CS linkage regions from human articular cartilage aggrecan of a 10-year-old (top panel) and a 72-year-old (bottom panel)*

The CS linkage regions isolated by HW-40s size-exclusion chromatography (see Figure 1) were examined by HPAEC on a CarboPac PA1 column (250 mm  $\times$  4 mm) with a PA1 guard column (50 mm  $\times$  4 mm) maintained at 30 °C and eluted at 1 ml/min. A 12 min isocratic period of 98% eluent A (0.1 M NaOH)/2% eluent B (1.3 M NaCl in 0.1 M NaOH) was followed by a linear gradient of 2-46% eluent B over 50 min; 46-87% eluent B over 8 min; 87-100% eluent B over 6 min; followed by a 4 min isocratic phase of 100% eluent B. The eluted oligosaccharides were monitored on-line by absorbance at 232 nm. \*Denotes a peak resulting from an eluent contaminant. The oligosaccharides are: Hexa-I,  $\Delta$ UA(β1-3)GalNAc(β1-4)GlcA(β1-3)Gal(β1-3)Gal(β1-4)Xyl-ol ; Hexa-II, ∆UA(β1-3)GalNAc*6S*(β1-4)GlcA(β1-3)Gal (β1-3)Gal(β1-4)Xyl-ol ; Hexa-III, ∆UA(β1-3)GalNAc*4S*(β1-4)GlcA(β1-3)Gal(β1-3)Gal(β1- 4)Xyl-ol ; and Hexa-IV, ∆UA(β1-3)GalNAc*6S*(β1-4)GlcA(β1-3)Gal*6S*(β1-3)Gal(β1-4)Xyl-ol.



#### *Figure 3 Partial one-dimensional <sup>1</sup> H 600 MHz NMR spectra of the CS linkage regions from human articular cartilage aggrecan of a 10-year-old and a 72-year-old*

The CS linkage regions isolated by HW-40s size-exclusion chromatography (see Figure 1) were examined by high-field NMR spectroscopy prior to analysis by HPAEC (see Figure 2). The abundance of 4-, 6- and un-sulphated linkage regions was estimated by integration of distinctive signals for H-4 in ∆UA at approx. 5.896 p.p.m. corresponding to an unsulphated linkage region (OS), at 5.886 p.p.m. corresponding to a linkage region containing a 6-sulphated GalNAc residue (6S), both with and without a sulphated Gal, and at 5.966 p.p.m. corresponding to a linkage region containing a 4-sulphated GalNAc residue (4S).

 $\Delta UA(\beta1-3)GalNAc[0S/4S/6S](\beta1-4)GlcA(\beta1-3)Gal[0S/6S](\beta1-3)Gal(\beta1-4)Xyl$ **B A**

#### *Figure 4 General structure of CS linkage region oligosaccharides from human articular cartilage*

other materials by HW-40s size-exclusion chromatography (see Figure 1) and the hexasaccharide linkage region mixture was subsequently examined by HPAEC (see Figure 2) and NMR spectroscopy (see Figure 3).

Four hexasaccharide linkage region structures, with the general structure shown in Figure 4, have been identified from human articular cartilage aggrecan. In each the GalNAc may be either 4-, 6- or un-sulphated and, in one structure, both the Gal towards the non-reducing terminus (Gal **B**) and the GalNAc residue are 6-sulphated. No other variants of Gal sulphation were identified.

Significant age-related changes in the abundance of the linkage regions have been identified, especially in the younger age ranges (Figure 5). In cartilage from 10- to 20-year-olds there was a significant increase in the abundance of GalNAc 6-sulphation (from approx. 43% up to approx. 75%) with a concomitant reduction in the incidence of 4-sulphated (from approx.  $25\%$ down to approx.  $6\%$ ) and unsulphated (from approx.  $30\%$  down to approx.  $20\%$ ) GalNAc residues. Over this age range there is also an increase in the incidence of Gal 6-sulphation (from approx.  $2\%$  up to approx.  $10\%$ ). In each sample the abundance of 4-sulphated GalNAc residues was lower than that of 6- or un-sulphated residues.

Beyond 20 years old the dramatic changes seen during the younger years cease and the data suggest that there are only slight changes in both sulphation levels and profiles. There appear to be only slight changes between 20 and 30 years old; however, during the later years (approx. 40–80 years old) there is a slight decrease in the level of GalNAc 6-sulphation with a concomitant increase in the levels of 4-sulphated residues, but little change in the level of unsulphated linkage regions.

CS linkage regions from the KS-rich portion of aggrecan (6B1 fraction) of selected samples were examined by the methods described for those from the CS-rich region. It was found that



*Figure 5 Age-related changes in the sulphation of CS linkage regions from human articular cartilage aggrecan*

D,∆UA(β1-3)GalNAc(β1-4)GlcA(β1-3)Gal(β1-3)Gal(β1-4)Xyl-ol ;E,∆UA(β1-3)GalNAc*6S-* (β1-4)GlcA(β1-3)Gal(β1-3)Gal(β1-4)Xyl-ol ; y, ∆UA(β1-3)GalNAc*4S*(β1-4)GlcA(β1-3)Gal- (β1-3)Gal(β1-4)Xyl-ol ; ~, ∆UA(β1-3)GalNAc*6S*(β1-4)GlcA(β1-3)Gal*6S*(β1-3)Gal(β1-4)Xylol.

*Table 1 Percentage abundance of linkage regions estimated by NMR and HPAEC*

	Percentage abundance			
	10-Year-old		72-Year-old	
GalNAc sulphation	<b>NMR</b>	HPAFC	NMR	<b>HPAEC</b>
Unsulphated	26.0	31.4	13.5	14 9
4-Sulphated	30.0	25.3	14.0	14.2
6-Sulphated	44.0	43.3	72.5	70.9

the linkage region sulphation profile did not differ from that seen following analysis of linkage regions from the CS-rich region (6B2 fraction) of the same aggrecan sample (results not shown).

We have examined the one-dimensional <sup>1</sup>H-NMR spectra of the heterogeneous mixtures of linkage regions, and partial spectra of samples from a 10-year-old and a 72-year-old may be seen in Figure 3. We have previously characterized the "H-NMR spectra for the octa-, hexa- and tetra-saccharide linkage regions of CS from articular cartilage aggrecan [11,24], and reported that the signals from H-1 and H-4 of the non-reducing terminal ∆UA were differently perturbed dependent upon the sulphation status of the adjacent GalNAc residue. Comparison of the onedimensional spectra obtained from human samples with our previous data [24] shows that the abundance of linkage regions in which the GalNAc is 6-, 4- or un-sulphated may be estimated by examination of H-4 signals from the adjacent non-reducing terminal ∆UA. The abundance of each structure was estimated by integration of H-4 signals from the terminal ∆UA and the data were found to be in close agreement (Table 1) with the data derived from HPAEC analysis (Figure 2) of the same samples.

## *DISCUSSION*

We have used an HPAEC fingerprinting method [9,11,23] for the analysis of CS linkage region hexasaccharides from human articular cartilage aggrecan of various ages. Linkage region structures have been identified (Figure 4) in which the GalNAc residue may be either 4-, 6- or un-sulphated; in addition, Gal **B** 6-sulphation, always found in conjunction with GalNAc 6-sulphation, has been identified.

It has been shown that there are significant age-related changes in the abundance of each linkage region identified (Figure 5). These dramatic changes occur primarily during the period up to 20 years old, and both the level and profile of sulphation of the CS linkage regions change over this period. There is a significant increase in the abundance of GalNAc 6-sulphation, with a concomitant decrease in the levels of both 4- and un-sulphated residues.

The total sulphation level (abundance of 4- plus 6-sulphated GalNAc residues) increases from approx. 70 $\%$  at 10 years old to approx. 80–90 $\%$  at 20 years old, although through this period the level of 4-sulphation falls. These data are in agreement with those of Cheng et al. [13] who found that this residue was sulphated in almost  $100\%$  of chains from old human articular cartilage. The level of GalNAc 6-sulphation for older humans compares well with data from both bovine and equine samples [11].

The sharp rise in GalNAc 6-sulphation at the linkage region slows at approx. 20 years old (Figure 5). Structural changes in other cartilage GAGs have also been found to occur primarily during the period from birth to 20 years old, with a dramatic slowing in the rate of change after this age. Examples of structures which undergo such changes include CS cap structure [25,26] and repeat region sulphation [27], and KS cap, linkage [28,29] and repeat region sulphation, fucosylation and chain length [29]. The pattern of age-related changes seen at the linkage region in the present study, i.e. an increase in GalNAc 6-sulphation, is also observed within the repeat region [27] and at the chain caps [25,26].

No data are available for linkage region sulphation at ages below 10 years old; however, the sharp rise in GalNAc 6 sulphation seen between 10 and 20 years old suggests that at younger ages there may be an even lower level of GalNAc 6 sulphation. This would be in accord with age-related changes in both the repeat region [27] and chain caps [25,26], which both show a low level of GalNAc 6-sulphation that increases from birth to approx. 20 years old.

The data suggest that between 20 years old and approx. 40 years old there are few significant changes in either the total level of sulphation or distribution between 4-sulphation and 6-sulphation; however, during the later years, approx. 40–80 years old, there is a slight decrease in the level of GalNAc 6-sulphation with a concomitant increase in the levels of 4-sulphated residues; there is little change in the abundance of unsulphated GalNAc residues. Further data are required to determine if this change is widely observed and reflects normal aging, or is an early response to a pathological condition such as osteoarthritis. Plaas et al. [25,26] have reported that in osteoarthritis the abundance of 4,6 disulphated CS chain caps falls with a concomitant increase in 4-sulphated residues. It is noteworthy that in the linkage region there is no significant change in unsulphated residues, which are abundant in young tissues.

Comparison of the level of linkage region GalNAc 6-sulphation observed at each age with the data of Bayliss et al. [27] for the levels of GalNAc 6-sulphation within the repeat region shows that the level of linkage region GalNAc 6-sulphation is always lower than the average level of GalNAc 6-sulphation along the chain. This preferential localization of 4- and unsulphated GalNAc residues close to the linkage region, at all ages [11], is in sharp contrast with the highly sulphated chain caps [25,26]. It is clear that the sulphation profile of the linkage region

of CS chains from articular cartilage aggrecan of human, as well as bovine and equine, origin does not reflect the composition of the remainder of the chains.

In human articular cartilage CS linkage region Gal **B** may be 6-sulphated, but only in conjunction with 6-sulphation of the GalNAc residue. Furthermore, we have shown that the abundance of this novel structure [11] increases with age.

There was no evidence of any uronic acid sulphation close to the linkage region at any age, in agreement with data for adult bovine and equine articular cartilage aggrecan [11], along with other studies of CS linkage region structure ([16,17] and see http://www.glycoforum.gr.jp/science/word/proteoglycan/ PGA06E.html), none of which report the observation of a sulphated uronic acid close to the linkage region of CS. However, in heparin and HS a sulphated uronic acid has been observed as the non-reducing terminal residue of linkage region oligosaccharides, which are of octasaccharide size or larger [38].

The sulphation pattern of the CS linkage regions from the CSrich region of aggrecan (6B2 fraction) and the KS-rich region (6B1 fraction) were examined in selected samples. There were no differences in the CS linkage regions of these two regions of aggrecan. These data suggest that there is no variation in the sulphation of CS linkage regions at different locations along the aggrecan monomer.

Data from the present study, and elsewhere [11], show that the linkage region of articular cartilage CS has a lower than average level of sulphation, whereas the chain cap has been shown to have a high level of sulphation [25,26]. KS from cornea [39] along with that from articular and tracheal cartilage fibromodulin [31,40–42] has also been shown to have levels of sulphation which are lower at the linkage region than towards the chain cap.

Many of the reported properties of CS, such as the cytoadherence of malaria-infected red blood cells [2], the adherence of the malaria parasite to human placenta [1,3,4] and the regulation of neurite outgrowth [5], involve GalNAc residues that are 4-sulphated. Indeed 6-sulphated GalNAc residues have been shown to abolish the ability of CS to inhibit malaria parasite binding to human placenta [3,4]. Thus the region of the CS chain which is closest to the linkage region, and which has a lower level of GalNAc 6-sulphation, may be able to participate in interactions and have properties and functions which differ from the rest of the chain. In addition, with aging, as the level of GalNAc 6-sulphation increases the extent of such interactions would change. The observation of altered sulphation at the linkage region has also been reported for HS [43], heparin [44] and KS [40].

In addition to the estimation of the abundance of each linkage region by HPAEC (Figure 2), selected linkage region mixtures have been examined using high-field NMR spectroscopy (Figure 3) and may be compared with the data from our previous NMR study of pure isolated linkage regions [24]. The data derived from NMR spectroscopy were found to be in close agreement (Table 1) with data from HPAEC analysis (Figure 2). This agreement confirms the validity of each method and demonstrates that NMR spectroscopy, although requiring greater amounts of material ( $>$  approx. 75  $\mu$ g), is useful in the non-destructive estimation of the sulphation pattern for the GalNAc residues close to the CS linkage regions. NMR spectroscopic approaches may allow the characterization of the abundance of discrete structures within a mixed population [45].

In conclusion, it is shown that the pattern of sulphation close to the linkage region of human articular cartilage aggrecan CS chains differs markedly with age. With increasing age, up to approx. 20 years old, there is an increase in the relative abundance of 6-sulphated GalNAc residues and a concomitant reduction in both 4- and un-sulphated residues. There is also an increase in the abundance of Gal 6-sulphation. Beyond 20 years old there is a slight reduction in the abundance of 6-sulphated GalNAc. This analysis of the normal age-related changes in sulphation of CS linkage regions will allow the interpretation of data from pathological samples.

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# *REFERENCES*

- 1 Fried, M., Lauder, R. M. and Duffy, P. E. (2000) *Plasmodium falciparum*: Adhesion of placental isolates modulated by the sulfation characteristics of the glycosaminoglycan receptor. Exp. Parasitol. *95*, 75–78
- 2 Cooke, B. M., Rogerson, S. J., Brown, G. V. and Coppel, R. L. (1996) Adhesion of malaria-infected red blood cells to chondroitin sulfate A under flow conditions. Blood *88*, 4040–4044
- 3 Fried, M. and Duffy, P. E. (1998) Maternal malaria and parasite adhesion. J. Mol. Med. *76*, 162–171
- 4 Fried, M., Lauder, R. M. and Duffy, P. E. (1998) Minimal chondroitin sulphate structures required to support *Plasmodium falciparum* adhesion. Glycobiology *8*, 1021
- 5 Margolis, R. U. and Margolis, R. K. (1997) Chondroitin sulfate proteoglycans as mediators of axon growth and pathfinding. Cell Tissue Res. *290*, 343–348
- 6 Nadanaka, S., Clement, A., Masayama, K., Faissner, A. and Sugahara, K. (1998) Characteristic hexasaccharide sequences in octasaccharides derived from shark cartilage chondroitin sulfate D with a neurite outgrowth promoting activity. J. Biol. Chem. *273*, 3296–3307
- 7 Mark, M. P., Baker, J. R., Kimata, K. and Roth, J.-V. (1990) Regulated changes in chondroitin sulfation during embryogenesis: an immunohistochemical approach. Int. J. Dev. Biol. *34*, 191–204
- 8 Herndon, M. E. and Lander, A. D. (1990) A diverse set of developmentally regulated proteoglycans is expressed in the rat central-nervous-system. Neuron *4*, 949–961
- 9 Lauder, R. M., Huckerby, T. N. and Nieduszynski, I. A. (2000) A high pH anion exchange chromatography fingerprinting method for chondroitin/dermatan sulphate and hyaluronan oligosaccharides. Glycobiology *10*, 393–401
- 10 Roden, L. (1980) Structure and metabolism of connective tissue proteoglycans. In The Biochemistry of Glycoproteins and Proteoglycans (Lennarz, W. J., ed.), pp. 267–371, Plenum Press, New York
- 11 Lauder, R. M., Huckerby, T. N. and Nieduszynski, I. A. (2000) Increased incidence of unsulphated and 4-sulphated residues in the chondroitin sulphate linkage region observed by high-pH anion-exchange chromatography. Biochem. J. *347*, 339–348
- 12 de Beer, T., Inui, A., Tsuda, H., Sugahara, K. and Vliegenthart, J. F. G. (1996) Polydispersity in sulfation profile of oligosaccharide-alditols isolated from the proteinlinkage region and the repeating disaccharide region of chondroitin 4-sulfate of bovine nasal septal cartilage. Eur. J. Biochem. *240*, 789–797
- 13 Cheng, F., Heinegard, D., Fransson, L. Å., Bayliss, M., Bielicki, J., Hopwood, J. and Yoshida, K. (1996) Variations in the chondroitin sulfate-protein linkage region of aggrecans from bovine nasal and human articular cartilages. J. Biol. Chem. *271*, 28572–28580
- 14 Shibata, S., Midura, R. J. and Hascall, V. C. (1992) Structural analysis of the linkage region oligosaccharides and unsaturated disaccharides from chondroitin sulfate using CarboPac PA1. J. Biol. Chem. *267*, 6548–6555
- 15 Sugahara, K., Masuda, M., Harada, T., Yamashina, I., de Waard, P. and Vliegenthart, J. F. G. (1991) Structural studies on sulfated oligosaccharides derived from the carbohydrate-protein linkage region of chondroitin sulfate proteoglycans of whale cartilage. Eur. J. Biochem. *202*, 805–811
- 16 de Waard, P., Vliegenthart, J. F. G., Harada, T. and Sugahara, K. (1992) Structural studies on sulfated oligosaccharides derived from the carbohydrate-protein linkage region of chondroitin-6-sulfate proteoglycans of shark cartilage. II. 7 compounds containing 2 or 3 sulfate residues. J. Biol. Chem. *267*, 6036–6043
- 17 Sugahara, K., Ohi, Y., Harada, T., de Waard, P. and Vliegenthart, J. F. G. (1992) Structural studies on sulfated oligosaccharides derived from the carbohydrate-protein linkage region of chondroitin 6-sulfate proteoglycans of shark cartilage. I. Six compounds containing 0 or 1 sulfate and/or phosphate residues. J. Biol. Chem. *267*, 6027–6035
- 18 Reference deleted
- 19 Doege, K., Chen, X. C., Cornuet, P. K. and Hassell, J. (1997) Non-glycosaminoglycan bearing domains of perlecan and aggrecan influence the utilization of sites for heparan and chondroitin sulfate synthesis. Matrix Biol. *16*, 211–221
- 20 Zhang, L. J. and Esko, J. D. (1994) Amino acid determinants that drive heparan sulfate assembly in a proteoglycan. J. Biol. Chem. *269*, 19295–19299
- Fritz, T. A., Lugemwa, F. N., Sarkar, A. K. and Esko, J. D. (1994) Biosynthesis of heparan sulfate on beta-D-xylosides depends on aglycone structure. J. Biol. Chem. *269*, 300–307
- 22 Sarkar, A. K. and Esko, J. D. (1995) Synthesis and glycosaminoglycan priming activity of three disaccharides related to the linkage region tetrasaccharide of proteoglycans. Carbohydr. Res. *279*, 161–171
- 23 Lauder, R. M., Huckerby, T. N. and Nieduszynski, I. A. (1999) Isolation and identification of an octasaccharide chondroitin sulphate linkage region fragment. Carbohydr. Lett. *3*, 381–388
- 24 Huckerby, T. N., Lauder, R. M. and Nieduszynski, I. A. (1998) Structural determination for octasaccharides derived from the carbohydrate-protein linkage region of chondroitin sulphate in aggrecan from bovine articular cartilage. Eur. J. Biochem. *258*, 669–676
- 25 Plaas, A. H. K., Wong-Palms, S., Roughley, P. J., Midura, R. J. and Hascall, V. C. (1997) Chemical and immunological assay of the nonreducing terminal residues of chondroitin sulfate from human aggrecan. J. Biol. Chem. *272*, 20603–20610
- 26 Plaas, A. H. K., West, L. A., WongPalms, S. and Nelson, F. R. T. (1998) Glycosaminoglycan sulfation in human osteoarthritis. Disease-related alterations at the non-reducing termini of chondroitin and dermatan sulfate. J. Biol. Chem. *273*, 12642–12649
- 27 Bayliss, M. T., Osborne, D., Woodhouse, S. and Davidson, C. (1999) Sulfation of chondroitin sulfate in human articular cartilage. The effect of age, topographical position and zone of cartilage on tissue composition. J. Biol. Chem. *274*, 15892–15900
- 28 Lauder, R. M., Huckerby, T. N., Nieduszynski, I. A. and Plaas, A. H. K. (1998) Agerelated changes in the structure of the keratan sulphate chains attached to fibromodulin isolated from bovine articular cartilage. Biochem. J. *330*, 753–757
- Brown, G. M., Huckerby, T. N., Bayliss, M. T. and Nieduszynski, I. A. (1998) Human aggrecan keratan sulfate undergoes structural changes during adolescent development. J. Biol. Chem. *273*, 26408–26414
- 30 Shinmei, M., Miyauchi, S., Machida, A. and Miyazaki, K. (1992) Quantitation of chondroitin 4-sulfate and chondroitin 6-sulfate in pathologic joint fluid. Arthritis Rheum. *35*, 1304–1308
- 31 Lauder, R. M., Huckerby, T. N. and Nieduszynski, I. A. (1997) The structure of keratan sulphate chains from fibromodulin of human articular cartilage. Glycoconjugate J. *14*, 651–660
- 32 Heinegård, D. and Axelsson, I. (1977) Distribution of keratan sulfate in cartilage proteoglycans. J. Biol. Chem. *252*, 1971–1979

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- 33 Thornton, D. J., Morris, H. G., Cockin, G. H., Huckerby, T. N., Nieduszynski, I. A., Carlstedt, I., Hardingham, T. E. and Ratcliffe, A. (1989) Structural and immunological studies of keratan sulphates from mature bovine articular cartilage. Biochem. J. *260*, 277–282
- 34 Carlson, D. M. (1968) Structures and immunochemical properties of oligosaccharides isolated from pig submaxillary mucins. J. Biol. Chem. *243*, 616–626
- Lauder, R. M. (2000) Analysis of proteoglycans and glycosaminoglycans. In Encyclopaedia of Analytical Chemistry (Myers, R. A. and Holyoake, A., eds.), pp. 860–895, John Wiley, London
- 36 Yamagata, T., Saito, H., Habuchi, O. and Suzuki, S. (1968) Purification and properties of bacterial chondroitinases and chondrosulfatases. J. Biol. Chem. *243*, 1523–1535
- 37 Huckerby, T. N., Lauder, R. M., Brown, G. M., Nieduszynski, I. A., Anderson, K., Boocock, J. B., Sandell, P. L. and Weeks, S. D. (2001) Characterisation of oligosaccharides from the chondroitin sulphates: <sup>1</sup>H and <sup>13</sup>C-NMR studies of diand tetrasaccharides. Eur. J. Biochem. *268*, 1181–1189
- Sugahara, K., Tsuda, H., Yoshida, K., Yamada, S., de Beer, T. and Vliegenthart, J. F. G. (1995) Structure determination of the octa- and decasaccharide sequences isolated from the carbohydrate-protein linkage region of porcine intestinal heparin. J. Biol. Chem. *270*, 22914–22923
- 39 Oeben, M., Keller, R., Stuhlsatz, H. W. and Greiling, H. (1987) Constant and variable domains of different disaccharide structure in corneal keratan sulphate chains. Biochem. J. *248*, 85–93
- Lauder, R. M., Huckerby, T. N. and Nieduszynski, I. A. (1994) Structure of the keratan sulphate chains attached to fibromodulin isolated from bovine tracheal cartilage. Oligosaccharides generated by keratanase digestion. Biochem. J. *302*, 417–423
- 41 Lauder, R. M., Huckerby, T. N. and Nieduszynski, I. A. (1995) The structure of the keratan sulphate attached to fibromodulin isolated from bovine tracheal cartilage. Oligosaccharides isolated following keratanase II digestion. Glycoconjugate J. *12*, 651–659
- 42 Lauder, R. M., Huckerby, T. N. and Nieduszynski, I. A. (1996) The structure of the keratan sulphate attached to fibromodulin isolated from bovine articular cartilage. Eur. J. Biochem. *242*, 402–409
- 43 Lyon, M., Deakin, J. A. and Gallagher, J. T. (1994) Liver heparan sulfate structure. A novel molecular design. J. Biol. Chem. *269*, 11208–11215
- Sugahara, K., Yamada, S., Yoshida, K., de Waard, P. and Vliegenthart, J. F. (1992) A novel sulfated structure in the carbohydrate-protein linkage region isolated from porcine intestinal heparin. J. Biol. Chem *267*, 1528–1533
- 45 Huckerby, T. N. and Lauder, R. M. (2000) Keratan sulphates from bovine tracheal cartilage. Structural studies of intact polymer chains using <sup>1</sup>H and <sup>13</sup>C-NMR spectroscopy. Eur. J. Biochem. *276*, 3360–3369