Supporting information for Etz et al. (May 7, 2002) Proc. Natl. Acad. Sci. USA,

10.1073/pnas.092569199.

ORF number	Name, putative function or homology	Localization motif*	Gene distribution: positive/total [†]	Hits per \mathbf{ORF}^{\ddagger}				Location of antigenic region	Reactivity with sera:
(TIGK)				LamB		FhuA		-	(MACS
				Patient	Healthy	Patient	Healthy	-	recovery) [§]
SA0095	Protein A	SP, LPXTG	30/30	63	-	53	36	aa 1–48	20/28
								aa 47–143	1/1
								aa 219–285	17/28
SA0209	Coomilase	SD	30/30				26	aa 345–424	3///1
	Coaguiase	51	30/30	_	_	_	20	aa 436-510 aa 505-570	3/3 1/7
								aa 569–619	2/7
SA0317	Lipase	SP	Variable (2)	45	_	42	3	aa 48–136	6/13
	1							aa 128–172	10/28 (100%)
								aa 201–258	7/28
SA0470	Putative Exotoxin 2	SP	29/30	-	3	-	101	aa 1–80	6/7
								aa 49–116	25/28
~								aa 98–190	13/17
SA0507	LysM domain	SP	30/30	-	-	2	1	aa 21–118	50/52
П	protein	CD	20/20			50	10	45 105	1 /1
SA0723	LysM domain	SP	30/30	-	-	38	18	aa 45 - 105	1/1
	with \$42581							aa 105-100 aa 66-153	13/27
SA0858	Emphy extracellular	SP	21/30	27	2	50	15	aa 22–56	58/71
5110050	matrix and plasma-	51	21/50	27	-	50	15	aa 23–99	59/59
	binding protein							aa 97–115	1/1
	01							aa 233–250	ND
								aa 245–265	ND
SA1062	Atl, autolysin	SP	29/30	47	5	22	61	aa 6–66	5/16
								aa 65–124	15/28
								aa 590–604	39/71
SA1472	Pathogenicity protein	SP	20/20	-	-	10	30	aa 8511–8640	5/26 (100%)
SA1781	LPXTGp5	SP, LPX1G	30/30	22	3	1	-	aa 37–49	1/1
								aa 63 - 1/1	12/70
\$42004	Putative leukocidin F					9		aa 274-334 aa 158-220	4/28
5A2004	precursor					,		aa 150–220	//2/(100/0)
SA2006	Putative aerolysin	SP	30/30	3	4	12	36	aa 31–61	19/27
								aa 58–74	49/71
SA2019	Putative SdrH	SP	30/30	7	9	_	_	aa 122–139	ND
								aa 164–182	39/71
SA2291	Homology with	SP	30/30	58	42	224	169	aa 137–237	13/27
	SA2581							aa 250–267	10/71
SA2295	Homology with	SP		4	-	5	-	aa 38–52	ND
	SA2581	CD	070((2)	11	22	1		aa 66–114	4/28
SA2418	Sbi, IgG-binding	SP	97% (3)	11	23	1	-	aa 208–287	38/46
\$42505	I PXTGp4	SP I PYTG			18	4	3	aa 200-514 aa 491-585	1//1
5A2505	LI XI Op4	51, LIXIO		_	10	-	5	aa 631–713	1/1
								aa 702–758	15/28
SA2509	FnbpB, Fibronection	SP	100% (4)	_	2	2	5	aa 693–769	26/26
	binding							aa 775–814	1/1
SA2511	FnbpA, Fibronection	SP	100% (4)	-	2	4	4	aa 710–787	19/25
	binding							aa 855–975	ND
		_						aa 916–983	15/28
SA2581	Staphyloxanthin	SP	26/30	51	43	130	13	aa 7–87	1/1
	biosynthesis protein,							aa 133–242	5/27 (100%)
	nomolog of								
	epidermidis SsaA								

Table 1. Immunogenic proteins frequently identified by bacterial surface display

1

ORF number (TIGR)	Name, putative function or homology	Localization motif*	Gene distribution: positive/total [†]	Hits per ORF [‡]				Location of R antigenic region w	Reactivity n with sera:
				LamB		FhuA		-	(MACS
				Patient	Healthy	Patient	Healthy	-	recovery) [§]
SA2584	IsaA	SP	30/30	69	2	8	3	aa 67–116 aa 98–184 aa 182–225	1/1 20/26 34/71
SA2659 SAA0001	Aureolysin repC	SP	100% (5)	_	- 6	1	6 18	aa 83–156 aa 9–42 aa 158–174	13/27 1/1 1/1

Screens of two libraries with two serum pools each are summarized: LSA250 library in FhuA with patient serum P1 (655 clones analyzed) and with serum from healthy individuals N1 (680); LSA50 library in LamB with patient sera P1 (498) and with serum from healthy individuals N2 (900). Only a selection of frequently identified antigens is shown. *, Motifs characteristic of secreted or surface exposed proteins: SP, signal peptide; LPXTG, sortase anchoring motif; †, PCR analysis was performed with oligonucleotides designed to amplify approximately 500- to 1,000-bp fragments covering at least one antigenic epitope. Twenty-four *Staphylococcus aureus* strains were isolated from patients and three strains from healthy individuals and typed according to restriction fragment length polymorphism (RFLP) of the spa and coa genes (1). The three laboratory strains, COL, Newman, and 8325-4, were used as controls. PCR products of variable size were obtained for the SA2019, SA0095, and SA0209 genes. Sequencing of the PCR products revealed that the size difference was due to a variable copy number of a repetitive DNA sequence in these three genes. ‡, The number of clones identified per ORF is given for the individual screens. Representative clones were analyzed further. §, One epitope of the indicated region was chosen for Western blot analysis with individual sera. Several epitopes identified frequently were not or were only very weakly reactive in Western blot analysis indicating the presence of conformational epitopes. These clones were therefore tested in MACS experiments where the rate of specific recovery with biotinylated patient serum P1 is expressed as percentage of the input

(shown in brackets). ¶, Identified 18 times of 33 in a FhuA prescreen with serum from healthy individuals N1, and was therefore eliminated from the library prior to screen FhuA with N1. ND, not determined; MACS, magnetic cell sorting.

References

- VandenBergh, M. F., Yzerman, E. P., van Belkum, A., Boelens, H. A., Sijmons, M. & Verbrugh, H. A. (1999) J. Clin. Microbiol. 37, 3133–3140.
- 2. Lee, C. Y. & Iandolo, J. J. (1986) J. Bacteriol. 166, 385–391.
- Zhang, L., Jacobsson, K., Ström, K., Lindberg, M. & Frykberg, L. (1999) *Microbiology* 145, 177–183.
- Smeltzer, M. S., Gillaspy, A. F., Pratt, F. L., Jr., Thames, M. D. & Iandolo, J. I. (1997) *Gene* 196, 249–259.
- Sabat, A., Kosowska, K., Poulsen, K., Kasprowicz, A., Sekowska, A., van Den Burg, B., Travis, J. & Potempa, J. (2000) *Infect. Immun.* 68, 973–976.