# Protein kinase CK2: A new view of an old molecular complex

Odile Filhol, Jean-Louis Martiel and Claude Cochet

## Supplementary data.

### Model assumptions and equations.

In the context of cellular signaling, it has been shown that the recruitment of proteins to the cell membrane increases the number of molecular complexes made with membrane-associated substrates (Kholodenko et al., 2000) (see also Haugh, 2002 for a full account of the biophysical model.) Using a similar approach, we show here, that the interaction of CK2 with its partners, including the dimeric CK2  $_2$  regulatory subunit or other multi-molecular targets C, leads to a large difference in the number of complexes CK2  $_2$  and CK2 C depending on the biophysical conditions prevailing in the cell.

Let A be the CK2 subunit, B<sub>2</sub> the dimeric CK2 <sub>2</sub> regulatory subunit and C a multimolecular complex. To reconcile *in vivo* and *in vitro* observations, we assume that:

1. interaction between CK2 and CK2  $_2$  subunits can take place everywhere in the cell bulk (Martel et al., 2002) (volume  $V_c$ .). At equilibrium, the law of mass action gives:

$$K = \frac{\left[A\right]^2 \left[B_2\right]}{\left[A_2 B_2\right]}$$

with  $K \approx 29.2 (\text{nM})^2$ .

2. CK2 can bind to the complex C only in a sub-compartment of the cell, which corresponds to the volume  $V_m$ . At equilibrium, concentrations are related to the dissociation constant  $K_O$  ('O' for 'other' CK2 partners) by:

$$K_{O} = \frac{\begin{bmatrix} A_{m} \end{bmatrix} \begin{bmatrix} C \end{bmatrix}}{\begin{bmatrix} A_{m} C \end{bmatrix}}$$

with  $K_0$  larger than  $\sqrt{K}$ . A<sub>m</sub> represents the CK2 subunit in the sub-compartment where binding occurs.

3. the CK2 subunit is at equilibrium between the different cell compartments:

$$\begin{bmatrix} A \end{bmatrix} = \begin{bmatrix} A_m \end{bmatrix}$$

In cells, about 20 to 30% of the total volume is occupied by large macromolecules, which accounts for a reduction of the diffusion coefficients for proteins by factors ranging from 3 to 11 (Ellis, 2001). Therefore, we assume that the number of CK2 subunits exchanged between the bulk and the sub-compartment are negligible.

4. the total number of the different molecular species is conserved according to

$$P_T = 2B_2 + 2A_2B_2$$
$$P_{c,T} = A + 2A_2B_2$$
$$P_{m,T} = A_m + A_mC_m$$
$$P_T = P_{c,T} + P_{m,T}$$
$$C_T = C + A_mC_m$$

where a symbol in italics designates the number of molecules of the corresponding chemical species. The first relation codes for conservation of the CK2 <sub>2</sub> complex throughout the cell. Because we assume zero flux between the bulk and the sub-compartment, the total number of CK2 subunits is constant in both compartments (second and third equations.) The fourth relation ensures the conservation of the total CK2 subunits. The conservation of the complex C is given in the final equation.

To compare the relative importance of the CK2 binding to CK2 <sub>2</sub> or to other binding patners, we use number of molecules instead of concentration. The equilibrium relations give:

$$A_2B_2 = \frac{A^2 \cdot B_2}{(V_c)^2 K}$$
$$A_mC = \frac{A_m \cdot C}{V_m K_o}$$
$$A_m = \frac{V_m}{V_c} A$$

Using the conservation relations and the equilibrium conditions, the number of CK2 subunits in both compartments is obtained as solution of two simultaneous equations:

(a) 
$$x + \frac{x^2}{\theta_1 + x^2} - \frac{V_c}{V_c + V_m} = 0$$

Eq. (1)

(b) 
$$y + \theta_2 \frac{y}{\theta_3 + y} - \frac{V_m}{V_c + V_m} = 0$$

where  $x = \frac{A}{P_T}$ ,  $y = \frac{A_m}{P_T}$ ,  $\theta_1 = K \frac{V_c}{P_T}^2$ ,  $\theta_2 = \frac{C_T}{P_T}$ ,  $\theta_3 = \frac{V_m K_o}{P_T}$ . Note that all the variables or

parameters are in dimensionless form. The complexes normalized number of complexes  $A_2B_2$ and  $A_mC$  are given by

$$\frac{A_2B_2}{P_T} = \frac{x^2}{\theta_1 + x^2}$$
$$\frac{A_mC}{P_T} = \theta_2 \frac{y}{\theta_3 + y}$$

where x and y are the unique solution of equation (1a,b) in the interval [0,1].

#### Model predictions.

The figure 3 (main text) is obtained for the following parameters:  $K=29.2(\text{nM})^2$ ,  $V_c=4.2\text{pl}$  (volume of a sphere of 10µm) and  $K_o = 100\sqrt{K}$ ;  $P_T = 100 V_c \sqrt{K}$ ,  $C_T = 100 V_c \sqrt{K}$ .  $V_m$ , the volume of the sub compartment, ranges from 4.2fl to 4.2pl. As the ratio  $V_c/V_m$  decreases ( $V_c = V_m$  in vitro), binding of CK2 to its two partners (CK2 <sub>2</sub> or C) favors the high affinity complex CK2 <sub>2 2</sub>. Conversely, when CK2 binding to its low-affinity partner is confined to a restricted compartment (large  $V_c/V_m$ ), equilibrium is shifted to the complex CK2 C. Equal amount of CK2 <sub>2 2</sub> and CK2 C (i.e. r=1) is obtained for  $V_c/V_m \approx 30$ .

#### **Figure legend**

Figure S1 shows how the ratio function, as defined above, changes with  $\theta = \frac{K_o}{\sqrt{K}}$ , the relative affinity of CK2 to its two partners. When  $\theta$  is in the range [1,10000] and for large  $V_c/V_m$ , CK2 is mainly engaged with its low-affinity partner, whatever the value of  $K_o$ . Conversely, at low  $V_c/V_m$ , CK2 <sub>2 2</sub> is predominant and represents the unique observable complex. These results support our conclusions and show the robustness of the prediction illustrated in figure 3 (main text). The different curves were obtained for  $K_o = 2.78\sqrt{K}$  (curve a) to

 $K_o = 10000\sqrt{K}$  (curve e); the data shown in figure 3 (main text) corresponds to curve (d) with  $K_o = 100\sqrt{K}$ . Other parameters are those of the figure 3 (main text).

To demonstrate the robustness of the behaviour shown in figure 3 (main text), we have determined the boundary between the domains where CK2  $_2 _2 < CK2$  C or CK2  $_2 _2 > CK2$  C by solving equation 1 supplemented with the constraint r=1 (i.e.  $A_2B_2=A_mC$ ) for different values of the parameters ( $V_c/V_m$ , ). Other parameters are those of figure 3 (main text). A limited space for the low-affinity reaction, representing 10% of the total bulk volume, results into an eighty-fold apparent decrease of the low-affinity dissociation constant. A further reaction confinement (5% and below) has a more drastic effect on the apparent dissociation constant which can counterpoise the tight association between CK2 and CK2 \_2.

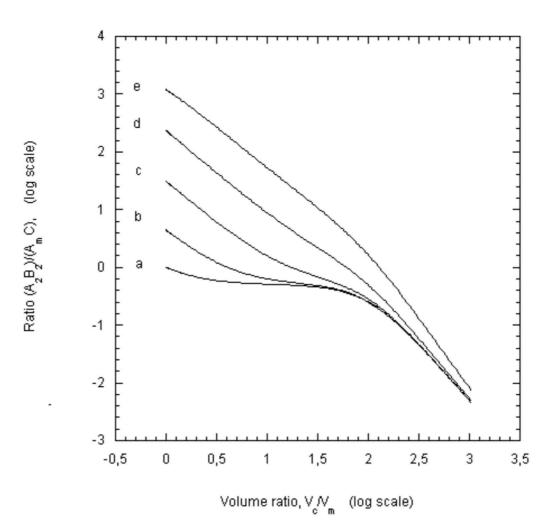


Figure S1

## References

Ellis, R.J., (2001) Macromolecular crowding: an important but neglected aspect of the intracellular environment. Curr. Opin. Struct. Biol. **11**, 114-119.

Haugh, J.M. (2002) A unified model for signal transduction reactions in cellular membranes.

Biophys. J. 82, 591-604.

Kholodenko, B.N., J.B. Hoek, & Westerhoff H.V. (2000) Why cytoplasmic signalling proteins should be recruited to cell membranes. Trends in Cell Biology **10**, 173-178.

Martel, V., Filhol O., Nueda A., & Cochet C. (2002) Dynamic localization/Association of Protein Kinase CK2 subunits in living cells: a role in its cellular regulation? Ann. N.Y. Acad. Sc. **973**, 272-277.