

## Supplementary Data

### Methods

**Plasmid construction** was done by ET cloning using homologous recombination in *E. coli* (Zhang et al., 1998) and standard cloning procedure. For the generation of APP fusion construct, a DNA fragment coding for amino acids 2-87 of LexA and amino acids 410-489 of VP16 (LV) together with a chloramphenicol resistance gene was amplified by PCR with primers containing homologies to the C-terminus of APP and its 3'UTR. To ensure that the released IC is translocated to the nucleus, we inserted the NLS from the SV 40 T-antigen between APP and LV. This fragment was inserted into a cDNA of APP695 or SPA4CT-DA (Lichtenthaler et al., 1999), fusing LV directly to the coding sequence of APP.

The Notch constructs were made by fusing amino acids 410-489 of VP16 to N<sup>LexA</sup> and N<sup>ΔEGF1-36-LexA</sup> (Kidd et al., 1998) again using recombination and selection for chloramphenicol resistance. For generation of N<sup>S2ΔCT/LV</sup>, the remaining extracellular domain until amino acids 1725 and the intracellular domain starting from amino acids 1771 of N<sup>ΔEGF/LV</sup> were deleted by recombination. After restriction enzyme mediated excision of the selection marker, all fusion constructs were subcloned into pUAST.

The reporter LexA-hrGFP was made by replacing the LacZ gene in LexA-lacZ (Kidd et al., 1998) with the hrGFP gene amplified from the vector phrGFP-C (Stratagene). pWIZ-PS/RNAi was made by cloning a PCR amplified fragment containing nucleotides 1281-1781 of the *Drosophila* PS cDNA into the pWIZ vector as suggested by Lee and Carthew (2003).

**Primer sequences**

*Italic* characters represent nucleotides homologues to APP or Notch, underlined characters the PCR primer and **bold** characters the inserted NLS.

APP<sup>LV</sup> and SPA4CT<sup>LV</sup>

up:

*GGCTACGAAAATCCAACCTACAAGTTCTTTGAGCAGATGCAGAACCCTC****CAAAAAA***  
***GAAGAGAAAGGTAATGAAAGCGTTAACGGCCAGGCAACAAG***

down:

*CCGATGGGTAGTGAAGCAATGGTTTTGCTGTCCAACCTCAGAGGCTGCTGTTACGC*  
*CCCGCCCTGCCACTCATC*

N<sup>LV</sup> and N<sup>ΔEGF/LV</sup>

up:

*GTTTGTCCAATTATGTCACACCACAGAAGTAAGGTTCTTCACAAAGATCGGCGCG*  
*CCTTACGCCCCGCCCCTGCCACTCATC*

down:

*AAGAGGAAGAAGGGTTGCCGCTGGTAGGTCGTGTGGCTGCCGGTGAACCTCTGTC*  
*GACGGCCCCCCCGAC*

N<sup>S2ΔCT/LV</sup>

Deletion of the EC

up:

*GCCGGCGGCTCACCATTGTCCTCATCCCCGGGATTCTTGATGCCCCTAACCGCAA*  
*ACGCCAGCGTCAACAGCAG*

down:

CTGAATAGGGAATTGGGAATTCGTTAACAGATCTGCGGCCGCGGTCTAGAGGTAC  
CTTACGCCCCGCCCTGCCACTCATC

Deletion of N<sup>ICD</sup>

up:

GCATTGGCCTTCTTTGGCATGGTCTTGAGTACGCAAAGAAAGCGGCCTCCAAAAA  
**AGAAGAGAAAGGTA**ATGAAAGCGTTAACGGCCAGGCAACAAG

down:

AAGAGGAAGAAGGGTTGCCGCTGGTAGGTCGTGTGGCTGCCGGTGAACCTCTGTC  
GACGGCCCCCCCGAC

lexA-hrGFP

up: GACGGTACCACCATGGTGAGCAAGCAGATCCTGAAG

down: TACCTGGTACCCTATTACACCCACTCG

PsnRNAi

up: GACTCAGACTGAGGCTGCGGGTTTCACGCAAG

down: GACTCTAGAGGATTACAAAAAGCTTTGTTAGACTATGATACTG

Supplementary Table I. Partial amino acid sequence of the LexA-VP16 fusion constructs

Construct	Amino acid sequence
N <sup>LV</sup>	...NNLYISGGHQANKGSEAIYI - <b>MKALTARQQEVFDLIRDHIS</b> ...
N <sup>ΔEGF/LV</sup>	<u>...LLTLAFANLPNTVRGTDAL</u> - IYDANYPGWNGGSGSGNDRY...NNLYISGGHQANKGSEAIYI - <b>MKALTARQQEVFDLIRDHIS</b> ...
N <sup>S2ΔCT/LV</sup>	<u>...LPASLPLLLLTLAFA</u> – VRGIKNPGDEDNGEPPANVKYVITGIILVIIALAFFGMVLSTQRKR – <i>PPKKKRKV</i> - <b>MKALTARQQEVFDLIRDHIS</b> ...
APP <sup>LV</sup>	...NPTYKFFEQMQRN - <i>PPKKKRKV</i> - <b>MKALTARQQEVFDLIRDHIS</b> ...
SPA4CT <sup>LV</sup>	<u>MLPGLALLLLAAWTARADA</u> - DAEFRHDSGYEVHHQKLVFF...KMQQNGYENPTYKFFEQMQRN - <i>PPKKKRKV</i> - <b>MKALTARQQEVFDLIRDHIS</b> ...

The partial sequences of the Notch- and APP-LV fusions are shown with signal sequences (underlined), the inserted nuclear localization sequences (*italic*) and LexA sequences (**bold**). For N<sup>S2ΔCT/LV</sup> the complete sequence of the remaining extracellular and transmembrane domain is shown.

## References

Kidd, S., Lieber, T. and Young, M.W. (1998). Ligand-induced cleavage and regulation of nuclear entry of Notch in *Drosophila melanogaster* embryos. *Genes Dev.*, **12**:3728-3740.

Lee, Y.S. and Carthew, R.W. (2003). Making a better RNAi vector for *Drosophila*: use of intron spacers. *Methods*, **30**:322-329.

Lichtenthaler, S.F., Multhaup, G., Masters, C.L. and Beyreuther, K. (1999). A novel substrate for analyzing Alzheimer's disease gamma-secretase. *FEBS Lett.*, **453**:288-292.

Zhang, Y., Buchholz, F., Muyrers, J.P. and Stewart, A.F. (1998). A new logic for DNA engineering using recombination in *Escherichia coli*. *Nat. Genet.*, **20**:123-128.