

## **CDYL : a new transcriptional repressor**

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### **Supplementary Materials**

**Figure 1.** Selection of *Xenopus* cDNAs based on their homology to mouse sequences.

*Xenopus* (tadpoles) double stranded cDNAs fragments homologous to cDNA fragments from differentiated mouse B16 cells were isolated as described in materials and methods. **A)** An ethidium bromide-stained gel containing *Xenopus* cDNAs before selection (lane 1), and cDNAs obtained after the first and second rounds of selection (lanes 2 and 3 respectively) is shown. The gel was then transferred onto a membrane and probed with a fraction of “selection 2” cDNAs (lower panel). **B)** Homology between the mouse CDYL cDNA and one of the *Xenopus* cDNA fragments from “selection 2”. Numbers indicate the position of nucleotides in the *Xenopus* and mouse cDNAs respectively.

**Figure 2.** Colocalisation of CDYL with HDAC1 and HDAC2.

HeLa cells were transfected with Flag-HDAC1(left) or Flag-HDAC2 (right) expression vectors, without (upper) or with (lower) a Ha-CDYL expression vector. Twenty four hours post-transfection, the cells were fixed and the intracellular localisations of the Flag-HDACs and Ha-CDYL were monitored by immunofluorescence using anti-flag and anti Ha-antibodies.

### **Supplementary Materials and Methods**

Isolation of *Xenopus* cDNAs with sequences highly homologous to mouse sequences

One µg polyA<sup>+</sup> RNA isolated from *Xenopus* tadpoles (taken at stage 45) or from mouse B16 cells induced to differentiate for twenty four hours with 10 mM butyrate was used to synthesize double stranded cDNAs using an appropriate kit (Amersham) according to the vendor's instruction. Each cDNA was digested either with RsaI or with AluI in separate tubes. Digested

cDNAs were pooled and ligated to either *Xenopus* or mouse specific linkers. Linkers were as follows : *Xenopus*1, tagtccgaattcaagcaagagcaca; *Xenopus* 2, ctcttgcttgaattcggacta ; mouse 1, gcggtgaccgggagatctgaattc ; mouse 2, gaattcagatc. Non-ligated linkers were then removed and *Xenopus* and mouse cDNAs were PCR amplified using *Xenopus* 2 and mouse 1 primers respectively. Ten µg of amplified mouse cDNA were incubated with 3' terminal transferase and dATP (Boehringer) according to the supplier's recommendations. After phenol-chloroform extraction, the tailed cDNAs were mixed with 1 µg of amplified *Xenopus* cDNA in 10 µl hybridization buffer (750 mM NaCl, 25 mM hepes pH, 7.6, 5 mM EDTA and 0.1% SDS), denatured at 100°C for 10 min, and incubated at 68°C for 20 hours. These materials were then diluted and loaded onto an oligo dT column according to the standard protocol for the selection of poly A+ RNAs. Retained cDNAs were PCR amplified using the *Xenopus* 1 primer. One µg of this cDNA was used to hybridize with 10 µg of mouse poly A-tailed cDNAs as above. Another round of selection was performed and the final *Xenopus* cDNAs were cloned in the bluescript vector.

### **Immunolocalisation**

Immunolocalisation of HA-tagged CDYL and Flag-tagged HDACs were carried out as previously described (Lemercier *et al.*, 2002).

### **Reference**

Lemercier, C., Brocard, M.P., Puvion-Dutilleul, F., Kao, H.Y., Albagli, O. and Khochbin, S. (2002). Class II histone deacetylases are directly recruited by BCL6 transcriptional repressor. *J Biol Chem*, **277**, 22045-22052.



