Supporting Information: DNA Interstrand Cross-link Formation Initiated by Reaction Between Singlet Oxygen and a Modified Nucleotide.

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**General Methods.** Oligonucleotides were synthesized as previously described.<sup>1</sup> Radiolabeling was carried out according to standard protocols.<sup>2</sup> U.V. photoreactions of oligonucleotides were carried out in Pyrex tubes in a Rayonet photoreactor fitted with 16 lamps having an output maximum at 350 nm. Rose Bengal sensitized white light photolyses were carried out in Pyrex tubes using tungsten lamp (250 W) with a 400 nm cut off filter. All photoreactions and thermoreactions were carried out in 10 mM potassium phosphate (pH 7.2) and 100 mM NaCl. After reaction, samples were precipitated from 0.3 N NaOAc pH 5.3 and calf thymus DNA (0.1 mM bp). Pellets were dried, and then resuspended in formamide loading buffer and subjected to 20% denaturing PAGE analysis. Percent ISC was determined by dividing the integrated volume

for the cross-linked product band by the total integrated volume of DNA in the lane.

**D**<sub>2</sub>**O effect on the consumption of monomeric 2.** White light photolyses of monomer **2** (50  $\mu$ M) containing dA (5  $\mu$ M) as an internal standard were carried out in H<sub>2</sub>O, 50 % D<sub>2</sub>O and D<sub>2</sub>O as a solvent in the presence of Rose Bengal (10  $\mu$ M). Sample was analyzed by reverse phase HPLC (C<sub>18</sub>- $\mu$ Bondpak column (7.8 × 300 mm)) with UV detection at 260 nm. The peaks of interest (retention time: **2** (21.1 min), dA (8.7 min), Rose Bengal (14.3 min)) were quantified using the following gradient conditions: 5% to 35% MeCN in water, linearly for 25 min.

## <sup>1</sup>H NMR analysis of oxidation of monomeric 2 by NaIO<sub>4</sub> or Rose Bengal photosensitization

<sup>1</sup>H NMR experiment of the reaction of 2(50 mM) with NaIO<sub>4</sub> (50 mM) was carried out in deuterated phosphate buffer (50 mM, pD 7.4) at room temperature. Reaction was initiated by adding periodate solution at room temperature. Data were collected prior to periodate addition, 10 min after addition and after 24 h at room temperature. Rose Bengal photosensitization was carried out in the same manner, except that the sensitizer was present at 50  $\mu$ M, 2 (200  $\mu$ M). The sample was irradiated for 5 min and the spectrum was immediately collected.

- (1) Hong, I. S.; Greenberg, M. M. J. Am. Chem. Soc. 2005, 127, 3692-3693.
- (2) Maniatis, T.; Fritsch, E. F.; Sambrook, J. *Molecular Cloning*; Cold Spring Harbor Laboratory, Cold Spring Harbor, NY., 1982.



UV photoreaction: 30 min (350 nm) White light (rose bengal, 100 μM): 30 min NaIO<sub>4</sub> (5 mM): 10 min at 37 °C

**Supporting Information Figure 1.** Phosphorimage autoradiogram of **3** (20 nM) decomposed by UV photolysis, Rose Bengal sensitized photolysis, reaction with  $NaIO_4$ .



**Supporting Information Figure 2.** Phosphorimage autoradiogram of ISC products subjected to piperidine and IrCl<sub>6</sub> treatment. Arrows indicate cleavage bands resulting from alkali treatment.



**Supporting Information Figure 3.** Phosphorimage autoradiogram of the effect of GSH on ISC formation in **3** exposed to Rose Bengal photosensitized photolysis. Photolysis time: 30 min, [Rose Bengal] =  $100 \mu$ M.



**Supporting Information Figure 4.** <sup>1</sup>H NMR analysis of the photolysis of **2** (500  $\mu$ M) in deuterated phosphate buffer (50 mM, pD 7.4) by Rose Bengal (50  $\mu$ M) for 5 min. (A) Before photolysis (B) Immediately after photolysis.



**Supporting Information Figure 5.** Hydroxyl radical footprinting of ISC produced by reaction of **3** with  $NaIO_4$ . (A) Strand containing phenyl selenide **2** (B) complementary strand.



**Supporting Information Figure 6.** Sample phosphorimage autoradiogram of ISC formation in **A** and **B** (10 nM) exposed to Rose Bengal sensitized photolysis. Photolysis time: 30 min, [Rose Bengal] =  $50 \mu$ M.

Supporting Information Table 1. Rose Bengal mediated ISC formation.

DNA duplex	$O_2$	ISC (%)
Α	yes	$55.2\pm0.02$
Α	no	$3.4\pm0.6$
В	yes	$2.4\pm0.1$