

## Supplementary methods

### **DNA primers used for quantitative real-time RT-PCR analysis**

$\beta$ -tubulin (nucleotides 634-709)

5'-TTCCGCACCCTGAAACTGA / 5'-TGACGCCGGACACAACAG

18S rRNA (nucleotides 1525-1588)

5'-CGGAATGGCACCACAAGAC / 5'-TGGTAAAGTTCCCCGTGTTGA

ND4 (nucleotides 566-654)

5'-CAATCTGACCATTCCATGTGTGA /  
5'-TTTCAGCACAATACTTGCTAATAAAACA

ATP synthase subunit  $\alpha$  (nucleotides 1330-1388)

5'-GGCCAGCAGGTACAAACGAT / 5'-TTGAACAGCGCGACGAATC

ATP synthase subunit  $\beta$  (nucleotides 1298-1366)

5'-CGCGTAAGTTGGTGAAGTTCCT / 5'-GGCCAGTCATTCTGTGAAGA

### **DNA primers used to amplify complete coding sequences of ATP synthase subunits $\alpha$ , $\beta$ , and $\gamma$ for sequence comparisons**

subunit  $\alpha$

5'-TATTGGTGGCACGCTT / 5'-TGCTACCGGAGGCTAA

subunit  $\beta$

5'-GAAGAAGTACCCGGAG / 5'-CCACGCTTTAAACAGC

subunit  $\gamma$

5'-GCTTTACTGCGTACTCTTC / 5'-CGCATACCTACCGCATT

### ***K. lactis* strains use in this study**

Strain	Relevant genotype	Source
CW35	Mata, adeT-600,uraA1, atp3::kan	Clark-Walker, unpublished
CW35-ATP3	CW35 plus LEU2::pCXJ4- ATP3	This study
CW35-atp3-2	CW35 plus LEU2::pCXJ4- atp3-2(Ile281Thr)	(Clark-Walker <i>et al.</i> , 2000)
CW35-atp3-3	CW35 plus LEU2::pCXJ4- atp3-3 (Leu265Pro)	This study

### **Yeast strains and media**

The genotypes and sources of *K. lactis* strains are listed in Table 1. GYP medium contains 2% glucose, 0.5% Bacto yeast extract and 1% Bacto peptone. EB medium is GYP with 16 mg/ml ethidium bromide, Gly YP contains 2% glycerol in place of glucose and G418 medium is GYP with 200 mg/ml of the antibiotic. For solidification, 2% Bacto agar was added.

### **Yeast plasmids**

pTZ19-KIATP3 was constructed by insertion of a 1.8kb *Hinf*I fragment of *K. lactis* genomic DNA containing the ATP3 gene (Chen and Clark-Walker, 1995) into the *Sma*I site of pTZ19U (Pharmacia). To create pCXJ4-KIATP3, a 1.8kb fragment was excised from the above plasmid by *Sac*I/*Bam*HI digestion at flanking sites in the vector and inserted into the same restriction sites in pCXJ4. pCXJ4 is an integrative vector containing the URA3 gene of *Saccharomyces cerevisiae* and the LEU2 gene of *K. lactis* (X.J. Chen, unpublished).

### ***In vitro* mutagenesis of *K. lactis* ATP3**

The Leu 265 Pro change in *K. lactis* corresponding to the Leu 262 Pro change in *T. brucei* was produced as described (Clark-Walker *et al.*, 2000). An oligonucleotide, KIATP3PRO1, carrying TTA(Leu) to CCA(Pro), flanked by genomic sequence for 27 nucleotides upstream and 21 nucleotides downstream, together with a 17-mer universal primer 1, 5'-GTAAAACGACGGCCATG-3', downstream of the *Eco*R1 site in pTZ19U, were included in a PCR reaction in the presence of pTZ19-KIATP3 cut with *Bgl*II. The PCR product was isolated from a gel, blunt ended with T4 DNA polymerase and used as a mega primer in a second PCR reaction together with *Sac*I cut pTZ19-KIATP3 and a primer, KIMG15P7, 5'-GTGACAAGGTTAAGGGTC-3', upstream of the *Bgl*II site in ATP3. The resulting PCR product was cut with *Sac*I/*Bgl*II and used to replace the *Sac*I/*Bgl*II fragment in pTZ19-KIATP3 containing wild-type sequence. A plasmid containing the desired mutation was identified by sequence determination and called pTZ19Klatp3-3.

### **Expression of mutagenized ATP3 in *K. lactis***

To examine expression of *atp3-3* containing Leu265Pro, plasmid pCXJ4-Klatp3-3 was targeted to LEU2 by cleavage at a unique *Hpa*I site followed by transformation of *K. lactis* CW35 *atp3::kan*. Transformants were selected for Ura<sup>+</sup>, screened for stability, then examined for single copy integration by Southern blotting of *Hind*III digested DNA using 32P-labeled LEU2 as a probe. Details of these procedures have been described previously (Clark-Walker *et al.*, 2000). The phenotype of CW35 *atp3::kan* containing a single integrated copy of mutagenized ATP3 was examined for suppression of  $\rho^0/\rho^-$ -lethality by resistance to ethidium bromide and production of petite colony mutants.

### **ATPase activity of *K. lactis* strains**

Kinetic parameters of  $F_1$ -ATPase were determined with freshly prepared mitochondria as described (Clark-Walker, 2003).  $V_{\max}$  and  $K_m$  estimates were obtained from non-linear, least-squares curve fits generated with the Kaleidagraph software using the Michaelis-Menten equation.