# The Cross-Reactivity of Ketha Gum and Pneumococcal Type I — Short Cut to a Constituent of a Polysaccharide\*

M. Heidelberger and Jean M. Tyler

Rutgers, The State University, New Brunswick, New Jersey

AND

## S. MUKHERJEE

National Sugar Institute, Kanpur

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Summary. Although ketha gum has been reported to contain arabinose, galactose, xylose, an unidentified neutral sugar and glucuronic acid, its marked crossreaction with Type I antipneumococcal horse sera led to the conclusion that Dgalacturonic acid must be an additional component. The capsular polysaccharide of Type I pneumococcus, the determinant of immunological specificity, consists of more than 50 per cent of D-galacturonic acid. Chromatography of hydrolysates of ketha gum confirmed the presence of arabinose, galactose and xylose, and furnished spots with the mobilities of galacturonic and 4-O-methylglucuronic acids. Traces of glucuronic acid appeared only under conditions such that demethylation of the 4-O-methyl acid might have occurred. Hydrolysis of larger amounts furnished D-galacturonic acid, as yet syrupy,  $[\alpha]_D + 32^\circ$  instead of  $+51^\circ$ , and 4-Omethyl-D-glucuronic acid,  $[\alpha]_D + 20^\circ$  instead of  $+35^\circ$ . The latter, especially if present as non-reducing end-groups, would account for the strong precipitation of ketha gum in Type II antipneumococcal horse sera. When ketha gum was esterified, reduced and hydrolysed, the two acids disappeared almost entirely, a component corresponding to 4-O-methylglucose was observed, and the hexose content was increased. Oxidation of the gum with periodate, followed by reduction and hydrolysis, caused the disappearance of the 4-O-methylglucuronic acid and most of the galacturonic acid. Comparison of analyses and chromatograms of the original gum with those of samples recovered from the specific precipitates with Types I and II antisera showed little or no fractionation.

# INTRODUCTION

When the composition of ketha gum, the exudate of *Feronia elephantum*, family *Rutaceae*, was originally described (Mathur and Mukherjee, 1952, 1954) it seemed possible that it might precipitate with Type II antipneumococcal sera owing to the stated presence of glucuronic acid. Tests with a number of antipneumococcal (anti-Pn) sera (Heidelberger, 1960a) not only showed that it reacted as expected (also, Heidelberger, 1960b), but that

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it precipitated anti-Pn I even more heavily and regularly. Since the type-specific capsular determinant of Type I pneumococcus contains more than 50 per cent of D-galacturonic acid (Heidelberger, Kendall and Scherp, 1936; Smith, Galloway and Mills, 1960), this pointed to the occurrence of the acid as a major constituent of ketha gum, particularly as other gums, such as those of *Khaya grandifolia* (Aspinall, Hirst and Matheson, 1956) and *K. senegalensis* (Aspinall, Johnston and Stephen, 1960), which contain much D-galacturonic acid, had been shown to precipitate anti-Pn I strongly (Heidelberger, 1960a). The chemical investigation of ketha gum was accordingly taken up once more, with results which not only confirmed the immunochemical prediction of the presence of D-galacturonic acid, but which also led to an alternative explanation of the reactivity of the gum with anti-Pn II.

# MATERIALS AND METHODS

The anti-Pn I sera used were furnished by the New York City Department of Health Laboratories through the kindness of Miss Annabel W. Walter and of Dr. M. Solotorovsky (Rutgers) and by Dr. R. Wahl, of the Institut Pasteur, Paris. Quantitative microestimations of antibody nitrogen precipitated by ketha gum from the various antisera were carried out in duplicate at  $0^{\circ}$  as in previous papers (Heidelberger and Rebers, 1958; Kabat and Mayer, 1961) with the use of a cold-box for drainage of the tubes after centrifugation; determination of N in the washed precipitates was by the Markham (1942) method. The data are collected in Table 1 and experiments on the inhibition of precipitation

Pneumococcal	$\mu g$ . Antibody N precipitated at $o^{\circ}$ from serum no.					
polysaccharide	592	884	1057	Pasteur Inst. 173		
С	86	70	288	21		
Type I (SI)	650	70 965	1024	1900		
Ketha gum						
0.1 mg.	213	107				
0.3	265	203				
0.4	Ū.	Ū		185		
1.0	246*	324	417, 404	398		
2.0		338		398 398		
3.0		346	442 463			
4.0		334		373		
5.0			440			
% type-specific						
antibody precipitated	41	36	45	21		

TABLE I

PRECIPITATION OF KETHA GUM IN C-ABSORBED TYPE I ANTIPNEUMOCOCCAL HORSE SERA

\* Supernatants at this level gave 420 µg. N with SI; total, 666 µg. N.

by sodium galacturonate and sodium glucuronate are shown in Table 2. These also confirm the importance of D-galacturonic acid in the cross-reaction.

For chromatography, Whatman No. 1 or 3 mm. paper was used with the solvent systems A, pyridine-ethyl acetate-acetic acid-water, 5:5:1:3 (Fischer and Dörfel, 1955), B, n-butanol-pyridine-water, 6:4:3 (Jeanes, Wise and Dimler, 1951) and C, n-butanol-benzene-pyridine-water, 5:3:3:1 (upper phase) (Laidlaw and Reid, 1952). Reducing

sugars were detected with aqueous saturated aniline oxalate to which acetic acid was added for ionophoretograms. Ionophoresis on 3 mm. paper (Foster, 1952) was carried out with 0.05 M borate at pH 10 at 500 V for 5 hours. Hexose (as galactose) was determined by the modified primary cysteine reaction (Dische, 1955) and by a modified secondary cysteine reaction (Dische and Danilchenko, 1962), pentose (as arabinose) by the modified basic cysteine reaction (Dische, 1949; Heidelberger, Adams and Dische, 1956) and uronic acid (as galacturonic acid) by the carbazole method (Dische, 1947). Specific rotations were measured in water at  $23^{\circ}$ .

TABLE	2
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Inhibitor		Total vol. (ml.)	Antibody N precipitated from		
	μ mol. added		I 592C 0.2 ml.	I 1057F,* 0.1 ml.	Inhibition (%)
			$(\mu g.)$	$(\mu g.)$	
None		0.4	33		
Na galacturonate	3.1	,,	II		67
,, ,,	0.3	,,	30		9 6
" pectate	(200 µg.)	,,	31		6
None		,,		40	
Na galacturonate	6.2	,,		18	55
,, ,,	0.6	,,		(35)†	55 (13)
None			30		
Na glucuronate	32.0	0.5	24		20
,, ,,	6.4	,,	24 28		7

INHIBITION OF PRECIPITATION OF KETHA GUM IN TYPE I ANTIPNEUMOCOCCAL SERA BY GALACTURONATE AND GLUCURONATE

\* Partially purified Felton antibody solution, at same anti-SI concentration as whole serum (Table 1).

† One determination only.

There was no inhibition of the SI-1057F reaction by  $6.2 \mu$  mol. Na galacturonate, nor of the SI-592C reaction by  $32 \mu$  mol. Na glucuronate.

## RESULTS

#### CHROMATOGRAPHIC EXAMINATION OF THE HYDROLYSATES OF KETHA GUM

The gum, 20 mg., was hydrolysed in sealed tubes at 100° with 2 ml. of 2N H<sub>2</sub>SO<sub>4</sub> for 5 and 10 hours. The hydrolysates were neutralized with BaCO<sub>3</sub> and the filtrates and washings evaporated to dryness and examined for neutral sugars with solvent *B*. Galactose and arabinose were found to be major components, xylose a minor component, and glucose was absent. Rhamnose was either absent, or present only in traces. No neutral oligosaccharides were seen. When the 5 hours' hydrolysate was deionized with Dowex 50W-X2(H) and examined for free acids in solvent *A* galacturonic acid was found, also a small proportion of glucuronic acid (glucurone was not detected on a paper run for 7 hours), a component with R<sub>G1u A</sub> 1.47 and traces of three components, presumably aldobiuronic acids, with R<sub>G1u A</sub> values 0.51, 0.59 and 0.64, which had disappeared from the 10 hours' hydrolysate. All components except the traces of glucuronic acid and aldobiuronic acids were also detected when 3 mg. of gum was similarly hydrolysed for 4 hours.

### LARGER-SCALE HYDROLYSIS OF KETHA GUM WITH ISOLATION OF THE URONIC ACIDS

1.5 g. gum was hydrolysed for 4 hours with  $2N H_2SO_4$  at  $95-100^\circ$ . The solution was cooled, neutralized with BaCO<sub>3</sub>, centrifuged and the residue washed twice with water. The combined supernatants were evaporated to dryness and treated with 25 ml. water and 25 ml. ethanol. Insoluble Ba salts were centrifuged off and the supernatant was evaporated to a syrup, taken up in 15 ml. water and passed through a column of Amberlite IR-120 (H). The eluate and washings were evaporated to small volume and applied to a column of Duolite A4(OH), to retain acidic material. The column was washed until the Molisch test was negative. When evaporated to dryness the eluate and washings gave a foam, 0.79 g., which on chromatography in solvent C showed galactose, arabinose and xylose, with possible traces of rhamnose. Uronic acids were eluted from the resin with 0.5N NaOH and the free acids obtained by passage through IR-120 (H) and evaporation of the eluate to dryness. Neutral sugars were absent (solvent C). In solvent A, galacturonic acid. identical with an authentic sample, and an acid with  $R_{Glu A}$  1.49 were the main components, together with slower-moving products. No glucuronic acid was present, and, indeed, this acid appeared only under conditions such that demethylation of 4-O-methylglucuronic acid might have occurred (Croon, Herrström, Kull and Lindberg, 1960).

Separation of the two main acidic components on paper with solvent A gave 26 mg. of Fraction 1 and 18 mg. of Fraction 2. Fraction 1 had  $[\alpha]_D + 26^\circ$  (c, 1.29), and was chromatographically identical with D-galacturonic acid. The methyl glycoside of the methyl ester was prepared but also failed to crystallize. Fraction 2,  $[\alpha]_D + 20^\circ$  (c, 0.89),  $(+35^\circ, Bowering and Timell, 1960)$ ,  $R_{Glu A}$  1.49, was chromatographically identical with authentic 4-O-methyl-D-glucuronic acid (kindly supplied by Dr. E. E. Percival).

Similar hydrolysis and fractionation of a second 2 g. yielded 17 mg. of galacturonic acid with  $[\alpha]_D + 32^\circ$  (c, 0.85), (lit., +51°) m.p. 168–171° (decomp.), which afforded *p*bromophenyl-hydrazine *p*-bromophenylhydrazone galacturonate of m.p. and mixed m.p. 145–146° (decomp.) with an authentic sample of m.p. 145–146° (decomp.) (Niemann, Schoeffel and Link, 1933).

Material from an additional hydrolysis yielded a galacturonic acid-containing fraction which was fractionated on charcoal-Celite (1:1) (Whistler and Durso, 1950) and further purified by paper chromatography. The product, 10 mg., had  $[\alpha]_D + 24^\circ$  (c, 0.59). Its methyl ester methyl glycoside, reduced with NaBH<sub>4</sub>, hydrolysed and chromatographed in solvent C, gave a strong spot with the R<sub>1</sub> of galactose and several other faint spots.

Reduction and hydrolysis (Hamilton and Smith, 1956) of periodate-oxidized ketha gum followed by chromatography showed that, of the acid components, the 4-O-methylglucuronic acid had been destroyed and that only minor amounts of galacturonic acid remained.

#### ESTERIFICATION AND REDUCTION OF KETHA GUM WITH IDENTIFICATION OF 4-O-METHYLGLUCOSE

The gum, 108 mg., was converted to the methyl ester methyl glycoside by refluxing for 5 hours with 10 ml. 1.5 per cent methanolic HCl and, after neutralization with Ag<sub>2</sub>CO<sub>3</sub>, the isolated product was reduced with NaBH<sub>4</sub>, 30 mg. Sodium was removed by Dowex 50W-X<sub>2</sub>(H) and borate by repeated distillation with methanol. The product, 58 mg., was re-esterified and again reduced. Chromatographic examination of a 4-hour hydrolysate of an aliquot in solvent *B* showed in addition to galactose, arabinose, and xylose, R<sub>Rh</sub> 0.74, a new spot with R<sub>Rh</sub> 0.89, which was identical with authentic 4-O-methylglucose. After treatment

with Dowex 50(H), and examination in solvent A some galacturonic acid was still found. The bulk of the product was hydrolysed but only a trace of 4-O-methylglucose was recovered after fractionation on paper with solvent B. The sugar had  $M_G$  0.30 (Foster, 1953) and after periodate oxidation (Lemieux and Bauer, 1953) gave the yellow spot,  $R_t$  0.6 (approx.), characteristically produced by a 4-O-methylhexose. Chromatography of the recovered galactose in solvent C, using multiple development ( $3 \times 12$  hours), showed that it was free of glucose.

# ATTEMPT TO FRACTIONATE KETHA GUM BY PRECIPITATION WITH ANTI-PN I AND ANTI-PN II

Fifty ml. of type I C-absorbed serum 592 was diluted with 50 ml. of saline at 0° and 50 mg. of ketha gum in 2.5 ml. saline was added (cf. Table 1). After several days at 0° the precipitate was centrifuged, washed several times with chilled saline and finally with ice-cold water, and dissociated with 5 per cent trichloroacetic acid (Heidelberger, Dische, Neely and Wolfrom, 1955). The dissolved polysaccharide was precipitated in the cold with ethanol, washed with ethanol and acetone, and dried over  $P_2O_5$  in vacuo. The recovered polysaccharide, 15 mg., K I, had  $[\alpha]_D + 5^\circ$  (c, 0.41) (cf. Table 3).

TABLE	3
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ANALYSIS OF KETHA GUM, ITS FRACTIONS PRECIPITATED BY ANTISERA AND ITS PRODUCT OF REDUCTION

Sample	[¤] <sub>D</sub> in water	as % an gala		Pentose, as % anhvdro- arabinose (Dische)	Uronic acid, as % anhydro- galacturonic acid (Dische)	Total anhydro- sugars, (%)
K K I K II K Red	$+4^{\circ}$ $+5^{\circ}$ $+2^{\circ}$	24 29 44	35 33 35 52	36 34 34 29	30 25 28 9	101 92 97 90

\*By primary cysteine-sulphuric acid colour-reaction.

+By secondary cysteine-sulphuric acid colour-reaction. These values were used in computing totals.

Ketha II, 15 mg., was similarly obtained from the specific precipitate given by 23 ml. of Type II antiserum 513 (Heidelberger, 1960b) and had  $[\alpha]_D + 2^\circ$  (c, 0.56) (cf. Table 3).

Ketha I and ketha II were heated in  $2N H_2SO_4$  at 100° for 6 hours and worked up as before. Chromatograms of both in solvent A showed galacturonic acid, 4-O-methyl-glucuronic acid, and traces of glucuronic acid, possibly heavier in ketha II. There was thus no obvious fractionation.

# DISCUSSION

The experiments described in the present paper demonstrate once more how immunochemical methods may point unerringly to the nature of an unknown constituent of a complex polysaccharide. In this instance it was anticipated that ketha gum, said to contain glucuronic acid, would show cross-precipitation in Type II antipneumococcal sera. Unexpected, however, was the consistent reactivity in Type I antipneumococcal

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sera and, as this involved 20 to 45 per cent of the type-specific antibody, it was evident that ketha gum contained multiples, in terms of the quantitative precipitin theory of Heidelberger and Kendall (1935), of a major antigenic grouping in common with the specific capsular polysaccharide of Type I pneumococcus, the determinant of Type I specificity. Since more than one-half of this substance consists of D-galacturonic acid and sugar acids, like other organic acids (Landsteiner, 1945), have been shown to be major determinants of immunological specificity (Goebel, 1936; Goebel and Hotchkiss, 1937; Marrack and Carpenter, 1938), it was a foregone conclusion that D-galacturonic acid would be found as a component of ketha gum. Its identification among the products of hydrolysis is outlined above, as is also that of 4-O-methyl-D-glucuronic acid, which, instead of the previously reported glucuronic acid, appears to be the other principal acidic constituent and is responsible for the cross-reactivity of the gum in Type II antipneumococcal sera. An O-methyl group in the 4-position of D-glucuronic acid, as in the hemicelluloses, does not prevent cross-precipitation of such substances in Type II antipneumococcal sera (Heidelberger, 1960 a, b), although the D-glucuronic acid is unsubstituted in the specific capsular polysaccharide of Type II pneumococcus, the determinant of Type II specificity.

Studies with other gums which precipitate Type I antipneumococcal sera will appear in another communication and work is in progress on the constitution of the Type I pneumococcal polysaccharide.

#### REFERENCES

- ASPINALL, G. O., HIRST, E. L. and MATHESON, N. K. (1956). 'Plant gums of the genus Khaya. The structure of Khaya grandifolia gum.' J. chem. Soc., 989-97.
- GOG-GY.
  ASPINALL, G. O., JOHNSTON, MARGARET J. and STEPHEN, A. M. (1960). 'Plant gums of the genus *Khaya*. Part II. The major component of *Khaya* senegalensis gum.' J. chem. Soc., 4918-27.
  BOWERING, W. D. S. and TIMELL, T. E. (1960). 'A chemical synthesis of 4-O-methyl-D-glucuronic acid.' Constal J. Chem. of A. L. L.
- Canad. J. Chem., 38, 311-12. CROON, I., HERSTRÖM, G., KUIL, G. and LINDBERG, B. (1960). 'Demethylation and degradation of sugars in acid hydrolysis.' Acta chem. scand., 14, 1338-42.
- in acid hydrolysis.' Acta chem. scand., 14, 1338-42. Dische, Z. (1947). 'A new specific color reaction of hexuronic acids.' J. biol. Chem., 167, 189-98. Dische, Z. (1949). 'Spectrophotometric method for the determination of free pentose and pentose in nucleotides.' J. biol. Chem., 181, 379-92. DISCHE, Z. (1955). In Glick, D., Meth. biochem. Anal. 2,
- 327.
- DISCHE, Z. and DANILCHENKO, A. (1962). 'Modifications of two color reactions of hexoses with cysteine
- and sulphuric acid.' Arch. Biochem. Biophys. (In press.) FISCHER, F. G. and DÖRFEL, H. (1955). 'Die papier-chromatographische Trennung und Bestimmung der

- chromatographische Frennung und Destimmung der Uronsäuren.' Hoppe-Seyl. Z., 301, 224-34.
  Foster, A. B. (1952). 'A simplified apparatus for filter paper ionophoresis.'Chem. and Ind. (Rev.), 1050-1.
  Foster, A. B. (1953). 'Ionophoresis of some di-saccharides.' J. chem. Soc., 982-6.
  GOEBEL, W. F. (1936). 'Chemo-immunological studies on conjugated carbohydrate-proteins. X. The immunological properties of an artificial The immunological properties of an artificial

antigen containing glucuronic acid.' 7. exp. Med., 64, 29-38

- GOEBEL, W. F. and HOTCHKISS, R. D. (1937). 'XI. The specificity of azo protein antigens containing glucuronic and galacturonic acids.' J. exp. Med., 66, 191-205.
- HAMILTON, J. K. and SMITH, F. (1956). 'Reduction of the products of periodate oxidation of carbohydrates. II. A new method for the end-group assay of
- II. A new method for the end-group assay of amylopectin.' J. Amer. chem. Soc., 78, 5907-9.
  HEIDELBERGER, M. (1960a). 'Structure and immunological specificity of polysaccharides.' Progress in the Chemistry of Organic Natural Products, vol. XVIII, pp. 503-36, Springer-Verlag, Wien.
  HEIDELBERGER, M. (1960b). 'The immunological specificity of type II pneumococcus and its separation into partial specificities. II.' J. exp. Med., 111, 33-43.
  HEIDELBERGER, M., ADAMS, J. and DISCHE, Z. (1956). 'Fractionation of gum arabic by chemical and
- 'Fractionation of gum arabic by chemical and immunological procedures.' J. Amer. chem. Soc., 78, 2853-5.
- HEIDELBERGER, M., DISCHE, Z., NEELY, W. B. and WOLFROM, M. L. (1955). 'Immunchemistry and the structure of lung galactan.' J. Amer. chem. Soc., 77, 3511-14.
- HEIDELBERGER, M. and KENDALL, F. E. (1935). 'The precipitin reaction between Type III pneumococcus polysaccharide and homologous antibody. III.
- Polysacchardte and homologous antibody. III.
  A quantitative study and a theory of the reaction mechanism.' J. exp. Med., 61, 563-91.
  HEIDELBERGER, M., KENDALL, F. E. and SCHERP, H. W. (1936). 'The specific polysaccharides of types I, II, and III pneumococcus. A revision of methods and data.' J. exp. Med., 64, 559-72.

- HEIDELBERGER, M. and REBERS, P. A. (1958). 'Crossreactions of polyglucoses in antipneumococcal sera. VI. Precipitation of type VIII and type III antisera by β-glucans.' *J. Amer. chem. Soc.*, **80**, 116-18.
- by β-glucans.' J. Amer. chem. Soc., 80, 116-18. JEANES, A., WISE, C. S. and DIMLER, R. J. (1951). 'Improved techniques in paper chromatography of carbohydrates.' Analyt. Chem., 23, 415-20.
- KABAT, E. A. and MAYER, M. M. (1961). Experimental Immunochemistry, 2nd ed. Charles C Thomas, Springfield, Ill.
- LAIDLAW, R. A. and REID, S. G. (1952). 'Analytical studies on the carbohydrates of grasses and clovers. I.—Development of methods for the estimation of the free sugar and fructosan contents.' *J. Sci. Agric.*, 3, 19-25.
- LANDSTEINER, K. (1945). The Specificity of Serological Reactions. Harvard Univ. Press, Cambridge, Mass. LEMIEUX, R. U. and BAUER, H. F. (1953). 'A method for
- LEMIEUX, R. U. and BAUER, H. F. (1953). 'A method for the identification of the mono-O-methyl glucoses.' *Canad. J. Chem.*, **31**, 814-20.
- Canad. J. Chem., 31, 814-20. Маккнам, R. (1942). 'A steam distillation apparatus suitable for micro-Kjeldahl analysis.' Biochem. J., 36, 790-1.

- MARRACK, J. and CARPENTER, BLANCHE R. (1938). 'The cross reactions of vegetable gums with Type II antipneumococcal serum.' Brit. J. exp. Path., 19, 53-65.
- MATHUR, G. P. and MUKHERJEE, S. (1952). 'Investigations on the structure of ketha (Feronia elephantum: Family Rutaceae) gum.' J. sci. industr. Res., 11B, 544-5.
- MATHUR, G. P. and MUKHERJEE, S. (1954). 'A new aldobiouronic acid 3-(glucuronopyranosyl)-D-galactopyranose from ketha (*Feronia elephantum*) gum.' *J. sci. industr. Res.*, 13B, 452-3.
- J. sci. industr. Res., 13B, 452-3. NIEMANN, C., SCHOEFFEL, E. and LINK, K. P. (1933). 'Substituted hydrazine derivatives of the hexuronic acids.' J. biol. Chem., 101, 337-58. SMITH, EVELYN E. B., GALLOWAY, B. and MILLS, G. T.
- SMITH, EVELYN E. B., GALLOWAY, B. and MILLS, G. T. (1960). 'The purification and composition of type I pneumococcal capsular polysaccharide.' *Biochem. J.*, 76, 35P.
- WHISTLER, R. L. and DURSO, D. F. (1950). 'Chromatographic separation of sugars on charcoal.' J. Amer. chem. Soc., 72, 677-9.