

**Supplemental Figure 1: Accumulation of transgenic *ndhA* transcripts in lines SylCEE-2.8 and 10.2**

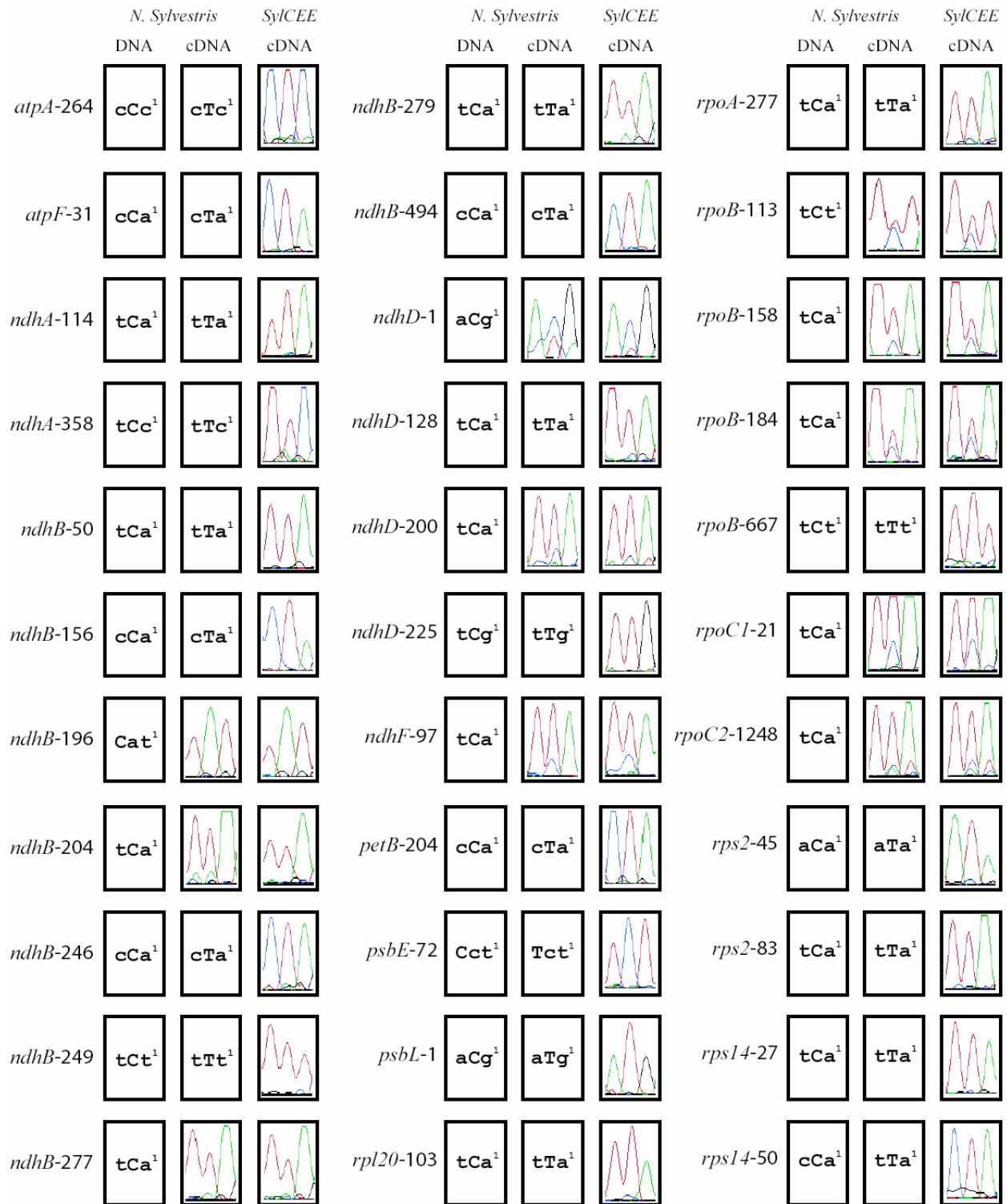
Five micrograms of total plant RNA from the indicated plant lines were analysed by RNA blot hybridization using strand-specific *aadA*, *psbL/J* and *ndhA* probes. Arrow heads = monocistronic *aadA*/CEE transcripts. Note the stronger signal intensity of the band corresponding to this transcript in line SylCEE-2.8 in comparison to line 10.2. Quantification of the signals for the monocistronic *aadA* transcripts indicated a 4-fold higher amount of these transcripts in SylCEE-10.2. Importantly, the difference in transcript levels is accompanied by the differences in editing efficiency in the two lines (Fig. 1D). Transcripts of ~2.1 kb length detected in all three

hybridization experiments (marked by asterisks) result from read-through transcription initiating at the *psbE* promoter (Bock et al., 1994). Methylene blue stainings of the 16S rRNA shown below each panel demonstrate comparable sample loading in all lanes.

CEE <i>ndhA</i> -189	ATCTCT- <b>ACTATCTAATA</b> -GTTCAA
tobacco <i>ndhF</i> -97:	CC <b>CACTTACT</b> - <b>TCTATTATG</b> -TCAA

**Supplemental Figure 2: Homology of the introduced spinach *ndhA* editing site to the endogenous tobacco *ndhF* site.**

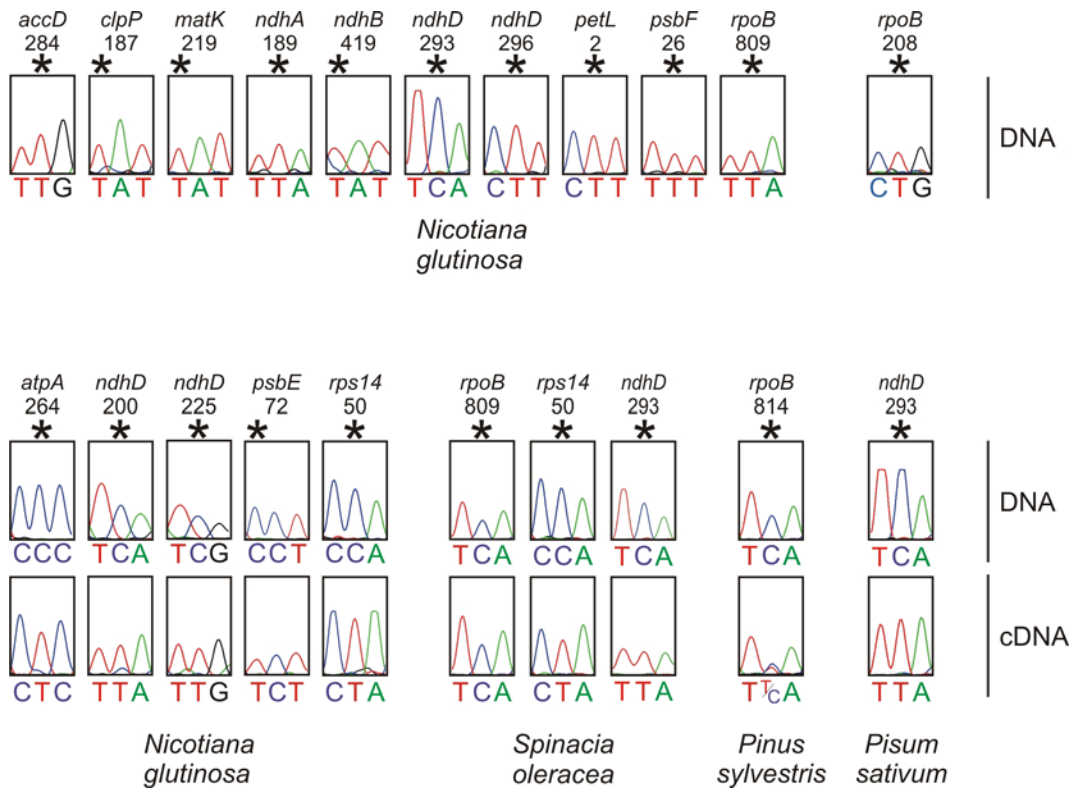
The nucleotide regions surrounding the described editing sites of *N. tabacum* and *N. sylvestris* were extracted and subsequently compared with the 20 nt. upstream of CEE *ndhA*-189. Allowing gaps (“-“), the highest similarity was found for site *ndhF*-97 (editing site: bold “C”, identical nt labelled in grey). However, the fact that editing of *ndhF*-97 is not reduced in either *N. tabacum* (Schmitz-Linneweber et al., 2001) or *N. sylvestris* plants (Supplementary Figure 4) overexpressing CEE *ndhA*-189 argues against the two sites being co-edited by the same factors.



**Supplemental Figure 3: Analysis of all known editing sites in SylCEE plants.**

Analysis of all known editing sites from the SylCEE transcriptome according to Sasaki *et al.* (2003). Editing status was determined by sequencing cDNA-derived PCR products covering the respective areas of interest. Where partial editing was found, complementary data from wild-type

*N. sylvestris* plants were obtained as well. Altogether, no significant reduction in editing relative to wild-type was observed in SylCEE plants.



**Supplemental Figure 4: Test for the presence and processing status of several plastid editing sites in different plant species.**

Sequence analysis of *Nicotiana glutinosa* DNA. The chromatogram excerpts show the indicated codon of the given gene. Asterisks mark the position of editing sites known from different angiosperm species and tested for their presence and – if necessary - processing status in species indicated below the chromatograms. The sites analysed in *N. glutinosa* are absent from *Nicotiana tabacum*. For cDNA analysis of *ndhD*-293 in *N. glutinosa* see Fig. 2A.

## Supplemental Table 1: Oligonucleotides

Plant(s)	Target	Name	Sequence	Purpose
<i>SylCEE</i>	ORF103	P1a	CACACAATTTAAGTAGATGCG	check of integration at left border
	<i>aadA</i>	P1b	ACTGCGGAGCCGTACAAATG	
	<i>aadA</i>	P2a	CTGTTCTTATTTTACCGGAGG	check of Integration at right border
	<i>petA</i>	P2b	CTTTCCAAAAGCTTCAATCTCTCGCATTCTG	
	<i>psbL</i>	ApsbL	TACGACACAATCAAACCCGA	probe <i>psbL/J</i>
	<i>psbJ</i>	AT7psbJ	GATAATACGACTCACTATAGGGTACTAGAGGGATGAACCCAAT	
	<i>aadA</i>	aadAli	GAAGCGGTTATCGCCGAAG	probe <i>aadA</i>
	<i>aadA</i>	aadAT7	GTAATCGACTCACTATAGGGAACCGGATCAAAGAGT	
	<i>ndhA</i>	ndhAfor	ACAGGAGATACTCGTTTATT	probe <i>ndhA</i>
	<i>ndhA</i>	AT7ndhA	GATAATACGACTCACTATAGGGACGAGGTTGTCAATAATAGAT	
	<i>aadA</i>	P2a	see above	amplification and sequencing of introduced <i>ndhA</i> editing site
	CEE	hindndhAex2rev	CTTTCCAAAAGCTTCAATCTCTCGCATTCTG	
<i>Syl CEE and Nicotiana sylvestris</i>	<i>atpA</i>	EcoatpAfor	TTCCAGAATTCACATTACAATACCTTGCTCC	amplification and sequencing of <i>N. sylvestris</i> and <i>N. sylvestris</i> cDNA
		Ecoatparev	TTCCAGAATTTCTTTCCAAAAGGCGTGAATGC	
	<i>atpF</i>	atpFforP	ATGAAAAACGTAACCGATTCC	
		atpFrevP	TTGTCGGACTTGATTAATCG	
	<i>ndhA</i>	ndhAfor	ACAGGAGATACTCGTTTATT	
		ndhAntex2173 rev	TGCTACTAATTCTTCTTGCTTCTG	
		tommunedfor	GATTATCTAACAGTTTAAGTACAGTTGATATAG	
		ndhA3' rev	CTTGTTTGAGACAAGTCGTG	
	<i>ndhB</i>	ndhBfor2	TCATGATCTGGCATGTACAG	
		ndhBedIII	ATTTCTTGAAGCTCAATCTCTCCCCGGAT	
		nb11	TTCATGCTTGTTTGAGTAATAGC	
		ndhBP12	GATATACCAAGAAAGATGTACG	
		AndhB	GTCGTTGCTTTTCTTTCTG	
		ndhB3for	TCTCTTATCCCTAGGAGGTC	
		ndhBrev2	CTAAAAAAGGCTATCCTGAGC	
		<i>ndhD</i>	ndhDStart5'	GGTCCAAGTGATCTTGTC
	Zagreb7		GCTCCCATTTTTAATAAAAATTCC	
	ndhDMitterev		CTAAAGTAGTGATAAATCCTG	
	<i>ndhF</i>	ndhFforP	GGAACAGACATATGAATATG	
		ndhFrevP	ATCCCAACATGGAAGTACTG	
	<i>petB</i>	petBPfor	TTCTGTAAATAGGATCACCC	
		petB3' revP	AATGATTGATTACAAATATTTAG	
	<i>psbE/L</i>	ApsbE	AAGCACAGGAGAACGTTCTG	
		AT7psbL	GATAATACGACTCACTATAGGGTCTTAATTGAAGAAATAATTGGA	
	<i>rpl20</i>	rpl20edfor	ATTCGCGAAAGGGGAGTATC	
		rpl20rev3	ATTTAAATTTATTCTGGTGG	
	<i>rpoA</i>	ArpoA	GGATTACAAATAGAGAGAAAAT	
		AT7rpoA	GTAATACGACTCACTATAGGGATTCAAAGGTTCCAACAATGT	
	<i>rpoB</i>	rpoBforP	GGAAAAACAGTAGGGATATGC	
		rpoBrevP	GAAAAATACCGGATCGCCACC	
		rpoBfor2	TCTTTTAGCAGGTAATGGAG	
		rpoCb4	GGTTCAAATACCCATGGATT	
	<i>rpoC1</i>	rpoC15' for	AATAATTTTCTTCTATGATCG	
rpoC15' rev		AAATAATCCATCTTTTCCG		
<i>rpoC2</i>	ArpoC2	AGCTAAGCCTTATTTGGCCAC		

		AT7rpoC2	GTAATACGACTCACTATAGGGCTCGCTTCAGATATGAAACTTTG	
<i>rps2</i>		rps2forP	AATGGAATCCTAAAATGGCG	
		rps2revP	CTGGAATTGAAATATCTGCG	
<i>rps14</i>		Arps14	ATGGCAAGGAAAAGTTTGAT	
		AT7rps14	GTAATACGACTCACTATAGGGCCTTACCAACTTGATCTTGTT	
<i>Solanaceans and cybrids</i>	<i>ndhD</i>	ndhDrev	CTCCATTAAGTCCCGTGTCG	editing of <i>ndhD</i> -293
		ndhDmittefor	CTAAAGTAGTGATAAATCCTG	
	<i>ndhA</i>	ndhA.nt.ex2.173. rev	TGCTACTAATTCTTCTTCTGCTTCTG	editing of <i>ndhA</i> -189
		ndhAfor	ACAGGAGATACTCGTTTATT	
<i>Nicotiana glutinosa</i>	<i>accD</i>	psbL14	TTCTCCGAATGAAATTCAAT	amplification and sequencing of <i>N. glutinosa</i> plastid DNA
		accDrev	AATGGGATCTAGAGAGACC	
		accDfor2	TAATGATCTCGAGGTAACCT	
		AT7accD	GTAATACGACTCACTATAGGGTGATTTTCTCTCCGACTAC	
	<i>atpA</i>	atpAMittefor	GAGATCAGAAGCAAATTGTGC	
		EcoatpAfor	TTCCAGAATTCACATTACAATACCTTGCTCC	
		EcoatpArev	TTCCAGAATTCTTTCCAAAAGGCGTGAATGC	
	<i>clpP</i>	clpPtKo	GGGTTGGTTTAGATTGATCC	
		clpPex2for	GTGCGACCAGATGTCCATAC	
		clpP.AT.for	GTAATGATCCATCAACCCGC	
		clpP.AT.2rev	TGAACCGCTACAAGATCAAC	
	<i>matK</i>	AbmatKfor	CTTTATCTTCTTTTGAAGGC	
		K7	AAAAATATCCAAATACCAA	
	<i>ndhA</i>	ndhA.nt.ex2.173. rev	TGCTACTAATTCTTCTTCTGCTTCTG	
		ndhA.nt.in.174. rev	CCTTTTTACTGCAAATTTAGAAGCCG	
		Tommunedfor	GATTATCTAACAGTTTAAGTACAGTTGATATAG	
		ndhA3'rev	CTTGTTTGAGACAAGTCGTG	
		ndhAfor	ACAGGAGATACTCGTTTATT	
	<i>ndhB</i>	AndhB	GTCGTTGCTTTTCTTCTG	
		ndhBrev2	CTAAAAAAGGCTATCCTGAGC	
		ndhB3for	TCTCTTATCCCTAGGAGGTC	
	<i>ndhD</i>	ndhDstart5'	GGTCCAAGTGATCTTGTG	
		ndhDrev	CTCCATTAAGTCCCGTGTCG	
		Zagreb7	GCTCCCATTTTTAATAAAAATTCC	
		ndhDMittefor	AGTACTTGATGCTTCTAGC	
		ndhDrev	CTCCATTAAGTCCCGTGTCG	
		Zagreb6	GAATTTTATTAATAAATGGGAGCG	
	<i>petL</i>	petL5'forP	ATTTGAAACTTAGGTAAGTGC	
		AT7petL	GATAATACGACTCACTATAGGGAATCTCAATGACAATGAACA	
		AT7petG	GATAATACGACTCACTATAGGGTAAATTAATCAAAGGTCCAA	
	<i>psbF</i>	ApsbE	AAGCACAGGAGAACGTTCTG	
		AT7psbL	GATAATACGACTCACTATAGGGTCTTAATTGAAGAAATAATTG GA	
	<i>rpoB</i>	rpoBforP	GGAAAAACAGTAGGGATATGC	
		rpoBrevP	GAAAATACCGGATCGCCACC	
		rpoBfor2	TCTTTTAGCAGGTAATGGAG	
		rpoCb4	GGTTCAAATACCCATGGATT	
	<i>rps14</i>	Arps14	ATGGCAAGGAAAAGTTTGAT	
		AT7rps14	GTAATACGACTCACTATAGGGCCTTACCAACTTGATCTTGTT	